- 1 Supplementary File
- 2 A modeling approach for assessing ecological risks of neonicotinoid insecticides from
- 3 emission to non-target organisms: A case study of cotton plant
- 4 Zijian Li<sup>a</sup>\*, Minmin Li<sup>b</sup>, Shan Niu<sup>c</sup>
- <sup>a</sup> School of Public Health (Shenzhen), Sun Yat-sen University, Shenzhen, Guangdong, 518107,
- 6 China.

- 7 b Key Laboratory of Agro-products Quality and Safety Control in Storage and Transport Process,
- 8 Chinese Academy of Agricultural Sciences, Beijing 100000, China.
- 9 <sup>c</sup> Department of Civil & Environmental Engineering, University of Pittsburgh, Pittsburgh, PA
- 10 15260, United States.
- \*Corresponding information: ORCID: 0000-0002-9291-5966; Phone: +86 136 4430 2865;
- 12 Email: <u>lizijian3@mail.sysu.edu.cn</u>

## 14 Table of contents

- S1. Plant uptake model
- 16 S2. Insecticide dissipation kinetics in soil
- 17 S3. Chemical-specific model inputs
- 18 S4. Variability analysis
- 19 S4.1 Soil properties
- S4.2 Dissipation half-lives of insecticides in soil
- 21 S5. Supplementary figures
- 22 S6. Model refinement for site-specific assessment
- S6.1 Weather input variables
- S6.2 Exposure input variables
- S6.3 Toxicological input variables
- 26 References

### S1. Plant uptake model

- 28 The plant uptake model for simulating the residue in leaves, pollen, and nectar for the vegetative
- development and flowing stages was introduced by Li (2022). In this section, the main modeling
- 30 processes associated with the input values specified for cotton plants are provided.
- In the vegetative development stage, the modified one-compartment uptake model (Li, 2021;
- 32 Trapp and Matthies, 1995), including the transpiration (i.e., residue uptake from soil) and surface
- deposition (i.e., residue uptake via penetration through leaf cuticle) uptake routes (Fantke et al.,
- 34 2011), was applied to describe the uptake of neonicotinoid insecticides by cotton plants. The
- 35 uptake process can be expressed in the following equations.
- 36 The residue fate in the leaf compartment:

$$\frac{dC_{Leaf}(t)}{dt} = k_{Up,Soil}C_{Soil}(t) + k_{Up,Surface}C_{Surface}(t) - (k_{El,Air} + k_{El,Degra} + k_{El,Grow})C_{Leaf}(t)$$

$$\forall t \in [0, t_F)$$
(S1)

- 37 where,
- 38  $C_{Leaf}(t)$  (mg kg<sup>-1</sup>) is the residue level in the leaf as a function of time (t, d);
- 39  $C_{Soil}(t)$  (mg kg<sup>-1</sup>) is the residue level in soil (surface) as a function of t;
- 40  $C_{Surface}(t)$  (mg kg<sup>-1</sup>) is the residue level on the leaf surface as a function of t;
- 41  $k_{Up,Soil}$  (d<sup>-1</sup>) is the uptake rate constant of the residue in the leaf via transpiration from soil water;
- 42  $k_{Up,Surface}$  (d<sup>-1</sup>) is the uptake rate constant of the residue in the leaf via surface deposition;
- 43  $k_{El,Air}$  (d<sup>-1</sup>) is the elimination rate constant of the residue in the leaf via volatilization;

- 44  $k_{El,Degra}$  (d<sup>-1</sup>) is the elimination rate constant of the residue in the leaf via degradation, which was
- approximated using the dissipation half-life in plant matrix (i.e., RL50) (Lewis et al., 2016) due to
- 46 the information limitation;
- 47  $k_{El,Grow}$  (d<sup>-1</sup>) is the elimination rate constant of the residue in the leaf via dilution due to the plant
- 48 growth;
- 49 "t = 0" denotes the time when insecticide application occurs;
- 50  $t_F$  (d) denotes the time when the cotton plant enters the flowering stage.
- The residue fate in the soil compartment:

$$C_{Soil}(t) = C_{Soil}(0) \exp(-k_{Diss,Soil}t) \ \forall \ t \in [0, \infty]$$
 (s2a)

$$C_{Soil}(0) = \frac{AR \times f_{Soil} \times cf_1}{\rho_{Soil} \times H_{Soil} \times cf_2}$$
 (s2b)

- 52 where,
- 53  $\rho_{Soil}$  (kg L<sup>-1</sup>) is the bulk density of soil;
- 54  $f_{Soil}$  (unitless) is the mass fraction of the residue in soil (initial distribution);
- 55  $H_{Soil}$  (m) is the height of the surface soil;
- 56 cf is the unit conversion factor.
- 57 The residue fate in the leaf surface:

$$C_{Surface}(t) = C_{Surface}(0) \exp(-k_{Diss,Surface}t) \ \forall \ t \in [0, t_H]$$
 (s3a)

$$C_{Surface}(0) = \frac{AR \times f_{Surface} \times cf_1}{LAI \times LMA \times \frac{1}{1 - W_{Leaf}}}$$
(s3b)

58 where,

- 59  $k_{Diss,Surface}$  (d<sup>-1</sup>) is the dissipation rate constant of the residue on the leaf surface, which can be
- approximated as four times as the dissipation rate constant in soil (Juraske et al., 2008);
- 61  $f_{Surface}$  (unitless) is the mass fraction of the residue on the leaf surface (initial distribution);
- 62  $LAI \text{ (m}^2 \text{ m}^{-2}\text{)}$  is the leaf area index of the cotton plant;
- 63 LMA (kg m<sup>-2</sup>) is the leaf mass (dry) per area of the cotton plant;
- 64  $W_{Leaf}$  (g g<sup>-1</sup>) is the water content of the leaf.
- Then, Eq. (1) can be solved with the initial condition " $C_{Leaf}(0) = 0$ " as follows:

$$C_{Leaf}(t)$$

$$= \left(\frac{AR \times f_{Soil} \times cf_{1}}{\rho_{Soil} \times H_{Soil} \times cf_{2}}\right) \left(\frac{k_{Up,Soil}}{k_{El,Air} + k_{El,Degra} + k_{El,Grow} - k_{Diss,Soil}}\right) \left\{ \exp\left(-k_{Diss,Soil}t\right) - \exp\left[-\left(k_{El,Air} + k_{El,Degra} + k_{El,Grow}\right)t\right] \right\}$$

$$+ \left(\frac{AR \times f_{Surface} \times cf_{1}}{LAI \times LMA \times \frac{1}{1 - W_{Leaf}}}\right) \left(\frac{k_{Up,Surface}}{k_{El,Air} + k_{El,Degra} + k_{El,Grow} - k_{Diss,Surface}}\right) \left\{ \exp\left(-k_{Diss,Surface}t\right) - \exp\left[-\left(k_{El,Air} + k_{El,Degra} + k_{El,Grow}\right)t\right] \right\} \forall t \in [0, t_{F})$$

In the flowering stage, the flower compartment was integrated into the plant uptake model, and the simple partitioning method (i.e., the partitioning of the residue between pollen or nectar and phloem sap) was applied to simulate the residue levels in pollen and nectar (Li, 2022), which can be expressed as follows:

$$\frac{C_{Pollen}(t)}{C_{Phloem}(t)} = K_{P-Ph} \ \forall \ t \in [t_F, t_H]$$
 (s5a)

$$\frac{C_{Nectar}(t)}{C_{Phloem}(t)} = K_{N-Ph} \forall \ t \in [t_F, t_H]$$
 (s5b)

$$\frac{C_{Leaf}(t)}{C_{Phloem}(t)} = K_{L-Ph} \,\forall \, t \in [t_F, t_H]$$
(S5c)

70 where,

- $C_{Pollen}(t)$  (mg kg<sup>-1</sup>) is the residue level in pollen as a function of t;
- $C_{Nectar}(t)$  (mg kg<sup>-1</sup>) is the residue level in nectar as a function of t;
- $C_{Pholem}(t)$  (mg L<sup>-1</sup>) is the residue level in phloem sap as a function of t;
- $K_{P-Ph}$  (L kg<sup>-1</sup>) is the partition coefficient of the residue between pollen and phloem sap;
- $K_{N-Ph}$  (L kg<sup>-1</sup>) is the partition coefficient of the residue between nectar and phloem sap;
- $K_{L-Ph}$  (L kg<sup>-1</sup>) is the partition coefficient of the residue between leaf and phloem sap;
- $t_H$  (d) is the time at harvest, which equals the preharvest interval (PHI, d) of the neonicotinoid
- 78 insecticide.
- Therefore, RUDs of the neonicotinoid insecticide in pollen ( $RUD_{Pollen}$ ) and nectar ( $RUD_{Nectar}$ ) can be
- 80 expressed as follows:

$$RUD_{Pollen}(t)$$

$$= \left(\frac{K_{P-L} \times f_{Soil} \times cf_1}{\rho_{Soil} \times H_{Soil} \times cf_2}\right) \left(\frac{k_{Up,Soil}}{k_{El,Air} + k_{El,Degra} + k_{El,Grow} - k_{Diss,Soil}}\right) \left\{\exp\left(-k_{Diss,Soil}t\right)\right\}$$
(s6a)

$$-\exp[-(k_{El,Air} + k_{El,Degra} + k_{El,Grow})t]\}$$

$$+ \left(\frac{K_{P-L} \times f_{Surface} \times cf_{1}}{LAI \times LMA \times \frac{1}{1 - W_{Leaf}}}\right) \left(\frac{k_{Up,Surface}}{k_{El,Air} + k_{El,Degra} + k_{El,Grow} - k_{Diss,Surface}}\right) \left\{ \exp\left(-k_{Diss,Surface}t\right) \right\}$$

$$-\exp\left[-\left(k_{El,Air}+k_{El,Degra}+k_{El,Grow}\right)t\right]\right\} \forall \ t \in [t_F,t_H]$$

 $RUD_{Nectar}(t)$ 

$$= \left(\frac{K_{N-L} \times f_{Soil} \times cf_1}{\rho_{Soil} \times H_{Soil} \times cf_2}\right) \left(\frac{k_{Up,Soil}}{k_{El,Air} + k_{El,Degra} + k_{El,Grow} - k_{Diss,Soil}}\right) \left\{ \exp\left(-k_{Diss,Soil}t\right) \right\}$$
(s6b)

$$-\exp\bigl[-\bigl(k_{El,Air}+k_{El,Degra}+k_{El,Grow}\bigr)t\bigr]\bigr\}$$

$$+ \left(\frac{K_{N-L} \times f_{Surface} \times cf_{1}}{LAI \times LMA \times \frac{1}{1 - W_{Leaf}}}\right) \left(\frac{k_{Up,Surface}}{k_{El,Air} + k_{El,Degra} + k_{El,Grow} - k_{Diss,Surface}}\right) \left\{ \exp\left(-k_{Diss,Surface}t\right) \right\}$$

$$-\exp\bigl[-\bigl(k_{El,Air}+k_{El,Degra}+k_{El,Grow}\bigr)t\bigr]\bigr\}\,\forall\,\,t\in[t_F,t_H]$$

- 81 where,
- 82  $K_{P-L}$  (unitless) is the partition coefficient of the residue between pollen and leaf, which can be
- 83 calculated by dividing  $K_{P-Ph}$  by  $K_{L-Ph}$ ;
- 84  $K_{N-L}$  (unitless) is the partition coefficient of the residue between nectar and leaf, which can be
- 85 calculated by dividing  $K_{N-Ph}$  by  $K_{L-Ph}$ .
- Values of the non-chemical-specific model inputs are provided in Table S1. Calculation processes
- of the residue-bioconcentration-related (leaf) rate constants are provided in the following equations.
- For the uptake process via transpiration,  $k_{Up,Soil}$  can be estimated using the transpiration rate of the
- cotton plant and the transpiration stream conversion factor (Trapp and Matthies, 1995) as follows:

$$k_{Up,Soil} = \frac{Q_{Leaf} \times TSCF}{M_{Leaf} \times K_{SW}}$$
 (s7)

- 90 where,
- 91  $Q_{Leaf}$  (L d<sup>-1</sup>) is the transpiration rate of the single leaf;
- 92  $M_{Leaf}$  (kg) is the mass of a single leaf;
- 93 TSCF (unitless) is the transpiration stream conversion factor;
- 94  $K_{SW}$  (L kg<sup>-1</sup>) is the partition coefficient of the residue between soil and water.
- 95 As the transpiration of plant leaves is dependent on the relative humidity and temperature of air,
- 96  $Q_{Leaf}$  can be estimated using the weather-dependent model (Li, 2020) as follows:

$$Q_{Leaf}(T_A, RH_A) = \eta A_{Leaf} LAI \left[ \frac{2.50 \times 10^6 (31.145 - 0.1T_A)}{(T_A - 35.85)^2} + 610.78 \left( 1 - \frac{RH_A}{100} \right) \right] exp \left[ \frac{17.2 (T_A - 273.15)}{T_A - 35.85} \right]$$
(\$8)

97 where,

- 98  $\eta$  (L m<sup>-2</sup>d<sup>-1</sup>) is the transpiration rate coefficient, calculated from the vapor pressure deficit and the
- 99 stomatal resistances (Li, 2020; Li and Niu, 2021);
- 100  $A_{Leaf}$  (m<sup>2</sup>) is the area of the single cotton leaf;
- 101  $T_A$  (K) is the air temperature;
- 102  $RH_A$  (%) is the relative humidity of air.
- For a general estimate, the reference state (i.e., 298.15 K and 50 %) (Li and Niu, 2021) was applied
- to calculate the  $Q_{Leaf}$  value.
- For the uptake process via surface deposition,  $k_{Up,Surface}$  can be estimated, using the residue
- mobility in the leaf cuticle and the partitioning process of the residue between cuticle and water
- 107 (Fantke et al., 2011; Juraske et al., 2007) as follows:

$$k_{Up,Surface} = k_{Cuticle} K_{CW}$$
 (S9)

- where,
- $k_{Cuticle}$  (d<sup>-1</sup>) is the rate constant of the residue mobility in the leaf cuticle, which can be estimated
- using the molecular weight (MW, g mol<sup>-1</sup>) (Fantke et al., 2011; Juraske et al., 2007);
- 111  $K_{CW}$  (L L<sup>-1</sup>) is the partition coefficient of the residue between leaf cuticle and water, which can be
- estimated using the partition coefficient between octanol and water  $(K_{OW}, L L^{-1})$  (Fantke et al.,
- 113 2011; Juraske et al., 2007).
- For the elimination process via volatilization,  $k_{El,Air}$  can be estimated by modeling the one-side
- residue exchange with the air (Trapp and Matthies, 1995) as follows:

$$k_{El,Air} = \frac{g}{L_{Leaf}\rho_{Leaf}K_{LA}}$$
 (s10)

- 116 where,
- 117  $g (m d^{-1})$  is the gas conductance;
- 118  $L_{Leaf}$  (m) is the leaf thickness;
- 119  $\rho_{Leaf}$  (kg L<sup>-1</sup>) is the fresh leaf density;
- 120  $K_{LA}$  (L kg<sup>-1</sup>) is the partition coefficient of the residue between leaf and air.
- For the residue partitioning among plant tissues, the composition-based approach was applied to
- approximate the partition coefficients (Chiou et al., 2001; Trapp et al., 2007), of which the general
- formula can be expressed as follows:

$$K_{i-j} \approx \frac{1.22 Lip_i (K_{OW})^{0.77} + Carb_i K_{ch} + \frac{W_i}{\rho_{Water}}}{1.22 Lip_j (K_{OW})^{0.77} + Carb_j K_{ch} + \frac{W_j}{\rho_{Water}}}$$
(S11)

- where,
- 125  $K_{i-j}$  (L kg<sup>-1</sup>) is the partition coefficient of the residue between compartments i and j;
- 126 Lip (L kg<sup>-1</sup>) is the lipid content;
- 127 Carb (kg kg<sup>-1</sup>) is the carbohydrate content;
- 128  $K_{ch}$  (L kg<sup>-1</sup>) is the partition coefficient of the residue between carbohydrate and water;
- 129  $\rho_{Water}$  (kg L<sup>-1</sup>) is the density of water.
- For the residue partitioning between leaf and air,  $K_{LA}$  can be estimated according to the
- composition of the leaf (Trapp, 2007) as follows:

$$K_{LW} = \frac{W_{Leaf}}{\rho_{Water}} + Lip_{Leaf} \times \frac{(K_{OW})^{0.95}}{\rho_{Octanol}}$$
 (s12a)

$$K_{LA} = \frac{K_{LW}}{K_{AW}} \tag{s12a}$$

- 132 where,
- $Lip_{Leaf}$  (g g<sup>-1</sup>) is the lipid content of the leaf;
- $\rho_{Octanol}$  (kg L<sup>-1</sup>) is the density of octanol;
- $K_{LW}$  (L kg<sup>-1</sup>) is the partition coefficient of the residue between leaf and water.
- For the residue partitioning in soil environment,  $K_{SW}$  can be estimated according to the
- composition of soil (Paraíba and Kataguiri, 2008; Trapp et al., 2007) as follows:

$$K_{SW} = \frac{f_{OC} \times K_{OC} \times \rho_{Soil,dry} + f_{Water} + f_{Air} \times K_{AW}}{\rho_{Soil,wet}}$$
(S13)

- 138 where,
- $f_{oc}$  (L L<sup>-1</sup>) is the organic matter volumetric fraction of soil;
- $f_{water}$  (L L<sup>-1</sup>) is the water volumetric fraction of soil;
- $f_{Air}$  (L L<sup>-1</sup>) is the air volumetric fraction of soil;
- $K_{AW}$  (L L<sup>-1</sup>) is the partition coefficient of the residue between air and water;
- $K_{oc}$  (L kg<sup>-1</sup>) is the partition coefficient of the residue between organic matter and water;
- $\rho_{Soil,dry}$  (kg L<sup>-1</sup>) is the density of dry soil;
- $\rho_{Soil,wet}$  (kg L<sup>-1</sup>) is the density of wet soil;
- Values of the model inputs are provided in Table S1.
- Table S1. Summary of inputs of the plant uptake model.

Variables	Variables	Units	Values	References
Bulk density of soil (wet)	$ ho_{Soil}$	kg·L <sup>-1</sup>	1.5	Generic
Depth of surface soil	$H_{Soil}$	m	0.05	Default
Area of a single leaf	$A_{Leaf}$	m <sup>2</sup>	0.01	Estimated (Mukherjee et al., 2015).
Mass of a single leaf	$M_{Leaf}$	kg	8 × 10 <sup>-4</sup>	Estimated using the leaf area of 0.01 m <sup>2</sup> (Mukherjee et al., 2015).
Transpiration rate coefficient	η	L·m⁻²d⁻¹	0.0025	(Li, 2020; Li and Niu, 2021)
Leaf area index	LAI	m²⋅m⁻²	3.0	Estimated for 50 days after germination (Monteiro et al., 2006).
Leaf mass (dry) per area	LMA	kg·m <sup>-2</sup>	0.1	Estimated (Meng et al., 2021).
Thickness of leaf	$L_{Leaf}$	m	0.0002	Estimated (Meng et al., 2021).
Mass fraction of residues on soil after application	f <sub>Soil</sub>	unitless	0.223	Estimated based on the capture coefficient of 0.5 and LAI of 3.0 m <sup>2</sup> ·m <sup>-2</sup> (Fantke et al., 2011; Juraske et al., 2007).
Mass fraction of residues on leaf surfaces after application	fsurface	unitless	0.612	Estimated based on the fraction of the loss to air of 16.5% and $f_{Soil}$ (Fantke et al., 2011; Juraske et al., 2007).
Water content of leaf	$W_{Leaf}$	kg·kg <sup>-1</sup>	0.8	Default
Lipid content of leaf	$Lip_{Leaf}$	kg·kg-1	0.02	Default (Trapp and Matthies, 1995)
Carbohydrate content of leaf	$Carb_{Leaf}$	kg·kg <sup>-1</sup>	0.1	Default (Wilson and Mannetje, 1978)
Water content of nectar	$W_{Nectar}$	kg·kg <sup>-1</sup>	0.7	Default
Lipid content of nectar	$Lip_{Nectar}$	kg·kg <sup>-1</sup>	0.0	Default
Carbohydrate content of nectar	$Carb_{Nectar}$	kg·kg <sup>-1</sup>	0.3	Default
Water content of pollen	$W_{Pollen}$	kg·kg <sup>-1</sup>	0.13	Estimated (Human and Nicolson, 2006)
Lipid content of pollen	$Lip_{Pollen}$	kg·kg <sup>-1</sup>	0.087	Estimated by converting from the dry mass fraction of 0.1 (Human and Nicolson, 2006).
Carbohydrate content of pollen	$Carb_{Pollen}$	kg·kg <sup>-1</sup>	0.75	Estimated (Human and Nicolson, 2006).
Volumetric fraction of organic matter in soil	$f_{oc}$	unitless	0.023	(Trapp and Matthies, 1995).
Volumetric fraction of water in soil	fwater	unitless	0.35	(Trapp and Matthies, 1995).
Volumetric fraction of air in soil	$f_{Air}$	unitless	0.12	(Trapp et al., 2007)

Density of dry soil	$ ho_{Soil,dry}$	kg·L <sup>-1</sup>	1.6	(Trapp et al., 2007)
Density of wet soil	$ ho_{Soil,wet}$	kg·L <sup>-1</sup>	1.95	(Trapp et al., 2007)
Density of leaf	$ ho_{Leaf}$	kg·L <sup>-1</sup>	0.4	Estimated (Meng et al., 2021).
Density of water	$ ho_{Water}$	kg·L <sup>-1</sup>	1.0	Default
Density of octanol	$ ho_{Octanol}$	kg·L <sup>-1</sup>	0.83	(U.S. National Library of Medicine, 2018)
Gas conductance	g	m·d⁻¹	86.4	(Trapp and Matthies, 1995)
Duration between insecticide application and the flowering stage	$t_F$	d	7	Default
Duration between insecticide application and harvest	$t_H$	d	50	Default (estimated according to the 6-wk flowering stage).
Elimination rate constant via dilution (plant growth)	$k_{El,Degra}$	d <sup>-1</sup>	0.035	Default value for the plant leaf (Trapp and Matthies, 1995)
Unit conversion factor 1	$cf_1$	mg·kg <sup>-1</sup> per m <sup>2</sup> ·ha	100	Calculated
Unit conversion factor 2	$cf_2$	L m <sup>-3</sup>	1000	Calculated

### S2. Insecticide dissipation kinetics in soil

The overall dissipation of the neonicotinoid insecticide from the surface soil comprises the volatilization, plant uptake, water-induced (i.e., rainfall), and degradation processes, which can be described using the first-order kinetics because the loss rates of the insecticide via these processes are proportional to the residue level in soil (Li and Niu, 2021). Thus,  $k_{Diss,Soil}$  can be further expressed as follows:

$$k_{Diss,Soil} = k_{Diss,Air} + k_{Diss,Plant} + k_{Diss,Water} + k_{Diss,Deg}$$
(S14)

where,

 $k_{Diss,Air}$  (d<sup>-1</sup>) is the dissipation rate constant of the residue via volatilization;

 $k_{Diss,Plant}$  (d<sup>-1</sup>) is the dissipation rate constant of the residue via plant uptake;

- $k_{Diss,Water}$  (d<sup>-1</sup>) is the dissipation rate constant of the residue via surface runoff and infiltration of
- the precipitation;
- 160  $k_{Diss,Deg}$  (d<sup>-1</sup>) is the dissipation rate constant of the residue via degradation.
- 161 The estimation of rate constants for the sub-dissipation processes were taken from (Li and Niu,
- 162 2021). For the volatilization process,  $k_{Diss,Air}$  can be estimated as follows:

$$k_{Diss,Air} = \frac{D_{Air}^0}{K_{SA} \times \rho_{Soil}^* \times d^2}$$
 (s15)

- where,
- $D_{Air}^{0}$  (m<sup>2</sup> d<sup>-1</sup>) is the diffusivity of the residue in gas under the reference state (298.15 K), which can
- be estimated using the MW of the residue and the diffusivity of H<sub>2</sub>O vapor in air (Trapp, 2007);
- 166  $K_{SA}$  (L L<sup>-1</sup>) is the partition coefficient of the residue between soil and air, which can be estimated
- using  $K_{SW}$  and  $K_{AW}$ ;
- 168  $\rho_{Soil}^*$  is the dimensionless bulk density of soil (wet);
- d (m) is the effective diffusion length in air (i.e., air boundary layer).
- For the plant uptake process,  $k_{Diss,Plant}$  can be estimated as follows:

$$k_{Diss,Plant} = \frac{Q_{Leaf} \times LAI \times 0.001}{A_{Leaf} \times H_{Soil} \times \rho_{Soil} \times K_{SW}}$$
(s16)

For the water-induced dissipation process,  $k_{Diss,Water}$  can be estimated as follows:

$$k_{Diss,Water} = \frac{I}{H_{Soil} \times \rho_{Soil} \times K_{SW}}$$
 (s17)

- where,
- 173 I (m d<sup>-1</sup>) is the average rainfall intensity.

For the degradation process,  $k_{Diss,Air}$  can be estimated as follows: 174

$$k_{Diss,Deg} = k_{Diss,Deg}^{0} \times \exp\left[\frac{E_a}{R} \times \left(\frac{1}{298.15K} - \frac{1}{T_{Soil}}\right)\right]$$
 (s18)

where, 175

- $k_{Diss,Deg}^{0}$  (d<sup>-1</sup>) is the degradation rate constant in the reference state (i.e., 298.15 K), which was 176
- taken from the USEtox database (Fantke et al., 2017); 177
- $E_a$  (kJ mol<sup>-1</sup>) is the activation energy; 178
- R (kJ mol<sup>-1</sup>K<sup>-1</sup>) is the gas constant; 179
- $T_{Soil}$  (K) is the soil temperature. 180
- The RUD<sub>Soil</sub>(0) value can be estimated according to Eq. (s2b) as follows: 181

$$RUD_{Soil}(0) = \frac{C_{Soil}(0)}{AR} = \frac{f_{Soil} \times cf_1}{\rho_{Soil} \times H_{Soil} \times cf_2}$$
(S19)

- For a general estimate of the  $k_{Diss,Soil}$  value of the neonicotinoid insecticide, default or generic 182 values were applied to simulate rate constants of the sub-dissipation processes, which are provided 183 in the following table.
- Table S2. Summary of inputs of the dissipation kinetics model. 185

Variables	Variables	Units	Values	References
Soil temperature	$T_{Soil}$	K	298.15 In the reference state	
Gas constant	R	kJ mol <sup>-</sup> <sup>1</sup> K <sup>-1</sup>	8.314	Default
Activation energy	$E_a$	kJ mol <sup>-1</sup>	60	Default (due to information limitation) (Li and Niu, 2021).
Average rainfall intensity	I	m d <sup>-1</sup>	0.0027	Default (estimated based on an average annual rainfall intensity of 1,000 mm yr <sup>-1</sup> ) (Li and Niu, 2021).
Effective diffusion length in air	d	m	0.01	Default (Wolters et al., 2003).

## S3. Chemical-specific model inputs

The chemical-specific model inputs, including partition coefficients and rate constants, are provided in the Supplementary database. The basic physicochemical properties (e.g., K<sub>OW</sub>) were taken from current studies (Fantke et al., 2017; Lewis et al., 2016).

### S4. Variability analysis

#### **S4.1 Soil properties**

Soil properties have a significant impact on the estimated  $\kappa_{SW}$  value of the insecticide residue, which further affect the simulated residue levels in plant tissues and soil. We varied soil properties shown in Table S3 to generate uncertainty intervals of  $\kappa_{SW}$ . The selected ranges of soil properties were taken from current studies (Guo et al., 2021). The simulation results are provided in the Supplementary database.

Table S3. Selected ranges of soil properties for estimating uncertainty intervals of K<sub>SW</sub>.

Input variables	Unit	Range	References
$f_{oc}$	g g <sup>-1</sup>	0.007-0.063	(Sudduth and Hummel, 1991)
$f_{Water}$	g g <sup>-1</sup>	0.2-0.4	(Datta et al., 2008)
$f_{Air}$	g g <sup>-1</sup>	0.2-0.3	(The University of Hawaii, 2021)
$ ho_{Soil,wet}$	kg L <sup>-1</sup>	1.1-1.6	(Rai et al., 2017)

### S4.2 Dissipation half-lives of insecticides in soil

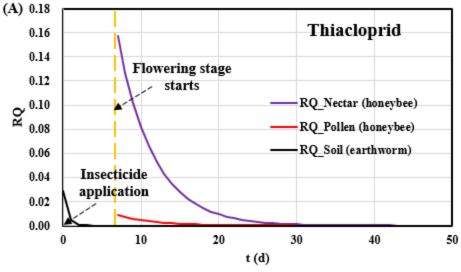
The  $k_{Diss,Soil}$  value of the insecticide is highly dependent on weather conditions (i.e., temperature and relative humidity), which further has the spatiotemporal pattern (Li and Niu, 2021). We estimated the weather-dependent  $k_{Diss,Soil}$  value by investigating temperature- or humidity-based sub-dissipation processes of the residue in soil.

For the volatilization process,  $k_{Diss,Air}$  is a function of  $T_{Air}$  and  $RH_{Air}$  (Davie-Martin et al., 2015, 2013), for which the full model can be found in (Li and Niu, 2021). For the plant uptake process,  $k_{Diss,Plant}$  is a function of  $T_{Air}$  and  $RH_{Air}$ , for which the weather-dependent value can be estimating using Eq. (s8). For the water-induced dissipation process,  $k_{Diss,Water}$  is a function of precipitation intensity, and the spatiotemporal pattern and uncertainty interval of the value can be estimated by varying the I value in Eq. (s17). For the degradation process,  $k_{Diss,Air}$  is a function of  $T_{Soil}$  that has a linear relationship with  $T_{Air}$  (Islam et al., 2015).

We used annual average values to evaluate the variability and spatiotemporal pattern of the simulated RQs of neonicotinoid insecticides for honeybees and earthworms, for which the simulation results are provided in the Supplementary database.

### S5. Supplementary figures

Supplementary figures are provided in this section.



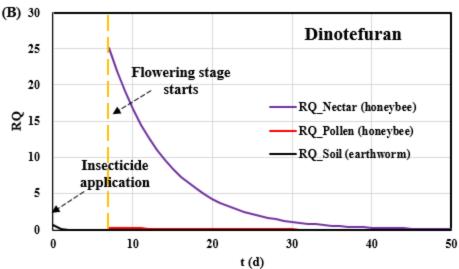
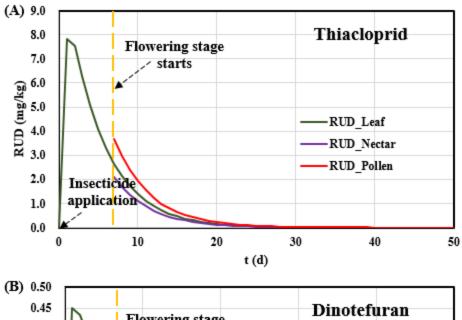


Figure S1. Simulated risk quotients (RQs) of (A) thiacloprid and (B) dinotefuran for honeybees (dietary intake of nectar and pollen) and earthworms (being exposed to soil) as functions of time (t, d) after insecticide application (t = 0). The insecticide application is assumed to occur one week before the flowering stage of the cotton plant. The RQs were simulated based on the unit application rate of the insecticide  $(1.0 \text{ kg ha}^{-1})$  to the cotton plant.



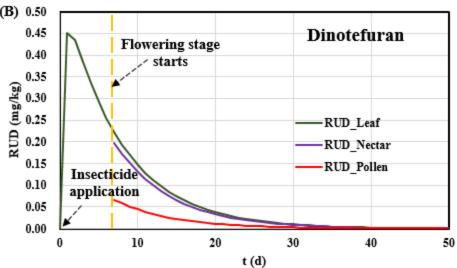


Figure S2. Simulated residue unit doses (RUDs, mg kg<sup>-1</sup>) of (A) thiacloprid and (B) dinotefuran in plant tissues (leaf, nectar, and pollen) of cotton as functions of time (t, d). The insecticide application is assumed to occur one week before the flowering stage of the cotton plant.

# Overall dissipation rate constant in soil

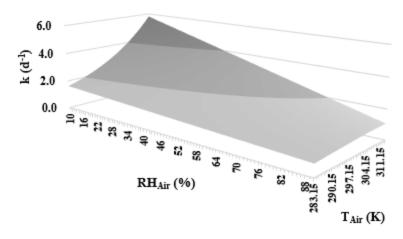
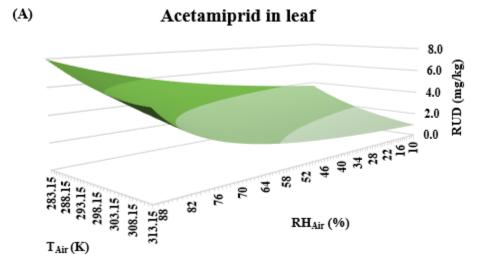
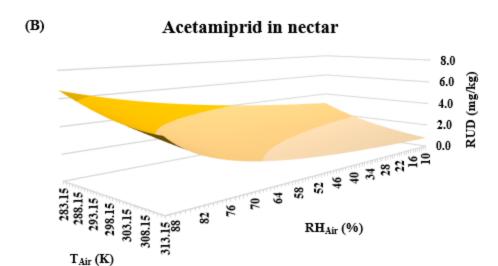
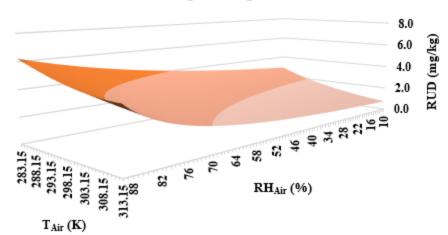


Figure S3. Simulated overall dissipation rate constant of acetamiprid in soil ( $k_{Diss,Soil}$ , d<sup>-1</sup>) as a function of the relative humidity of air ( $RH_{Air} \in [10\%, 90\%]$ ) and the temperature of air ( $T_{Air} \in [283.15K, 313.15K]$ ). A default value of 1,000 mm yr<sup>-1</sup> (0.0027 m d<sup>-1</sup>) was used for the average precipitation intensity.





(C) Acetamiprid in pollen



(D) Acetamiprid in soil

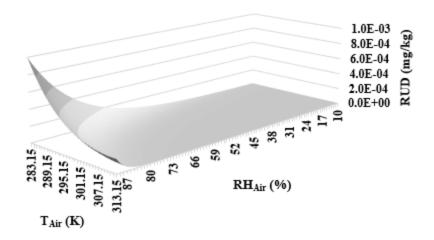


Figure S4. Simulated residue unit doses (RUDs, mg kg<sup>-1</sup>) (based on the insecticide application rate of 1.0 kg ha<sup>-1</sup> via foliar spray) of acetamiprid in (A) cotton leaves, (B) nectar, (C) pollen, and (D) soil as functions of the relative humidity of air ( $RH_{Air} \in [10\%, 90\%]$ ) and the temperature of air ( $T_{Air} \in [283.15K, 313.15K]$ ). The RUDs were simulated on 7 d after the insecticide application to the cotton plant. A default value of 1,000 mm yr<sup>-1</sup> (0.0027 m d<sup>-1</sup>) was used for the average precipitation intensity.

#### S6. Model refinement for site-specific assessment

### **S6.1** Weather input variables

For weather conditions, users can modify the values for weather input variables (i.e., temperature, relative humidity, or rainfall intensity) in sheet "NNIs - LBs of RQs (weather)" or "NNIs - UBs of RQs (weather)" of the Supplementary Database in order to generate site-specific simulation results. In Europe, cotton is typically grown in the south, where the average temperature during cotton-growing seasons ranges from 20°C to 30°C (i.e., from 293.15K to 303.15K). Consequently, the site-specific risk assessment for these regions of southern Europe can be conducted using the following procedures:

In the "NNIs - LBs of RQs (weather)" sheet of the Supplementary File, users can enter 293.15K in the "temperature" cell and leave other weather input variables at their default or generic values (e.g., relative humidity as 50% and rainfall intensity as 0.0027 m d-1) as depicted in the screenshot below.

	X	Υ	Z	AA	AB
		Tempreture	Relative humidity	Rainfall intensity	
1)	k <sup>0</sup> <sub>Diss,Deg</sub>				$\mathbf{k}_{\mathrm{Diss,Deg}}$
	đ⁻¹	K	%	m/đ	đ⁻¹
)	4.33E-01	293.15	50.00	2.70E-03	4.33E-01
2	1.27E-03	293.15	50.00	2.70E-03	1.27E-03
1	8.45E-03	293.15	50.00	2.70E-03	8.45E-03
)	2.24E-01	293.15	50.00	2.70E-03	2.24E-01
2	3.63E-03	293.15	50.00	2.70E-03	3.63E-03
L	7.88E-01	293.15	50.00	2.70E-03	7.87E-01
1	1.39E-02	293.15	50.00	2.70E-03	1.39E-02
U	Bs of Ksw	NNIs - I	Bs of RQs	(weather)	NNIs -

262

263

Screenshot 1. Illustration of modifying weather input variables (temperature: 293.15K) for sitespecific assessment.

Then, as shown in the screenshot below, users can have the simulated ecological risks in their respective cells.

	AO	AP	AQ	AR	AS	AT	AU	AV
r )		Aerial ap	pliaction			Risk assessm	ent (based on	l kg per ha)
	RUD <sub>Leaf</sub>	RUD <sub>Nectar</sub>	RUD <sub>Pollen</sub>	RUD <sub>Soil</sub>		RQ <sub>Nectar</sub>	RQ <sub>Pollen</sub>	RQ <sub>Soil</sub>
H	mg.kg <sup>-1</sup>	mg.kg <sup>-1</sup>	mg.kg <sup>-1</sup>	mg.kg <sup>-1</sup>				-
П	3.50E+00	2.89E+00	2.73E+00	2.31E-06		1.04E+00	3.21E-02	2.56E-06
П	8.89E+00	7.18E+00	7.63E+00	9.08E-05		4.76E+02	1.65E+01	6.87E-05
П	2.83E-01	2.53E-01	8.48E-02	4.95E-07		3.21E+01	3.50E-01	1.01E-06
П	3.44E-01	3.04E-01	1.14E-01	1.73E-07		8.87E-03	1.08E-04	1.73E-09
П	2.68E+00	2.30E+00	1.73E+00	1.28E-05		8.28E+01	2.03E+00	1.19E-05
П	3.06E+00	2.41E+00	4.24E+00	3.60E-06		1.81E-01	1.04E-02	3.42E-07
	5.56E-01	4.90E-01	1.94E-01	9.37E-07		5.96E+01	7.68E-01	9.37E-09
Ц								
Ц								
Ц								
Ц								
Ц								
Ц								
Ц								
	NNIs - UBs of Ksw NNIs - LBs of RQs (weather) NNIs - UBs of RQs (weather)							

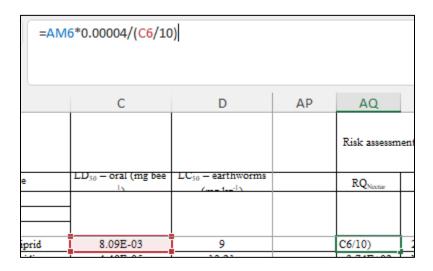
Screenshot 2. Illustration of simulated ecological risks by varying weather input variables (temperature: 293.15K).

The same method can be used to generate simulation results at a temperature of 303.15K.

### **S6.2** Exposure input variables

The operational tool proposed allows users to modify exposure input variables (i.e., honeybee daily nectar or pollen intake rates) to perform species-specific risk assessment for honeybees. The aforementioned sequential procedure can be utilized by users to conduct the simulation. Users can modify the numeric values in the mathematical formulas of the cells "RQ Nectar" and "RQ Pollen" to determine their specific honeybee daily nectar and pollen consumption rates, respectively. The simulation process is depicted in the diagram below.

If the nectar consumption rate is set to 40 mg/bee/d (i.e., 0.00004 kg/bee/d) rather than 292 mg/bee/d (i.e., 0.000292 kg/bee/d) as stated in the main text, then users can enter the numeric value 0.00004 in the mathematical formula of the cell "RQ Nectar" in the sheet "NNIs - default values" (or any calculation sheet), as shown in the screenshot below.



Screenshot 3. Illustration of modifying exposure input variables for site-specific assessment.

Then, users can obtain the simulated ecological risks (i.e., RQ Nectar) corresponding to the modified nectar consumption rates, as depicted in the following screenshot.

0	AQ	AR	AS	AT
	Risk assessm	ent (based on	l kg per ha)	
	RQ <sub>Nectar</sub>	RQ <sub>Pollen</sub>	RQ <sub>Soil</sub>	
	1.18E-01	2.65E-02	2.71E-07	
	5.12E+01	1.29E+01	8.58E-06	
	3.44E+00	2.74E-01	3.34E-08	
	9.68E-04	8.64E-05	6.41E-11	
	8.78E+00	1.57E+00	9.05E-07	
	2.16E-02	9.01E-03	7.67E-08	
	6.35E+00	5.97E-01	3.69E-10	

Screenshot 4. Illustration of simulated ecological risks by modifying exposure input variables.

## **S6.3 Toxicological input variables**

In this study, a 10 uncertainty factor was used to account for inter- or intra-species variability; however, this value may be an overestimate or underestimate of the actual toxicity of pesticides to non-target organisms. In order to conduct a more accurate assessment of the ecological risk, the toxicological data should be updated. Users can modify the uncertainty factor and toxicological data for honeybees or earthworms in the spreadsheet. As shown in the screenshot below, users can enter an uncertainty factor of 5, rather than 10, in the mathematical formula tabulated for the cell "RQ."

=AO	6/(D6/5)						
	С	D	AP	AQ	AR	AS	
				Risk assessm	ent (based on	l kg per ha)	
	LD <sub>50</sub> – oral (mg bee	LC <sub>50</sub> — earthworms		RQ <sub>Nectar</sub>	RQ <sub>Pollen</sub>	RQ <sub>Soil</sub>	
							L
	1						$\vdash$
prid	8.09E-03	9		1.18E-01	2.65E-02	5)	Ţ
idin	4.40E-05	13.21		5.12E+01	1.29E+01	8.58E-06	
ran	2.30E-05	4.9		3.44E+00	2.74E-01	3.34E-08	
nid	1.00E-01	1000		9.68E-04	8.64E-05	6.41E-11	

Screenshot 5. Illustration of modifying toxicological input variables for species-specific risk assessment.

#### 297 References

- 298 Chiou, C.T., Sheng, G., Manes, M., 2001. A partition-limited model for the plant uptake of organic
- contaminants from soil and water. Environmental Science and Technology.
- 300 https://doi.org/10.1021/es0017561
- Datta, S., Taghvaeian, S., Stivers, J., 2008. Understanding Soil Water Content and Thresholds for
- 302 Irrigation Management [WWW Document]. URL https://extension.okstate.edu/fact-
- 303 sheets/understanding-soil-water-content-and-thresholds-for-irrigation-management.html (accessed
- 304 5.26.21).
- Davie-Martin, C.L., Hageman, K.J., Chin, Y.P., 2013. An improved screening tool for predicting
- volatilization of pesticides applied to soils. Environmental Science and Technology.
- 307 https://doi.org/10.1021/es3020277
- Davie-Martin, C.L., Hageman, K.J., Chin, Y.P., Rougé, V., Fujita, Y., 2015. Influence of Temperature,
- Relative Humidity, and Soil Properties on the Soil-Air Partitioning of Semivolatile Pesticides:
- Laboratory Measurements and Predictive Models. Environmental Science and Technology.
- 311 https://doi.org/10.1021/acs.est.5b02525
- Fantke, P., Charles, R., Alencastro, L.F. de, Friedrich, R., Jolliet, O., 2011. Plant uptake of pesticides and
- 313 human health: Dynamic modeling of residues in wheat and ingestion intake. Chemosphere.
- 314 https://doi.org/10.1016/j.chemosphere.2011.08.030
- Fantke, P. (Ed.), Bijster, M., Guignard, C., Hauschild, M., Huijbregts, M., Jolliet, O., Kounina, A.,
- Magaud, V., Margni, M., McKone, T.E., Posthuma, L., Rosenbaum, R.K., van de Meent, D., van
- Zelm, R., 2017. USEtox® 2.0 Documentation (Version 1).
- 318 Guo, Y., Xiao, S., Li, Z., 2021. A screening model for managing periodic pesticide application in
- residential lawn soils. Environmental Advances. https://doi.org/10.1016/j.envadv.2021.100078
- Human, H., Nicolson, S.W., 2006. Nutritional content of fresh, bee-collected and stored pollen of Aloe
- greatheadii var. davyana (Asphodelaceae). Phytochemistry 67.
- 322 https://doi.org/10.1016/j.phytochem.2006.05.023
- 323 Islam, K.I., Khan, A., Islam, T., 2015. Correlation between Atmospheric Temperature and Soil
- Temperature: A Case Study for Dhaka, Bangladesh. Atmospheric and Climate Sciences.
- 325 https://doi.org/10.4236/acs.2015.53014
- Juraske, R., Antón, A., Castells, F., 2008. Estimating half-lives of pesticides in/on vegetation for use in
- multimedia fate and exposure models. Chemosphere.
- 328 https://doi.org/10.1016/j.chemosphere.2007.08.047
- Juraske, R., Antón, A., Castells, F., Huijbregts, M.A.J., 2007. Human intake fractions of pesticides via
- greenhouse tomato consumption: Comparing model estimates with measurements for Captan.
- Chemosphere. https://doi.org/10.1016/j.chemosphere.2006.11.047
- Lewis, K.A., Tzilivakis, J., Warner, D.J., Green, A., 2016. An international database for pesticide risk
- assessments and management. Human and Ecological Risk Assessment.
- 334 https://doi.org/10.1080/10807039.2015.1133242

- Li, Z., 2022. Modeling pesticide residues in nectar and pollen in support of pesticide exposure assessment
- for honeybees: A generic modeling approach. Ecotoxicology and Environmental Safety Accepted.
- Li, Z., 2021. Approximate Modeling of the Uptake of Pesticides by Grass for Grazing Risk Assessment
- and Pasture Management. ACS Agricultural Science & Technology.
- https://doi.org/10.1021/acsagscitech.1c00036
- Li, Z., 2020. Spatiotemporal pattern models for bioaccumulation of pesticides in common herbaceous and
- woody plants. Journal of Environmental Management.
- 342 https://doi.org/10.1016/j.jenvman.2020.111334
- Li, Z., Niu, S., 2021. Modeling pesticides in global surface soils: Evaluating spatiotemporal patterns for
- 344 USEtox-based steady-state concentrations. Science of The Total Environment 791.
- 345 https://doi.org/10.1016/j.scitotenv.2021.148412
- Meng, H., Lei, Z., Zhang, W., Zhang, Y., 2021. Variation mechanism of leaf mass per area in cotton
- under the systematic regulation. Cotton Science 33, 144–154.
- Monteiro, J.B.E.A., Sentelhas, P.C., Chiavegato, E.J., 2006. Microclimate and ramulosis occurrence in a
- cotton crop under three plant population densities in southern Brazil. AgriScientia 23.
- 350 https://doi.org/10.31047/1668.298x.v23.n2.2691
- 351 Mukherjee, T., Ivanova, M., Dagda, M., Kanayama, Y., Granot, D., Holaday, A.S., 2015. Constitutively
- overexpressing a tomato fructokinase gene (LeFRK1) in cotton (Gossypium hirsutum L. cv. Coker
- 353 312) positively affects plant vegetative growth, boll number and seed cotton yield. Functional Plant
- 354 Biology 42. https://doi.org/10.1071/FP15035
- Paraíba, L.C., Kataguiri, K., 2008. Model approach for estimating potato pesticide bioconcentration
- factor. Chemosphere. https://doi.org/10.1016/j.chemosphere.2008.07.026
- Rai, R.K., Singh, V.P., Upadhyay, A., 2017. Chapter 17 Soil Analysis, in: Planning and Evaluation of
- 358 Irrigation Projects.
- 359 Sudduth, K.A., Hummel, J.W., 1991. EVALUATION OF REFLECTANCE METHODS FOR SOIL
- 360 ORGANIC MATTER SENSING. Transactions of the ASAE 34, 1900–1909.
- The University of Hawaii, 2021. Soil Composition [WWW Document]. URL
- https://www.ctahr.hawaii.edu/mauisoil/a comp.aspx (accessed 5.26.21).
- Trapp, S., 2007. Fruit tree model for uptake of organic compounds from soil and air. SAR and QSAR in
- Environmental Research. https://doi.org/10.1080/10629360701303693
- Trapp, S., Cammarano, A., Capri, E., Reichenberg, F., Mayer, P., 2007. Diffusion of PAH in potato and
- carrot slices and application for a potato model. Environmental Science and Technology.
- 367 https://doi.org/10.1021/es0624180
- 368 Trapp, S., Matthies, M., 1995. Generic One-Compartment Model for Uptake of Organic Chemicals by
- Foliar Vegetation. Environmental Science and Technology. https://doi.org/10.1021/es00009a027
- 370 U.S. National Library of Medicine, 2018. National Center for Biotechnology Information Chemicals &
- 371 Bioassays [WWW Document].

372 373 374	Wilson, J.R., Mannetje, L., 1978. Senescence, digestibility and carbohydrate content of buffel grass and green panic leaves in swards. Australian Journal of Agricultural Research 29. https://doi.org/10.1071/AR9780503
375 376 377	Wolters, A., Linnemann, V., Herbst, M., Klein, M., Schäffer, A., Vereecken, H., 2003. Pesticide volatilization from soil: Lysimeter measurements versus predictions of European registration models. Journal of Environmental Quality. https://doi.org/10.2134/jeq2003.1183
378	