

Retinoic Acid Treatment Affects Gene Expression of Immature Neuronal Markers and Up-Regulates SNAP25 Expression in SH-SY5Y cells

Eline Poinsignon-Clavel

1006765528

HMB310 UofT

INTRODUCTION

- SH-SY5Y cells (neuroblast, immature neuron) ¹
 - Epithelial structure, grow in clumps, not fully differentiated when used for qPCR
 - Neuronal model

What is RA?

- Active form of vit A
- **Increases cell differentiation**
- Decrease cell proliferation
 - Can suppress tumors
- Act as transcription factor in nucleus – gene expression²

RA side effect:

- Oxidative stress
- Less susceptible to get Parkinson's³

- RA Tx: homogeneous differentiated SH-SY5Y neurons
- no RA Tx: high proliferation of undifferentiated SH-SY5Y cells⁴

Research Question/Aim:

How does RA treatment affect gene expression in immature neurons?

Group data

- Collected from 3 Thursday PM classes
- 12 genes -> neuronal markers
- RA-affected because increases differentiation
- binds to receptors in promoting region
- regulates gene transcription in early development⁵ -> (state of SH-SY5Y cells)
- Down-regulated by differentiation -> cell-cell interaction which stops cellular processes, hence decreasing cell proliferation
- Up-regulated by differentiation -> for cell differentiation

Hypothesis:

RA will change gene expression of different neuronal markers

SNAP25

- Collected from 3 Thursday PM classes
- Presynaptic plasma membrane protein part of SNARE complex
 - **Negatively regulates neurotransmitter release**
 - Negatively regulates intracellular Ca^{2+} from V-gated Ca^{2+} channels⁶
- Normal postsynaptic surface expression of NMDARs⁷
- RA Tx -> levels double⁸
 - Increased differentiation & decrease proliferation -> decrease cellular synaptic pathways

Hypothesis:

RA will increase SNAP25 expression (up-regulation)

METHODOLOGY

Independent variable: RA treatment

Dependent variable: Gene expression level

1. SH-SY5Y treated with 10 μ M RA (control = no RA) for 72h
 2. qPCR steps applied as in Figure 1
- Same treatment done on other sample using RNA sequencing (by technicians)

Baselines:

- qPCR normalized w GAPDH
- RNAseq normalized w TPM

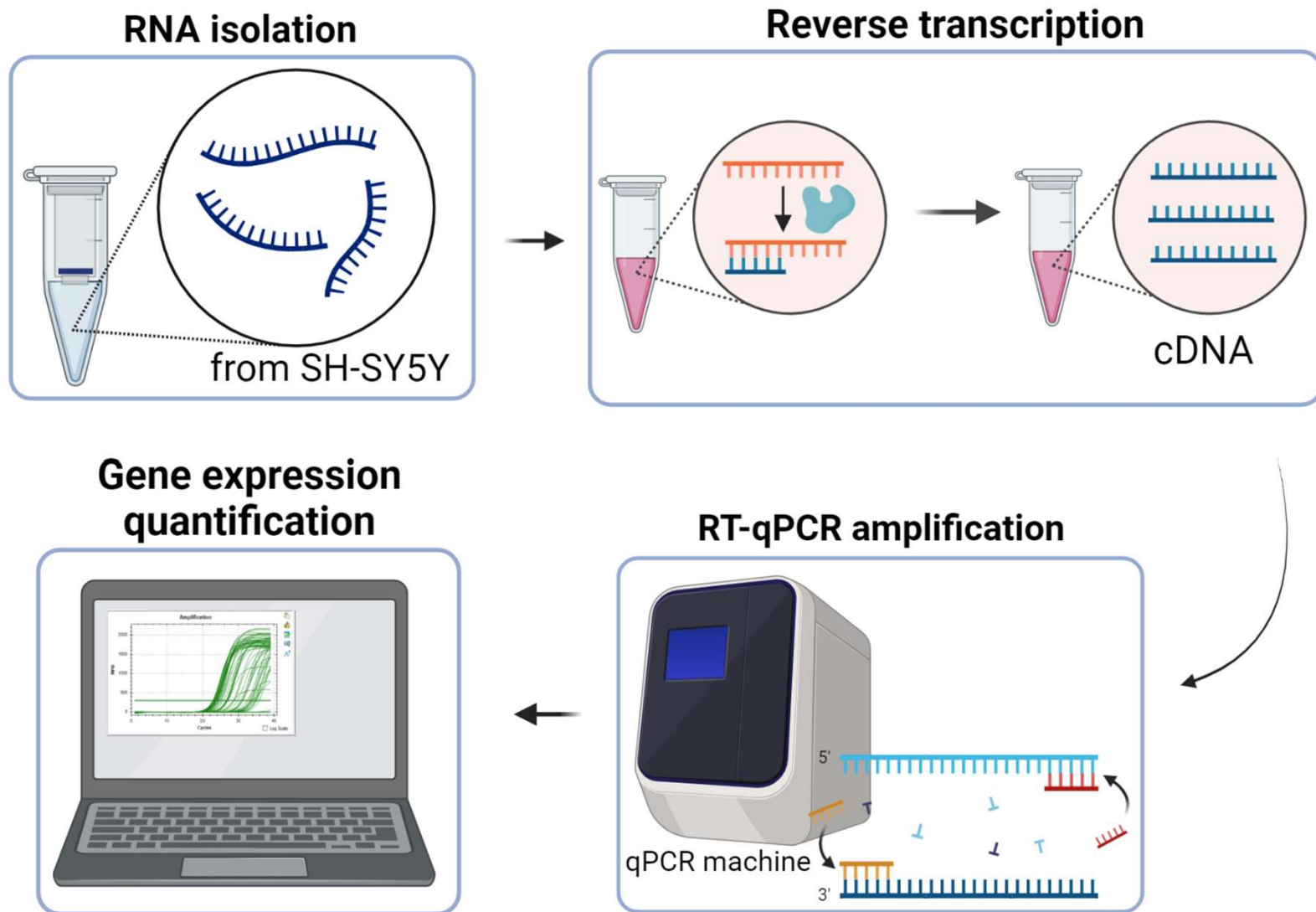
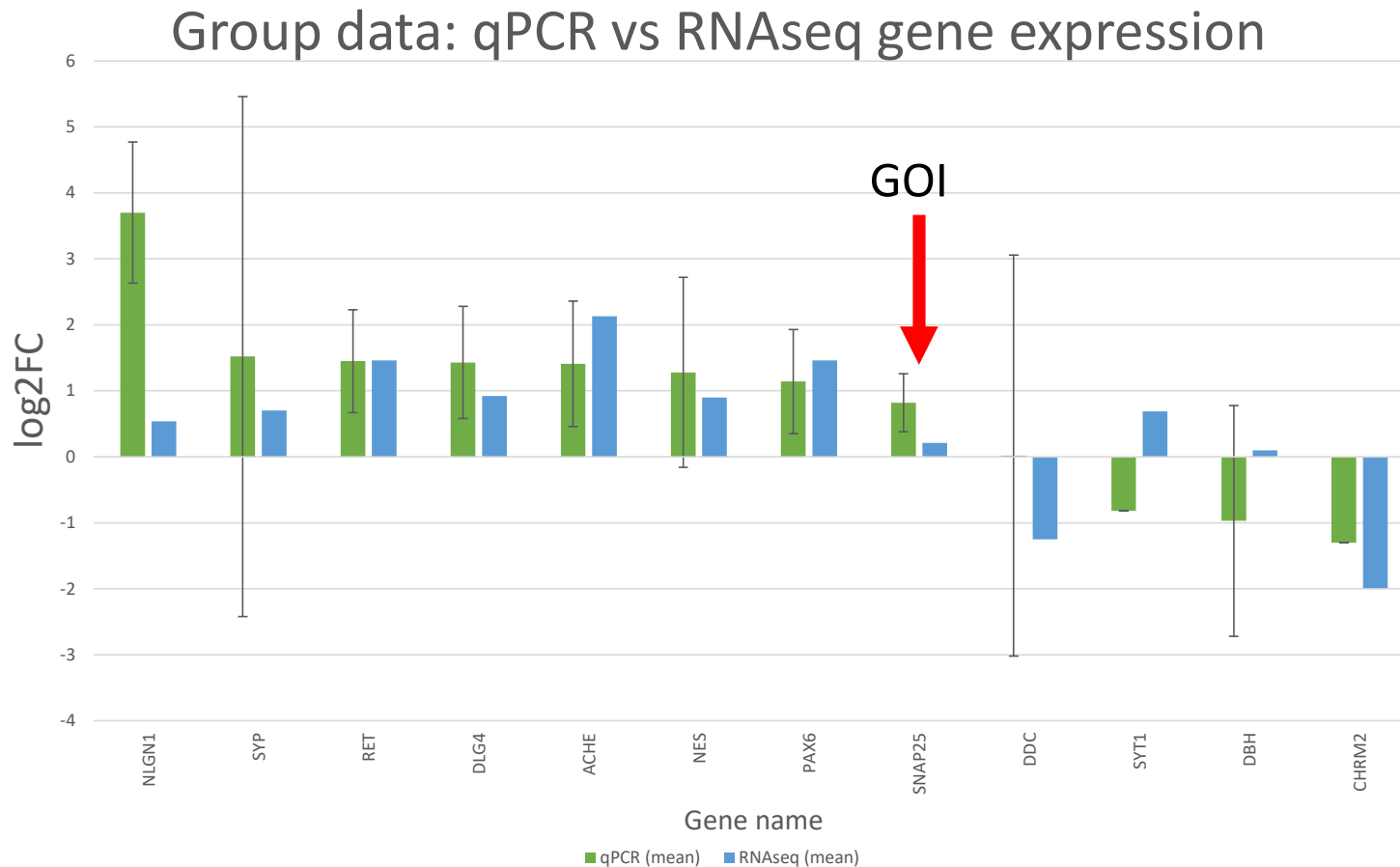


Figure 1: qPCR steps (made with BioRender⁹)

RESULTS

Group Data

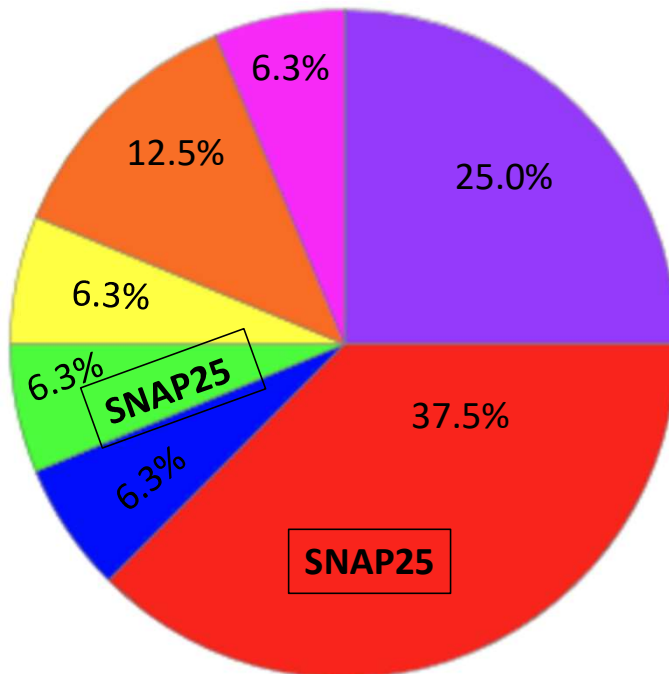


- qPCR data used SD
- DDC, SYT1, DBH had different directionalities (qPCR vs RNAseq)
 - Up-regulated vs down-regulated

Figure 2: Expression level of 12 neuronal markers after RA Tx for qPCR and RNAseq (made with Excel) – data taken from mean of 3 Thursday PM classes

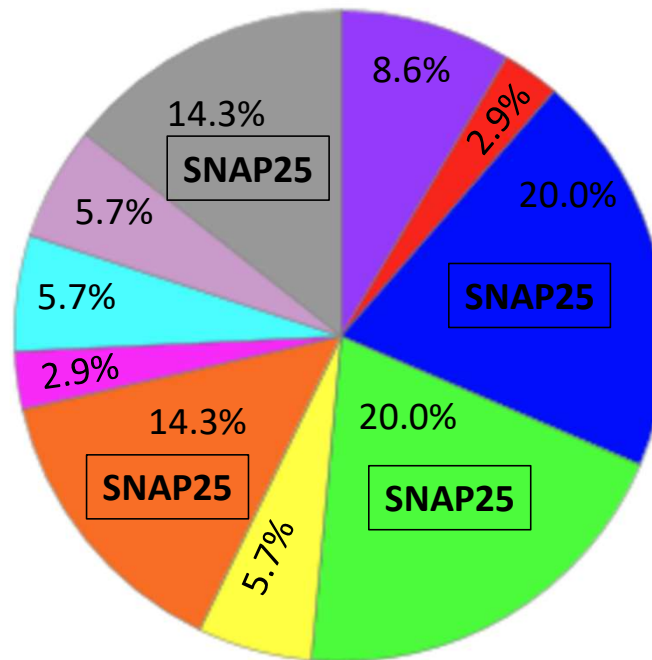
Figure 3: Up-regulated genes (based on qPCR) – Ontological domains (made by PANTHER¹⁰)

Molecular function



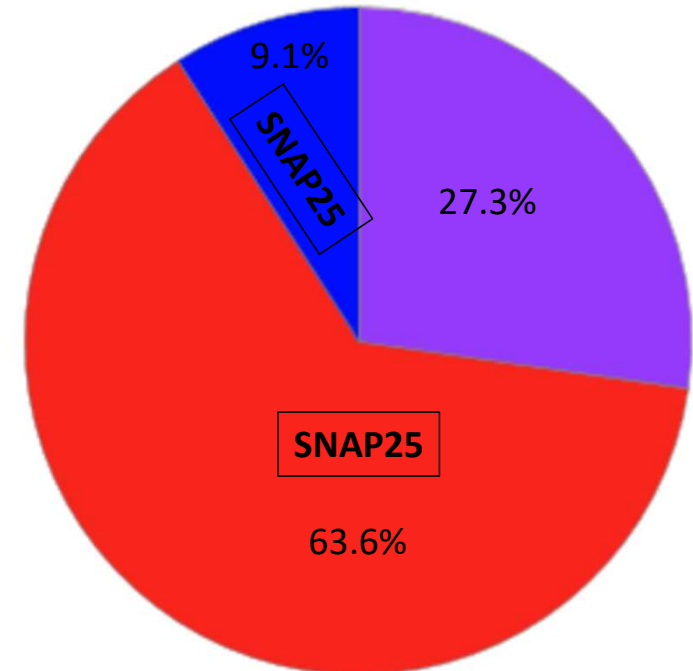
[No PANTHER category is assigned \(UNCLASSIFIED\)](#)
[binding \(GO:0005488\)](#)
[catalytic activity \(GO:0003824\)](#)
[molecular adaptor activity \(GO:0060090\)](#)
[molecular function regulator \(GO:0098772\)](#)
[molecular transducer activity \(GO:0060089\)](#)
[transcription regulator activity \(GO:0140110\)](#)

Biological process



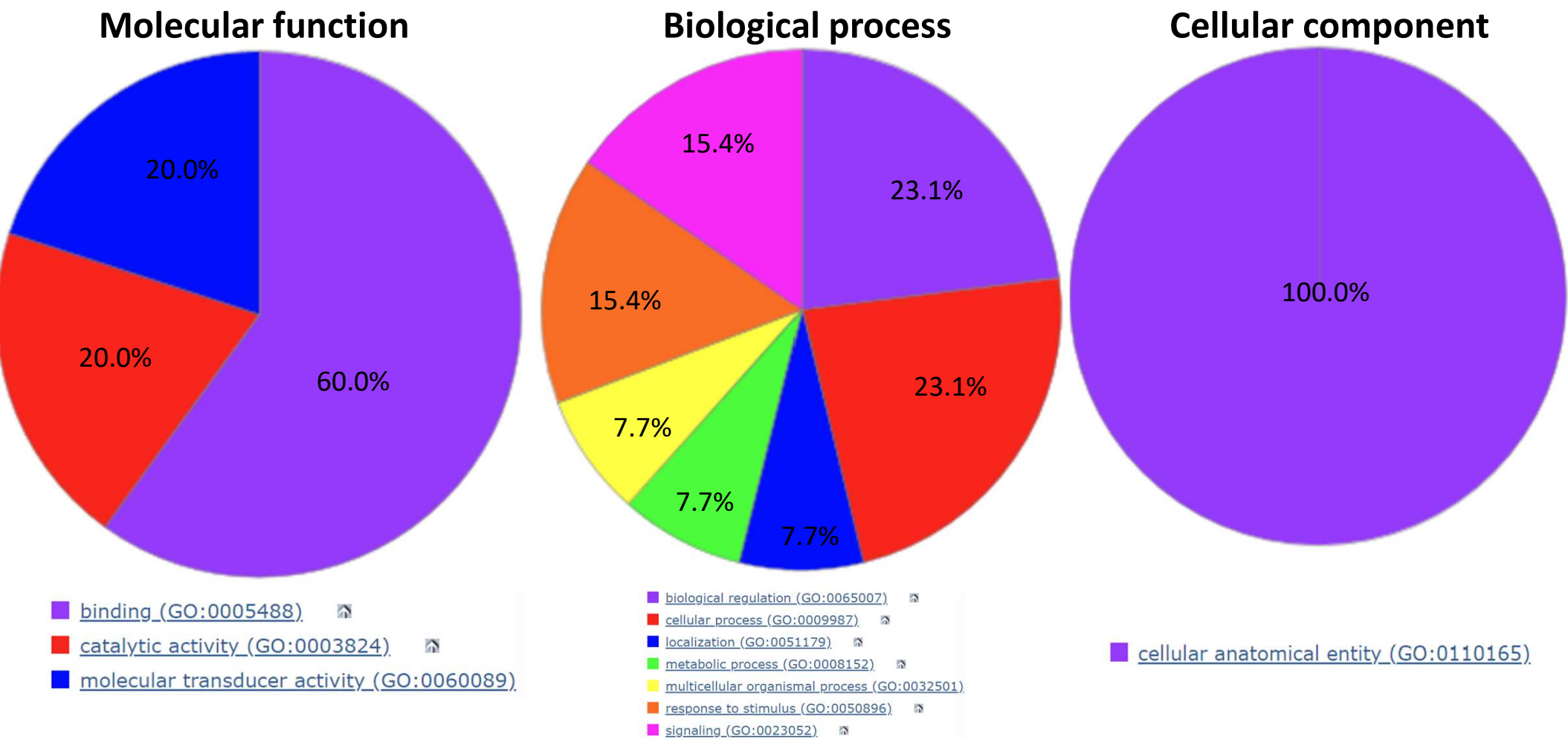
[No PANTHER category is assigned \(UNCLASSIFIED\)](#)
[biological adhesion \(GO:0022610\)](#)
[biological regulation \(GO:0065007\)](#)
[cellular process \(GO:0009987\)](#)
[developmental process \(GO:0032502\)](#)
[localization \(GO:0051179\)](#)
[metabolic process \(GO:0008152\)](#)
[multicellular organismal process \(GO:0032501\)](#)
[response to stimulus \(GO:0050896\)](#)
[signaling \(GO:0023052\)](#)

Cellular component



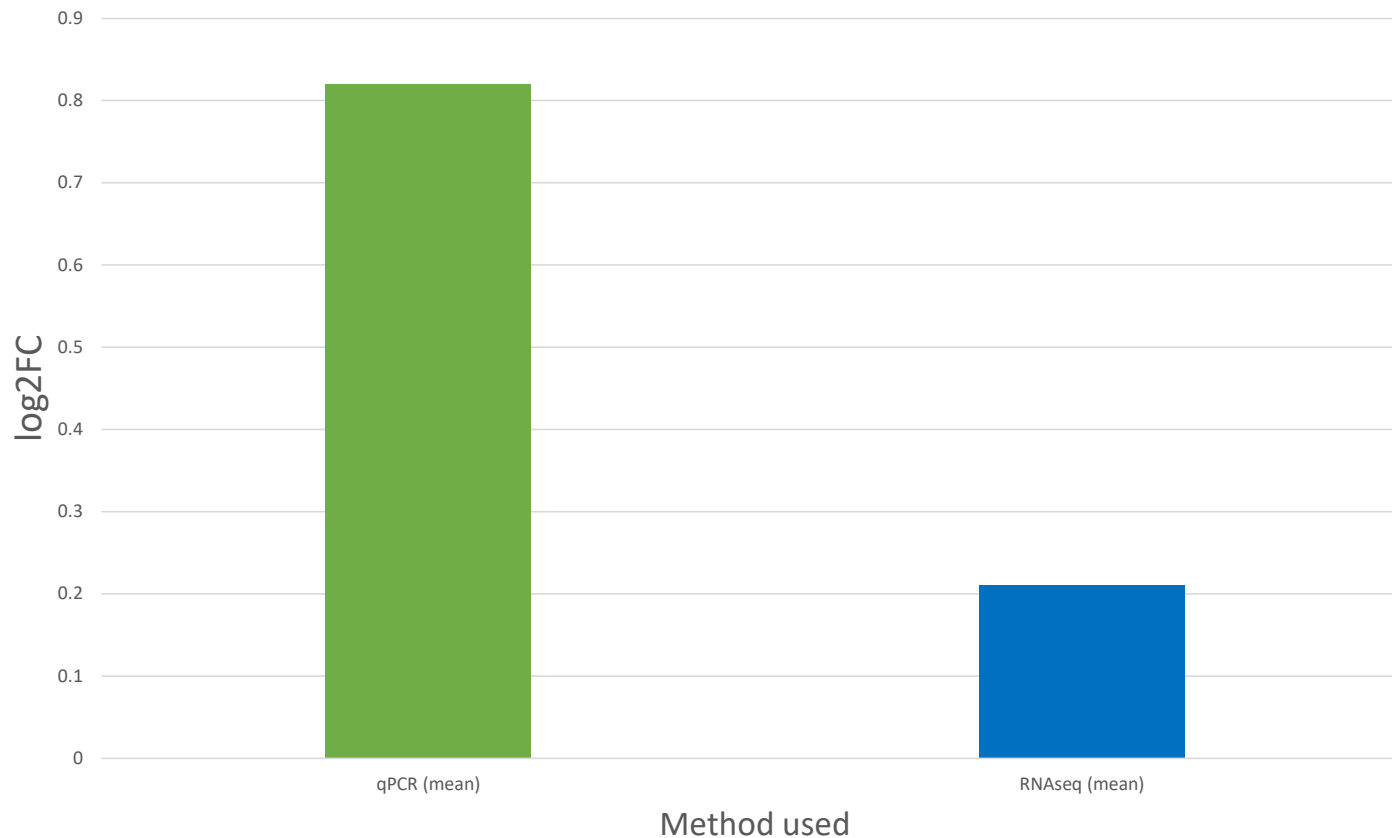
[No PANTHER category is assigned \(UNCLASSIFIED\)](#)
[cellular anatomical entity \(GO:0110165\)](#)
[protein-containing complex \(GO:0032991\)](#)

Figure 4: Down-regulated genes (based on qPCR) – Ontological domains (made by PANTHER¹⁰)



SNAP25 - GOI

SNAP25 expression in qPCR vs RNAseq

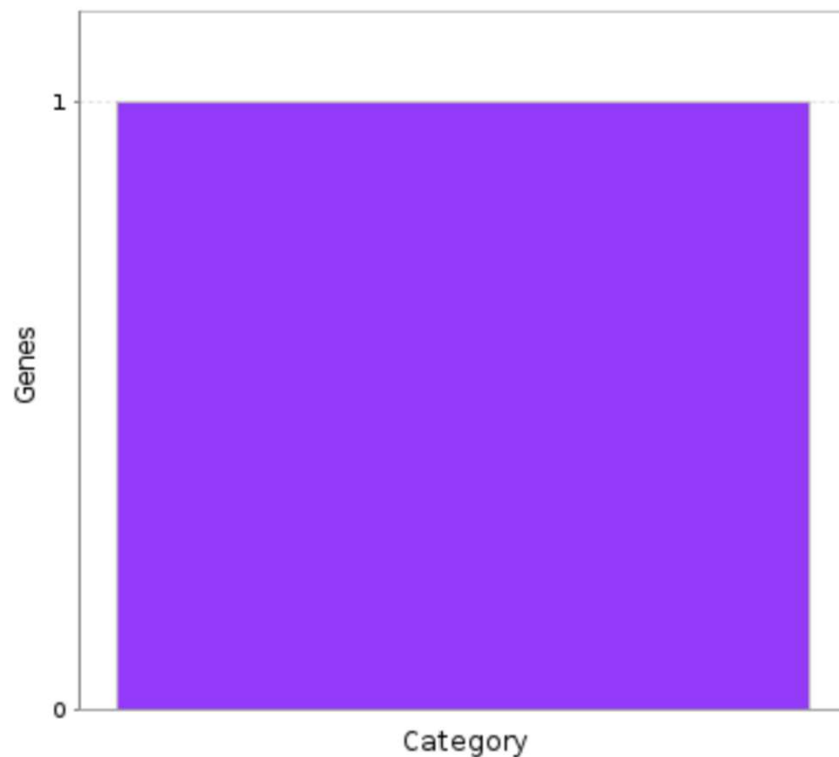


- Same directionality
-> **up-regulated**
- qPCR expression level = 0.82 log2FC
 - increase compared to control

Figure 5: SNAP25 expression level after RA Tx for qPCR and RNAseq (made with Excel) - data taken from mean of 3 Thursday PM classes

Ontological subdomain

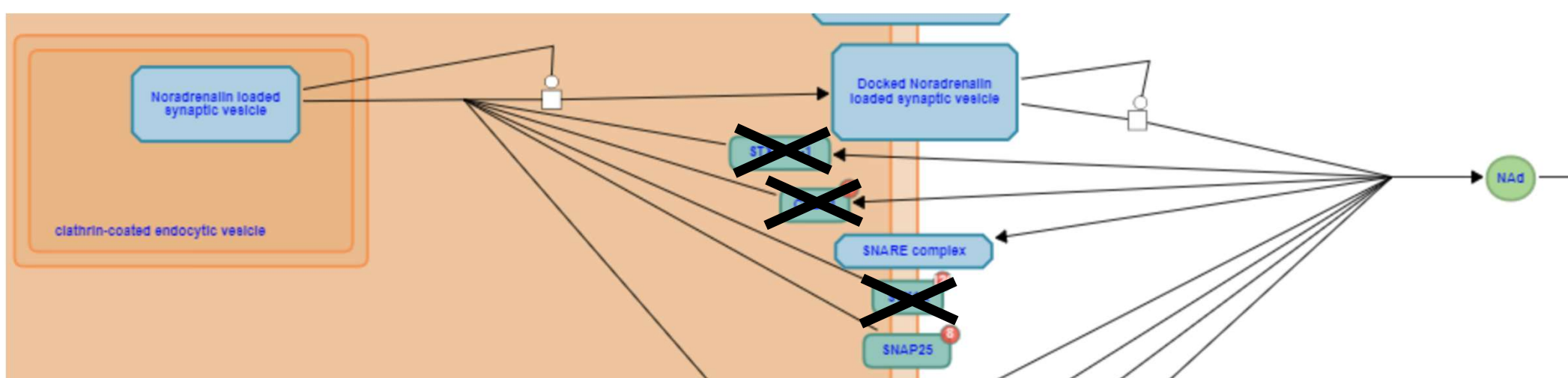
Figure 6: Up-regulated genes - **Protein-containing complex subdomain** of Cellular components (made by PANTHER¹⁰)



- Only SNAP25 (out of up-regulated genes) in protein-containing complex subdomain

■ [membrane protein complex \(GO:0098796\)](#) → SNARE complex

Figure 7: Neurotransmitter release cycle pathway involving SNAP25 (from Reactome¹¹)



- Noradrenaline (NA) vesicle input to SNAP25 (part of SNARE)
- NA gets released and transitioned to extracellular NA
 - Outputs back to SNARE

DISCUSSION - CONCLUSION

- **RA affected gene expression of 12 genes (up/down regulation)**
- **SNAP25 upregulated by RA Tx**
- > **both hypotheses were correct**
- SNAP25 had low variability (qPCR)
- SNAP25 upregulation -> more neurotransmitters released⁸
- BUT it negatively regulates intracellular Ca²⁺, so upregulation should decrease neurotransmitter release
- CHRM2, SYT1, DBH were down-regulated (qPCR) -> involved in cell signalling and synaptic activity
- Different directionality for some genes in qPCR vs RNAseq
 - > qPCR not always homogenous amplification¹²
 - > difference amplification efficiencies¹³

Limitations

- RA treatment not long enough
 - neuroblastoma cells have to be differentiated *in vitro* for at least 7 days for SH-SY5Y cells to be used for experiments¹⁴
- Gene expression change throughout cell differentiation, but was only measured at 72h

Future directions

- Let SH-SY5Y cells differentiate for 1 week
- Samples at different time-points along differentiation process
 - For accurate idea of RA Tx effect on SNAP25

References

1. Kovalevich J, Langford D. 2013. Considerations for the Use of SH-SY5Y Neuroblastoma Cells in Neurobiology. *Neuronal Cell Culture*. 1078:9–21. doi:10.1007/978-1-62703-640-5_2.
2. Grenier E, Maupas FS, Beaulieu J-F, Seidman E, Delvin E, Sane A, Tremblay E, Garofalo C, Levy E. 2007. Effect of retinoic acid on cell proliferation and differentiation as well as on lipid synthesis, lipoprotein secretion, and apolipoprotein biogenesis. *American Journal of Physiology-Gastrointestinal and Liver Physiology*. 293(6):G1178–G1189. doi:10.1152/ajpgi.00295.2007.
3. Cheung Y-T, Lau WK-W, Yu M-S, Lai CS-W, Yeung S-C, So K-F, Chang RC-C. 2009. Effects of all-trans-retinoic acid on human SH-SY5Y neuroblastoma as in vitro model in neurotoxicity research. *NeuroToxicology*. 30(1):127–135. doi:10.1016/j.neuro.2008.11.001.
4. Shipley MM, Mangold CA, Szpara ML. 2016. Differentiation of the SH-SY5Y Human Neuroblastoma Cell Line. *Journal of Visualized Experiments*.(108). doi:10.3791/53193.
5. Balmer JE, Blomhoff R. 2002. Gene expression regulation by retinoic acid. *Journal of Lipid Research*. 43(11):1773–1808. doi:10.1194/jlr.r100015-jlr200.
6. Antonucci F, Corradini I, Fossati G, Tomasoni R, Menna E, Matteoli M. 2016. SNAP-25, a Known Presynaptic Protein with Emerging Postsynaptic Functions. *Frontiers in Synaptic Neuroscience*. 8:7. doi:10.3389/fnsyn.2016.00007. <https://www.ncbi.nlm.nih.gov/pubmed/27047369>.
7. Jurado S, Goswami D, Zhang Y, Molina Alfredo J, Miñano, Südhof Thomas C, Malenka Robert C. 2013. LTP Requires a Unique Postsynaptic SNARE Fusion Machinery. *Neuron*. 77(3):542–558. doi:10.1016/j.neuron.2012.11.029.
8. Andres D, Keyser BM, Petrali J, Benton B, Hubbard KS, McNutt PM, Ray R. 2013. Morphological and functional differentiation in BE(2)-M17 human neuroblastoma cells by treatment with Trans-retinoic acid. *BMC Neuroscience*. 14(1):49. doi:10.1186/1471-2202-14-49. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3639069/>.
9. BioRender. 2022. appbiorendercom. <https://app.biorender.com/>.
10. PANTHER - Gene List Analysis. 2018. Pantherdborg. <http://www.pantherdb.org/>.
11. Reactome Pathway Database. 2019. Reactomeorg. <https://reactome.org/>.
12. Fassbinder-Orth CA. 2014. Methods for Quantifying Gene Expression in Ecoimmunology: From qPCR to RNA-Seq. *Integrative and Comparative Biology*. 54(3):396–406. doi:10.1093/icb/icu023.
13. Everaert C, Luypaert M, Maag JLV, Cheng QX, Dinger ME, Hellemans J, Mestdagh P. 2017. Benchmarking of RNA-sequencing analysis workflows using whole-transcriptome RT-qPCR expression data. *Scientific Reports*. 7(1):1559. doi:10.1038/s41598-017-01617-3. [accessed 2021 Mar 7]. <https://www.nature.com/articles/s41598-017-01617-3>.
14. Sarkanen J-R, Nykky J, Siikanen J, Selinummi J, Ylikomi T, Jalonon TO. 2007. Cholesterol supports the retinoic acid-induced synaptic vesicle formation in differentiating human SH-SY5Y neuroblastoma cells. *Journal of Neurochemistry*. 102(6):1941–1952. doi:10.1111/j.1471-4159.2007.04676.x.