Script to analyze behavior data for *C. elegans* manuscript

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updated 04/27/2018 to include data and analysis for additional behavior experiments and update figure panels

Purpose:

To plot all behavior data for manuscript.

Background:

This is an R Markdown document. http://rmarkdown.rstudio.com.

The code required to get these graphs is in the gray boxes, along with information on how to run the statistical tests reported in the paper and qqPlots that were used to determine if these statistical tests were generally appropriate.

This was run in RStudio, using R version 3.3.1 (2016-10-31), Sincere Pumpkin Patch on a x86_64-apple-darwin13.4.0

Before these data can be analyzed, I used the following code block to:

- 1. Load all libraries required for analysis and plotting
- 2. Load these data
- 3. Subset data to use later to plot

```
#load libraries
library(plyr)
library(dplyr)
library(ggplot2)
library(ggthemes)
library(multcompView)
library(car)
allbeh <- read.csv('2018Dennis DEETandCelegans behaviordata.csv')
#start subsetting data:
hawaiian <- subset(allbeh,expt=='hawaiian')</pre>
#hawaiiand didn't have any positive controls that
# had enough responding animals to record
#so I excluded the day. temp in lab was also low that day.
hawaiian <- subset(hawaiian,date!=20160920)
hawaiiane <- subset(hawaiian,percentDEET==0)</pre>
hawaiiand <- subset(hawaiian,percentDEET==0.15)
```

```
rescue <- subset(allbeh,expt=='rescue'|expt=='rescue2')
rescueE <- subset(rescue,percentDEET==0)
rescueD <- subset(rescue,percentDEET==0.15)
spots <- subset(allbeh,expt=='spots')
screen <- subset(allbeh,expt=='screen2')
bacteria <- subset(allbeh,expt=='bacterial')
ASH <- subset(allbeh,expt=='ASH')
odordrc <- subset(allbeh,expt=='odordrc')</pre>
```

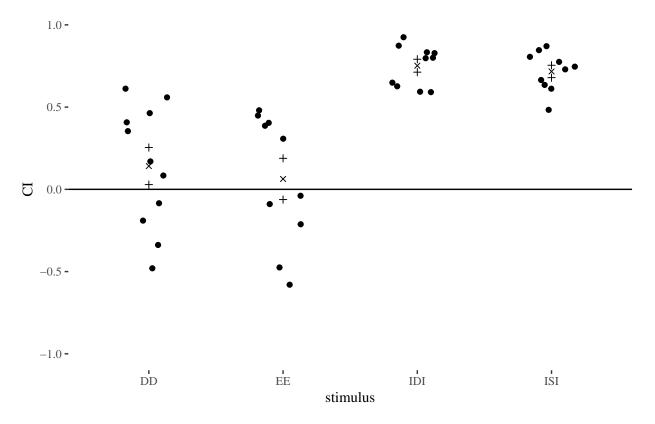
First, I tested if DEET is a volatile repellent (1uL 50% DEET in two spots, just as a volatile odorant). Next, I tested if DEET can inhibit attraction to isoamyl alochol if presented as a point stimulus (2uL 50% DEET or solvent was added between each of the two spots of isoamyl alochol (IAA).) Both are plotted below:

 $DD = 2 \times 1$ uL DEET as point stimulus $EE = 2 \times 1$ uL solvent (ethanol) as point stimulus $IDI = 2 \times 1$ uL spots IAA as point stimulus with 2uL 50% DEET between $IEI = 2 \times 1$ uL spots IAA as point stimulus with 2uL solvent/ethanol between ## Fig 1 b-c

```
# make summary data
spotssum <- ddply(spots,c('stimulus'),summarise,N=length(stimulus),mean=mean(CI),sd=sd(CI),se=sd/sqrt(N

# plot each point, xs are means, +s are standard error
ggplot(spots,aes(x=stimulus,y=CI)) +
    ggtitle('Testing volatile repellency of DEET') +
    geom_jitter(width=0.2,height=0) +
    geom_point(data=spotssum,aes(x=stimulus,y=mep),shape=3) +
    geom_point(data=spotssum,aes(x=stimulus,y=mem),shape=3) +
    geom_point(data=spotssum,aes(x=stimulus,y=mean),shape=4) +
    theme_tufte() +
    ylim(-1,1) +
    geom_hline(yintercept=0)</pre>
```

Testing volatile repellency of DEET



DEET does not look like a volatile point stimulus: p = 0.6478

DEET does not interrupt chemotaxis to isoamyl alochol when presented as a point stimulus: p=0.5286

```
IDI <- subset(spots,stimulus=="IDI")
ISI <- subset(spots,stimulus=="ISI")</pre>
```

```
t.test(x=ISI$CI,y=IDI$CI,"two.sided")

##

## Welch Two Sample t-test

##

## data: ISI$CI and IDI$CI

## t = -0.64263, df = 17.967, p-value = 0.5286

## alternative hypothesis: true difference in means is not equal to 0

## 95 percent confidence interval:

## -0.14927950 0.07935372

## sample estimates:

## mean of x mean of y

## 0.7166651 0.7516280
```

Dose-response-like curve for effect of DEET in agarose on IAA chemotaxis

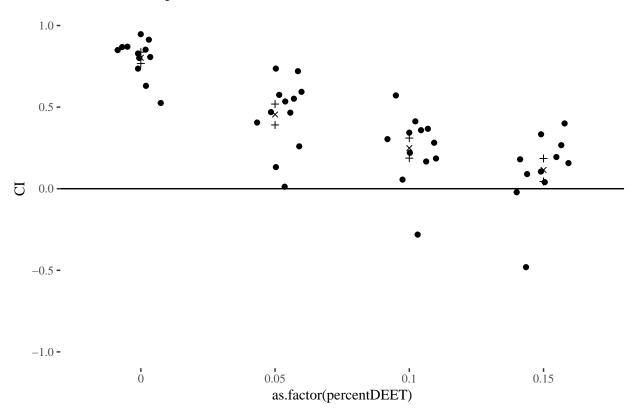
Fig 1e

```
drc <- subset(allbeh,expt=='drc')
drc <- subset(drc,percentDEET<0.2)
drcN2 <- subset(drc,genotype=='N2')

drcN2summary <- ddply(drcN2,c("percentDEET"),summarise,N=length(percentDEET),mean=mean(CI),sd=sd(CI),se
drcN2summary$mep <- drcN2summary$mean + drcN2summary$se
drcN2summary$mem <- drcN2summary$mean - drcN2summary$se

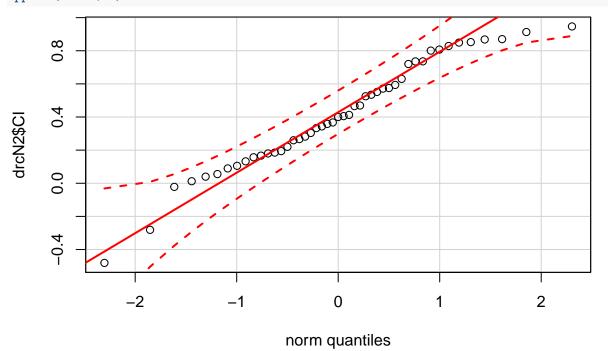
ggplot(drcN2,aes(x=as.factor(percentDEET),y=CI)) +
    ggtitle('DEET dose response') +
    geom_jitter(width=0.2,height=0) +
    theme_tufte() +
    ylim(-1,1) +
    geom_hline(yintercept=0) +
    geom_point(data=drcN2summary,aes(x=as.factor(percentDEET),y=mep),shape=3)+ geom_point(data=drcN2summary,aes(x=as.factor(percentDEET),y=mep),shape=3)</pre>
```

DEET dose response



Below find the stats for these dose response data

#plot QQ plot for model used in 2 way anova, if looks like a ~ straight line the residuals are probably qqPlot(drcN2\$CI)



```
tukeydrc <- TukeyHSD(aov(formula = CI ~ as.factor(percentDEET), data = drcN2))

Tukeydrc.levels <- tukeydrc$`as.factor(percentDEET)`[,4]
multcompLetters(Tukeydrc.levels)['Letters']

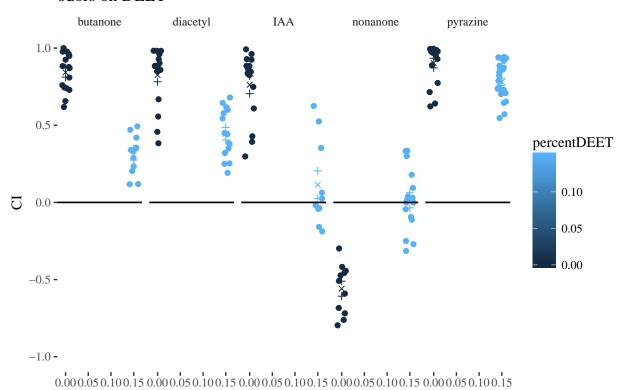
## $Letters
## 0.05 0.1 0.15 0
## "a" "ab" "b" "c"</pre>
```

What other odors are affected by DEET

Fig 1f (not in same order)

```
odor <- subset(allbeh,expt=='odor')</pre>
odorsum <- ddply(odor,c("stimulus", "percentDEET"), summarise, N=length(stimulus), mean=mean(CI), sd=sd(CI),
odorsum$mep <- odorsum$mean + odorsum$se
odorsum$mem <- odorsum$mean - odorsum$se
ggplot(odor,aes(x=percentDEET,y=CI,color=percentDEET)) +
  ggtitle('odors on DEET') +
  geom_jitter(width=0.01,height=0) +
 theme_tufte() +
 ylim(-1,1) +
  geom_hline(yintercept=0) +
  geom_point(data=odorsum,aes(x=percentDEET,y=mep),shape=3) +
  geom_point(data=odorsum,aes(x=percentDEET,y=mem),shape=3) +
  geom_point(data=odorsum,aes(x=percentDEET,y=mean),shape=4) +
  theme_tufte() +
  geom_hline(yintercept=0) +
  facet_grid(~stimulus)
```

odors on DEET



percentDEET

statistics for these data:

#plot QQ plot for model used in 2 way anova, if looks like a ~ straight line the residuals are probably
odors.aov = aov(CI~as.factor(percentDEET)*stimulus,data=odor)
plot(odors.aov,which=2)

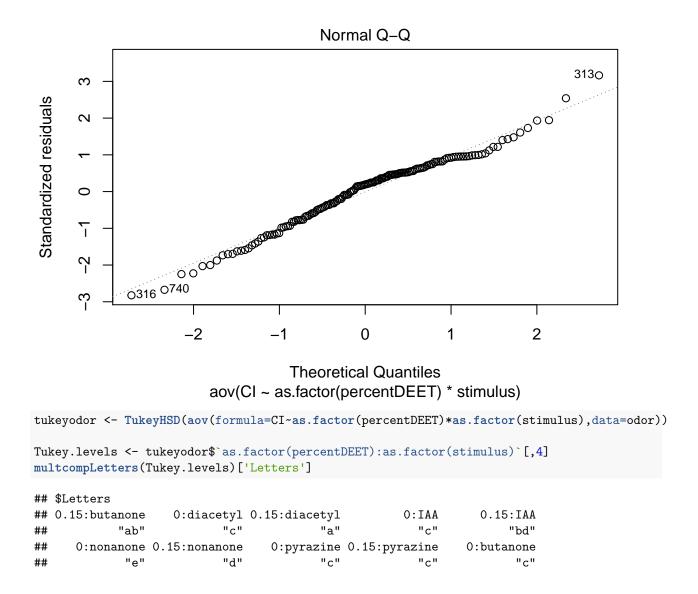


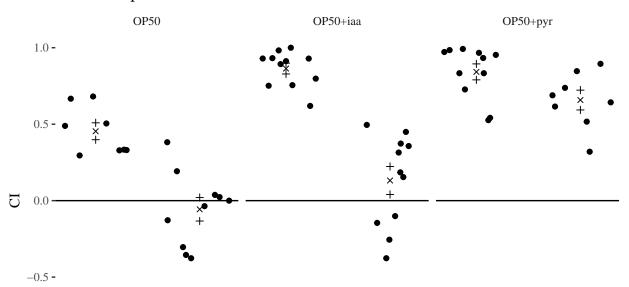
Fig 1g plotting data for animals chemotaxing to 20uL of bacteria in LB +/-pyrazine or isoamyl alcohol vs LB alone.

```
# make summary data for bacteria
bacteriasummary <-ddply(bacteria,c("stimulus", "percentDEET"),summarise,N=length(CI),mean=mean(CI),sd=s.
bacteriasummary$mep <- bacteriasummary$mean + bacteriasummary$se
bacteriasummary$mem <- bacteriasummary$mean - bacteriasummary$se

ggplot(bacteria,aes(x=percentDEET,y=CI)) +
    getitle('bacteria experiment') +
    geom_point(data=bacteriasummary,aes(x=percentDEET,y=mep),shape=3) +
    geom_point(data=bacteriasummary,aes(x=percentDEET,y=mem),shape=3) +
    geom_point(data=bacteriasummary,aes(x=percentDEET,y=mem),shape=4) +
    geom_jitter(width=0.05,height=0) +
    theme_tufte() +
    ylim(-1,1) +
    geom_hline(yintercept=0) +</pre>
```

facet_grid(~stimulus)

bacteria experiment



-1.0 --0.05 0.00 0.05 0.10 0.15 0.200.05 0.00 0.05 0.10 0.15 0.200.05 0.00 0.05 0.10 0.15 0.200.05 0.00 0.05 0.10 0.15 0.20 percentDEET

bacteria.aov = aov(CI~as.factor(percentDEET)*stimulus,data=bacteria)
plot(bacteria.aov,which=2)

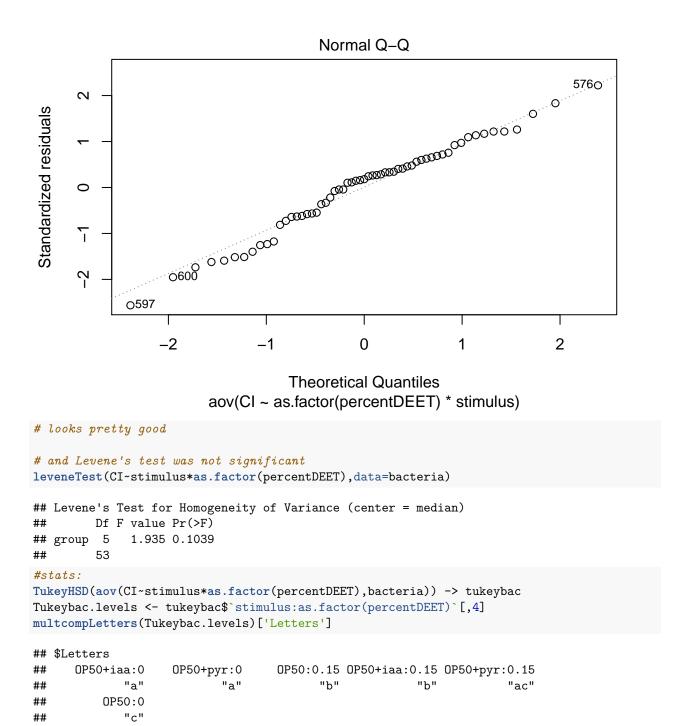
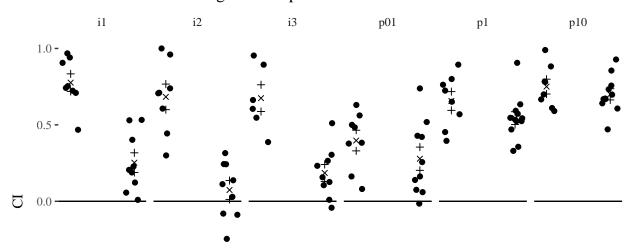


Fig 1h-i

```
# make summary data for bacteria
odordrcsummary <-ddply(odordrc,c("stimulus","percentDEET"),summarise,N=length(CI),mean=mean(CI),sd=sd(C
odordrcsummary$mep <- odordrcsummary$mean + odordrcsummary$se
odordrcsummary$mem <- odordrcsummary$mean - odordrcsummary$se
ggplot(odordrc,aes(x=percentDEET,y=CI)) +</pre>
```

```
ggtitle('odor over orders of magnitude experiment') +
geom_point(data=odordrcsummary,aes(x=percentDEET,y=mep),shape=3) +
geom_point(data=odordrcsummary,aes(x=percentDEET,y=mem),shape=3) +
geom_point(data=odordrcsummary,aes(x=percentDEET,y=mean),shape=4) +
geom_jitter(width=0.02,height=0) +
theme_tufte() +
ylim(-1,1) +
geom_hline(yintercept=0) +
facet_grid(~stimulus)
```

odor over orders of magnitude experiment



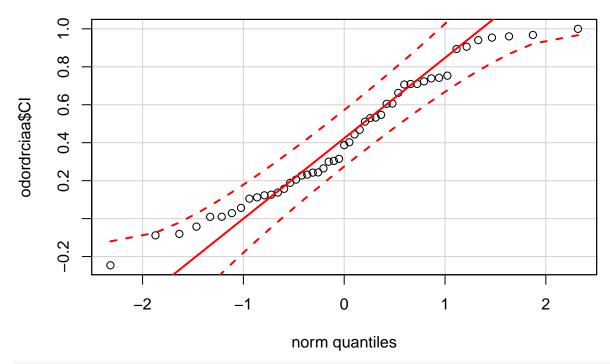
-0.5 -

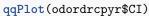
−1.0 **-**

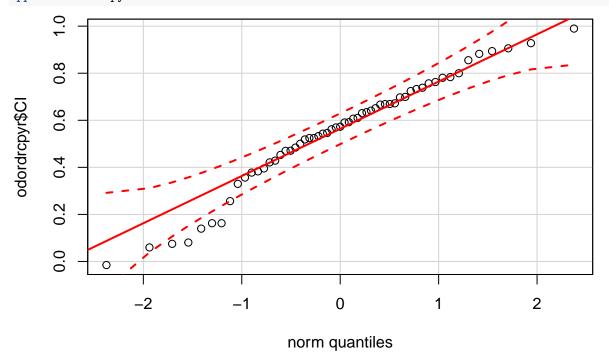
```
odordrciaa <- subset(odordrc,stimulus!="p01")
odordrciaa <- subset(odordrciaa,stimulus!="p1")
odordrciaa <- subset(odordrciaa,stimulus!="p10")

odordrcpyr <- subset(odordrc,stimulus!="i1")
odordrcpyr <- subset(odordrcpyr,stimulus!="i2")
odordrcpyr <- subset(odordrcpyr,stimulus!="i3")

#qqPlots
qqPlot(odordrciaa$CI)</pre>
```







```
#levene's tests
leveneTest(CI~stimulus*as.factor(percentDEET),data=odordrciaa)
```

```
## Levene's Test for Homogeneity of Variance (center = median)
## Df F value Pr(>F)
## group 5 0.2278 0.9484
## 43
```

```
leveneTest(CI~stimulus*as.factor(percentDEET),data=odordrcpyr)
## Levene's Test for Homogeneity of Variance (center = median)
        Df F value Pr(>F)
## group 5 1.5338 0.1959
##
         51
#stats
tukeyiaadrc <- TukeyHSD(aov(CI~stimulus*as.factor(percentDEET),odordrciaa))</pre>
Tukeyiaadrc.levels <- tukeyiaadrc$`stimulus:as.factor(percentDEET)`[,4]</pre>
multcompLetters(Tukeyiaadrc.levels)['Letters']
## $Letters
##
      i2:0
              i3:0 i1:0.15 i2:0.15 i3:0.15
                                               i1:0
                                                "a"
                               "b"
                                        "b"
##
       "a"
               "a"
                       "b"
tukeypyrdrc <- TukeyHSD(aov(CI~stimulus*as.factor(percentDEET),odordrcpyr))</pre>
Tukeypyrdrc.levels <- tukeypyrdrc$`stimulus:as.factor(percentDEET)`[,4]</pre>
multcompLetters(Tukeypyrdrc.levels)['Letters']
## $Letters
##
       p1:0
               p10:0 p01:0.15 p1:0.15 p10:0.15
                                                    p01:0
                                                     "bc"
        "a"
                 "a"
                        "b" "ac" "a"
##
```

Forward genetic screen

I completed a forward genetic screen with help from Wendy Wang in Shai Shaham's lab (and lots of guidance from Shai on both the screen and follow up) plot of genetic screen data:

Figure 2b

```
ed03 == LBV003 in paper ed03 was our in-house name as it was the third mutant I (__E__mily __D__ennis) found ggplot(screen,aes(x=genotype,y=CI)) + ggtitle('screen data') + geom_boxplot(outlier.shape=NA,coef=0) + ggtitle('screen data')
```

screen data 1.0 -0.5 -0.0 --0.5 **-−1.0 -**N2 ed03 genotype

EMS-isolated strain LBV003 is DEET-resistant compared to N2 (p-value = 0.003101)

note that in the spreadsheet, all screen and rescue experiments

```
N2screen <- subset(screen,genotype=="N2")
ed03screen <- subset(screen,genotype=="ed03")
t.test(x=N2screen$CI,y=ed03screen$CI,"two.sided")
##
##
   Welch Two Sample t-test
## data: N2screen$CI and ed03screen$CI
## t = -4.1191, df = 8.2912, p-value = 0.003101
\#\# alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## -0.5890473 -0.1678773
## sample estimates:
## mean of x mean of y
## 0.03958319 0.41804553
```

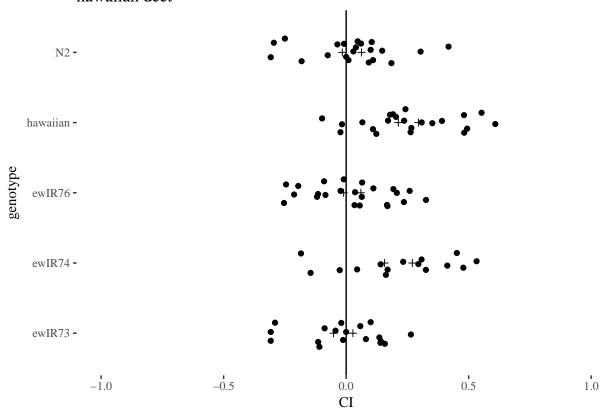
Figure 2e: a wild-isolate is naturally DEET-resistant

```
# make summary data (means, standard errors) to plot
hdsummary <-ddply(hawaiiand,c("genotype"),summarise,N=length(genotype),mean=mean(CI),sd=sd(CI),se=sd/sq
hdsummary$mep <- hdsummary$mean + hdsummary$se
```

```
hdsummary$mem <- hdsummary$mean - hdsummary$se

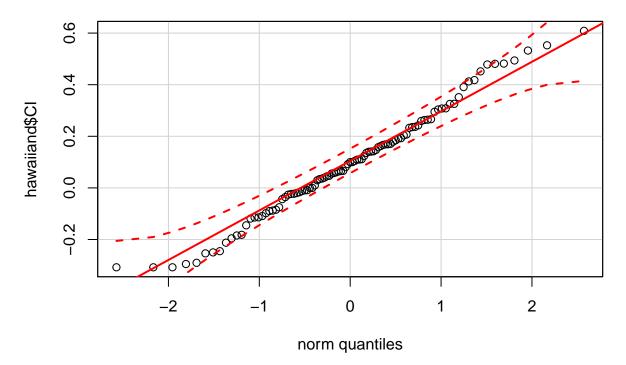
#plot hawaiian strains chemotaxing to IAA on DEET agar
ggplot(hawaiiand,aes(x=genotype,y=CI)) +
    ggtitle('hawaiian deet') +
    geom_point(data=hdsummary,aes(x=genotype,y=mep),shape=3) +
    geom_point(data=hdsummary,aes(x=genotype,y=mem),shape=3) +
    geom_jitter(width=0.2,height=0) + theme_tufte() + ylim(-1,1) +
    geom_hline(yintercept=0) +
    coord_flip()</pre>
```

hawaiian deet



First I did a qqPlot for these data to test if residuals are roughly normal and an ANOVA is appropriate. Dashed lines on the plot indicate 95% confidence interval.

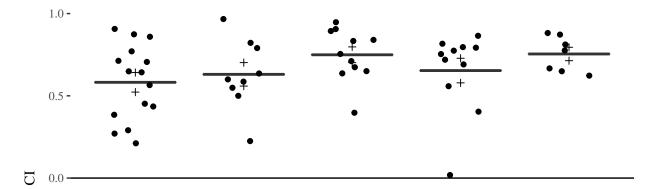
```
qqPlot(hawaiiand$CI)
```



I then ran an ANOVA on these data

```
tukeyhawaiian <- TukeyHSD(aov(formula = CI ~ genotype, data = hawaiiand))</pre>
hawaiian.levels <- tukeyhawaiian$genotype[,4]
multcompLetters(hawaiian.levels)['Letters']
##
  $Letters
##
     ewIR74
              ewIR76 hawaiian
                                     N2
                                          ewIR73
        "a"
                 "b"
                           "a"
                                    "b"
                                             "b"
##
# make summary data for hawaiiane
hawaiiane <- subset(hawaiiane,genotype!='ewIR69')</pre>
hesummary <-ddply(hawaiiane,c("genotype"),summarise,N=length(genotype),mean=mean(CI),sd=sd(CI),se=sd/sq
hesummary$mep <- hesummary$mean + hesummary$se
hesummary$mem <- hesummary$mean - hesummary$se
ggplot(hawaiiane,aes(x=genotype,y=CI)) +
  ggtitle('hawaiian etoh') +
  geom_boxplot(data=hesummary,aes(x=genotype,y=mean)) +
  geom_point(data=hesummary,aes(x=genotype,y=mep),shape=3)
  geom_point(data=hesummary,aes(x=genotype,y=mem),shape=3)
  geom_jitter(width=0.2,height=0) +
  theme_tufte() +
  ylim(-1,1) +
  geom_hline(yintercept=0)
```

hawaiian etoh

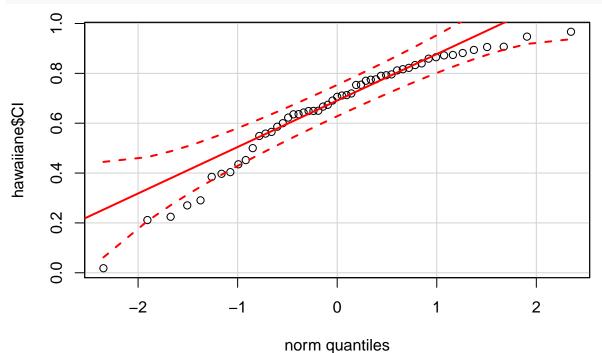


-0.5 -

ewIR73 ewIR74 ewIR76 hawaiian N2
genotype

Below find the stats for these hawaiian ethanol plate data

qqPlot(hawaiiane\$CI)



tukeyhawaiiane <- TukeyHSD(aov(formula=CI~genotype,data=hawaiiane))
Tukeyhe.levels <- tukeyhawaiiane\$`genotype`[,4]</pre>

```
multcompLetters(Tukeyhe.levels)['Letters']

## $Letters
## ewIR74 ewIR76 hawaiian N2 ewIR73
## "a" "a" "a" "a" "a" "a"
```

Fig 2g-h and 3h

Animals expressing either str-217 rescue construct (str-217p::str-217::SL2::GFP) or ADL rescue construct (srh-220p::str-217::SL2::mCherry) return to being DEET sensitive, indicating str-217 is required in each of these strains to confer DEET-sensitivity

```
# make summary data for rescue
rescue2 <- subset(allbeh,expt=='rescue2')
rescue <- subset(allbeh,expt=='rescue')</pre>
rescueall <- rbind(rescue2,rescue)</pre>
rescueD <- subset(rescueall,percentDEET==0.15)</pre>
rescueDsummary <-ddply(rescueD,c("genotype"),summarise,N=length(genotype),mean=mean(CI),sd=sd(CI),se=sd
rescueDsummary$mean + rescueDsummary$se
rescueDsummary$mem <- rescueDsummary$mean - rescueDsummary$se
ggplot(rescueD,aes(x=genotype,y=CI)) +
  ggtitle('rescue') +
  geom_boxplot(data=rescueDsummary,aes(x=genotype,y=mean)) +
  geom_point(data=rescueDsummary,aes(x=genotype,y=mep),shape=3) +
  geom_point(data=rescueDsummary,aes(x=genotype,y=mem),shape=3) +
  geom_jitter(width=0.1,height=0) +
  theme_tufte() +
  ylim(-1,1) +
  geom_hline(yintercept=0)
```

1.0 -

```
0.5 -
0.0
  -0.5 -
  −1.0 -
                                                      ed03-r
                                                                    ewIR74
                                                                                 ewIR74r
                                         ed03
             crispr
                          crispr-r
                                              genotype
crisprnoR <- subset(rescueD,genotype=='crispr')</pre>
crisprR <- subset(rescueD,genotype=='crispr-r')</pre>
ed03noR <- subset(rescueD,genotype=='ed03')
ed03R <- subset(rescueD,genotype=='ed03-r')</pre>
ewir74noR <- subset(rescueD,genotype=='ewIR74')</pre>
ewir74R <- subset(rescueD,genotype=='ewIR74r')</pre>
t.test(crisprnoR$CI,crisprR$CI)
##
  Welch Two Sample t-test
##
##
## data: crisprnoR$CI and crisprR$CI
## t = 3.4747, df = 12.49, p-value = 0.004341
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## 0.1477553 0.6388575
## sample estimates:
## mean of x mean of y
## 0.2857813 -0.1075251
t.test(ed03noR$CI,ed03R$CI)
##
   Welch Two Sample t-test
##
## data: ed03noR$CI and ed03R$CI
```

t = 2.6252, df = 7.4678, p-value = 0.03226

```
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## 0.02040963 0.34879833
## sample estimates:
## mean of x mean of y
## 0.28164730 0.09704332
t.test(ewir74noR$CI,ewir74R$CI)
##
##
   Welch Two Sample t-test
## data: ewir74noR$CI and ewir74R$CI
## t = 4.3087, df = 9.6589, p-value = 0.001669
\#\# alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## 0.1607155 0.5083839
## sample estimates:
     mean of x
                   mean of y
## 0.3355008562 0.0009511747
```

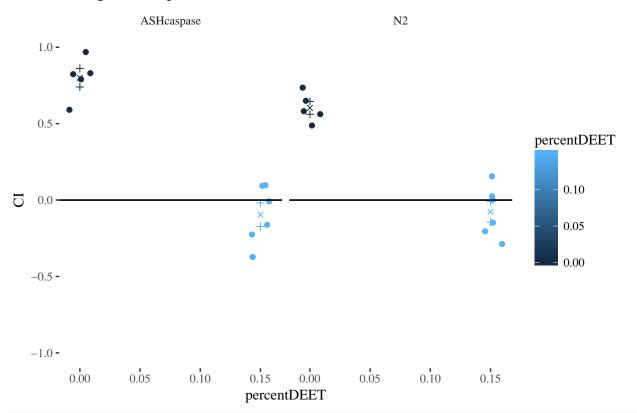
Fig 3d neurons are not required for complete DEET-sensitivity

```
ASHsum <- ddply(ASH,c("genotype","percentDEET"),summarise,N=length(genotype),mean=mean(CI),sd=sd(CI),se
ASHsum$mep <- ASHsum$mean + ASHsum$se

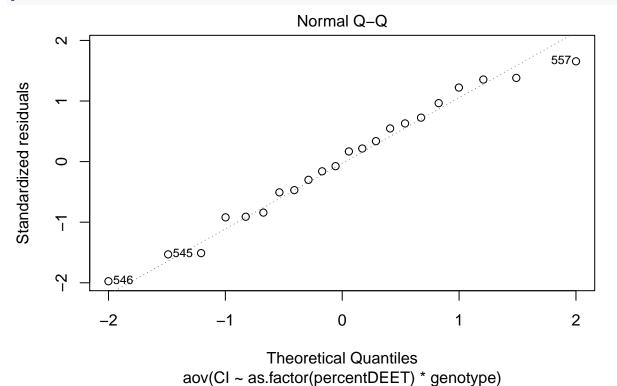
ASHsum$mem <- ASHsum$mean - ASHsum$se

ggplot(ASH,aes(x=percentDEET,y=CI,color=percentDEET)) +
    ggtitle('testing ASH requirement') +
    geom_jitter(width=0.01,height=0) +
    theme_tufte() +
    ylim(-1,1) +
    geom_hline(yintercept=0) +
    geom_point(data=ASHsum,aes(x=percentDEET,y=mep),shape=3) +
    geom_point(data=ASHsum,aes(x=percentDEET,y=mem),shape=3) +
    geom_point(data=ASHsum,aes(x=percentDEET,y=mean),shape=4) +
    theme_tufte() +
    geom_hline(yintercept=0) +
    facet_grid(-genotype)
```

testing ASH requirement



ASH.aov = aov(CI~as.factor(percentDEET)*genotype,data=ASH)
plot(ASH.aov,which=2)

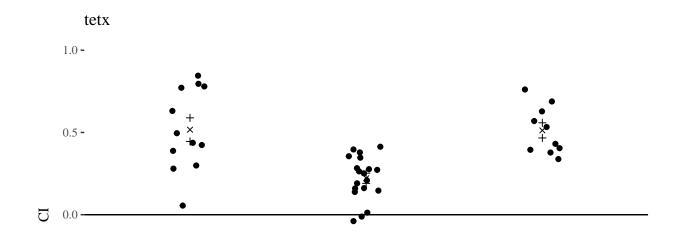


```
leveneTest(CI~genotype*as.factor(percentDEET),data=ASH)
## Levene's Test for Homogeneity of Variance (center = median)
        Df F value Pr(>F)
## group 3 1.6038 0.2235
##
         18
tukeyASH <- TukeyHSD(aov(formula=CI~as.factor(percentDEET)*as.factor(genotype),data=ASH))</pre>
TukeyASH.levels <- tukeyASH$`as.factor(percentDEET):as.factor(genotype)`[,4]</pre>
multcompLetters(TukeyASH.levels)['Letters']
## $Letters
## 0.15:ASHcaspase
                                            0.15:N2
                               0:N2
                                                       0:ASHcaspase
               "a"
                                "b"
                                                "a"
```

ADL chemical synapses are required for complete DEET-sensitivity

Figure 3i

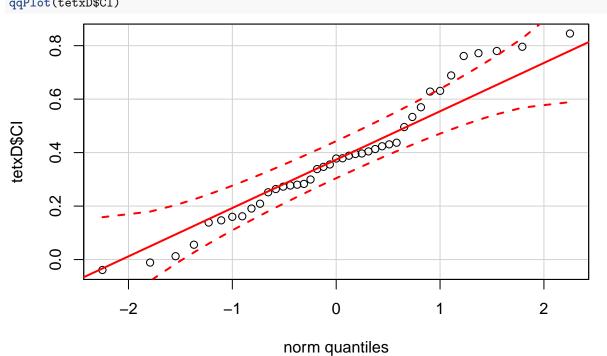
```
# make summary data for tetx
tetx <- subset(allbeh,expt=='tetx')</pre>
tetx <- subset(tetx,genotype!='ed03')</pre>
#removes one data point where I had an ed03 plate to check stocks after thawing, done at the same time
tetxD <- subset(tetx,percentDEET==0.15)</pre>
tetxDsummary <-ddply(tetxD,c("genotype"),summarise,N=length(genotype),mean=mean(CI),sd=sd(CI),se=sd/sqr
tetxDsummary$mep <- tetxDsummary$mean + tetxDsummary$se</pre>
tetxDsummary$mem <- tetxDsummary$mean - tetxDsummary$se</pre>
ggplot(tetxD,aes(x=genotype,y=CI)) +
  ggtitle('tetx') +
  geom_point(data=tetxDsummary,aes(x=genotype,y=mep),shape=3) +
  geom_point(data=tetxDsummary,aes(x=genotype,y=mem),shape=3) +
  geom_point(data=tetxDsummary,aes(x=genotype,y=mean),shape=4) +
  geom_jitter(width=0.1,height=0) +
  theme_tufte() +
  ylim(-1,1) +
  geom_hline(yintercept=0)
```



-0.5 **-**

−1.0 tetx crispr N2 genotype

qqPlot(tetxD\$CI)



TukeyHSD(aov(CI~genotype,tetxD))

Tukey multiple comparisons of means 95% family-wise confidence level