

Practical section II: Introduction to R and RNAseq data analysis

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Outline

Part I: Introduction to R

- S1: Data structures and basic operations.
- S2: Data import and export.
- S3: Summary statistics and data visualization.

Part II: Introduction to RNAseq data analysis

- S4: RNAseq data analysis with DESeq2.
- S5: Exploring and visualizing RNAseq data.

Course materials on GitHub

What is GitHub?

GitHub is like a digital library where people store and share their writing projects, allowing others to view, discuss, or contribute to them. Think of it as a collaborative workspace for writers, but instead of stories or essays, people work on computer programs.



GitHub repository:

<https://github.com/enriquea/ZebraQ>

Downloading the repo:

```
$ git clone https://github.com/enriquea/ZebraQ.git
```

```
eam:~ eam$ git clone https://github.com/enriquea/ZebraQ.git
Cloning into 'ZebraQ'...
remote: Enumerating objects: 75, done.
remote: Counting objects: 100% (75/75), done.
remote: Compressing objects: 100% (61/61), done.
remote: Total 75 (delta 18), reused 61 (delta 11), pack-reused 0
Receiving objects: 100% (75/75), 9.98 MiB | 1.10 MiB/s, done.
Resolving deltas: 100% (18/18), done.
```

Course materials (Dataset)

Disclaimer: The data used in this course is intended to be used for educational purposes only.

`/data` folder contains the following files:

- Genes associated with CHD with functional annotations:

`/data/chd_genes.annotations.tsv`

- Gene counts from RNA-seq data of wild type and mutant zebrafish (*Danio Rerio*) hearts:

`/data/salmon.merged.gene_counts.tsv`

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Part I: Introduction to R

RStudio



1. Text editor (scripts)

```
6  html_document: default
7  pdf_document: default
...
11 knitr::opts_chunk$set(echo = FALSE)
12
13
14 ## 1. Load libraries
15
16 ```{r load libraries, echo = TRUE, message=FALSE, warning=FALSE}
17
18 library(DESeq2, quietly = TRUE)
19 library(ggplot2, quietly = TRUE)
20 library(RColorBrewer, quietly = TRUE)
21 library(EnhancedVolcano, quietly = TRUE)
22
```

2. R console

```
R 4.1.3 ~ ~/projects/local/ZebraQ/
R version 4.1.3 (2022-03-10) -- "One Push-Up"
Copyright (C) 2022 The R Foundation for Statistical Computing
Platform: x86_64-apple-darwin17.0 (64-bit)

R is free software and comes with ABSOLUTELY NO WARRANTY.
You are welcome to redistribute it under certain conditions.
Type 'license()' or 'licence()' for distribution details.

Natural language support but running in an English locale

R is a collaborative project with many contributors.
Type 'contributors()' for more information and
'citation()' on how to cite R or R packages in publications.

help()' for on-line help, or
type 'q()' to quit R.

>
```

3. R environment

Environment History Connections Git

R Global Environment

- package:stats
- package:graphics
- package:grDevices
- package:utils
- package:datasets
- package:methods
- package:base

Environment is empty

4. Files explorer

Name	Size	Modified
.DS_store	10 KB	Jun 23, 2024, 2:27 PM
.git		
.gitignore	260 B	Sep 5, 2024, 2:25 PM
.Rhistory	14.9 KB	Sep 15, 2023, 1:39 PM
.Rproj.user		
data		
docs		
lessons		
local		
README.md		
results		
setup		
slides		
ZebraQ.Rproj	205 B	Sep 5, 2024, 2:17 PM

Part I: Introduction to R

Base R Cheat Sheet

Getting Help

Accessing the help files

?mean

Get help of a particular function.

help.search('weighted mean')

Search the help files for a word or phrase.

help(package = 'dplyr')

Find help for a package.

More about an object

str(iris)

Get a summary of an object's structure.

class(iris)

Find the class an object belongs to.

Using Libraries

install.packages('dplyr')

Download and install a package from CRAN.

library(dplyr)

Load the package into the session, making all its functions available to use.

dplyr::select

Use a particular function from a package.

data(iris)

Load a built-in dataset into the environment.

Working Directory

getwd()

Find the current working directory (where inputs are found and outputs are sent).

setwd('C://file/path')

Change the current working directory.

Use projects in RStudio to set the working directory to the folder you are working in.

Vectors

Creating Vectors

<code>c(2, 4, 6)</code>	<code>2 4 6</code>	Join elements into a vector
<code>2:6</code>	<code>2 3 4 5 6</code>	An integer sequence
<code>seq(2, 3, by=0.5)</code>	<code>2.0 2.5 3.0</code>	A complex sequence
<code>rep(1:2, times=3)</code>	<code>1 2 1 2 1 2</code>	Repeat a vector
<code>rep(1:2, each=3)</code>	<code>1 1 1 2 2 2</code>	Repeat elements of a vector

Vector Functions

sort(x)

Return x sorted.

table(x)

See counts of values.

rev(x)

Return x reversed.

unique(x)

See unique values.

Selecting Vector Elements

By Position

<code>x[4]</code>	The fourth element.
<code>x[-4]</code>	All but the fourth.
<code>x[2:4]</code>	Elements two to four.
<code>x[-(2:4)]</code>	All elements except two to four.
<code>x[c(1, 5)]</code>	Elements one and five.

By Value

<code>x[x == 10]</code>	Elements which are equal to 10.
<code>x[x < 0]</code>	All elements less than zero.
<code>x[x %in% c(1, 2, 5)]</code>	Elements in the set 1, 2, 5.

Named Vectors

<code>x['apple']</code>	Element with name 'apple'.
-------------------------	----------------------------

Programming

For Loop

```
for (variable in sequence) {
  Do something
}
```

Example

```
for (i in 1:4) {
  j <- i + 10
  print(j)
}
```

While Loop

```
while (condition) {
  Do something
}
```

Example

```
while (i < 5) {
  print(i)
  i <- i + 1
}
```

If Statements

```
if (condition) {
  Do something
} else {
  Do something different
}
```

Example

```
if (i > 3) {
  print('Yes')
} else {
  print('No')
}
```

Functions

```
function_name <- function(var) {
  Do something
  return(new_variable)
}
```

Example

```
square <- function(x) {
  squared <- x*x
  return(squared)
}
```

Reading and Writing Data

Input	Output	Description
<code>df <- read.table('file.txt')</code>	<code>write.table(df, 'file.txt')</code>	Read and write a delimited text file.
<code>df <- read.csv('file.csv')</code>	<code>write.csv(df, 'file.csv')</code>	Read and write a comma separated value file. This is a special case of read.table/write.table.
<code>load('file.Rdata')</code>	<code>save(df, file = 'file.Rdata')</code>	Read and write an R data file, a file type special for R.

Conditions	a == b	Are equal	a > b	Greater than	a >= b	Greater than or equal to	is.na(a)	Is missing
	a != b	Not equal	a < b	Less than	a <= b	Less than or equal to	is.null(a)	Is null

Part I: Introduction to R

Types

Converting between common data types in R. Can always go from a higher value in the table to a lower value.

as.logical	TRUE, FALSE, TRUE	Boolean values (TRUE or FALSE).
as.numeric	1, 0, 1	Integers or floating point numbers.
as.character	'1', '0', '1'	Character strings. Generally preferred to factors.
as.factor	'1', '0', '1', levels: '1', '0'	Character strings with preset levels. Needed for some statistical models.

Maths Functions

log(x)	Natural log.	sum(x)	Sum.
exp(x)	Exponential.	mean(x)	Mean.
max(x)	Largest element.	median(x)	Median.
min(x)	Smallest element.	quantile(x)	Percentage quantiles.
round(x, n)	Round to n decimal places.	rank(x)	Rank of elements.
signif(x, n)	Round to n significant figures.	var(x)	The variance.
cor(x, y)	Correlation.	sd(x)	The standard deviation.

Variable Assignment

```
> a <- 'apple'
> a
[1] 'apple'
```







The Environment

ls()	List all variables in the environment.
rm(x)	Remove x from the environment.
rm(list = ls())	Remove all variables from the environment.

You can use the environment panel in RStudio to browse variables in your environment.

Matrixes

```
m <- matrix(x, nrow = 3, ncol = 3)
Create a matrix from x.
```

 m[2,]	Select a row	 t(m)	Transpose
 m[, 1]	Select a column	 m %*% n	Matrix Multiplication
 m[2, 3]	Select an element	 solve(m, n)	Find x in: m * x = n

Lists

```
l <- list(x = 1:5, y = c('a', 'b'))
A list is collection of elements which can be of different types.
```

 l[[2]]	Second element of l.	 l[1]	New list with only the first element.
 l\$x	Element named x.	 l['y']	New list with only element named y.




Also see the **dplyr** library.

Data Frames



```
df <- data.frame(x = 1:3, y = c('a', 'b', 'c'))
A special case of a list where all elements are the same length.
```

x	y
1	a
2	b
3	c

Matrix subsetting

 df[, 2]	
 df[2,]	
 df[2, 2]	

List subsetting

 df\$x		 df[[2]]	
---	--	---	--

Understanding a data frame

View(df) See the full data frame.

head(df) See the first 6 rows.

nrow(df)
Number of rows.

ncol(df)
Number of columns.

dim(df)
Number of columns and rows.

cbind - Bind columns.

rbind - Bind rows.

Strings

Also see the **stringr** library.

paste(x, y, sep = ' ')	Join multiple vectors together.
paste(x, collapse = ' ')	Join elements of a vector together.
grep(pattern, x)	Find regular expression matches in x.
gsub(pattern, replace, x)	Replace matches in x with a string.
toupper(x)	Convert to uppercase.
tolower(x)	Convert to lowercase.
nchar(x)	Number of characters in a string.

Factors

factor(x)	Turn a vector into a factor. Can set the levels of the factor and the order.
cut(x, breaks = 4)	Turn a numeric vector into a factor but 'cutting' into sections.

Statistics

lm(x ~ y, data=df)	Linear model.	t.test(x, y)	Perform a t-test for difference between means.	prop.test	Test for a difference between proportions.
glm(x ~ y, data=df)	Generalised linear model.	pairwise.t.test	Perform a t-test for paired data.	aov	Analysis of variance.
summary	Get more detailed information out a model.				

Distributions

	Random Variates	Density Function	Cumulative Distribution	Quantile
Normal	rnorm	dnorm	pnorm	qnorm
Poisson	rpois	dpois	ppois	qpois
Binomial	rbinom	dbinom	pbinom	qbinom
Uniform	runif	dunif	punif	qunif

Plotting

Also see the **ggplot2** library.

 plot(x)	Values of x in order.	 plot(x, y)	Values of x against y.	 hist(x)	Histogram of x.
---	-----------------------	--	------------------------	---	-----------------

Dates

See the **lubridate** library.

Outline

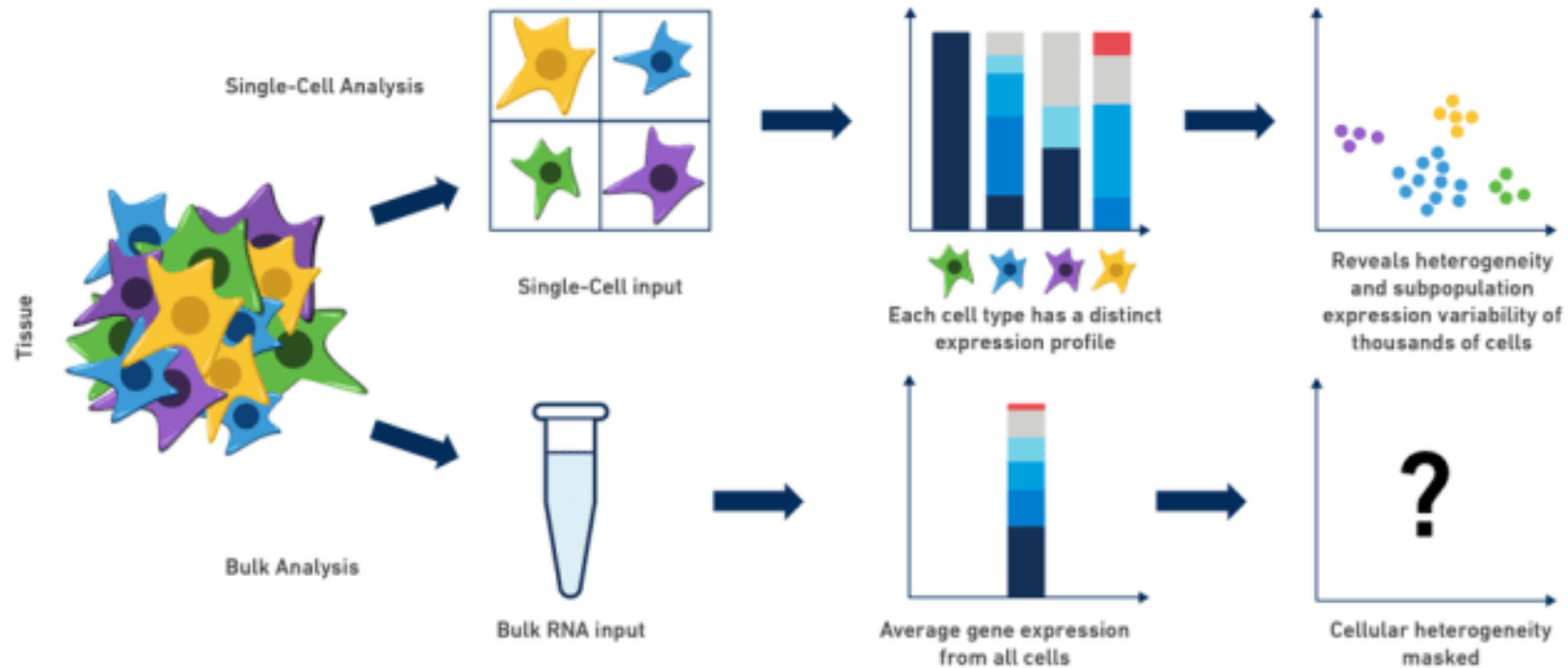
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Part II: Introduction to RNAseq data analysis (Supp. slides)

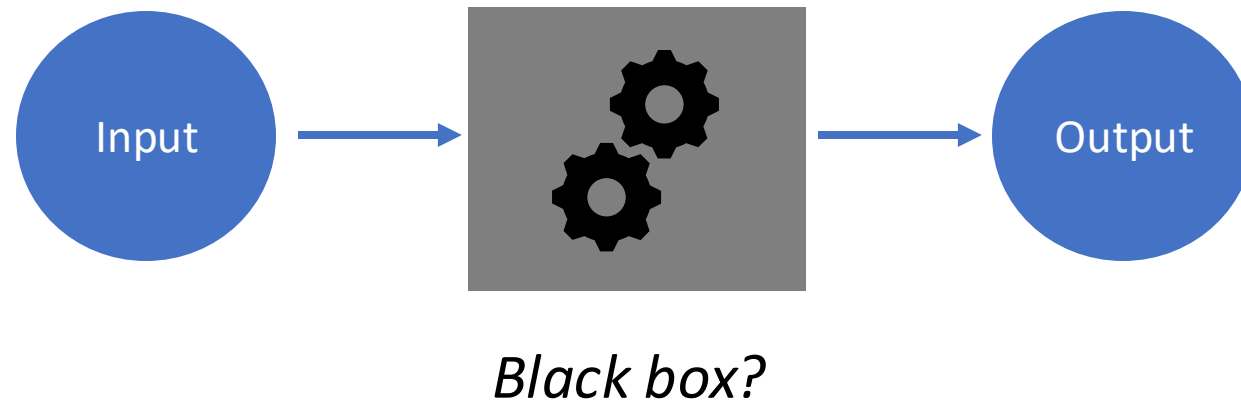
- S4: RNAseq data analysis with DESeq2.
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Bulk vs scRNA-seq

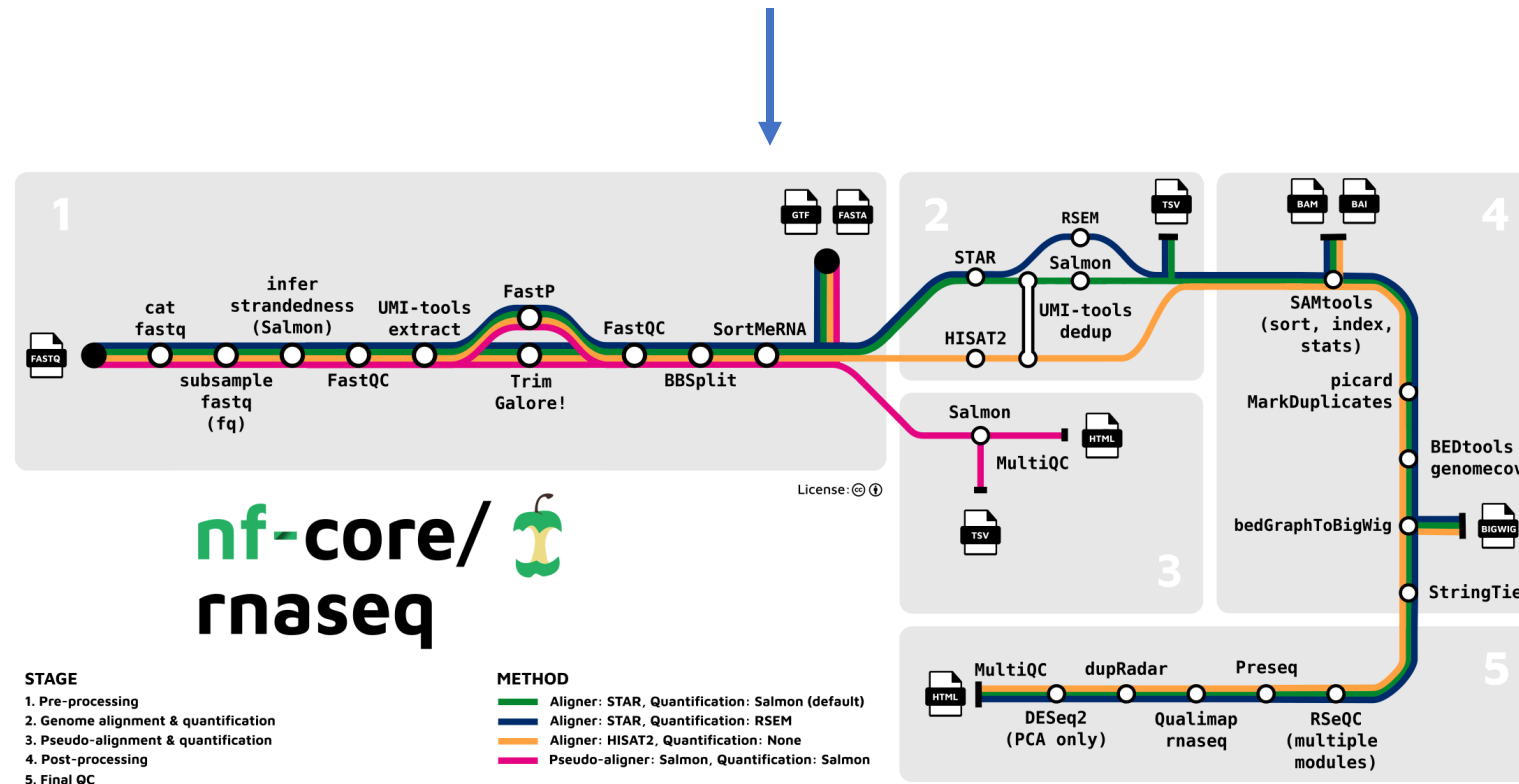
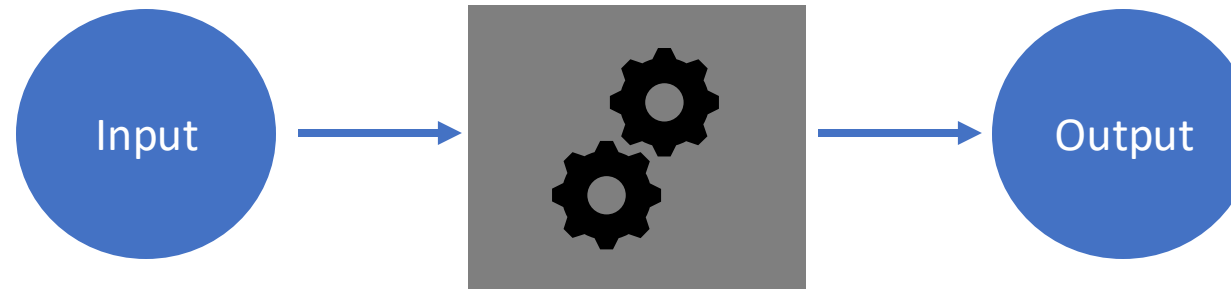


<https://www.10xgenomics.com/blog/single-cell-rna-seq-an-introductory-overview-and-tools-for-getting-started>

RNAseq pipeline

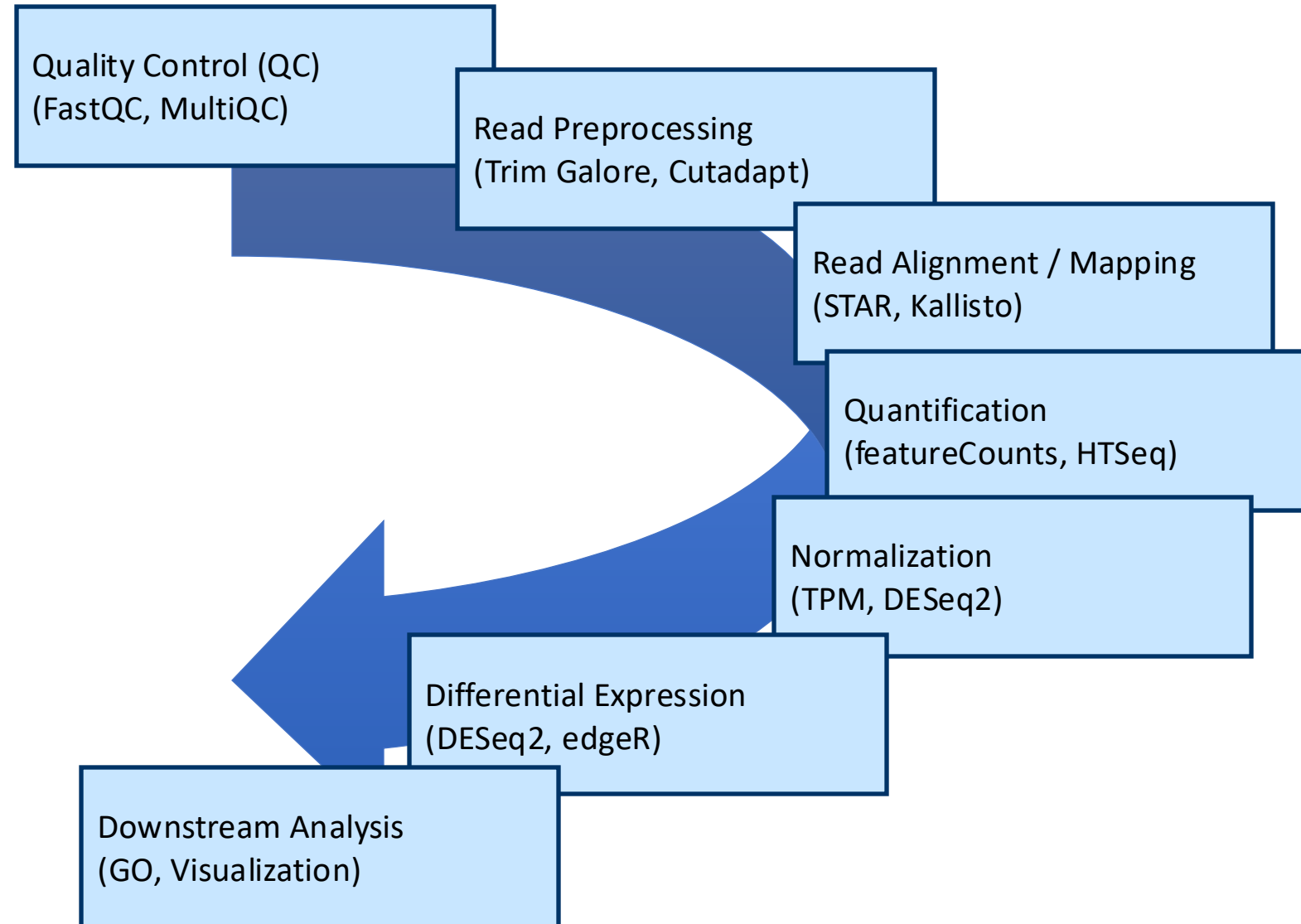


RNAseq pipeline



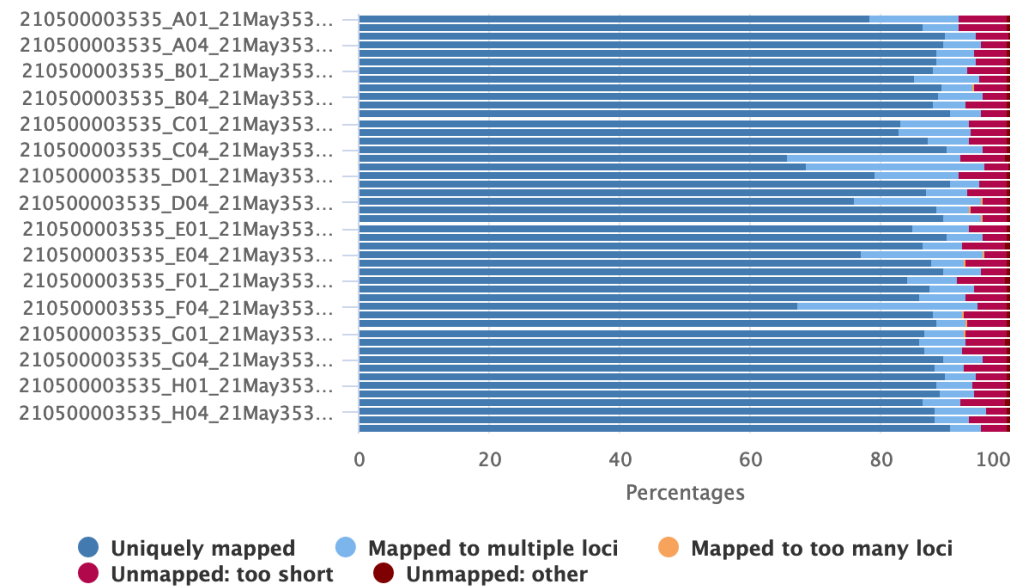
<https://nf-co.re/rnaseq>

RNA-seq pipeline: main bioinformatics steps



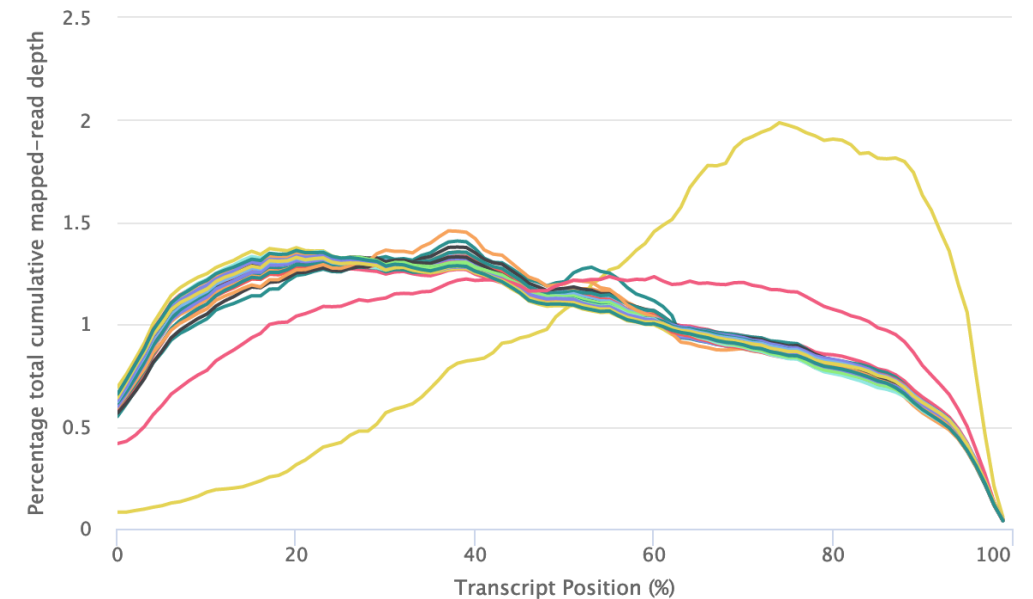
QC visualization and monitoring

STAR: Alignment Scores



Created with MultiQC

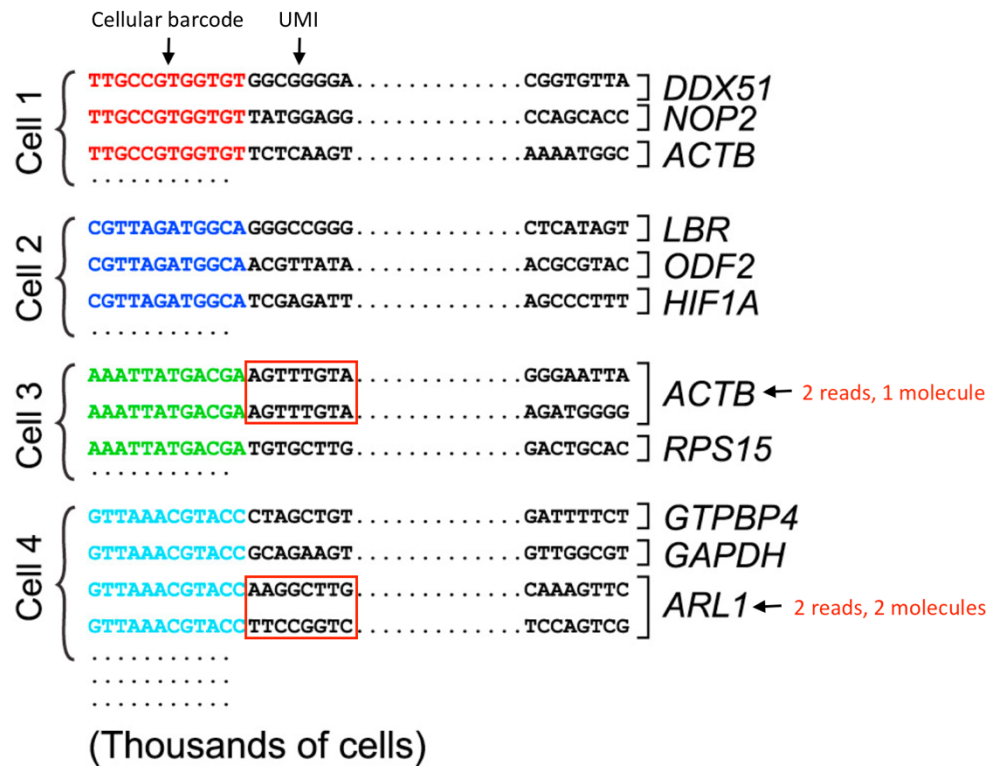
Qualimap RNAseq: Coverage Profile Along Genes (total)



Created with MultiQC

“Matrix count”

“The outcome of this procedure is a gene/cell count matrix, which is used as an estimate of the number of RNA molecules in each cell for each gene”



	Cell1	Cell2	...	CellN
<i>Gene1</i>	3	2	.	13
<i>Gene2</i>	2	3	.	1
<i>Gene3</i>	1	14	.	18
...
...
...
<i>GeneM</i>	25	0	.	0

Source: <https://github.com/hbctraining/scRNA-seq>

Results from DESeq experiment

The results table when printed will provide the information about the comparison, e.g. "log2 fold change (MAP): condition treated vs untreated", meaning that the estimates are of $\log_2(\text{treated} / \text{untreated})$.

```
[1] "Comparison performed: condition_MT_vs_WT"
log2 fold change (MLE): condition MT vs WT
Wald test p-value: condition MT vs WT
DataFrame with 13755 rows and 6 columns
```

	baseMean	log2FoldChange	lfcSE	stat	pvalue	padj
	<numeric>	<numeric>	<numeric>	<numeric>	<numeric>	<numeric>
LOC100000024	35.01346	1.2466174	0.670687	1.8587185	0.063067	0.125215
LOC100000576	401.25055	-0.2115917	0.220422	-0.9599394	0.337086	0.467804
LOC100000851	2.21568	0.0301926	1.226557	0.0246157	0.980361	0.988411
LOC100001344	61.36695	-0.1027153	0.246557	-0.4165978	0.676973	0.774125
LOC100001550	42.24091	0.5415869	0.334623	1.6184984	0.105555	0.190337
...
zwilch	400.60805	0.5873992	0.121817	4.821984	1.42137e-06	1.21568e-05
zyg11	1465.05339	-0.5332838	0.168022	-3.173889	1.50411e-03	5.47396e-03
zymnd12	8.45217	-0.6658308	0.683108	-0.974708	3.29705e-01	4.60536e-01
zyx	1029.10390	-0.0871254	0.148073	-0.588395	5.56267e-01	6.73980e-01
zzz3	1124.36740	-0.4385312	0.207517	-2.113229	3.45811e-02	7.64093e-02

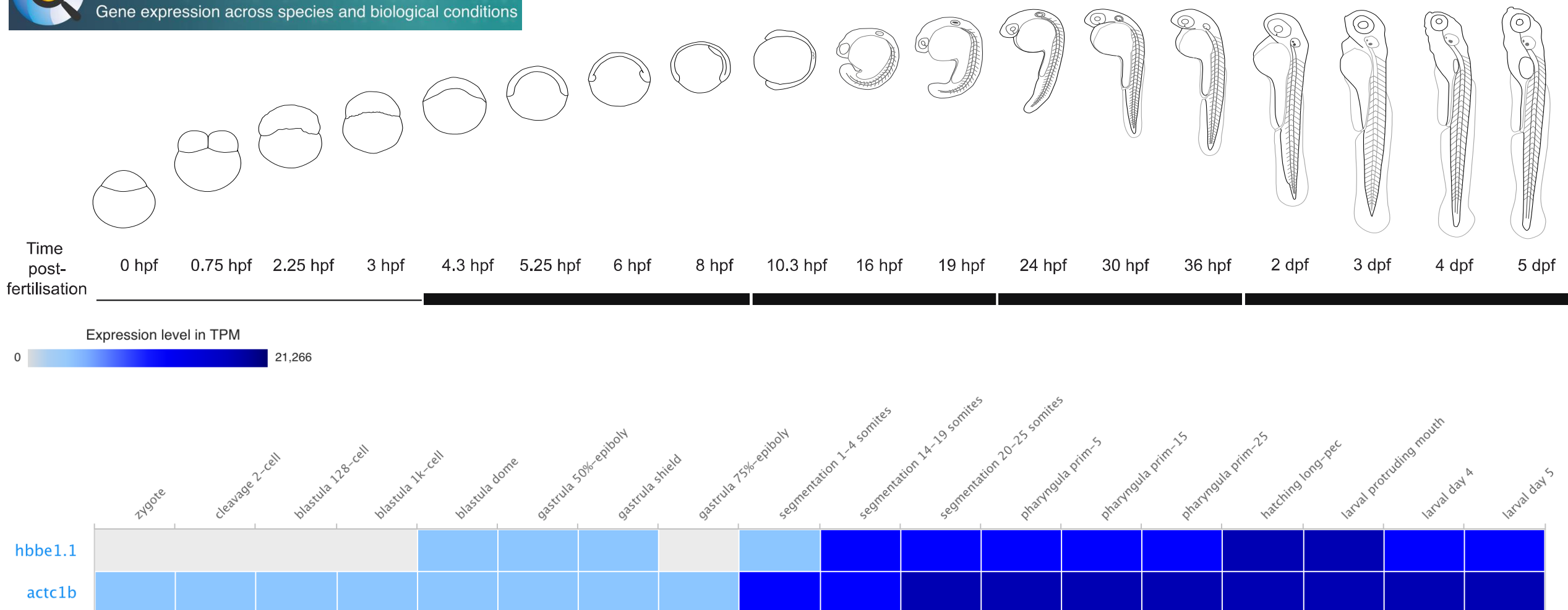
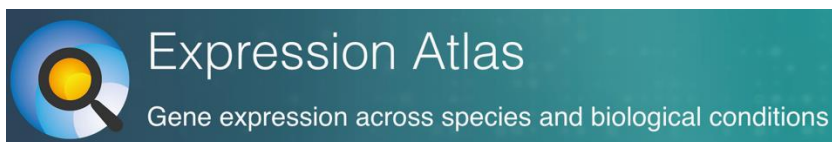
Log2FoldChange: For a given comparison, a positive fold change value indicates an increase of expression, while a negative fold change indicates a decrease in expression.

P-value: Indicates whether the gene analysed is likely to be differentially expressed in that comparison.

Adj. p-value: The p-value obtained for each gene above is re-calculated to correct for multiple testing (n genes).

https://biocorecrg.github.io/CRG_Bioinformatics_for_Biologists/differential_gene_expression.html

Comparing results with external resources/databases

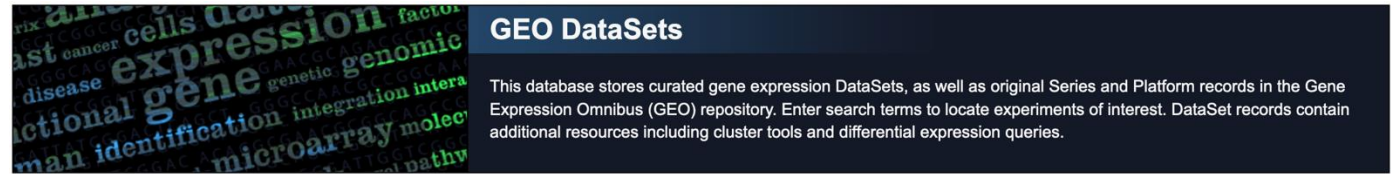


<https://www.ebi.ac.uk/gxa/experiments/E-ERAD-475/Results;>

<https://doi.org/10.7554/eLife.30860>

Other resources & databases

- Co-expression analysis



<https://www.ncbi.nlm.nih.gov/geo/>

- Protein-protein interaction analysis



STRING

<https://string-db.org/>

- Pathway enrichment analysis



reactome

<https://reactome.org/>

