

Class 19: Pertussis Mini Proj

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Background

Pertussis (aka Whooping cough) is a highly infectious lung infection caused by the bacteria *B. pertussis*.

The CDC tracks case numbers in the US and makes this data available online:

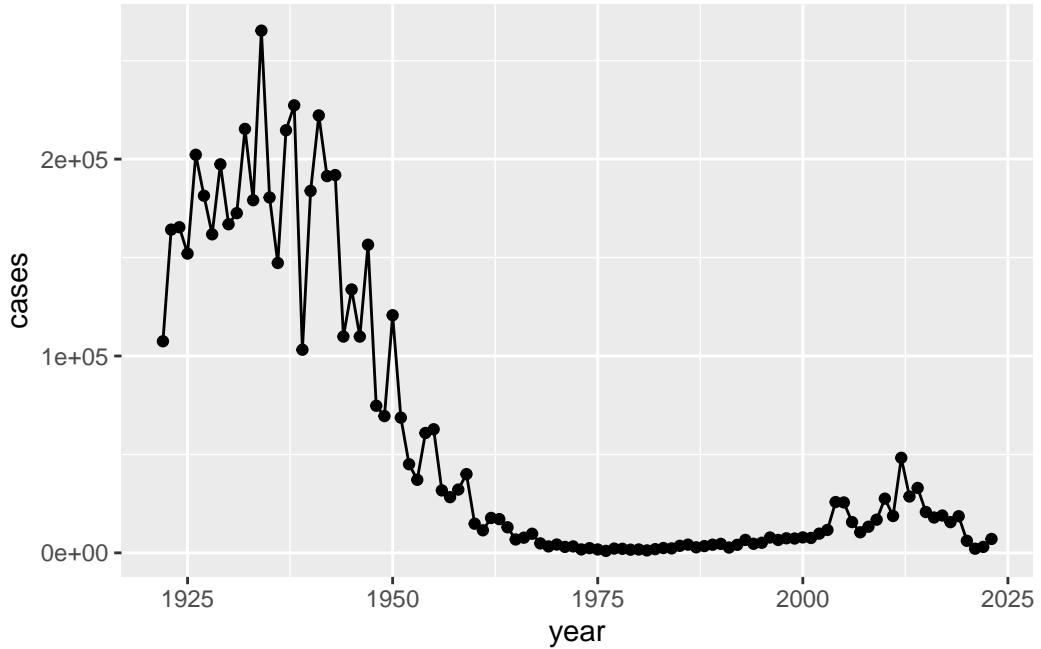
copied table from website initiate variable assign it to Addins>paste as data.frame -> pastes in the table in data.frame format highlight all and go to Code>reformat selection (cmd+shift+A)

Q1. Make a plot of pertussis cases per year with ggplot

```
library(ggplot2)
```

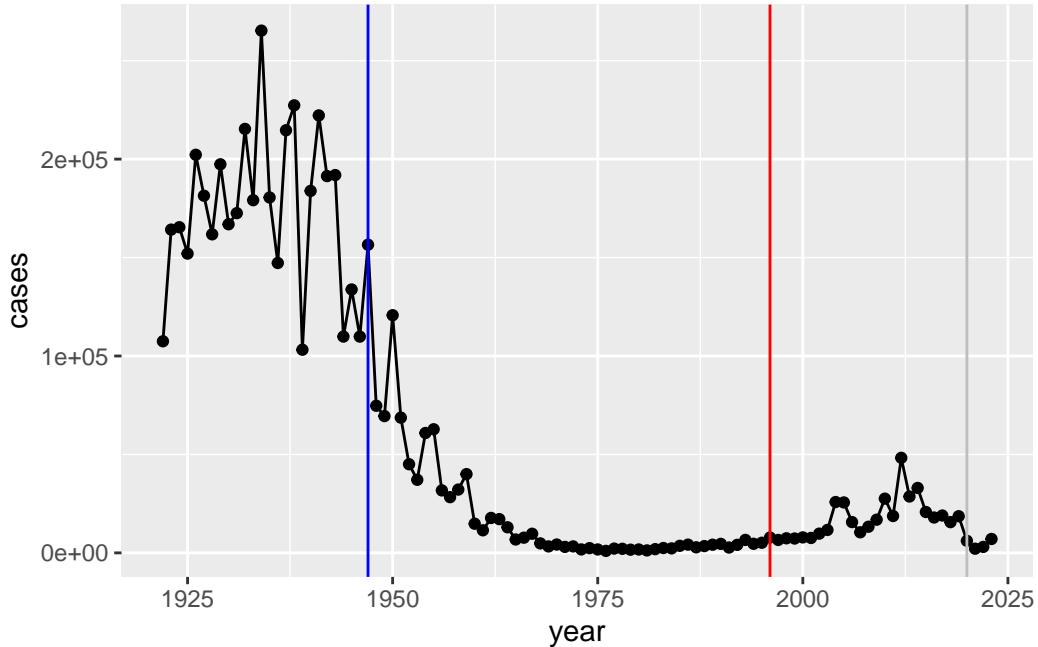
```
Warning: package 'ggplot2' was built under R version 4.5.2
```

```
ggplot(cdc) + aes(x=year, y=cases) + geom_point() + geom_line()
```



Q2. Add some annotation (lines on the plot) for some major milestones in our interaction with pertussis. The original wP deployment in 1947 and the newer aP vaccine roll-out in 1996. Finally a line for 2020

```
ggplot(cdc) + aes(x=year, y=cases) + geom_point() + geom_line() + geom_vline(xintercept = 1947)
```



This red line seems to precede a spike in cases. This is likely related to a few different factors, including anti-vaccine movements, slight changes in pertussis strain (and the vaccine needing to catch up), and most importantly how the aP vaccine was shown to require boosters for continued efficacy. aP is the cell-free vaccine, which was an improvement on the wP vaccine which used the whole bacterial cell to inoculate.

The CMI-PB Project

The CMI-Pertussis Boost (PB) project focuses on gathering data on this same topic. What is distinct between aP and wP individuals. aP induced protection wanes faster than wP...Why?

CMI-PB makes their data available via a JSON format returning API. We can read JSON format with the `read_json()` function from the **jsonlite** package

```
# install.packages("jsonlite")
library(jsonlite)

subject <- read_json("http://cmi-pb.org/api/v5_1/subject", simplifyVector = TRUE)
head(subject)
```

	subject_id	infancy_vac	biological_sex	ethnicity	race
1	1	wP	Female	Not Hispanic or Latino	White

```

2      2      wP      Female Not Hispanic or Latino White
3      3      wP      Female          Unknown White
4      4      wP      Male Not Hispanic or Latino Asian
5      5      wP      Male Not Hispanic or Latino Asian
6      6      wP      Female Not Hispanic or Latino White
year_of_birth date_of_boost     dataset
1  1986-01-01  2016-09-12 2020_dataset
2  1968-01-01  2019-01-28 2020_dataset
3  1983-01-01  2016-10-10 2020_dataset
4  1988-01-01  2016-08-29 2020_dataset
5  1991-01-01  2016-08-29 2020_dataset
6  1988-01-01  2016-10-10 2020_dataset

```

Q3. How many “subjects” are in this db?

```
nrow(subject)
```

```
[1] 172
```

Q4. How many wP and aP primed subjects are there in the dataset?

```
table(subject$infancy_vac)
```

```
aP wP
87 85
```

Q5. What is the **biological sex** and **race** breakdown of these subjects? Ie, is this a representative population?

```
table(subject$race, subject$biological_sex)
```

		Female	Male
American Indian/Alaska Native		0	1
Asian		32	12
Black or African American		2	3
More Than One Race		15	4
Native Hawaiian or Other Pacific Islander		1	1
Unknown or Not Reported		14	7
White		48	32

Not a representative sample :(

Let's read in more tables from the CMI-PB database API (go to website and get some more links)

```
specimen <- read_json("http://cmi-pb.org/api/v5_1/specimen", simplifyVector = TRUE)
ab_titer <- read_json("http://cmi-pb.org/api/v5_1/plasma_ab_titer", simplifyVector = TRUE)
```

A wee peak at these:

```
head(specimen)
```

	specimen_id	subject_id	actual_day_relative_to_boost
1	1	1	-3
2	2	1	1
3	3	1	3
4	4	1	7
5	5	1	11
6	6	1	32

	planned_day_relative_to_boost	specimen_type	visit
1	0	Blood	1
2	1	Blood	2
3	3	Blood	3
4	7	Blood	4
5	14	Blood	5
6	30	Blood	6

```
head(ab_titer)
```

	specimen_id	isotype	is_antigen_specific	antigen	MFI	MFI_normalised
1	1	IgE	FALSE	Total	1110.21154	2.493425
2	1	IgE	FALSE	Total	2708.91616	2.493425
3	1	IgG	TRUE	PT	68.56614	3.736992
4	1	IgG	TRUE	PRN	332.12718	2.602350
5	1	IgG	TRUE	FHA	1887.12263	34.050956
6	1	IgE	TRUE	ACT	0.10000	1.000000

	unit	lower_limit_of_detection
1	UG/ML	2.096133
2	IU/ML	29.170000
3	IU/ML	0.530000
4	IU/ML	6.205949
5	IU/ML	4.679535
6	IU/ML	2.816431

Join using the `inner_join()` function from `dplyr`

```
library(dplyr)
```

Attaching package: 'dplyr'

The following objects are masked from 'package:stats':

filter, lag

The following objects are masked from 'package:base':

intersect, setdiff, setequal, union

```
meta <- inner_join(subject, specimen)
```

Joining with `by = join_by(subject_id)`

```
head(meta)
```

	subject_id	infancy_vac	biological_sex	ethnicity	race
1	1	wP	Female	Not Hispanic or Latino	White
2	1	wP	Female	Not Hispanic or Latino	White
3	1	wP	Female	Not Hispanic or Latino	White
4	1	wP	Female	Not Hispanic or Latino	White
5	1	wP	Female	Not Hispanic or Latino	White
6	1	wP	Female	Not Hispanic or Latino	White

	year_of_birth	date_of_boost	dataset	specimen_id
1	1986-01-01	2016-09-12	2020_dataset	1
2	1986-01-01	2016-09-12	2020_dataset	2
3	1986-01-01	2016-09-12	2020_dataset	3
4	1986-01-01	2016-09-12	2020_dataset	4
5	1986-01-01	2016-09-12	2020_dataset	5
6	1986-01-01	2016-09-12	2020_dataset	6

	actual_day_relative_to_boost	planned_day_relative_to_boost	specimen_type
1	-3	0	Blood
2	1	1	Blood
3	3	3	Blood
4	7	7	Blood

```

5           11          14      Blood
6           32          30      Blood
visit
1     1
2     2
3     3
4     4
5     5
6     6

```

```
ab_data <- inner_join(meta, ab_titer)
```

Joining with `by = join_by(specimen_id)`

```
head(ab_data)
```

	subject_id	infancy_vac	biological_sex	ethnicity	race		
1	1	wP	Female	Not Hispanic or Latino	White		
2	1	wP	Female	Not Hispanic or Latino	White		
3	1	wP	Female	Not Hispanic or Latino	White		
4	1	wP	Female	Not Hispanic or Latino	White		
5	1	wP	Female	Not Hispanic or Latino	White		
6	1	wP	Female	Not Hispanic or Latino	White		
	year_of_birth	date_of_boost	dataset	specimen_id			
1	1986-01-01	2016-09-12	2020_dataset	1			
2	1986-01-01	2016-09-12	2020_dataset	1			
3	1986-01-01	2016-09-12	2020_dataset	1			
4	1986-01-01	2016-09-12	2020_dataset	1			
5	1986-01-01	2016-09-12	2020_dataset	1			
6	1986-01-01	2016-09-12	2020_dataset	1			
	actual_day_relative_to_boost	planned_day_relative_to_boost	specimen_type				
1	-3		0	Blood			
2	-3		0	Blood			
3	-3		0	Blood			
4	-3		0	Blood			
5	-3		0	Blood			
6	-3		0	Blood			
	visit	isotype	is_antigen_specific	antigen	MFI	MFI_normalised	unit
1	1	IgE	FALSE	Total	1110.21154	2.493425	UG/ML
2	1	IgE	FALSE	Total	2708.91616	2.493425	IU/ML
3	1	IgG	TRUE	PT	68.56614	3.736992	IU/ML

```

4      1    IgG           TRUE     PRN  332.12718      2.602350 IU/ML
5      1    IgG           TRUE     FHA 1887.12263      34.050956 IU/ML
6      1    IgE           TRUE     ACT   0.10000      1.000000 IU/ML
lower_limit_of_detection
1                  2.096133
2                  29.170000
3                  0.530000
4                  6.205949
5                  4.679535
6                  2.816431

```

Q6. How many different Ab isotypes are there?

```
unique(ab_data$isotype)
```

```
[1] "IgE"  "IgG"  "IgG1" "IgG2" "IgG3" "IgG4"
```

2 major isotypes (6 including IgG subtypes)

Q7. How many different antigens are there in the dataset?

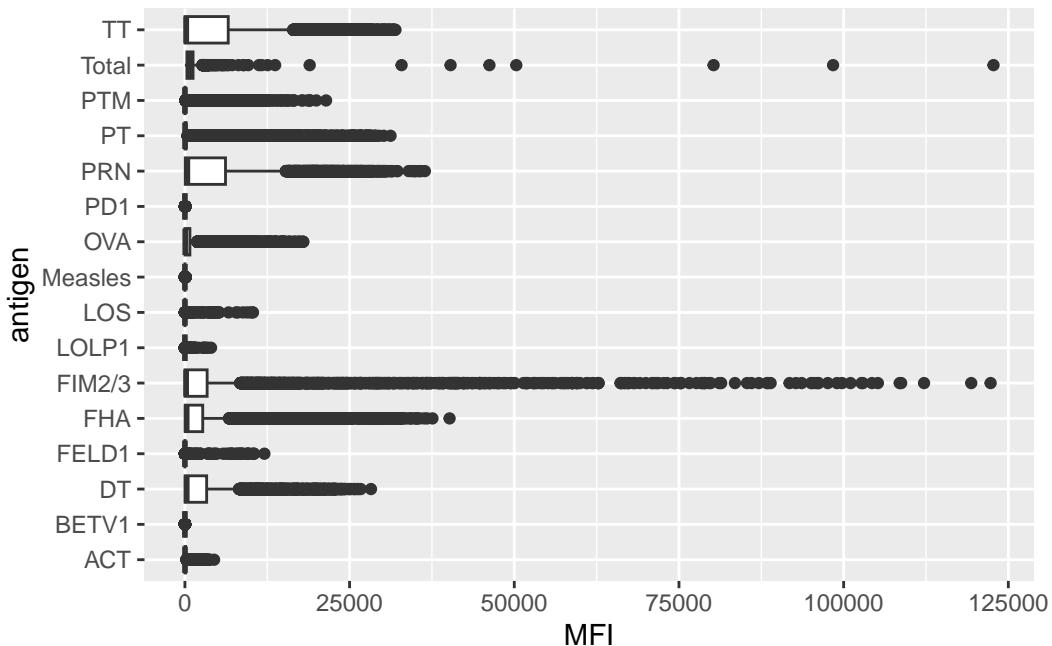
```
unique(ab_data$antigen)
```

```
[1] "Total"    "PT"       "PRN"      "FHA"      "ACT"      "LOS"      "FELD1"
[8] "BETV1"    "LOLP1"    "Measles"   "PTM"      "FIM2/3"   "TT"      "DT"
[15] "OVA"      "PD1"
```

Q8. Let's plot MFI vs antigen

```
ggplot(ab_data) + aes(x=MFI, y=antigen) + geom_boxplot()
```

Warning: Removed 1 row containing non-finite outside the scale range
(`stat_boxplot()`).



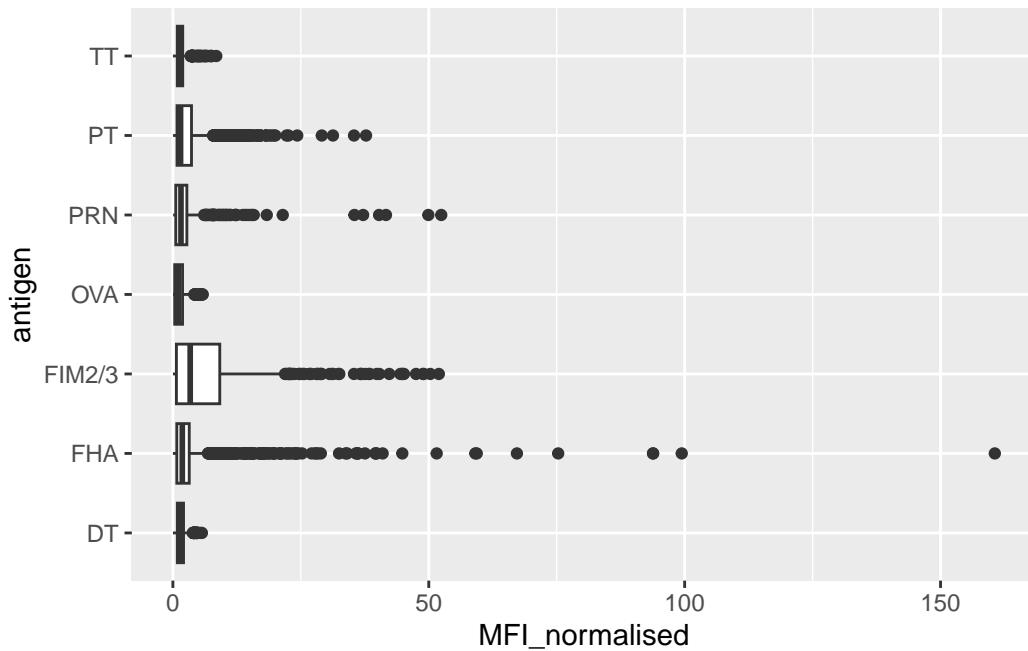
Focus in IgG

IgG is crucial for long-term immunity and responding to the bacterial and viral infections.

```
igg <- ab_data |>
  filter(isotype == "IgG")
```

Plot of antigen levels again but for IgG only

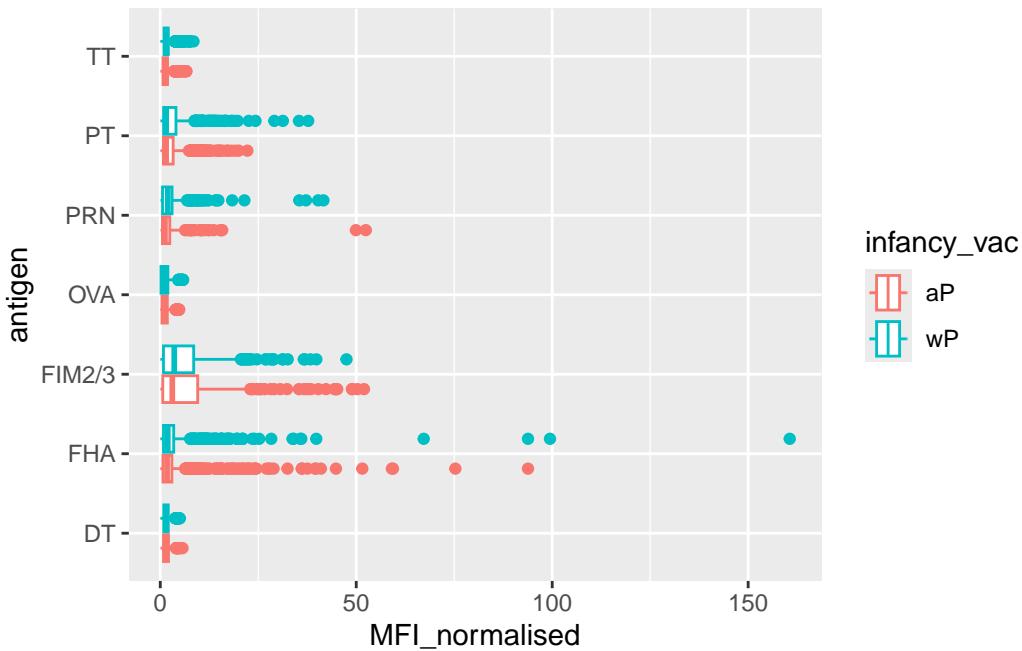
```
ggplot(igg) + aes(MFI_normalised, antigen) + geom_boxplot()
```



Differences between aP and wP?

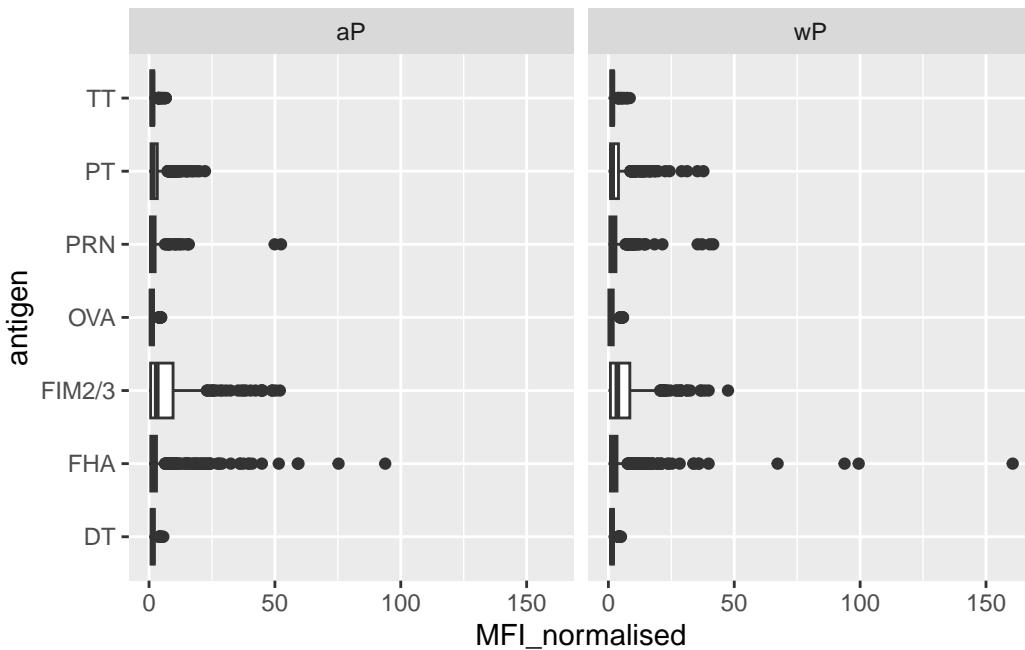
We can color up by the `infancy_vac` values of “wP” or “aP”

```
ggplot(igg) + aes(MFI_normalised, antigen, col=infancy_vac) + geom_boxplot()
```



We could also “facet” by the “aP” vs “wP” column

```
ggplot(igg) + aes(MFI_normalised, antigen) + geom_boxplot() + facet_wrap(~infancy_vac)
```



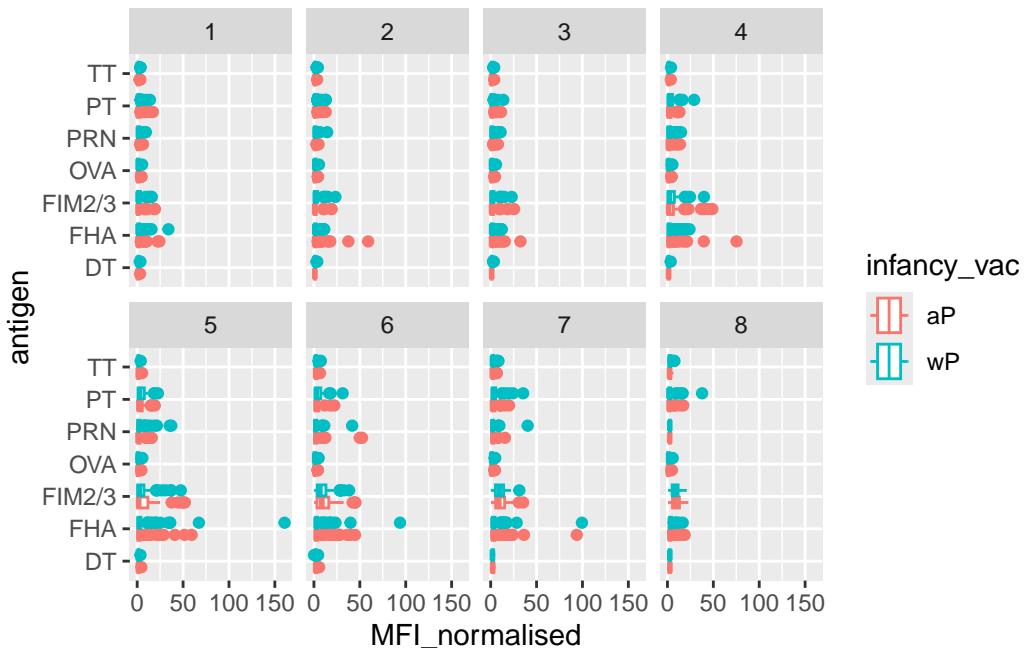
Time course analysis

We can use ‘visit’ as a proxy for time and facet our plots by this value 1 to 8 ...

```
table(ab_data$visit) # it's an ongoing study so more recent visits are still collecting data
```

1	2	3	4	5	6	7	8	9	10	11	12
8280	8280	8420	8420	8420	8100	7700	2670	770	686	105	105

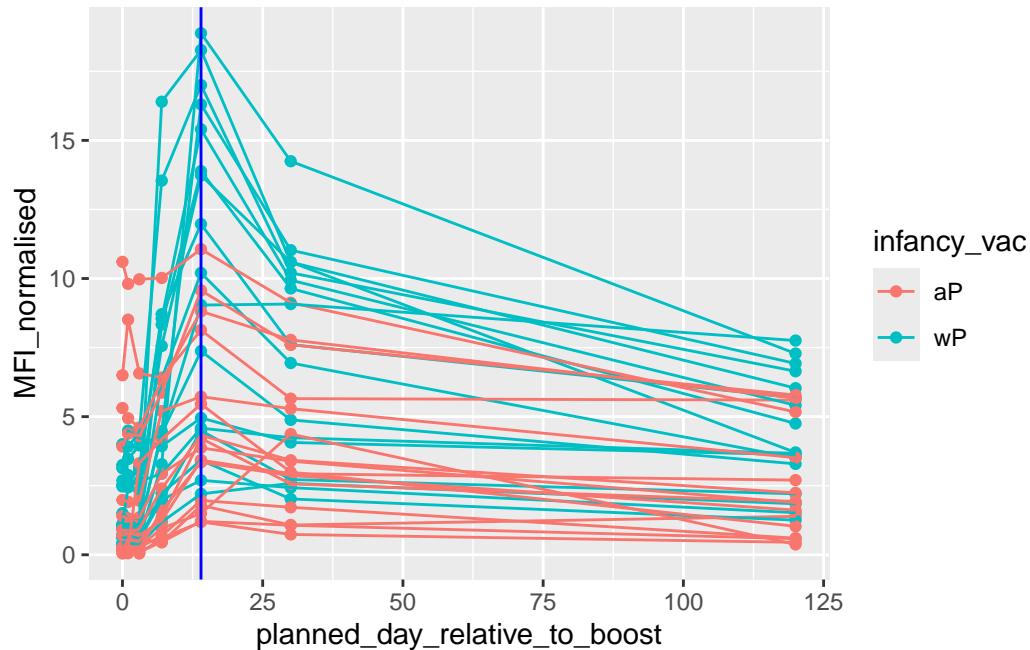
```
igg |>  
  filter(visit %in% 1:8) |>  
  ggplot() + aes(MFI_normalised, antigen, col=infancy_vac) + geom_boxplot() + facet_wrap(~vi
```



Time course of PT (Virulence Factor: Pertussis Toxin)

```
pt <- igg |>  
  filter(antigen == "PT") |>  
  filter(dataset == "2021_dataset")
```

```
ggplot(pt) +
  aes(planned_day_relative_to_boost,
      MFI_normalised,
      col=infancy_vac,
      group=subject_id) +
  geom_point() +
  geom_line() +
  geom_vline(xintercept = 14, col="blue")
```



System setup

```
sessionInfo()
```

```
R version 4.5.1 (2025-06-13)
Platform: aarch64-apple-darwin20
Running under: macOS Sonoma 14.6.1
```

```
Matrix products: default
BLAS:    /Library/Frameworks/R.framework/Versions/4.5-arm64/Resources/lib/libRblas.0.dylib
LAPACK:  /Library/Frameworks/R.framework/Versions/4.5-arm64/Resources/lib/libRlapack.dylib; 1
```

```
locale:  
[1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8  
  
time zone: America/Los_Angeles  
tzcode source: internal  
  
attached base packages:  
[1] stats      graphics   grDevices utils      datasets   methods    base  
  
other attached packages:  
[1] dplyr_1.1.4   jsonlite_2.0.0 ggplot2_4.0.1  
  
loaded via a namespace (and not attached):  
[1] vctrs_0.6.5          cli_3.6.5           knitr_1.50          rlang_1.1.6  
[5] xfun_0.54            generics_0.1.4       S7_0.2.1           labeling_0.4.3  
[9] glue_1.8.0           htmltools_0.5.8.1    scales_1.4.0        rmarkdown_2.30  
[13] grid_4.5.1           evaluate_1.0.5       tibble_3.3.0        fastmap_1.2.0  
[17] yaml_2.3.11          lifecycle_1.0.4      compiler_4.5.1     RColorBrewer_1.1-3  
[21] pkgconfig_2.0.3      rstudioapi_0.17.1    farver_2.1.2       digest_0.6.39  
[25] R6_2.6.1             tidyselect_1.2.1     pillar_1.11.1      magrittr_2.0.4  
[29] withr_3.0.2          tools_4.5.1          gtable_0.3.6
```