**Abstract**

**Introduction**

Diving ectotherms are crucial to the functioning of aquatic ecosystems and represent a taxonomically diverse group (Butler and Jones, 1982; Costa, 2007). The evolution of air-breathing organs made terrestrial environs habitable, but many species returned to aquatic habitats, leading to the reinvasion of air-breathing species (Butler and Jones, 1982; Seymour, 1982). This transition occurred in all groups of tetrapod vertebrates across multiple points in evolutionary history, leading to a diverse array of diving ectotherms (turtles, snakes, crocodiles, iguanas, frogs and salamanders) living in freshwater and marine habitats (Butler and Jones, 1982). The ecology of these species is closely tied to the underwater environment and submergences are necessary to fulfill a range of critical tasks. For example, the viperine water snake (*Natrix maura*) dives to hunt fish by actively chasing them or by using a sit-and-wait strategy (Aubret et al., 2015). Juvenile estuarine crocodiles (*Crocodylus porosus*) dive to avoid both aerial and subsurface predators, whilst in adult crocodiles, diving is more important for social interactions, facilitating sit-and-wait hunting and avoiding rapid surface currents (Campbell et al., 2010c; Grigg and Kirshner, 2015). Time available for these underwater tasks is, however, constrained by the need to surface to replenish oxygen stores (Butler and Jones, 1982).

The maximum amount of time an air-breathing animal can remain underwater prior to the switch to anaerobic metabolism and concomitant lactate accumulation is termed the aerobic dive limit (ADL; Butler, 2006). A diver’s ADL can be calculated by dividing its total body oxygen stores (stored in the lungs, blood and muscle tissue) by the rate at which oxygen is consumed (i.e. metabolic rate). The ADL of ectotherms is inversely related to water temperature, due to the thermal sensitivity of metabolism (Rodgers and Franklin, 2017). As the body temperature of a diving ectotherm equilibrates with water temperature, the rate at which body oxygen stores are consumed becomes directly related to water temperature, with oxygen stores consumed more rapidly as temperatures rise. For example, ectotherm metabolic rates typically double (or even triple) with every 10°C increase in body temperature, meaning that an ectotherm diving at 30°C is predicted to dive for only half as long compared to a conspecific diving at 20°C. For example, the ADL of a freshwater turtle (*Mauremys caspica leprosa*) fell from 221 min at 15°C to 58 min at 25°C (Fuster et al., 1997). Anthropogenically forced warming in marine and freshwater habitats therefore poses a threat to ectothermic divers, with the duration of time performing critical underwater tasks potentially being greatly reduced.

The threat of climate warming to diving ectotherms has been largely overlooked, despite many studies showing substantial decreases in dive durations as temperatures rise. Dive durations in the Arafura file snake (*Acrochordus arafurae)*, for example, decreased by 63% in response to an acute temperature increase from 20°C to 32°C (Pratt and Franklin, 2010). Similarly, during a one-hour continual predator threat juvenile *C. porosus* were able to remain submerged for 36.8 min at 28°C and only 21.3 min at 35°C (Rodgers et al., 2015). Dive durations are also influenced by seasonal temperature increases. Field studies have shown marked declines in dive durations in summer months compared to winter months in turtles (Bentivegna et al., 2003; Bradshaw et al., 2007; Gordos et al., 2003) and crocodilians (Campbell et al., 2010a). Warmer waters, brought about by climate change, could have deleterious consequences for diving ectotherms. Elevated temperatures may cut short time available for critical underwater activities and force animals to spend more time at the water’s surface where predation risk is highest (Heithaus and Dill, 2002; Heithaus and Dill, 2003; Heithaus et al., 2002).

Gas-exchange systems may play an important role in mediating the effects of elevated temperatures on diving behaviour. Many divers rely solely on pulmonary gas exchange and have a negligible capacity for aquatic respiration (Wright, 1986). In contrast, bimodal breathers can supplement aerial respiration with aquatic respiration. Aquatic gas exchange typically occurs via diffusion across the skin (i.e. cutaneous respiration; Feder and Burggren, 1985), but some divers also have specialised organs (e.g. buccopharyngeal cavity and cloacal bursae) to facilitate the exchange of respiratory gases with the aquatic environment (Maina, 2002). For example, several freshwater turtles (e.g. *Elseya albagula, Elusor macrurus* and *Rheodytes leukops*) have cloacal bursae which are highly vascularised, gill-like evaginations of the cloaca, that are ventilated with water by the contraction of cloacal muscles (FitzGibbon and Franklin, 2010; Gordos and Franklin, 2002; Storey et al., 2008). The ability to respire aquatically allows bimodal breathers to prolong dive durations and it has been suggested that supplemental aquatic respiration may reduce the energetic cost of diving, prolong time available for underwater tasks and facilitate the exploitation of oxygen-rich riffle zones (Mathie and Franklin, 2006; Storey et al., 2008; Tucker et al., 2001). Due to these advantages, it has been proposed that bimodal breathers may be less sensitive to the effects of temperature on dive durations compared to divers solely reliant on pulmonary gas exchange. Moreover, bimodal breathers are predicted to upregulate their reliance on aquatic respiration at high temperatures to meet increased metabolic demands and defend dive durations. Studies testing this prediction have returned mixed findings, with some studies suggesting reliance on aquatic respiration is independent of temperature (Bruton et al., 2012; Prassack et al., 2001; Pratt and Franklin, 2010; Udyawer et al., 2016).

Body size is also a key determinant of ectotherm dive durations (Hayward et al., 2016). The influence of body mass on dive durations was originally questioned in ectotherms (Brischoux et al., 2008; Campbell et al., 2010b), however; Hayward et al. (2016) found that after controlling for the effect of temperature, dive durations increase as a power law with body mass. Body oxygen stores typically scale linearly (i.e. isometrically, scaling exponent of 1) with body mass. For example, estuarine crocodiles can undergo a 20, 000-fold increase in body size throughout a lifetime (e.g. 0.05 kg hatchling - 1000 kg adult; Grigg and Kirshner, 2015) and a similar increase in lung oxygen stores (scaling exponent = 0.9; Wright and Kirshner, 1987). In contrast to body oxygen stores, metabolic rate scales allometrically with body mass with an exponent of approximately 0.75. The implications of the differential scaling of oxygen stores and oxygen usage means that larger divers not only have larger oxygen stores, but they consume these stores more efficiently. Therefore, ADLs are mass-dependent in terms of both oxygen stores and usage, such that larger animals can remain submerged far longer than smaller animals before needing to surface. Smaller divers may therefore be disproportionately affected by increases in temperature due to small body oxygen stores and high mass-specific metabolic rates, however the potential moderating effects of body size remain unexplored.

The deleterious effects of climate change on ectotherms diving behaviour may, however, be counteracted by phenotypic plasticity in the form of thermal acclimatization responses (Seebacher et al., 2015). The capacity to responsively alter the thermal sensitivity of underlying physiology in response to a changing environment is termed acclimatization when observed in the field under natural conditions, or alternatively termed acclimation, when observed under experimentally controlled conditions (Wilson and Franklin, 2002). Thermal acclimation/acclimatization can involve physiological remodelling across multiple levels of organismal organisation; from the up- or down-regulation of particular genes (Podrabsky and Somero, 2004), to alterations in enzyme reaction rates, mitochondria density and function, cell membrane thickness, muscle contractility and cardiovascular functioning (Glanville and Seebacher, 2006; Keen et al., 2017; Shuman and J., 2018). In the context of diving at elevated temperatures, thermal acclimation/acclimatization involves a dampening of thermodynamic effects on metabolism at high temperatures, so that body oxygen stores are consumed at a slower rate, and/or an increase in body oxygen stores via an upregulation of oxygen-binding proteins (e.g. haemoglobin, myoglobin). Increases in global temperatures and thermal variability are predicted to create or strengthen selection pressure favouring plastic phenotypes, particularly in long-lived species, like crocodilians, marine turtles and marine iguanas (Kawecki, 2000). A changing environment may also favour an increase in phenotypic variability, such that the probability of a beneficial phenotype arising is maximised (Hansen et al., 2006). Populations with greater phenotypic variation are predicted to cope better with extreme climatic events, like heatwaves, because they are more likely to contain individuals, who by chance, can tolerate the changed conditions (Hansen et al., 2006). Indeed, increases in developmental temperatures have been shown to increase phenotypic variability in fishes, with larger temperature changes seeing greater increases in phenotypic variability (O'Dea et al., 2018). However, it remains unknown if rises in temperature increase phenotypic variability with respect to diving, despite the adaptive potential of this response.

Although it is recognised that climate warming presents a major threat to the persistence of many ectothermic species, the effects of elevated temperature on diving has been largely overlooked. Diving species depend on their capacity to remain submerged to perform a myriad of fitness-related activities, such as foraging, prey-pursuit, predator-avoidance and key social interactions. Two reviews show that dive durations tend to decrease as temperatures rise in ectotherms (Hayward et al., 2016; Jackson, 2007), but a systematic review, with a focus on climate warming, has not yet been conducted. Here, we test whether diving is thermally sensitive in ectotherms using meta-analytic methods. Specifically, we tested the following predictions: (i) acute increases in temperature decrease mean dive durations, (ii) larger increases in temperature will exert a stronger effect on mean dive durations, (iii) the effects of temperature on mean dive durations will be weaker in bimodal breathers compared to aerial breathers, (iv) the effects of temperature on mean dive durations will be weaker in larger (in terms of body mass) divers compared to smaller divers, and (v) chronic exposure to elevated temperature will blunt (i.e. lessen) the effects of temperature on mean dive durations. Because changing environments can also impact phenotypic variability, we explored how elevated temperatures and moderating variables (i.e. magnitude of temperature increase, breathing mode and body size) affect variability in dive durations. These finding were then used to predict how much underwater time could be lost to climate warming and how diving, ectothermic vertebrates may fare in a warmer world.

**Methods**

*Literature search and data extraction*

We searched for experimental studies that measured dive durations (i.e. minutes submerged) in diving vertebrates at two more water temperatures. Searches were conducted using Scopus and Web of Science's (WoS) core collection on 25 February 2020. We used the following search strings: TOPIC: (“dive” OR “diving”) AND (“temperature” OR “thermal” OR “season\*”) AND (“ectotherm\*” OR “reptile\*” OR “\*snake\*” OR “turtle\*” OR “crocod\*” OR “newt\*”) in WoS and TITLE-ABS-KEY (“dive” OR “diving”) AND (“temperature” OR “thermal” OR “season\*”) AND (“ectotherm\*” OR “reptile\*” OR “\*snake\*” OR “turtle\*” OR “crocod\*” OR “newt\*”) in Scopus.

We identified 225 and 222 studies meeting the search criteria in Scopus and WoS, respectively. A total of 149 duplicates (115 identified in EndNote, 32 identified in Rayyan) were removed leaving us with 298 papers for title and abstract screening. We cross-referenced this search with three major reviews: Jackson (2007), Brischoux et al. (2008) and Hayward et al. (2016), and included any papers missed in our initial search (N = x). Title and abstract screening was conducted in Rayyan (Ouzzani et al., 2016) and we excluded studies that: (1) were reviews or commentaries (N = 28), (2) were not on air-breathing, diving ectotherms (N = 33) or (3) did not measure voluntary dive durations across two or more experimental temperatures or field seasons under laboratory or field conditions (N =115). We included field studies where dive durations were measured in free-ranging animals across one or more seasons using biotelemetry tags (e.g. **list examples**). These field studies needed to couple dive duration recordings with either direct body temperature measures or water temperature measurements (assuming animal body temperature equaled water temperature). Only one study met the above criteria for invertebrates (Calosi et al., 2012), so we chose to focus exclusively on vertebrates. We also excluded studies where animals were forced to dive due to atypical physiological alterations induced by forced submergence (**ref**). Studies on leatherback turtles (*Dermochelys coriacea*) were excluded because they are functionally endothermic (N=24; Bostrom and Jones, 2007; Southwood et al., 2005).

Following these exclusion steps, we identified 100 papers with extractable data (Fig. 2). From these papers we extracted means, variance (standard errors, standard deviations or confidence intervals) and sample sizes. We avoided extracting maximum dive durations because maximum dive durations were rarely reported and likely have anaerobic contributions. Data presented in figures were extracted using *metaDigitise* (Pick et al., 2018). Authors were contacted to request missing summary statistics where necessary and if they could not be obtained, we excluded these studies (N=x). We only included comparisons for treatment groups or field recording when they had three or more animals.

*Effect size calculation*

To test the effect of temperature on the mean and variability in dive duration, we calculated two effect sizes, the log response ratio (*ln*RR; Lajeunesse, 2011; Lajeunesse, 2015) and the log coefficient of variation ratio (*ln*CVR; Nakagawa et al., 2015) using the *escalc* function in the *metafor* package (Viechtbauer, 2010) in *R*. To assess differences in mean dive duration we used *ln*RR, which is the natural logarithm of the ratio between mean dive durations between treatments. Changes in the variability in dive duration across treatments was quantified using *ln*CVR, which is the ratio between the coefficients of variation (i.e., SD divided by the mean) for treatments. We used *ln*CVR because we observed strong mean-variance relationships in our data. When dive durations were measured at multiple temperatures, we took each pairwise temperature comparison. For both logged ratios (*ln*RR and *ln*CVR), we specified the control temperature (i.e. cooler temperature) as the denominator and the treatment temperature (i.e. warmer temperature) as the numerator, so that negative effect sizes indicate a decrease in dive duration means or variance at the warmer (treatment) temperature, whereas positive effect sizes indicate an increase in dive duration means or variance at the warmer (treatment) temperature.

*Moderators*

We extracted a series of moderator variables from each study that we predicted would affect both the magnitude and direction of effect sizes. These included: 1) respiration mode (i.e. aerial or bimodal); 2) body mass (g) and 3) the magnitude of temperature change between treatments (i.e. ΔT) and 4) the average temperature of the two treatments (T\_mean). Including the temperature difference between treatments, as well as the mean temperature of treatments, was important because we expected that the magnitude of effects would be larger for experimental treatments farther apart in temperature. Additionally, effect size and magnitude are expected to depend on the general temperature at which dive performance was measured; if treatments were conducted at high temperatures only smaller differences in dive duration are expected. When body mass data were not presented (N =1), we obtained body mass estimates for the same species and life-stage from published literature (Denoel et al., 2005).

*Meta-analysis*

We fit meta-analytic and meta-regression multilevel mixed-effects models using the *rma.mv* function in the *metafor* package in *R.* When the 95% confidence intervals of model estimates did not cross zero, they were considered statistically significant. Respiration mode (i.e. aerial or bimodal), logged body mass (g) and magnitude of temperature change (i.e. ΔT) were included as moderator variables. To visualize the impact of moderator variables, effect sizes were calculated for different magnitudes of temperature increases (i.e. + 3°C, + 5-7°C, + 8-9°C and + ≥ 10°C) and for aerial versus bimodal breathers. Our data contained several sources of non-independence, such as shared sampling covariances between effect sizes, phylogenetic relationships among taxa and study-level non-independence (Noble et al. 2017). We accounted for these sources of non-independence by including random-effects that estimate study and phylogenetic variance as well as a modified sampling (co)variance matrix that explicitly accounted for the covariance between effect sizes sharing treatment groups in their calculation. To obtain a phylogenetic correlation matrix, we first generated a phylogeny by searching for species names in the TimeTree database (timetree.org; Hedges et al., 2006), which uses published molecular data to build a phylogenetic tree with branch lengths (Fig. x). We also estimated a residual variance by including an observation-level random effect as this is not estimated by default in *metafor*. There was one outlier (XX) and we conducted sensitivity analyses to ensure that our results did not change when excluding it from our models. They did not, as such we only report models including the full dataset.

In addition to analyzing contrast-based effect sizes, we also estimated the effect of temperature on *lnMean and lnSD* dive durations directly using a Bayesian arm-based meta-analytic model in the *MCMCglmm* package (Hadfield, 2010). Given all data was in the same units, arm-based models have fewer issues with respect to complex patterns of non-independence and allow for greater flexibility in estimating overall within species and across species temperature effects. Our models accounted for sampling variance for *lnMean* and *lnSD* (see Nakagawa et al 2015 for equations to calculate sampling variance). They also included a within species temperature slope, which we refer to as Tw. This was calculated by centering temperature treatments around the mean for each species (i.e. by subtracting the mean test temperature for each species from each value of the input variable) (van de Pol and Wright, 2009). In addition to these variables, we fit separate models that included fixed effects of log body mass and respiration mode. We also included a random species slope and intercept for Tw to estimate how much variation in slope and intercept exists across species. To evaluate the effect of temperature on variability in dive duration, we modelled the logged standard deviation (*lnSD*) of each observation as a function of logged mean dive duration, Tw, logged body mass and respiration mode as fixed effects. Log mean dive duration was included as a fixed effect to account for mean-variance relationships. We included study- and species-specific random effects. For our species-level random effects we used the phylogenetic covariance matrix and estimated both a random slope and intercept for Tw. MCMC chains were run for 130,000 iterations, with a 30,000 burnin and a thinning interval of 50 (effective sample size = 2000). We visually checked if chains were mixing well and there were no problems of autocorrelation.

*Publication bias*

We explored our data for evidence of publication bias and publication lag effects. Publication bias occur when studies with null or surprising results (generally because of low power) go unpublished (i.e., the ‘file-drawer’ problem). Lag effects can also exist where initial studies provide strong evidence for an effect only to be followed by studies showing weaker effects. We explored publication bias by plotting funnel plots of residuals from our mutilevel regression models that account for known sources of heterogeneity and non-independence (Nakagawa & Santos, 2012), and used Egger’s regression. We used model residuals because strong effect size heterogeneity can lead to what looks to be apparent publication bias (refs). Lag effects were explored by plotting effect size by year of publication, and fitting a multilevel model with publication year. Lag effects would be supported if year had a negative effect on the magnitude of effect size.

*Availability of data and code*

Raw data and code used in the meta-analysis is all located on the Open Science Framework (OSF) webpage (doi).

**Results**

Our laboratory-based dataset includes 15 papers reporting data on 16 species spanning four orders (serpentes, testudines, crocodilians and urodela). We were able to disentangle study effects from species effects because three studies tested more than one species and two species were replicated across multiple studies. Figure x shows the spread of data across the four orders and the number of aerial and bimodal breathing species. We calculated 55 effect sizes for pairwise temperature treatment comparisons. Across species we had between 2 – 4 temperature treatment manipulations within a species with the average temperature difference being 7.4 ± 3.6°C (mean ± S.D.; range = 6 – 20°C) and test temperatures ranging from 9 – 35°C. The average body mass of the diving species was 779.4 ± 940.9 g (mean ± S.D.) and ranged from 2.5 g in juvenile *Natrix maura* to 3813 g in adult *Elseya irwini*. Body masses were comparable between aerial (23.9 ± 7.6 g mean ± S.D.) and bimodal (24.3 ± 2.9 g mean ± S.D.) breathers. We identified an outlier (study 23, observation 48) and verified extracted values with values reported in the publication (Clark et al., 2008). We refit all models excluding the outlier and this did not change model outputs, so the outlier was included in all final models.

*(i) Did acute increases in temperature reduce overall dive duration mean and variability?*

Acute increases in temperature decreased mean dive duration overall by 63% (*ln*RR: -1.003, 95% confidence interval, 95% CI: -1.444 – - 0.563; Figure 1A), but there was high between study heterogeneity (I2study = 73%, 95% CI: 0.533 – 0.860). In contrast, acute increases in temperature had a weak effect on the variability in overall dive duration between temperature treatments (*ln*CVR: 0.021 95% CI: -0.309 – 0.351; an approximately 2% increase in variance, Fig. 1B).

*(ii) Do larger increases in temperature exert a strong effect on dive duration means and variability?*

The magnitude of temperature increase had a significant moderating effect on dive duration means, with a larger increase exerting stronger effects (*lnRR*: -0.111, 95% CI: -0.141 – -0.081). Our arm-based meta-analysis also supported this finding, suggesting that temperature, on average, decreased log mean dive duration within species (Tw = -0.114, 95% 95% CI: -0.602 to 0.302), with an 11% decrease in dive duration for every 1°C increase in test temperature. Within-species slopes and intercepts were also highly variable, suggesting the impact of temperature depends heavily on species-specific responses (σ2slope = 0.18, 95% CI: 0.07–0.311; σ2Intercept = 2.49, 95% CI = 0.369–5.74). Effect size estimates for temperature increases of different magnitudes showed that a + 3°C, + 5-7°C, + 8-9°C and + ≥10°C reduced mean dive durations by 23.1%, 51.2%, 66.7% and 74.9%, respectively (Fig. 1C). In contrast, the magnitude of temperature increase did not have a significant moderating effect on dive duration variability (*ln*CVR: 0.012, 95% CI: -0.031 – 0.054; Figure 1C). Arm-based meta-analytic models also supported this finding suggesting a weak effect on variance (*lnSD*, when controlling for the mean) with changes in temperature (Tw = -0.065, 95% CI: -0.637 – 0.587).

*(iii) Are the effects of temperature on dive duration means and variability weaker in bimodal breathers compared to aerial breathers?*

The effects of temperature on dive duration means were comparable between bimodal breathers (*ln*RR -0.372, 95% CI: -1.473 – 0.73) and aerial breathers (*ln*RR -0.144, CI: - 1.205 – 0.92; Fig. 2A) when controlling for temperature differences and average temperature between groups. No significant difference between these groups was detectable (Contrast-based models: -0.23, 95% CI: -1.053 – 0.597; Arm-based models: -0.45, 95% CI: -2.05 – 1.11). Both groups experienced a 15 – 45% reduction in mean dive durations as temperatures increased. Acute increases in temperature had a tendency (although not significantly so) to increase dive duration variability by similar magnitudes in both bimodal breathers (*ln*CVR 0.451, 95% CI: -0.911 – 1.813) and aerial breathers (*ln*CVR 0.633, 95% CI: -0.541 – 1.808; Fig. 2B). However, there were no significant differences in variability between these groups (Contrast-based models: -0.182, 95% CI: -0.776 – 0.411; Arm-based models: 1.31, 95% CI -0.428 – 2.956).

*(iv) Are the effects of temperature on dive duration means and variability weaker in larger compared to smaller divers?*

Body mass (mean ± S.D.= 779.4 ± 940.9 g) had a borderline significant moderating effect on dive duration means (*ln*RR; 0.126, 95% CI: - 0.009 – 0.262), when controlling for the temperature difference, mean temperature and respiration mode. However, body mass had no effect on the magnitude of differences observed in dive duration variability (*ln*CVR -0.077, 95% CI: -0.200 – 0.0468), when controlling for the temperature difference, mean temperature and respiration mode. This was backed up by results from the arm-based models (*ln*Mean: 0.0513, 95%CI: -0.191 – 0.266; *ln*SD: 0.01, 95% CI: -0.362 – 0.361).

*(v) Does chronic exposure to elevated temperature blunt (i.e. lessen) the effects of temperature on dive duration means and variability?*

There were insufficient data to assess the effects of chronic exposure to elevated temperatures on dive durations. Only four studies (Bruton et al., 2012; Clark et al., 2008; Rodgers and Franklin, 2017; Rodgers et al., 2015) on three species (*Acrochordus arafurae, Elusor macrurus* and *Crocodylus porosus*) assessed the thermal plasticity of dive durations, yielding 10 effect sizes. Since a quantitative analysis was not possible, a qualitative assessment of these data is provided in the discussion.

*(vi) Are effects of temperature on dive duration means and variability more pronounced under laboratory compared to field conditions?*

**Discussion**

Underwater excursions are crucial to the survival and ecological success of many diving species, but we show here that dive durations can be cut short by temperature increases. Our arm-based model showed that for every 1°C increase in water temperature, diving ectotherms experience an 11% decrease in dive durations. Average water temperatures in marine and freshwater habitats are projected to increase between 1.5 – 4°C in the next century, and marine heatwaves are already increasing in intensity, duration and frequency worldwide (Cheng et al., 2019; Hobday and Lough, 2011; Hughes et al., 2017; Stillman, 2019). Our data suggest that this magnitude of warming could translate to substantial cuts in dive durations, by approximately 16 – 44%. Climate warming may therefore reduce time for obligate underwater activities (e.g. predator avoidance, foraging, social interactions) and diving species may be forced to spend more time at the water’s surface.

*The magnitude of temperature increase matters*

As predicted, larger increases in temperature exerted stronger effects on dive duration means, but not variability. Our contrast-based model estimated that + 3°C, + 5–7°C, + 8–9°C and + ≥10°C reduced dive durations by approximately 23%, 51%, 67% and 75%, respectively. This finding is intuitive because larger increases in temperature generally correspond with greater increases in ectothermic oxygen demands, consequently reducing aerobic dive limits by a larger magnitude. Our contrast-based model estimates align well for with our pairwise *Q*10 values for dive durations. *Q*10 values clustered around 0.36 (*Q*10 = 0.36 ± 0.25, mean ± S.D.) which equates to an approximate 64% decrease in dive durations for every 10°C increase in body temperature. Moreover, our estimates are similar to predictions generated from the temperature sensitivity of ectotherm metabolism. Ectotherm metabolic rates typically double or triple with every 10°C increase in body temperature, with *Q*10 values ranging between two and three. Thus, dive durations are expected to reduce by 50 – 67%. Our estimates are on high end this range (75 –77% reductions) but this may attributable to diving metabolic rates sometimes being more temperature sensitive than resting metabolic rates (Rodgers and Franklin, 2017).

The magnitude of warming that aquatic habitats experience moving into the future may therefore moderate how strongly diving ectotherms are impacted. Species or populations inhabiting shallow, exposed (little shade) habitats, such as small lakes and ponds, may suffer from greater declines in dive durations due to their greater heating potential, compared to more thermally stable marine habitats. Human demands placed on freshwater ecosystems, like water extraction and flow regulation, may exacerbate warming in these habitats, as natural flows and depths are often greatly reduced. For example, several freshwater turtles (e.g. *Emydura macquarii*, *Chelodina longicollis* and *Chelodina expansa*) inhabit Australia’s Murray-Darling Basin which is subject to intense water extraction for irrigation purposes and flow regulation (e.g. damming) (Chessman, 2011; Leblanc et al., 2012). Consequently, this habitat experiences reduced flows which increase daily maximum temperatures and exacerbate the impacts of heatwaves (Chessman, 2011; Leblanc et al., 2012). Alternatively, species inhabiting deep, aquatic habitats may be somewhat buffered from the effects of warming if they can dive to cooler, thermally stratified regions of the water column.

*Bimodal breathers are just as vulnerable as aerial breathers*

Although bimodal breathers are predicted to be more resilient to the effects of temperature on dive durations than aerial breathers, we found no significant difference between these groups. Both groups were similarly affected by temperature increases and experienced a 15 – 45% reduction in mean dive durations (mean temperature increase: 7.4 ± 3.6°C). Bimodal breathers have been predicted to defend dive durations at elevated temperatures by increasing their reliance on aquatic respiration to meet increased metabolic demands (Pratt and Franklin, 2010). Bimodal breathers may be able upregulate aquatic respiration by increasing blood flow to the skin to maximise capillary recruitment (Burggren and Moalli, 1984). Alternatively, the rate of diffusion of respiratory gases between the water and skin may be increased by lowering blood PO2 by, for example, initiating a cardiovascular shunt which allows blood to circumvent the lungs and recirculate to the systemic system (Lillywhite and Donald, 1989). However, several studies have shown that bimodal breathers do not upregulate aquatic respiration at high temperatures (Prassack et al., 2001; Pratt and Franklin, 2010; Udyawer et al., 2016). Moreover, there is a marked difference in the temperature-sensitivity of tissue gas diffusion and ectotherm metabolic rate (Jackson, 2007). Tissue gas diffusion increases by only 10% with every 10°C increase in body temperature (*Q*10 = 1.1; Dejours, 1981), whereas ectotherm metabolic rate typically doubles or triples for the same 10°C increase in body temperature (*Q*10 = 2 – 3; Seebacher et al., 2015). Consequently, aquatic respiration rates cannot keep pace with this exponential rise in oxygen demands and the relative contribution of aquatic respiration to total respiration decreases as temperatures increase (Jackson, 2007). Additionally, the solubility of oxygen in water declines as temperature increases, further reducing the efficiency of aquatic respiration. Several studies have shown that the relative contribution of aquatic respiration to total respiration either decreases with rising temperature or remains temperature-independent (Prassack et al., 2001; Pratt and Franklin, 2010; Udyawer et al., 2016). Rather, bimodal respiration becomes advantageous at cold temperatures when aquatic respiration can meet resting or routine metabolic demands, allowing some species to remain submerged for days, weeks and even months at a time as an overwintering strategy (Penney, 1987). The benefits of aquatic respiration in prolonging dive durations therefore only appear to be realized at cooler temperatures (Clark et al., 2008; Prassack et al., 2001). Our findings show that bimodal and aerial breathers are equally sensitive to the negative effects of rising temperatures on dive durations, and we can predict that the diving behaviour of both groups will be disrupted as the climate continues to warm.

*Body mass had a weak moderating effect*

Our contrast-based model showed that body mass had a borderline significant moderating effect on the magnitude of differences observed with increased temperature. As such, the effects of temperature on mean dive duration were weaker in larger divers. This supports our prediction that smaller divers would be disproportionately affected by temperature increases due to their smaller body oxygen stores and higher mass-specific metabolic rates. The effect we observed was weak, likely due to the small body mass range in our data (2.5 – 3813 g). Although this reflects an ~1500-fold change in body mass, very large diving ectotherms, like adult sea turtles and crocodilians, were not included likely owing to the difficulty of maintaining large specimens in the laboratory. The moderating effect of body mass may have been further weakened by unusual scaling relationships between body mass and dive durations in bimodal breathers. For example, the freshwater white-throated snapping turtle (*Elseya albagula*) utilizes cloacal bursae to respire aquatically and small turtles are able to dive for just as long as large turtles (size range: 19 – 6725 g; Mathie and Franklin, 2006). Their capacity to defend dive durations is attributable to smaller turtles having higher mass-specific cloacal bursae surface areas (scaling exponent: Mb0.77), allowing them to extract more oxygen from the water (Mathie and Franklin, 2006). Similar observations have been documented in freshwater turtles (*Trionyx spiniferus, Sternotherus odoratus* and *Kinosternon subrubrum*) utilizing cutaneous respiration, with mass-specific cutaneous surface area decreasing with body mass, with scaling exponents ranging between 0.40 – 0.91 (Stone et al., 1992). These findings suggest that body mass has a stronger moderating effect in aerial compared bimodal breathers, however our contrast-based model showed that there was no significant interaction between respiration mode and body mass. Our findings suggest that large divers may be slightly buffered to the effects of warming on diving durations, but this effect is weak.

*Limited thermal phenotypic plasticity in dive duration*

Thermal acclimation capacity remains unassessed in most ectothermic divers and represents a pressing knowledge gap. Only four studies (Bruton et al., 2012; Clark et al., 2008; Rodgers and Franklin, 2017; Rodgers et al., 2015) have assessed thermal acclimation capacity in diving ectotherms. Nonetheless, the findings from these studies are similar and suggest diving ectotherms have a limited capacity for thermal phenotypic plasticity in dive durations. For example, Bruton et al. (2012) examined the thermal acclimation capacity of the Arafura filesnake (*Acrochordus arafurae*) by holding two groups of snakes at 24°C or 32°C for three months before assessing diving performance at both temperatures. Partial thermal acclimation was observed in maximum dive durations and warm-acclimated snakes dived for longer at both test temperatures. However, this acclimation response was very weak and warm-acclimated snakes still experienced a halving in dive durations between 24°C or 32°C (cool-acclimated *Q*10: 0.39; warm-acclimated *Q*10: 0.41). Moreover, no acclimation response was observed in mean dive durations (Bruton et al., 2012). In a similar study, Clark et al. (2008) exposed hatchling Mary River turtles (*Elusor macrurus*) to either 17°C or 28°C for eight weeks, finding that turtles had the capacity to partially acclimate mean and maximum dive durations to the cool temperature (512% increase in dive duration), but acclimation to the warm temperature was absent. Acclimation of diving performance to temperatures mimicking climate warming scenarios has also been investigated (Rodgers and Franklin, 2017; Rodgers et al., 2015). For example, Rodgers et al. (2015) acclimated juvenile estuarine crocodiles (*C. porosus*) to three climate warming scenarios (current summer, 28°C; moderate climate warming, 31.5°C; high climate warming, 35°C) for 30 days, and found that dive durations decreased dramatically at high temperatures (*Q*10: 0.17) in all three treatment groups. A follow-up study showed that this absence of plasticity in dive durations was underscored by an inability to adjust diving metabolic at elevated temperatures (Rodgers and Franklin, 2017).

*Talk about variability next.*

**Figures**

**A close up of a map

Description automatically generated**

**Figure 1:** Overall meta-analytic results for the effect of an increase in temperature (mean ± S.D. = 7.4 ± 3.6°C) on dive duration means (*ln*RR, **A**) and variability (*ln*CVR, **B**) in diving ectothermic vertebrates. Temperature increases decreased dive mean duration when controlling for the average temperature of the temperature treatment pair and body mass of the species (*lnRR* **C)**. While there was a tendency for the variability in dive duration to increase these effects were not statistically distinguishable form no effect (0) (*ln*CVR, **D**). Data are presented as effect sizes (*ln*RR or *ln*CVR) with 95% confidence intervals. The vertical, dotted zero line indicates no effect and effect sizes are considered significant when 95% confidence intervals do not cross with the zero line. Sample sizes for each group (k) and number of species (Sp) are provided. Abbreviations: ‘NS’ = not statistically significant; ‘\*’ statistically significant deviation from effect size of 0. Between study (I2study) and phylogenetic heterogeneity (I2phylogeny) provided for MLMA models.

**A close up of a map

Description automatically generated**

**Figure 2:** Effects of temperature increase (mean ± S.D. = 7.4 ± 3.6°C) on dive duration means (*ln*RR, **A**)and variability (*ln*CVR, **B**) in all diving ectothermic vertebrates (i.e. overall), aerial breathers and bimodal breathers. The effects of temperature increases on dive duration means and variability were comparable between aerial and bimodal breathers, with dive duration means decreasing and dive duration variability increasing. Data are presented as effect sizes (*ln*RR or *ln*CVR) with 95% confidence intervals. The vertical, dotted zero line indicates no effect and effect sizes are considered significant when 95% confidence intervals do not cross with the zero line. Sample sizes for each group are included in parentheses. Different lowercase letters indicate where groups are statistically different from each other.

**References**

**Aubret, F., Tort, M. and Sarraude, T.** (2015). Evolution of alternative foraging tactics driven by water temperature and physiological constraints in an amphibious snake. *Biological Journal of the Linnean Society* **115**, 411-422.

**Bentivegna, F., Hochscheid, S. and Minucci, C.** (2003). Seasonal variability in voluntary dive duration of the Mediterranean loggerhead turtle, Caretta caretta. *Scientia Marina* **67**, 371-375.

**Bostrom, B. L. and Jones, D. R.** (2007). Exercise warms adult leatherback turtles. *Comparative Biochemistry & Physiology Part A: Molecular and Integrative Physiology* **147**, 323-331.

**Bradshaw, C. J. A., McMahon, C. R. and Hays, G. C.** (2007). Behavioral inference of diving metabolic rate in free-ranging leatherback turtles. *Physiological and Biochemical Zoology* **80**, 209-219.

**Brischoux, F., Bonnet, X., Cook, T. R. and Shine, R.** (2008). Allometry of diving capacities: ectothermy vs. endothermy. *Journal of Evolutionary Biology* **21**, 324-329.

**Bruton, M. J., Cramp, R. L. and Franklin, C. E.** (2012). Benefits of thermal acclimation in a tropical aquatic ectotherm, the Arafura filesnake, Acrochordus arafurae. *Journal of Comparative Physiology B-Biochemical Systems and Environmental Physiology* **182**, 541-551.

**Burggren, W. and Moalli, R.** (1984). ‘Active’ regulation of cutaneous gas exchange by capillary recruitment in amphibians: experimental evidence and a revised model for skin respiration. . *Respiration Physiology* **55**, 379-392.

**Butler, P. J.** (2006). Aerobic dive limit. What is it and is it always used appropriately? *Comparative Biochemistry and Physiology, Part A* **145**, 1-6.

**Butler, P. J. and Jones, D. R.** (1982). The comparative physiology of diving in vertebrates. *Advances in Comparative Physiology and Biochemistry* **8**, 179-364.

**Calosi, P., Bilton, D. T., Spicer, J. I., Verberk, W., Atfield, A. and Garland, T.** (2012). The comparative biology of diving in two genera of European Dytiscidae (Coleoptera). *Journal of Evolutionary Biology* **25**, 329-341.

**Campbell, H. A., Dwyer, R. G., Gordos, M. and Franklin, C. E.** (2010a). Diving through the thermal window: implications for a warming world. *Proceedings: Biological Sciences* **277**, 3837-3844.

**Campbell, H. A., Sullivan, S., Read, M. A., Gordos, M. A. and Franklin, C. E.** (2010b). Ecological and physiological determinants of dive duration in the freshwater crocodile. *Functional Ecology* **24**, 103-111.

**Campbell, H. A., Watts, M. E., Sullivan, S., Read, M. A., Choukroun, S., Irwin, S. R. and Franklin, C. E.** (2010c). Estuarine crocodiles ride surface currents to facilitate long-distance travel. *Journal of Animal Ecology* **79**, 955-964.

**Cheng, L., Abraham, J., Hausfather, Z. and Trenberth, K. E.** (2019). How fast are the oceans warming? Observational records of ocean heat content show that ocean warming is accelerating. *Science* **363**, 128.

**Chessman, B. C.** (2011). Declines of freshwater turtles associated with climatic drying in Australia’s Murray–Darling Basin. *Wildlife Reserach* **38**, 664-671.

**Clark, N. J., Gordos, M. A. and Franklin, C. E.** (2008). Thermal plasticity of diving behavior, aquatic respiration, and locomotor performance in the Mary River turtle Elusor macrurus. *Physiological and Biochemical Zoology* **81**, 301-309.

**Costa, D. P.** (2007). Diving physiology of marine vertebrates. In *Encyclopedia of Life Sciences* pp. 1-7. Chichester: John Wiley & Sons Ltd.

**Dejours, P.** (1981). Principles of Camparative Respiratory Physiology. Amsterdam: Elseview-North Holland Biomedical Press.

**Denoel, M., Mathieu, M. and Poncin, P.** (2005). Effect of water temperature on the courtship behavior of the Alpine newt *Triturus alpestris*. *Behavioral Ecology and Sociobiology* **58**, 121-127.

**Feder, M. E. and Burggren, W. W.** (1985). Cutaneous gas exchange in vertebrates: Design, patterns, control and implications. *Biological Reviews* **60**, 1-45.

**FitzGibbon, S. I. and Franklin, C. E.** (2010). The importance of the cloacal bursae as the primary site of aquatic respiration in the freshwater turtle, Elseya albagula. *Australian Zoologist* **35**, 276-282.

**Fuster, J. F., Pages, T. and Palacios, L.** (1997). Effect of temperature on oxygen stores during aerobic diving in the freshwater turtle Mauremys caspica leprosa. *Physiological Zoology* **70**, 7-18.

**Glanville, E. J. and Seebacher, F.** (2006). Compensation for environmental change by complementary shifts of thermal sensitivity and thermoregulatory behaviour in an ectotherm. *Journal of Experimental Biology* **209**, 4869-4877.

**Gordos, M. and Franklin, C. E.** (2002). Diving behaviour of two Australian bimodally respiring turtles, Rheodytes leukops and Emydura macquarii, in a natural setting. *Journal of Zoology* **258**, 335-342.

**Gordos, M. A., Franklin, C. E. and Limpus, C. J.** (2003). Seasonal changes in the diel surfacing behaviour of the bimodally respiring turtle Rheodytes leukops. *Canadian Journal of Zoology-Revue Canadienne De Zoologie* **81**, 1614-1622.

**Grigg, G. C. and Kirshner, D.** (2015). Biology and evolution of crocodylians. . Clayton South, Australia: CSIRO Publishing.

**Hadfield, J. D.** (2010). MCMC Methods for Multi-Response Generalized Linear Mixed Models: The MCMCglmm R Package. *Journal of Statistical Software* **33**, 1-22.

**Hansen, T. F., Carter, A. J. R. and Pélabon, C.** (2006). On adaptive accuracy and precision in natural populations. *The American Naturalist* **168**, 168-181.

**Hayward, A., Pajuelo, M., Haase, C. G., Anderson, D. M. and Gillooly, J. F.** (2016). Common metabolic constraints on dive duration in endothermic and ectothermic vertebrates. *Peerj* **4**, 9.

**Hedges, S. B., Dudley, J. and Kumar, S.** (2006). TimeTree: A public knowledge-base of divergence times among organisms. *Bioinformatics* **22**, 2971-2972.

**Heithaus, M. R. and Dill, L. M.** (2002). Food availability and predation risk influence bottlenose dolphin habitat use. *Ecology* **83**, 480-491.

**Heithaus, M. R. and Dill, L. M.** (2003). Optimal diving under the risk of predation. *Journal of Theoretical Biology* **223**, 79-92.

**Heithaus, M. R., Dill, L. M., Marshall, G. J. and Buhleier, B. M.** (2002). Habitat use and foraging behavior of tiger sharks (Galeocerdo cuvier) in a seagrass ecosystem. *Marine Biology* **140**, 237-248.

**Hobday, A. J. and Lough, J. M.** (2011). Projected climate change in Australian marine and freshwater environments. *Marine and Freshwater Research* **62**, 1000-1014.

**Hughes, T. P., Kerry, J. T., Álvarez-Noriega, M., Álvarez-Romero, J. G., Anderson, K. D., Baird, A. H., Babcock, R. C., Beger, M., Bellwood, D. R., Berkelmans, R. et al.** (2017). Global warming and recurrent mass bleaching of corals. *Nature* **543**, 373-377.

**Jackson, D. C.** (2007). Temperature and hypoxia in ectothermic tetrapods. *Journal of Thermal Biology* **32**, 125-133.

**Kawecki, T. J.** (2000). The evolution of canalization under fluctuating selection. *Evolution* **54**, 1-12.

**Keen, A. N., Jordan, M. K., Holly, S. A. and Gillis, T. E.** (2017). Temperature-induced cardiac remodelling in fish. *Journal of Experimental Biology* **220**, 147-160.

**Lajeunesse, M. J.** (2011). On the meta-analysis of response ratios for studies with correlatedand multi-group designs. *Ecology* **92**, 2049-2055.

**Lajeunesse, M. J.** (2015). Bias and correction for the log-response ratio in ecological meta-analysis. *Ecology* **96**, 2056-2063.

**Leblanc, M., Tweed, S., Van Dijk, A. and Timbal, B.** (2012). A review of historic and future hydrological changes in the Murray-Darling Basin. *Global and Planetary Change* **80**, 226-246.

**Lillywhite, H. B. and Donald, J. A.** (1989). Pulmonary blood flow regulation in an aquatic snake. *Science* **245**, 293-295.

**Maina, J. N.** (2002). Structure, function and evolution of the gas exchangers: comparative perspectives. *Journal of Anatomy* **201**, 281-304.

**Mathie, N. J. and Franklin, C. E.** (2006). The influence of body size on the diving behaviour and physiology of the bimodally respiring turtle, Elseya albagula. *Journal of Comparative Physiology B-Biochemical Systemic and Environmental Physiology* **176**, 739-747.

**Nakagawa, S., Poulin, R., Mengersen, K., Reinhold, K., Engqvist, L., Lagisz, M. and Senior, A. M.** (2015). Meta-analysis of variation: Ecological and evolutionary applications and beyond. *Methods in Ecology and Evolution* **6**, 143-152.

**O'Dea, R. E., Lagisz, M., Hendry, A. P. and Nakagawa, S.** (2018). Developmental temperature affects phenotypic means and

variability: A meta‐analysis of fish data. *Fish and Fisheries* **20**, 1005-1022.

**Ouzzani, M., Hammady, H., Fedorowicz, Z. and Elmagarmid, A.** (2016). Rayyan — a web and mobile app for systematic reviews. *Systematic Reviews* **5**, 210.

**Penney, D. G.** (1987). Frogs and turtles: Different ectotherm overwintering strategies. *Comparative Biochemistry & Physiology Part A: Molecular and Integrative Physiology* **86**, 609-615.

**Pick, J. L., Nakagawa, S. and Noble, D. W. A.** (2018). Reproducible, flexible and high-throughput data extraction from primary literature: The metaDigitise R package. *Methods in Ecology and Evolution* **10**, 426-431.

**Podrabsky, J. E. and Somero, G. N.** (2004). Changes in gene expression associated with acclimation to constant temperatures and fluctuating daily temperatures in an annual killifish Austrofundulus limnaeus. *Journal of Experimental Biology* **207**, 2237-2254.

**Prassack, S. L., Bagatto, B. and Henry, R. P.** (2001). Effects of temperature and aquatic Po-2 on the physiology and behaviour of Apalone ferox and Chrysemys picta. *Journal of Experimental Biology* **204**, 2185-2195.

**Pratt, K. L. and Franklin, C. E.** (2010). Temperature independence of aquatic oxygen uptake in an air-breathing ectotherm and the implications for dive duration. *Comparative Biochemistry and Physiology a-Molecular & Integrative Physiology* **156**, 42-45.

**Rodgers, E. M. and Franklin, C. E.** (2017). Physiological mechanisms constraining ectotherm fright-dive performance at elevated temperatures. *Journal of Experimental Biology* **220**, 3556-3564.

**Rodgers, E. M., Schwartz, J. J. and Franklin, C. E.** (2015). Diving in a warming world: the thermal sensitivity and plasticity of diving performance in juvenile estuarine crocodiles (Crocodylus porosus). *Conservation Physiology* **3**, 9.

**Seebacher, F., White, C. R. and Franklin, C. E.** (2015). Physiological plasticity increases resilience of ectothermic animals to climate change. *Nature Climate Change* **5**, 61-66.

**Seymour, R. S.** (1982). Physiological adaptations to aquatic life. In *Biology of the Reptilia, Physiological Ecology*, vol. 13 eds. C. Gans and F. H. Pough), pp. 1-51: Academic Press.

**Shuman, J. L. and J., C. D.** (2018). Red muscle function and thermal acclimation to cold in rainbow smelt, Osmerus mordax, and rainbow trout, Oncorhynchus mykiss. *Journal of Experimenta Zoology Part A* **329**, 547-556.

**Southwood, A. L., Andrews, R. D., Paladino, F. V. and Jones, D. R.** (2005). Effects of diving and swimming behavior on body temperatures of Pacific leatherback turtles in tropical seas. *Physiological and Biochemical Zoology* **78**, 285-297.

**Stillman, J. H.** (2019). Heat Waves, the New Normal: Summertime Temperature Extremes Will Impact Animals, Ecosystems, and Human Communities. *Physiology (Bethesda, Md.)* **34**, 86-100.

**Stone, P. A., Dobie, J. L. and Henry, R. P.** (1992). Cutaneous surface-area and bimodal respiration in soft-shelled (Trionyx-spiniferus), stinkpot (Sternotherus-odoratus), and mud turtles (Kinosternon-subrubrum). *Physiological Zoology* **65**, 311-330.

**Storey, E. M., Kayes, S. M., De Vries, I. and Franklin, C. E.** (2008). Effect of water depth, velocity and temperature on the surfacing frequency of the bimodally respiring turtle Elseya albagula. *Functional Ecology* **22**, 840-846.

**Tucker, A. D., Limpus, C. J., Priest, T. E., Cay, J., Glen, C. and Guarino, E.** (2001). Home ranges of Fitzroy River turtles (Rheodytes leukops) overlap riffle zones: potential concerns related to river regulation. *Biological Conservation* **102**, 171-181.

**Udyawer, V., Simpfendorfer, C. A., Heupel, M. R. and Clark, T. D.** (2016). Coming up for air: thermal dependence of dive behaviours and metabolism in sea snakes. *Journal of Experimental Biology* **219**, 3447-3454.

**van de Pol, M. and Wright, J.** (2009). A simple method for distinguishing within- versus between-subject effects using mixed models. *Animal Behaviour* **77**, 753-758.

**Viechtbauer, W.** (2010). Conducting meta-analyses in R with the metafor package. *Journal of Statistical Software* **36**, 1-48.

**Wilson, R. S. and Franklin, C. E.** (2002). Testing the beneficial acclimation hypothesis. *Trends in Ecology & Evolution* **17**, 66-70.

**Wright, J. C.** (1986). Low to negligible cutaneous oxygen uptake in juvenile Crocodylus porosus. *Comparative Biochemistry and Physiology a-Molecular & Integrative Physiology* **84**, 479-481.

**Wright, J. C. and Kirshner, D.** (1987). Allometry of lung volume during voluntary submergence in the saltwater crocodile Crocodylus porosus. *Journal of Experimental Biology* **130**, 433-436.