Translation control tunes Drosophila oogenesis

A Thesis
Presented to
The Division of Biology
University at Albany

In Partial Fulfillment of the Requirements for the Degree Doctor of Philosophy

Elliot T. Martin

December 2021

Approved for the Division (Biology)

Prashanth Rangan

Gabriele Fuchs

Acknowledgements

'The work herein was only able to be completed thanks to the contribution of others. Foremost, my wife Allison Martin, without whom I would have given up countless times along the way to my PhD. She has been a sounding board, a life-coach, and my best friend for the years this work has taken.

Secondly, my family including my son, Levi, who from childhood supported my curiosity and enabled me to pursue my interests and passions. Knowing that I have always had them to fall back on provided a cushion that ahs helped me from struggling in undergrad to the completion of my PhD.

For direction, motivation, and guidance, I thank my mentors Dr. Prash Rangan and Dr. Gaby Fuchs. They agreed to mentor a disorganized student with less than stellar academics. Since that point they have helped me not only in developing a successful project, but also in maturing as an academic, a bench scientist, and generally, into adulthood.

A thank you to my labmates who were always there to talk me through a failed experiment or get excited about an interesting result.

To my collaborators, Elaine Nguyen, Roni Lahr, Dr. Andrea Berman, Dr. Shamsi Emtenani, and Dr. Daria Siekhaus, that contributed to this work I thank you for your expertise and beautiful results.

Finally, to my committee members, Dr. Thomas Begley, Dr. Paolo Forni, and Dr. Joesph Wade for their guidance and advice throughout my graduate studies.

```
r if(!knitr:::is_latex_output()) '## Abstract {-}''
```

Preface

This is an example of a thesis setup to use the reed thesis document class (for LaTeX) and the R bookdown package, in general.

Table of Contents

${f Introduction}$

List of Tables

List of Figures

Abstract

All dynamic biological processes require control over transcription, translation, or posttranslational products. Stem cells in particular require dynamic control of gene expression. My work has focused on characterizing this control, primarily at the translation level, to better understand how stem cell differentiation occurs. Stem cells are cells with the unique ability to develop into more specialized cell types in a process called differentiation. Some stem cell, including those focused on in my work, also have the ability to "self-renew," a process that allows one stem cell to copy itself giving rise to two stem cells. These processes must be carefully balanced as excess self-renewal will result in cells that do not give rise to differentiated cells necessary for further development or biological function. However, excess differentiation will result in the lack of an available pool of stem cells, preventing future differentiation and development. The decision of a stem cell to either self renew or differentiate is controlled by specific cellular pathways that can act at the level of transcription, translation, or post-translation. To study the regulation of these pathways in-vivo I have used the female *Drosophila* germline as a model system. The female *Drosophila* germline is contained within two pairs of ovaries. Ovaries consist of two main types of tissue, soma and germline. Each ovary is made up of strands called ovarioles. Ovarioles represent an assembly line of successive development. At the anterior tip of each ovariole a structure called a germarium is present. At the anterior of the germarium two to three stem cells are housed in a somatic niche. These germline stem cells (GSCs) can self-renew, or differentiate giving rise to a daughter cell called a cystoblast (CB). The CB turns on a differentiation factor called bag of marbles (bam). This CB then undergoes four incomplete cellular divisions, resulting in interconnected cysts consisting of two, four, eight, and finally sixteen cells. One of these cells is designated as the oocyte while the rest of the cells will become nurse cells. The sixteen cell cyst is then encapsulated by somatic cells, forming egg chambers. Egg chambers successively grow in size in fourteen stages. During this time the nurse cells produce mRNAs and proteins that are transported to the oocyte. The oocyte continues to grow, while the nurse cells eventually die, dumping their contents into the oocyte. Once the oocyte reaches the final, 14th stage it is known as an egg. Each of the steps from GSC to egg require changes in cellular pathways. These changes can occur at the level of transcription, translation, or post-translation. Decades of research has elucidated many of the changes that occur during oogenesis, however, many players in this process still remain mysterious. My work has helped to identify and characterize novel developmental mechanisms that are required for the successive developmental transitions that take place during orgenesis. I have leveraged RNAseq and polysome-seq to probe the global transcriptional and translational landscape over development. I have also used the power of *Drosophila* genetics in concert with these sequencing techniques to identify and characterize misregulated pathways.

Dedication

Dedicated to my wife and best friend Alli.

Introduction