2024/11/11 晚上10:04 「Module_5b_Adni_PatData_cleaned_4SNPs.ipynb」的副本 - Colab

```
    Welcome to the notebook where we will include data on 4 SNPs on chromosome 19.
```

This notebook was created at San Francisco State University for the PINC and gSTAR programs by Dr Pleuni Pennings, Lucy Moctezuma Tan and Lorena Benitez Rivera. We acknowledge help from Dr Adegoke Ojewole and Dr Hector Corrada Bravo.

Loading libraries and data from Github repository

```
# Importing packages for data handling
import pandas as pd
 import numpy as np
# importing packages for making Random Forest Model and evaluating performance
from sklearn.model_selection import train_test_split
from sklearn.ensemble import RandomForestClassifier
from xgboost import XGBClassifier
from sklearn import preprocessing
from sklearn import metrics
from sklearn.metrics import accuracy_score
# importing packages for ploting
from matplotlib import pyplot as plt
from sklearn.metrics import confusion_matrix, ConfusionMatrixDisplay
```

Load the "PatData_cleaned_4SNPs.csv" version of the dataset that has only 4 SNP locations on chromosome 19.

```
# Loading data from github repository
url = "https://raw.githubusercontent.com/pleunipennings/CSC508Data/main/PatData_cleaned_4SNPs.csv"
data = pd. read_csv(url)
data.head()
```

→	PTID	AGE	PTGENDER	PTEDUCAT	PTETHCAT	PTRACCAT	PTMARRY	APOE4	ı DX	Ventricles	Hippocampus	WholeBrain	Entorhinal	Fusiform	MidTemp	ICV	rs4147929	rs41289512	rs76320948	rs3865444	
	0 002_S_0295	84.8	2	18	1	1	1	,	l NL	43332.500000	6805.125000	1.071568e+06	3752.625000	17693.875000	19420.125000	1.649602e+06	1.000000	0.002014	0.0	0	11.
	1 002_S_0413	76.3	1	16	1	1	1	() NL	31936.454545	6824.636364	1.055413e+06	4131.090909	20095.909091	20235.545455	1.600009e+06	1.003998	0.000000	0.0	1	
	2 002_S_0559	79.3	2	16	1	1	2	,	l NL	38410.666667	7496.666667	1.092807e+06	3998.333333	18993.000000	22226.000000	1.703968e+06	2.000000	0.000000	0.0	1	
	3 002_S_0685	89.6	1	16	1	1	1	() NL	40921.571429	7063.250000	9.800458e+05	3894.375000	14152.250000	18133.625000	1.521331e+06	1.000000	0.000000	0.0	0	
	4 002_S_0729	65.1	1	16	1	1	1	,	l Dementia	23871.666667	5552.833333	8.942806e+05	2057.428571	16579.142857	16136.714286	1.298887e+06	2.000000	NaN	0.0	1	

Dropping Missing values

from plotnine import *

As mentioned before we have new features we have not used before, we will see if these will help improve our model's predictions for Alzheimers. Below we have a list of column names.

```
# Loading column feature names
data.columns
 Index(['PTID', 'AGE', 'PTGENDER', 'PTEDUCAT', 'PTETHCAT', 'PTRACCAT',
            'PTMARRY', 'APOE4', 'DX', 'Ventricles', 'Hippocampus', 'WholeBrain',
            'Entorhinal', 'Fusiform', 'MidTemp', 'ICV', 'rs4147929', 'rs41289512',
            'rs76320948', 'rs3865444'],
           dtype='object')
# Check for missing data
data.isna().sum()
           PTID
           AGE
        PTGENDER
```

PTEDUCAT 0 PTETHCAT 0 PTRACCAT 0 PTMARRY APOE4 DX **Ventricles** Hippocampus 0 WholeBrain **Entorhinal Fusiform** MidTemp **rs4147929** 10 **rs41289512** 81 rs76320948 33 **rs3865444** 0 dtype: int64

#Let's drop missing data present in the location features data = data.dropna() # Check how much data you have left

data. shape

Exploring Genetic Data (APOE4 and SNPs Data)

APOE4 Data

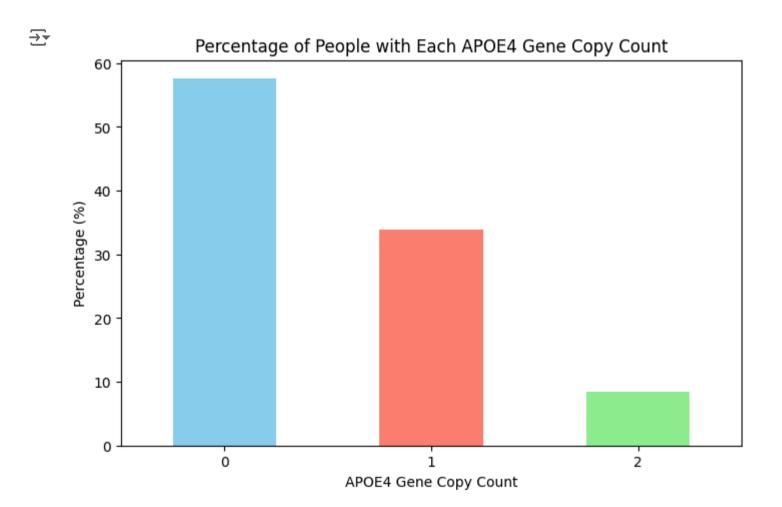
 \longrightarrow (698, 20)

We have work with this mysterious feature before. But it is worth taking a closer look into it. The APOE4 gene has been associated with an increased risk of Alzheimer's disease. APOE4 has values for each patient that range from 0 to 2. We see here that 0 is most common, followed by 1 and then 2. When a patient has a 0 here, it means that the patient doesn't carry the APOE4 allele, 1 means they carry one copy of APOE4 and 2 means they carry two copies.

Task 1: plotting APOE4 frequencies.

• Create a barplot to check what percentage of people in this dataset carry both copies of this gene!

```
import matplotlib.pyplot as plt
# Count the frequencies of each APOE4 value
apoe4_counts = data['APOE4'].value_counts(normalize=True) * 100
# Plot the frequencies as a bar chart
plt.figure(figsize=(8, 5))
apoe4_counts.plot(kind='bar', color=['skyblue', 'salmon', 'lightgreen'])
plt.title('Percentage of People with Each APOE4 Gene Copy Count')
plt.xlabel('APOE4 Gene Copy Count')
plt.ylabel('Percentage (%)')
plt.xticks(rotation=0)
plt.show()
```



SNP Locations Data

Recall from the text that SNP (Single Nucleotide Polymorphism) are variants that consist of just one nucleotide in the same location. For our

```
dataset we are considering 4 locations. These are:
   • rs4147929
  • rs41289512
```

• rs76320948 rs3865444

These variables represent the location of SNP (snips) that we have interest in, essentially it is the equivalent of a genetic address. These are official names you can google to find more about each of these sites.

Normally, for each SNP, each chromosome can carry one genetic letter (A, C, G or T). Because we each have two copies of each of our chromosomes, we also have two letters for a SNP.

On a website called SNPedia, I looked up rs4147929. It tells me that we can have AA, AG or GG at this location. Carrying one or two A's at this location is associated with higher Alzheimer's risk.

Now, in the data we have, the letters are not listed, but instead a number is listed that we can interpret as the estimated number of G (non-risk) alleles in that person. If this number is close to 2, the person has genotype GG, if the number is close to 1, they have AG and if the number is close to 0, they have the AA genotype.

The reason that the numbers are not exactly 0, 1, and 2 has to do with how the data is collected.

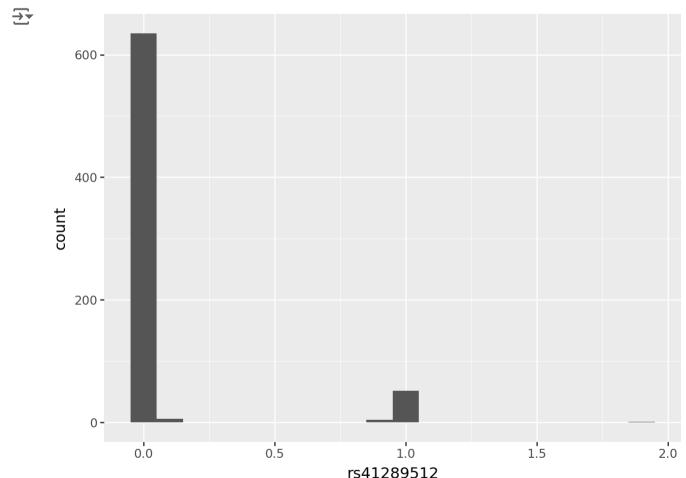
https://www.snpedia.com/index.php/Rs4147929

Looking at values in location "rs4147929"

data["rs4147929"].value_counts()

```
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          rs4147929
          2.000000 455
           1.000000
          0.000000
          1.001007
          1.997986
          1.003998
           1.996002
           1.002991
           1.981995
           1.998993
          1.002014
          1.928009
          1.042999
           1.001984
           1.009003
          1.053986
          1.019012
          1.973999
          1.092010
          1.997009
           1.005005
          0.998993
           1.026001
          0.006012
          1.031006
          1.010010
          0.996002
          1.994995
           1.949005
          1.902008
          0.001007
          1.946014
           1.993011
           1.988007
          0.011993
         dtype: int64
   # Let's round to have no decimals.
   data["rs4147929"] = data["rs4147929"].round(decimals = 0)
   data["rs4147929"].value_counts()
          rs4147929
                       474
             2.0
             1.0
                       203
             0.0
         dtype: int64
    Below is some terminoly for those who are not familiar with Genetic jargon!
       • Minor Frequency Alleles (MAF): is the frequency at which the second most common allele in a population occur. They are important to
         study in population genetics because they provide information to distinguish comon versus rare variants in a population.
       • Homozygotes: The SNPs Minor Frequency Alleles are taken from Table 1 in this <u>paper</u>. Each SNP comes in two letters (two alleles). The
         one that is less common is called the minor allele.
    Note that for this SNP in location rs4147929, the Minor Allele Frequency is listed as 0.161. We therefore expect the square of that frequency to
    be the number of people who carry the allele 2x (homozygotes). In other words: if around 16% of all chromosome 19s in humans have an A at
    this location, then the chance that someone has two As is 16% of 16% = 2.6%. In our dataset, that would translate to just 18 people out of 698
    people. Is this actually what the data shows?
   698*(0.161^2) = 18 \ people.
    As you can see in one of the previous cells, we have 21 peope in the dataset with two times the minor allele. That is pretty close to 18!
   # Plotting counts for location rs4147929
    (ggplot(data=data) +
     aes(x='rs4147929') +
     geom_histogram(stat='bin', binwidth = 0.1) +
     labs(x = "rs4147929")
    \overline{\Rightarrow}
             300 -
                    0.0
                                                                      1.5
                                                 rs4147929
   Now for this SNP in location rs41289512 the Minor Allele Frequency (MAF) listed was 0.039. Notice that there are no homozygotes of the
    second type. We would expect 0.039^2\ homozygotes=0.15 . Out of 698 patients, we had expected just 1 patient with two times the minor
    allele. It is not so surprising that we don't see any.
   # Plotting counts for location rs41289512
    (ggplot(data=data) +
     aes(x='rs41289512') +
     geom_histogram(stat='bin', binwidth = 0.1) +
     labs(x = "rs41289512")
             200 -
                    0.0
                                                       1.0
                                      0.5
                                                 rs41289512
```

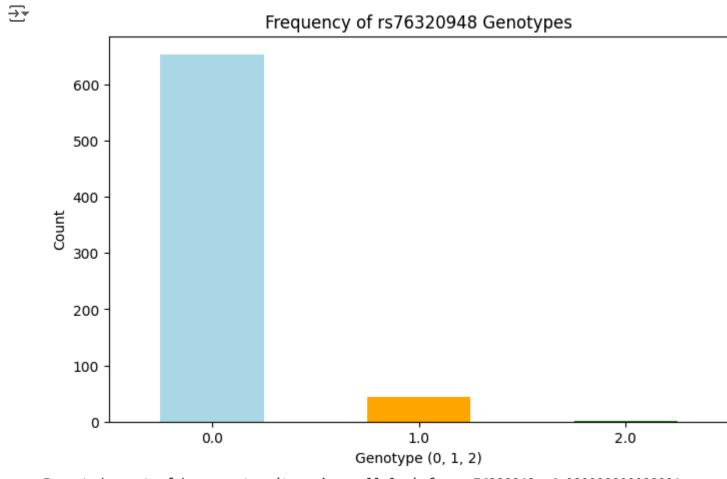
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Task 2: Looking at SNP genotypes.

• Plot the frequency barplots for the other two sites (rs76320948 and rs3865444). • Calculate the expected and observed number of people in the dataset with twice the minor allele for these SNPs. Is it what was expected?

```
# Plot frequency for SNP site rs76320948
rs76320948_counts = data['rs76320948'].round().value_counts()
# Plotting
plt.figure(figsize=(8, 5))
rs76320948_counts.plot(kind='bar', color=['lightblue', 'orange', 'green'])
plt.title('Frequency of rs76320948 Genotypes')
plt.xlabel('Genotype (0, 1, 2)')
plt.ylabel('Count')
plt.xticks(rotation=0)
plt.show()
# Calculate expected number with twice the minor allele
# Minor Allele Frequency (MAF) for rs76320948 is assumed; adjust with actual MAF if known
maf_rs76320948 = 0.1 # Example MAF, replace if known
expected_homozygotes = len(data) * (maf_rs76320948 ** 2)
print(f"Expected count of homozygotes (two minor alleles) for rs76320948: {expected_homozygotes}")
print(f"Observed count: {rs76320948_counts.get(0, 0)}")
```



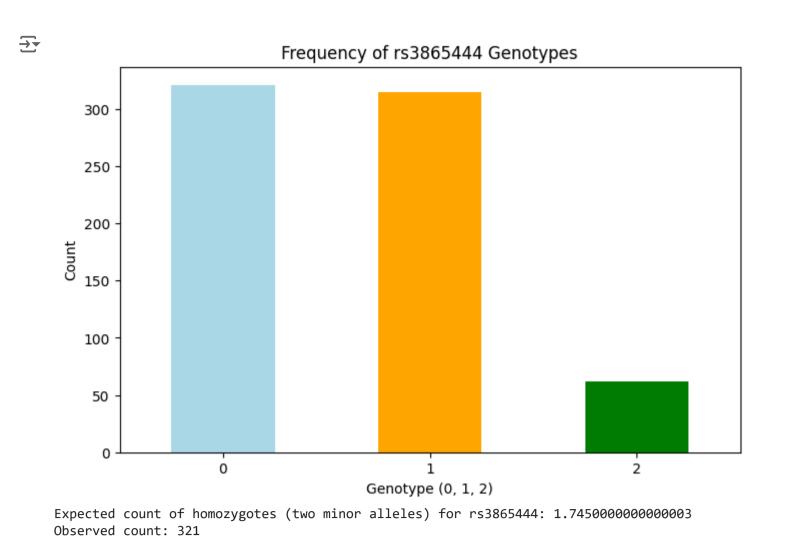
Expected count of homozygotes (two minor alleles) for rs76320948: 6.98000000000001 Observed count: 653

Plot frequency for SNP site rs3865444 rs3865444_counts = data['rs3865444'].round().value_counts()

https://colab.research.google.com/drive/1D9RgSofVUiqrXxtH78hG2GpNK9Ce1b3k#scrollTo=9rX3uwfM_fUS&printMode=true

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```
# Plotting
plt.figure(figsize=(8, 5))
rs3865444_counts.plot(kind='bar', color=['lightblue', 'orange', 'green'])
plt.title('Frequency of rs3865444 Genotypes')
plt.xlabel('Genotype (0, 1, 2)')
plt.ylabel('Count')
plt.xticks(rotation=0)
plt.show()
# Calculate expected number with twice the minor allele
# Minor Allele Frequency (MAF) for rs3865444 is assumed; adjust with actual MAF if known
maf_rs3865444 = 0.05  # Example MAF, replace if known
expected_homozygotes = len(data) * (maf_rs3865444 ** 2)
print(f"Expected count of homozygotes (two minor alleles) for rs3865444: {expected_homozygotes}")
print(f"Observed count: {rs3865444_counts.get(0, 0)}")
```



Random forest with genetic data for 4 SNPs

In this part we will create a Random Forest Model from this data set and valuate the model.

- Split the data in labels (the diagnosis) and features (the other columns)
- Use about 70-80% of our data as the training data and the rest as test data. In the code, 70% training and 30% test.
- Create our Randdom Fores model and training it
- Making our predictions

NOTE: Notice that we have set the *Random seed = 1* for splitting our data and into testing and when creating our Random Forest object. This was done for reproducibility of results

Splitting data into labels and features labels = data["DX"] features = data.drop(columns=['PTID', 'DX']) # splitting data into training and testing sets features_train, features_test, labels_train, labels_test = train_test_split(features, labels, test_size=0.3, random_state=1) # creating Random Fores Model and Training it rf = RandomForestClassifier(n_estimators = 1000, bootstrap = True, random_state = 1) rf.fit(features_train, labels_train) # #Predict the response for test dataset labels_pred = rf.predict(features_test)

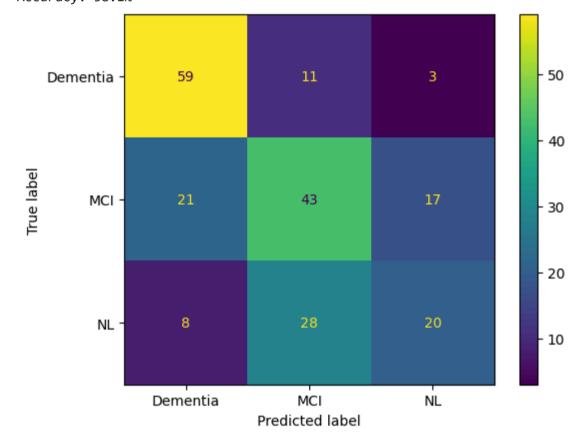
Let's now take a look at what was predicted and have a sense of how we did by creating a Confusion Matrix Plot. We can also plot an accuracy score along with it.

Look at the predicted values. print(labels_pred[:10])

#Let's visualize how well the RF does.

['MCI' 'MCI' 'MCI' 'Dementia' 'MCI' 'Dementia' 'MCI' 'NL' 'MCI']

accuracy = accuracy_score(labels_test, labels_pred) print("Accuracy: %.1f%%" % (accuracy * 100)) plt2 = metrics.ConfusionMatrixDisplay.from_estimator(rf, features_test, labels_test) plt.grid(False) → Accuracy: 58.1%



We can see that our model seems to predict fairly well for people that have Alzheimer's, 13 mistakes versus 60 correct predictions. For Mild Cognitive impairment the model does not do as well with 42 correct predictions versus 39 erros and finally for Normal is doesn't do well at all since its 21 versus 35. We can actually obtain individual measures for how well a model does for a particular class using F-scores, more on this in later modules.

Checking Feature Importance for our model

Below we will check the feature importance for our model. Recall that this basically shows us what features were most relevant in making predictions. Let's also visualize this to see where does SNP features rank in terms of importance for the model.

getting feature importance values from our model importance = rf.feature_importances_

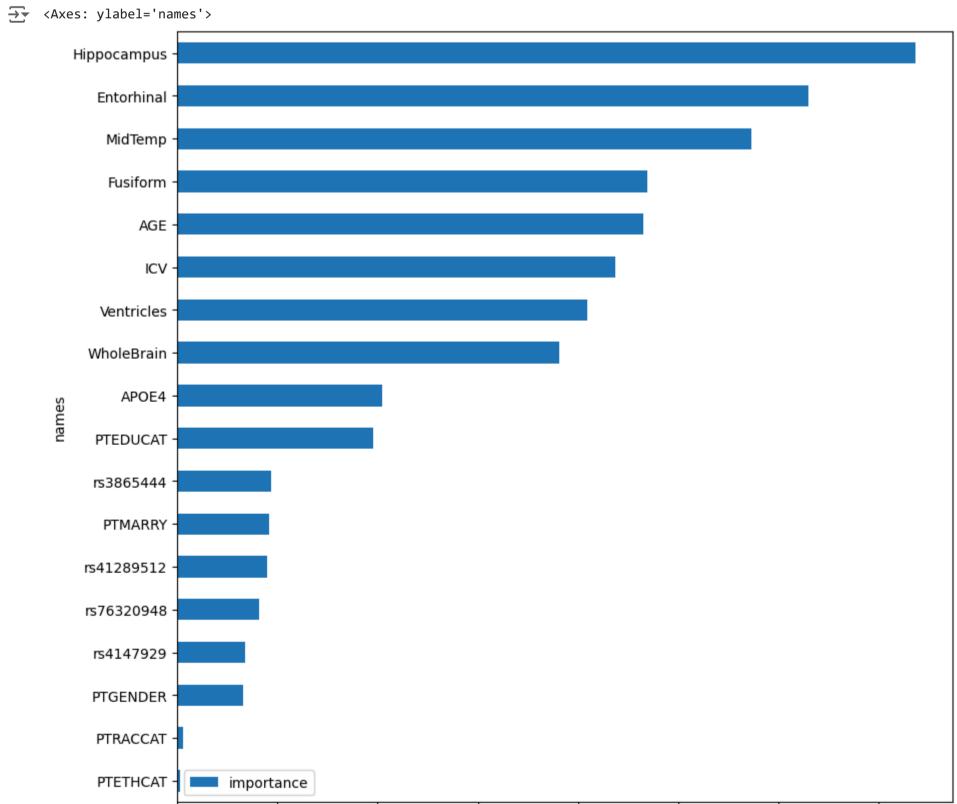
summarize feature importance / see if the SNP columns are important for the RF names = features.columns.to_numpy(dtype=object)

Creating a dataframe for feature importance

importanceDF = pd. DataFrame({'names':names, 'importance':importance}) # Sort the dataframe with feature importances

importanceDF = importanceDF.sort_values(by=['importance'])

Creating barplot for feature importance importanceDF.plot.barh(x='names', y='importance', figsize = (10,10))



The SNP locations do not place very high in terms of aiding prediction for our model. We can see that the presence of the gene APOE4 is more relevant! The main important feature still is the hypocampus volume though by far.

0.10

0.12

0.14

So the question remains if adding genetic data actually help with the predictions at all? Below we will redo the Random Forest Model this time without the genetic data to see changes in the performance of the model and find out!

Does Removing Genetic Data worsen the results for Random Forest Model?

In this iteration we will refit our model twice:

- First by removing the SNP location data Model 1
- Then by removing both SNP location data as well as the APOE4 gene feature **Model 2**

Creating 2 lists of column features one with just SNP sites and another with all genetic data SNP_cols = ['rs4147929', 'rs41289512', 'rs76320948', 'rs3865444'] Genetics_cols = ['rs4147929', 'rs41289512', 'rs76320948', 'rs3865444', 'APOE4']

Now we will create 2 sets of features for both our models as described above. Our labels are still the same as before so we don't need to redefine them here.

Features for model 1 features_no_snp = features[features.columns.drop(SNP_cols)]

Features for model 2 features_no_genetics = features[features.columns.drop(Genetics_cols)]

We will proceed with doing 2 different splits of training and testing data. Notice that i am keeping the random seed as 1 just to be consistent with the prior model containing all genetic features.

Split of testing and training for Model 1

X_train1, X_test1, Y_train1, Y_test1 = train_test_split(features_no_snp, labels, test_size=0.3, random_state=1)

https://colab.research.google.com/drive/1D9RgSofVUiqrXxtH78hG2GpNK9Ce1b3k#scrollTo=9rX3uwfM_fUS&printMode=true

Now we will be creating 2 random forest models and train them separately. We have mantain again the same hyperparameter settings than before so we can make a fairer comparison.

. . .

```
# Creating and training Random forest with model 1
rf1 = RandomForestClassifier(n_estimators = 1000, bootstrap = True, random_state = 1)
rf1.fit(X_train1, Y_train1)
# Creating and training Random forest with model 2
rf2 = RandomForestClassifier(n_estimators = 1000, bootstrap = True, random_state = 1)
rf2.fit(X_train2, Y_train2)
```

RandomForestClassifier RandomForestClassifier(n_estimators=1000, random_state=1)

Split of testing and training for Model 2

We will make 2 sets of predictions, one with model1 and another with model2

Predictions using model 1 Y_pred1 = rf1.predict(X_test1) # Predictions using model 2 Y_pred2 = rf2.predict(X_test2)

Finally we will be taking a look at each models confusion matrix and accuracy scores, and what differences we get if any!

plot for model 1 fig, (ax1, ax2) = plt.subplots(1, 2, figsize=(10, 10), sharey= True)rfl_matrix = confusion_matrix(Y_test1, Y_pred1) plt1 = ConfusionMatrixDisplay(rf1_matrix, display_labels=["Dementia", "MCI", "NL"]) plt1. plot (ax=ax1) plt1.ax_.set_title("Model 1: No SNP Location data") plt1.im_.colorbar.remove() # plot for Model 2 rf2_matrix = confusion_matrix(Y_test2, Y_pred2) plt2 = ConfusionMatrixDisplay(rf2_matrix, display_labels=["Dementia","MCI","NL"]) plt2. plot (ax=ax2)plt2.ax_.set_title("Model 2: No SNP Location & APOE4 data") plt2.im_.colorbar.remove() plt.grid(False)

Exception ignored in: <function WeakMethod.__new__.<locals>._cb at 0x7e962d0ca320> Traceback (most recent call last):

File "/usr/lib/python3.10/weakref.py", line 61, in _cb File "/usr/local/lib/python3.10/dist-packages/matplotlib/cbook.py", line 248, in _remove_proxy del self.callbacks[signal][cid]



printing accuracies for both models acc_1 = accuracy_score(Y_test1, Y_pred1) print("Accuracy for model 1: %.1f%%" % (acc_1 * 100)) acc_2 = accuracy_score(Y_test2, Y_pred2) print("Accuracy for model 2: %.1f%%" % (acc_2 * 100))

Accuracy for model 1: 60.0% Accuracy for model 2: 59.5%

Looking at both of these Confusion matrices side by side, it looks like they are not that different. Model 2 does not include APOE4 and it seemed to only do a bit worse than Model 1. Surprisingly however removing the genetic information seems to have increased it's accuracy, albeit just by a minuscule amount!

How will a Gradient Boosted Tree Model do?

Below we can quickly check the accuracies had we tried using a Gradient Boosted Model rather than a Random Forest. We will do a check of accuracies for 3 models:

Including all the original data Model GBT

• First by removing the SNP location data Model GBT1

• Then by removing both SNP location data as well as the APOE4 gene feature **Model GBT2**

Recall that first we need to recode our labels into numbers for XGBoost modeling before we split the data. In the code below we will transform our Diagnosis Category into the values 0, 1 and 2 (Dementia, MCI and NL).

Separating labels from the general dataframe labels = data["DX"] # Creating a label encoder object le = preprocessing.LabelEncoder() # Fitting the label encoder into the labels columns le.fit(data["DX"]) # Transforming the classes into numbers labels_t = le.transform(data["DX"])

Splitting the Data again after **recoding the label** into numbers.

Split of testing and training for Model GBT X_train, X_test, Y_train, Y_test = train_test_split(features, labels_t, test_size=0.3, random_state=1) # Split of testing and training for Model GBT1 X_train1, X_test1, Y_train1, Y_test1 = train_test_split(features_no_snp, labels_t, test_size=0.3, random_state=1)

Split of testing and training for Model GBT2 X_train2, X_test2, Y_train2, Y_test2 = train_test_split(features_no_genetics, labels_t, test_size=0.3, random_state=1)

Now we can quickly run Model GBT, Model GBT1 and Model GBT2 to compare accuracies.

Checking accuracy for Model GBT GBT = XGBClassifier(random_state=1) GBT.fit(X_train, Y_train) $Y_pred_GBT = GBT.predict(X_test)$ acc_GBT = accuracy_score(Y_test, Y_pred_GBT) print("Accuracy: %.1f%%" % (acc_GBT * 100)) Accuracy: 61.0%

GBT1 = XGBClassifier() GBT1.fit(X_train1,Y_train1)

Checking accuracy for Model GBT1

 $Y_pred_GBT1 = GBT1.predict(X_test1)$ acc_GBT1 = accuracy_score(Y_test1, Y_pred_GBT1) print("Accuracy: %.1f%%" % (acc_GBT1 * 100)) → Accuracy: 61.4%

Checking accuracy for Model GBT2 GBT2 = XGBClassifier() GBT2.fit(X_train2,Y_train2) Y_pred_GBT2 = GBT2.predict(X_test2) acc_GBT2 = accuracy_score(Y_test2, Y_pred_GBT2) print("Accuracy: %.1f%%" % (acc_GBT2 * 100))

Accuracy: 59.5%

Overall It seems that Gradient Boosted Trees perform better than the Random Forest one. However, it seems that for Gradient Boosted Trees, including less Genetic data seems to worsen the performance, unlike what we saw for Random Forests.

Task 3: Hyperparameters.

1. Go back to the results of your hyper parameter tuning work. Plug in some better parameters in the models I made above. Can you make them perform better? 2. Does the genetic information about the patients help us make a better predictive model?

from sklearn.preprocessing import LabelEncoder

Split the data into training and testing sets

rf_best_accuracy = accuracy_score(y_test, rf_best_pred) print(f"Tuned Random Forest Accuracy: {rf_best_accuracy}")

Encode target variable as numeric values label_encoder = LabelEncoder() y = label_encoder.fit_transform(y) # This will convert 'Dementia', 'MCI', 'NL' to numeric labels

Train the Random Forest model with tuned hyperparameters rf_best_model = RandomForestClassifier(random_state=1, **best_rf_params) rf_best_model.fit(X_train, y_train) rf_best_pred = rf_best_model.predict(X_test)

X_train, X_test, y_train, y_test = train_test_split(X, y, test_size=0.3, random_state=1)

Train the XGBoost model with tuned hyperparameters xgb_best_model = XGBClassifier(random_state=1, **best_xgb_params) xgb_best_model.fit(X_train, y_train) xgb_best_pred = xgb_best_model.predict(X_test) xgb_best_accuracy = accuracy_score(y_test, xgb_best_pred) print(f"Tuned XGBoost Accuracy: {xgb_best_accuracy}")

To interpret the numeric predictions, you can use label_encoder.inverse_transform()

Tuned Random Forest Accuracy: 0.5619047619047619 Tuned XGBoost Accuracy: 0.6142857142857143

1. Evaluate Model Performance: Train models both with and without genetic data. An increase in accuracy and other relevant metrics when genetic data is included indicates that it offers valuable predictive insights.

2. Assess Feature Importance: Analyze the significance of genetic features, such as APOE4, in models like Random Forest. If these features rank high in importance, it means they play a crucial role in shaping predictions.

3. Examine Specific Predictions: Investigate whether the inclusion of genetic data enhances the classification of particular categories, such as distinguishing Alzheimer's from Mild Cognitive Impairment.

https://colab.research.google.com/drive/1D9RgSofVUiqrXxtH78hG2GpNK9Ce1b3k#scrollTo=9rX3uwfM fUS&printMode=true