

STAT 331 Final Project

Maxine, Estella, Judy, Weiwei

04/12/2021

1. Summary

A maximum of 200 words describing the objective of the report, an overview of the statistical analysis, and summary of the main results.

2. Objective

The goal of this project is to analyze the pollutants.csv data and write a report on your analysis. The specific goals of your analysis are up to you to decide.

3. Exploratory Data Analysis

Conduct exploratory data analyses: report summary statistics, visualize data (histograms, scatter plots, etc.). Report on any interesting findings and comment on how these inform the rest of your analysis.

can use this as a tutorial <https://r4ds.had.co.nz/exploratory-data-analysis.html>

Import dataset

```
# CHANGE ABSOLUTE PATH
```

```
# setwd("~/Desktop/stat341/R331project/data")
setwd("~/School/4A/STAT 331/R331project/data")
# setwd("~/Desktop/R331project/data")
```

```
pollutants_raw <- read.csv("pollutants.csv", header = TRUE)
names(pollutants_raw)
```

```
## [1] "X"                "length"          "POP_PCB1"        "POP_PCB2"
## [5] "POP_PCB3"         "POP_PCB4"        "POP_PCB5"        "POP_PCB6"
## [9] "POP_PCB7"         "POP_PCB8"        "POP_PCB9"        "POP_PCB10"
## [13] "POP_PCB11"        "POP_dioxin1"     "POP_dioxin2"     "POP_dioxin3"
## [17] "POP_furan1"       "POP_furan2"     "POP_furan3"     "POP_furan4"
## [21] "whitecell_count"  "lymphocyte_pct"  "monocyte_pct"    "eosinophils_pct"
## [25] "basophils_pct"    "neutrophils_pct" "BMI"             "edu_cat"
## [29] "race_cat"         "male"           "ageyrs"          "yrssmoke"
## [33] "smokenow"         "ln_lbxcot"
```

Note that “edu_cat”, “race_cat”, “male”, “smokenow” are categorical data and X is the index column.

```
# Man's work
# clean the pollutants dataframe
pollutants <- subset(pollutants_raw , select = -X)
```

```

# deal with categorical data

# 1 = Less Than 9th Grade or 9-11th Grade (Includes 12th grade with no diploma)
# 2 = High School Grad/GED or Equivalent
# 3 = Some College or AA degree
# 4 = College Graduate
edu_factor=factor(pollutants$edu_cat)

# 1 = Other Race (Including Multi-Racial);
# 2 = Mexican American;
# 3 = Non-Hispanic Black;
# 4 = Non-Hispanic White
race_factor=factor(pollutants$race_cat,
                    labels = c("Other", "Mexican", "Black", "White"))

# 0 = does not currently smoke;
# 1 = currently smokes
smoke_factor=factor(pollutants$smokenow, labels = c("Non-Smoker", "Smoker"))

# 0 = female, 1 = male
gender_factor=factor(pollutants$male, labels = c("female", "male"))

pollutants$edu_cat = edu_factor
pollutants$race_cat = race_factor
pollutants$smokenow = smoke_factor
pollutants$male = gender_factor

```

Get the names of covariates after we have performed some cleaning on the data

```

names(pollutants)

## [1] "length"          "POP_PCB1"         "POP_PCB2"         "POP_PCB3"
## [5] "POP_PCB4"         "POP_PCB5"         "POP_PCB6"         "POP_PCB7"
## [9] "POP_PCB8"         "POP_PCB9"         "POP_PCB10"        "POP_PCB11"
## [13] "POP_dioxin1"      "POP_dioxin2"      "POP_dioxin3"      "POP_furan1"
## [17] "POP_furan2"      "POP_furan3"      "POP_furan4"      "whitecell_count"
## [21] "lymphocyte_pct"   "monocyte_pct"     "eosinophils_pct"  "basophils_pct"
## [25] "neutrophils_pct" "BMI"              "edu_cat"          "race_cat"
## [29] "male"             "ageyrs"           "yrssmoke"         "smokenow"
## [33] "ln_lbxcot"

```

Data Distribution

We investigate the distribution of covariates from the supplied data.

```

# Max's work
# put bargraphs for categorical data onto one picture
par(mfrow=c(2,2))

plot(edu_factor,
     main="Distribution of Education",
     xlab="Education Level Count")

plot(race_factor,

```

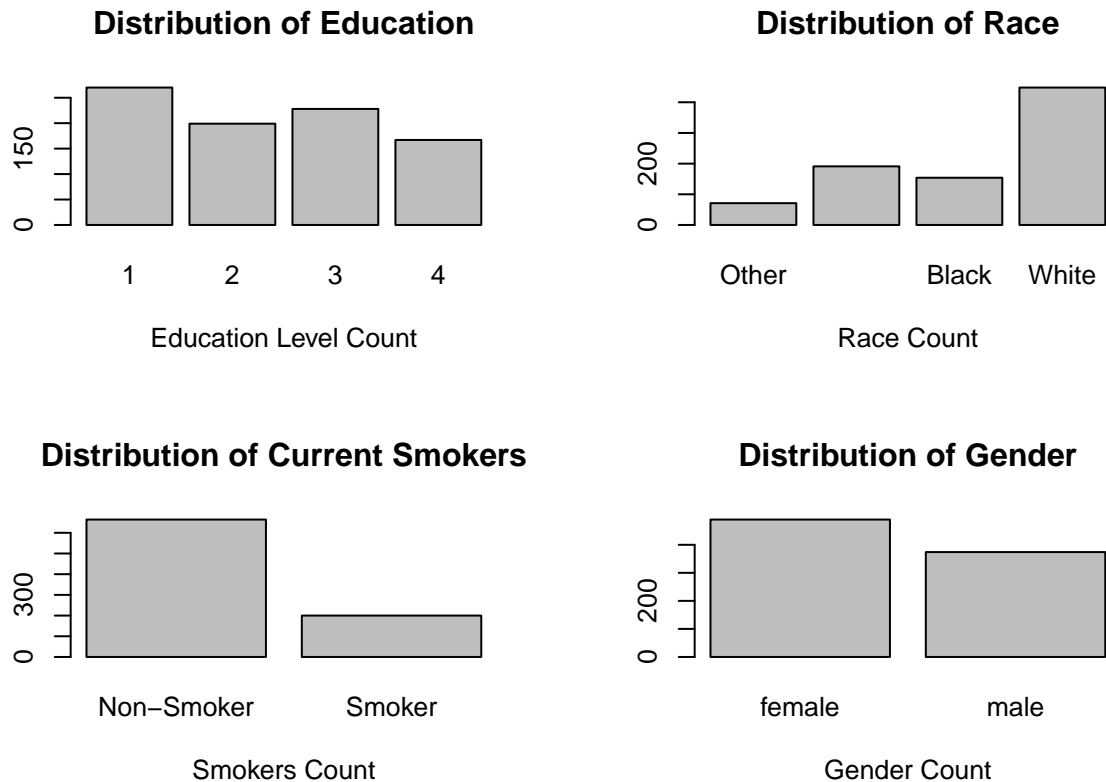
```

    main="Distribution of Race",
    xlab="Race Count")

plot(smoke_factor,
     main="Distribution of Current Smokers",
     xlab="Smokers Count")

plot(gender_factor,
     main="Distribution of Gender",
     xlab="Gender Count")

```



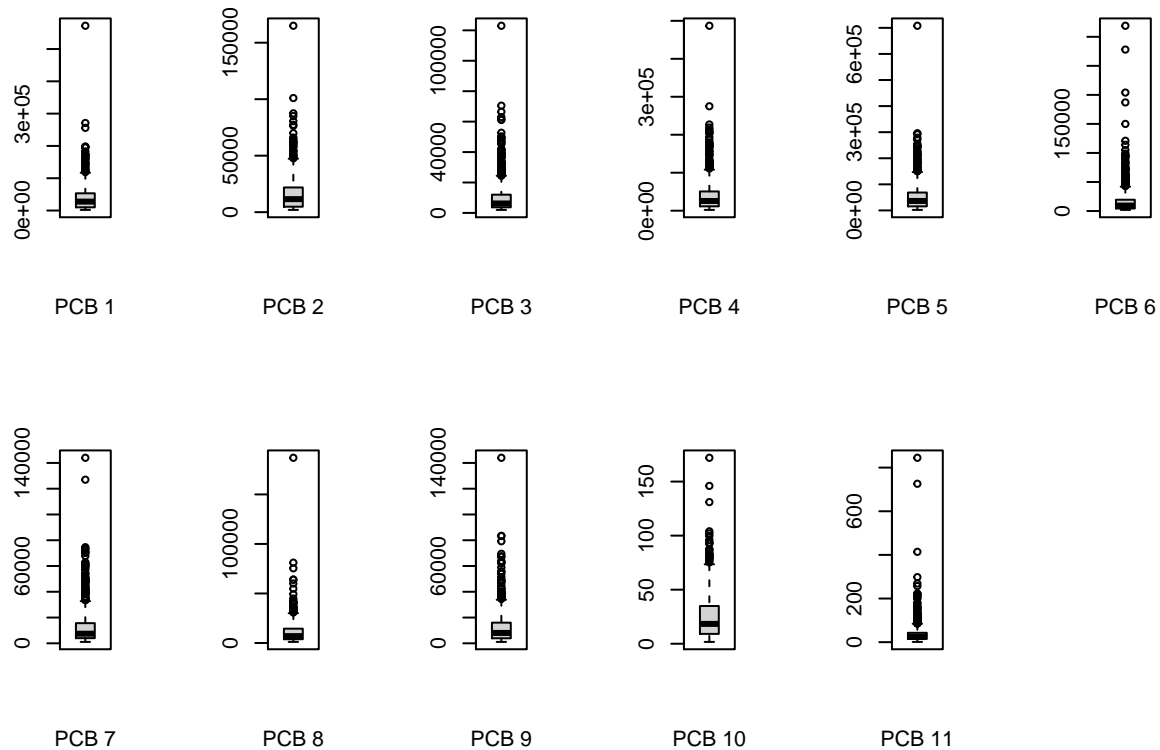
We see that we have more data about non-smokers than smokers and white people than other races. There are more entries for lower-education than higher, and more female than male. However, the distribution of gender and education are relatively close.

```

# Men's work
# PCB 1-6
par(mfrow=c(2,6))
boxplot(pollutants[, 2], xlab="PCB 1")
boxplot(pollutants[, 3], xlab="PCB 2")
boxplot(pollutants[, 4], xlab="PCB 3")
boxplot(pollutants[, 5], xlab="PCB 4")
boxplot(pollutants[, 6], xlab="PCB 5")
boxplot(pollutants[, 7], xlab="PCB 6")
boxplot(pollutants[, 8], xlab="PCB 7")
boxplot(pollutants[, 9], xlab="PCB 8")
boxplot(pollutants[, 10], xlab="PCB 9")
boxplot(pollutants[, 11], xlab="PCB 10")
boxplot(pollutants[, 12], xlab="PCB 11")

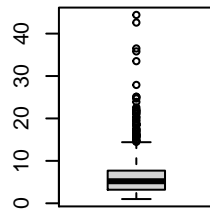
```

```
# Dioxin
par(mfrow=c(2,4))
```

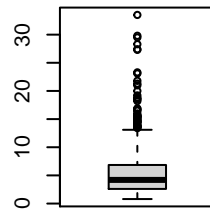


```
boxplot(pollutants[, 16], xlab="Furan 1")
boxplot(pollutants[, 17], xlab="Furan 2")
boxplot(pollutants[, 18], xlab="Furan 3")
boxplot(pollutants[, 19], xlab="Furan 4")
# Furan
boxplot(pollutants[, 13], xlab="Dioxin 1")
boxplot(pollutants[, 14], xlab="Dioxin 2")
boxplot(pollutants[, 15], xlab="Dioxin 3")

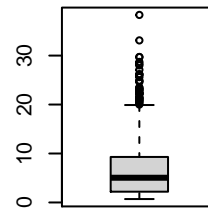
# white blood cells and concentrations
par(mfrow=c(2,6))
```



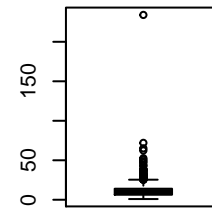
Furan 1



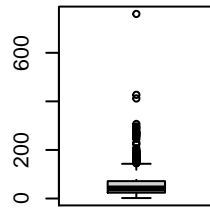
Furan 2



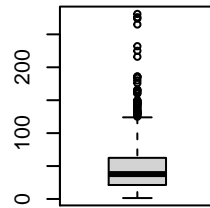
Furan 3



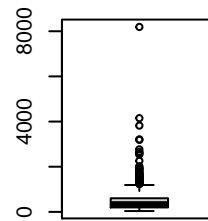
Furan 4



Dioxin 1



Dioxin 2

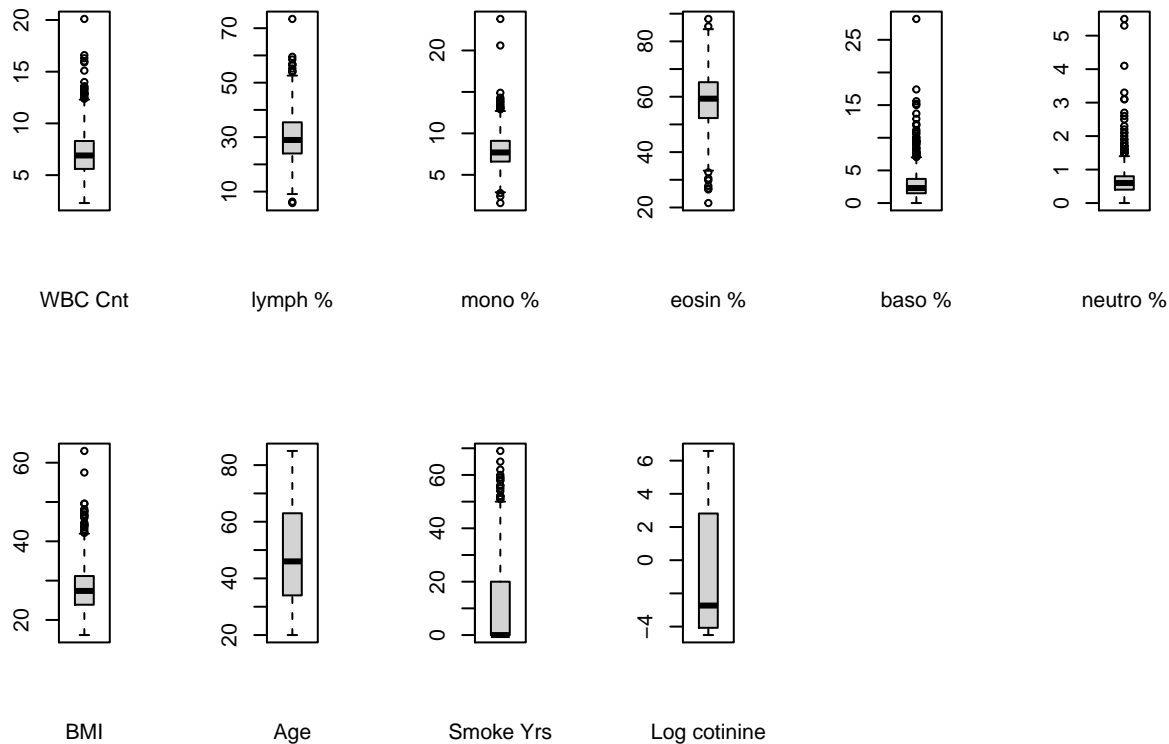


Dioxin 3

```

boxplot(pollutants[, 20], xlab="WBC Cnt")
boxplot(pollutants[, 21], xlab="lymph %")
boxplot(pollutants[, 22], xlab="mono %")
boxplot(pollutants[, 23], xlab="eosin %")
boxplot(pollutants[, 24], xlab="baso %")
boxplot(pollutants[, 25], xlab="neutro %")
# others
boxplot(pollutants[, 26], xlab="BMI")
boxplot(pollutants[, 30], xlab="Age")
boxplot(pollutants[, 31], xlab="Smoke Yrs")
boxplot(pollutants[, 33], xlab="Log cotinine")

```



We see that there are some extreme outliers in some concentration of PCBs, Dioxins, and Furan. The maximum values are sometimes over double the magnitude of the second largest.

However with a little investigation in `@ref(#outlier-entries)`, we see that outliers for PCB values mostly came from one observation.

```
pollutants[436, 3:12]
```

```
##      POP_PCB2 POP_PCB3 POP_PCB4 POP_PCB5 POP_PCB6 POP_PCB7 POP_PCB8 POP_PCB9
## 436  165000  123000  487000   708000  319000  127000  187000  144000
##      POP_PCB10 POP_PCB11
## 436      131      137
```

Similarly, the most extreme outliers for different types of Dioxin and Furan also came from the same entry of data:

- Entry 285 contain the highest value for Dioxin 1 and 3, which are extreme outliers as we can see from the boxplots
- Entry 559 contain the highest value for Furan 2 and 4, where Furan 4 has an extreme outlier

Other covariates do not have a common entry that contribute to the outliers.

Multicollinearity

Correlation among PCB Concentrations

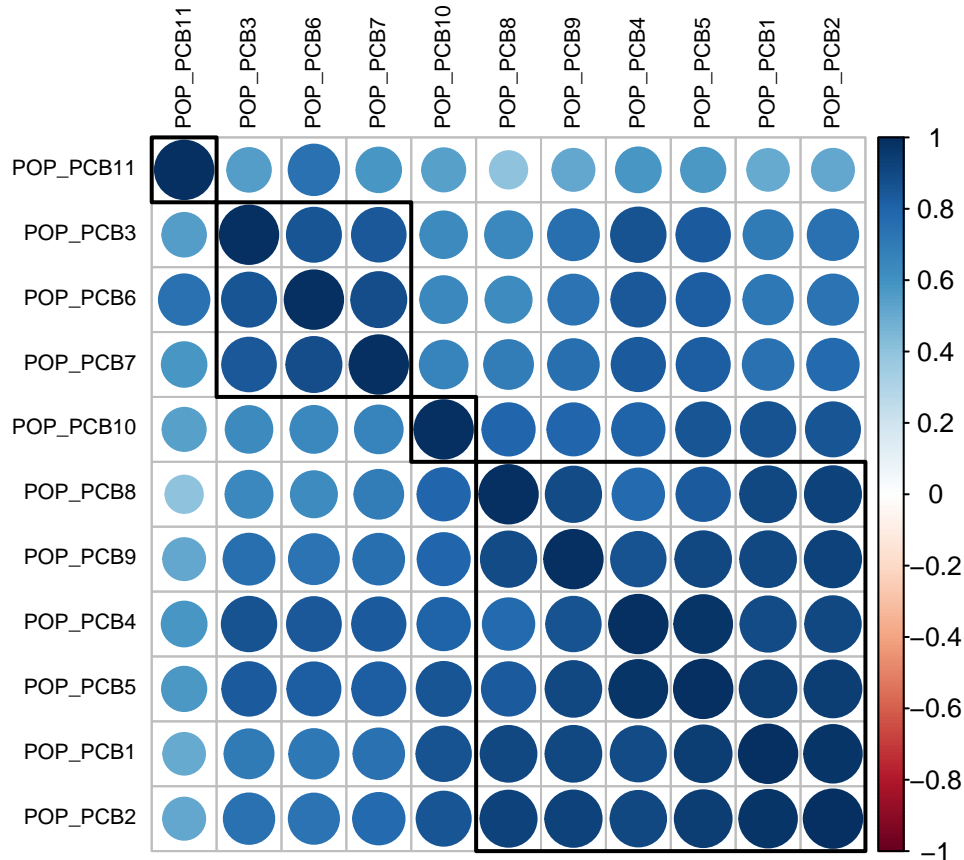
```
# Estella's work 1
library(corrplot)
```

```
## corrplot 0.84 loaded
```

```
library(ggplot2)

POP_PCB = c("POP_PCB1", "POP_PCB2", "POP_PCB3", "POP_PCB4",
            "POP_PCB5", "POP_PCB6", "POP_PCB7", "POP_PCB8",
            "POP_PCB9", "POP_PCB10", "POP_PCB11")
POP_PCB_data <- pollutants[, POP_PCB]
cc = cor(POP_PCB_data, method = "spearman")

# cluster my POP_PCB so that those with similar patterns
# of correlation coefficients are closer together.
# https://jkeorcz.github.io/2019/06/11/Correlation-heatmaps.html
corrplot(cc, tl.col = "black", order = "hclust", hclust.method = "average",
         addrect = 4, tl.cex = 0.7)
```



Correlation among PCB Concentrations

4. Methods:

Describe your statistical analysis: What is your model? Did you use any transformations or extensions of the basic multiple linear regression model? How did you select a model? Does the model fit the data well? Are the necessary assumptions met? Be sure to explain and justify your decisions.

```
train_data <- pollutants[1:600,]
test_data <- pollutants[601:nrow(pollutants),]
```

```

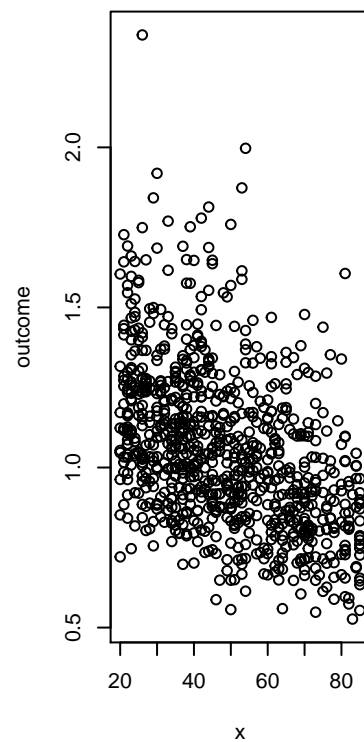
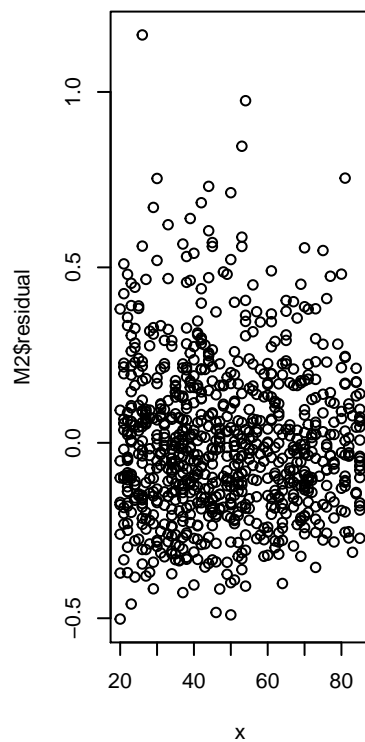
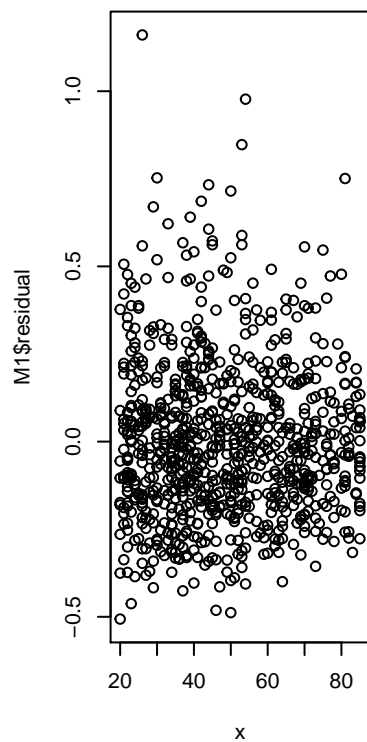
# Judy's work Part 1
# testing non-linearity in SLR
# if for any covariate, residual vs x for M1 has a pattern and
# residual vs x for M2 seems random, then y has a nonlinear
# relationship with with x.
# M1: fitting y to x
# M2: fitting y to x^2

par(mfrow=c(1, 3))
outcome <- pollutants$length
check <- function(x) {
  M1 <- lm(outcome ~ x)
  print(paste("residual for M1: ", sigma(M1)))
  M2 <- lm(outcome ~ x + I(x^2))
  print(paste("residual for M2: ", sigma(M2)))
  plot(x, M1$residual)
  plot(x, M2$residual)
  plot(x, outcome)
}

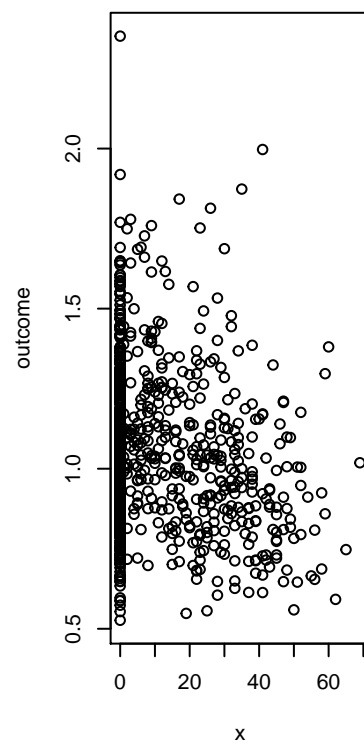
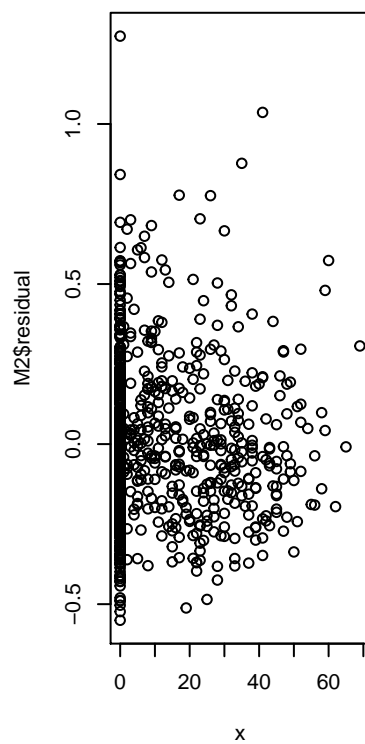
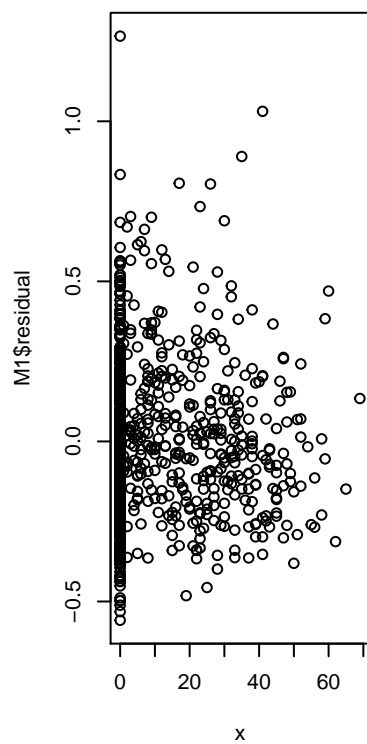
list <- list(pollutants$ageyrs, pollutants$yrssmoke,
             pollutants$BMI, pollutants$ln_lbxcot,
             pollutants$whitecell_count, pollutants$lymphocyte_pct,
             pollutants$monocyte_pct, pollutants$eosinophils_pct,
             pollutants$basophils_pct, pollutants$neutrophils_pct)
for (column in list) {
  check(column)
}

## [1] "residual for M1:  0.224172364185412"
## [1] "residual for M2:  0.22429269961392"

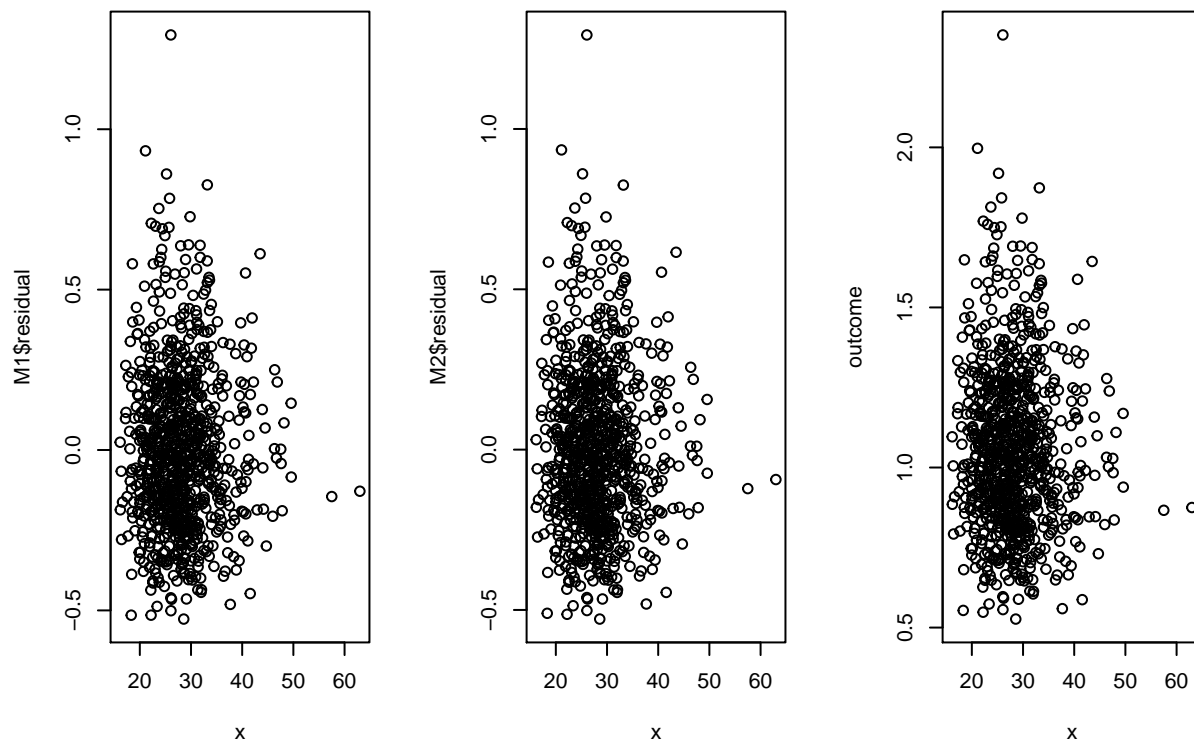
```

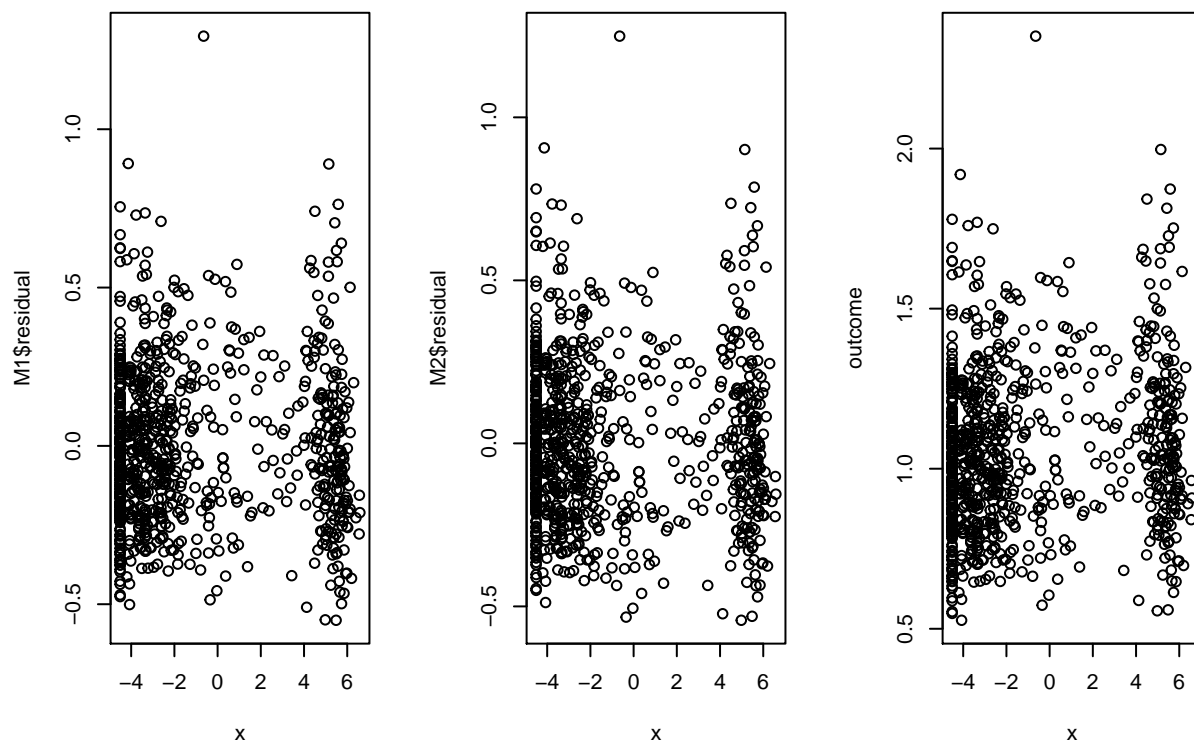
```
## [1] "residual for M1: 0.246320733146214"
## [1] "residual for M2: 0.245622720856213"
```



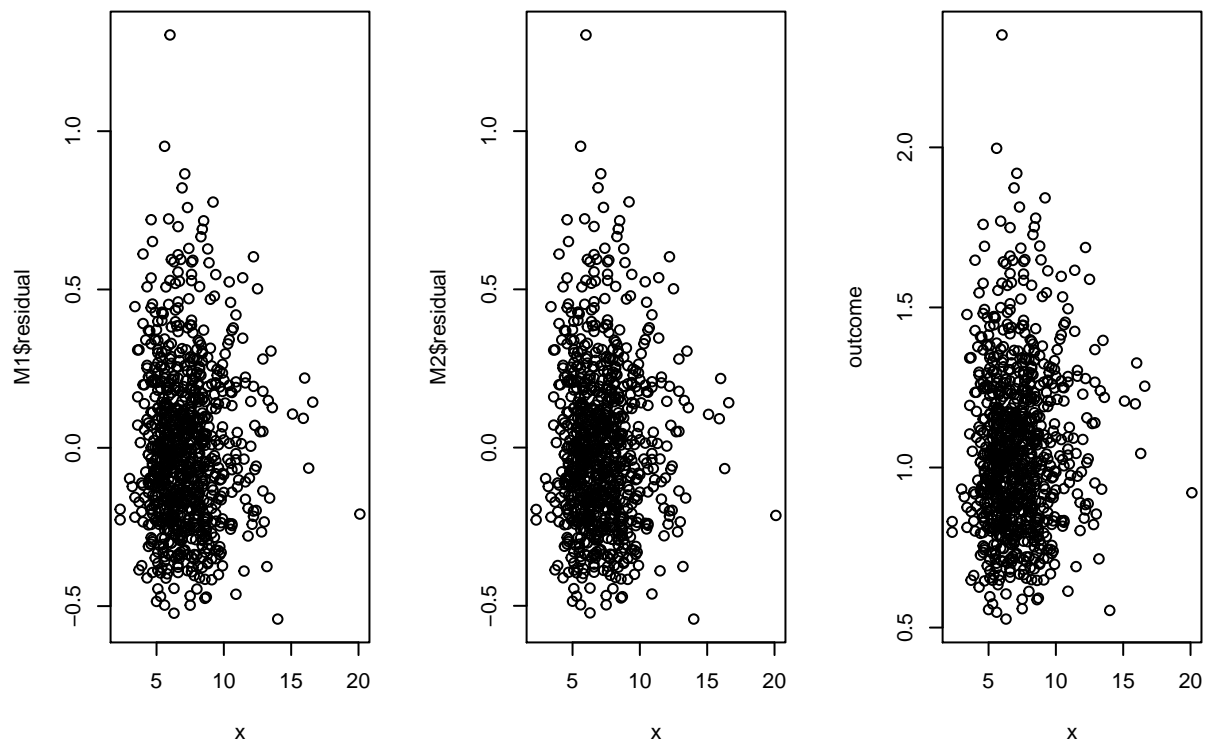
```
## [1] "residual for M1: 0.250228706427173"
## [1] "residual for M2: 0.25036248052387"
```



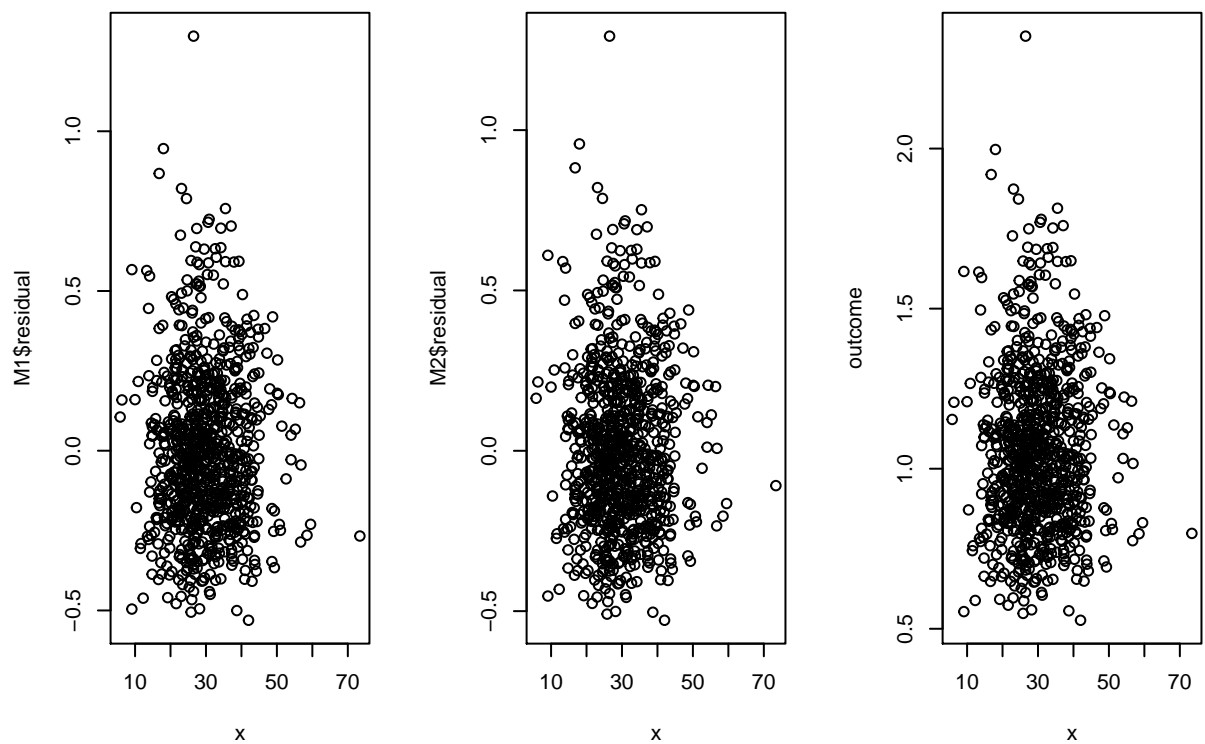
```
## [1] "residual for M1: 0.248212063673837"
## [1] "residual for M2: 0.24710732733351"
```



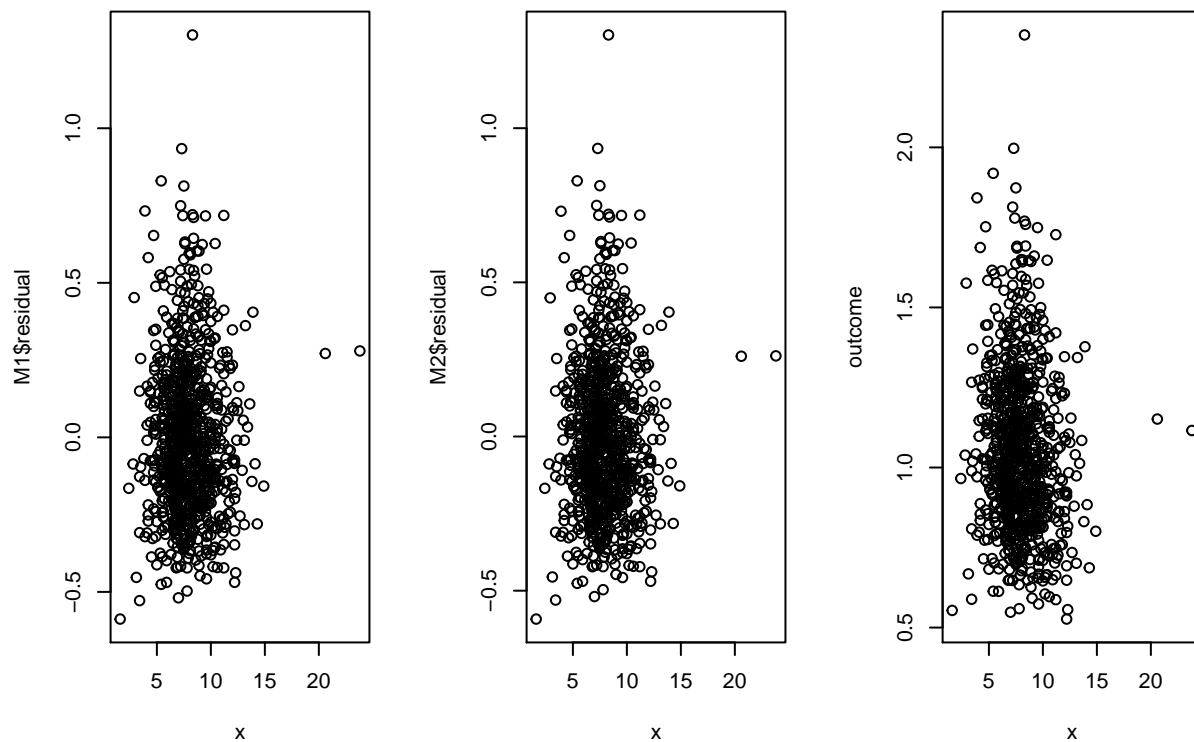
```
## [1] "residual for M1: 0.250065445847753"
## [1] "residual for M2: 0.250210403543218"
```



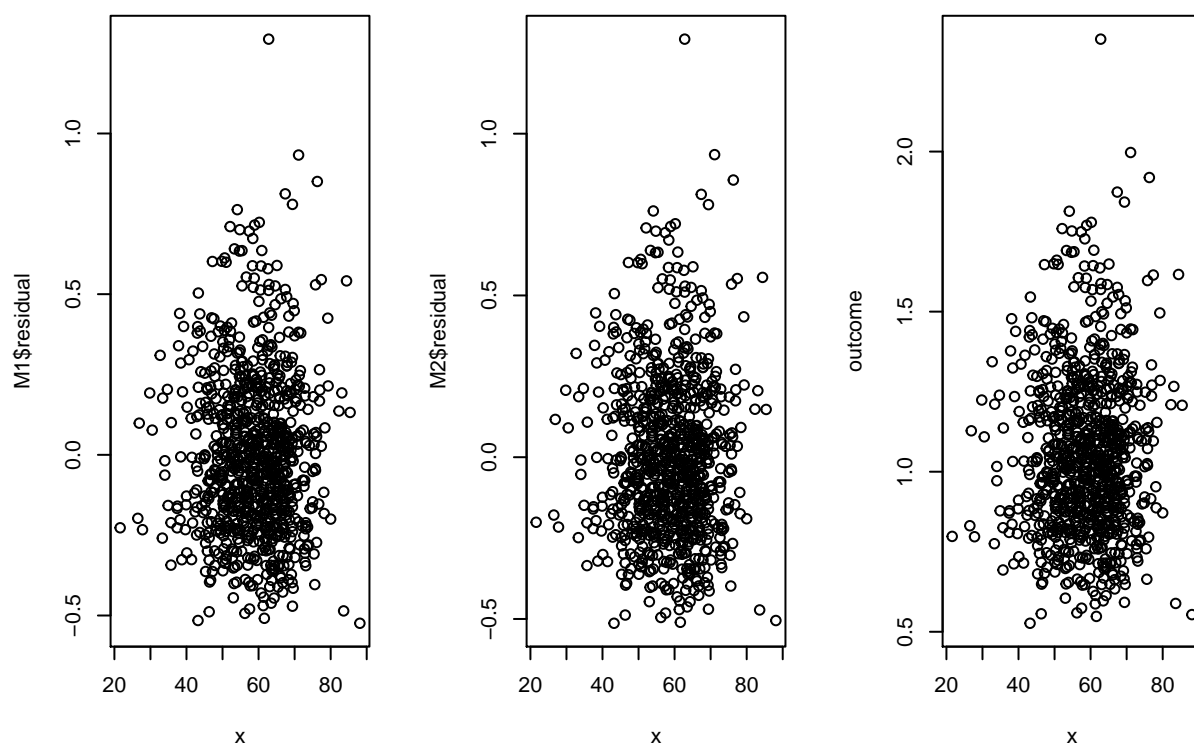
```
## [1] "residual for M1: 0.250373616826691"
## [1] "residual for M2: 0.250255208638358"
```



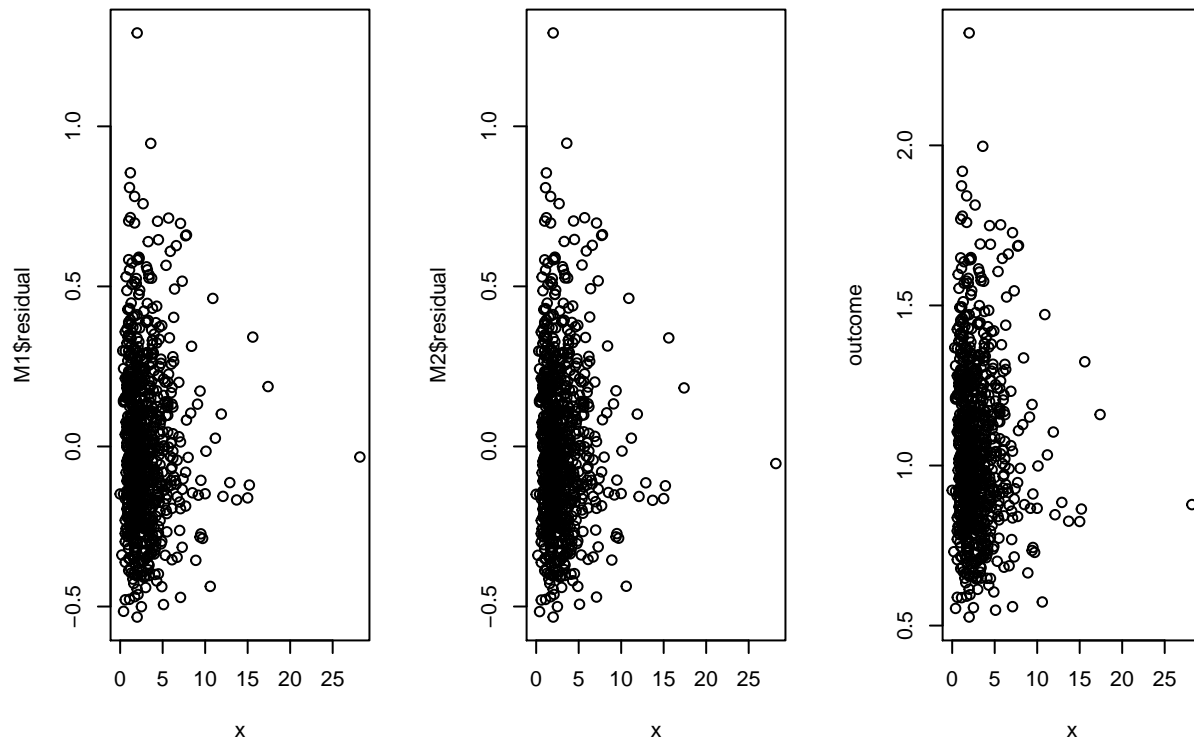
```
## [1] "residual for M1: 0.248704466454944"
## [1] "residual for M2: 0.248847192837983"
```



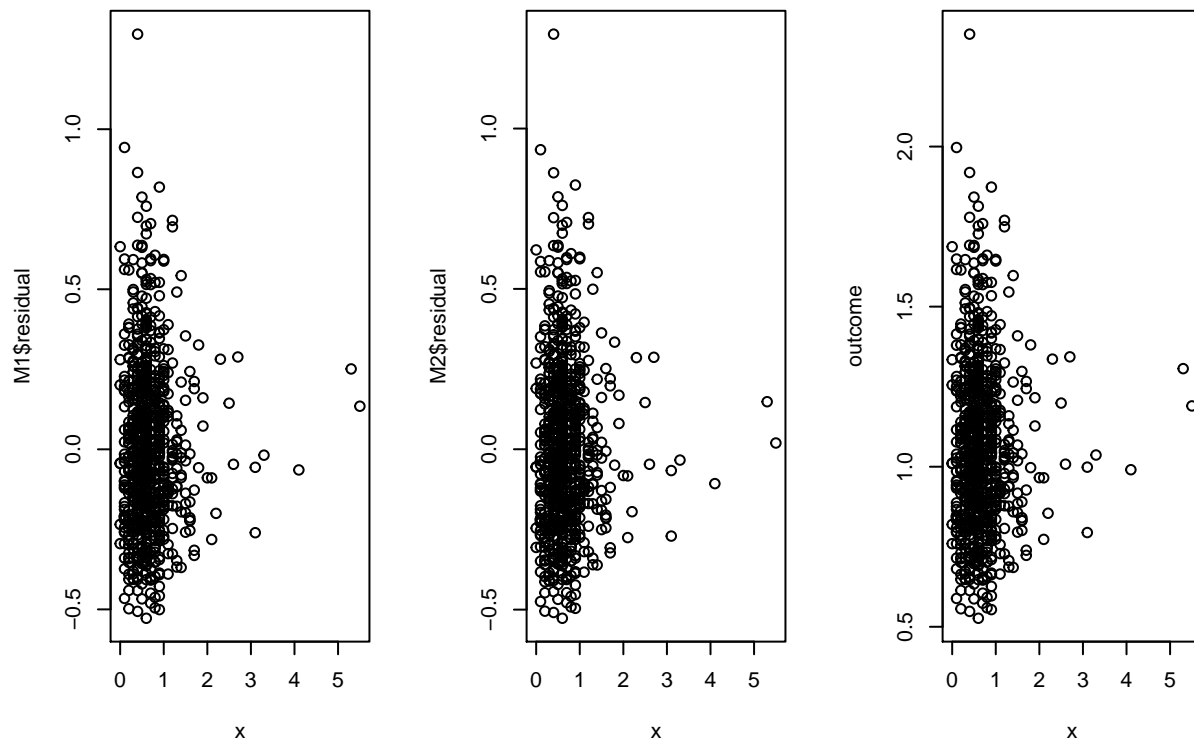
```
## [1] "residual for M1: 0.25026710930793"
## [1] "residual for M2: 0.250393729526099"
```



```
## [1] "residual for M1: 0.250043388210667"
## [1] "residual for M2: 0.25018695270193"
```



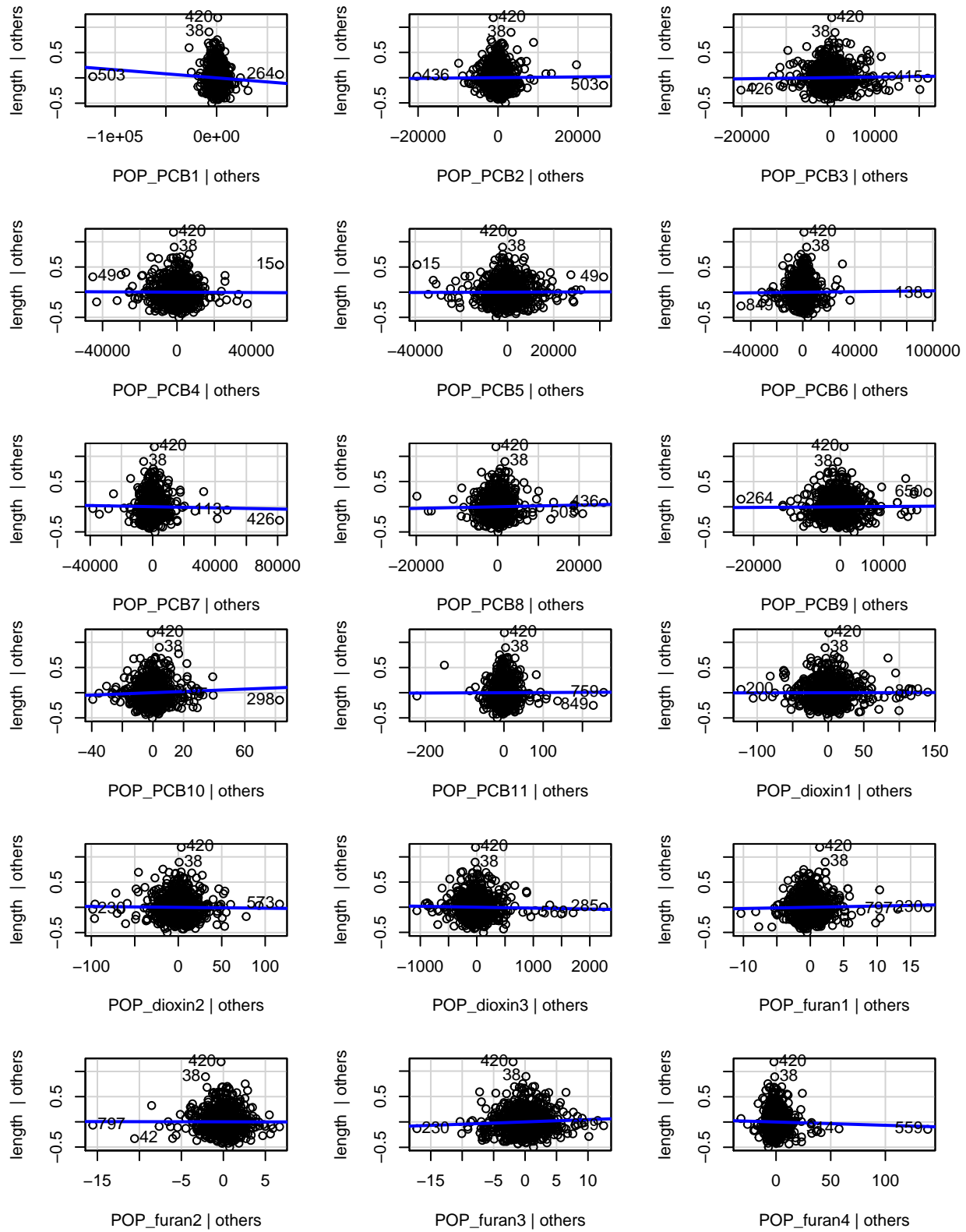
```
## [1] "residual for M1: 0.250382476371691"
## [1] "residual for M2: 0.25042580861039"
```

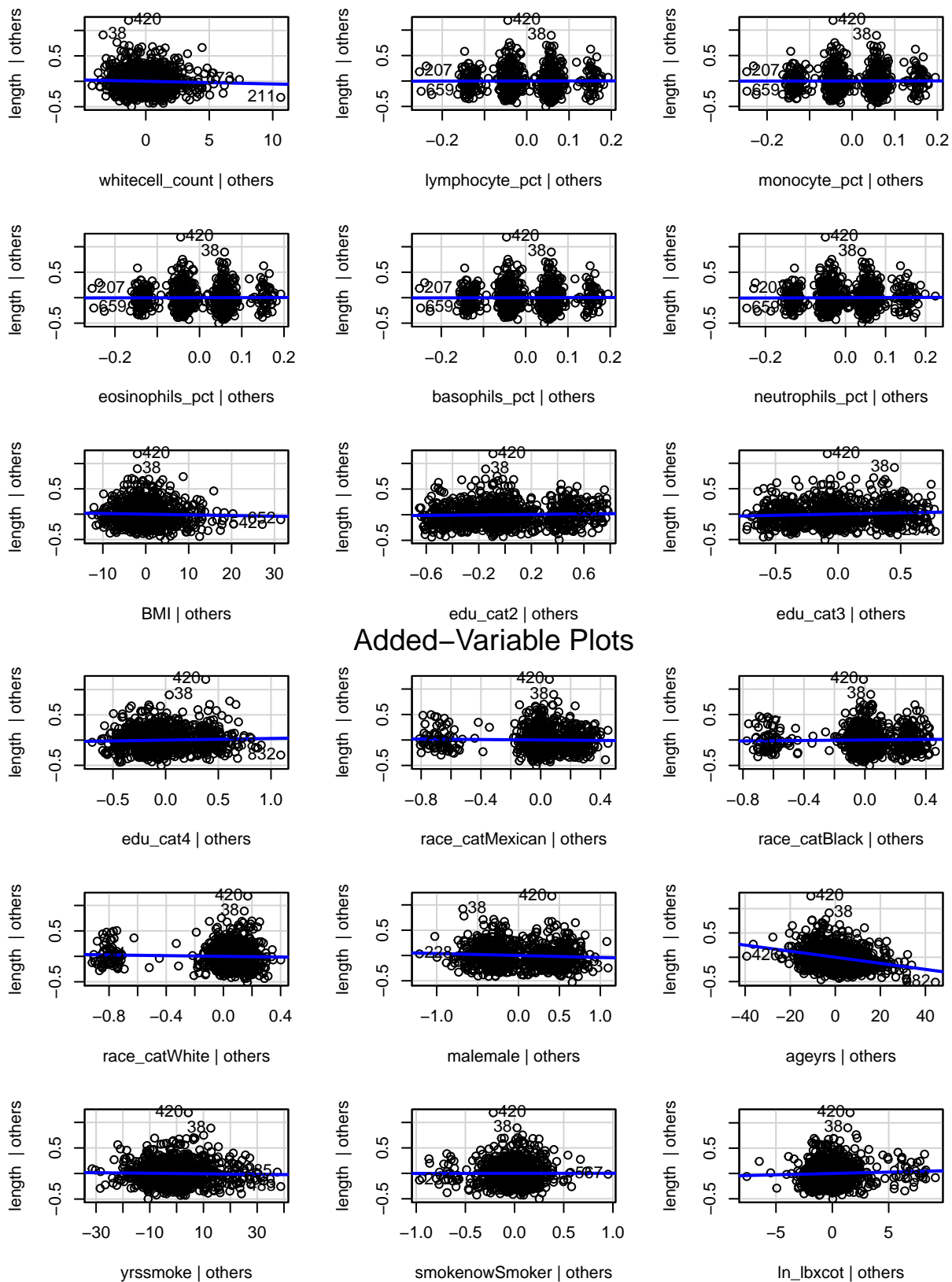


```
# Judy's work Part 2
# testing non-linearity in MLR
library(car)
```

```
## Loading required package: carData
```

```
M <- lm (length ~ ., data=pollutants)
avPlots(M)
```





Added-Variable Plots

```
# Estella's work 3
f <- as.formula(
```

```

paste("length", paste("(", paste(POP_PCB, collapse = "+"), ")^2"), sep="~"))

m <- lm(f, data = pollutants)
summary(m)

```

```

##
## Call:
## lm(formula = f, data = pollutants)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.53819 -0.16080 -0.01896  0.12149  1.20671
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)    1.153e+00  2.892e-02  39.876 < 2e-16 ***
## POP_PCB1      -6.741e-06  3.521e-06  -1.915  0.05591 .
## POP_PCB2       3.801e-06  9.328e-06   0.407  0.68378
## POP_PCB3       6.747e-06  6.701e-06   1.007  0.31431
## POP_PCB4       1.373e-06  3.278e-06   0.419  0.67539
## POP_PCB5       1.920e-06  3.267e-06   0.588  0.55680
## POP_PCB6      -3.673e-06  4.336e-06  -0.847  0.39729
## POP_PCB7      -5.281e-06  4.697e-06  -1.124  0.26126
## POP_PCB8      -1.073e-05  8.331e-06  -1.288  0.19796
## POP_PCB9      -1.833e-06  5.806e-06  -0.316  0.75232
## POP_PCB10     2.720e-03  2.088e-03   1.303  0.19311
## POP_PCB11     4.644e-04  9.916e-04   0.468  0.63969
## POP_PCB1:POP_PCB2  9.529e-11  2.113e-10   0.451  0.65216
## POP_PCB1:POP_PCB3 -6.580e-10  4.156e-10  -1.583  0.11377
## POP_PCB1:POP_PCB4  1.116e-10  1.917e-10   0.582  0.56080
## POP_PCB1:POP_PCB5 -1.621e-11  1.318e-10  -0.123  0.90218
## POP_PCB1:POP_PCB6  6.244e-11  2.176e-10   0.287  0.77423
## POP_PCB1:POP_PCB7  2.221e-11  2.742e-10   0.081  0.93548
## POP_PCB1:POP_PCB8 -5.209e-10  2.693e-10  -1.935  0.05340 .
## POP_PCB1:POP_PCB9  4.146e-10  2.287e-10   1.813  0.07020 .
## POP_PCB1:POP_PCB10 1.675e-07  1.311e-07   1.277  0.20183
## POP_PCB1:POP_PCB11 -6.663e-08  7.321e-08  -0.910  0.36303
## POP_PCB2:POP_PCB3  1.673e-09  8.717e-10   1.919  0.05537 .
## POP_PCB2:POP_PCB4 -6.761e-10  4.688e-10  -1.442  0.14963
## POP_PCB2:POP_PCB5  3.840e-10  3.632e-10   1.057  0.29069
## POP_PCB2:POP_PCB6 -1.426e-09  5.834e-10  -2.444  0.01474 *
## POP_PCB2:POP_PCB7  1.532e-09  6.770e-10   2.264  0.02387 *
## POP_PCB2:POP_PCB8  2.135e-09  8.207e-10   2.602  0.00945 **
## POP_PCB2:POP_PCB9 -1.356e-09  7.249e-10  -1.870  0.06183 .
## POP_PCB2:POP_PCB10 -1.232e-06  4.242e-07  -2.904  0.00378 **
## POP_PCB2:POP_PCB11 3.388e-07  2.013e-07   1.683  0.09270 .
## POP_PCB3:POP_PCB4 -3.996e-11  1.199e-10  -0.333  0.73900
## POP_PCB3:POP_PCB5  4.665e-11  2.413e-10   0.193  0.84674
## POP_PCB3:POP_PCB6 -3.741e-10  2.662e-10  -1.405  0.16029
## POP_PCB3:POP_PCB7  6.438e-10  2.896e-10   2.223  0.02649 *
## POP_PCB3:POP_PCB8  7.340e-10  8.821e-10   0.832  0.40563
## POP_PCB3:POP_PCB9 -4.221e-10  5.470e-10  -0.772  0.44059
## POP_PCB3:POP_PCB10 -4.835e-07  2.555e-07  -1.892  0.05885 .
## POP_PCB3:POP_PCB11 7.155e-08  7.874e-08   0.909  0.36382

```



```

## POP_PCB4:POP_PCB5      3.002e-12  6.669e-11   0.045  0.96410
## POP_PCB4:POP_PCB6      1.788e-10  1.543e-10   1.159  0.24694
## POP_PCB4:POP_PCB7     -2.117e-10  1.579e-10  -1.341  0.18019
## POP_PCB4:POP_PCB8     -4.525e-11  3.961e-10  -0.114  0.90908
## POP_PCB4:POP_PCB9      1.217e-10  2.625e-10   0.464  0.64294
## POP_PCB4:POP_PCB10     1.345e-07  8.933e-08   1.505  0.13265
## POP_PCB4:POP_PCB11     1.685e-08  5.047e-08   0.334  0.73861
## POP_PCB5:POP_PCB6      4.714e-11  1.390e-10   0.339  0.73458
## POP_PCB5:POP_PCB7     -1.555e-10  1.446e-10  -1.076  0.28244
## POP_PCB5:POP_PCB8     -4.639e-10  3.185e-10  -1.457  0.14562
## POP_PCB5:POP_PCB9     -1.626e-11  1.822e-10  -0.089  0.92890
## POP_PCB5:POP_PCB10     9.703e-08  9.241e-08   1.050  0.29406
## POP_PCB5:POP_PCB11    -5.549e-08  4.079e-08  -1.360  0.17407
## POP_PCB6:POP_PCB7     -2.248e-11  1.147e-10  -0.196  0.84474
## POP_PCB6:POP_PCB8      7.086e-10  3.808e-10   1.861  0.06310 .
## POP_PCB6:POP_PCB9      4.295e-10  3.267e-10   1.315  0.18895
## POP_PCB6:POP_PCB10     2.152e-07  1.182e-07   1.820  0.06909 .
## POP_PCB6:POP_PCB11    -4.299e-08  2.038e-08  -2.109  0.03523 *
## POP_PCB7:POP_PCB8     -1.029e-09  4.279e-10  -2.404  0.01645 *
## POP_PCB7:POP_PCB9     -2.467e-10  3.622e-10  -0.681  0.49603
## POP_PCB7:POP_PCB10    -3.893e-08  1.308e-07  -0.298  0.76608
## POP_PCB7:POP_PCB11     4.226e-08  3.690e-08   1.145  0.25246
## POP_PCB8:POP_PCB9      1.317e-10  5.297e-10   0.249  0.80373
## POP_PCB8:POP_PCB10     5.264e-07  3.029e-07   1.738  0.08265 .
## POP_PCB8:POP_PCB11    -5.764e-08  1.285e-07  -0.449  0.65382
## POP_PCB9:POP_PCB10    -2.240e-08  1.448e-07  -0.155  0.87712
## POP_PCB9:POP_PCB11     7.916e-08  6.811e-08   1.162  0.24548
## POP_PCB10:POP_PCB11  -5.384e-05  2.694e-05  -1.999  0.04599 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.2377 on 797 degrees of freedom
## Multiple R-squared:  0.1666, Adjusted R-squared:  0.09763
## F-statistic: 2.415 on 66 and 797 DF,  p-value: 1.316e-08

# Estella's work 4
# setting threshold of pvalue to be 0.05 and assess possible interaction terms
pvalues <- summary(m)$coefficients[,4]
p_threshold = 0.05
selected <-which(pvalues<= p_threshold)
names(selected)

## [1] "(Intercept)"      "POP_PCB2:POP_PCB6"  "POP_PCB2:POP_PCB7"
## [4] "POP_PCB2:POP_PCB8"  "POP_PCB2:POP_PCB10" "POP_PCB3:POP_PCB7"
## [7] "POP_PCB6:POP_PCB11" "POP_PCB7:POP_PCB8"  "POP_PCB10:POP_PCB11"

#stepwise parameters selection without any interaction terms
M0 <- lm(length ~ 1, data = train_data) # minimal model
Mfull <- lm(length ~ ., data= train_data)

## 2 corresponds to AIC
## log(n) corresponds to BIC

# stepwise AIC
Mstart <- lm(length ~ ., data= train_data)

```

```

system.time({
  MAIC <- step(object = Mstart,
               scope = list(lower = M0, upper = Mfull),
               direction = "both", trace = 0, k = 2)
})

##    user  system elapsed
##  0.812   0.085   0.904

#stepwiseBIC
system.time({
  MBIC <- step(object = Mstart,
               scope = list(lower = M0, upper = Mfull),
               direction = "both", trace = 0, k = log(nrow(train_data)))
})

##    user  system elapsed
##  0.826   0.078   0.912

#stepwiseB_Adjusted R2
MAIC

##
## Call:
## lm(formula = length ~ POP_PCB1 + POP_PCB10 + POP_furan1 + POP_furan2 +
##    whitecell_count + monocyte_pct + edu_cat + race_cat + male +
##    ageyrs + ln_lbxcot, data = train_data)
##
## Coefficients:
##      (Intercept)      POP_PCB1      POP_PCB10      POP_furan1
##      1.443e+00     -5.602e-07      1.780e-03     -6.532e-03
##      POP_furan2  whitecell_count  monocyte_pct      edu_cat2
##      8.968e-03     -1.029e-02     -6.643e-03      4.105e-02
##      edu_cat3      edu_cat4  race_catMexican  race_catBlack
##      6.188e-02      8.254e-02     -3.635e-03      3.584e-02
##      race_catWhite      malemale      ageyrs      ln_lbxcot
##      -4.701e-02     -4.513e-02     -5.820e-03      7.573e-03

MBIC

##
## Call:
## lm(formula = length ~ POP_furan3 + ageyrs, data = train_data)
##
## Coefficients:
## (Intercept)  POP_furan3      ageyrs
##    1.355743    0.005969   -0.006922

# stepwise parameters selection with any interaction terms
M0 <- lm(length ~ 1, data = train_data) # minimal model

# tail to remove length column
single <- paste(tail(colnames(train_data),-1), collapse = " + ")
# tail to remove intercept column
interaction <- paste(tail(names(selected),-1), collapse = " + ")
f_interaction <- as.formula(
  paste("length", paste("(", single,"+", interaction, ")"), sep = " ~"))

```

```

Mfull <- lm(f_interaction, data = train_data)
Mstart <- lm(f_interaction, data = train_data)

# stepwise AIC
Mstart <- lm(length ~ ., data= train_data)
system.time({
  MAIC_Interaction <- step(object = Mstart,
                           scope = list(lower = M0, upper = Mfull),
                           direction = "both", trace = 0, k = 2)
})

##      user  system elapsed
##    0.833    0.079    0.915

#stepwiseBIC
system.time({
  MBIC_Interaction <- step(object = Mstart,
                           scope = list(lower = M0, upper = Mfull),
                           direction = "both", trace = 0,
                           k = log(nrow(train_data)))
})

##      user  system elapsed
##    0.875    0.080    0.956

#stepwiseB_Adjusted R2
MAIC_Interaction

##
## Call:
## lm(formula = length ~ POP_PCB1 + POP_PCB6 + POP_PCB10 + POP_PCB11 +
##     POP_dioxin2 + POP_furan3 + whitecell_count + monocyte_pct +
##     BMI + edu_cat + race_cat + male + ageyrs + ln_lbxcot + POP_PCB10:POP_PCB11,
##     data = train_data)
##
## Coefficients:
##      (Intercept)      POP_PCB1      POP_PCB6
##      1.473e+00     -8.511e-07     1.150e-06
##      POP_PCB10      POP_PCB11      POP_dioxin2
##      2.839e-03      9.157e-04     -6.180e-04
##      POP_furan3      whitecell_count      monocyte_pct
##      4.745e-03     -9.472e-03     -6.707e-03
##      BMI      edu_cat2      edu_cat3
##     -2.272e-03      4.205e-02      5.902e-02
##      edu_cat4      race_catMexican      race_catBlack
##      7.656e-02      1.408e-03      4.927e-02
##      race_catWhite      malemale      ageyrs
##     -3.842e-02     -3.208e-02     -6.126e-03
##      ln_lbxcot      POP_PCB10:POP_PCB11
##      7.374e-03     -2.457e-05

MBIC_Interaction

##
## Call:
## lm(formula = length ~ POP_furan3 + ageyrs, data = train_data)

```

```
##
## Coefficients:
## (Intercept)    POP_furan3      ageyrs
##      1.355743      0.005969     -0.006922

# man's work
predAIC <- predict(MAIC, newdata=test_data)
predBIC <- predict(MBIC, newdata=test_data)
predAICInteraction <- predict(MAIC_Interaction, newdata=test_data)
predBICInteraction <- predict(MBIC_Interaction, newdata=test_data)

mean((test_data$length - predAIC)^2)

## [1] 0.05336494

mean((test_data$length - predBIC)^2)

## [1] 0.04804827

mean((test_data$length - predAICInteraction)^2)

## [1] 0.05230268

mean((test_data$length - predBICInteraction)^2)

## [1] 0.04804827
```

5. Results:

Report on the findings of your analysis

6. Discussion:

Comment on your findings/conclusions; describe any limitations of your analysis.

7. Appendix

Data Summary

Looking at the useful metrics for the data

```
summary(pollutants)
```

##	length	POP_PCB1	POP_PCB2	POP_PCB3
##	Min. :0.5266	Min. : 2000	Min. : 2000	Min. : 2000
##	1st Qu.:0.8754	1st Qu.: 9975	1st Qu.: 4800	1st Qu.: 3700
##	Median :1.0286	Median : 27600	Median : 11500	Median : 6200
##	Mean :1.0543	Mean : 38082	Mean : 15637	Mean : 10158
##	3rd Qu.:1.2095	3rd Qu.: 53325	3rd Qu.: 21825	3rd Qu.: 12000
##	Max. :2.3512	Max. :572000	Max. :165000	Max. :123000
##	POP_PCB4	POP_PCB5	POP_PCB6	POP_PCB7
##	Min. : 2100	Min. : 2100	Min. : 2000	Min. : 1100
##	1st Qu.: 11475	1st Qu.: 15600	1st Qu.: 4400	1st Qu.: 4000
##	Median : 25550	Median : 36300	Median : 9400	Median : 7450
##	Mean : 38456	Mean : 52650	Mean : 16820	Mean : 12682
##	3rd Qu.: 50650	3rd Qu.: 68625	3rd Qu.: 19500	3rd Qu.: 15625

```

## Max. :487000 Max. :708000 Max. :319000 Max. :144000
## POP_PCB8 POP_PCB9 POP_PCB10 POP_PCB11
## Min. : 1100 Min. : 1100 Min. : 1.70 Min. : 1.30
## 1st Qu.: 3800 1st Qu.: 3900 1st Qu.: 9.10 1st Qu.: 14.80
## Median : 6950 Median : 8050 Median : 18.35 Median : 24.50
## Mean : 10530 Mean : 12220 Mean : 24.49 Mean : 38.15
## 3rd Qu.: 14425 3rd Qu.: 16025 3rd Qu.: 34.90 3rd Qu.: 42.95
## Max. :187000 Max. :144000 Max. :172.00 Max. :845.00
## POP_dioxin1 POP_dioxin2 POP_dioxin3 POP_furan1
## Min. : 1.90 Min. : 1.40 Min. : 36.8 Min. : 1.000
## 1st Qu.: 23.90 1st Qu.: 21.27 1st Qu.: 197.0 1st Qu.: 3.200
## Median : 41.35 Median : 37.80 Median : 342.5 Median : 5.200
## Mean : 57.65 Mean : 47.81 Mean : 494.4 Mean : 6.371
## 3rd Qu.: 71.62 3rd Qu.: 62.42 3rd Qu.: 603.0 3rd Qu.: 7.700
## Max. :760.00 Max. :281.00 Max. :8190.0 Max. :44.400
## POP_furan2 POP_furan3 POP_furan4 whitecell_count
## Min. : 0.800 Min. : 0.700 Min. : 0.90 Min. : 2.300
## 1st Qu.: 2.600 1st Qu.: 2.200 1st Qu.: 6.40 1st Qu.: 5.600
## Median : 4.200 Median : 5.050 Median : 9.65 Median : 6.900
## Mean : 5.390 Mean : 6.669 Mean : 11.54 Mean : 7.191
## 3rd Qu.: 6.825 3rd Qu.: 9.300 3rd Qu.: 14.00 3rd Qu.: 8.300
## Max. :33.500 Max. :38.300 Max. :234.00 Max. :20.100
## lymphocyte_pct monocyte_pct eosinophils_pct basophils_pct
## Min. : 5.80 Min. : 1.600 Min. :21.60 Min. : 0.000
## 1st Qu.:24.00 1st Qu.: 6.600 1st Qu.:52.35 1st Qu.: 1.500
## Median :28.95 Median : 7.700 Median :59.30 Median : 2.300
## Mean :29.92 Mean : 7.936 Mean :58.62 Mean : 2.903
## 3rd Qu.:35.42 3rd Qu.: 9.100 3rd Qu.:65.22 3rd Qu.: 3.700
## Max. :73.40 Max. :23.800 Max. :88.10 Max. :28.200
## neutrophils_pct BMI edu_cat race_cat male
## Min. :0.0000 Min. :16.16 1:270 Other : 71 female:490
## 1st Qu.:0.4000 1st Qu.:23.88 2:199 Mexican:191 male :374
## Median :0.6000 Median :27.38 3:228 Black :154
## Mean :0.6669 Mean :28.09 4:167 White :448
## 3rd Qu.:0.8000 3rd Qu.:31.17
## Max. :5.5000 Max. :62.99
## ageyrs yrssmoke smokenow ln_lbxcot
## Min. :20.00 Min. : 0.0 Non-Smoker:664 Min. : -4.5099
## 1st Qu.:34.00 1st Qu.: 0.0 Smoker :200 1st Qu.: -4.0745
## Median :46.00 Median : 0.0 Median : -2.7334
## Mean :48.36 Mean :10.6 Mean : -0.9804
## 3rd Qu.:63.00 3rd Qu.:20.0 3rd Qu.: 2.8000
## Max. :85.00 Max. :69.0 Max. : 6.5848

```

Outlier Entries

Here we will find entries where outliers for different covariate occurred.

```

pollutant_mat = data.matrix(pollutants, rownames.force = NA)

max_PCB_idx = c()
for (c in 2:12) {
  max_PCB_idx[c-1] = which.max(pollutant_mat[, c])
}

```

```

max_PCB_idx

## [1] 436 436 436 436 436 436 426 436 436 298 272

max_dioxin_idx = c()
for (c in 13:15) {
  max_dioxin_idx[c-12] = which.max(pollutant_mat[, c])
}
max_dioxin_idx

## [1] 285 573 285

max_furan_idx = c()
for (c in 16:19) {
  max_furan_idx[c-15] = which.max(pollutant_mat[, c])
}
max_furan_idx

## [1] 230 559 590 559

max_WBC_idx = c()
for (c in 20:25) {
  max_WBC_idx[c-19] = which.max(pollutant_mat[, c])
}
max_WBC_idx

## [1] 211 766 440 782 739 415

```