# class18

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```
#BiocManager::install("tximport")
  #BiocManager::install("rhdf5")
  library(tximport)
  # setup the folder and filenames to read
  folders <- dir(pattern="SRR21568*")</pre>
  samples <- sub("_quant", "", folders)</pre>
  files <- file.path( folders, "abundance.h5" )</pre>
  names(files) <- samples</pre>
  txi.kallisto <- tximport(files, type = "kallisto", txOut = TRUE)</pre>
1 2 3 4
  head(txi.kallisto$counts)
                SRR2156848 SRR2156849 SRR2156850 SRR2156851
ENST00000539570
                                    0
                                         0.00000
ENST00000576455
                                    0 2.62037
ENST00000510508
                                    0.00000
ENST00000474471
                       0
                                   1 1.00000
ENST00000381700
                         0
                                   0.00000
                                                          0
ENST00000445946
                                         0.00000
  colSums(txi.kallisto$counts)
SRR2156848 SRR2156849 SRR2156850 SRR2156851
   2563611
              2600800
                         2372309
                                    2111474
```

```
sum(rowSums(txi.kallisto$counts)>0)
```

#### [1] 94561

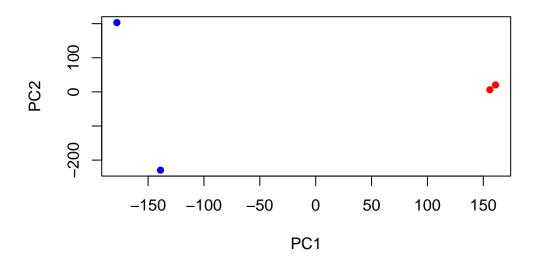
```
to.keep <- rowSums(txi.kallisto$counts) > 0
kset.nonzero <- txi.kallisto$counts[to.keep,]

keep2 <- apply(kset.nonzero,1,sd)>0
x <- kset.nonzero[keep2,]

pca <- prcomp(t(x), scale=TRUE)
summary(pca)</pre>
```

#### Importance of components:

```
PC1 PC2 PC3 PC4
Standard deviation 183.6379 177.3605 171.3020 1e+00
Proportion of Variance 0.3568 0.3328 0.3104 1e-05
Cumulative Proportion 0.3568 0.6895 1.0000 1e+00
```

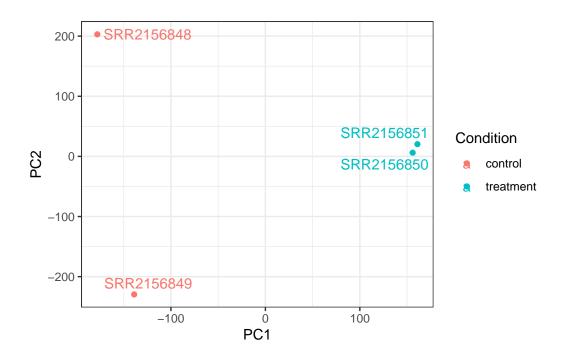


```
library(ggrepel)

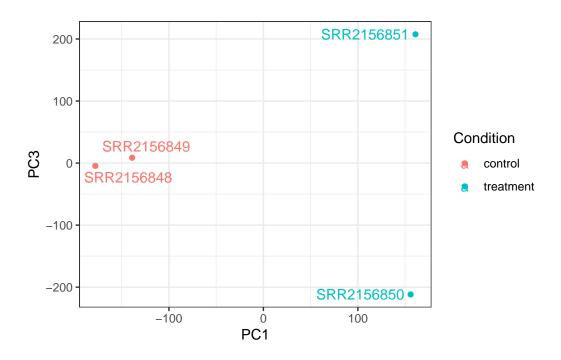
# Make metadata object for the samples
colData <- data.frame(condition = factor(rep(c("control", "treatment"), each = 2)))
rownames(colData) <- colnames(txi.kallisto$counts)

# Make the data.frame for ggplot
y <- as.data.frame(pca$x)
y$Condition <- as.factor(colData$condition)

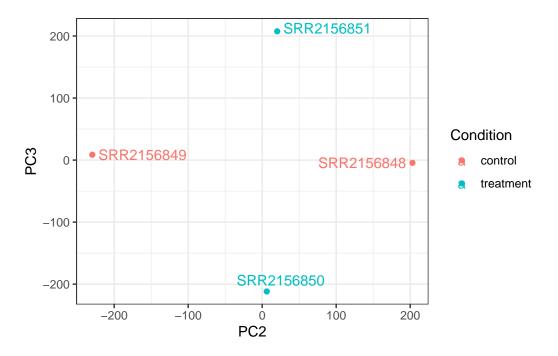
ggplot(y) +
   aes(PC1, PC2, col=Condition) +
   geom_point() +
   geom_text_repel(label=rownames(y)) +
   theme_bw()</pre>
```



```
ggplot(y) +
  aes(PC1, PC3, col=Condition) +
  geom_point() +
  geom_text_repel(label=rownames(y)) +
  theme_bw()
```



```
ggplot(y) +
  aes(PC2, PC3, col=Condition) +
  geom_point() +
  geom_text_repel(label=rownames(y)) +
  theme_bw()
```



### **DEA**

### library(DESeq2)

Loading required package: S4Vectors

Loading required package: stats4

Loading required package: BiocGenerics

Attaching package: 'BiocGenerics'

The following objects are masked from 'package:stats':

IQR, mad, sd, var, xtabs

The following objects are masked from 'package:base':

anyDuplicated, aperm, append, as.data.frame, basename, cbind, colnames, dirname, do.call, duplicated, eval, evalq, Filter, Find, get, grep, grepl, intersect, is.unsorted, lapply, Map, mapply, match, mget, order, paste, pmax, pmax.int, pmin, pmin.int, Position, rank, rbind, Reduce, rownames, sapply, setdiff, sort, table, tapply, union, unique, unsplit, which.max, which.min

Attaching package: 'S4Vectors'

The following object is masked from 'package:utils':

findMatches

The following objects are masked from 'package:base':

expand.grid, I, unname

Loading required package: IRanges

Loading required package: GenomicRanges

Loading required package: GenomeInfoDb

 ${\tt Loading\ required\ package:\ SummarizedExperiment}$ 

Warning: package 'SummarizedExperiment' was built under R version 4.3.2

Loading required package: MatrixGenerics

Loading required package: matrixStats

Attaching package: 'MatrixGenerics'

```
The following objects are masked from 'package:matrixStats':
```

colAlls, colAnyNAs, colAnys, colAvgsPerRowSet, colCollapse, colCounts, colCummaxs, colCummins, colCumprods, colCumsums, colDiffs, colIQRDiffs, colIQRs, colLogSumExps, colMadDiffs, colMads, colMaxs, colMeans2, colMedians, colMins, colOrderStats, colProds, colQuantiles, colRanges, colRanks, colSdDiffs, colSds, colSums2, colTabulates, colVarDiffs, colVars, colWeightedMads, colWeightedMeans, colWeightedMedians, colWeightedSds, colWeightedVars, rowAlls, rowAnyNAs, rowAnys, rowAvgsPerColSet, rowCollapse, rowCounts, rowCummaxs, rowCummins, rowCumprods, rowCumsums, rowDiffs, rowIQRDiffs, rowIQRs, rowLogSumExps, rowMadDiffs, rowMads, rowMaxs, rowMeans2, rowMedians, rowMins, rowOrderStats, rowProds, rowQuantiles, rowRanges, rowRanks, rowSdDiffs, rowSds, rowSums2, rowTabulates, rowVarDiffs, rowVars, rowWeightedMads, rowWeightedMeans, rowWeightedMedians, rowWeightedMedians, rowWeightedVars

Loading required package: Biobase

Welcome to Bioconductor

```
Vignettes contain introductory material; view with 'browseVignettes()'. To cite Bioconductor, see 'citation("Biobase")', and for packages 'citation("pkgname")'.
```

Attaching package: 'Biobase'

The following object is masked from 'package:MatrixGenerics':

rowMedians

The following objects are masked from 'package:matrixStats':

anyMissing, rowMedians

```
sampleTable <- data.frame(condition = factor(rep(c("control", "treatment"), each = 2)))
rownames(sampleTable) <- colnames(txi.kallisto$counts)</pre>
```

```
dds <- DESeqDataSetFromTximport(txi.kallisto,</pre>
                                   sampleTable,
                                   ~condition)
using counts and average transcript lengths from tximport
  dds <- DESeq(dds)
estimating size factors
using 'avgTxLength' from assays(dds), correcting for library size
estimating dispersions
gene-wise dispersion estimates
mean-dispersion relationship
-- note: fitType='parametric', but the dispersion trend was not well captured by the
   function: y = a/x + b, and a local regression fit was automatically substituted.
   specify fitType='local' or 'mean' to avoid this message next time.
final dispersion estimates
fitting model and testing
  res <- results(dds)</pre>
  head(res)
log2 fold change (MLE): condition treatment vs control
Wald test p-value: condition treatment vs control
DataFrame with 6 rows and 6 columns
                 baseMean log2FoldChange
                                              lfcSE
                                                         stat
                                                                 pvalue
```

NA

<numeric> <numeric> <numeric> <numeric>

NA

NA

NA

3.155061 4.86052 0.6491203 0.516261

<numeric>

ENST00000539570 0.000000

ENST00000576455 0.761453

ENST00000510508	0.000000	NA	NA	NA	NA
ENST00000474471	0.484938	0.181923	4.24871	0.0428185	0.965846
ENST00000381700	0.000000	NA	NA	NA	NA
ENST00000445946	0.000000	NA	NA	NA	NA
	padj				
	<numeric></numeric>				
ENST00000539570	NA				
ENST00000576455	NA				
ENST00000510508	NA				
ENST00000474471	NA				
ENST00000381700	NA				
ENST00000445946	NA				