CavPred 1.0.0

Examples

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Master in Bioinformatics for Health Sciences

Contents

Example 1: detailed analysis of results for the protein 1acj			
a. Preprocessing step	2		
b. Grid step	3		
c. Voronoi step	4		
d. Combination step	5		
Example 2: Results for protein 1mbn	6		

Example 1: detailed analysis of results for the protein 1acj

The pdb file for protein 1acj was downloaded from the RCSB PDB database (https://files.rcsb.org/download/1ACJ.pdb). Then the program is run with the following command (in the same folder where it has been installed):

python3 CavPred-1.0.0/cavpred/get_binding.py 1acj.pdb

```
esults_1acj/
   1acj_binding_coloured.pdb
   1acj_binding_residues.txt
   1acj H.pdb
   1acj.log
   1acj.pqr
   1acj_protein.pdb
   grid_files
       cavity_with_depth.pdb
       hydropathy.pdb
       results.toml
   voronoi files
       1acj_binding_sites.txt
       1acj_delaunay.png
       1acj_voronoi.png
       1acj_voronoi.txt
```

After running the program, many files are generated. We will run through each one of them to better understand the pipeline of CavPred and the results it gives.

a. Preprocessing step

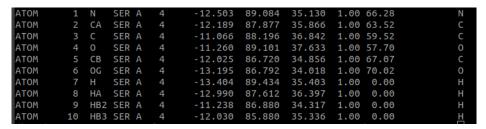
The first part of the program consists in modulating the initial pdb file so it can be processed correctly by the prediction algorithms.

1acj_protein.pdb

```
ATOM 1 N SER A 4 -12.503 89.084 35.130 1.00 66.28 N
ATOM 2 CA SER A 4 -12.189 87.877 35.866 1.00 63.52 C
ATOM 3 C SER A 4 -11.066 88.196 36.842 1.00 59.52 C
ATOM 4 O SER A 4 -11.260 89.101 37.633 1.00 57.70 O
ATOM 5 CB SER A 4 -12.025 86.720 34.856 1.00 67.07 C
ATOM 6 OG SER A 4 -13.195 86.792 34.018 1.00 70.02 O
ATOM 7 N GLU A 5 -9.920 87.515 36.768 1.00 54.99 N
ATOM 8 CA GLU A 5 -8.763 87.673 37.662 1.00 47.42 C
ATOM 9 C GLU A 5 -9.082 86.841 38.872 1.00 43.05 C
ATOM 10 O GLU A 5 -8.332 85.956 39.236 1.00 44.40 O
```

This file is generated after extracting only the protein information from the pdb file, in other words, all lines starting with the word "ATOM". As a result, if the initial file had a ligand (which is our case), it is no longer present in this one.

1acj H.pdb





This file has Hydrogen molecules added to the pdb file, which is useful for the analysis later on.

1acj.pqr

/								,		_
ATOM	1	N	SER	Α	4	-12.503	89.084	35.130	0.1849	1.8240
ATOM	2	CA	SER	Α	4	-12.189	87.877	35.866	0.0567	1.9080
ATOM	3	C	SER	Α	4	-11.066	88.196	36.842	0.6163	1.9080
ATOM	4	0	SER	Α	4	-11.260	89.101	37.633	-0.5722	1.6612
ATOM	5	CB	SER	Α	4	-12.025	86.720	34.856	0.2596	1.9080
ATOM	6	OG	SER	Α	4	-13.195	86.792	34.018	-0.6714	1.7210
ATOM	7	Н	SER	Α	4	-13.404	89.434	35.403	0.1898	0.6000
ATOM	8	HA	SER	Α	4	-12.990	87.612	36.397	0.0782	1.1000
ATOM	9	HB2	SER	Α	4	-11.238	86.880	34.317	0.0273	1.3870
ATOM	10	HB3	SER	Α	4	-12.030	85.880	35.336	0.0273	1.3870

This file is the exact same as 1acj_H.pdb but converted to pqr format.

b. Grid step

This step consists in predicting binding sites thanks to geometrical information and some additional information on hydrophobicity.

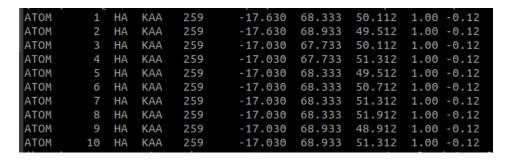
In total, the grid step found a total number of 16 cavities. The most voluminous and deep one being the cavity named KAI by the program.

1acj_cavity_with_depth.pdb

ATOM	1	HA	KAA	259	-17.630	68.333	50.112	1.00	0.00
ATOM	2	HA	KAA	259	-17.630	68.933	49.512	1.00	0.00
ATOM	3	HA	KAA	259	-17.030	67.733	50.112	1.00	0.85
ATOM	4	HA	KAA	259	-17.030	67.733	51.312	1.00	1.34
ATOM	5	HA	KAA	259	-17.030	68.333	49.512	1.00	0.85
ATOM	6	Н	KAA	259	-17.030	68.333	50.112	1.00	0.60
ATOM	7	HA	KAA	259	-17.030	68.333	50.712	1.00	0.60
ATOM	8	HA	KAA	259	-17.030	68.333	51.312	1.00	0.85
ATOM	9	HA	KAA	259	-17.030	68.333	51.912	1.00	1.34
ATOM	10	HA	KAA	259	-17.030	68.933	48.912	1.00	0.85

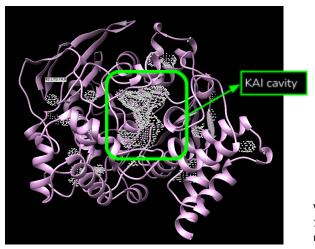
This file stores the coordinates for the predicted cavities based on geometrical and depth information about the 3D disposition of the amino acids. It only contains information on the cavities, not the actual protein, so, when visualized with any graphic software, such as Chimera, it has to be visualized together with 1acj_protein.pdb.

1acj_hydropathy.pdb



Similarly to the previous file, this one contains additional information on hydropathy for each predicted cavity. Again, it only contains the coordinates of the cavities, so it has to be visualized together with the 1acj_protein.pdb file.





Visualization of hydropathy.pdb (white) and 1acj_protein.pdb (pink) in the Chimera software. The most relevant cavity (KAI) is highlighted in green.

1acj_results.toml

```
# pykVFinder results

[FILES]
INPUT = "/home/anna/Downloads/results_1acj/1acj_H.pdb"
OUTPUT = "/home/anna/Downloads/results_1acj/grid_files/cavity_with_depth.pd

[PARAMETERS]
STEP = 0.6

[RESULTS.VOLUME]
KAA = 19.01
KAB = 29.16
KAC = 6.7
KAD = 12.31
KAE = 12.31
KAF = 16.85
KAG = 30.89
KAH = 12.31
KAI = 604.15
KAJ = 6.05

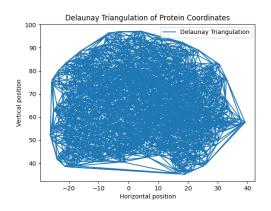
[RESULTS.FREQUENCY.KAI.RESIDUES]
ARG = 1
ASN = 1
ASP = 1
GLN = 1
GL
```

This file stores information about all the cavities, with volume, depth and area information. Additionally, it contains information about which residues are found in each of the cavities.

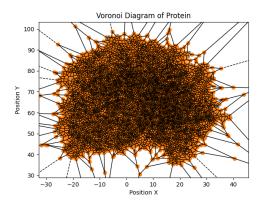
c. Voronoi step

This step found a total of 357 residues to be possible protein pockets.

1acj_delaunay.png



1acj_voronoi.png





These two files contain the graphical results of the Voronoi analysis, which are later used to calculate potential binding sites.

1acj_binding_sites.txt

```
GLU 5
LEU 7
VAL 8
ASN 9
LYS 11
SER 12
GLY 13
LYS 14
VAL 15
MET 16
```

This file contains all residues considered potential binding sites by the Voronoi algorithm.

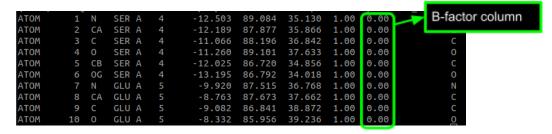
d. Combination step

Lastly, the program keeps only the aminoacid positions that are considered potential cavities by both the grid and Voronoi calculations. In the case of 1acj, 60.51% of the amino acids predicted to be in a binding site according to the grid method are also likely to be in a binding site according to the Voronoi method.

1acj_binding_residues.txt

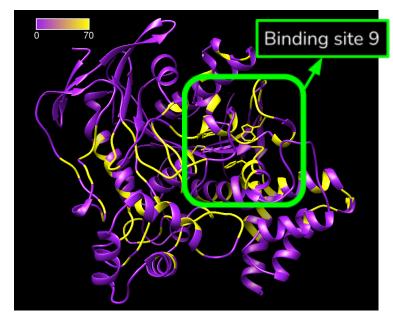
This file contains all the different cavities found in the protein and the list of residues corresponding to them. The previously identified KAI cavity is now binding site 9.

1acj_binding_coloured.pdb



This file contains all the residues in the protein, and those potential to be part of a binding site have b-factor value (column 11) set to 70, while the rest have 0. This way, these residues can be visually differentiated with visualization softwares.



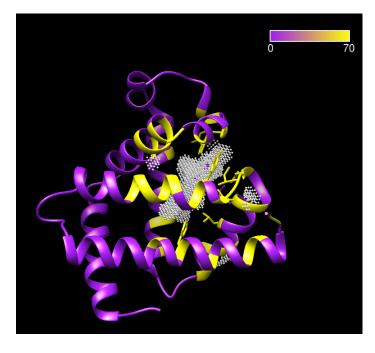


Visualization of1acj_binding_coloured.pdb in the Chimera software. The yellow-colored residues are those which have b-factor 70, so these correspond to potential binding sites. Binding site 9 (highlighted in green), which is the largest, correpsonds to the part where the ligand interacts in the original protein.

Example 2: Results for protein 1mbn

The pdb file for protein 1mbn was downloaded from the RCSB PDB database (https://files.rcsb.org/download/1MBN.pdb). This is a small protein, which helps in viusalizing the results.

python3 CavPred-1.0.0/cavpred/get_binding.py 1mbn.pdb



Visualization of 1mbn_binding_coloured.pdb together with 1mbn_hydropathy.pdb in the Chimera software. The yellow-colored residues are those which have b-factor 70, corresponding to potential binding sites. As it can be seen, the predicted binding sites correspond to where the ligand binds in the protein.

