### Lab 18: Plot

AUTHOR Emily Hickey (A15575724)

## **Downstream Analysis**

```
library(tximport)
```

```
folders <- dir(pattern="SRR21568*")
samples <- sub("_quant", "", folders)
files <- file.path( folders, "abundance.h5" )
names(files) <- samples

txi.kallisto <- tximport(files, type = "kallisto", txOut = TRUE)</pre>
```

#### 1 2 3 4

```
head(txi.kallisto$counts)
```

```
SRR2156848 SRR2156849 SRR2156850 SRR2156851
ENST00000539570
                         0
                                    0
                                          0.00000
ENST00000576455
                                     0
                                          2.62037
                                                           0
ENST00000510508
                                    0
                                          0.00000
                                                           0
                         0
                                    1
ENST00000474471
                         0
                                       1.00000
ENST00000381700
                         0
                                         0.00000
                                                           0
ENST00000445946
                                          0.00000
```

```
colSums(txi.kallisto$counts)
```

```
SRR2156848 SRR2156849 SRR2156850 SRR2156851
2563611 2600800 2372309 2111474
```

```
sum(rowSums(txi.kallisto$counts)>∅)
```

#### [1] 94561

```
to.keep <- rowSums(txi.kallisto$counts) > 0
kset.nonzero <- txi.kallisto$counts[to.keep,]</pre>
```

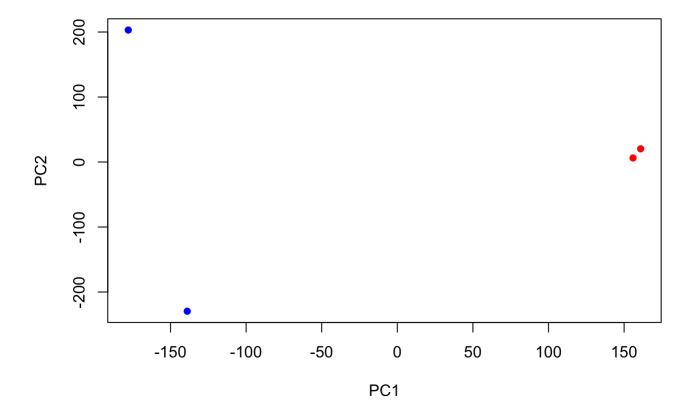
```
keep2 <- apply(kset.nonzero,1,sd)>0
x <- kset.nonzero[keep2,]</pre>
```

# **Principal Component Analysis**

```
pca <- prcomp(t(x), scale=TRUE)
summary(pca)</pre>
```

Importance of components:

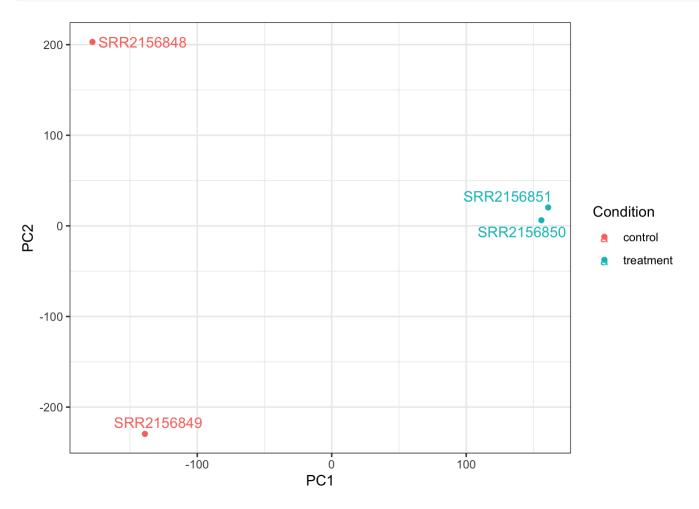
```
PC1 PC2 PC3 PC4 Standard deviation 183.6379 177.3605 171.3020 1e+00 Proportion of Variance 0.3568 0.3328 0.3104 1e-05 Cumulative Proportion 0.3568 0.6895 1.0000 1e+00
```



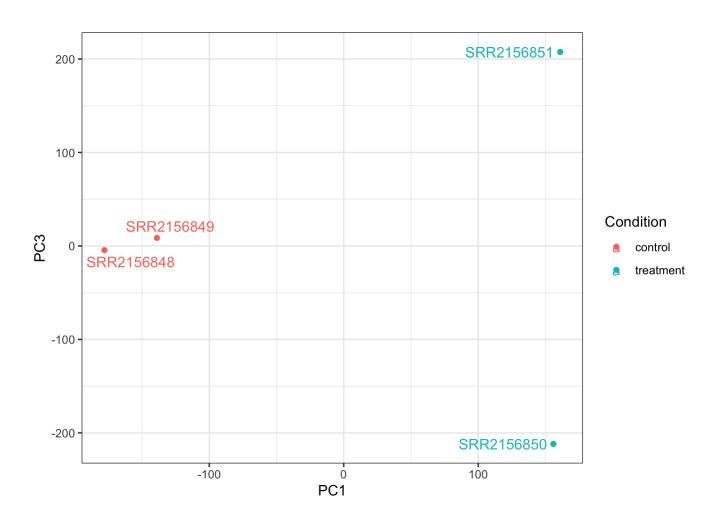
```
library(ggplot2)
library(ggrepel)
```

```
colData <- data.frame(condition = factor(rep(c("control", "treatment"), each = 2)))
rownames(colData) <- colnames(txi.kallisto$counts)</pre>
```

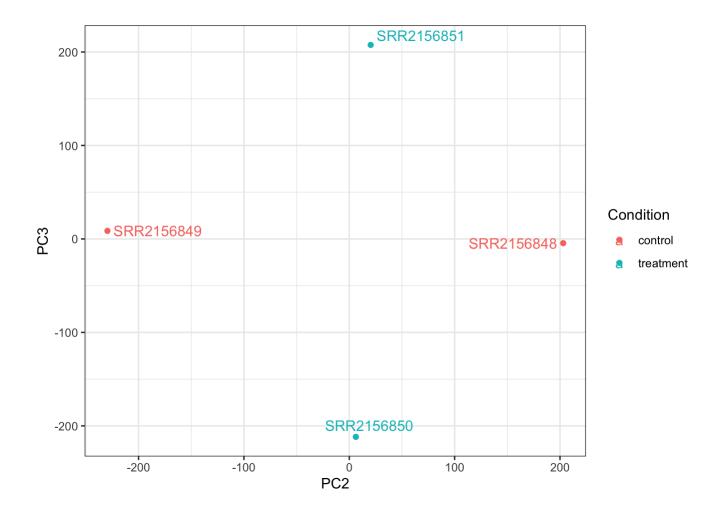
```
y <- as.data.frame(pca$x)
y$Condition <- as.factor(colData$condition)
ggplot(y) +
  aes(PC1, PC2, col=Condition) +
  geom_point() +
  geom_text_repel(label=rownames(y)) +
  theme_bw()</pre>
```



```
y <- as.data.frame(pca$x)
y$Condition <- as.factor(colData$condition)
ggplot(y) +
  aes(PC1, PC3, col=Condition) +
  geom_point() +
  geom_text_repel(label=rownames(y)) +
  theme_bw()</pre>
```



```
y <- as.data.frame(pca$x)
y$Condition <- as.factor(colData$condition)
ggplot(y) +
   aes(PC2, PC3, col=Condition) +
   geom_point() +
   geom_text_repel(label=rownames(y)) +
   theme_bw()</pre>
```



# Differential-expression analysis

```
Loading required package: S4Vectors

Warning: package 'S4Vectors' was built under R version 4.3.2

Loading required package: stats4

Loading required package: BiocGenerics

Attaching package: 'BiocGenerics'

The following objects are masked from 'package:stats':

IQR, mad, sd, var, xtabs

The following objects are masked from 'package:base':

anyDuplicated, aperm, append, as.data.frame, basename, cbind,
```

colnames, dirname, do.call, duplicated, eval, evalq, Filter, Find, get, grep, grepl, intersect, is.unsorted, lapply, Map, mapply, match, mget, order, paste, pmax, pmax.int, pmin, pmin.int, Position, rank, rbind, Reduce, rownames, sapply, setdiff, sort, table, tapply, union, unique, unsplit, which.max, which.min

Attaching package: 'S4Vectors'

The following object is masked from 'package:utils':

findMatches

The following objects are masked from 'package:base':

expand.grid, I, unname

Loading required package: IRanges

Loading required package: GenomicRanges

Loading required package: GenomeInfoDb

Warning: package 'GenomeInfoDb' was built under R version 4.3.2

Loading required package: SummarizedExperiment

Loading required package: MatrixGenerics

Loading required package: matrixStats

Attaching package: 'MatrixGenerics'

The following objects are masked from 'package:matrixStats':

colAlls, colAnyNAs, colAnys, colAvgsPerRowSet, colCollapse, colCounts, colCummaxs, colCummins, colCumprods, colCumsums, colDiffs, colIQRDiffs, colIQRs, colLogSumExps, colMadDiffs, colMads, colMaxs, colMeans2, colMedians, colMins, colOrderStats, colProds, colQuantiles, colRanges, colRanks, colSdDiffs, colSds, colSums2, colTabulates, colVarDiffs, colVars, colWeightedMads, colWeightedMeans, colWeightedMedians, colWeightedSds, colWeightedVars, rowAlls, rowAnyNAs, rowAnys, rowAvgsPerColSet, rowCollapse, rowCounts, rowCummaxs, rowCummins, rowCumprods, rowCumsums, rowDiffs, rowIQRDiffs, rowIQRs, rowLogSumExps, rowMadDiffs, rowMads, rowMaxs, rowMeans2, rowMedians, rowMins, rowOrderStats, rowProds, rowQuantiles, rowRanges, rowRanks, rowSdDiffs, rowSds, rowSums2, rowTabulates, rowVarDiffs, rowVars, rowWeightedMads, rowWeightedMeans, rowWeightedMedians, rowWeightedSds, rowWeightedVars

```
Loading required package: Biobase
Welcome to Bioconductor
    Vignettes contain introductory material; view with
    'browseVignettes()'. To cite Bioconductor, see
    'citation("Biobase")', and for packages 'citation("pkgname")'.
Attaching package: 'Biobase'
The following object is masked from 'package:MatrixGenerics':
    rowMedians
The following objects are masked from 'package:matrixStats':
    anyMissing, rowMedians
sampleTable <- data.frame(condition = factor(rep(c("control", "treatment"), each = 2)))</pre>
rownames(sampleTable) <- colnames(txi.kallisto$counts)</pre>
dds <- DESeqDataSetFromTximport(txi.kallisto,</pre>
                                  sampleTable,
                                  ~condition)
using counts and average transcript lengths from tximport
dds <- DESeq(dds)</pre>
estimating size factors
using 'avgTxLength' from assays(dds), correcting for library size
estimating dispersions
gene-wise dispersion estimates
mean-dispersion relationship
-- note: fitType='parametric', but the dispersion trend was not well captured by the
   function: y = a/x + b, and a local regression fit was automatically substituted.
   specify fitType='local' or 'mean' to avoid this message next time.
final dispersion estimates
fitting model and testing
```

# res <- results(dds) head(res)</pre>

log2 fold change (MLE): condition treatment vs control
Wald test p-value: condition treatment vs control

DataFrame with 6 rows and 6 columns

	baseMean	log2FoldChange	lfcSE	stat	pvalue
	<numeric></numeric>	<numeric></numeric>	<numeric></numeric>	<numeric></numeric>	<numeric></numeric>
ENST00000539570	0.000000	NA	NA	NA	NA
ENST00000576455	0.761453	3.155061	4.86052	0.6491203	0.516261
ENST00000510508	0.000000	NA	NA	NA	NA
ENST00000474471	0.484938	0.181923	4.24871	0.0428185	0.965846
ENST00000381700	0.000000	NA	NA	NA	NA
ENST00000445946	0.000000	NA	NA	NA	NA
	padj				

7 ....

<numeric>

ENST00000539570 NA
ENST00000576455 NA
ENST00000510508 NA
ENST00000474471 NA
ENST00000381700 NA
ENST00000445946 NA