

	Environmental Analysis Teaching and Research Laboratory	Date: 7/12/2015	Number: 32 v0.1
	Standard Operating Procedure	Title: Texture Analysis using a Hydrometer	
	Approved By: Los Huertos	Revision Date: August 15, 2016	

## 1. Scope and Application

1.1 The scope of this SOP is train researchers...

1.2 The applications of this SOP are for...

## 2. Summary of Method

2.1 This SOP does this...

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### **3. Acknowledgements**

### **4. Definitions**

4.1 Term1: is...

### **5. Interferences**

5.1 Biases and interferences can come from...

### **6. Health and Safety**

6.1 Sodium hexametaphosphate is a non-hazardous material but may be irritating to skin and eyes. Dust may be irritating to lungs if inhaled.

#### **Safety and Personnel Protective Equipment**

6.2 Care should be taken to avoid inhalation. Handle powdered HMP in a well-ventilated area or fume hood. Chemical-safety goggles should be worn while pouring HMP solution and during blending to avoid contact with eyes. In case of contact, flush eyes with cold water for 15 minutes. Latex gloves should be worn to avoid contact with skin. Hands and contaminated skin should be washed thoroughly with soap and water after handling. Sweep, vacuum, scoop or remove spilled sodium hexametaphosphate. Flush residual area with water. In case of spills on clothes, wash thoroughly before wearing again. See the SDS for more information on contamination and spills.

### **7. Personnel & Training Responsibilities**

7.1 Researchers training is required before this the procedures in this method can be used...

7.2 Researchers using this SOP should be trained for the following SOPs:

- SOP01 Laboratory Safety
- SOP02 Field Safety
- SOP03 Handling Hardous Materials

Task	Time	Elapsed Time	Time until completion
Collect Soil	1-3 hours	1-3 hours	

## 8. Required Materials and Apparati

1. Sieve No. 10 and No. 200.
2. 250-ml and 400-ml beakers
3. Hot plate
4. Soil tins
5. Drying oven
6. Milkshake Mixer and Dispersion Cup (metal milkshake cup)
7. 151H Hydrometer
8. 1 L Sedimentation Cylinder and No. 13 stopper
9. Stopwatch
10. Balance sensitive to 0.01 g for weighing the material passing sieves.

## 9. Reagents and Standards

### 10. Reagents

1. Hydrogen peroxide, 30%.
2. Hydrochloric acid, 1N.
3. Sodium hexametaphosphate

### 11. Estimated Time

**11.1** This procedure requires several days to complete.

## 12. Sample Collection, Preservation, and Storage

### 13. Procedure

**13.1** Before beginning, we need to organize the procedures by day, since we can't complete the analysis during a single lab period.

**Session 1:** Air dry 200 grams of soil (could require more than one day);

**Session 2:** Pass air-dry soils through No. 10 sieve; dry sub-sample for moisture content;

**Session 3:** Calculate and weigh  $SW_e$  for pretreatment and/or dispersion;

**Session 4:** Conduct hydrometer measurements; and

**Session 5:** Analyze particles retained on No. 200 sieve and complete data entry.

### 13.1. Air-Dried Soil

**13.2** After collecting a soil, we allow them to air dry by putting them in a tin or beaker and let them equalibrate with the laboratory for 24 or more hours. If the soils are very wet, this may take several days.

### 13.2. Determining Proportion of Soil Passing No. 10 Sieve

**13.3** Soils are defined as particles that are less than 2 mm. Thus, we begin by removing the gravel, particles that exceed the sand-sized particles,  $>2.0$  mm. These particles, however, influence the soil texture, so we will quantify the contribution of particles that exceed 2 mm.

**13.4** We can do this by analyzing the soil that can pass through the No. 10 sieve, which has a mesh size of 2 mm.

1. Homogenize the soil by removing it from the bag and mix thoroughly;
2. Weigh out approximately 65 g of silt or clay soils or 120 g of a sandy soil, record the value in Box 10.
3. Prepare soil with air-drying and pulverizing to pass a No. 10 mesh sieve ( $< 2$  mm)–pick out the rocks and use a mortar and pestle to break up the soil so that it all passes through the sieve.
4. Weigh the soil that passed through the sieve and record the weight in Box 11.

### 13.3. Measuring and Correcting for Soil Water Content

**13.5** We will use air-dried soil for our test. However, even air-dried soil contains water. This moisture is associated with clay particles, salts, and organic matter and is often referred to as hygroscopic water. To calculate the soil moisture content, we weigh the soil before and after oven drying the soil at  $110^{\circ}$  C. Once we determine the moisture content, we can use the moisture content to correct for the water in the air-dried soil. Because this method relies on the change in soil mass, the water content is often referred to as gravimeter water content as opposed to a volumetric water content.

**13.6** This temperature is somewhat arbitrary, and clay minerals in particular may contain 10-15% water (dry basis) at  $400^{\circ}$  C (Gardner 1986). As temperature increases, first water in soil pores evaporates, then water adsorbed to mineral surfaces, followed by water between lattice layers and that which forms part of the mineral lattice itself. The exact quantities and patterns of release in a heterogeneous mixture like soil depends on the particular mix of minerals making up a sample. Water adsorbed to organic components (as well as other volatile organic substances) will also evaporate

over a range of temperatures. The key point is to specify the temperature used when reporting moisture data.

**13.7** To determine the hygroscopic water content,

1. Weigh and record the weight of a soil tin (Appendix A, Box 13) and the tin # (Appendix A, Box 12);
2. Remove a 10-15 gram sub-sample from the air-dried soil that passed through the No. 10 sieve, put the sub-sample into the pre-weighed soil tin and record the weight (Appendix, Box 14);
3. Dry the sample in a drying oven set at 110 ° C for approximately 24 hours;
4. Allow the tins to cool and record the oven-dried weight (Appendix, Box 15).

**13.8** We calculate the hygroscopic moisture content we subtract the mass of oven-dried sample and the air-dried mass before drying, usually less than one, unless there is no hygroscopic moisture, then is is one. Use the following equation to calculate the water content

$$\text{Moisture}_H = (\text{air-dry mass w/tin} - \text{oven-dried mass w/tin}) / (\text{air-dried mass} - \text{tin mass}) \quad (1)$$

*Moisture<sub>H</sub>* can be calculated using R code developed for this SOP. See Appendix B for a user guide for the program.

### 13.4.Calculating Soil Sample Mass

**13.9** We then calculate the “Total Sample Represented” in the hydrometer test as the mass of soil in hydrometer test as the oven dry mass used by the percent passed through 2 mm sieve and multiply by 100.

**13.10** We will analyze air dried soil – that has passed through a No. 10 sieve.

$$W = \text{ovendrymass} * \text{percentpassing2mmsieve} * 100 \quad (2)$$

$$WB = \frac{M_{air} * 10000}{Per_{No.10} * (100 + Moisture_H)} \quad (3)$$

**13.11** But since the sample has hygroscopic water, we will use the correction factor above to weigh a “oven-dried equivalent”,  $DW_e$ .

**13.12** If the soils are coarse textured (>70% sand), will will use  $DW_e$  of 100 g, otherwise 50 g is sufficient.

**13.13** Since air dried soil contains water, the soil sample used will weigh slightly more than 50g or 100 g for sandy soils where  $DW_e = DW_{air} * (1 + DW_c)$ ,<sup>1</sup>, thus we can re-arrange the equation to determine the amount of air-dried soil to use for our test:

$$DW_{air} = 50 / (1 + DW_c) \quad (4)$$

and for sandy soils,

$$DW_{air} = 100 / (1 + DW_c) \quad (5)$$

## Pre-Treatment

**13.14** Soils should be pre-treated for soil high in organic matter ...

1. Transfer 50 or 100 grams of dry weight equivalent ( $DW_e$ ) grams of soil to a 400-ml glass beaker.
2. Removal of carbonates: Add 50 mL DI water and sufficient 1 M HCl to reduce the soil pH to between 3.0 and 4.0. Stir and allow to equilibrate 10 minutes until there is no effervescence.
3. Removal of organic matter:<sup>2</sup> Carefully add 10 ml of 30% hydrogen peroxide ( $H_2O_2$ ), a few milliliters at a time, allowing effervescence to subside before adding more, until no more frothing occurs. If necessary, make the suspension acidic to litmus paper with a few drops of 1N HCl. Oxidation with  $H_2O_2$  requires an acidic medium. When frothing subsides heat to 90 °C and add  $H_2O_2$  until frothing subsides. Rinse down walls of beaker and continue heating until no excess  $H_2O_2$  is consumed.
4. Removal of iron oxides Add 150 mL of citrate-bicarbonate buffer to sample in beaker. Add 3 g of sodium hydrosulfite ( $Na_2S_2O_3$ ) gradually as samples may froth. Place in water bath at 80°C for 20 minutes. Place the blender cup on electrical mixer and stir for 10 minutes.
5. Removal of soluble salts: Place soil in centrifuge tube and add 100 mL water and centrifuge for 10 min at 1500 rpm until supernatant is clear. If the centrifuge is not clear, repeat the process until it is clear.

## Dispersing Soil Sample

1. Using the datasheet and the R function (`SoilEquiv()`) or web app (<http://>) to calculate  $DW_e$ ;
2. Weigh approximately  $DW_e$ g of soil (From Box 18) into the dispersion cup and record to the nearest 0.1gram (Box 6).

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<sup>1</sup>I need to check this!

<sup>2</sup>may need the previous treatment to wet and acidify the soil before this is done. Not clear.

3. Record the dispersion cup number (Box 6).
4. Add 125 mL of sodium hexametaphosphate (40 g/L). Stir so the slurry is thoroughly wetted. Cover and allow to soak for at least 16 hours.
5. Add distilled water up to the first “indentation” of the dispersion cup.
6. Attach the dispersion cup to the mixer; mix 5 minutes for sandy soils, 15 minutes for fine-textured soils.

### 13.5. Conducting Hydrometer Tests

1. Make up a blank cylinder with 125 mL of sodium hexa-metaphosphate and add distilled water to the 1000 mL mark. Record the blank hydrometer reading and temperature. If the reading is above 0 (zero) on the hydrometer scale (in other words, if the zero mark is below the surface), record the blank correction as a negative number. Read at the top of the meniscus.
2. Transfer soil suspension to sedimentation cylinder; use distilled water from squirt bottle to get all of sample from mixing cup.
3. Fill cylinder to 1000-mL mark with distilled water.
4. Cover the cylinder with a stopper (No. 13) and invert the cylinder 180 degrees for a full minute, where the cylinder should be turned upside-down and back should take 2 seconds. You may need to vigorously mix the cylinder for the first two turns to loosen the soil at the bottom.
5. After the minute of inverting, begin timing. While slowing spinning the hydrometer remove air bubbles, carefully lower the hydrometer into suspension after 20s; record the hydrometer reading at 40 seconds. Remove the hydrometer and rinse the hydrometer between each reading.
6. This 40-second reading should be **repeated to obtain 3 accurate** after repeating the inversion process. Because the suspension is opaque, read the hydrometer at the top of the meniscus rather than at the bottom. There is a correction for this!
7. After final 40-second reading, remove hydrometer, carefully lower a thermometer into the suspension and record the temperature (degrees C). Mixing raises temperature by 3-5°C, so it is important to record the temperature for both hydrometer readings (See Table 1).
8. The hydrometer must be removed and rinsed and dried after each reading. If this is not followed, the hydrometer will accumulate particles on the glass walls and become increasingly inaccurate.
9. Take a hydrometer reading at the following times:
10. Be sure that the cylinder is back from the edge of the counter and in a location where it won't be disturbed.



Obs.	Soil Reading Times	Temperture	Blank Hydrometer
0	Before Inverting (0 seconds)	X	X
1	40 seconds (Repeat twice)		
2	2.0 minutes		
3	5.0 minutes	X	X
4	15 minutes		
5	30 minutes		
6	60 minutes	X	X
7	2 hours	X	X
8	3 hours (optional)	X	X
9	6 or 8 hours (optional)	X	X
10	24 hours	X	X

Table 1: Reading times modified from **citetstandard2007d422**. These times can be adjusted, be sure to record the actual times in Boxes 19 and 20.

### Determining Proportion of Soil Retained No. 200 Sieve

**13.15** Particles retained on the No. 200 sieve are larger are often categorized as sand.

As the hydrometer does very little to distinsin between size fractionation of sand – very fine, fine, medium, course, and very course sand.

**13.16** But collecting the particles on the No. 200 sieve, we can use seives to better understand the soil characteristics. We can do this by analyzing the mass of the soils retained on the No. 200 sieve, which has a mesh size of XX 0.125 mm.

1. Poor the contents of the cylinder through a No. 200 sieve;
2. Using the sink with a sediment trap, wash the soil in the sieve until the water leaving the sieve is clear.
3. Select a soil tin, and record the tin # (Box XX) and tin mass (Box XX).
4. Carefully wash the soil from the tin into the soil, without damaging the sieve screen. The screens are quite delicate, so using a squeeze bottle is probably best. Using a brush or metal device will force particles into the screen, which can get stuck, distort and stretch the mesh openings or otherwise damage them.
5. Weigh the dry soil with the soil tin and record the results in Box Xx.

### 14. Recording the Results

Record the results in Appendix A, Then enter the data into the data entry form (<http://test>). Other columns calculations will be produced with the R code from Appendix B.

## 15. Particle Size Distribution Calculations

Once the data has been entered in the online form, you can extract the data as a csv file. This csv file can then be called by R to calculate the data automatically, using the following functions.

### 15.1.Importing Data

Import and check data integrity

```
> #hydrometer = import()
> #head(hydrometer)
> #str(hydrometer)
```

What should you be looking for? We want to make sure the data shown are the same as expected. There are several ways to look at the data to determine this. I have shown two ways. 1) Looking at the header (first six observations), then we evaluate the structure of the dataframe.

### 15.2.Calculating effective diameter, $D_e$

### 15.3.Calculating the Percent Finer, PF

### 15.4.Graphing and Interpreting the Results

## 16. Conclusion

### Other Citations of Interest:

Wilde, S.A., R.B. Corey, J.G. Iyer, and G.K. Voigt. 1979. Soil and Plant Analysis for Tree Culture. Oxford and IBH Publishing Co., New Delhi. Pp. 12-13.

Anderson, J.M., and J.S. Ingram, eds. 1993. Tropical Soil Biology and Fertility: A Handbook of Methods. 2nd ed. CAB International, Wallingford, UK. P. 93-94.

## 17. Appendix A. Data Sheet

Grain Size Analysis–Hydrometer Datasheet

1. Project		2. Researcher:		3. Date	
4. Sample ID		5. Hydrometer Number/Type			
Sample Preparation					
6. Pre-Treatment		<input type="checkbox"/> HCl		<input type="checkbox"/> H <sub>2</sub> O <sub>2</sub>	
				<input type="checkbox"/> Fe–O	
7. Beaker ID		8. Dispersing Agent		9. Specific Gravity of Particles $GS_p =$	
				10. Desired Sample <input type="checkbox"/> 50 g <input type="checkbox"/> 100 g	
11. Total Soil for Split (g)			12. Soil Passing Sieve No. 10 (g)		
13. Tin #:		14. Tin Tare Weight		15. Tin w/Air-Dried Soil	
				16. Tin w/Oven-Dried Soil	
Calculated Values					
17. % Passing No. 10		18. % Hygroscopic Correction Factor		19. Effective Soil Weight $WS_e$	
				20. Actual Air-Dried Soil Weight $WS_a$	

Hydrometer Readings and Calculations \*K is a constant that is calculated by the temperature and density of the suspension.

21. Time	22. Elapsed Time (t) sec/min	23. Actual Hydrometer Reading ( $R_a$ )	24. Blank Hydrometer Reading ( $R_b$ )	25. Temp. °C	26. K constant*	27. Effective Depth (L)	28. Particle Diameter ( $D_e$ )	29. PF Partial	30. PF Total

No. 200 Sieve Processing

31. Tin #	32. Tin Mass (g)	33. Dry Soil w/Tin (g)
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Quality Control/Quality Assurance

34. Researcher (Signature)	35. Data Entry By (Signature)	36. Quality Check By (Signature)
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**18. Appendix B: R code to calculate parameters for the SOP**

**19. Data Analysis and Calculations**

**20. QC/QA Criteria**

**21. References**

- 21.1** APHA, AWWA, WEF. (2012) Standard Methods for examination of water and wastewater. 22nd American Public Health Association (Eds.). Washington. 1360 pp. (2014).