## Supplementary Material:

# Model-Driven Spatial Evaluation of Nutrient Recovery from Livestock Leachate for Struvite Production

Edgar Martín-Hernández<sup>a,b</sup>, Gerardo J. Ruiz-Mercado<sup>c</sup>, and Mariano Martín<sup>a,\*</sup>

<sup>a</sup>Department of Chemical Engineering, University of Salamanca, Plza. Caídos 1-5, 37008 Salamanca, Spain

<sup>b</sup>Oak Ridge Institute for Science and Education, hosted by Office of Research & Development, U.S.

Environmental Protection Agency, Cincinnati, Ohio 45268, United States

<sup>c</sup>Center for Environmental Solutions and Emergency Response (CESER), U.S. Environmental

Protection Agency, 26 West Martin Luther King Drive, Cincinnati, Ohio 45268, United States

\*mariano.m3@usal.es

### Contents

1	Cat	ttle organic waste composition	2			
2	Dis	tribution functions of elements from cattle waste	5			
3	The	ermodynamic modeling of the carbonates system	7			
4	4 Surrogate models to estimate the formation of precipitates from livestock					
	orga	anic waste	11			
	4.1	Influence of magnesium	11			
		4.1.1 Influence of calcium	12			

4.1.2	Influence of alkalinity	 	 	 13
References				14

### 1 Cattle organic waste composition

To calculate the distribution functions of the different compunds of the cattle organic waste, 37 data sets for cattle organic waste compositions from 20 bibliographic references were evaluated. Table 1 collects the main statistical parameters of the data, and Table 2 shown the data obtained from literature.

Table 1: Statistical summary of cattle organic waste composition. Data from (Alburquerque et al., 2012; Bolzonella et al., 2018; Seppälä et al., 2013; Gell et al., 2011; Normak et al., 2012; Sørensen et al., 2011; Tampere & Viiralt, 2014; Moset et al., 2017; Zheng et al., 2016; Xia et al., 2012; Möller & Müller, 2012; Rigby & Smith, 2011; Smith et al., 2007; Risgberg et al., 2017; Ledda et al., 2016; Kirchmann & Witter, 1992; Möller et al., 2008; Walsh et al., 2012; Løes et al., 2017; Martin, 2003).

	DM	C	Ca	K	Z	Ь	$Ca^{2+}$	$\mathrm{K}^{+}$	N-NH <sub>4</sub>	$P-PO_4^{3-}$
	(%  mass)	Total Ca	Total K	Total N	Total P					
count	36	9	6	12	35	24	1	1	31	13
mean		2.478	0.117	0.253	0.384	0.059	0.154	1.000	0.590	0.541
$\operatorname{std}$		0.946	0.027	0.158	0.133	0.038	NaN	NaN	0.117	0.159
mim		1.200	0.080	0.084	0.114	0.005	0.154	1.000	0.348	0.216
25.00%		1.719	0.100	0.148	0.299	0.032	0.154	1.000	0.516	0.421
50.00%		2.788	0.110	0.223	0.399	0.048	0.154	1.000	0.616	0.597
75.00%	7.362	3.194	0.129	0.276	0.449	0.080	0.154	1.000	0.666	0.671
max		3.400	0.165	0.635	0.789	0.164	0.154	1.000	0.820	0.700

Table 2: Data obtained from literature.

7.35 4.90 7.40	(	(% wt)	(% wt)	(% wt)	(% wt)	ro4	nn4/n ratio	Ca2+/Ca ratio	K+/K ratio	Reference
4.90 7.40	0.52	0.15	0.64	2.98	0.17	0.35	0.63			(Möller & Müller, 2012)
7.40	0.79	0.04	0.16		0.13	0.33	0.68	0.15	П	(Rigby & Smith, 2011)
0	0.42	0.03	0.23				0.38			(Smith et al., 2007)
09.9	0.45						09.0			(Smith et al., 2007)
5.58	0.46						0.64			(Smith et al., 2007)
4.49	0.51	0.05	0.12				0.82			(Smith et al., 2007)
8.47	0.55	0.05				0.67	0.56			(Smith et al., $2007$ )
5.69	0.35	0.04				0.70	0.65			(Smith et al., 2007)
8.30	0.57	0.05				0.64	0.50			(Smith et al., $2007$ )
6.75	0.37	0.03				0.57	0.64			(Smith et al., 2007)
3.80	0.40	0.03				0.70	0.07			(Smith et al., $2007$ )
5.50	0.35	0.03				0.60	0.45			(Smith et al., 2007)
7.99	0.45	0.03				0.62	0.55			(Smith et al., $2007$ )
7.52	0.41	0.03				0.54	0.58			(Smith et al., $2007$ )
7.40	0.34			3.40			0.65			(Risgberg et al., 2017)
6.10	0.41			2.60			0.68			(Risgberg et al., 2017)
4.80	0.27	0.05	0.36			0.22	0.35			(Seppälä et al., 2013)
4.12	0.20						0.52			(Alburquerque et al., 2012)
7.84	0.25						0.38			(Alburquerque et al., 2012)
2.62	0.15						0.53			(Alburquerque et al., 2012)
7.00	0.34	0.16				0.42	0.52			(Bolzonella et al., 2018)
6.10	0.20									(Gell et al., 2011)
6.24	0.47						0.42			(Moset et al., $2017$ )
7.20	0.36	80.0					0.51			(Ledda et al., 2016)
	0.27	0.06	0.08		0.13					(Kirchmann & Witter, 1992)
9.20	0.40	0.06	0.43	3.27						(Möller et al., 2008)
5.20	0.11	0.01	0.09	1.42	0.10					(Walsh et al., 2012)
4.40										(Zheng et al., 2016)
4.65	0.29						0.62			(Sørensen et al., 2011)
4.82	0.31						0.07			(Sørensen et al., 2011)
3.50	0.43	80.0	0.22		0.11		0.72			(Tampere & Viiralt, 2014)
3.80	0.44	80.0	0.22		0.08		0.73			(Tampere & Viiralt, 2014)
3.20	0.43	80.0	0.24		0.10		0.77			(Tampere & Viiralt, 2014)
3.00	0.24	0.03		1.20	0.09		0.63			$(L\phi es et al., 2017)$
9.11	0.55	0.09				0.67	0.56			(Martin, 2003)
4.70	0.40	0.08	0.25		0.15		0.72			(Normak et al., $2012$ )
5.65										(Xia et al., 2012)

### 2 Distribution functions of elements from cattle waste

The kernel density estimation (KDE) method is used to fit the probability function to data distribution and to determine the probability density distributions of calcium, nitrogen, phosphorus, potassium, ammonia/total nitrogen, and phosphate/total phosphorus. As histograms, KDE is a non-parametric way to estimate the probability density function of a random variable. However, in the KDE method, kernels are functions associated with each data set. Thus, the unknown density function can be computed as the weighted sum of these functions (Duong, 2001).

The calculation procedure is divided into three steps. First, cattle organic waste composition data are collected from the literature. Second, the KDEs of all compounds are estimated, and finally, distribution functions are fitted, using the kernel density estimations to validate the chosen distribution. The probability density distributions which best fit the data distribution are normal distribution functions for the distribution of nitrogen as in Figure 1a, ammonia/total nitrogen molar ratio as observed in Figure 1b, and phosphorus as shown in Figure 1c. Lognormal distribution functions are the best fit for phosphate/total phosphorus molar ratio as in Figure 1d, calcium, as shown in Figure 1e, and potassium as observed in Figure 1f.

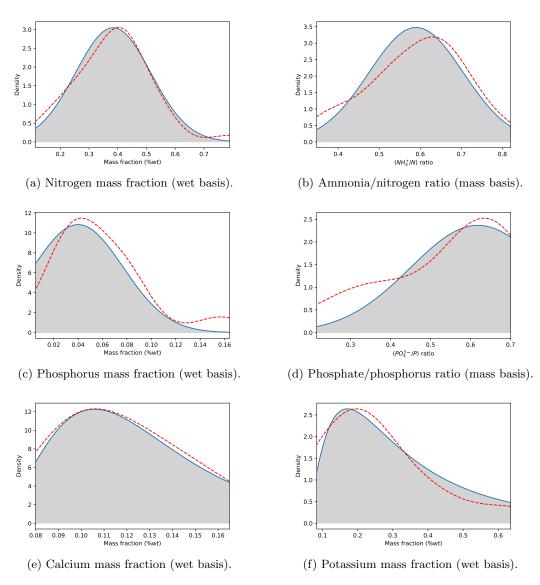


Figure 1: Kernel density estimation (red dashed line) and probability density distribution (blue solid line) for the evaluated components in cattle organic waste.

### 3 Thermodynamic modeling of the carbonates system

To calculate the total carbonate content, the concept of defining alkalinity will be used. The formal definition of total alkalinity is the capacity of a solution to buffer changes in pH that would make the solution more acidic. From the perspective of charge balance, total alkalinity can be considered as a measure of the number of protons that can be accepted by the proton acceptors present in the dissolution. Therefore, total alkalinity, TA, is calculated as the excess of proton acceptors over donors with respect to a chosen zero level of protons, as in Eq. 1:

$$TA = \text{proton acceptors} - \text{proton donors}$$
 (1)

Note that the distribution of different species for various systems, such as carbonic acid and phosphoric acid, depend on pH, and the contribution of each species to alkalinity, is a function of its electrical charge. For our case, the chemical systems considered are water, carbonic acid, phosphoric acid, and ammonia systems

A zero level of protons is a reference system defined by choosing a certain chemical compound for each system at a certain pH selected arbitrarily. This reference chemical is the dominant species at the selected pH. The chemical species above this zero level of protons will be proton acceptors while species below this level will be proton donors in a determined chemical system. There is a standard defined by Dickson (1981), where the reference pH value is set to 4.5 and chosen as the zero level of protons. For a more detailed discussion about alkalinity, we refer to Wolf-Gladrow et al. (2007). The dominant chemical substances for each system at the reference pH of 4.5 are  $H_2O$ ,  $H_2CO_3$ ,  $H_2PO_4^-$  and  $NH_4^+$ . With these considerations, the proton donors and acceptors for each system are collected in Table 3:

Table 3: Proton donors and acceptors for each chemical system.

Chemical system	Proton donors	Proton acceptors
Water	$\{H^+\}$	$\{OH^-\}$
Carbonic acid	$\{H^+\}$	$\{HCO_3^-\}, 2 \cdot \{CO_3^{2-}\} $ $\{HPO_4^{2-}\}, 2 \cdot \{PO_4^{3-}\}$
Phosphoric acid	$\{H_3PO_4\}, \{H^+\}$	$\{HPO_4^{2-}\}, 2 \cdot \{PO_4^{3-}\}$
Ammonia system	$\{H^+\}$	$\{NH_3\}$

Therefore, the alkalinity due to carbonates, called carbonate alkalinity  $(Alk_{carb})$ , as described in Eq. 2, can be calculated from total alkalinity value minus the alkalinity contribution of the other compounds, Eq. 4. The activities of the chemical species from water, phosphoric acid, and ammonia systems are obtained by solving the equilibrium and mass balance equations described previously.

The formulation of the alkalinity problem is described next. First, the alkalinity given in milligrams of CaCO<sub>3</sub> per liter is transformed into equivalents per liter by Eq. 3 and the carbonate alkalinity is calculated through Eq. 4.

$$Alk_{carb} = \{HCO_3^-\} + 2\{CO_3^{2-}\}$$
 (2)

$$Alk\left(\frac{Eq}{L}\right) = \frac{mg_{CacO_3}}{L} \cdot \frac{1}{1000} \frac{g}{mg} \cdot \frac{1Eq}{50}$$
(3)

$$Alk_{carb} = Alk - \{OH^{-}\} - \{HPO_{4}^{2-}\} - 2\{PO_{4}^{3-}\}$$

$$-\{NH_{3}\} + \{H_{3}PO_{4}\} + \{H^{+}\}$$

$$(4)$$

To compute the distribution of carbonate species, the fractions of each compound ( $\alpha_{CO_2}$ ,  $\alpha_{HCO_3^-}$  and  $\alpha_{CO_3^{2-}}$ ), which only depend on the pH, are calculated by employing Eqs. 5 - 7 (Wolf-Gladrow et al., 2007). As only  $HCO_3^-$ , and  $CO_3^{2-}$  contribute to alkalinity, the first one with one equivalent and the second one with two equivalents, as shown in Eq. 2, it is necessary to add Eq. 8. Because cattle waste has basic pH and the major species are  $HCO_3^-$  and  $CO_3^{2-}$ , it has been considered that total carbonates calculated in Eq. 8, are equal to the sum of all

carbonate species in the organic waste.

$$\alpha_{CO_2} = \frac{\left\{H^+\right\}^2}{\left\{H^+\right\}^2 + \left\{H^+\right\} K_{sp_{H_2CO_3}} + K_{sp_{H_2CO_3}} K_{sp_{HCO_2^-}}}$$
 (5)

$$\alpha_{HCO_{3}^{-}} = \frac{\{H^{+}\} K_{sp_{H_{2}CO_{3}}}}{\{H^{+}\}^{2} + \{H^{+}\} K_{sp_{H_{2}CO_{3}}} + K_{sp_{H_{2}CO_{3}}} K_{sp_{HCO_{2}^{-}}}}$$
(6)

$$\alpha_{CO_3^{2-}} = \frac{K_{sp_{H_2CO_3}} K_{sp_{HCO_3^-}}}{\{H^+\}^2 + \{H^+\} K_{sp_{H_2CO_3}} + K_{sp_{H_2CO_3}} K_{sp_{HCO_3^-}}}$$
(7)

$$Total carbonates = \frac{Alk_{carb}}{\alpha_{HCO_3^-} + 2\alpha_{CO_3^{2-}}}$$
(8)

Once the total carbonates concentration is known, the thermodynamic models for carbonate species existing in aqueous phase can be defined for the following species

$$H_2CO_3 \leftrightarrow HCO_3^- + H^+$$
  
 $HCO_3^- \leftrightarrow CO_3^{2-} + H^+$ 

Two elements are necessary to define the thermodynamic models for carbante species, the therodynamic equilibria defined by the Eq. 9, computed using activity coefficients, and mass balance defined by Eq. 11. Finally, solving the equations 2 to 11 the concentrations of the different carbonate species are determined.  $pK_{sp}$  values for all systems evaluated in this work are collected in Table 4.

$$K_{sp} = \frac{\prod \{Products\}}{\prod \{Reactants\}}$$

$$\tag{9}$$

$$[i]_{initial} = \sum_{products} [i]_{products} \tag{10}$$

$$i \in \left\{CO_3^{2-}\right\}$$

Total carbonates = 
$$[H_2CO_3] + [HCO_3^-] + [CO_3^{2-}]$$
 (11)

Table 4:  $pK_{sp}$  values for the considered aqueous phase chemical systems.

Name	Chemical system	$pK_{sp}$	Source
Ammonia	$NH_4^+ \leftrightarrow NH_3 + H^+$	9.2	(Bates & Pinching, 1949)
Water	$H_2O \leftrightarrow OH^- + H^+$	14	(Skoog, West, Holler, & Crouch, 2014)
	$H_3PO_4 \leftrightarrow H_2PO_4^- + H^+$	2.1	(Ohlinger, Young, & Schroeder, 1998)
Phosphoric acid	$H_2PO_4^- \leftrightarrow HPO_4^{2-} + H^+$	7.2	(Ohlinger et al., 1998)
	$HPO_4^{2-} \leftrightarrow PO_4^{3-} + H^+$	12.35	(Ohlinger et al., 1998)
Carbonic acid	$H_2CO_3 \leftrightarrow HCO_3^- + H^+$	6.35	(Skoog et al., 2014)
Carbonic acid	$HCO_3^- \leftrightarrow CO_3^{2-} + H^+$	10.33	(Skoog et al., 2014)

# 4 Surrogate models to estimate the formation of precipitates from livestock organic waste

### 4.1 Influence of magnesium

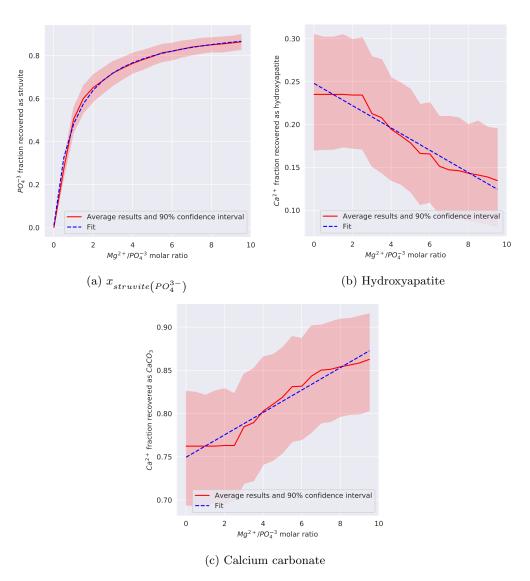


Figure 2: Evolution phosphorus as phosphate fraction recovered as struvite and calcium fraction recovered as precipitates along  ${\rm Mg^{2+}/PO_4^{3-}}$  molar ratio values considering 50 different composition data sets.

#### 4.1.1 Influence of calcium

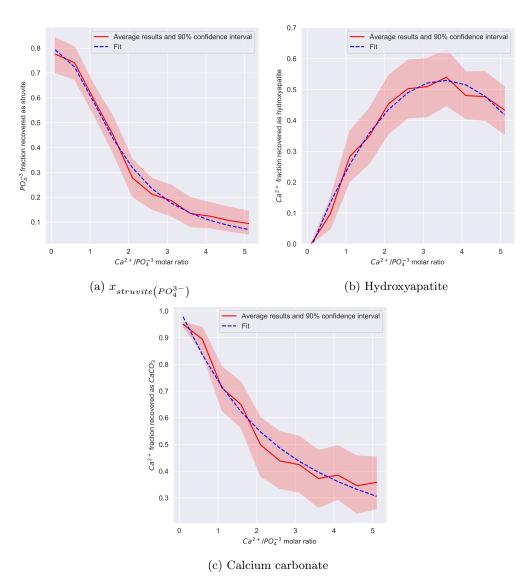


Figure 3: Evolution of phosphorus as phosphate fraction recovered as struvite and calcium fraction recovered as precipitates along  $\mathrm{Ca}^{2+}/\mathrm{PO}_4^{3-}$  molar ratio values considering 50 different composition data sets.

### 4.1.2 Influence of alkalinity

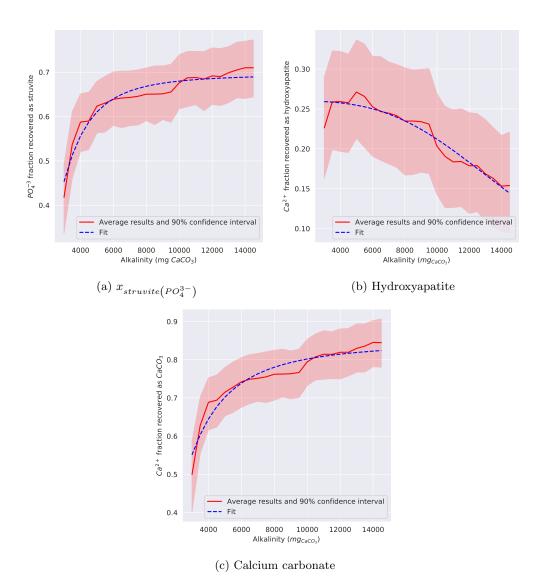


Figure 4: Evolution of phosphorus as phosphate fraction recovered as struvite and calcium fraction recovered as precipitates among alkalinity values considering 50 different composition data sets.

### References

- Alburquerque, J., de la Fuente, C., & Bernal, M. (2012). Chemical properties of anaerobic digestates affecting c and n dynamics in amended soils. Agric. Ecosyst. Environ., 160, 15-22.
- Bates, R. G., & Pinching, G. (1949). Acid dissociation constant of ammonium ion at 0-50 °C, and the base strength of ammonia. U.S. J. of Res. of the National Bureau of Standard, 42, 419-430.
- Bolzonella, D., Fatone, F., Gottardo, M., & Frison, N. (2018). Nutrients recovery from anaerobic digestate of agro-waste: Techno-economic assessment of full scale applications. *J. Environ. Manage.*, 216, 111-119.
- Dickson, A. (1981). An exact definition of total alkalinity and a procedure for the estimation of alkalinity and total inorganic carbon from titration data. *Deep-Sea Res.*, 28(6), 609-623.
- Duong, T. (2001). An introduction to kernel density estimation. University of Western Australia.
- Gell, K., van Groenigen, J., & Cayuela, M. (2011). Residues of bioenergy production chains as soil amendments: Immediate and temporal phytotoxicity. J. Hazard. Mater., 186, 2017-2025.
- Kirchmann, H., & Witter, E. (1992). Composition of fresh, aerobic and anaerobic farm animal dungs. Bioresour. Technol., 40(2), 137-142.
- Ledda, C., Schievano, A., Scaglia, B., Rossoni, M., G., A. F. F., & Adani, F. (2016). Integration of microalgae production with anaerobic digestion of dairy cattle manure: an overall mass and energy balance of the process. J. Clean. Prod., 112, 103-112.
- Løes, A., Pommeresche, R., & Khalil, R. (2017). Effects of marble application to manure and anaerobic digestates (Tech. Rep.). NORSØK.
- Martin, J. (2003). A comparison of dairy cattle manure management with and without anaerobic digestion and biogas utilization (Tech. Rep.). AgSTAR Program, U.S. Environmental Protection Agency.
- Möller, K., & Müller, T. (2012). Effects of anaerobic digestion on digestate nutrient availability and crop growth: A review. *Eng. Life Sci.*, 12(3), 242-257.
- Möller, K., Stinner, W., Deuker, A., & Leithold, G. (2008). Effects of different manuring systems with and without biogas digestion on nitrogen cycle and crop yield in mixed organic dairy

- farming. Nutr. Cycl. Agroecosyst., 82(3), 209-232.
- Moset, V., Fontaine, D., & Møller, H. (2017). Co-digestion of cattle manure and grass harvested with different technologies. effect on methane yield, digestate composition and energy balance. *Energy*, 141, 451-460.
- Normak, A., Suurpere, J., Orupõld, K., Jõgi, E., & Kokin, E. (2012). Simulation of anaerobic digestion of cattle manure. Agronomy Research Biosystem Engineering Special Issue, 1, 167-174.
- Ohlinger, K. N., Young, T., & Schroeder, E. (1998). Predicting struvite formation in digestion.

  Wat. Res., 32(12), 3607-3614.
- Rigby, H., & Smith, S. (2011). New markets for digestate from anaerobic digestion (Tech. Rep.). WRAP (Waste and Resources Action Programme).
- Risgberg, K., Cederlund, H., Pell, M., Arthurson, V., & Schnürer, A. (2017). Comparative characterization of digestate versus pig slurry and cow manure chemical composition and effects on soil microbial activity. *Waste Manag.*, 61, 529-538.
- Seppälä, M., Pyykkönen, V., Väisänen, A., & Rintala, J. (2013). Biomethane production from maize and liquid cow manure - effect of share of maize, post-methanation potential and digestate characteristics. Fuel, 107, 209-216.
- Skoog, D., West, D., Holler, F., & Crouch, S. (2014). Fundamentals of analytical chemistry. Cengage Learning.
- Smith, K., Grylls, J., Metcalfe, P., Jeffrey, B., & Sinclair, A. (2007). Nutrient value of digestate from farm-based biogas plants in scotland (Tech. Rep.). Scottish Executive Environment and Rural Affairs Department.
- Sørensen, P., Mejnertsen, P., & Møller, H. (2011). Nitrogen fertilizer value of digestates from anaerobic digestion of animal manures and crops. *NJF Report*, 7(8), 42-44.
- Tampere, M., & Viiralt, R. (2014). The efficiency of biogas digestate on grassland compared to mineral fertilizer and cattle slurry. *Research for Rural Development*, 1, 89-94.
- Walsh, J., Jones, D., Edwards-Jones, G., & Williams, A. (2012). Replacing inorganic fertilizer with anaerobic digestate may maintain agricultural productivity at less environmental cost. J. Plant. Nutr. Soil Sci., 175(6), 840-845.
- Wolf-Gladrow, D., Zeebe, R., Klaas, C., Körtzinger, A., & Dickson, A. (2007). Total alkalinity:

- The explicit conservative expression and its application to biogeochemical processes. Mar. Chem., 106, 287-300.
- Xia, M., Tao, W., Wang, Z., & Pei, Y. (2012). Treatment of anaerobically digested dairy manure in a two-stage biofiltration system. Water Sci. Technol., 65 (11), 1975-1981.
- Zheng, Y., Ke, L., Xia, D., Zheng, Y., Wang, Y., Li, H., & Li, Q. (2016). Enhancement of digestates dewaterability by ctab combined with cfa pretreatment. Sep. Purif. Technol., 163, 282-289.