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Spiking Neural Networks on EEG data processing

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Abstract

Among the methods of Brain-Computer Interface (BCI), Electroencephalogram (EEG) stands out as the most cost-effective and simple system to implement. However, it does have some quirks, such as high sensitivity to electromagnetic interference and difficulty in capturing the signal due to suboptimal scalp placement of the electrodes.

To address these issues, numerous Neural Networks (NNs) have been developed to handle such data. Despite their effectiveness, conventional NNs still require a significant amount of computational power for operation and training. This work proposes an alternative approach to this problem by utilizing Spiking Neural Networks (SNNs), which hypothetically consume less power. The aim is to assess the viability of SNNs in this context.

The results indicate a very close similarity in performance, demonstrating that SNNs are indeed a promising option for future research endeavors.

The sources used in this work are available in https://github.com/ensismoebius/deepLearnning)

Keywords: EEG, Spiking Neural Networks, SNN, Signal Processing

1. Introduction

Standard Neural Networks (NNs), as defined here—multilayered, fully connected, with or without recurrent or convolutional layers—require that all neurons are activated in both the forward and backward passes. This implies that every unit (neuron) in the network must process some data, leading to power consumption **CITATION HERE**.

In an era where responsible use of power resources is mandatory, coupled with the growing interest in Brain-Computer Interface (BCI) devices, a new problem arises: How to create NN models that are power-efficient both in usage and training, enabling their deployment even on portable devices?

In light of the considerations, the objective of this paper is to test if the current state of Spiking Neural Networks (SNNs) can be an effective method in the processing of Electroencephalogram (EEG) data.

In this work PUT THE NUMBER AND KINDS OF NEURAL NETWORKS CREATE HERE and use the PUT THE MEASUMENT USED HERE

The experiments and results described hereafter, relevantly complemented with tables and graphics lead to a set of interesting conclusions from the point of view of digital signal processing and intelligent systems. Thus, this piece of work provides a relevant contribution, supporting future investigations.

The remaining of this document is organized as follows: Section 2 presents a short literature review, Section 3 describes the proposed approach, Section 4 details the tests and the results and, lastly, Section 5 is dedicated to the conclusions that are followed by the references.

2. Short Review Notes

2.1. Brain-Computer Interface and EEG

Among the methods of Brain-Computer Interface (BCI), Electroencephalogram (EEG) stands out as the most cost-effective and simple system to implement. However, it does have some quirks, such as high sensitivity to electromagnetic interference and difficulty in capturing the signal due to suboptimal scalp placement of the electrodes. So one of the fundamental aspect that any EEG processing system must have is the tolerance to noise [1].

The *wet* electrodes are placed using conductive gel and are less prone to movement artifacts which are electromagnetic interferences caused by some motion like blinking. *Dry* electrodes does not need the gel but is more sensible to artifacts.

EEG records the electrical activities of the brain, typically placing along the scalp surface electrodes. These electrical activities result from ionic current flows induced by the synchronized synaptic activation of the brain's neurons. They manifest as rhythmic voltage fluctuations ranging from 5 to $100\mu V$ in amplitude and between 0.5 and 40 Hz in frequency[1].

The operational frequencies bands in the brain are following [2]:

- **Delta** (1–4Hz): The slowest and usually the highest amplitude waveform. The Delta band is observed in babies and during deep sleep in adults.
- Theta (4–8Hz): Observed in children, drowsy adults, and during memory recall. Theta wave amplitude is typically less than $100\mu V$.
- Alpha (8–12Hz): Usually the dominant frequency band, appearing during relaxed awareness or when eyes are closed. Focused attention or relaxation with open eyes reduces the amplitude of the Alpha band. These waves are normally less than 50 μV .
- **Beta** (12–25Hz): Associated with thinking, active concentration, and focused attention. Beta wave amplitude is normally less than $30 \,\mu V$.
- Gamma (over 25Hz): Observed during multiple sensory processing. Gamma patterns have the lowest amplitude.

According to [1], for most of the tasks brain do, there is regions associated with it. These ones are described in Table 1.

Region	Channels	Tasks
Frontal lobe	Fp1, Fp2, Fpz, Pz, F3, F7, F4, F8	Memory, concentration, emotions.
Parietal lobe	P3, P4, Pz	Problem Solving, attention, grammar, sense of
		touch.
Temporal lobe	T3, T5, T4, T6	Memory, recognition of faces, hearing, word
		and social clues.
Occipital lobe	O1, O2, Oz	Reading, vision.
Cerebellum		Motor control, balance.
Sensorimotor Cortex	C3, C4, Cz	Attention, mental processing, fine motor con-
		trol, sensory integration.

Table 1: Brain tasks and its corresponding regions. See Figure 1 for more information.

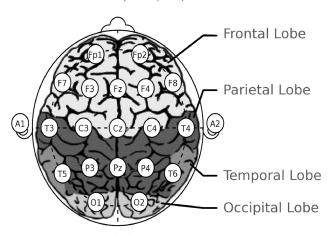


Figure 1: Electrodes placement according to 10-20 standard [3] and brain's lobes. Odd numbers are assigned to the electrodes on the left hemisphere, and even numbers are assigned to the electrodes on the right. Source [1]

2.2. Spiking Neural Networks

Neural Networks (NNs), as defined here as *a multilayered, fully connected, with or without recurrent or convolutional layers network*, require that all neurons are activated in both the forward and backward passes. This implies that every unit in the network must process some data, leading to power consumption **CITATION HERE**.

The sensory system of biological neurological systems converts external data, such as light, odors, touch, flavors, and others, into **spikes**. A spike is a voltage that convey information [4]. These ones are then transmitted along the neuronal chain to be processed, generating a response to the environment.

The biological neuron only spikes when a certain level of excitatory signals get accumulated above some threshold in its soma staying inactive when there is no signal, therefore, this type of cell is very efficient in terms of energy consumption and processing.

In order to get the aforementioned advantages the Spiking Neural Networks (SNNs) instead of employing continuous activation values, like NNs, SNNs utilize **spikes** at the input, hidden and outputs layers. SNNs can have continuous inputs as well and keep its properties.

An SNN **is not** a one-to-one simulation of neurons. Instead, it approximates certain computational capabilities of specific biological properties. Some studies like [6] created models way closer to natural neurons exploring the nonlinearity of dendrites and another neuron features yielding remarkable results in the classification.

As can be seen in Figure 2 SNNs neurons, given the correct parameters, are very noise tolerant. Generating spikes even when a considerably level of interference is present. This units are very time-oriented too, being great when it comes to process streams of data **CITATION HERE**.

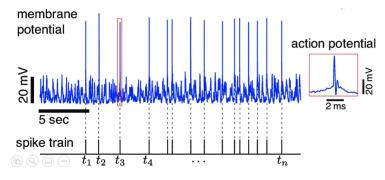


Figure 2: Spikes from a noisy signal. Source [7]

2.3. Spiking Neuron

Although this work focus in just *Leaky Integrate and Fire Neurons* (LIF) because is simpler, more efficient and currently generalize better for most of the problems [7], there is one interesting enough, because it paves the way to another ones, that worth to mention:

2.3.1. Hodgkin-Huxley neuron

The Hodgkin-Huxley neuron [8] was modeled conducting experiments on a giant axon of a squid and identified three distinct types of ion currents: Na^+ (sodium), K^+ (Potassium), and a leak current primarily composed of Cl^- (chloride) ions.

Like can be seen in Figure 3 this neuron is modeled as conjunction of a resistor R, potentiometers R_{Na} , R_K and a capacitor C along with batteries (ion concentrators) E_L , E_{Na} , E_K in which an current I is injected. The potentiometers indicate the channels of Na^+ and K^+ which can ben opened or closed, the batteries represents Nernst potential [9] for each channel.

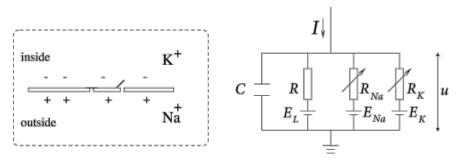


Figure 3: Schematic diagram for the Hodgkin-Huxley model. Source: [8]

Given that:

- C_m is the membrane capacitance that represents the ability of the neuronal membrane to store charge.
- g_{Na} is the sodium conductance that describes its fast opening permeability.
- g_K is the potassium conductance that describes its slow opening permeability.
- g_{leak} is the background conductance that describes ion movements not related to Na^+ or K^+ channels.
- *V* is the membrane potential.
- *I* is the injected current.
- *m*, *h*, and *n* are dimensionless gating variables representing the probability that the sodium and potassium channels are open.
- α and β are voltage-dependent rate constants.

Than the neuron's behavior is ruled by the Equations 1, 2, 3, 4 and Listings 1, 2, 3.

The equation 1 models how rate of change $\frac{dV}{dt}$ of membrane potential (V) is determined by the injected current (I) and the currents through Na^+ , K^+ , and g_{leaky} channels.

$$\frac{dV}{dt} = \frac{I - g_{\text{Na}} m^3 h (V - E_{\text{Na}}) - g_{\text{K}} n^4 (V - E_{\text{K}}) - g_{\text{leak}} (V - E_{\text{leak}})}{C_m} \tag{1}$$

```
# Equation 1
2 dVdt = (I_total - I_Na - I_K - I_leak) / Cm
```

Listing 1: Python implementation of the Hodgkin-Huxley neuron's first equation

Equations 2, 3, 4 are the rate of change of the gating variables m, h, and n, which represent the probability that the Na^+ and K^+ channels are open.

$$\frac{dm}{dt} = \alpha_m (1 - m) - \beta_m m \tag{2}$$

$$\frac{dh}{dt} = \alpha_h (1 - h) - \beta_h h \tag{3}$$

$$\frac{dn}{dt} = \alpha_n (1 - n) - \beta_n n \tag{4}$$

```
# Equations 2-4

dmdt = alpha_m(V) * (1 - m) - beta_m(V) * m

dhdt = alpha_h(V) * (1 - h) - beta_h(V) * h

dndt = alpha_n(V) * (1 - n) - beta_n(V) * n
```

Listing 2: Python implementation of the Hodgkin-Huxley neuron's second, third and fourth equation

Next are the helper functions representing the α and β . They are used to calculate the rate constants for the gating variables in the differential equations.

$$\alpha_m(V) = 0.1 \frac{V + 40.0}{1.0 - \exp\left(-\frac{V + 40.0}{10.0}\right)}$$
 (5)

$$\beta_m(V) = 4.0 \exp\left(-\frac{V + 65.0}{18.0}\right) \tag{6}$$

$$\alpha_h(V) = 0.07 \exp\left(-\frac{V + 65.0}{20.0}\right) \tag{7}$$

$$\beta_h(V) = \frac{1.0}{1.0 + \exp\left(-\frac{V + 35.0}{10.0}\right)} \tag{8}$$

$$\alpha_n(V) = 0.01 \frac{V + 55.0}{1.0 - \exp\left(-\frac{V + 55.0}{10.0}\right)}$$
(9)

$$\beta_n(V) = 0.125 \exp\left(-\frac{V + 65.0}{80.0}\right) \tag{10}$$

```
# Helper functions for rate constants

def alpha_m(V):
    return 0.1 * (V + 40.0) / (1.0 - math.exp(-(V + 40.0) / 10.0))

def beta_m(V):
    return 4.0 * math.exp(-(V + 65.0) / 18.0)

def alpha_h(V):
    return 0.07 * math.exp(-(V + 65.0) / 20.0)

def beta_h(V):
    return 1.0 / (1.0 + math.exp(-(V + 35.0) / 10.0))

def alpha_n(V):
    return 0.01 * (V + 55.0) / (1.0 - math.exp(-(V + 55.0) / 10.0))
```

```
17 def beta_n(V):
18 return 0.125 * math.exp(-(V + 65.0) / 80.0)
```

Listing 3: Helper functions representing the α and β

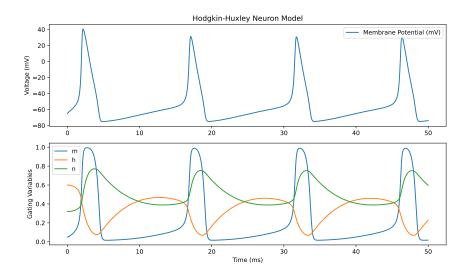


Figure 4: Hodgkin-Huxley neuron behavior under constant input current. Source: The author

And finally the full code that plots the Figure 4 which shows the neuron behavior:

```
import math
2 import matplotlib.pyplot as plt
4 # Hodgkin-Huxley model parameters
5 Cm = 1.0 # Membrane capacitance (uF/cm^2)
g_Na = 120.0 # Sodium conductance (mS/cm^2)
 g_K = 36.0
               # Potassium conductance (mS/cm^2)
g_leak = 0.3 # Leak conductance (mS/cm^2)
9 E_Na = 50.0
               # Sodium reversal potential (mV)
E_K = -77.0
              # Potassium reversal potential (mV)
E_{leak} = -54.387 # Leak reversal potential (mV)
# Time parameters
dt = 0.01 # Time step (ms)
15
 t_max = 50.0 # Maximum simulation time (ms)
# Initial conditions
18 V, m, h, n = -65.0, 0.05, 0.6, 0.32 # V, m, h, n
19
_{\rm 20} # Lists to store results for plotting
time_points = []
22 membrane_potential = []
23 gating_variables_m = []
gating_variables_h = []
25 gating_variables_n = []
27 # Helper functions for rate constants
28 def alpha_m(V):
   return 0.1 * (V + 40.0) / (1.0 - math.exp(-(V + 40.0) / 10.0))
30
def beta_m(V):
  return 4.0 * math.exp(-(V + 65.0) / 18.0)
32
```

```
34 def alpha_h(V):
   return 0.07 * math.exp(-(V + 65.0) / 20.0)
37 def beta_h(V):
   return 1.0 / (1.0 + math.exp(-(V + 35.0) / 10.0))
40 def alpha_n(V):
   return 0.01 * (V + 55.0) / (1.0 - math.exp(-(V + 55.0) / 10.0))
43 def beta_n(V):
   return 0.125 * math.exp(-(V + 65.0) / 80.0)
46 # Simulation loop
for t in range(int(t_max / dt)):
   # Membrane currents
48
   I_Na = g_Na * m**3 * h * (V - E_Na)
   I_K = g_K * n**4 * (V - E_K)
50
   I_leak = g_leak * (V - E_leak)
51
52
53
    # Total membrane current
    I_total = 10.0 # Injected current (adjust as needed)
54
    # Update variables using Euler method
56
57
    dVdt = (I_total - I_Na - I_K - I_leak) / Cm
    dmdt = alpha_m(V) * (1 - m) - beta_m(V) * m
58
    dhdt = alpha_h(V) * (1 - h) - beta_h(V) * h
59
60
    dndt = alpha_n(V) * (1 - n) - beta_n(V) * n
61
   V += dt * dVdt
    m += dt * dmdt
63
   h += dt * dhdt
64
   n += dt * dndt
66
   # Store results for plotting
67
   time_points.append(t * dt)
68
   membrane_potential.append(V)
69
70
    gating_variables_m.append(m)
    gating_variables_h.append(h)
71
72
    gating_variables_n.append(n)
74 # Plot results
plt.figure(figsize=(10, 6))
76 plt.subplot(2, 1, 1)
77 plt.plot(time_points, membrane_potential, label='Membrane Potential (mV)')
78 plt.title('Hodgkin-Huxley Neuron Model')
79 plt.ylabel('Voltage (mV)')
80 plt.legend()
82 plt.subplot(2, 1, 2)
plt.plot(time_points, gating_variables_m, label='m')
84 plt.plot(time_points, gating_variables_h, label='h')
85 plt.plot(time_points, gating_variables_n, label='n')
86 plt.xlabel('Time (ms)')
plt.ylabel('Gating Variables')
88 plt.legend()
90 plt.tight_layout()
plt.show()
```

Listing 4: Full implementation of a Hodkin-Huxley neuron

2.3.2. Leaky Integrate and Fire Neuron

The Leaky Integrate and Fire Neuron (LIF) is one of the simplest neuron models in SNNs, still, it can applied successfully to most of the problems in with SNNs can be used.

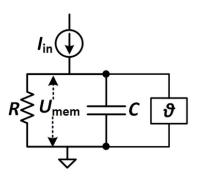


Figure 5: RC model: R is resistance to the leakage, I_{in} the input current, C the capacitance, U_{mem} means the accumulated action potential and v is a switch that lets the capacitor discharge (i.e. emit a spike) when some potential threshold is reached. Source: [5]

LIF, like a NN neuron takes the sum of weighted inputs but, rather than pass it directly to its activation function, some *leakage* is applied which descreases in some degree the sum. LIF behave much like Resistor-Capacitor circuits as can be seen in Figure 5.

Unlike the Hodgkin-Huxley neuron, spikes are represented as **sparsely** distributed **ones** in a train of **zeros**, as illustrated in Figure 6 and 7. This approach simplify the models and reduces the computational power and needed storage to run an SNN.

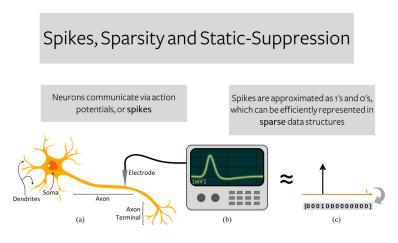


Figure 6: Sparsity on Spike Neuron Networks. Source: [5]

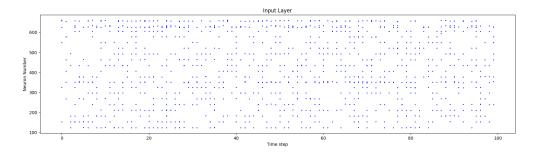


Figure 7: Sparse activity of a SNN: The horizontal axis represents the time step of some data being processed the vertical are the SNN neuron number. Note that, most of the time, very few neurons gets activated. Source: The author.

As result of the mentioned above, in SNN information is coded in format of *timing* and/or *rate* of spikes giving consequently great capabilities of processing streams of data but limiting the processing of static data.

LIF model is governed by the Equations.

$$C\frac{dV}{dt} = \frac{(E_{rest} - V + R.I_{syn} + R.I_{leak})}{\tau_m}$$
 (11)

$$V(t + \Delta t) = V(t) + \Delta t. \frac{dV}{dt}$$
(12)

$$C\frac{dV}{dt} = \frac{\text{(terms as shown)}}{\tau_m} \tag{13}$$

3. The Proposed Approach

- 3.1. Speech Signals Database
- 3.2. Database Organization
- 3.3. Proposed Approach Structure

4. Tests and Results

- 4.1. Metrics
- 4.2. Experiments
- 4.3. Complementary Tests

5. Conclusions

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