# Directional NK Paper Supplement

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## Contents

## 0.1 Introduction

This document (created using Rmarkdown) contains: 1) all analysis code used to generate figures for this paper, 2) supplemental figures that did not fit in the main body of the paper.

First, we set up our R environment:

```
library(ggplot2)
library(reshape)
library(dplyr)
library(tidyr)
library(readr)
library(ggraph)
library(tidygraph)
```

Next, we load in all our data, which are expected to be in the directory above the one where this script is.

```
# NOTE: This cell will take quite a while to run

base <- "../"

# Read in the file that contains the names of the output documents
files <- read.table(paste(base, "files.txt", sep = ""), stringsAsFactors = F)
nval <- 15
ntimes = nrow(files)

# Compile the summary statistics, populations, and (reproductive) landscapes
# into individual data frames row <- 1
filename <- files[1,]
subfolder <- strsplit(filename, "/")[[1]][1]
subfile <- strsplit(filename, "/")[[1]][2]

# read in the statistics
trace <- read.table(paste(base, subfolder, "/stats-", subfile, sep = ""),
                    header = T, stringsAsFactors = F)

# Read in the population output
pops <- read.table(paste(base, subfolder, "/popt-", subfile, sep = ""),
                    header = T, stringsAsFactors = F,
```

```r
                        col.names =
                          c("Gen", "Genome", "Count", "Fitness", "ShockFitness"))

# get N, K, mu, replicate (seed), and regime arguments
subpieces <- strsplit(subfile, "-")
kval <- as.numeric(substring(subpieces[[1]][2], 2))

# if seed is followed by another argument
# seed <- as.numeric(substring(subpieces[[1]][3], 2))

# if seed ends argument list
seed <- as.numeric(substring(strsplit(subpieces[[1]][3], "[.]")[[1]][1], 2))
muval <- ifelse(length(subpieces[[1]]) == 5,
                as.numeric(substring(subpieces[[1]][4], 2)),
                as.numeric(paste(substring(subpieces[[1]][4], 2),
                                 subpieces[[1]][5], sep = "-")))
regparam <- subpieces[[1]][length(subpieces[[1]])]
regval <- ifelse(length(regmatches(regparam, gregexpr("S", regparam))[[1]]) > 0,
                 "Sudden",
                 "Gradual")

# read in the reproductive landscape (requires the N value and the K value).
# For the shock landscape, use "/sland-" instead of "/land-".
peaks <- read.table(paste(base, subfolder, "/land-", subfile, sep = ""),
                    skip = (6+nval+2^(kval+1)),
                    col.names = c("Genome", "Fitness"))
shock_peaks <- read.table(paste(base, subfolder, "/sland-", subfile, sep = ""),
                          skip = (6+nval+2^(kval+1)),
                          col.names = c("Genome", "Fitness"))

# Assign treatment parameters to each data frame
trace$k <- kval
pops$k <- kval
peaks$k <- kval
shock_peaks$k <- kval
trace$n <- nval
pops$n <- nval
peaks$n <- nval
shock_peaks$n <- nval
trace$seed <- seed
pops$seed <- seed
peaks$seed <- seed
shock_peaks$seed <- seed
trace$regime <- regval
pops$regime <- regval
peaks$regime <- regval
shock_peaks$regime <- regval
trace$mu <- muval
pops$mu <- muval
peaks$mu <- muval
shock_peaks$mu <- muval
for (f in 2:nrow(files)){
  filename <- files[f,]
```

```r
subfolder <- strsplit(filename, "/")[[1]][1]
subfile <- strsplit(filename, "/")[[1]][2]
thistrace <- read.table(paste(base, subfolder, "/stats-", subfile, sep = ""),
                        header = T,
                        stringsAsFactors = F)
thispop <- read.table(paste(base, subfolder, "/popt-", subfile, sep = ""),
                      header = T,
                      stringsAsFactors = F,
                      col.names =
                        c("Gen", "Genome", "Count",
                          "Fitness", "ShockFitness"))
subpieces <- strsplit(subfile, "-")
kval <- as.numeric(substring(subpieces[[1]][2], 2))
#seed <- as.numeric(substring(subpieces[[1]][3], 2))
seed <- as.numeric(substring(strsplit(subpieces[[1]][3], "[.]")[[1]][1], 2))
regparam <- subpieces[[1]][length(subpieces[[1]])]
regval <- ifelse(length(regmatches(regparam,
                                   gregexpr("S", regparam))[[1]]) > 0,
                 "Sudden",
                 "Gradual")
#print(paste(seed, regval))
muval <- ifelse(length(subpieces[[1]]) == 5,
                as.numeric(substring(subpieces[[1]][4], 2)),
                as.numeric(paste(substring(subpieces[[1]][4], 2),
                                 subpieces[[1]][5], sep = "-")))
thispeak <- read.table(paste(base, subfolder, "/land-", subfile, sep = ""),
                       skip = (6+nval+2^(kval+1)),
                       col.names = c("Genome", "Fitness"))
thisshockpeak <- read.table(paste(base,
                                  subfolder,
                                  "/sland-",
                                  subfile,
                                  sep = ""),
                            skip = (6+nval+2^(kval+1)),
                            col.names = c("Genome", "Fitness"))
thistrace$k <- kval
thispop$k <- kval
thispeak$k <- kval
thisshockpeak$k <- kval
thistrace$n <- nval
thispop$n <- nval
thispeak$n <- nval
thisshockpeak$n <- nval
thistrace$seed <- seed
thispop$seed <- seed
thispeak$seed <- seed
thisshockpeak$seed <- seed
thistrace$regime <- regval
thispop$regime <- regval
thispeak$regime <- regval
thisshockpeak$regime <- regval
thistrace$mu <- muval
thispop$mu <- muval
```

```
  thispeak$mu <- muval
  thisshockpeak$mu <- muval
  trace <- rbind(trace, thistrace)
  pops <- rbind(pops, thispop)
  peaks <- rbind(peaks, thispeak)
  shock_peaks <- rbind(shock_peaks, thisshockpeak)
}

# Create regime label for future plotting
shock_peaks$Landscape <- "shock"
peaks$Landscape <- "reproduction"
```

Next, we do some additional post-processing:

```
# Check number of persisting generations
ntimes <- length(unique(trace$seed))
survivals <- data.frame(n = rep(NA, ntimes),
                        k = rep(NA, ntimes),
                        seed = rep(NA, ntimes),
                        regime = rep(NA, ntimes),
                        cutoff = rep(NA, ntimes),
                        gen = rep(NA, ntimes),
                        uniques = rep(NA, ntimes),
                        average = rep(NA, ntimes),
                        stdev = rep(NA, ntimes),
                        max = rep(NA, ntimes),
                        shockcut = rep(NA, ntimes),
                        shockAvg = rep(NA, ntimes),
                        shockStd = rep(NA, ntimes),
                        shockMax = rep(NA, ntimes),
                        mu = rep(NA, ntimes),
                        diversity = rep(NA, ntimes),
                        evenness = rep(NA, ntimes),
                        population = rep(NA, ntimes))
for(s in 1:ntimes){
  sd <- unique(trace$seed)[s]
  sub <- subset(trace, seed == sd)
  survivals[s, ] <- subset(sub,
                           gen == max(sub$gen),
                           select = c(n, k, seed, regime, cutoff, gen, uniques,
                                      average, stdev, max, shockCut, shockAvg,
                                      shockStd, shockMax, mu,
                                      diversity, evenness, population))
}

# Adjust significant digits on mutation rate for plotting
survivals$mu <- format(survivals$mu, digits=3, drop0trailing = TRUE, format="g")
survivals <- survivals %>% mutate(mu = case_when(
  mu=="0.066667" ~ "0.0667",
  TRUE ~ mu
  )
)
```

## 0.2 Analysis

### 0.2.1 Survival probability

First, we assess the effect of different evolutionary regimes on the probability of a population surviving until
the end of the experiment:

```r
pd <- position_dodge(.5)
ggplot(data = survivals %>%  # Do some additional processing of the data
                # Only grab data points from the final generation
                # (eliminates populations that went extinct)
                filter(gen == 51) %>%
                # Calculate percentages per condition
                # (combo of k, regime, and mu)
                group_by(k, regime, mu) %>%
                summarise(
                  # The sudden regime had 40 replicates
                  # while the gradual regime had 20
                  percent_survive = ifelse(
                                        regime == "Sudden",
                                        n()/40,
                                        n()/20
                                    )
                ),
       # Plot each regime as a different color with k as x axis
       aes(x = as.factor(k), fill = regime)) +
  scale_x_discrete() +
  # Plot data as bars indicating proportion of population that survived
  geom_bar(
    aes(y=percent_survive),
    position = position_dodge(.92),  # Put bars for each regime side by side
    stat = "identity"
  ) +
  theme_bw(base_size = 14) +
  theme(panel.grid.minor = element_blank(),
    panel.grid.major = element_line(color = "grey91"),
    strip.text.y = element_text(face = "italic"),
    legend.position = c(.85, .25)) +
  # Make a panel for each mutation rate
  facet_wrap(~mu, labeller = labeller(mu=label_both)) +
  scale_fill_viridis_d(name = "Treatment") +
  ylab("Proportion of persisting populations") +
  xlab("K")
```

### 0.2.2 Final fitness

Next, we compare the maximum fitness at the end of the run (in both landscapes) across conditions (for runs
that did not go extinct):

```r
# Pivot shock and reproductive fitness into a single column
# so we can easily plot a comparison
ggplot(data = survivals %>%
                # Grab only final generation,
                # to exclude populations that went extinct early
                filter(gen==51) %>%
                pivot_longer(c(max, shockMax),
```

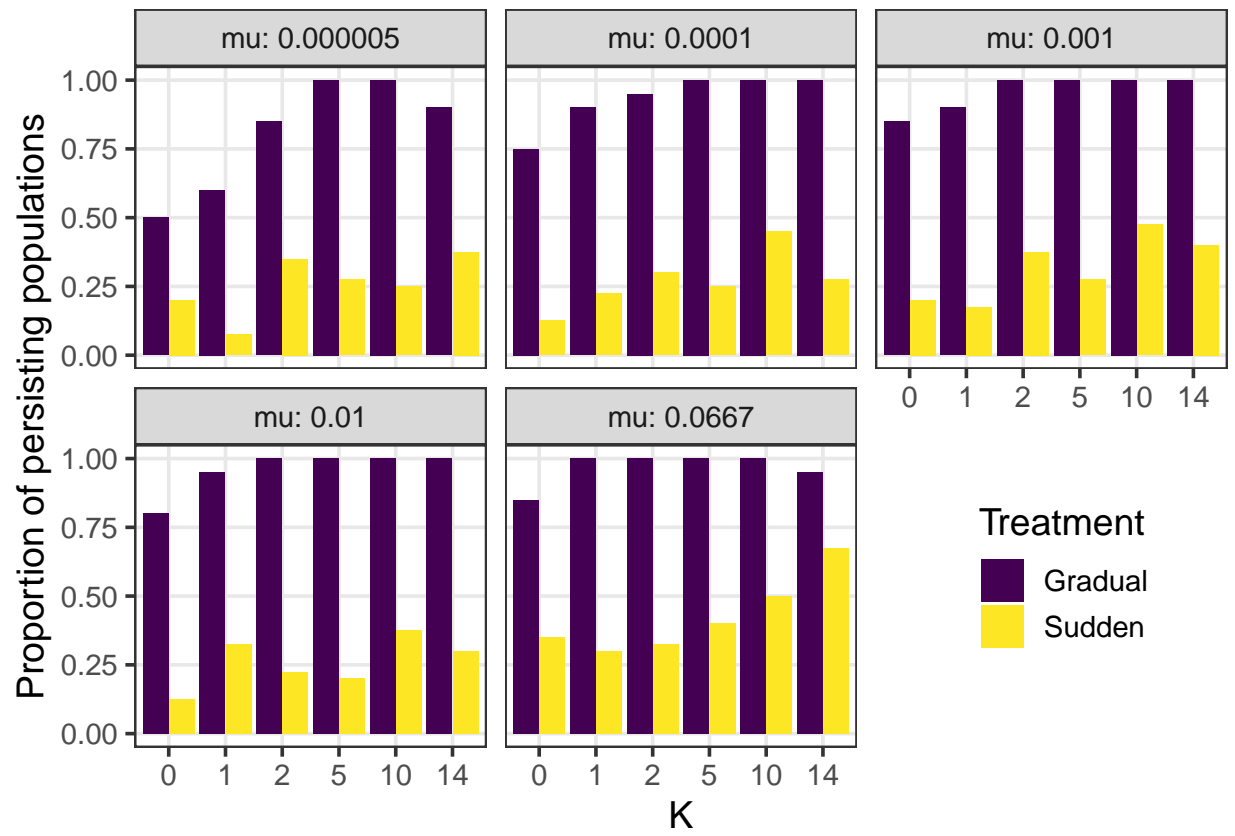Figure 1: Proprtion of replicates surviving until end

```
                        names_to="Landscape",
                        values_to="max_fitness") %>%
               mutate(Landscape = case_when(
                                  Landscape == "max" ~ "Reproduction",
                                  Landscape=="shockMax" ~ "Shock")
                       ),
       aes(x = as.factor(k),
           y = max_fitness,
           color = regime,
           shape=Landscape,
           linetype=regime, # Give regimes different linetype for greyscale
           alpha=Landscape  # Give landscapes different opacity for greyscale
       )
) +
# Plot a bootstrapped confidence interval around the mean
stat_summary(fun.data = "mean_cl_boot", geom = "pointrange", position = pd) +
theme_bw(base_size = 14) +
theme(panel.grid.minor = element_blank(),
      panel.grid.major = element_line(color = "grey91"),
      strip.text.y = element_text(face = "italic"),
      legend.position = c(.85, .20)) +
scale_color_hue(name = "Treatment") +
scale_alpha_discrete(range = c(1,.6)) +  # Set alpha levels the landscapes
facet_wrap(~mu, labeller = labeller(mu = label_both)) +
ylab("Final maximum population fitness") +
scale_linetype("Treatment") +
xlab("K")
```

We make the same plot for average fitness (recall that there the shock landscape is the only source of death, so there are many low fitness individuals in the population at any point in time):

```
ggplot(data = survivals %>%
                 filter(gen==51) %>%
                 pivot_longer(c(average, shockAvg),
                              names_to="Landscape",
                              values_to="max_fitness") %>%
                 mutate(Landscape = case_when(
                                    Landscape == "average" ~ "Reproduction",
                                    Landscape=="shockAvg" ~ "Shock")
                         ),
       aes(x = as.factor(k),
           y = max_fitness,
           color = regime,
           shape=Landscape,
           linetype=regime, alpha=Landscape)
       ) +
 stat_summary(fun.data = "mean_cl_boot", geom = "pointrange", position = pd) +
 theme_bw(base_size = 14) +
 theme(panel.grid.minor = element_blank(),
       panel.grid.major = element_line(color = "grey91"),
       strip.text.y = element_text(face = "italic"),
       legend.position = c(.85, .2)) +
 scale_color_hue(name = "Treatment") +
 facet_wrap(~mu, labeller = labeller(mu = label_both)) +
```
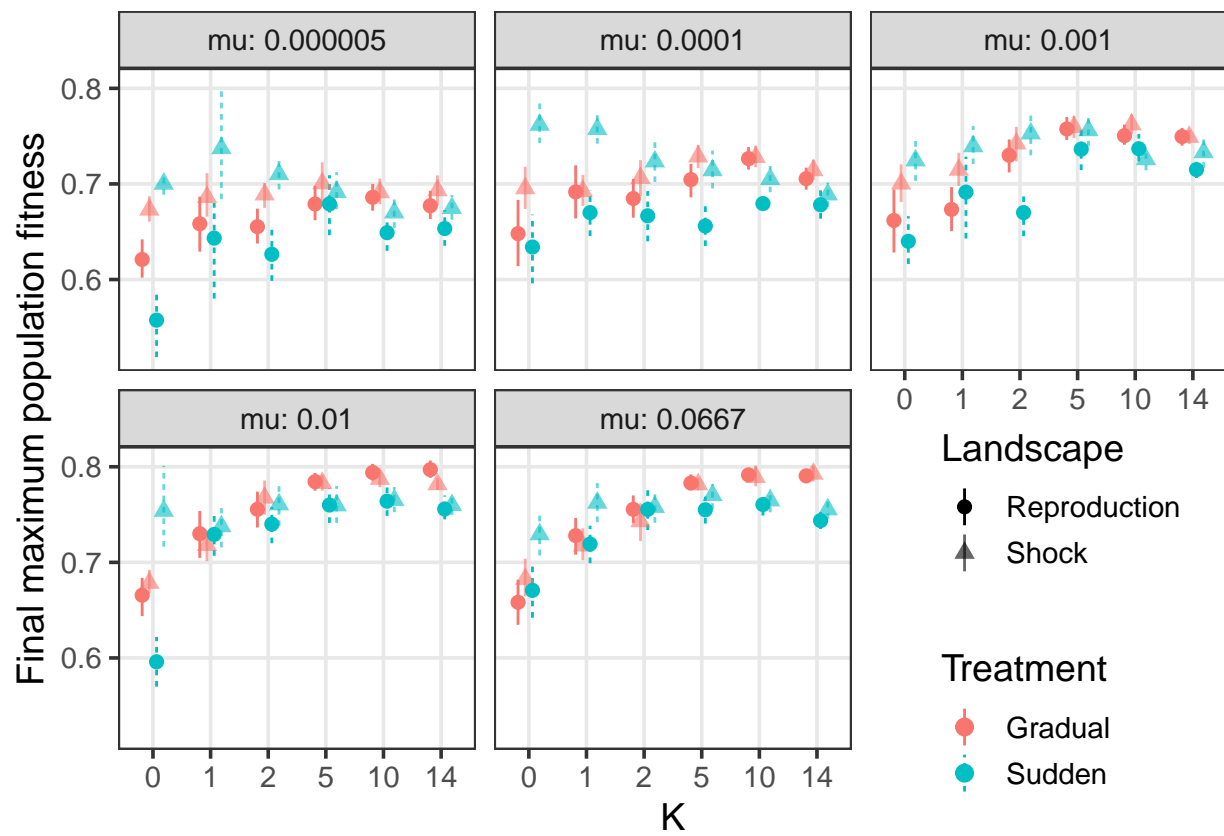
Figure 2: Final maximum fitness

```
ylab("Final average population fitness") +
scale_alpha_discrete(range = c(1,.6)) +
scale_linetype("Treatment") +
xlab("K")
```
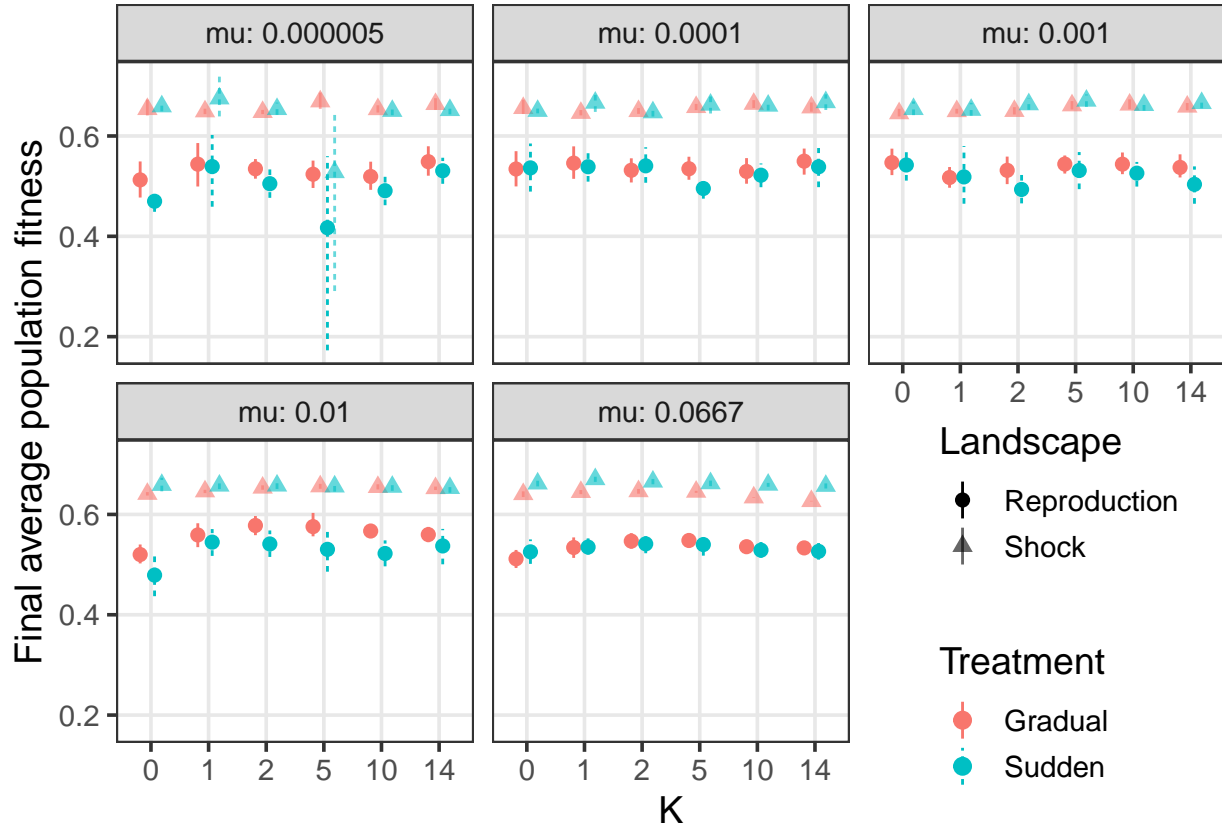


Figure 3: Final average fitness

### 0.2.3   Fitness timeseries analysis

Next, we analyze the behavior of maximum fitness (within each individual population) over time:

```
ggplot(data = trace %>%
          filter(mu==.01, k==10),   # Plot a single representative combination of mu and k
       aes(x = gen, y = max)
) +
facet_wrap(~regime) +   # A panel for each shock regime
# Bootstrapped 95% CI for repro landscape
stat_summary(fun.data="mean_cl_boot",
             geom="ribbon",
             alpha=.5, aes(fill="Reproduction")) +
# Avg max fitness for repro landscape
stat_summary(fun.data="mean_cl_boot",
             geom="line",
             alpha=.5,
             aes(color="Reproduction")) +
# Bootstrapped 95% CI for shock landscape
stat_summary(aes(y=shockMax, fill="Shock"),
```

```
                fun.data="mean_cl_boot",
                geom="ribbon",
                alpha=.5) +
    # Avg max fitness for shock landscape
    stat_summary(aes(y=shockMax, color="Shock"),
                fun.data="mean_cl_boot",
                geom="line",
                alpha=.5) +
theme_bw(base_size = 14) +
theme(panel.grid.minor = element_blank(),
      panel.grid.major = element_line(color = "grey91"),
      strip.text.y = element_text(face = "italic"),
      legend.position = "bottom")  +
# Clean up legend
scale_color_manual("Landscape",
                    values = c("Reproduction"="#440154", "Shock"="#fde725")) +
scale_fill_manual("Landscape",
                    values = c("Reproduction"="#440154", "Shock"="#fde725")) +
ylab("Average Maximum Fitness") +
xlab("Generation")
```
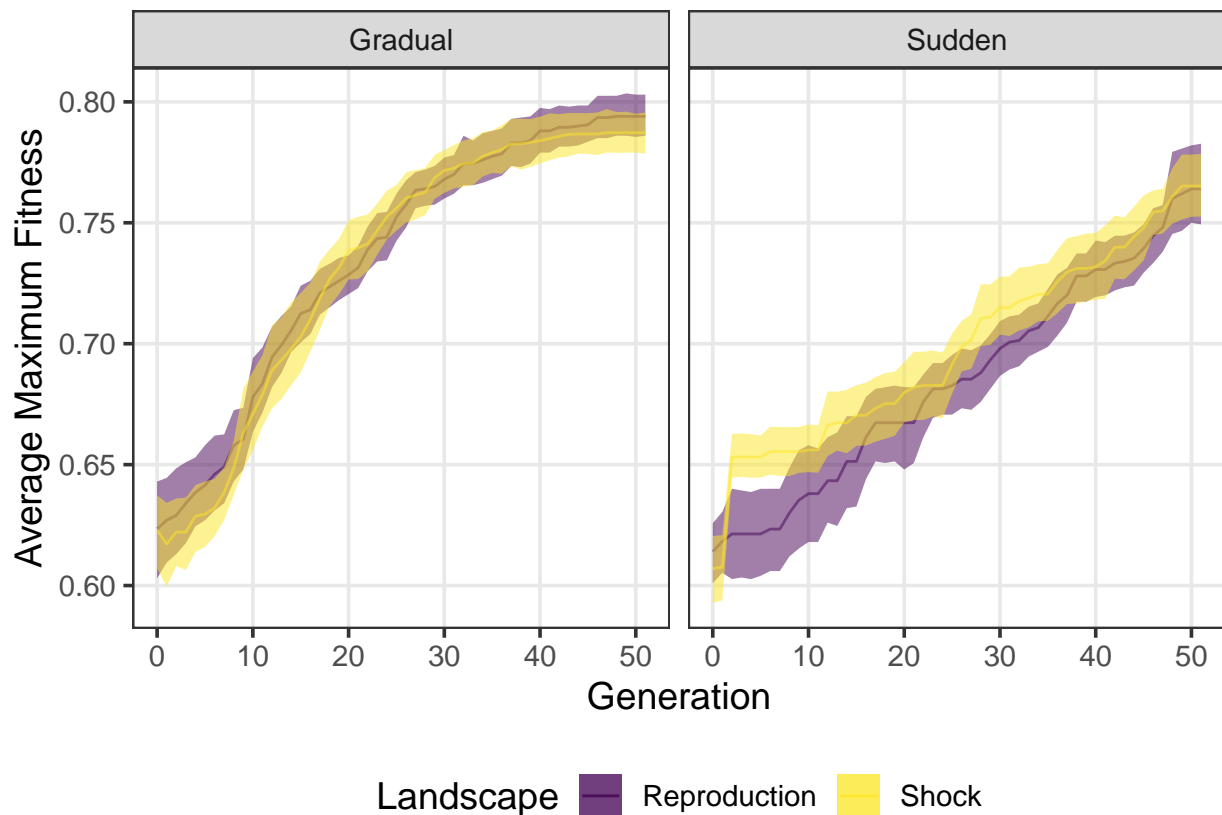


Figure 4: Maximum fitness over time

And we do the same for average fitness:

```
ggplot(data = trace %>%
        filter(mu==.01, k==10),
    aes(x = gen, y = average)) +
```

```r
  facet_wrap(~regime) +
  stat_summary(fun.data="mean_cl_boot",
               geom="ribbon",
               alpha=.5,
               aes(fill="Reproduction")) +
  stat_summary(fun.data="mean_cl_boot",
               geom="line",
               alpha=.5,
               aes(color="Reproduction")) +
  stat_summary(aes(y=shockAvg, fill="Shock"),
               fun.data="mean_cl_boot",
               geom="ribbon",
               alpha=.5) +
  stat_summary(aes(y=shockAvg, color="Shock"),
               fun.data="mean_cl_boot",
               geom="line",
               alpha=.5) +
  theme_bw(base_size = 14) +
  theme(panel.grid.minor = element_blank(),
        panel.grid.major = element_line(color = "grey91"),
        strip.text.y = element_text(face = "italic"),
        legend.position = "bottom")  +
  scale_color_manual("Landscape",
                     values = c("Reproduction"="#440154", "Shock"="#fde725")) +
  scale_fill_manual("Landscape",
                     values = c("Reproduction"="#440154", "Shock"="#fde725")) +
  ylab("Average Mean Fitness") +
  xlab("Generation")
```

### 0.2.4   Diversity

Diversity clearly plays an important role in the dynamics we observe. Here, we plot the number of unique genotypes over time in each condition:

```r
ggplot(data = trace %>%
         filter(mu==.01, k==10),
       aes(x = gen, y = uniques)) +
  facet_wrap(~regime) +  # One panel for each regime
  # Bootstrapped 95% CI around mean count of unique genotypes
  stat_summary(fun.data="mean_cl_boot", geom="ribbon", alpha=.5) +
  # Mean count of unique genotypes
  stat_summary(fun.data="mean_cl_boot", geom="line", alpha=.5) +
  theme_bw(base_size = 14) +
  theme(panel.grid.minor = element_blank(),
        panel.grid.major = element_line(color = "grey91"),
        strip.text.y = element_text(face = "italic"),
        legend.position = "none")   +
  ylab("Unique genotypes") +
  xlab("Generation")
```

A lot of this trend is likely being driven by population size, so it may also be worth looking at the same plot normalized by population:

```r
ggplot(data = trace %>%
         filter(mu==.01, k==10),
```
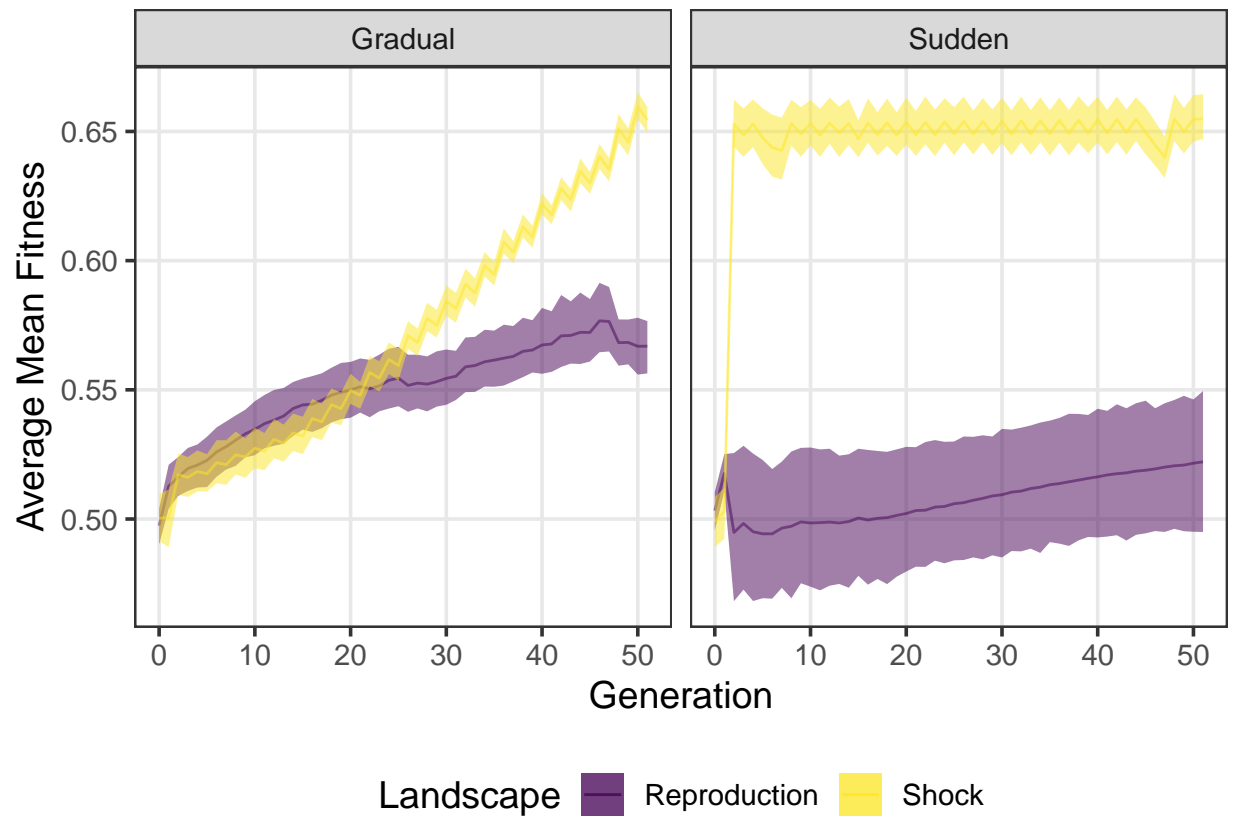
Figure 5: Average fitness over time
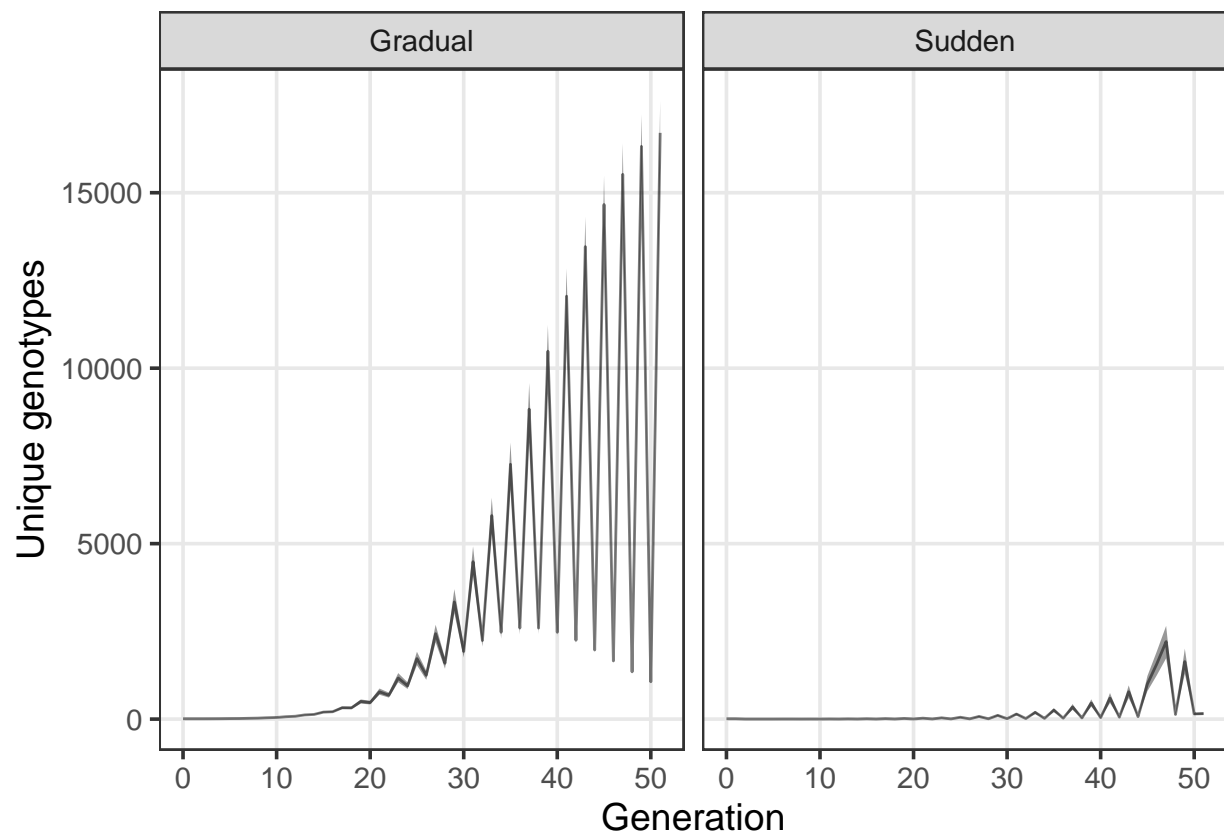
Figure 6: Unique genotypes over time

```
        aes(x = gen, y = uniques/population)) +
    facet_wrap(~regime) +  # One panel for each regime
    # Bootstrapped 95% CI around mean count of unique genotypes
    stat_summary(fun.data="mean_cl_boot", geom="ribbon", alpha=.5) +
    # Mean count of unique genotypes
    stat_summary(fun.data="mean_cl_boot", geom="line", alpha=.5) +
    theme_bw(base_size = 14) +
    theme(panel.grid.minor = element_blank(),
          panel.grid.major = element_line(color = "grey91"),
          strip.text.y = element_text(face = "italic"),
          legend.position = "none")   +
    ylab("Unique genotypes") +
    xlab("Generation")
```
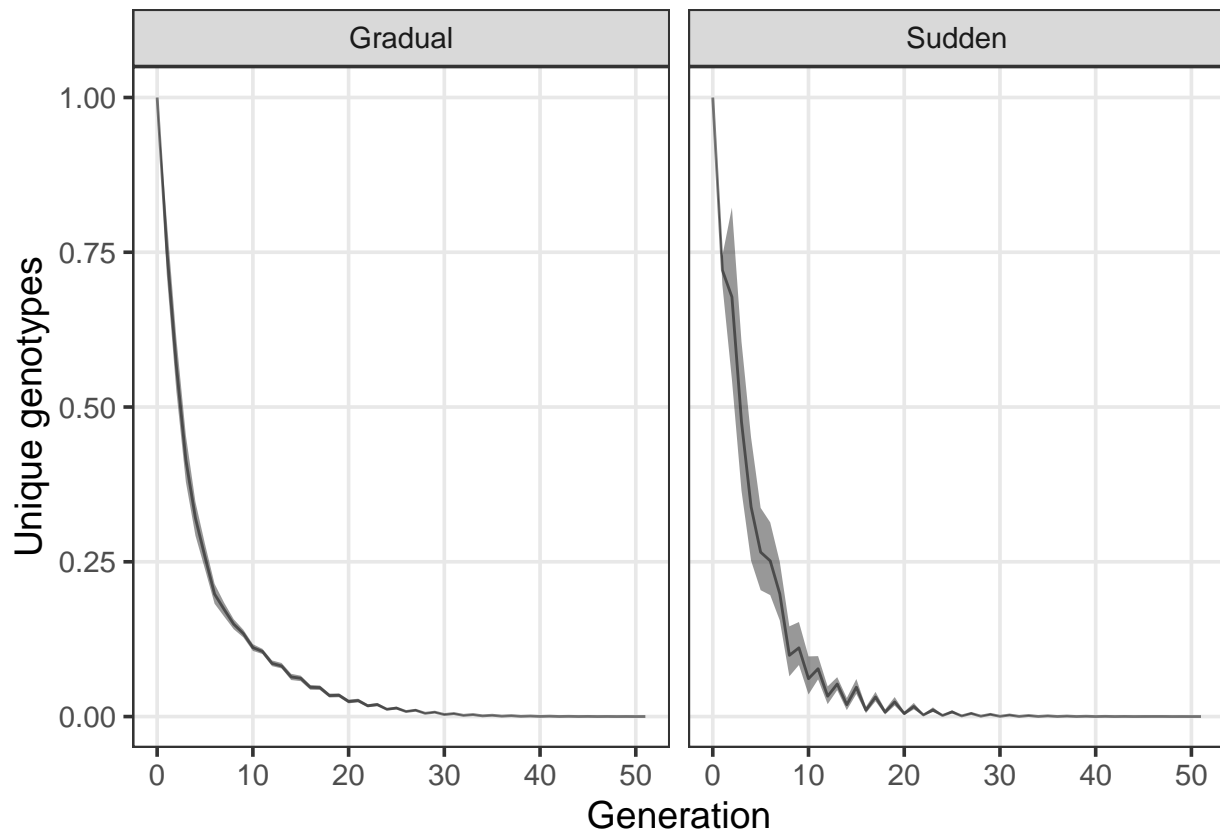


Figure 7: Unique genotypes over time, normalized by population size

### 0.2.5    Landscape analysis

We see relatively high fitnesses in both landscapes. The next obvious set of questions to ask concern the way the population is traversing the two landscapes.

One possibility is that successful populations are finding regions of the fitness landscape that are highly fit in both landscapes. To assess this possibility, we measure the correlation between reproductive fitness and shock fitness among evolved genotypes in the population. We measure this correlation for each population:

```
# Make new dataframe with all the correlations
df <- pops %>%
```

```r
        # Group by run (as indicated by random seed used for each run)
        # and by genome within run
        group_by(seed, Genome) %>%
        arrange(Gen) %>%  # Sort by generation so time series are in order
        # Group by run (as indicated by random seed used for each run)
        # and by genome within run
        filter(last(Gen)==50, first(Gen) < 48, row_number() == 1) %>%
        # Condense each genome into a single row -
        # these values should all be identical across rows, so first is as good
        # as any other value
        summarise(Gen = first(Gen),
                  Fitness= first(Fitness),
                  ShockFitness=first(ShockFitness),
                  regime=first(regime),
                  k=first(k),
                  mu=first(mu),
                  .groups = "drop_last"  # Keep data frame grouped by run
                  ) %>%
        # Calculate correlation between fitness in each environment for run
        summarise(r=cor(Fitness, ShockFitness, method="spearman"),
                  regime=first(regime),
                  k=first(k),
                  mu=first(mu)
                  )

# Clean up significant digits on mutation rate for label
df$mu <- format(df$mu, digits=3, drop0trailing = TRUE, format="g")
df <- df %>% mutate(mu = case_when(
  mu=="0.066667" ~ "0.0667",
  TRUE ~ mu
  )
)

# Actually plot the data
ggplot(df %>% filter(k== 10)) +
  geom_boxplot(aes(x=as.factor(mu), y=r, fill=regime)) +
  xlab("Mu") +
  scale_fill_viridis_d("Regime") +
  ylab("Correlation between reproductive\nand shock fitness") +
  theme_bw(base_size = 14) +
  theme(panel.grid.minor = element_blank(),
        panel.grid.major = element_line(color = "grey91"),
        strip.text.y = element_text(face = "italic"),
        legend.position = "bottom")
```

This implies that different genotypes have high fitness in each landscape rather than a single genotype being good at both. To understand better what is happening, it would be helpful to more directly examine the traversal of these landscapes. Unfortunately, doing so is very challenging with an N=15 landscape due to the high dimensionality. To build intuition, we undertake a case study in a lower-dimensional landscape.

## 0.3 Lower dimensional case study

Here, we use a pair of N=6, K=1 fitness landscapes. We plot these landscapes as graphs in which noddes represent genotypes and edges represent single-mutation adjaceny between genotypes.
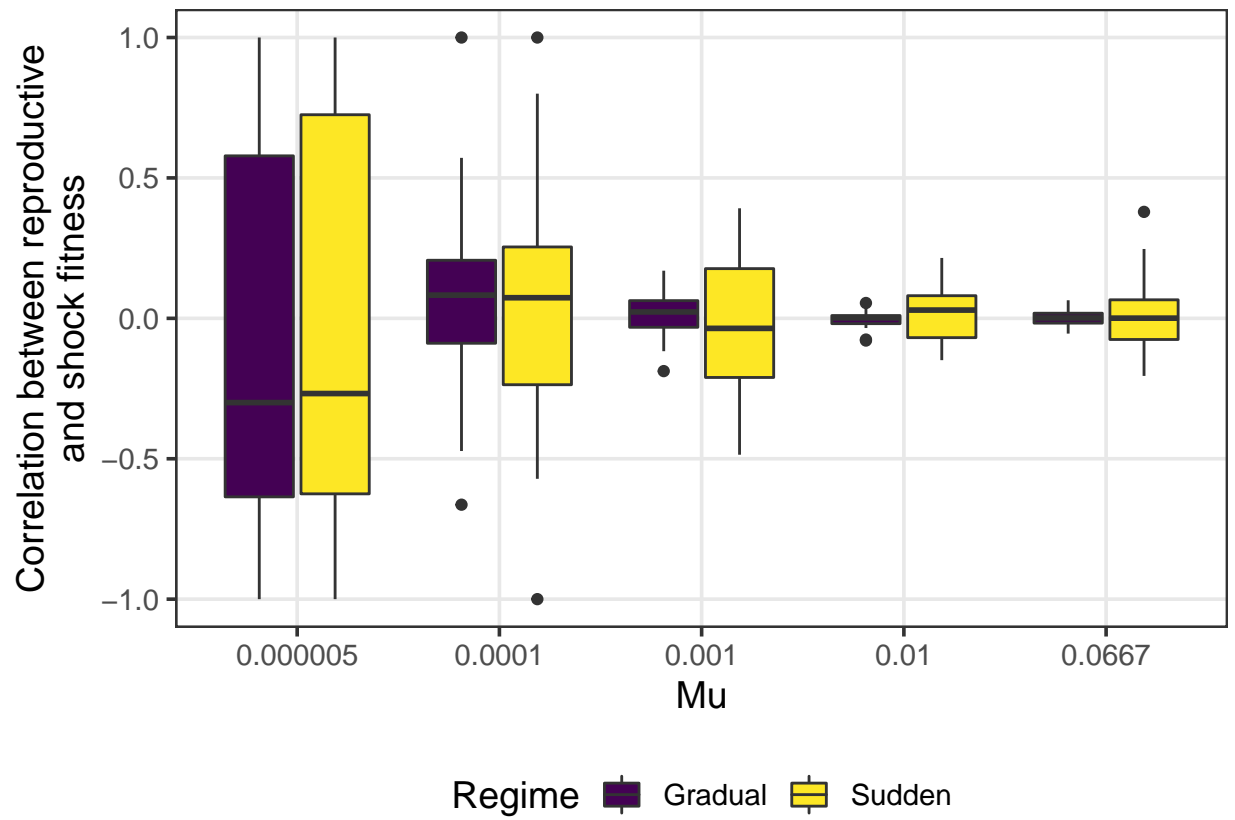
Figure 8: Correlation between reproductive and shock fitness in evolved genomes

First, we set up the data:

```r
# Read in a file containing all the edges for this fitness landscape graph
edges <- read_csv("edges_6.csv")

# Read in fitnesses from reproduction fitness landscape
repro_fits <- read_csv("repro_fits_1963.csv")

# Read in fitnesses from shock landscape
shock_fits <- read_csv("shock_fits_1963.csv")

# Clean up dataframes
colnames(repro_fits)[1] <- "name"
colnames(shock_fits)[1] <- "name"
colnames(shock_fits)[3] <- "shock_fitness"

# Make sure name is the same type in all data frames for easy joining
repro_fits$name <- as.character(repro_fits$name)
shock_fits$name <- as.character(shock_fits$name)
edges$from <- as.character(edges$from)
edges$to <- as.character(edges$to)

# Read in population time series data from run of evolutioon
pop <- read_table("../outs/popt-N6-k1-s1963-r0.001-Gradual.txt",skip = 3)

# Clean up data frame
colnames(pop) <- c("gen", "genome", "count", "fitness",
                   "shockfitness", "placeholder")
pop$placeholder <- NULL
pop$genome <- as.character(pop$genome)

# For now we're only worrying about the last generation
pop <- pop %>%filter(gen == max(gen))

# Build tidygraph data frame containing all the data we need
graph <- as_tbl_graph(edges)  # Make graph object
graph <- graph %>% full_join(repro_fits) %N>%  # Join in repro landscape data
                   # Set up data for positioning nodes in an intelligible way
                   group_by(ones) %>%
                   arrange(as.numeric(name)) %>%
                   mutate(pos = row_number(), total = n()) %>%
                   ungroup()
# Join in shock landscape data
graph <- graph %>% full_join(shock_fits, by = c("name", "ones"))
graph <- graph %>%  # Join in population data frame
         full_join(pop %>%  # Only want one row per genome so we summarize
                       group_by(genome) %>%
                       arrange(gen) %>%
                       summarise(count=max(count), gen=first(gen)),
                              by = c("name" = "genome")) %N>%
                       mutate(count=ifelse(is.na(count), -1, count),
                              gen=ifelse(is.na(gen), -1, gen)) %>%
                       mutate(count=ifelse(count > 1000, 1000, count))
```

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```
# Set up edge data
graph <- graph %E>%
        mutate(
          fitness = if_else(.N()$fitness[to] > .N()$fitness[from],
                            .N()$fitness[to],
                            .N()$fitness[from]),
          shock_fitness = if_else(.N()$shock_fitness[to] >
                                          .N()$shock_fitness[from],
                                  .N()$shock_fitness[to],
                                  .N()$shock_fitness[from]),
          followed = if_else(is.na(.N()$count[to]) |
                                  is.na(.N()$count[from]),
                              0,
                              if_else(.N()$count[to] > .N()$count[from],
                                      .N()$count[to],
                                      .N()$count[from]))
        )
```

Next, we visualize the data for the reproductive landscape:

```
ggraph(graph, layout = 'manual', x=max(ones)/2 * (pos)/(total+1), y=ones) +
    geom_edge_link(aes(color=fitness)) +
    geom_node_circle(aes(fill=fitness,
                         r = (.05*(count>0)) + sqrt(count)/200,
                         alpha=shock_fitness>=.66,
                         color=shock_fitness>=.66)) +
    theme(legend.position = "none") +
    # Set up legend and appearance
    theme_graph(base_family = 'Helvetica') +
    scale_fill_viridis_c("Fitness", limits=c(.3,.75)) +
    coord_fixed(ratio = .4) +
    scale_edge_color_viridis("Fitness", limits=c(.3,.75)) +
    scale_alpha_discrete("Can survive highest shock", range = c(.5,1)) +
    scale_color_manual("Can survive highest shock",
                       values=c("FALSE"="white", "TRUE"="black"),
                       guide = guide_legend(override.aes =
                                                  list(fill = "#008080") )) +
    guides(edge_color=FALSE)  # Edge color legend is same as fill
```

And the shock landscape:

```
ggraph(graph, layout = 'manual', x=max(ones)/2 * (pos)/(total+1), y=ones) +
    geom_edge_link(aes(color=shock_fitness)) +
    geom_node_circle(aes(fill=shock_fitness,
                         r = (.05*(count>0)) + sqrt(count)/200,
                         alpha=shock_fitness>=.66,
                         color=shock_fitness>=.66)) +
    theme(legend.position = "none") +
    # Set up legend and appearance
    theme_graph(base_family = 'Helvetica') +
    scale_fill_viridis_c("Fitness", limits=c(.3,.75)) +
    coord_fixed(ratio = .4) +
    scale_edge_color_viridis("Fitness", limits=c(.3,.75)) +
    scale_alpha_discrete("Can survive highest shock", range = c(.5,1)) +
    scale_color_manual("Can survive highest shock",
```
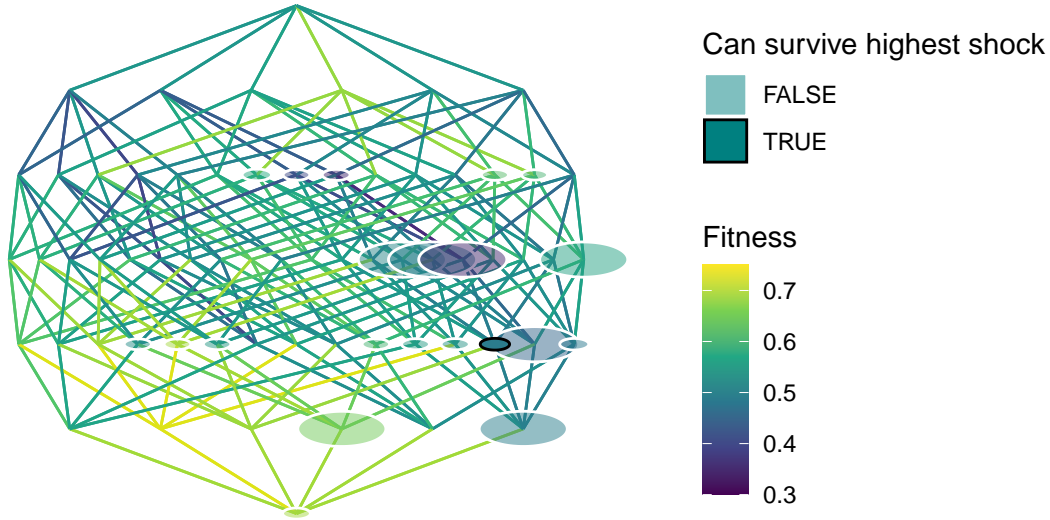
Figure 9: Graph of the full reproductive fitness landscape with final population size represented by node size

```
                    values=c("FALSE"="white", "TRUE"="black"),
                    guide = guide_legend(override.aes =
                                          list(fill = "#008080") )) +
    guides(edge_color=FALSE)  # Edge color legend is same as fill
```
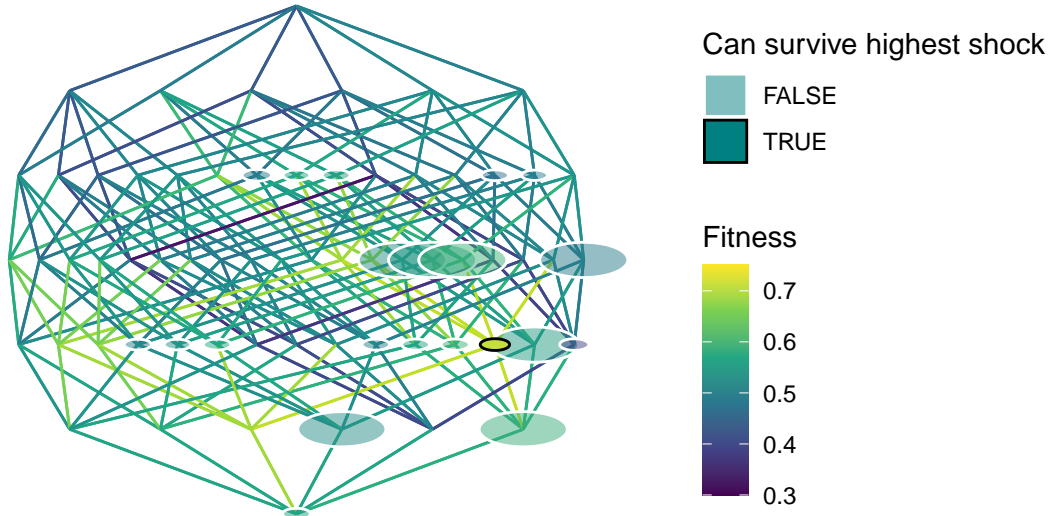


Figure 10: Graph of the full shock fitness landscape with final population size represented by node size

Note that in these graphs: node color indicates fitness in the reproductive and shock fitness landscapes respectively, edge color also indicates fitness in the landscape being visualized (specifically, the fitness of the fitter of the two genotypes being connected by the edge), and node size indicates final population size of that genotype. Node opacity indicates whether that genotype is capable of surviving the maximum shock in the shock landscape, as does node outline color.

From this data visualization, it appears that (in this run at least) the genotypes that are successful in each landscape are indeed very different. The population of genotypes that are fit in the reproductive landscape grows quickly but is wiped out at each shock event and most then be repopulated by mutations from the genotypes that can survive the shocks. This phenomenon explains the pattern of oscillations with increasing

amplitude observed in the graphs of unique genotypes and population size.