Sapphire manual

1. Introduction

This document provides a manual of the software named Sapphire which is a part of DIAMonDs (Drosophila Individual Activity Monitoring and Detection System). The software is a web application working on Dash framework in Python. Installation and usage of Sapphire is described as follows.

2. System requirement

- Any OS (Ubuntu 16.04/18.04, OS X 10.11 El Capitan, Windows 7 are validated)
- Google chrome (web browser)
- Python 3.6

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3-1. Python environment

Sapphire requires Python as base environment. Some operation systems such as Ubuntu and MacOSX includes Python, however, Anaconda or Miniconda is recommended because it provides convenience in creation and management of multiple virtual environments.

3-1-1. Installation of Python

Python could be automatically installed by installation of Anaconda or Miniconda.

Anaconda:

http://www.anaconda.com

Miniconda:

https://docs.conda.io/en/latest/miniconda.html

3-1-2. Creation of virtual environment by Anaconda

To create a new environment, you can type at the command in Terminal:

```
conda create -n sapphire python=3.6
```

The command specify that the name of the environment is sapphire and the version of Python is 3.6. The latest version of Sapphire requires Python 3.6 or later version.

You can activate the environment by typing:

```
source activate sapphire
```

Then, your terminal prompt shows like this:

```
(sapphire) your_machine:your_directory username$
```

3-1-3. Installation of modules

Modules are installed by conda install command.

(Example 1) Install of Numpy

conda install numpy

(Example 2) Install of Numpy and Scipy (multiple install)

conda install numpy scipy

(Example 3) Install of Numpy ver. 1.15.0 (version specification)

```
conda install numpy==1.15.0
```

(Example 4) Installation of dash (and related modules).

Execute the install command specifying the version:

```
conda install -c conda-forge dash==0.43.0
```

3-1-4. Module dependencies

The application requires Python 3.6.

It also needs Python modules as described bellow.

Following minimum modules are required if you want to use minimum functions of Sapphire such as viewing images, signals, etc (Table 1).

Module name	Version	Description		
dash	0.43.0	You have not to install following modules because		
		it is automatically installed with dash.		
		\cdot dash-core-component		
		\cdot dash-html-component		
		\cdot dash-renderer		
		\cdot dash-table		
		· plotly		
dash-core-components	0.48.0			
dash-html-components	0.16.0			
dash-renderer	0.24.0			
dash-table	3.7.0			
numpy	1.16.5			
pandas	0.25.1			
Pillow	6.1.0			
plotly	3.2.0			
scipy	1.3.1			

Table 1. Minimum module requirement of Sapphire

Additional modules are required if you want to advanced functions of Sapphire such as the inference and/or further analysis with the scripts in this repository (Table 2).

Module name	Version	Description		
changefinder	0.3	Please use "pip install" command if you are using Anaconda		
		or Miniconda environment, because this is not provided		
		in Anaconda repository at this point (8th Apr 2020).		
cudatoolkit	9.0	Automatically installed with install of tensorflow-gpu		
cudnn	7.6.0	Automatically installed with install of tensorflow-gpu		
keras	2.2.4			
tensorflow/tensorflow-gpu	1.9.0	You can accelerate inference of a neural network by		
		GPU computing. In this case, please install tensoflow-gpu,		
		not tensorflow. In installation of tensoflow-gpu, cudatoolkit		
		and cudnn will be installed automatically.		
tqdm	4.32.1			

Table 2. Additional module requirement for advanced operation of Sapphire

3-2. Data preparation

3-2-1. Original images

Sequential raw images should be gray-scale and individual well including animals also should be under 56 x 56 pixel size.

3-2-2. 'Mask maker': making of mask file

Mask file should be made for access to individual well. Sapphire provides semiautomatic mask creation whose procedure is as follows:

- (A) Target dataset selection
- (B) Well selection
- (C) Mask parameter tuning
- (D) Conformation and save a proper parameters

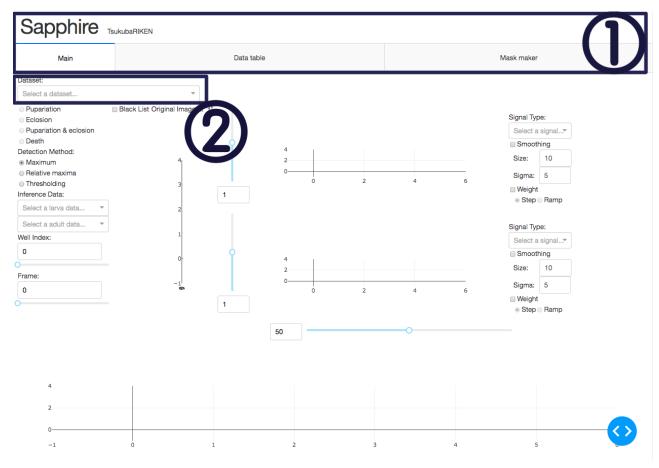


Fig 1. Layout of 'Main' tab

Sapphire TSUKUBARIKEN

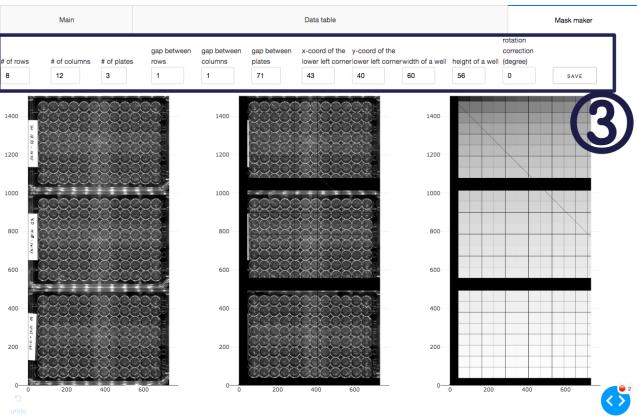


Fig 2. Layout of 'Mask maker' tab

(A) Target dataset selection

Select target datasets in 'Main' tab, and click 'Mask maker' tab after the selection of dataset.

(B) Well selection

A first image of sequential images is displayed after dataset selection. Users can choose an arbitral well within a population image and frame on a well as a reference by right click.

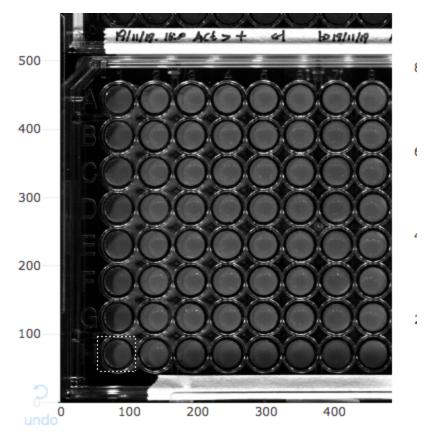


Fig 3. Reference well surrounded by white dots square

(C) Mask parameter tuning

Users can modify the mask parameters with monitoring a generated mask image (Fig. 4)



Fig 4. Parameters for mask creation

Each parameter descriptions are as follows:

No.	Parameter	Description		
1	# of rows	# of well row in a population image		
2	# of cols	# of well colimn in a population image		
3	# of plates	# of plate		
4	gap between rows	Horizontal gap between wells		
5	gap between columns	Vertical gap between wells		
6	gap between plates	Gap between plates		
7	x-coord of the lower left corner	Horizontal coordinate (pixel) of a reference well		
8	x-coord of the lower left corner	Vertical coordinate (pixel) of a reference well		
9	width of a well	Width of a well		
10	height of a well	Height of a well		
11	rotation correction (degree)	Angle from vertical line		

Table 3. Parameters for mask creation

(D) Confirmation and save a proper parameters

If mask parameters were proper and all the wells were correctly surrounded by square-shape reference box, save the parameters by press SAVE button.

3-2-3. Data directory structure

Sapphire loads data such as config files, original images, etc. from dataset directory tree. The directory tree is rigorously defined as follow. Note that the files/directories beginning with asterisk can be arbitrarily named, although the others should have the names exactly same as given below.

```
- config.json
grouping.csv
inference
   — adult
      └── *profile1
            - *cf_r0.003_signals.npy
            - probs
                - 000.npz
                - 001.npz
                - 002.npz
             probs.npz
             - *signals.npy
    - larva
      └─ *profile1
            - probs
              -- 000.npz
                — 001.npz
              └─ 002.npz
             probs.npz
            - *signals.npy
 mask.npy
 mask_params.json
 *network
    - adult
      --- *profile1
      └─ *profile2
   — larva
      -- *profile1
      └── *profile2
- original
```

- dataset1, dataset2: Dataset directory.
- blacklist.csv: written in CSV (comma separated value) format.
- config.json: Configuration file for a dataset.
- grouping.csv: Defines groups of flies (CSV format).
- inference: Directory for storing results of inference by a trained neural network.
- inference/adult or inference/larva: Stores inference results for adult/larva flies.
- inference/*/profile1: The name of training profile indicating which trained network is used for the inference. The directory name is same as network/*/profile1.
- inference/*/cf_r0.003_signals.npy: ChangeFinder signal.
- inference/*/probs: Stores inference results of each fly in Numpy archive format. The number in file names indicates fly ID.
- inference/*/probs.npz: Numpy archive including inference results of all the flies.
- inference/*/signals.npy: Label diference signal.
- mask.npy: Definition of pixels of each fly in an original image. You can create this file with Sapphire's mask maker tab.
- mask_params.json: Parameters for creating the mask. You can create this file with Sapphire's mask maker tab.
- network: Stores neural networks trained for semantic segmentation.
- network/adult or network/larva: Stores networks for adult/larva flies.
- network/*/profile1: The name of training profile.
- original: Stores original images.

3-3. Automatic detection

3-3-1. Animal body segmentation (inference by trained neural network)

To perform animal body inference by trained network, you can type as follows:

python inference.py PATH_OF_TARGET_DATASET FILE_OF_TRAINED_NET

WORK

3-3-2. Getting change point signal

You can get ChangeFinder signal by running make_CF_signals.py script. The script requires signals.py file which has signal data calculated by sequential segmentation data.

```
python make_CF_signals.py PATH/signal.py TARGET_EVENT
```

TARGET_EVENT takes an event name 'pupariation', 'eclosion', or 'death'

The script also accepts the parameter of ChangeFinder, r.

(Example) Calculation with r=0.009

```
python make_CF_signals.py PATH/signal.py TARGET_EVENT -r0.009
```

Note that r should take between zero and one.

3-4. Launching of Sapphire as a viewer

Sapphire can be launched in arbitral web browser.

The web application has three tabs.

'Main' tab

- Viewing original image data
- Viewing inference data
- Visualization of individual activity signals and summary data

'Data table' tab

 Browsing all the event timing obtained by automatic detection algorithm, and exporting the result as csv file.

'Mask maker' tab

• Making mask file

3-4-1. Usage

You can launch Sapphire by typing:

```
python sapphire.py
```

Sapphire is a web application, so you can open Sapphire in your web browser by accessing localhost:8050 (Fig 4).

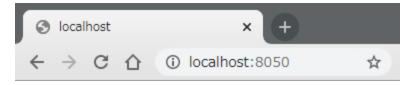


Fig 4. URL to Sapphire

3-4-2. 'Main' tab

Original images, inference results, summary could be visualized in 'Main' tab (Fig 5).

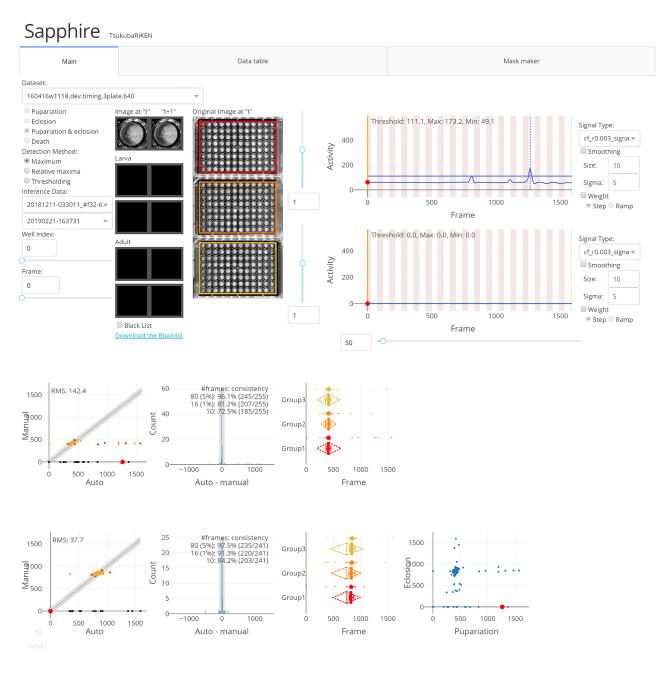


Fig 5. Screenshot of 'Main' tab

3-4-3. Dataset and target event

By the pull-down list at left top in 'Main' tab, users can choose the dataset to be visualized. A dataset directory includes a configuration file (config.json) that defines which type of event should be detected by the system (e.g. pupariation, eclosion, and death). Such target event will be automatically shown in the corresponding radio button.

The current version of Sapphire provides three types of detection methods.

Maximum, relative maxima, and simple thresholding of ChageFinder signal can be chosen at the radio button named 'Detection method'.

3-4-4. Inference data

By pull-down list in 'Inference data', users could select an inference data if they had several inference data obtained by various methods or conditions.

3-4-5. Visualization of well example

Individual well image and inference result at specific time step (t) and consecutive time step (t+1) are displayed (Fig 6). The well example can be selected by textbox named 'Well index' and sliderbar under the textbox, or direct clicking of target well on a population image.

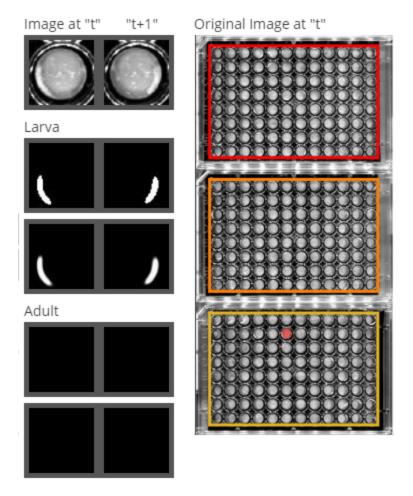


Fig 6. Well example

Time step t is also modifiable by textbox named 'Frame' and sliderbar under the textbox, or direct clicking on time series of activity signal desplayed in right top panel. The time series data will be explained in following section.

3-4-6. Visualization of time series data

Sapphire can visualize several types of time series data (Fig 7). The signal users would like to show can be selected by pull-down list named 'Signal Type'. Visualized time series data are modifiable by 'smoothing' checkbox and textbox defining the smoothing parameters. The signals can be weighted by step or ramp function. Usually, upper panel describes the signal obtained from larva data and lower panel shows the signal from adult animal.

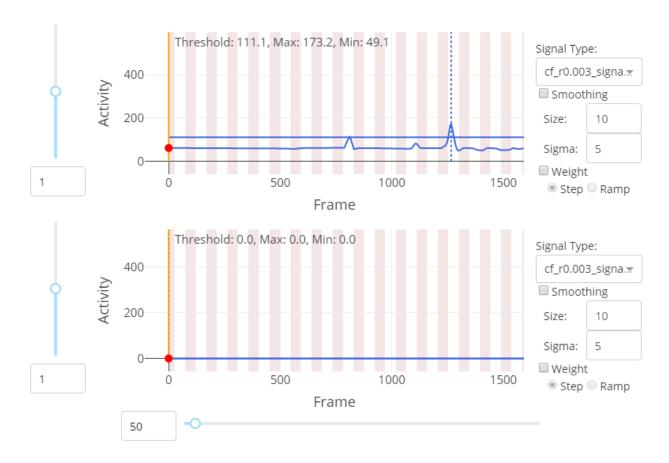


Fig 7. Time series data

A vertical orange line indicates in the graph is a timing of manual detection if it was loaded. A horizontal blue line describes a threshold value which is manually modifiable by textbox at left side and sliderbar above it. A vertical blue dot line corresponds to a timing of automatic detection by the system.

3-4-7. Visualization of summary data

Sapphire provides summary visualization by several styles (Fig 8). Scatter plot describes a comparison between automatic- and manual-detections (left panel). A colored dot corresponds to an individual well and the black line is y = x.

The gray region indicates 5% consistency between auto and manual for entire frames. The frame difference between auto and manual is displayed as a histogram (middle panel). Summary for individual group, defined by user via group.csv, is shown as box plots (right panel).

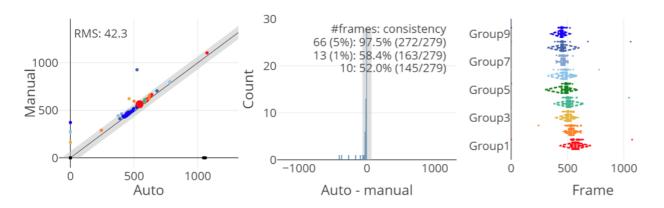


Fig 8. Comparison between automatic- and manual-detections

3-4-8. 'Data table' tab

This tab provides some table data (Fig 9). The 'Timestamp' table shows the correspondence of an image file name to the captured date and time. 'Event timings' tables exhibit animal event (pupariation, eclosion, and death) timings evaluated by human experts (manual) and the system (auto). You can get CSV files of the tables by clicking 'download' link.

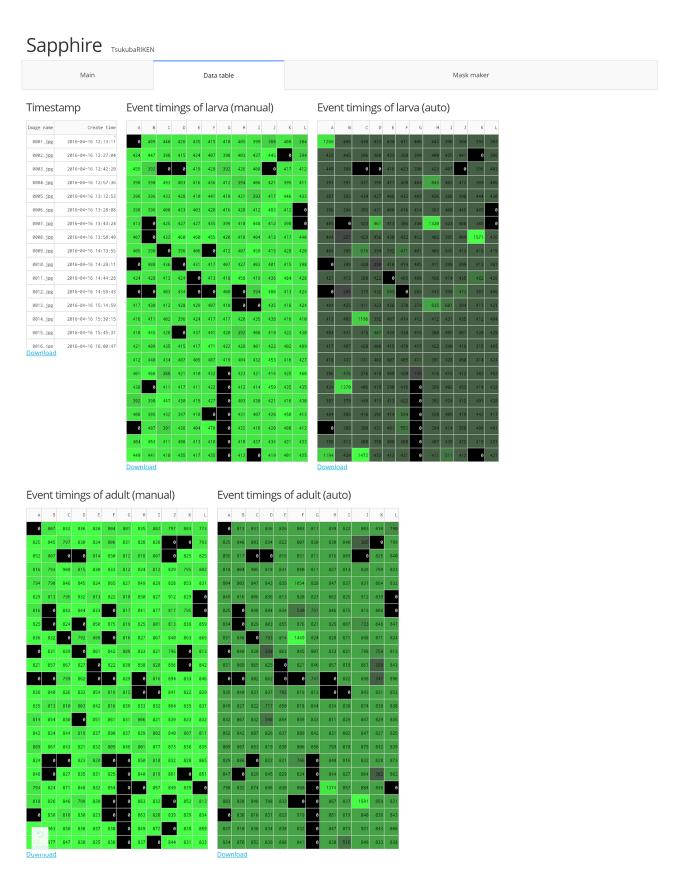


Fig 9. Summary table in 'Data table' tab