DivDiv Trait Data Citations

Eric Crandall

# Generational\_Structure

*1*

NA

*2*

*2*

*2*

*2*

*2*

NA

*3*

*4*

*5*

*6*

*7*

NA

*8*

NA

NA

NA

*9*

*10*

NA

*11*

NA

*12*

*13*

NA

*14*

NA

*15*

NA

*2*

NA

*16*

NA

NA

NA

*17*

*18*

*19*

*20*

*2*

NA

NA

*2*

*10*

*21*

*22*

*23*

*20*

*20*

*24*

*25*

*26*

NA

NA

*20*

*27*

NA

*28*

*2*

*29*

*30*

*31*

*10*

*32*

*1*

NA

NA

*33*

*33*

*33*

*33*

*33*

*33*

*34*

*35*

*20*

NA

NA

*2*

*36*

*37*

*37*

*10*

*38*

*39*

NA

NA

# ReturnToSpawningGround

NA

*40*

*2*

*2*

*2*

*2*

*2*

NA

NA

*41*

*5*

*6*

NA

NA

*42*

NA

NA

NA

NA

NA

NA

NA

NA

NA

*13*

NA

NA

NA

*43*

*44*

*2*

NA

*16*

NA

NA

NA

*17*

*45*

NA

*20*

*2*

NA

NA

*2*

NA

*2*

*46*

NA

*20*

*20*

*47*

*25*

*48*

NA

NA

*20*

*2*

NA

*2*

*2*

*41*

*41*

*41*

NA

NA

NA

NA

*49*

NA

NA

NA

NA

*50*

*50*

NA

NA

*51*

NA

NA

*2*

NA

*52*

*53*

NA

*38*

NA

NA

NA

# Spawning\_mode

*1*

*54*

*55*

*55*

*55*

*56*

*56*

*54*

*3*

NA

NA

NA

*7*

*54*

*57*

*57*

*58*

*59*

*60*

*61*

*54*

*54*

*62*

*54*

NA

*63*

*64*

*54*

*65*

*66*

*67*

*68*

NA

*54*

*69*

*54*

NA

*70*

*71*

*20*

*72*

*54*

*54*

*73*

*74*

*75*

*76*

*77*

*54*

*78*

*47*

NA

NA

*79*

*80*

*78*

*27*

*54*

*81*

*82*

NA

NA

NA

*83*

*32*

*1*

*84*

*20*

*33*

*33*

*33*

*33*

*33*

*33*

*34*

*35*

*20*

*54*

*54*

*85*

*36*

NA

NA

*86*

NA

*54*

*54*

*60*

# Larval\_feeding

*87*

*54*

*88*

*88*

*88*

*88*

*88*

*54*

*89*

*90*

*5*

*6*

*91*

*54*

*57*

*57*

*92*

*59*

*93*

*61*

*54*

*54*

NA

*54*

*13*

*63*

*94*

*54*

*65*

*95*

*96*

*68*

*16*

*54*

*97*

*54*

*17*

*45*

*71*

*20*

*98*

*54*

*54*

*99*

*100*

*99*

*101*

*102*

*54*

*78*

*47*

*25*

*48*

*103*

*104*

*78*

*99*

*54*

*99*

*99*

*90*

*90*

*90*

*105*

*106*

*1*

*107*

*20*

*33*

*33*

*33*

*33*

*33*

*33*

*34*

*35*

*20*

*54*

*54*

*99*

*36*

*52*

*53*

*86*

*38*

*54*

*54*

*60*

# isPlanktonic\_atanypoint

*1*

*54*

*55*

*55*

*55*

*56*

*56*

*54*

*89*

NA

NA

NA

*7*

*54*

*57*

*57*

NA

*59*

*93*

*61*

*54*

*54*

*62*

*54*

NA

*108*

*109*

*54*

*65*

*110*

*111*

*68*

NA

*54*

*97*

*54*

NA

*45*

*71*

*20*

*72*

*54*

*54*

*73*

*100*

*112*

*113*

*77*

*54*

*78*

*47*

NA

NA

*103*

*104*

*78*

*114*

*54*

*115*

*116*

NA

NA

NA

*83*

NA

*1*

NA

*117*

*118*

*118*

*118*

*118*

*118*

*50*

*34*

*35*

*20*

*54*

*54*

*119*

*36*

NA

NA

NA

NA

*54*

*54*

*60*

# isBenthic

# PLD\_point

*1*

*120*

*55*

*55*

*55*

*56*

*56*

*121*

*89*

NA

NA

NA

NA

NA

NA

NA

NA

*59*

NA

*61*

*122*

*123*

*62*

*124*

NA

*125*

*109*

NA

NA

*126*

*127*

NA

NA

*128*

*97*

*129*

NA

NA

*71*

*130*

*131*

*132*

*132*

*133*

*134*

*75*

*113*

*135*

*136*

*137*

*47*

NA

NA

NA

*138*

*139*

*140*

*141*

*142*

*143*

NA

NA

NA

*144*

NA

*1*

*145*

*146*

*147*

*147*

*147*

*147*

*147*

*147*

*34*

*148*

NA

*149*

*120*

NA

*36*

NA

NA

*150*, *151*

NA

*152*

*153*

*60*

# Body\_Size

*154*

*155*

*156*

*157*

*158*

*159*

*160*

*20*

*89*

*90*

*5*

*6*

*161*

*162*

*163*

*163*

*164*

*165*

*93*

*166*

*167*

*20*

*168*

*163*

*13*

*125*

*169*

*20*

*163*

*20*

*170*

*20*

*16*

*20*

*171*

*20*

*17*

*18*

*172*

*20*

*173*, *174*

*20*

*20*

*175*

*176*

*177*

*178*

*179*

*20*

*163*

24

*25*

*180*

*181*

*182*

*163*

*183*

*20*

*21*

*184*

*90*

*90*

*90*

*185*

*186*

*187*

*163*

*188*

*163*

*163*

*163*

*163*

*189*

*20*

*35*

*163*

*20*

*20*

*21*

*190*

*191*

*37*

*10*, *192*

*38*

*20*

*20*

*20*

# Fecundity\_EggSize

*193*, *194*

*152*

*195*

*195*

*195*

*56*

*56*

*196*

*197*

*90*

NA

*6*

*198*

*199*

*163*

*163*

*200*

*59*

*93*

NA

*163*

*201*

NA

*163*

*13*

NA

*109*

NA

*163*

*20*

*202*

NA

*16*

NA

*203*

NA

*17*

NA

*71*

*204*

*98*

*205*

*206*

*93*

*207*

*208*

*93*

*23*

NA

*163*

*209*

*25*

*48*

*103*

NA

*163*

*210*

*211*

*212*

*90*

*90*

*90*

*213*

*214*

NA

NA

*163*

*215*

*215*

*215*

*215*

*215*

*215*

*34*

*35*

*215*

NA

NA

*216*

*36*

*191*

*53*

*86*

*38*

*217*

*218*

*219*

1. L. E. Timm, L. M. Isma, M. W. Johnston, H. D. Bracken-Grissom, Comparative population genomics and biophysical modeling of shrimp migration in the gulf of mexico reveals current-mediated connectivity. *Frontiers in Marine Science*. **7** (2020), doi:[10.3389/fmars.2020.00019](https://doi.org/10.3389/fmars.2020.00019).

2. J. S. Madin, K. D. Anderson, M. H. Andreasen, T. C. L. Bridge, S. D. Cairns, S. R. Connolly, E. S. Darling, M. Diaz, D. S. Falster, E. C. Franklin, R. D. Gates, A. M. T. Harmer, M. O. Hoogenboom, D. Huang, S. A. Keith, M. A. Kosnik, C.-Y. Kuo, J. M. Lough, C. E. Lovelock, O. Luiz, J. Martinelli, T. Mizerek, J. M. Pandolfi, X. Pochon, M. S. Pratchett, H. M. Putnam, T. E. Roberts, M. Stat, C. C. Wallace, E. Widman, A. H. Baird, The coral trait database, a curated database of trait information for coral species from the global oceans. *Scientific Data*. **3** (2016), doi:[10.1038/sdata.2016.17](https://doi.org/10.1038/sdata.2016.17).

3. K. P. Sebens, [Reproductive ecology of the intertidal sea anemones anthopleura xanthogrammica (brandt) and a. Elegantissima (brandt): Body size, habitat, and sexual reproduction](https://doi.org/10.1016/0022-0981(81)90159-3). *Journal of Experimental Marine Biology and Ecology*. **54**, 225–250 (1981).

4. Aptenodytes patagonicus: BirdLife international: The IUCN red list of threatened species 2020: e.T22697748A184637776. *IUCN Red List of Threatened Species* (2020), doi:[10.2305/iucn.uk.2020-3.rlts.t22697748a184637776.en](https://doi.org/10.2305/iucn.uk.2020-3.rlts.t22697748a184637776.en).

5. Arctocephalus forsteri: Chilvers, b.l. & goldsworthy, s.d.: The IUCN red list of threatened species 2015: e.T41664A45230026. *IUCN Red List of Threatened Species* (2014), doi:[10.2305/iucn.uk.2015-2.rlts.t41664a45230026.en](https://doi.org/10.2305/iucn.uk.2015-2.rlts.t41664a45230026.en).

6. Aythya marila: BirdLife international: The IUCN red list of threatened species 2018: e.T22680398A132525108. *IUCN Red List of Threatened Species* (2018), doi:[10.2305/iucn.uk.2018-2.rlts.t22680398a132525108.en](https://doi.org/10.2305/iucn.uk.2018-2.rlts.t22680398a132525108.en).

7. E. O’Reilly, B. M. Titus, M. W. Nelsen, S. Ratchford, N. E. Chadwick, [Giant ephemeral anemones? Rapid growth and high mortality of corkscrew sea anemones bartholomea annulata (le sueur, 1817) under variable conditions](https://doi.org/10.1016/j.jembe.2018.08.013). *Journal of Experimental Marine Biology and Ecology*. **509**, 44–53 (2018).

8. D. A. Ebert, [Reproductive biology of skates, bathyraja(ishiyama), along the eastern bering sea continental slope](https://doi.org/10.1111/j.0022-1112.2005.00628.x). *Journal of Fish Biology*. **66**, 618–649 (2005).

9. M. R. Gaither, G. A. Gkafas, M. de Jong, F. Sarigol, F. Neat, T. Regnier, D. Moore, D. R. Gr?cke, N. Hall, X. Liu, J. Kenny, A. Lucaci, M. Hughes, S. Haldenby, A. R. Hoelzel, [Genomics of habitat choice and adaptive evolution in a deep-sea fish](https://doi.org/10.1038/s41559-018-0482-x). *Nature Ecology & Evolution*. **2**, 680–687 (2018).

10. H.-J. T. Hoving, V. V. Laptikhovsky, B. H. Robison, [Vampire squid reproductive strategy is unique among coleoid cephalopods](https://doi.org/10.1016/j.cub.2015.02.018). *Current Biology*. **25**, R322–R323 (2015).

11. E. M. Salas, G. Bernardi, M. L. Berumen, M. R. Gaither, L. A. Rocha, [RADseq analyses reveal concordant indian ocean biogeographic and phylogeographic boundaries in the reef fishDascyllus trimaculatus](https://doi.org/10.1098/rsos.172413). *Royal Society Open Science*. **6**, 172413 (2019).

12. B. A. Fach, *Turkish Journal of Fisheries and Aquatic Sciences*. **14** (2014), doi:[10.4194/1303-2712-v14\_2\_06](https://doi.org/10.4194/1303-2712-v14_2_06).

13. Eudyptes chrysolophus: BirdLife international: The IUCN red list of threatened species 2020: e.T22697793A184720991. *IUCN Red List of Threatened Species* (2020), doi:[10.2305/iucn.uk.2020-3.rlts.t22697793a184720991.en](https://doi.org/10.2305/iucn.uk.2020-3.rlts.t22697793a184720991.en).

14. D. Grawunder, E. A. Hambleton, M. Bucher, I. Wolfowicz, N. Bechtoldt, A. Guse, Induction of gametogenesis in the cnidarian endosymbiosis model aiptasia sp. *Scientific Reports*. **5** (2015), doi:[10.1038/srep15677](https://doi.org/10.1038/srep15677).

15. A. J. Edwards, [FAO species catalogue. Vol. 10. Gadiform fishes of the world (order gadiformes). An annotated and illustrated catalogue of cods, hakes, grenadiers and other gadiform fishes known to date](https://doi.org/10.1016/0025-326x(92)90599-2). *Marine Pollution Bulletin*. **24**, 326–327 (2024).

16. Halichoerus grypus: Bowen, d.: The IUCN red list of threatened species 2016: e.T9660A45226042. *IUCN Red List of Threatened Species* (2016), doi:[10.2305/iucn.uk.2016-1.rlts.t9660a45226042.en](https://doi.org/10.2305/iucn.uk.2016-1.rlts.t9660a45226042.en).

17. <https://animaldiversity.org/accounts/Lagenorhynchus_acutus/#lifespan_longevity>

18. G. C. D. Estrada, C. H. Callado, M. L. G. Soares, C. S. Lisi, [Annual growth rings in the mangrove laguncularia racemosa (combretaceae)](https://doi.org/10.1007/s00468-008-0224-9). *Trees*. **22**, 663–670 (2008).

19. B.-D. Zhang, D.-X. Xue, Y.-L. Li, J.-X. Liu, [RAD genotyping reveals fine-scale population structure and provides evidence for adaptive divergence in a commercially important fish from the northwestern pacific ocean](https://doi.org/10.7717/peerj.7242). *PeerJ*. **7**, e7242 (2019).

20. Froese, R. and D. Pauly, Editors. 2000. [FishBase 2000: concepts, design and data sources](https://www.fishbase.se/). ICLARM, Los Baños, Laguna, Philippines. 344 p.

21. M. E. Brandt, [The effect of species and colony size on the bleaching response of reef-building corals in the florida keys during the 2005 mass bleaching event](https://doi.org/10.1007/s00338-009-0548-y). *Coral Reefs*. **28**, 911–924 (2009).

22. A. E. Hopkins, [Ecological observations on spawning and early larval development in the olympia oyster (ostrea lurida)](https://doi.org/10.2307/1932760). *Ecology*. **17**, 551–566 (1936).

23. D. Vafidis, C. Antoniadou, K. Kyriakouli, [Reproductive cycle of the edible sea urchin paracentrotus lividus (echinodermata: Echinoidae) in the aegean sea](https://doi.org/10.3390/w11051029). *Water*. **11**, 1029 (2019).

24. <https://www.irlspecies.org/taxa/index.php?taxon=9671#:~:text=Individuals%20reaching%20sexual%20maturity%20may,highly%20abundant%20throughout%20its%20range>

25. Phocoena phocoena: Braulik, g., minton , g., amano, m. & bjørge, a.: The IUCN red list of threatened species 2020: e.T17027A50369903. *IUCN Red List of Threatened Species* (2020), doi:[10.2305/iucn.uk.2020-2.rlts.t17027a50369903.en](https://doi.org/10.2305/iucn.uk.2020-2.rlts.t17027a50369903.en).

26. <https://www.iucnredlist.org/species/17028/50370296>

27. P. G’elin, C. Fauvelot, V. Mehn, S. Bureau, H. Rouz’e, H. Magalon, [Superclone expansion, long-distance clonal dispersal and local genetic structuring in the coral pocillopora damicornis type β in reunion island, south western indian ocean](https://doi.org/10.1371/journal.pone.0169692). *PLOS ONE*. **12**, e0169692 (2017).

28. K. Soong, J. C. Lang, [Reproductive integration in reef corals](https://doi.org/10.2307/1542018). *The Biological Bulletin*. **183**, 418–431 (1992).

29. Pygoscelis adeliae: BirdLife international: The IUCN red list of threatened species 2020: e.T22697758A157660553. *IUCN Red List of Threatened Species* (2020), doi:[10.2305/iucn.uk.2020-3.rlts.t22697758a157660553.en](https://doi.org/10.2305/iucn.uk.2020-3.rlts.t22697758a157660553.en).

30. Pygoscelis antarcticus: BirdLife international: The IUCN red list of threatened species 2020: e.T22697761A184807209. *IUCN Red List of Threatened Species* (2020), doi:[10.2305/iucn.uk.2020-3.rlts.t22697761a184807209.en](https://doi.org/10.2305/iucn.uk.2020-3.rlts.t22697761a184807209.en).

31. Pygoscelis papua: BirdLife international: The IUCN red list of threatened species 2020: e.T22697755A157664581. *IUCN Red List of Threatened Species* (2019), doi:[10.2305/iucn.uk.2020-3.rlts.t22697755a157664581.en](https://doi.org/10.2305/iucn.uk.2020-3.rlts.t22697755a157664581.en).

32. <https://www.cabi.org/isc/datasheet/47509>

33. J. P. Wourms, "Reproduction and development of sebastes in the context of the evolution of piscine viviparity" in *Rockfishes of the genus Sebastes: Their reproduction and early life history* (1991; <http://dx.doi.org/10.1007/978-94-011-3792-8_12>), pp. 111–126.

34. D. Moran, C. K. Smith, B. Gara, C. W. Poortenaar, [Reproductive behaviour and early development in yellowtail kingfish (seriola lalandi valenciennes 1833)](https://doi.org/10.1016/j.aquaculture.2006.10.005). *Aquaculture*. **262**, 95–104 (2007).

35. A. L. Gould, K. E. Dougan, S. T. Koenigbauer, P. V. Dunlap, [Life history of the symbiotically luminous cardinalfish siphamia tubifer (perciformes: apogonidae)](https://doi.org/10.1111/jfb.13063). *Journal of Fish Biology*. **89**, 1359–1377 (2016).

36. C. Bartilotti, A. Dos Santos, [The secret life of deep-sea shrimps: Ecological and evolutionary clues from the larval description ofSystellaspis debilis(caridea: oplophoridae)](https://doi.org/10.7717/peerj.7334). *PeerJ*. **7**, e7334 (2019).

37. Uria aalge: BirdLife international: The IUCN red list of threatened species 2018: e.T22694841A132577296. *IUCN Red List of Threatened Species* (2018), doi:[10.2305/iucn.uk.2018-2.rlts.t22694841a132577296.en](https://doi.org/10.2305/iucn.uk.2018-2.rlts.t22694841a132577296.en).

38. Zalophus wollebaeki: Trillmich, f.: The IUCN red list of threatened species 2015: e.T41668A45230540. *IUCN Red List of Threatened Species* (2014), doi:[10.2305/iucn.uk.2015-2.rlts.t41668a45230540.en](https://doi.org/10.2305/iucn.uk.2015-2.rlts.t41668a45230540.en).

39. M. E. Bushnell, J. T. Claisse, C. W. Laidley, [Lunar and seasonal patterns in fecundity of an indeterminate, multiple‐spawning surgeonfish, the yellow tang zebrasoma flavescens](https://doi.org/10.1111/j.1095-8649.2010.02569.x). *Journal of Fish Biology*. **76**, 1343–1361 (2010).

40. J. H. Choat, "Spawning aggregations in reef fishes; ecological and evolutionary processes" in *Reef Fish Spawning Aggregations: Biology, Research and Management* (2012; <http://dx.doi.org/10.1007/978-94-007-1980-4_4>), pp. 85–116.

41. G. V. Clucas, J. L. Younger, D. Kao, L. Emmerson, C. Southwell, B. Wienecke, A. D. Rogers, C. Bost, G. D. Miller, M. J. Polito, P. Lelliott, J. Handley, S. Crofts, R. A. Phillips, M. J. Dunn, K. J. Miller, T. Hart, [Comparative population genomics reveals key barriers to dispersal in southern ocean penguins](https://doi.org/10.1111/mec.14896). *Molecular Ecology*. **27**, 4680–4697 (2018).

42. G. R. Hoff, [Identification of multiple nursery habitats of skates in the eastern bering sea](https://doi.org/10.1111/jfb.12939). *Journal of Fish Biology*. **88**, 1746–1757 (2024).

43. B. Planque, [Projecting the future state of marine ecosystems, “la grande illusion”?](https://doi.org/10.1093/icesjms/fsv155) *ICES Journal of Marine Science: Journal du Conseil*. **73**, 204–208 (2024).

44. J. M. Gonz’alez-Irusta, P. J. Wright, [Spawning grounds of atlantic cod (gadus morhua) in the north sea](https://doi.org/10.1093/icesjms/fsv180). *ICES Journal of Marine Science: Journal du Conseil*. **73**, 304–315 (2015).

45. Laguncularia racemosa: Ellison, a., farnsworth, e. & moore, g.: The IUCN red list of threatened species 2010: e.T178798A7609219. *IUCN Red List of Threatened Species* (2007), doi:[10.2305/iucn.uk.2010-2.rlts.t178798a7609219.en](https://doi.org/10.2305/iucn.uk.2010-2.rlts.t178798a7609219.en).

46. B. A. McIntyre, E. E. McPhee-Shaw, M. B. A. Hatch, S. M. Arellano, [Location matters: Passive and active factors affect the vertical distribution of olympia oyster (ostrea lurida) larvae](https://doi.org/10.1007/s12237-020-00771-8). *Estuaries and Coasts*. **44**, 199–213 (2020).

47. L. E. Timm, T. L. Jackson, J. A. Browder, H. D. Bracken-Grissom, Population genomics of the commercially important gulf of mexico pink shrimp farfantepenaeus duorarum (burkenroad, 1939) support models of juvenile transport around the florida peninsula. *Frontiers in Ecology and Evolution*. **9** (2021), doi:[10.3389/fevo.2021.659134](https://doi.org/10.3389/fevo.2021.659134).

48. A. A. Hohn, A. J. Read, S. Fernandez, O. Vidal, L. T. Findley, [Life history of the vaquita, phocoena sinus (phocoenidae, cetacea)](https://doi.org/10.1111/j.1469-7998.1996.tb05450.x). *Journal of Zoology*. **239**, 235–251 (1996).

49. T. Brunel, C. J. G. van Damme, M. Samson, M. Dickey?Collas, [Quantifying the influence of geography and environment on the northeast atlantic mackerel spawning distribution](https://doi.org/10.1111/fog.12242). *Fisheries Oceanography*. **27**, 159–173 (2017).

50. K. Takano, A. Takemura, M. Furihata, T. Nakanishi, A. Hara, "Annual reproductive and spawning cycles of female sebastiscus marmoratus" in *Developments in environmental biology of fishes* (Springer Netherlands, 1991; <http://dx.doi.org/10.1007/978-94-011-3792-8_5>), pp. 39–48.

51. W. B. Driggers, B. S. Frazier, D. H. Adams, G. F. Ulrich, C. M. Jones, E. R. Hoffmayer, M. D. Campbell, [Site fidelity of migratory bonnethead sharks sphyrna tiburo (l. 1758) to specific estuaries in south carolina, USA](https://doi.org/10.1016/j.jembe.2014.05.006). *Journal of Experimental Marine Biology and Ecology*. **459**, 61–69 (2014).

52. D. E. LEE, C. L. ABRAHAM, P. M. WARZYBOK, R. W. BRADLEY, W. J. SYDEMAN, [AGE-SPECIFIC SURVIVAL, BREEDING SUCCESS, AND RECRUITMENT IN COMMON MURRES (URIA AALGE) OF THE CALIFORNIA CURRENT SYSTEM](https://doi.org/10.1525/auk.2008.07007). *The Auk*. **125**, 316–325 (2008).

53. <https://en.wikipedia.org/wiki/Thick-billed_murre>

54. R. K. Cowen, S. Sponaugle, "Relationships between early life history traits and recruitment among coral reef fishes" in *Early Life History and Recruitment in Fish Populations* (1997; <http://dx.doi.org/10.1007/978-94-009-1439-1_15>), pp. 423–449.

55. M. W. Miller, A. J. Bright, R. E. Pausch, D. E. Williams, [Larval longevity and competency patterns of caribbean reef-building corals](https://doi.org/10.7717/peerj.9705). *PeerJ*. **8**, e9705 (2020).

56. D. E. Morse, N. Hooker, A. N. C. Morse, R. A. Jensen, [Control of larval metamorphosis and recruitment in sympatric agariciid corals](https://doi.org/10.1016/0022-0981(88)90027-5). *Journal of Experimental Marine Biology and Ecology*. **116**, 193–217 (1988).

57. N. K. Dulvy, J. D. Reynolds, [Evolutionary transitions among egg–laying, live–bearing and maternal inputs in sharks and rays](https://doi.org/10.1098/rspb.1997.0181). *Proceedings of the Royal Society of London. Series B: Biological Sciences*. **264**, 1309–1315 (2024).

58. P. Jivoff, A. H. Hines, [Effect of female molt stage and sex ratio on courtship behavior of the blue crab callinectes sapidus](https://doi.org/10.1007/s002270050345). *Marine Biology*. **131**, 533–542 (2024).

59. A. L. Moran, [Spawning and larval development of the black turban snail tegula funebralis (prosobranchia: trochidae)](https://doi.org/10.1007/s002270050074). *Marine Biology*. **128**, 107–114 (1997).

60. J. S. Nelson, T. C. Grande, M. V. H. Wilson, Fishes of the world (2024), doi:[10.1002/9781119174844](https://doi.org/10.1002/9781119174844).

61. A. Arkhipkin, [Age and growth of planktonic squids cranchia scabra and liocranchia reinhardti (cephalopoda, cranchiidae) in epipelagic waters of the central-east atlantic](https://doi.org/10.1093/plankt/18.9.1675). *Journal of Plankton Research*. **18**, 1675–1683 (1996).

62. E. S. Chang, M. E. Orive, P. Cartwright, [Nonclonal coloniality: Genetically chimeric colonies through fusion of sexually produced polyps in the hydrozoanEctopleura larynx](https://doi.org/10.1002/evl3.68). *Evolution Letters*. **2**, 442–455 (2018).

63. J. J. Grigor, M. S. Schmid, L. Fortier, [Growth and reproduction of the chaetognaths eukrohnia hamata and parasagitta elegans in the canadian arctic ocean: Capital breeding versus income breeding](https://doi.org/10.1093/plankt/fbx045). *Journal of Plankton Research*. **39**, 910–929 (2017).

64. A. Schlesinger, E. Kramarsky-Winter, H. Rosenfeld, R. Armoza-Zvoloni, Y. Loya, [Sexual plasticity and self-fertilization in the sea anemone aiptasia diaphana](https://doi.org/10.1371/journal.pone.0011874). *PLoS ONE*. **5**, e11874 (2010).

65. J. H. Churchill, J. P. Kritzer, M. J. Dean, J. H. Grabowski, G. D. Sherwood, [Patterns of larval-stage connectivity of atlantic cod (gadus morhua) within the gulf of maine in relation to current structure and a proposed fisheries closure](https://doi.org/10.1093/icesjms/fsw139). *ICES Journal of Marine Science*. **74**, 20–30 (2016).

66. A. Md. Ye, F. Md. Yu, N. F. M. Ikhsan, Z. Hassan, Lunar cycle drives family-specific larval fish assemblages in the tropical nearshore ecosystem (2024), doi:[10.21203/rs.3.rs-2652251/v1](https://doi.org/10.21203/rs.3.rs-2652251/v1).

67. Y. Nakajima, Y. Zayasu, C. Shinzato, N. Satoh, S. Mitarai, [Genetic differentiation and connectivity of morphological types of the broadcast‐spawning coral galaxea fascicularis in the nansei islands, japan](https://doi.org/10.1002/ece3.1981). *Ecology and Evolution*. **6**, 1457–1469 (2016).

68. <https://fishesofaustralia.net.au/home/species/2129#moreinfo>

69. E. M. Moore, T. G. Langley, J. S. Goldstein, W. H. Watson, [American lobster, homarus americanus, reproduction and recruitment in a new england estuary](https://doi.org/10.1007/s12237-020-00759-4). *Estuaries and Coasts*. **43**, 2141–2151 (2020).

70. R. I. Lonard, F. W. Judd, H. R. DeYoe, R. Stalter, "Biology and ecology of the halophyte laguncularia racemosa (l.) gaertn. F.: A review" in *Handbook of halophytes* (Springer International Publishing, 2020; <http://dx.doi.org/10.1007/978-3-030-17854-3_71-1>), pp. 1–15.

71. ???, Y.-U. Kim, ???, B.-G. Kim, J.-M. Kim, H.-T. Heo, Embryonic developmen larvae and juveniles of the small yellow croaker (larimichthys polyactis) reared in aquarium. *Korean Journal of Fisheries and Aquatic Sciences*. **37**, 478–484 (2004).

72. S. Goffredo, V. Airi, J. Radeti’c, F. Zaccanti, [Sexual reproduction of the solitary sunset cup coral leptopsammia pruvoti (scleractinia, dendrophylliidae) in the mediterranean. 2. Quantitative aspects of the annual reproductive cycle](https://doi.org/10.1007/s00227-005-0137-8). *Marine Biology*. **148**, 923–931 (2005).

73. J. L. Padilla-Gamiño, R. R. Bidigare, D. J. Barshis, A. Alamaru, L. H’edouin, X. Hern’andez-Pech, F. Kandel, S. Leon Soon, M. S. Roth, L. J. Rodrigues, A. G. Grottoli, C. Portocarrero, S. A. Wagenhauser, F. Buttler, R. D. Gates, [Are all eggs created equal? A case study from the hawaiian reef-building coral montipora capitata](https://doi.org/10.1007/s00338-012-0957-1). *Coral Reefs*. **32**, 137–152 (2012).

74. [R.t. Hanlon, j.b. Messenger cephalopod behaviour. Xvi, 232 p. Cambridge university press, 1996. Price £50.00 (£20.95 paperback)](https://doi.org/10.1017/s0025315400044623). *Journal of the Marine Biological Association of the United Kingdom*. **78**, 1392–1392 (1998).

75. S. W. Davies, M. E. Strader, J. T. Kool, C. D. Kenkel, M. V. Matz, [Modeled differences of coral life-history traits influence the refugium potential of a remote caribbean reef](https://doi.org/10.1007/s00338-017-1583-8). *Coral Reefs*. **36**, 913–925 (2017).

76. J. S. Barber, J. E. Dexter, S. K. Grossman, C. M. Greiner, J. T. Mcardle, [Low temperature brooding of olympia oysters (ostrea lurida) in northern puget sound](https://doi.org/10.2983/035.035.0209). *Journal of Shellfish Research*. **35**, 351–357 (2016).

77. C. Carreras, A. Garc’ıa?Cisneros, O. S. Wangensteen, V. Ord’oñez, C. Palac’ın, M. Pascual, X. Turon, [East is east and west is west: Population genomics and hierarchical analyses reveal genetic structure and adaptation footprints in the keystone species paracentrotus lividus (echinoidea)](https://doi.org/10.1111/ddi.13016). *Diversity and Distributions*. **26**, 382–398 (2019).

78. J. M. Leis, "The pelagic stage of reef fishes: The larval biology of coral reef fishes" in *The Ecology of Fishes on Coral Reefs* (1991; <http://dx.doi.org/10.1016/b978-0-08-092551-6.50013-1>), pp. 183–230.

79. J. Grahame, [<I>reproduction and development of marine invertebrates of the northern pacific coast. Data and methods for the study of eggs, embryos, and larvae</i>. Megumi f. strathmann](https://doi.org/10.1086/416265). *The Quarterly Review of Biology*. **64**, 197–197 (2024).

80. M. G. Frick, H. R. Martins, A. B. Bolten, K. A. Bjorndal, K. L. Williams, [Diet and fecundity of columbus crabs, planes minutus, associated with oceanic-stage loggerhead sea turtles, caretta caretta, and inanimate flotsam](https://doi.org/10.1651/c-2440). *Journal of Crustacean Biology*. **24**, 350–355 (2004).

81. F. G. W. Smith, Atlantic reef corals, a handbook of the common reef and shallow-water corals of bermuda, florida, the west indies, and brazil; photos. By fredrick m. bayer. (1948), doi:[10.5962/bhl.title.6809](https://doi.org/10.5962/bhl.title.6809).

82. and William N S Arlidge, Coral reef viruses in kane’ohe bay, hawai’i: Abundance, diversity &amp; environmental drivers (2021), doi:[10.26686/wgtn.17008594](https://doi.org/10.26686/wgtn.17008594).

83. <https://www.sealifebase.ca/summary/Pyroteuthis-margaritifera.html>

84. P. Kerrison, H. N. Le, [Environmental factors on egg liberation and germling production of sargassum muticum](https://doi.org/10.1007/s10811-015-0580-y). *Journal of Applied Phycology*. **28**, 481–489 (2015).

85. M. J. A. Vermeij, K. L. Barott, A. E. Johnson, K. L. Marhaver, [Release of eggs from tentacles in a caribbean coral](https://doi.org/10.1007/s00338-010-0595-4). *Coral Reefs*. **29**, 411–411 (2024).

86. <https://animaldiversity.org/accounts/Vampyroteuthis_infernalis>

87. P. Foxton, [Observations on the early development and hatching of the eggs of acanthephyra purpurea a. Milne-edwards](https://doi.org/10.1163/156854064x00641). *Crustaceana*. **6**, 235–237 (1964).

88. E. M. Graham, A. H. Baird, S. R. Connolly, M. A. Sewell, B. L. Willis, [Rapid declines in metabolism explain extended coral larval longevity](https://doi.org/10.1007/s00338-012-0999-4). *Coral Reefs*. **32**, 539–549 (2013).

89. B. H. Cornwell, [Gene flow in the anemone anthopleura elegantissima limits signatures of local adaptation across an extensive geographic range](https://doi.org/10.1111/mec.15506). *Molecular Ecology*. **29**, 2550–2566 (2020).

90. G. L. Kooyman, [The penguins spheniscidae. Bird families of the world, volume 2.tony d. williams](https://doi.org/10.1086/419312). *The Quarterly Review of Biology*. **71**, 133–134 (1996).

91. I. Wolfowicz, S. Baumgarten, P. A. Voss, E. A. Hambleton, C. R. Voolstra, M. Hatta, A. Guse, Aiptasia sp. Larvae as a model to reveal mechanisms of symbiont selection in cnidarians. *Scientific Reports*. **6** (2016), doi:[10.1038/srep32366](https://doi.org/10.1038/srep32366).

92. D. Macedo, I. Caballero, M. Mateos, R. Leblois, S. McCay, L. A. Hurtado, [Population genetics and historical demographic inferences of the blue crabCallinectes sapidusin the US based on microsatellites](https://doi.org/10.7717/peerj.7780). *PeerJ*. **7**, e7780 (2019).

93. E. Haber, [Rare and endangered plants of canada: Report of the plants subcommittee, the committee on the status of endangered wildlife in canada (COSEWIC)](https://doi.org/10.5962/p.355667). *The Canadian field-naturalist*. **100**, 400–403 (1986).

94. <https://www.nature.com/articles/s41598-019-45167-2>

95. A. Md. Ye, F. Md. Yu, N. F. M. Ikhsan, Z. Hassan, Lunar cycle drives family-specific larval fish assemblages in the tropical nearshore ecosystem (2024), doi:[10.21203/rs.3.rs-2652251/v1](https://doi.org/10.21203/rs.3.rs-2652251/v1).

96. C. P. Marquis, A. H. Baird, R. de Nys, C. Holmström, N. Koziumi, [An evaluation of the antimicrobial properties of the eggs of 11 species of scleractinian corals](https://doi.org/10.1007/s00338-005-0473-7). *Coral Reefs*. **24**, 248–253 (2005).

97. E. R. Annis, L. S. Incze, R. S. Steneck, N. Wolff, [Estimates of in situ larval development time for the lobster, homarus americanus](https://doi.org/10.1651/s-2758.1). *Journal of Crustacean Biology*. **27**, 454–462 (2007).

98. S. D. Cairns, [Species richness of recent scleractinia](https://doi.org/10.5479/si.00775630.459.1). *Atoll Research Bulletin*. **459**, 1–46 (1999).

99. A. H. Baird, J. R. Guest, B. L. Willis, [Systematic and biogeographical patterns in the reproductive biology of scleractinian corals](https://doi.org/10.1146/annurev.ecolsys.110308.120220). *Annual Review of Ecology, Evolution, and Systematics*. **40**, 551–571 (2009).

100. R. Villanueva, E. A. G. Vidal, F. ’A. Fern’andez-’Alvarez, J. Nabhitabhata, [Early mode of life and hatchling size in cephalopod molluscs: Influence on the species distributional ranges](https://doi.org/10.1371/journal.pone.0165334). *PLOS ONE*. **11**, e0165334 (2016).

101. A. Hettinger, E. Sanford, T. M. Hill, J. D. Hosfelt, A. D. Russell, B. Gaylord, [The influence of food supply on the response of olympia oyster larvae to ocean acidification](https://doi.org/10.5194/bg-10-6629-2013). *Biogeosciences*. **10**, 6629–6638 (2013).

102. L. Fenaux, C. Cellario, M. Etienne, [Croissance de la larve de l’oursin paracentrotus lividus](https://doi.org/10.1007/bf00399021). *Marine Biology*. **86**, 151–157 (1985).

103. [J. C. VON VAUPEL KLEIN, *Crustaceana*. **55**, 320 (1988)](https://doi.org/10.1163/156854088x00456).

104. J. B. Pfaller, A. C. Payton, K. A. Bjorndal, A. B. Bolten, S. F. McDaniel, [Hitchhiking the high seas: Global genomics of rafting crabs](https://doi.org/10.1002/ece3.4694). *Ecology and Evolution*. **9**, 957–974 (2019).

105. A. Moreno, A. Dos Santos, U. Piatkowski, A. M. P. Santos, H. Cabral, [Distribution of cephalopod paralarvae in relation to the regional oceanography of the western iberia](https://doi.org/10.1093/plankt/fbn103). *Journal of Plankton Research*. **31**, 73–91 (2008).

106. E. J. Guerra?Castro, J. E. Conde, A. Barcelo, J. J. Cruz?Motta, [Variation in fouling assemblages associated with prop roots of rhizophora mangle l. In the caribbean: The role of neutral and niche processes](https://doi.org/10.1111/aec.13071). *Austral Ecology*. **46**, 991–1007 (2021).

107. P. Baweja, S. Kumar, D. Sahoo, I. Levine, "Biology of seaweeds" in *Seaweed in health and disease prevention* (Elsevier, 2016; <http://dx.doi.org/10.1016/B978-0-12-802772-1.00003-8>), pp. 41–106.

108. K. N. Kosobokova, A. I. Isachenko, [The gonad maturation and size structure of the population of abundant planktonic predator eukrohnia hamata (möbius, 1875) (chaetognatha) in the eurasian basin of the arctic ocean in summer](https://doi.org/10.1134/s1063074017010072). *Russian Journal of Marine Biology*. **43**, 25–33 (2017).

109. M. Bucher, I. Wolfowicz, P. A. Voss, E. A. Hambleton, A. Guse, Development and symbiosis establishment in the cnidarian endosymbiosis model aiptasia sp. *Scientific Reports*. **6** (2016), doi:[10.1038/srep19867](https://doi.org/10.1038/srep19867).

110. A. Md. Ye, F. Md. Yu, N. F. M. Ikhsan, Z. Hassan, Lunar cycle drives family-specific larval fish assemblages in the tropical nearshore ecosystem (2024), doi:[10.21203/rs.3.rs-2652251/v1](https://doi.org/10.21203/rs.3.rs-2652251/v1).

111. R. C. Babcock, A. J. Heyward, [Larval development of certain gamete-spawning scleractinian corals](https://doi.org/10.1007/bf00298178). *Coral Reefs*. **5**, 111–116 (1986).

112. B. D. Limer, J. Bloomberg, D. M. Holstein, The influence of eddies on coral larval retention in the flower garden banks. *Frontiers in Marine Science*. **7** (2020), doi:[10.3389/fmars.2020.00372](https://doi.org/10.3389/fmars.2020.00372).

113. S. K. Grossman, E. E. Grossman, J. S. Barber, S. K. Gamblewood, S. C. Crosby, [Distribution and transport of olympia oyster ostrea lurida larvae in northern puget sound, washington](https://doi.org/10.2983/035.039.0204). *Journal of Shellfish Research*. **39**, 215 (2020).

114. J. H. Vandermeulen, [Studies on reef corals. III. Fine structural changes of calicoblast cells in pocillopora damicornis during settling and calcification](https://doi.org/10.1007/bf00390649). *Marine Biology*. **31**, 69–77 (1975).

115. F. G. W. Smith, Atlantic reef corals, a handbook of the common reef and shallow-water corals of bermuda, florida, the west indies, and brazil; photos. By fredrick m. bayer. (1948), doi:[10.5962/bhl.title.6809](https://doi.org/10.5962/bhl.title.6809).

116. and William N S Arlidge, Coral reef viruses in kane’ohe bay, hawai’i: Abundance, diversity &amp; environmental drivers (2021), doi:[10.26686/wgtn.17008594](https://doi.org/10.26686/wgtn.17008594).

117. H. M. Austin, [The eggs and planktonic stages of british marine fishes.f. S. russell](https://doi.org/10.1086/409937). *The Quarterly Review of Biology*. **52**, 216–216 (1977).

118. J. P. Wourms, [Reproduction and development ofSebastes in the context of the evolution of piscine viviparity](https://doi.org/10.1007/bf02296882). *Environmental Biology of Fishes*. **30**, 111–126 (1991).

119. <https://www.sealifebase.ca/summary/Stephanocoenia-intersepta.html>).

120. D. T. Wilson, M. I. McCormick, [Microstructure of settlement-marks in the otoliths of tropical reef fishes](https://doi.org/10.1007/s002270050522). *Marine Biology*. **134**, 29–41 (2024).

121. G. B. Nanninga, P. Saenz-Agudelo, P. Zhan, I. Hoteit, M. L. Berumen, [Not finding nemo: Limited reef-scale retention in a coral reef fish](https://doi.org/10.1007/s00338-015-1266-2). *Coral Reefs*. **34**, 383–392 (2015).

122. E. Macpherson, N. Raventos, [Relationship between pelagic larval duration and geographic distribution of mediterranean littoral fishes](https://doi.org/10.3354/meps327257). *Marine Ecology Progress Series*. **327**, 257–265 (2006).

123. Ronald. E. Thresher, P. L. Colin, L. J. Bell, [Planktonic duration, distribution and population structure of western and central pacific damselfishes (pomacentridae)](https://doi.org/10.2307/1445439). *Copeia*. **1989**, 420 (1989).

124. I. Palomera, B. Morales-Nin, J. Lleonart, [Larval growth of anchovy, engraulis encrasicolus, in the western mediterranean sea](https://doi.org/10.1007/bf00391991). *Marine Biology*. **99**, 283–291 (2024).

125. <https://www.sealifebase.ca/summary/Eukrohnia-hamata.html>

126. S. Campana, [Year-class strength and growth rate in young atlantic cod gadus morhua](https://doi.org/10.3354/meps135021). *Marine Ecology Progress Series*. **135**, 21–26 (1996).

127. R. SUWA, M. NAKAMURA, [A precise comparison of developmental series of oocyte growth and oocyte maturation between real-oocytes and pseudo-oocytes in the coral &lt;i&gt;galaxea fascicularis&lt;/i&gt;](https://doi.org/10.3755/galaxea.20.1_1) *Galaxea, Journal of Coral Reef Studies*. **20**, 1–7 (2018).

128. R. E. Thresher, E. B. Brothers, [REPRODUCTIVE ECOLOGY AND BIOGEOGRAPHY OF INDO-WEST PACIFIC ANGELFISHES (PISCES: POMACANTHIDAE)](https://doi.org/10.1111/j.1558-5646.1985.tb00429.x). *Evolution*. **39**, 878–887 (1985).

129. R. N., M. E., [Planktonic larval duration and settlement marks on the otoliths of mediterranean littoral fishes](https://doi.org/10.1007/s002270000535). *Marine Biology*. **138**, 1115–1120 (2001).

130. [reported for Lateolabrax japonicus : https://en.wikipedia.org/wiki/Japanese\_sea\_bass](file:///Users/eric/github/DivDiv_Refs/reported%20for%20Lateolabrax%20japonicus%20:%20https:/en.wikipedia.org/wiki/Japanese_sea_bass)

131. <https://www.marlin.ac.uk/species/detail/1285>

132. J. B. Puritz, J. R. Gold, D. S. Portnoy, Fine-scale partitioning of genomic variation among recruits in an exploited fishery: Causes and consequences. *Scientific Reports*. **6** (2016), doi:[10.1038/srep36095](https://doi.org/10.1038/srep36095).

133. G. Alexander, J. R. Hancock, A. S. Huffmyer, S. B. Matsuda, [Larval thermal conditioning does not improve post-settlement thermal tolerance in the dominant reef-building coral, montipora capitata](https://doi.org/10.1007/s00338-022-02234-x). *Coral Reefs*. **41**, 333–342 (2022).

134. R. C. Williams, B. C. Jackson, L. Duvaux, D. A. Dawson, T. Burke, W. Sinclair, [The genetic structure of nautilus pompilius populations surrounding australia and the philippines](https://doi.org/10.1111/mec.13255). *Molecular Ecology*. **24**, 3316–3328 (2015).

135. S. L’opez, X. Turon, E. Montero, C. Palac’ın, C. Duarte, I. Tarjuelo, [Larval abundance, recruitment and early mortality in paracentrotus lividus (echinoidea). Interannual variability and plankton-benthos coupling](https://doi.org/10.3354/meps172239). *Marine Ecology Progress Series*. **172**, 239–251 (1998).

136. M. L. Reaka-Kudla, and, The ecology of coral reefs : Results of a workshop on coral reef ecology held by the american society of zoologists, philadelphia, pennsylvania, december 1983 / edited by marjorie l. reaka. (1985), doi:[10.5962/bhl.title.4015](https://doi.org/10.5962/bhl.title.4015).

137. M. Oshima, D. Robert, Y. Kurita, M. Yoneda, O. Tominaga, T. Tomiyama, Y. Yamashita, S. Uehara, [Do early growth dynamics explain recruitment success in japanese flounder paralichthys olivaceus off the pacific coast of northern japan?](https://doi.org/10.1016/j.seares.2009.12.002) *Journal of Sea Research*. **64**, 94–101 (2010).

138. J. B. Pfaller, M. A. Gil, [Sea turtle symbiosis facilitates social monogamy in oceanic crabs via refuge size](https://doi.org/10.1098/rsbl.2016.0607). *Biology Letters*. **12**, 20160607 (2016).

139. A. L. Primo, A. C. Vaz, D. Crespo, F. Costa, M. Pardal, F. Martinho, [Contrasting links between growth and survival in the early life stages of two flatfish species](https://doi.org/10.1016/j.ecss.2021.107314). *Estuarine, Coastal and Shelf Science*. **254**, 107314 (2021).

140. V. R. Cumbo, T. Y. Fan, P. J. Edmunds, [Effects of exposure duration on the response of pocillopora damicornis larvae to elevated temperature and high pCO2](https://doi.org/10.1016/j.jembe.2012.10.019). *Journal of Experimental Marine Biology and Ecology*. **439**, 100–107 (2013).

141. M. L. Reaka-Kudla, and, The ecology of coral reefs : Results of a workshop on coral reef ecology held by the american society of zoologists, philadelphia, pennsylvania, december 1983 / edited by marjorie l. reaka. (1985), doi:[10.5962/bhl.title.4015](https://doi.org/10.5962/bhl.title.4015).

142. E. P., G. R., G. D., [The biology of larvae from the reef coral porites astreoides , and their response to temperature disturbances](https://doi.org/10.1007/s002270100634). *Marine Biology*. **139**, 981–989 (2001).

143. C. D. Storlazzi, M. van Ormondt, Y.-L. Chen, E. P. L. Elias, Modeling fine-scale coral larval dispersal and interisland connectivity to help designate mutually-supporting coral reef marine protected areas: Insights from maui nui, hawaii. *Frontiers in Marine Science*. **4** (2017), doi:[10.3389/fmars.2017.00381](https://doi.org/10.3389/fmars.2017.00381).

144. L. E. Timm, H. D. Bracken-Grissom, A. Sosnowski, M. Breitbart, M. Vecchione, H. Judkins, [Population genomics of three deep-sea cephalopod species reveals connectivity between the gulf of mexico and northwestern atlantic ocean](https://doi.org/10.1016/j.dsr.2020.103222). *Deep Sea Research Part I: Oceanographic Research Papers*. **158**, 103222 (2020).

145. Rafting to bombay. *Rafting to Bombay* (2009), doi:[10.5040/9781350919587](https://doi.org/10.5040/9781350919587).

146. B. Villamor, M. Bernal, C. Hern’andez, [Models describing mackerel (scomber scombrus) early life growth in the north and northwest of the iberian peninsula in 2000](https://doi.org/10.3989/scimar.2004.68n4571). *Scientia Marina*. **68**, 571–583 (2004).

147. B. W. Coad, ["The rockfishes of the northeast pacific" by milton s. Love, mary yoklavich, and lyman thorsteinson. 2002. [Book review]](https://doi.org/10.22621/cfn.v117i2.715). *The Canadian Field-Naturalist*. **117**, 323 (2024).

148. A. L. Gould, P. V. Dunlap, Shedding light on specificity: Population genomic structure of a symbiosis between a coral reef fish and luminous bacterium. *Frontiers in Microbiology*. **10** (2019), doi:[10.3389/fmicb.2019.02670](https://doi.org/10.3389/fmicb.2019.02670).

149. G. M. Wellington, B. C. Victor, [Planktonic larval duration of one hundred species of pacific and atlantic damselfishes (pomacentridae)](https://doi.org/10.1007/bf00541659). *Marine Biology*. **101**, 557–567 (1989).

150. P. R. Stephens, J. Z. Young, [The statocyst of vampyroteuthis infernalis (mollusca: cephalopoda)](https://doi.org/10.1111/j.1469-7998.1976.tb04704.x). *Journal of Zoology*. **180**, 565–588 (1976).

151. G. E. Pickford, "Vertical distribution." in *Vampyroteuthis infernalis Chun* (2024; <http://dx.doi.org/10.1163/9789004628397_006>), pp. 18–24.

152. C. K. Callan, A. I. Burgess, C. R. Rothe, R. Touse, [Development of improved feeding methods in the culture of yellow tang, zebrasoma flavescens](https://doi.org/10.1111/jwas.12496). *Journal of the World Aquaculture Society*. **49**, 493–503 (2018).

153. D. T. Wilson, M. I. McCormick, [Microstructure of settlement-marks in the otoliths of tropical reef fishes](https://doi.org/10.1007/s002270050522). *Marine Biology*. **134**, 29–41 (1999).

154. S. A. Sudnik, T. Falkenhaug, [Maturation, fecundity and embryos development in three deep-water shrimps (decapoda: Caridea: Pasiphaeidae, oplophoridae) along the mid-atlantic ridge from iceland to the azores](https://doi.org/10.15298/arthsel.24.4.03). *rej*. **24**, 401416–0 (2015).

155. J. N. Grim, [The vestibuliferan ciliate balantidium acanthuri n. Sp. From two species of the surgeonfish, genus acanthurus](https://doi.org/10.1016/s0003-9365(89)80006-5). *Archiv für Protistenkunde*. **137**, 157–160 (2024).

156. E. A. Agudo-Adriani, J. Cappelletto, F. Cavada-Blanco, A. Croquer, [Colony geometry and structural complexity of the endangered speciesAcropora cervicornispartly explains the structure of their associated fish assemblage](https://doi.org/10.7717/peerj.1861). *PeerJ*. **4**, e1861 (2016).

157. A. L. Zubillaga, L. M. M’arquez, A. Cr’oquer, C. Bastidas, [Ecological and genetic data indicate recovery of the endangered coral acropora palmata in los roques, southern caribbean](https://doi.org/10.1007/s00338-007-0291-1). *Coral Reefs*. **27**, 63–72 (2007).

158. <https://en.wikipedia.org/wiki/Elkhorn_coral>

159. J. Voss, L. Richardson, [Coral diseases near lee stocking island, bahamas: Patterns and potential drivers](https://doi.org/10.3354/dao069033). *Diseases of Aquatic Organisms*. **69**, 33–40 (2006).

160. <https://coralpedia.bio.warwick.ac.uk/en/corals/agaricia_undata>

161. <https://en.wikipedia.org/wiki/Bartholomea_annulata>

162. J. A. Thia, K. McGuigan, L. Liggins, W. F. Figueira, C. E. Bird, A. Mather, J. L. Evans, C. Riginos, [Genetic and phenotypic variation exhibit both predictable and stochastic patterns across an intertidal fish metapopulation](https://doi.org/10.1111/mec.15829). *Molecular Ecology*. **30**, 4392–4414 (2021).

163. E. Beukhof, T. S. Dencker, M. L. D. Palomares, A. Maureaud, A trait collection of marine fish species from north atlantic and northeast pacific continental shelf seas (2019), doi:[10.1594/PANGAEA.900866](https://doi.org/10.1594/PANGAEA.900866).

164. <https://www.nwf.org/Educational-Resources/Wildlife-Guide/Invertebrates/Blue-Crab>

165. <https://en.wikipedia.org/wiki/Tegula_funebralis>

166. <https://en.wikipedia.org/wiki/Cranchiidae>

167. E. Jansson, F. Besnier, K. Malde, C. Andr’e, G. Dahle, K. A. Glover, Genome wide analysis reveals genetic divergence between goldsinny wrasse populations ﻿ (2020), doi:[10.21203/rs.2.22229/v1](https://doi.org/10.21203/rs.2.22229/v1).

168. <https://en.wikipedia.org/wiki/Ectopleura_larynx>

169. <https://en.wikipedia.org/wiki/Exaiptasia>

170. J. E. N. V. and, Hermatypic corals of japan / (1992), doi:[10.5962/bhl.title.60634](https://doi.org/10.5962/bhl.title.60634).

171. <https://en.wikipedia.org/wiki/American_lobster>

172. H. K. Lim, M. H. Le, C. M. An, S. Y. Kim, M. S. Park, Y. J. Chang, [Reproductive cycle of yellow croaker larimichthys polyactis in southern waters off korea](https://doi.org/10.1007/s12562-010-0288-5). *Fisheries Science*. **76**, 971–980 (2010).

173. S. Goffredo, J. Radeti’c, V. Airi, F. Zaccanti, [Sexual reproduction of the solitary sunset cup coral leptopsammia pruvoti (scleractinia: Dendrophylliidae) in the mediterranean. 1. Morphological aspects of gametogenesis and ontogenesis](https://doi.org/10.1007/s00227-005-1567-z). *Marine Biology*. **147**, 485–495 (2005).

174. E. Caroselli, F. Zaccanti, G. Mattioli, G. Falini, O. Levy, Z. Dubinsky, S. Goffredo, [Growth and demography of the solitary scleractinian coral leptopsammia pruvoti along a sea surface temperature gradient in the mediterranean sea](https://doi.org/10.1371/journal.pone.0037848). *PLoS ONE*. **7**, e37848 (2012).

175. A. Nishikawa, R. A. Kinzie, K. Sakai, [Fragmentation and genotypic diversity of the scleractinian coralMontipora capitatain kaneohe bay, hawaii](https://doi.org/10.1017/s0025315408002865). *Journal of the Marine Biological Association of the United Kingdom*. **89**, 101–107 (2008).

176. <https://en.wikipedia.org/wiki/Chambered_nautilus>

177. R. Bak, E. Meesters, [Coral population structure:the hidden information of colony size-frequency distributions](https://doi.org/10.3354/meps162301). *Marine Ecology Progress Series*. **162**, 301–306 (1998).

178. P. S. E. zu Ermgassen, M. W. Gray, C. J. Langdon, M. D. Spalding, R. D. Brumbaugh, [Quantifying the historic contribution of olympia oysters to filtration in pacific coast (USA) estuaries and the implications for restoration objectives](https://doi.org/10.1007/s10452-013-9431-6). *Aquatic Ecology*. **47**, 149–161 (2013).

179. C. F. Boudouresque, M. Verlaque, "Paracentrotus lividus" in *Sea urchins: Biology and ecology* (Elsevier, 2013; <http://dx.doi.org/10.1016/B978-0-12-396491-5.00021-6>), pp. 297–327.

180. R. L. Brownell, L. T. Findley, O. Vidal, A. Robles, N. Silvia Manzanilla, [EXTERNAL MORPHOLOGY AND PIGMENTATION OF THE VAQUITA, PHOCOENA SINUS (CETACEA: MAMMALIA)](https://doi.org/10.1111/j.1748-7692.1987.tb00149.x). *Marine Mammal Science*. **3**, 22–30 (1987).

181. <https://en.wikipedia.org/wiki/Pisaster_ochraceus>

182. <https://en.wikipedia.org/wiki/Planes_minutus>

183. R. H. Richmond, [Energetics, competency, and long-distance dispersal of planula larvae of the coral pocillopora damicornis](https://doi.org/10.1007/bf00392790). *Marine Biology*. **93**, 527–533 (2024).

184. M. Sudek, T. M. Work, G. S. Aeby, S. K. Davy, [Histological observations in the hawaiian reef coral, porites compressa, affected by porites bleaching with tissue loss](https://doi.org/10.1016/j.jip.2012.07.004). *Journal of Invertebrate Pathology*. **111**, 121–125 (2012).

185. M. E. Flock, T. L. Hopkins, [Species composition, vertical distribution, and food habits of the sergestid shrimp assemblage in the eastern gulf of mexico](https://doi.org/10.2307/1549076). *Journal of Crustacean Biology*. **12**, 210–223 (2024).

186. <https://en.wikipedia.org/wiki/Rhizophora_mangle>

187. (2024), doi:[10.1371/journal.pone.0112057.g001](https://doi.org/10.1371/journal.pone.0112057.g001).

188. F. Vaux, L. K. Rasmuson, L. A. Kautzi, P. S. Rankin, M. T. O. Blume, K. A. Lawrence, S. Bohn, K. G. O’Malley, [Sex matters: Otolith shape and genomic variation in deacon rockfish (sebastes diaconus)](https://doi.org/10.1002/ece3.5763). *Ecology and Evolution*. **9**, 13153–13173 (2019).

189. G. W. Baeck, J. M. Jeong, Y. M. Yeo, S.-H. Huh, J. M. Park, [Length-weight and length-length relationships for 10 species of scorpionfishes (scorpaenidae) on the south coast of korea: LWRs and LLRs for 10 species of scorpionfishes](https://doi.org/10.1111/j.1439-0426.2012.01956.x). *Journal of Applied Ichthyology*. **28**, 677–679 (2012).

190. F. Griffiths, S. Brandt, [Distribution of mesopelagic decapod crustacea in and around a warm-core eddy in the tasman sea](https://doi.org/10.3354/meps012175). *Marine Ecology Progress Series*. **12**, 175–184 (1983).

191. <https://en.wikipedia.org/wiki/Common_murre>

192. A. V. Golikov, F. R. Ceia, R. M. Sabirov, J. D. Ablett, I. G. Gleadall, G. Gudmundsson, H. J. Hoving, H. Judkins, J. P’alsson, A. L. Reid, R. Rosas-Luis, E. K. Shea, R. Schwarz, J. C. Xavier, The first global deep-sea stable isotope assessment reveals the unique trophic ecology of vampire squid vampyroteuthis infernalis (cephalopoda). *Scientific Reports*. **9** (2019), doi:[10.1038/s41598-019-55719-1](https://doi.org/10.1038/s41598-019-55719-1).

193. L. E. Timm, L. M. Isma, M. W. Johnston, H. D. Bracken-Grissom, Comparative population genomics and biophysical modeling of shrimp migration in the gulf of mexico reveals current-mediated connectivity. *Frontiers in Marine Science*. **7** (2024), doi:[10.3389/fmars.2020.00019](https://doi.org/10.3389/fmars.2020.00019).

194. S. A. Sudnik, T. Falkenhaug, [Maturation, fecundity and embryos development in three deep-water shrimps (decapoda: Caridea: Pasiphaeidae, oplophoridae) along the mid-atlantic ridge from iceland to the azores](https://doi.org/10.15298/arthsel.24.4.03). *rej*. **24**, 401416–0 (2015).

195. T. Foster, J. Gilmour, Egg size and fecundity of biannually spawning corals at scott reef. *Scientific Reports*. **10** (2020), doi:[10.1038/s41598-020-68289-4](https://doi.org/10.1038/s41598-020-68289-4).

196. <http://www.ipcbee.com/vol22/11-CAAS2011-X026.pdf>

197. K. P. SEBENS, [THE REGULATION OF ASEXUAL REPRODUCTION AND INDETERMINATE BODY SIZE IN THE SEA ANEMONE<i>ANTHOPLEURA ELEGANTISSIMA</i>(BRANDT)](https://doi.org/10.2307/1540863). *The Biological Bulletin*. **158**, 370–382 (2024).

198. B. L. Jennison, [Reproduction in three species of sea anemones from key west, florida](https://doi.org/10.1139/z81-235). *Canadian Journal of Zoology*. **59**, 1708–1719 (1981).

199. <https://jmstt.ntou.edu.tw/journal/vol21/iss7/27>

200. D. J. Graham, R. Fulford, P. Biesiot, H. Perry, [Fecundity and egg diameter of primiparous and multiparous blue crab callinectes sapidus (brachyura: Portunidae) in mississippi waters](https://doi.org/10.1163/193724011x615325). *Journal of Crustacean Biology*. **32**, 49–56 (2012).

201. N. G. Emel’yanova, D. A. Pavlov, L. T. B. Thuan, [Hormonal stimulation of maturation and ovulation, gamete morphology, and raising of larvae in dascyllus trimaculatus (pomacentridae)](https://doi.org/10.1134/s0032945209030059). *Journal of Ichthyology*. **49**, 249–263 (2009).

202. N. Okubo, T. Mezaki, Y. Nozawa, Y. Nakano, Y.-T. Lien, H. Fukami, D. C. Hayward, E. E. Ball, [Comparative embryology of eleven species of stony corals (scleractinia)](https://doi.org/10.1371/journal.pone.0084115). *PLoS ONE*. **8**, e84115 (2013).

203. P. Ouellet, F. Plante, [An investigation of the sources of variability in american lobster (homarus americanus) eggs and larvae: Female size and reproductive status, and interannual and interpopulation comparisons](https://doi.org/10.1651/c-2467). *Journal of Crustacean Biology*. **24**, 481–495 (2004).

204. C.-B. Kang, J.-G. Myoung, Y.-U. Kim, H.-C. Kim, [Early osteological development and squamation in the spotted sea bass lateolabrax maculates (pisces: lateolabracidae)](https://doi.org/10.5657/kfas.2012.0271). *Korean Journal of Fisheries and Aquatic Sciences*. **45**, 271–282 (2024).

205. K. Kasimatis, C. Riginos, [A phylogenetic analysis of egg size, clutch size, spawning mode, adult body size, and latitude in reef fishes](https://doi.org/10.1007/s00338-015-1380-1). *Coral Reefs*. **35**, 387–397 (2015).

206. "Early life" in *Encyclopedia of Astrobiology* (2011; <http://dx.doi.org/10.1007/978-3-642-11274-4_2362>), pp. 461–461.

207. K. De Baets, C. Klug, D. Korn, N. H. Landman, Data from: Early evolutionary trends in ammonoid embryonic development (2015), doi:[10.5061/DRYAD.K84C3K8J](https://doi.org/10.5061/DRYAD.K84C3K8J).

208. D. M. Holstein, T. B. Smith, J. Gyory, C. B. Paris, Fertile fathoms: Deep reproductive refugia for threatened shallow corals. *Scientific Reports*. **5** (2015), doi:[10.1038/srep12407](https://doi.org/10.1038/srep12407).

209. M. P. Velazquez, A. Gracia, Fecundity of litopenaeus setiferus, farfantepenaeus aztecus and f. Duorarum, in the southwestern gulf of mexico. *Gulf and Caribbean Research*. **12** (2000), doi:[10.18785/gcr.1201.01](https://doi.org/10.18785/gcr.1201.01).

210. J. Stoddart, R. Black, [Cycles of gametogenesis and plantation in the coral pocillopora damicornis](https://doi.org/10.3354/meps023153). *Marine Ecology Progress Series*. **23**, 153–164 (1985).

211. M.-Y. Leu, C.-H. Liou, W.-H. Wang, S.-D. Yang, P.-J. Meng, [Natural spawning, early development and first feeding of the semicircle angelfish [pomacanthus semicirculatus(cuvier, 1831)] in captivity](https://doi.org/10.1111/j.1365-2109.2009.02192.x). *Aquaculture Research*. **40**, 1019–1030 (2009).

212. and William N S Arlidge, Coral reef viruses in kane’ohe bay, hawai’i: Abundance, diversity &amp; environmental drivers (2021), doi:[10.26686/wgtn.17008594](https://doi.org/10.26686/wgtn.17008594).

213. <https://animalia.bio/pterygioteuthis-microlampas>

214. <https://www.sdnhm.org/oceanoasis/fieldguide/rhiz-man.html>

215. "Viviparous, adj." in *Oxford English Dictionary* (2024; <http://dx.doi.org/10.1093/oed/1117862896>).

216. J. R. Lueg, A. L. Moulding, V. N. Kosmynin, D. S. Gilliam, [Gametogenesis and spawning ofSolenastrea bournoniandStephanocoenia interseptain southeast florida, USA](https://doi.org/10.1155/2012/370247). *Journal of Marine Biology*. **2012**, 1–13 (2012).

217. M. E. Bushnell, J. T. Claisse, C. W. Laidley, [Lunar and seasonal patterns in fecundity of an indeterminate, multiple‐spawning surgeonfish, the yellow tang <i>zebrasoma flavescens</i>](https://doi.org/10.1111/j.1095-8649.2010.02569.x). *Journal of Fish Biology*. **76**, 1343–1361 (2024).

218. D. A. Pavlov, N. G. Emel’yanova, [Fertility of ovulated oocytes after their short-term storage in water in several species of marine tropical fishes](https://doi.org/10.1134/s0032945208080122). *Journal of Ichthyology*. **48**, 655–664 (2008).

219. B. Polidoro, Red list of marine bony fishes of the eastern central atlantic (2024), doi:[10.2305/iucn.ch.2016.04.en](https://doi.org/10.2305/iucn.ch.2016.04.en).

220. Neves, E.G., Histological analysis of reproductive trends of three Porites species from Kane'ohe Bay, Hawai'I (2000).