cirrhosis-prediction

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Cirrhosis Prediction

Cirrhosis is a late stage of scarring (fibrosis) of the liver caused by many forms of liver diseases and conditions, such as hepatitis and chronic alcoholism. The following data contains the information collected from the Mayo Clinic trial in primary biliary cirrhosis (PBC) of the liver conducted between 1974 and 1984. A description of the clinical background for the trial and the covariates recorded here is in Chapter 0, especially Section 0.2 of Fleming and Harrington, Counting

Processes and Survival Analysis, Wiley, 1991. A more extended discussion can be found in Dickson, et al., Hepatology 10:1-7 (1989) and in Markus, et al., N Eng J of Med 320:1709-13 (1989).

A total of 424 PBC patients, referred to Mayo Clinic during that ten-year interval, met eligibility criteria for the randomized placebo-controlled trial of the drug D-penicillamine. The first 312 cases in the dataset participated in the randomized trial and contain largely complete data. The additional 112 cases did not participate in the clinical trial but consented to have basic measurements recorded and to be followed for survival. Six of those cases were lost to follow-up shortly after diagnosis, so the data here are on an additional 106 cases as well as the 312 randomized participants.

Objectives

- 1. To build a model that will classify images in detecting the Liver Cirrhosis using Liver Ultrasound Data.
- 2. To compare or trade off four algorithms such as Logistic Regression (LR), Support Vector Machine (SVM), Multilayer Perceptron (MLP), and Convolutional Neural Networks (CNN) that is suitable in in detecting the Liver Cirrhosis using Liver Ultrasound Data.
- 3. To test the accuracy of four algorithms such as Logistic Regression (LR), Support Vector Machine (SVM), Multilayer Perceptron (MLP), and Convolutional Neural Networks (CNN) that will be fit in to detect Liver Cirrhosis Detection using Liver Ultrasound Data

1 - Packages

```
#install.packages("dplyr")
library(dplyr)
```

```
##
## Attaching package: 'dplyr'
## The following objects are masked from 'package:stats':
##
##
       filter, lag
## The following objects are masked from 'package:base':
##
       intersect, setdiff, setequal, union
##
library(corrplot)
## corrplot 0.92 loaded
library(gplots)
##
## Attaching package: 'gplots'
## The following object is masked from 'package:stats':
##
##
       lowess
library(tidyverse)
## — Attaching core tidyverse packages -
                                                                 – tidyverse 2.0.0 —
## ✓ forcats
             1.0.0
                         ✓ readr
## ✓ ggplot2
               3.4.4
                                     1.5.1

✓ stringr

## ✓ lubridate 1.9.3

✓ tibble

                                      3.2.1
## ✓ purrr
               1.0.2

✓ tidyr

                                      1.3.0
## - Conflicts -
                                                           - tidyverse_conflicts() --
## * dplyr::filter() masks stats::filter()
## * dplyr::lag()
                     masks stats::lag()
## i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflic
ts to become errors
```

```
library(finalfit)
library(ROSE)
```

```
## Loaded ROSE 0.0-4
```

library(caret)

```
## Loading required package: lattice
##
## Attaching package: 'caret'
##
## The following object is masked from 'package:purrr':
##
## lift
```

library(randomForest)

```
## randomForest 4.7-1.1
## Type rfNews() to see new features/changes/bug fixes.
##
## Attaching package: 'randomForest'
##
## The following object is masked from 'package:ggplot2':
##
## margin
##
## The following object is masked from 'package:dplyr':
##
## combine
```

```
library(ggplot2)
library(e1071)
library(RSNNS)
```

```
## Loading required package: Rcpp
##
## Attaching package: 'RSNNS'
##
## The following objects are masked from 'package:caret':
##
## confusionMatrix, train
```

library(tensorflow)

```
##
## Attaching package: 'tensorflow'
##
## The following object is masked from 'package:RSNNS':
##
## train
##
## The following object is masked from 'package:caret':
##
## train
```

```
library(keras)
library(smotefamily)

library(reticulate)
use_python("/usr/local/bin/python3")
```

2 - Data Preparation & EDA

```
cirrhosis_data <- read.csv("input/cirrhosis.csv")
print(head(cirrhosis_data))</pre>
```

##		ID	N_Days	Status		Drug	Αç	je Sex	Asc	cites	Hepatomegaly	Spiders	Edema
##	1	1	400	D	D-per	nicillamine	2146	54 F		Y	Y	Y	Y
##	2	2	4500	С	D-per	nicillamine	2061	17 F		N	Y	Y	N
##	3	3	1012	D	D-per	nicillamine	2559	94 M		N	N	N	S
##	4	4	1925	D	D-per	nicillamine	1999	94 F		N	Y	Y	S
##	5	5	1504	\mathtt{CL}		Placebo	1391	18 F		N	Y	Y	N
##	6	6	2503	D		Placebo	2420)1 F		N	Y	N	N
##		Bil	irubin	Cholest	terol	Albumin Co	pper	Alk_P	hos	SGO	T Trygliceri	des Plat	elets
##	1		14.5		261	2.60	156	171	8.0	137.9	5	172	190
##	2		1.1		302	4.14	54	739	4.8	113.5	2	88	221
##	3		1.4		176	3.48	210	51	6.0	96.1	0	55	151
##	4		1.8		244	2.54	64	612	1.8	60.6	3	92	183
##	5		3.4		279	3.53	143	67	1.0	113.1	5	72	136
##	6		0.8		248	3.98	50	94	4.0	93.0	0	63	NA
##		Pro	thrombi	n Stage	9								
##	1		12.	. 2	4								
##	2		10.	6 3	3								
##	3		12.	. 0	4								
##	4		10.	.3	4								
##			10.		3								
##			11.		3								

```
cirrhosis_data <- cirrhosis_data %>%
  select(-ID)
```

```
str(cirrhosis_data)
```

```
## 'data.frame':
                    418 obs. of 19 variables:
                   : int
                          400 4500 1012 1925 1504 2503 1832 2466 2400 51 ...
##
   $ N Days
                          "D" "C" "D" "D" ...
    $ Status
                   : chr
##
                          "D-penicillamine" "D-penicillamine" "D-pe
##
    $ Drug
                   : chr
nicillamine" ...
##
    $ Age
                   : int
                          21464 20617 25594 19994 13918 24201 20284 19379 15526 25772
##
    $ Sex
                   : chr
                          "F" "F" "M" "F"
                          "Y" "N" "N" "N"
    $ Ascites
                   : chr
##
    $ Hepatomegaly : chr
                          "Y" "Y" "N" "Y" ...
##
    $ Spiders
                          "Y" "Y" "N" "Y"
##
                   : chr
                          "Y" "N" "S" "S" ...
##
    $ Edema
                   : chr
    $ Bilirubin
                         14.5 1.1 1.4 1.8 3.4 0.8 1 0.3 3.2 12.6 ...
##
                   : num
                          261 302 176 244 279 248 322 280 562 200 ...
##
    $ Cholesterol : int
    $ Albumin
                   : num 2.6 4.14 3.48 2.54 3.53 3.98 4.09 4 3.08 2.74 ...
##
                          156 54 210 64 143 50 52 52 79 140 ...
    $ Copper
                   : int
##
    $ Alk Phos
                         1718 7395 516 6122 671 ...
##
                   : num
##
    $ SGOT
                   : num
                         137.9 113.5 96.1 60.6 113.2 ...
                         172 88 55 92 72 63 213 189 88 143 ...
##
    $ Tryglicerides: int
    $ Platelets
                          190 221 151 183 136 NA 204 373 251 302 ...
                   : int
##
    $ Prothrombin : num 12.2 10.6 12 10.3 10.9 11 9.7 11 11 11.5 ...
##
                          4 3 4 4 3 3 3 3 2 4 ...
##
    $ Stage
                   : int
```

```
summary(cirrhosis data)
```

##	N_Days	Status	Drug	Age
#	Min. : 41	Length:418	Length:418	Min. : 9598
##	1st Qu.:1093	Class :character	Class :character	1st Qu.:15644
##	Median :1730	Mode :character	Mode :character	Median :18628
##	Mean :1918			Mean :18533
##	3rd Qu.:2614			3rd Qu.:21272
##	Max. :4795			Max. :28650
##				
##	Sex	Ascites	Hepatomegaly	Spiders
##	Length:418	Length:418	Length: 418	Length:418
##	Class :charact	er Class:charac	ter Class :charac	cter Class :character
##	Mode :charact	er Mode :charac	ter Mode :charac	cter Mode :character
##				
##				
##				
##				
##	Edema	Bilirubin	Cholesterol	Albumin
##	Length:418	Min. : 0.30	0 Min. : 120.0	Min. :1.960
##	Class :charact	er 1st Qu.: 0.80	0 1st Qu.: 249.5	1st Qu.:3.243
##	Mode :charact			
##		Mean : 3.22	1 Mean : 369.5	Mean :3.497
##		3rd Qu.: 3.40	0 3rd Qu.: 400.0	
##		Max. :28.00		
##			NA's :134	
##	Copper	Alk_Phos	SGOT	Tryglicerides
##	Min. : 4.00	-	Min. : 26.35	Min. : 33.00
##	1st Qu.: 41.25			1st Qu.: 84.25
##	Median : 73.00			Median :108.00
" " ##	Mean : 97.65			Mean :124.70
" " ##	3rd Qu.:123.00			3rd Qu.:151.00
##	Max. :588.00			Max. :598.00
##	NA's :108	NA's :106	NA's :106	NA's :136
##	Platelets	Prothrombin	Stage	111 5 • 150
##	Min. : 62.0		Min. :1.000	
## ##	1st Qu.:188.5	1st Qu.:10.00	1st Qu.:2.000	
	Median :251.0		Median :3.000	
## ##	Mean :257.0			
## ##			Mean :3.024	
## ##	3rd Qu.:318.0	3rd Qu.:11.10	3rd Qu.:4.000	
## ##	Max. :721.0		Max. :4.000	
##	NA's :11	NA's :2	NA's :6	

Handling Missing Values

```
# drop the 6 rows with missing 'Stage'
cirrhosis data <- cirrhosis data[complete.cases(cirrhosis data$Stage), ]</pre>
# Numerical columns --> Median
# Impute missing values in numerical columns with the median.
numerical_columns <- sapply(cirrhosis_data, is.numeric)</pre>
for (c in names(cirrhosis_data[, numerical_columns])) {
  cirrhosis data[, c] <- ifelse(is.na(cirrhosis data[, c]), median(cirrhosis data[,</pre>
c], na.rm = TRUE), cirrhosis data[, c])
# Categorical columns --> Most Frequent
# Impute missing values in categorical columns with the most frequent value
categorical columns <- sapply(cirrhosis data, is.factor)</pre>
for (c in names(cirrhosis_data[, categorical_columns])) {
  cirrhosis data[, c] <- ifelse(is.na(cirrhosis data[, c]), levels(cirrhosis data[,</pre>
c])[which.max(table(cirrhosis_data[, c]))], cirrhosis_data[, c])
# Convert 'Stage' to integer
cirrhosis data$Stage <- as.integer(cirrhosis data$Stage)</pre>
cirrhosis data <- na.omit(cirrhosis data)</pre>
```

Missing Values

summary(cirrhosis_data)

##	N_Days	Status	Drug	Age
##	Min. : 41	Length:312	Length:312	Min. : 9598
##	1st Qu.:1191	Class :character	Class :character	1st Qu.:15428
##	Median :1840	Mode :character	Mode :character	Median :18188
##	Mean :2006			Mean :18269
##	3rd Qu.:2697			3rd Qu.:20715
##	Max. :4556			Max. :28650
##	Sex	Ascites	Hepatomegaly	Spiders
##	Length: 312	Length: 312	Length:312	Length: 312
##	Class :characte	er Class :charact	er Class :charac	cter Class :character
##	Mode :characte	er Mode :charact	er Mode :charac	cter Mode :character
##				
##				
##				
##	Edema	Bilirubin	Cholesterol	Albumin
##	Length: 312	Min. : 0.300	Min. : 120.0	Min. :1.96
##	Class :characte	er 1st Qu.: 0.800	1st Qu.: 255.8	1st Qu.:3.31
##	Mode :characte	er Median: 1.350	Median : 309.5	Median :3.55
##		Mean : 3.256	Mean : 364.1	Mean :3.52
##		3rd Qu.: 3.425	3rd Qu.: 392.2	3rd Qu.:3.80
##		Max. :28.000	Max. :1775.0	Max. :4.64
##	Copper	Alk_Phos	SGOT	Tryglicerides
##	Min. : 4.00	Min. : 289.0	Min. : 26.35	Min. : 33.0
##	1st Qu.: 41.75	1st Qu.: 871.5	1st Qu.: 80.60	1st Qu.: 87.0
##	Median : 73.00	Median : 1259.0	Median :114.70	Median :108.0
##	Mean : 97.49	Mean : 1982.7	Mean :122.56	Mean :123.1
##	3rd Qu.:123.00	3rd Qu.: 1980.0	3rd Qu.:151.90	3rd Qu.:145.2
##	Max. :588.00	Max. :13862.4	Max. :457.25	Max. :598.0
##	Platelets	Prothrombin	Stage	
##	Min. : 62.0		Min. :1.000	
##	1st Qu.:200.0	1st Qu.:10.00 1	lst Qu.:2.000	
##	Median :255.5	Median:10.60 M	Median :3.000	
##	Mean :261.8	Mean :10.73 M	Mean :3.032	
##	3rd Qu.:322.0	3rd Qu.:11.10 3	3rd Qu.:4.000	
##	Max. :563.0	Max. :17.10 M	Max. :4.000	

Recode and Mutate Data

```
cirrhosis_data <- cirrhosis_data %>%
  mutate(
    Sex = recode(Sex, 'M' = 0, 'F' = 1),
    Ascites = recode(Ascites, 'N' = 0, 'Y' = 1),
    Drug = recode(Drug, 'D-penicillamine' = 0, 'Placebo' = 1),
    Hepatomegaly = recode(Hepatomegaly, 'N' = 0, 'Y' = 1),
    Spiders = recode(Spiders, 'N' = 0, 'Y' = 1),
    Edema = recode(Edema, 'N' = 0, 'Y' = 1, 'S' = -1),
    Status = recode(Status, 'C' = 0, 'CL' = 1, 'D' = -1),
)
```

```
str(cirrhosis_data)
```

```
## 'data.frame':
                    312 obs. of 19 variables:
##
    $ N Days
                   : int 400 4500 1012 1925 1504 2503 1832 2466 2400 51 ...
##
   $ Status
                   : num
                          -1 0 -1 -1 1 -1 0 -1 -1 -1 ...
                          0 0 0 0 1 1 1 1 0 1 ...
## $ Drug
                   : num
                          21464 20617 25594 19994 13918 24201 20284 19379 15526 25772
##
    $ Age
                   : int
. . .
##
    $ Sex
                          1 1 0 1 1 1 1 1 1 1 ...
                   : num
##
   $ Ascites
                   : num
                         1 0 0 0 0 0 0 0 0 1 ...
    $ Hepatomegaly : num
                         1 1 0 1 1 1 1 0 0 0 ...
##
    $ Spiders
                         1 1 0 1 1 0 0 0 1 1 ...
##
                   : num
   $ Edema
                         1 0 -1 -1 0 0 0 0 0 1 ...
##
                   : num
                         14.5 1.1 1.4 1.8 3.4 0.8 1 0.3 3.2 12.6 ...
    $ Bilirubin
##
                   : num
##
    $ Cholesterol : num 261 302 176 244 279 248 322 280 562 200 ...
                         2.6 4.14 3.48 2.54 3.53 3.98 4.09 4 3.08 2.74 ...
##
   $ Albumin
                   : num
##
    $ Copper
                   : num
                         156 54 210 64 143 50 52 52 79 140 ...
## $ Alk Phos
                   : num 1718 7395 516 6122 671 ...
## $ SGOT
                   : num 137.9 113.5 96.1 60.6 113.2 ...
   $ Tryglicerides: num 172 88 55 92 72 63 213 189 88 143 ...
##
                   : int 190 221 151 183 136 249 204 373 251 302 ...
##
   $ Platelets
##
    $ Prothrombin : num 12.2 10.6 12 10.3 10.9 11 9.7 11 11 11.5 ...
##
    $ Stage
                   : int
                          4 3 4 4 3 3 3 3 2 4 ...
##
   - attr(*, "na.action")= 'omit' Named int [1:100] 313 314 315 316 317 318 319 320
     ... attr(*, "names")= chr [1:100] "314" "315" "316" "318" ...
##
```

3 - Preprocessing Data

```
#Upsampling
balancedData <- SMOTE(
   X = cirrhosis_data,
   target = cirrhosis_data$Stage,
   K = 3)

#Standardize
scaledData <- balancedData$data</pre>
```

```
X <- subset(scaledData, select = -c(Status, N_Days, Stage, class))
y <- scaledData$Stage

X_matrix <- as.matrix(sapply(X, as.numeric))</pre>
```

```
#Splitting the data 80/20
set.seed(23)
split <- createDataPartition(scaledData$Stage, p = 0.8, list = FALSE)
X_train <- X_matrix[split, ]
X_test <- X_matrix[-split, ]
y_train <- y[split]
y_test <- y[-split]</pre>
```

4 - Modelling

Random Forest

```
rf_model <- randomForest(x = X_train, y = factor(y_train), ntree = 500)

# Make predictions on the test set
predictions <- predict(rf_model, newdata = X_test)

# Evaluate the model
conf_matrix <- table(predictions, y_test)
print(conf_matrix)</pre>
```

```
## y_test
## predictions 1 2 3 4
## 1 58 4 3 0
## 2 0 1 0 2
## 3 0 7 15 6
## 4 0 1 6 13
```

```
accuracy <- sum(diag(conf_matrix)) / sum(conf_matrix)
print(paste("Accuracy:", round(accuracy, 4)))</pre>
```

```
## [1] "Accuracy: 0.75"
```

```
# Feature importance
importance(rf_model)
```

```
##
                 MeanDecreaseGini
## Drug
                        14.185748
## Age
                        22.565158
## Sex
                         6.064978
## Ascites
                        1.796973
## Hepatomegaly
                        36.645801
## Spiders
                       10.495472
## Edema
                        2.446834
## Bilirubin
                        35.717158
## Cholesterol
                        20.481980
## Albumin
                        28.290631
## Copper
                        18.491163
## Alk Phos
                        21.426696
## SGOT
                        24.696566
## Tryglicerides
                        25.902124
## Platelets
                        21.301023
## Prothrombin
                        21.171215
```

SVM

```
## y_test
## predictions 1 2 3 4
## 1 58 7 4 1
## 2 0 0 0 1
## 3 0 3 15 6
## 4 0 3 5 13
```

```
accuracy <- (sum(diag(conf_matrix)) / sum(conf_matrix))
print(paste("Accuracy:", round(accuracy, 4)))</pre>
```

```
## [1] "Accuracy: 0.7414"
```

```
# dataframe to matrix

matrixX_train <- as.matrix(X_train)
matrixy_train <- as.matrix(y_train)
matrixX_test <- as.matrix(X_test)
matrixy_test <- as.matrix(y_test)</pre>
```

MLP

```
model_mlp <- keras_model_sequential()
input_shape <- dim(X_train)[2]

model_mlp %>%
  layer_dense(units = 32, activation = 'relu', input_shape = input_shape) %>%
  layer_dense(units = 64, activation = 'relu') %>%
  layer_dense(units = 32, activation = 'relu') %>%
  layer_dense(units = 5, activation = 'relu') %>%
  compile(
    optimizer = 'adam',
    loss = 'sparse_categorical_crossentropy',
    metrics = c('accuracy'))

summary(model_mlp)
```

```
## Model: "sequential"
##
## Layer (type)
                            Output Shape
                                                    Param #
dense 3 (Dense)
                                                    544
##
                            (None, 32)
##
  dense 2 (Dense)
                            (None, 64)
                                                    2112
##
  dense 1 (Dense)
                            (None, 32)
                                                    2080
## dense (Dense)
                                                    165
                            (None, 5)
## Total params: 4901 (19.14 KB)
## Trainable params: 4901 (19.14 KB)
## Non-trainable params: 0 (0.00 Byte)
##
```

model_mlp %>% fit(matrixX_train, matrixy_train, epochs = 20, validation_data = list(m atrixX_test, matrixy_test))

```
## Epoch 1/20
## 15/15 - 0s - loss: 378.8661 - accuracy: 0.3141 - val_loss: 223.1509 - val_accurac
y: 0.5086 - 483ms/epoch - 32ms/step
## Epoch 2/20
## 15/15 - 0s - loss: 95.0060 - accuracy: 0.3205 - val loss: 70.0251 - val accuracy:
0.3103 - 70 \text{ms/epoch} - 5 \text{ms/step}
## Epoch 3/20
## 15/15 - 0s - loss: 51.6143 - accuracy: 0.3803 - val loss: 52.1501 - val accuracy:
0.4052 - 49ms/epoch - 3ms/step
## Epoch 4/20
## 15/15 - 0s - loss: 52.1113 - accuracy: 0.3312 - val loss: 73.0927 - val accuracy:
0.1724 - 50 \text{ms/epoch} - 3 \text{ms/step}
## Epoch 5/20
## 15/15 - 0s - loss: 45.1129 - accuracy: 0.3803 - val loss: 57.0884 - val accuracy:
0.2931 - 50 \text{ms/epoch} - 3 \text{ms/step}
## Epoch 6/20
## 15/15 - 0s - loss: 23.6788 - accuracy: 0.3868 - val_loss: 35.1046 - val_accuracy:
0.5000 - 48ms/epoch - 3ms/step
## Epoch 7/20
## 15/15 - 0s - loss: 33.0452 - accuracy: 0.3226 - val_loss: 76.1884 - val_accuracy:
0.5000 - 53 \text{ms/epoch} - 4 \text{ms/step}
## Epoch 8/20
## 15/15 - 0s - loss: 43.3555 - accuracy: 0.3462 - val loss: 33.5154 - val accuracy:
0.5431 - 49ms/epoch - 3ms/step
## Epoch 9/20
## 15/15 - 0s - loss: 36.5277 - accuracy: 0.3675 - val loss: 41.8665 - val accuracy:
0.1379 - 46ms/epoch - 3ms/step
## Epoch 10/20
```

```
## 15/15 - 0s - loss: 19.0711 - accuracy: 0.4017 - val loss: 35.9004 - val accuracy:
0.3276 - 48ms/epoch - 3ms/step
## Epoch 11/20
## 15/15 - 0s - loss: 24.9486 - accuracy: 0.3462 - val_loss: 43.3852 - val_accuracy:
0.5000 - 48 \text{ms/epoch} - 3 \text{ms/step}
## Epoch 12/20
## 15/15 - 0s - loss: 28.1007 - accuracy: 0.3739 - val_loss: 29.1176 - val_accuracy:
0.2241 - 48ms/epoch - 3ms/step
## Epoch 13/20
## 15/15 - 0s - loss: 23.3510 - accuracy: 0.4038 - val loss: 39.1374 - val accuracy:
0.2328 - 45 \text{ms/epoch} - 3 \text{ms/step}
## Epoch 14/20
## 15/15 - 0s - loss: 35.9711 - accuracy: 0.3419 - val_loss: 48.1245 - val_accuracy:
0.2586 - 48ms/epoch - 3ms/step
## Epoch 15/20
## 15/15 - 0s - loss: 36.2496 - accuracy: 0.4017 - val loss: 29.7412 - val accuracy:
0.2414 - 46ms/epoch - 3ms/step
## Epoch 16/20
## 15/15 - 0s - loss: 27.6098 - accuracy: 0.3504 - val loss: 53.3075 - val accuracy:
0.4828 - 46ms/epoch - 3ms/step
## Epoch 17/20
## 15/15 - 0s - loss: 32.7499 - accuracy: 0.3419 - val_loss: 39.1943 - val_accuracy:
0.2500 - 43 \text{ms/epoch} - 3 \text{ms/step}
## Epoch 18/20
## 15/15 - 0s - loss: 36.5610 - accuracy: 0.3440 - val loss: 82.3010 - val accuracy:
0.2500 - 46 \text{ms/epoch} - 3 \text{ms/step}
## Epoch 19/20
## 15/15 - 0s - loss: 48.4172 - accuracy: 0.3632 - val_loss: 44.0441 - val_accuracy:
0.2500 - 46 \text{ms/epoch} - 3 \text{ms/step}
## Epoch 20/20
## 15/15 - 0s - loss: 32.2664 - accuracy: 0.3547 - val_loss: 20.2731 - val_accuracy:
0.2414 - 44ms/epoch - 3ms/step
```

```
score <- model_mlp %>% evaluate(matrixX_test, matrixy_test)
```

```
## 4/4 - 0s - loss: 20.2731 - accuracy: 0.2414 - 24ms/epoch - 6ms/step
```

```
score
```

```
## loss accuracy
## 20.2731247 0.2413793
```

CNN

```
CNNmodel <- keras_model_sequential()</pre>
CNNmodel %>%
  layer conv 1d(filters = 32, kernel size = 3, padding = 'same', activation = 'relu',
input shape = c(16, 1)) %>%
  layer max pooling 1d(pool size = 2) %>%
  layer conv 1d(filters = 64, kernel size = 3, padding = 'same', activation = 'relu')
%>%
  layer max pooling 1d(pool size = 2) %>%
  layer conv 1d(filters = 32, kernel size = 3, padding = 'same', activation = 'relu')
%>왕
  layer_max_pooling_1d(pool_size = 2) %>%
  layer dropout(0.25) %>%
  layer flatten() %>%
  layer dense(units = 16, activation = 'relu') %>%
  layer_dense(units = 5, activation = 'softmax') %>%
 compile(
    loss = 'sparse categorical crossentropy',
    optimizer = 'adam',
    metrics = 'accuracy')
summary(CNNmodel)
```

```
## Model: "sequential_1"
##
##
  Layer (type)
                               Output Shape
                                                         Param #
## ------
##
  convld 2 (ConvlD)
                                (None, 16, 32)
                                                         128
## max pooling1d 2 (MaxPooling1D)
                               (None, 8, 32)
##
   convld 1 (ConvlD)
                               (None, 8, 64)
                                                         6208
## max pooling1d 1 (MaxPooling1D)
                               (None, 4, 64)
   convld (ConvlD)
                               (None, 4, 32)
                                                         6176
##
## max_pooling1d (MaxPooling1D)
                               (None, 2, 32)
## dropout (Dropout)
                               (None, 2, 32)
   flatten (Flatten)
##
                                (None, 64)
   dense 5 (Dense)
                               (None, 16)
                                                         1040
##
##
   dense 4 (Dense)
                               (None, 5)
                                                         85
## Total params: 13637 (53.27 KB)
## Trainable params: 13637 (53.27 KB)
## Non-trainable params: 0 (0.00 Byte)
##
```

```
CNNmodel %>%
  fit(
    x = matrixX_train,
    y = matrixy_train,
    epochs = 20,
    validation_data = list(matrixX_test, matrixy_test))
```

```
## Epoch 1/20
## 15/15 - 1s - loss: 259.8183 - accuracy: 0.3141 - val_loss: 67.8598 - val_accuracy:
0.5000 - 629 \text{ms/epoch} - 42 \text{ms/step}
## Epoch 2/20
## 15/15 - 0s - loss: 90.4796 - accuracy: 0.3504 - val loss: 41.9381 - val accuracy:
0.5086 - 57ms/epoch - 4ms/step
## Epoch 3/20
## 15/15 - 0s - loss: 32.0528 - accuracy: 0.2842 - val_loss: 3.6813 - val_accuracy:
0.2069 - 59ms/epoch - 4ms/step
## Epoch 4/20
## 15/15 - 0s - loss: 12.0261 - accuracy: 0.2286 - val_loss: 2.2647 - val_accuracy:
0.5086 - 65ms/epoch - 4ms/step
## Epoch 5/20
## 15/15 - 0s - loss: 6.8257 - accuracy: 0.4359 - val_loss: 1.5914 - val_accuracy: 0.
5000 - 60ms/epoch - 4ms/step
## Epoch 6/20
## 15/15 - 0s - loss: 5.0420 - accuracy: 0.4359 - val loss: 1.5841 - val accuracy: 0.
5000 - 65ms/epoch - 4ms/step
## Epoch 7/20
## 15/15 - 0s - loss: 4.7129 - accuracy: 0.4316 - val_loss: 1.5770 - val_accuracy: 0.
5000 - 64ms/epoch - 4ms/step
## Epoch 8/20
## 15/15 - 0s - loss: 4.5016 - accuracy: 0.4103 - val loss: 1.5685 - val accuracy: 0.
5000 - 64ms/epoch - 4ms/step
## Epoch 9/20
## 15/15 - 0s - loss: 3.2632 - accuracy: 0.4167 - val loss: 1.5613 - val accuracy: 0.
5000 - 62ms/epoch - 4ms/step
## Epoch 10/20
## 15/15 - 0s - loss: 3.3154 - accuracy: 0.4231 - val loss: 1.5533 - val accuracy: 0.
5000 - 61ms/epoch - 4ms/step
## Epoch 11/20
## 15/15 - 0s - loss: 2.7117 - accuracy: 0.3996 - val loss: 1.5457 - val accuracy: 0.
5000 - 63ms/epoch - 4ms/step
## Epoch 12/20
## 15/15 - 0s - loss: 2.7840 - accuracy: 0.4637 - val_loss: 1.5472 - val_accuracy: 0.
5000 - 62ms/epoch - 4ms/step
## Epoch 13/20
## 15/15 - 0s - loss: 2.3919 - accuracy: 0.4658 - val_loss: 1.5318 - val_accuracy: 0.
5000 - 63ms/epoch - 4ms/step
```

```
## Epoch 14/20
## 15/15 - 0s - loss: 2.2709 - accuracy: 0.4615 - val_loss: 1.5246 - val_accuracy: 0.
5000 - 62ms/epoch - 4ms/step
## Epoch 15/20
## 15/15 - 0s - loss: 2.4526 - accuracy: 0.4722 - val loss: 1.5172 - val accuracy: 0.
5000 - 61ms/epoch - 4ms/step
## Epoch 16/20
## 15/15 - 0s - loss: 2.1170 - accuracy: 0.4509 - val loss: 1.5100 - val accuracy: 0.
5000 - 61ms/epoch - 4ms/step
## Epoch 17/20
## 15/15 - 0s - loss: 2.2062 - accuracy: 0.4466 - val loss: 1.5031 - val accuracy: 0.
5000 - 60ms/epoch - 4ms/step
## Epoch 18/20
## 15/15 - 0s - loss: 2.4031 - accuracy: 0.4615 - val loss: 1.4962 - val accuracy: 0.
5000 - 62ms/epoch - 4ms/step
## Epoch 19/20
## 15/15 - 0s - loss: 2.4706 - accuracy: 0.4637 - val_loss: 1.4901 - val_accuracy: 0.
5000 - 61ms/epoch - 4ms/step
## Epoch 20/20
## 15/15 - 0s - loss: 2.0937 - accuracy: 0.4658 - val_loss: 1.4841 - val_accuracy: 0.
5000 - 85ms/epoch - 6ms/step
```

CNNmodel %>% evaluate(matrixX_test, matrixy_test)

```
## 4/4 - 0s - loss: 1.4841 - accuracy: 0.5000 - 26ms/epoch - 6ms/step
```

```
## loss accuracy
## 1.48409 0.50000
```

predictionsTest <- CNNmodel %>% evaluate(matrixX_test, matrixy_test)

```
## 4/4 - 0s - loss: 1.4841 - accuracy: 0.5000 - 22ms/epoch - 5ms/step
```

predictionsTest

```
## loss accuracy
## 1.48409 0.50000
```