Post-release survival of spot prawns (*Pandalus platyceros*) declines with increasing air exposure and temperature­

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*Running headline: Spot prawn survival declines with air exposure and high temperature*

**ABSTRACT**

*Prawns like to be cold*

*Release them lickety split*

*Back to life at depth*

**KEY WORDS**

fisheries management, spot prawn, post-release survival, population dynamics, marine invertebrates

**INTRODUCTION**

Marine invertebrate fisheries are growing faster than any other group of fisheries in the world, with catches increasing by six-fold since 1950 and double the taxa reported (Anderson et al. 2011). Within that growth, decapod crustacean fisheries are growing faster than any other major group of marine invertebrates (Boenish et al. 2022). Despite the pace of expansion, invertebrate fisheries receive relatively less scientific and stock assessment attention (CITE). This attention is pivotal to stewarding marine invertebrate populations because they play important ecosystem roles (e.g., Eddy et al. 2017, CITE, CITE) and because this trend may be an example of ‘fishing down the food web’ (Pauly et al. 1998; Pinsky et al. 2011) which is accompanied by risks to ecosystems and the human communities that depend on them (CITE). The global pattern of expanding invertebrate fisheries is mirrored on the Pacific coast of Canada where declining finfish fisheries have led to a redistribution of effort towards invertebrate fisheries (Perry et al. 1999) including the spot prawn fishery in British Columbia (BC).

Spot prawns (*Pandalus platyceros*) are sequential protandrous hermaphrodites, beginning their lives as males before transitioning to and reproducing as females. In BC, they are thought to live for four years (Butler 1964). In the spring, brooding females release hatched eggs which spend 2-3 months in a larval dispersal stage before settling and developing as juveniles in shallow waters (Marliave and Roth 1995). Spot prawns spend ~2 years as males before transitioning to females. They breed in the late summer through early fall and females brood eggs through the winter before releasing them the following spring. Timed to begin after most brooding females have released their eggs, the commercial spot prawn fishery is the largest shrimp fishery in BC.

With annual landed values of $33.5-39 million (DFO 2019), the commercial fishery supports over 250 licenses and a growing recreational fishery. While most shrimp fisheries are conducted by trawl with accompanying concerns for their negative ecosystem impacts (Andrew & Pepperell, 1992), the prawn-by-trap fishery represents a rare example of a relatively low-impact shrimp fishery (Boutillier and Bond 2000). The commercial fishery is managed as a derby style fishery, where in-season closures are implemented based on the average number of females-per-trap (the ‘Spawner Index’, Boutillier and Bond 2000) to ensure sufficient spawning females for the subsequent year. An additional management measure was introduced in 1985, implementing a minimum size limit of 30 mm carapace length. Since then, the size limit has increased to 33 mm and commercial traps must comply to a minimum mesh size. These size-based measures are designed to protect the small males in the population who will fertilise females and go on to transition and reproduce as females in subsequent years. Furthermore, the year-round recreational fishery mandates that egged females must not be retained. Despite numerous release-based management measures, the survival of released prawns is not well understood.

[add a paragraph on the body of research on post-release survival in fisheries]

To address the uncertainty in post-release survival of spot prawns caught in trap fisheries, we conducted an in-situ field experiment that evaluated the influence of air exposure, air temperature, and carapace length on the probability of survival after release. [flesh out transition paragraph]

**MATERIALS AND METHODS**

We conducted 23 experimental trials between May 22, 2022 and June 28, 2022 in the Broughton Archipelago, British Columbia, Canada (Fig. X). A given experimental trial consisted of three components conducted over three days: setting traps for field collection, collecting prawns and conducting the experimental treatment, and processing prawns at the end of the experiment to assess survival and condition. Each trial took 3-4 days to complete (the string of traps to initially collect prawns soaked for 24-48 hours).

EXPERIMENT SET-UP

To collect prawns for a given trial, we set a string of 10 prawn traps (insert technical specs) baited with pellets (insert technical specs) within a target depth range (55-110 meters) that aligns with the approximate depth range targeted by commercial and recreational fisheries. Depending on weather conditions and logistics, the string of traps soaked for 24-48 hours before we hauled the traps and began the experimental trial. On the day the trial began, at the trap setting site, we collected air temperature, water temperature (at 0 m and 10 m depths), and water salinity (at 0 m and 10 m depths) using a YSI (insert technical details). During some of the trials, the YSI was broken, and we collected temperature and salinity data using a thermometer and refractometer respectively. We hauled the string of traps using a hydraulic pot hauler (insert technical details). During trap hauling, as each trap came on the boat, we removed any bycatch and emptied the remaining prawns into a small square white bin (10 L, insert dimensions) with drilled holes that allowed water to flow through. We placed each white bin in a large fish tote (insert technical specs) filled with seawater (XX L). This method ensured that until the trial began, prawns experienced minimal air exposure (10-15 seconds as trap was emptied into white bin). After we finished hauling all traps, we assessed how many prawns to assign to each treatment (minimum 35 per treatment, maximum 70 per treatment to minimise density-dependent effects). We haphazardly assigned prawns to one of four or five treatments: ‘immediate release’ or air exposures of 30, 60, 90, or 120 minutes. In circumstances where numbers of prawns was a limiting factor, we did not include the 120 minute treatment. To implement the ‘immediate release’ treatment in an experimentally tractable manner, we hung prawns off the boat in a mesh drawstring bag (insert mesh specs) approximately 20 m below the surface of the water. This mimicked quick release while still allowing for the experimental ‘release’ of prawns from all treatments together in one string of traps. To minimise air exposure, we emptied the prawns of one of the white bins into a solid white bin filled with seawater. We counted out the appropriate number of individuals, using forceps to place a coloured orthodontic elastic band (insert technical specs) on the base of the rostrum (Figure X), and placing each prawn in the mesh bag which was submerged in a 20 L bucket of seawater. Once all prawns had been banded for the ‘immediate release’ treatment, we cinched the mesh bag and attached it to a weighted line hanging off the boat to a depth of 20 m. To begin the air exposure treatments, we removed the remaining white bins from the fish tote at the same time such that all prawns hit the air together. We started a timer for the first treatment (30 minutes) and began to distribute the appropriate number of prawns to each bin, distributing haphazardly by size. For trials with fewer prawns, we allotted one bin for treatment (e.g., ~35 prawns per bin) and for trials with more prawns, we allotted two bins per treatment (e.g., two bins, each with ~35 prawns). For the duration of the treatment time, we kept the bins under the canopy of the boat such that they received no direct sun exposure or direct precipitation. The spatial arrangement of the bins was haphazard with respect to treatment. Choosing different colours for different treatments (the exact colour varied trial to trial), we applied coloured bands to the rostrum of each individual prawn. As the timer for a given treatment went off, we emptied the prawns of a certain band colour into a weighted mesh bag and clipped it to the hanging line such that it descended to hang with the other treatment bags at ~20 m. At the end of the final treatment (90 minutes or 120 minutes), we placed the final group of prawns in a mesh bag hung off the side of the boat such that all treatments experienced the process of being lowered and raised in a mesh bag. Finally, we raised all the bags at the same time and distributed the prawns from all treatments across six baited prawn traps with the tunnels tied shut such that prawns could not escape. To avoid confounding treatment effect with trap effect, we distributed some prawns from each treatment to each trap such that traps contained a mix of all treatments (the coloured bands facilitated mixing prawns without losing information on treatment). Once prawns had been distributed, we closed the traps and reset the string of six traps in the same location and depth they had been hauled from initially. For each trial, we timed the length of time it took from the trial ending (mesh bags coming out of water) to the final trap hitting the water during re-setting.

EXPERIMENT WRAP-UP

Approximately 24 hours after we conducted the experimental treatment, we returned to the re-set string of traps to end the trial. We followed the same procedure as at the beginning of the experiment, collecting temperature and salinity measurements before re-hauling the string and placing the contents of each trap into a white square bin which we kept in the seawater-filled fish tote.

After re-hauling the string, we emptied one square bin (i.e., one trap’s worth of prawns) at a time into a sampling tray and collected the end-of-trial data. For each individual prawn, we recorded their band colour, stage (juvenile, male, transitional, female, egged female, or spent female), and their carapace length as well as whether they were alive, dead, or scavenged. We considered a prawn dead if their gill filaments were not moving at all (i.e., the individual was no longer breathing). A ‘scavenged’ prawn referred to an individual that was dead and missing some body parts. We returned dead and scavenged prawns to the ocean. As they were counted and measured, alive prawns were transferred from the sampling tray to a mesh bag submerged in a 20 L bucket of seawater. After processing a single trap, the mesh bag of live prawns was hung off the boat at 20 m to maintain the prawns as close to their initial condition as possible.

ASSESSING REFLEX BEHAVIOURS

After collecting survival data for all traps, we assayed each live prawn for a suite of ten reflex behaviours, based on the approach outlined in (Stoner 2012) that developed a set of ten reflex behaviours in spot prawns that cumulatively predicted long term survival in the lab. Processing one trap’s worth of prawns at a time, we assessed each prawn for how many of the reflex behaviours they displayed, resulting in a cumulative score that could range from 0-10. Here, a score of zero indicates a prawn that is alive but displays no other behaviours (poor condition) and a score of ten indicates a prawn that is alive and displays all assessed reflex behaviours (good condition). After we finish assessing the reflexes of the live prawns, we remove their nose band and return them to the ocean.

STATISTICAL ANALYSIS

To evaluate the influence of air exposure, air temperature, and carapace length on the post-release survival of spot prawnscaptured in trap fisheries, we used generalized linear mixed-effects models (GLMMs) that accommodate the hierarchical structure of the experiment and the non-linear distribution of the data. Our response data consist of 0 (dead or scavenged) and 1 (alive) so we used a binomial error structure to model the probability of survival. We included a random effect on the intercept for unique trap-within-trial to account for the shared variation within a trap of a given trial (where the number of random effect levels corresponds trials multiplied by traps).

In some cases (488 of 5053), we excluded data from prawns for which either treatment group or carapace length was unknown. A small portion of the prawns (273) lost their coloured band during the release stage of the experiment (Table 1). As the band colour denoted treatment group, prawns that lost their band could not be assigned to a treatment. We considered the possibility that small prawns may have been more likely to lose their band. To ensure we would not confound our results, we compared the size distribution of these prawns to that of the prawns that retained their band (Appendix 1). There was a statistically significant difference between the two groups (T=3.25, *p*=0.0013), however the difference was very small (1 mm, 3%). We therefore excluded these individuals from the final dataset. We excluded an additional 215 prawns that had damage on their carapace such that we could not measure length accurately.

We considered how the loss of prawns during the experiment may have influenced our results. We lost prawns in two ways: through the mesh of the bags used during the treatment stage or through the mesh of the traps during the release stage of the trial. We could not determine whether these individuals survived the treatment or not. To investigate whether there was a bias in prawn loss (i.e. if either dead or living prawns were more likely to be lost), we evaluated the percentage of prawns lost in each treatment. We found that, on average, more prawns were lost from treatment groups with longer air exposure (Figure X). To evaluate the influence of the potential bias in prawn loss, we simulated four scenarios for prawn loss: no prawns lost, only dead prawns lost, only surviving prawns lost, dead and surviving prawns lost at equal frequencies. We evaluated the difference in survival estimates among the four scenarios to address whether loss of prawns could confound our interpretation of how survival did or did not differ across treatment groups. We found that for a typical percentage of prawns lost (20%) (Figure X), the effect on the estimated percentage of prawns that survived was minor (maximum 6% for most trials) (Figure X) even if we lost living or dead prawns more frequently.

We took a model selection approach to evaluate the relative importance of three fixed effects and their two-way interactions: time out of water, air temperature, and carapace length. In total, we considered a suite of 18 candidate models estimating the probability of prawn survival ~24 hours after release (Table 2). All models included a normally distributed random effect on the intercept to account for variation in the probability of survival across trials and traps. The random effect includes 123 levels, accounting for 21 trials and 6 traps in most trials (Table 1). We expected survival may vary by trial and trap because location, time, and orientation on the ground varied between trial and trap. We conducted all analyses in R (R core team 2023). For completeness, we fit the models in two ways: Gaussian Quadrature (10 points) with lme4 (Bates et al. 2015), and Laplace approximation with glmmTMB (Brooks et al. 2017). To prioritise simplicity and interpretability, we compared models using Bayesian Information Criterion (BIC) (Table 2).

**RESULTS**

The 23 experimental trials included 5,052 prawns encompassing juvenile through female life stages. Due to the timing of the experimental period, we did not have access to egged or spent females to include in the experiment. The majority of the prawns were male or transitional stage and prawn carapace length ranged from 18.0 mm to 52.36 mm (Figure X). Air temperature varied throughout the experimental season with trials conducted in as cool a climate as 10.7oC and as warm as 25oC. We tried to maintain relatively constant high salinity conditions which required pumping water from below the freshwater layer during the freshet. The seawater that we kept prawns in during the experiment ranged from 24.5 ppt to 31.4 ppt, with the exception of two trials which we did not include in the final analysis (trial 11, 21.5 ppt during end-of-trial processing and trial 12, 14.0 ppt during end-of-trial processing).

The post-release survival of spot prawns declined with increasing length of air exposure (Figure X) and with increasing air temperature (Figure X). Of the models we fit to determine the best predictors of prawn survival probability, the model including two interaction effects – an interaction between treatment and temperature and between temperature and length – was best supported by BIC (Table 2). There was no definitively clear top model, with reasonable support for five models which all fell within 10 ∆BIC of the top model (CITE). The treatment-temperature interaction effect was common across all five top models. On average, the log-odds of survival declined by 0.096 per additional 10 minutes out of water (95% C.I. = -0.105, -0.086) and it declined by 0.318 per degree Celsius warmer (95% C.I. = -0.470, -0.167). Larger prawns had slightly lower probability of survival (insert prediction) than smaller prawns (insert prediction). Air exposure had a stronger effect on warm days with an additional decrease in log-odds of survival by 0.00189 (95% C.I. = -0.00253, -0.00125). Model-averaged predictions of survival probability were very similar to the predictions from the top model so for ease of interpretability we discuss the estimates from the top model in the main text (see Supplementary Material for model-averaged predictions).

The mean reflex score for surviving prawns, on a scale from 0-10, was 8.96 (Figure SX). Based on the reflex-mortality relationships defined in Stoner (2012), we estimated post-experiment mortality of surviving prawns at 6-14% (Figure X).

**DISCUSSION**

* Contextualise the results in terms of the fishery.
  + What is the critical window for survival, time-wise?
  + What are the implications of the temperature effect for the commercial fishery?
  + How might we speculate on the survival of egged females?
  + Commercial license requirements and enforcement of sorting tables
  + Increasing proportion of (under-sized) males caught towards end of season.
  + Evidence that it is worth making the effort
* Discussion of the physiological mechanisms for death and how we expect air exposure/temperature to affect that process
  + Acknowledge lack of consideration of density-dependence
  + Discuss possible dynamics re: temperature differential (per Dylan’s comments, cite sea lice research?)
  + Discuss reflex behaviours: suggestion of long-term survival, not just immediate.
* Discussion of limitations & considerations
  + Lost prawns
  + Issues comparing to ‘real world’ (if anything, we might be underestimating survival in some respects)
  + Accounting for predation (this keeps coming up, suggest follow up?). We have not accounted for the influence of predation but we may be under-estimating survival due to handling effects.
  + Could consider following up with a sensitivity analysis: what are the population-level consequences of higher or lower post-release survival?
* Couching results in broader picture of post-release survival studies.

**CONTRIBUTIONS**

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