

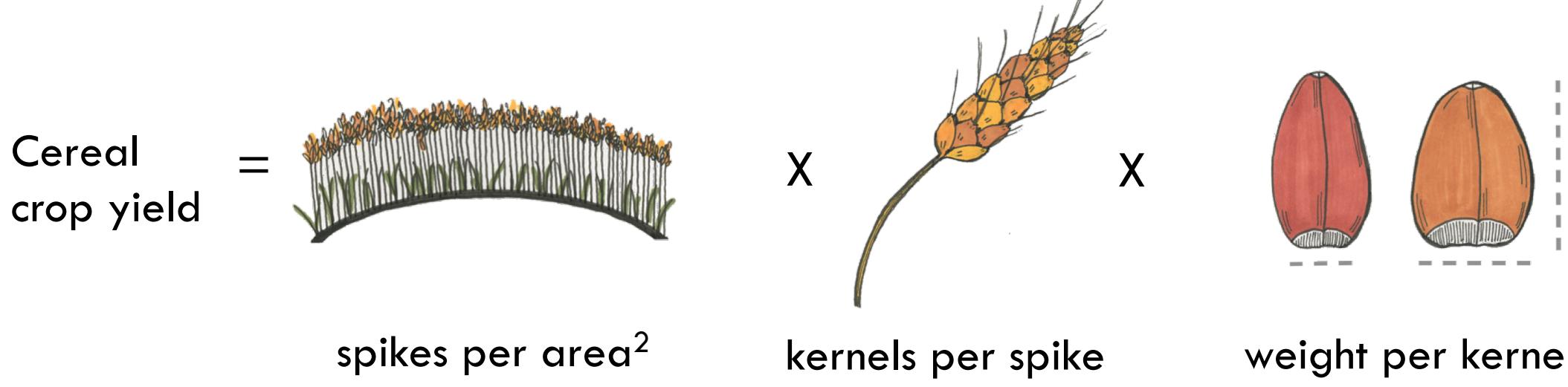
Counting on Crossovers: Fine-Mapping a Kernel Weight & Morphology Gene in Wheat

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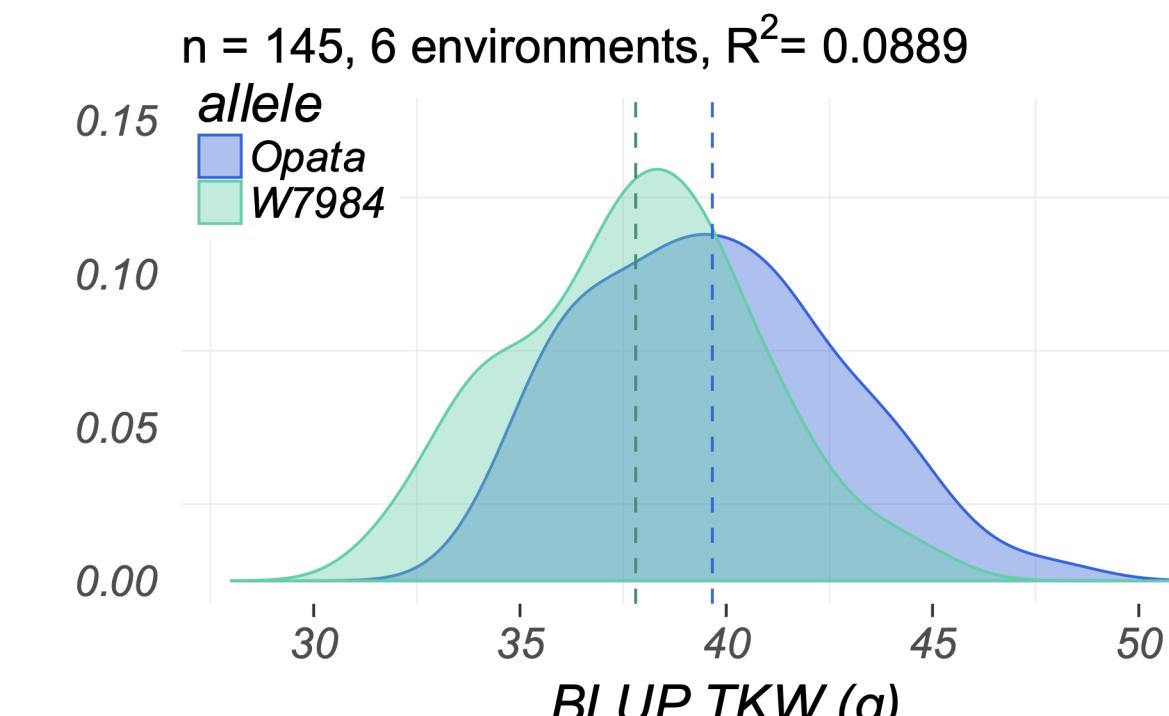
Kernel weight and width are both significantly associated with two nearby regions on chromosome 5A. The candidate gene(s) expression occurs prior to 10 DPA.

1 Yield is a balancing act



Recent advancement of the wheat reference genome assembly and genome editing tools can help facilitate the characterization of genes underlying quantitative trait loci (QTL) for yield components. Kernel weight and morphology are valuable traits to consider when releasing a new wheat variety because they can impact the number of kernels it takes to fill a bushel and milling quality.

5A QTL BLUP TKW SynOp DH distributions by allele type

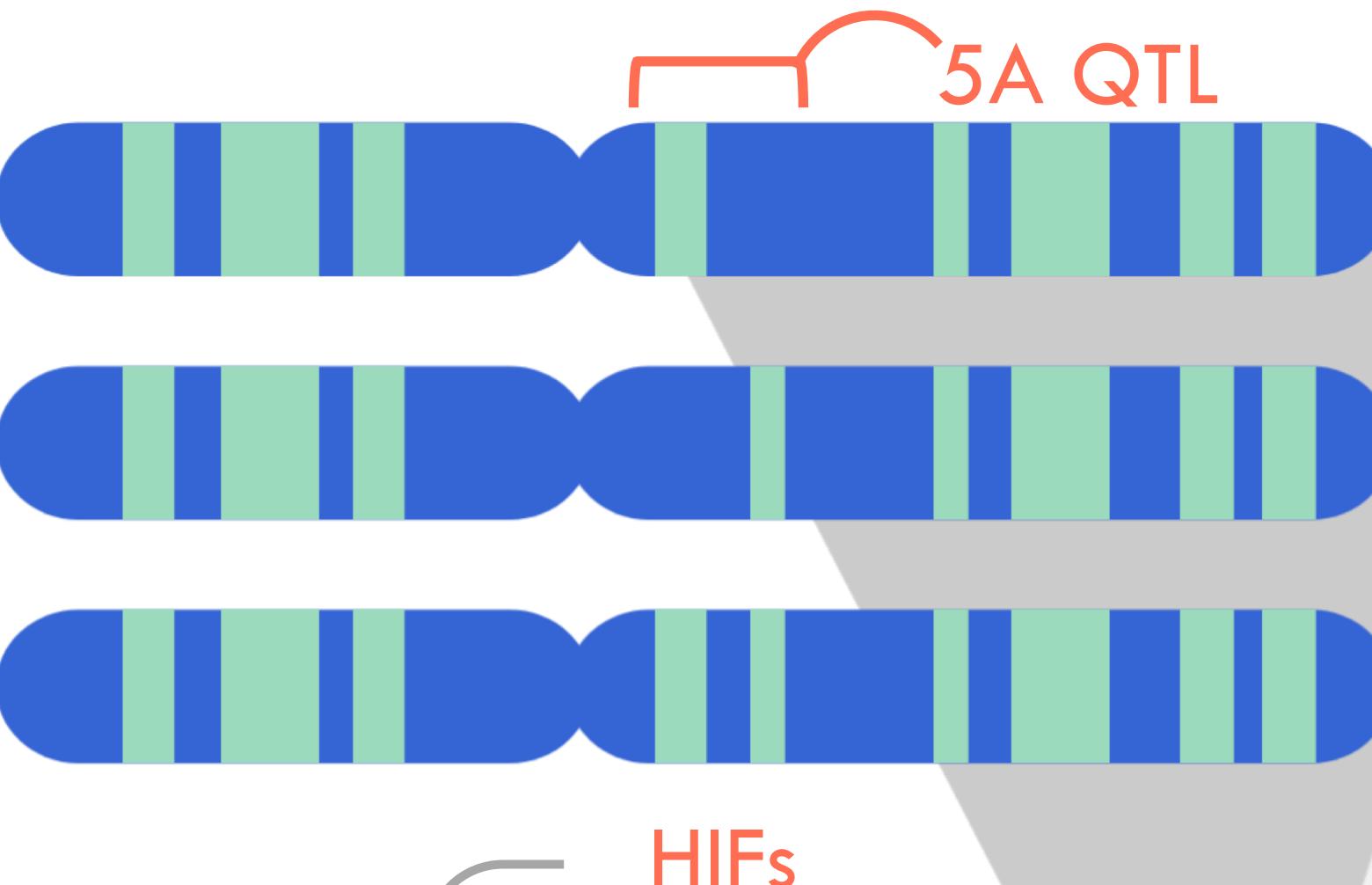


SynOp DH with Opata allele 5A QTL 4.9% heavier on average compared to W7984 allele

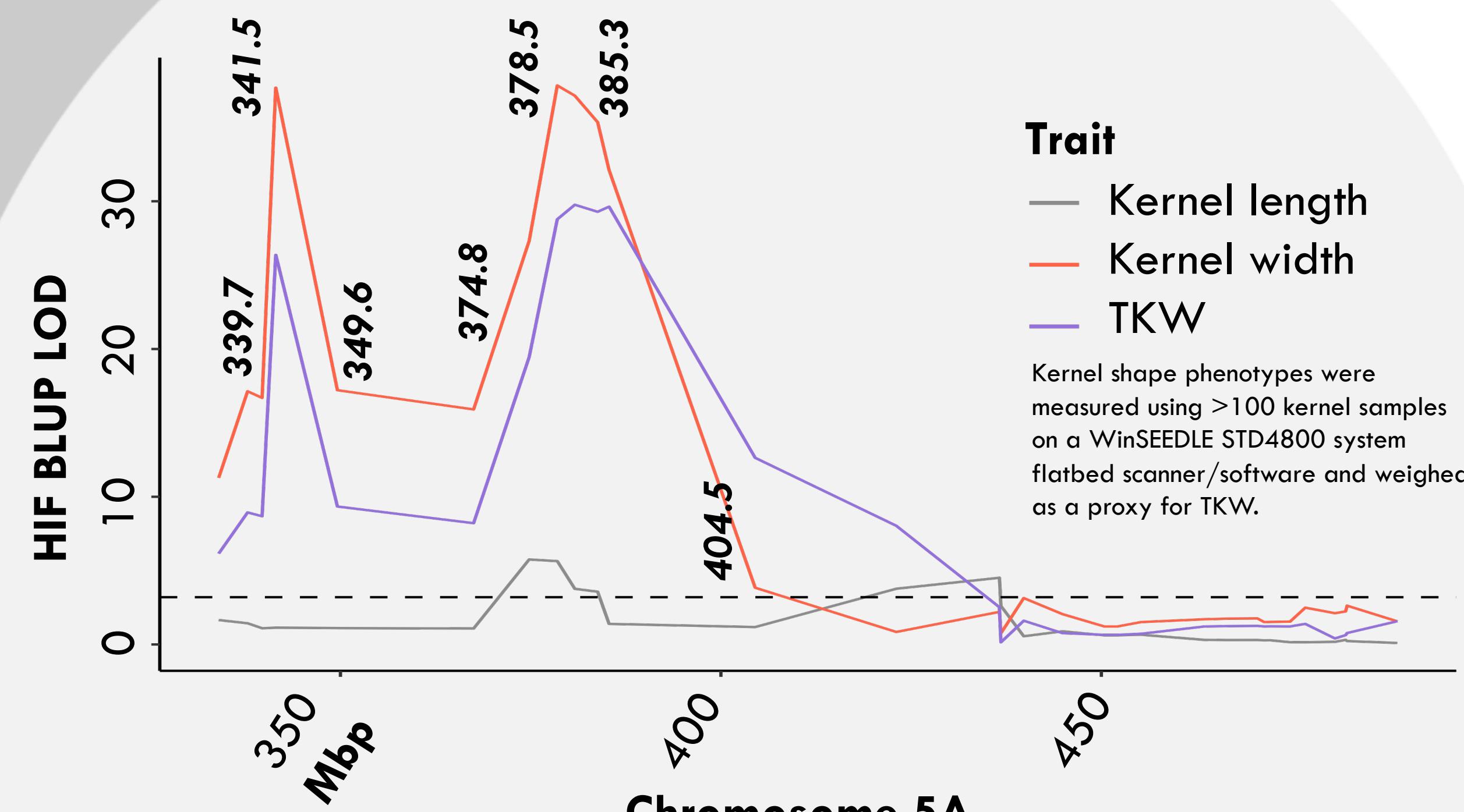
A QTL for thousand kernel weight (TKW) and kernel width was mapped using R/qtl (Broman et al., 2003) to a 100 Mbp region on chromosome 5AL in the W7984 Synthetic x Opata M85 spring bread wheat doubled haploid population (SynOp DH, 145-line subset of 215, Sorrells et al., 2011). Mixed models were fitted to extract BLUPs across 6 site-year combinations.

2 HIFs enhance genetic resolution of QTL

Fine-mapping a causal genetic variant is limited by crossovers that disrupt linked markers. We developed three heterogeneous inbred families (HIFs) F_{6:6} to increase genetic resolution of the 5A QTL. HIFs were selected from two SynOp recombinant inbred lines (SynOp RIL 2,039 F₆ lines, Sorrells et al., 2011) based on heterozygosity across the 5A QTL. The individual progeny were phenotyped and genotyped in order to track recombination events across the 5A QTL. The resulting HIFs have highly homogenous background genomes and distinct crossovers across the QTL that can be confidently associated with the line's phenotype (Tunstra et al., 1997).



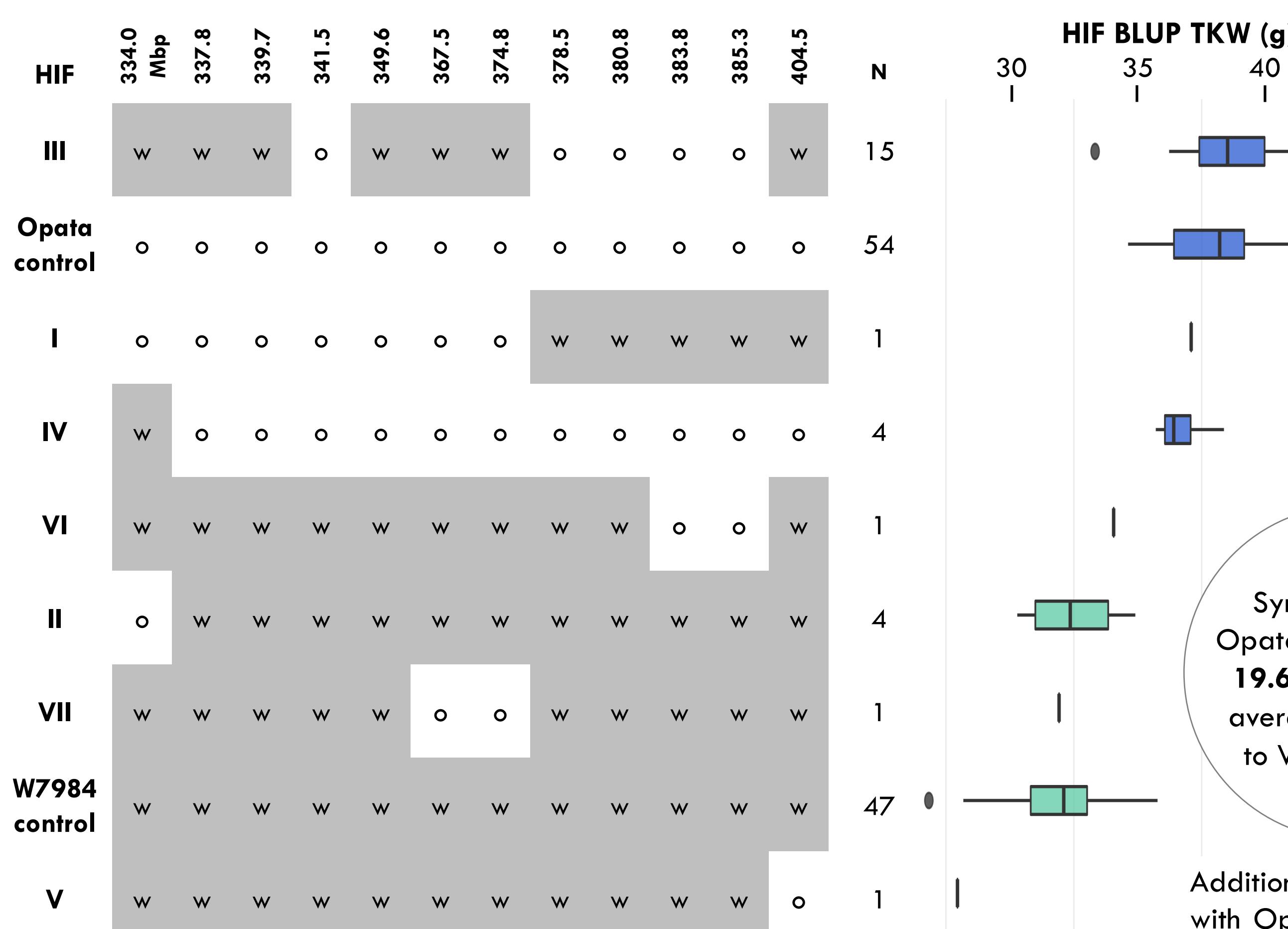
> 7,000 individual plants genotyped and phenotyped across six generations in the field and greenhouse were narrowed down to 128 lines with crossovers that limit the QTL region of significance. Mixed models were fitted to extract BLUPs across 2019 replicated headrows.



3 Comparing HIF crossovers further explains two QTL peaks

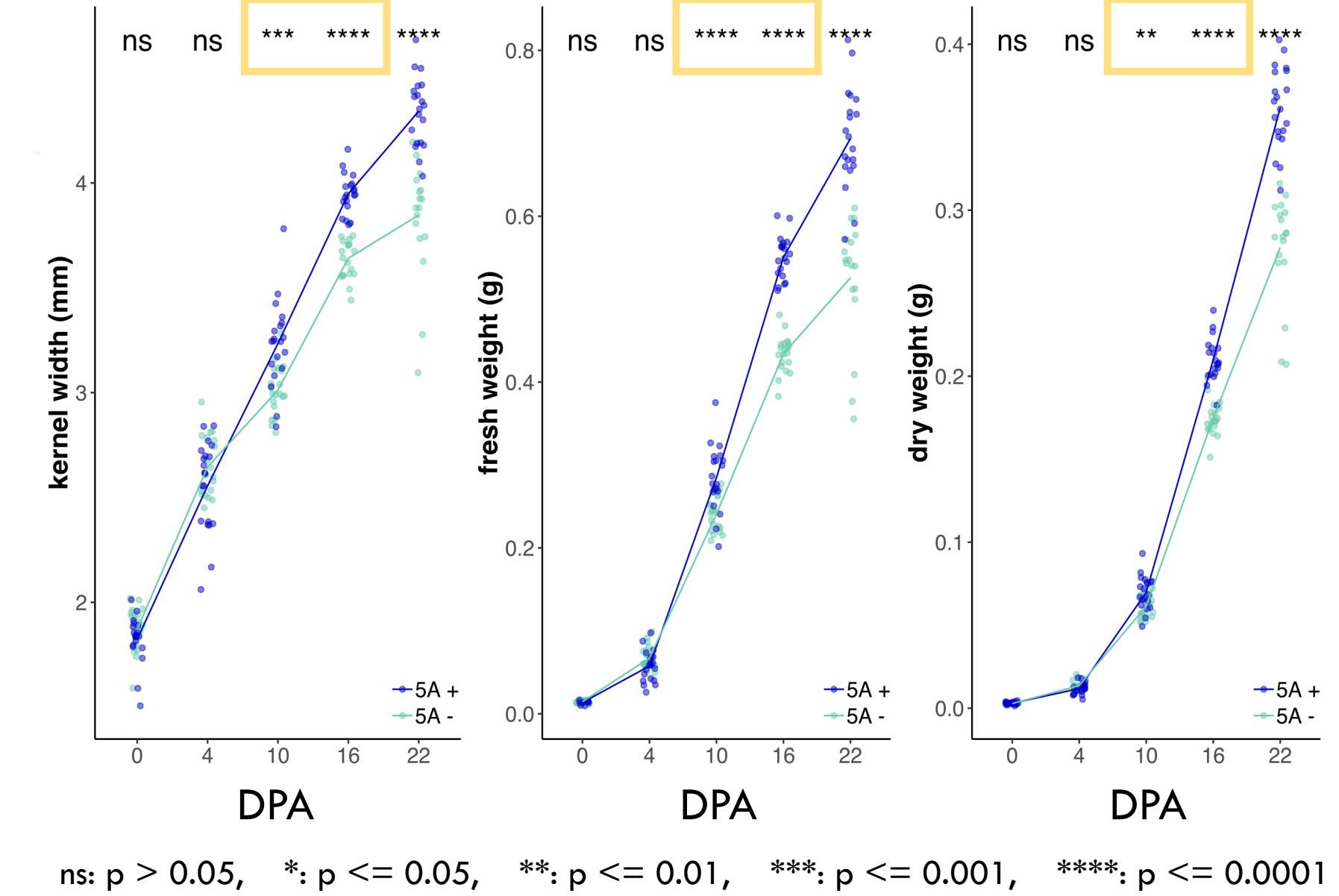
2019 HIFs grown in headrows and genotyped with KASP markers across the 5A QTL were analyzed with R/qtl, narrowing the significant sequence to a 9.9 and 10.5 Mbp region.

Tests for epistasis between the most significant markers were inconclusive because a simple linear interaction model cannot estimate an interaction from the present genotype frequencies (341.5 : 380.8 Mbp, 73 o:o, 54 w:w, 1 o:w, 0 w:o). The two QTL peaks could be due to multiple causal variants, or linkage. HIF I is the only line with a recombination event resulting in o:w genotype for the most significant markers and will be very useful for comparison and understanding the two-peak phenomenon in a gene expression study.



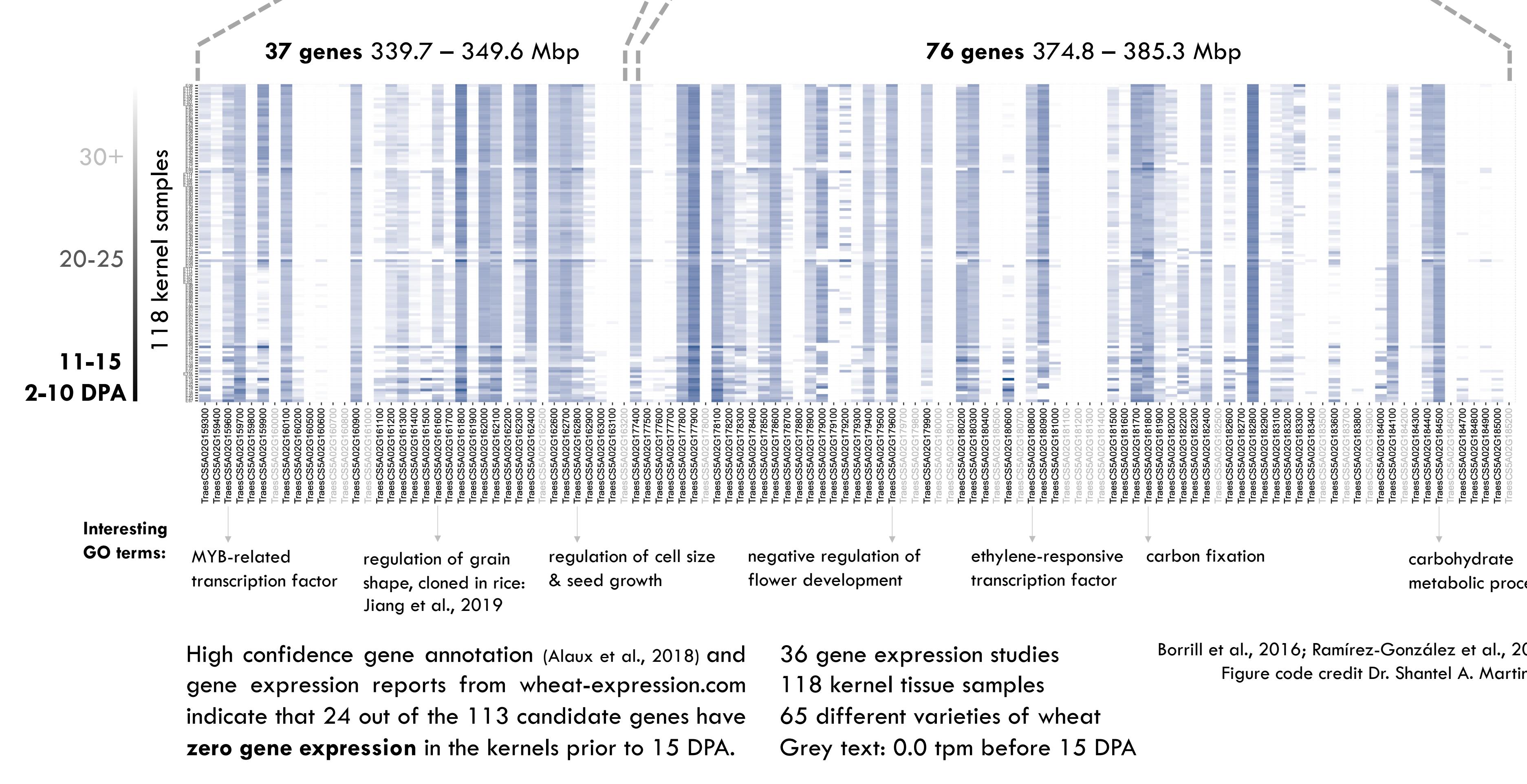
Additionally, SynOp HIF with Opata or W7984 allele 5A QTL have no significant difference in grain fill duration or spikelets / spike.

4 Kernel development time sequence analysis



Kernel morphology for 10 HIF genotypes with positive or negative 5A QTL causal variant alleles were tracked during the 2019 field season (Days post anthesis, DPA). Kernel width, fresh weight and dry weight were measured with 2 replicates / genotype, 10 spikes / time point and 10 kernels / spike. Significant difference in kernel morphology is measurable 10-16 DPA for 5A QTL HIFs grown in Ithaca, NY. The difference in phenotypes for the positive and negative causal variant genotypes is likely due to a gene expression event prior to 10 DPA, providing us with a high confidence window for timing RNA extraction.

5 Previous gene expression reports for 5A QTL peak regions



6 Next steps: 3'RNA-seq & genome editing

Relying on large populations over many generations to detect crossovers and capture finer resolution of the QTL is resource limiting. Our attention has now turned to gene expression and genome editing tools to facilitate characterization of the underlying causal grain weight and morphology gene.

3'RNA-seq experimental design

HIF grain tissue sample 4 & 8 DPA	334.0 Mbp	337.8	339.7	341.5	349.6	367.5	374.8	380.8	383.8	385.3	404.5
7-956-2-19-1-31-1	o	o	o	o	o	o	o	o	o	o	o
7-956-2-19-1-44	w	w	w	w	w	w	w	w	w	w	w
7-956-2-19-1-31-5	o	o	o	o	o	o	w	w	w	w	w
7-956-2-12-1-69-07	w	w	w	w	w	w	w	w	w	w	w

SynOp HIF tissue culture protocols

- Wan & Layton, 2006
- 1.5-2 mm embryos on CM4C callus induction medium, 2 months (98%)
- Callus transferred to MM50.2C / MM50C regeneration medium until shoots are >3"
- Regenerated shoots transferred to rooting medium (tba)

References

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