

Assignment 3: Data Exploration

Emma Brentjens

OVERVIEW

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

Directions

1. Rename this file `<FirstLast>_A03_DataExploration.Rmd` (replacing `<FirstLast>` with your first and last name).
2. Change “Student Name” on line 3 (above) with your name.
3. Work through the steps, **creating code and output** that fulfill each instruction.
4. Be sure to **answer the questions** in this assignment document.
5. When you have completed the assignment, **Knit** the text and code into a single PDF file.
6. After Knitting, submit the completed exercise (PDF file) to the dropbox in Sakai.

The completed exercise is due on Sept 30th.

Set up your R session

1. Check your working directory, load necessary packages (tidyverse), and upload two datasets: the ECOTOX neonicotinoid dataset (ECOTOX_Neonicotinoids_Insects_raw.csv) and the Niwot Ridge NEON dataset for litter and woody debris (NEON_NIWO_Litter_massdata_2018-08_raw.csv). Name these datasets “Neonics” and “Litter”, respectively. Be sure to include the subcommand to read strings in as factors.

```
#1.
##checking working directory
getwd()

## [1] "/home/guest/R/EDA-Fall2022/Assignments"

##setting working directory to raw data folder (???????)
#setwd("/home/guest/R/EDA-Fall2022/Data/Raw")

##loading tidyverse package
library(tidyverse)
library(ggplot2)

##uploading and naming datasets
Neonics <- read.csv("Copyof_ECOTOX_Neonicotinoids_Insects_raw.csv", stringsAsFactors = T)
Litter <- read.csv("Copyof_NEON_NIWO_Litter_massdata_2018-08_raw.csv", stringsAsFactors = T)
#####this seems to only work when running in console when I tried to set wd to raw data folder
```

Learn about your system

2. The neonicotinoid dataset was collected from the Environmental Protection Agency’s ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used

widely in agriculture. The dataset that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicology of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer: This research can provide information on the efficacy of neonicotinoids to protect crops from insect pests. It is important to understand which doses are effective as using too little of the insecticide would not produce the desired effect but using too much may be harmful to nontarget species, like bees (Texas A&M AgriLife Extension, n.d.). Citation: Texas A&M AgriLife Extension. (n.d.). What is a neonicotinoid? Insects in the City. Retrieved September 26, 2022, from <https://citybugs.tamu.edu/factsheets/ipm/what-is-a-neonicotinoid/>

3. The Niwot Ridge litter and woody debris dataset was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: The amount of leaf litter and debris on the ground can influence soil moisture and nutrient cycling. As leaf litter decomposes, it is broken down into compounds that plants can absorb as they grow (Giweta, 2020). Leaf litter types can also provide information about plant composition in ecosystems (Giweta, 2020). Citation: Giweta, M. (2020). Role of litter production and its decomposition, and factors affecting the processes in a tropical forest ecosystem: A review. *Journal of Ecology and Environment*, 44(1), 11. <https://doi.org/10.1186/s41610-020-0151-2>

4. How is litter and woody debris sampled as part of the NEON network? Read the `NEON_Litterfall_UserGuide.pdf` document to learn more. List three pieces of salient information about the sampling methods here:

Answer: 1. 2. 3.

Obtain basic summaries of your data (Neonics)

5. What are the dimensions of the dataset?

```
##obtaining summary statistics
summary(Neonics)
```

```
##      CAS.Number
## Min.      : 58842209
## 1st Qu.:138261413
## Median :138261413
## Mean    :147651982
## 3rd Qu.:153719234
## Max.    :210880925
##
##
##                                     Chemical.Name
## (2E)-1-[(6-Chloro-3-pyridinyl)methyl]-N-nitro-2-imidazolidinimine :2658
## 3-[(2-Chloro-5-thiazolyl)methyl]tetrahydro-5-methyl-N-nitro-4H-1,3,5-oxadiazin-4-imine: 686
## [C(E)]-N-[(2-Chloro-5-thiazolyl)methyl]-N'-methyl-N''-nitroguanidine      : 452
## (1E)-N-[(6-Chloro-3-pyridinyl)methyl]-N'-cyano-N-methylethanimidamide      : 420
## N''-Methyl-N-nitro-N'-[(tetrahydro-3-furanyl)methyl]guanidine             : 218
## [N(Z)]-N-[3-[(6-Chloro-3-pyridinyl)methyl]-2-thiazolidinylidene]cyanamide  : 128
## (Other)                                                                    : 61
##
##                                     Chemical.Grade
## Not reported                                                                :3989
## Technical grade, technical product, technical formulation: 422
## Pestanal grade                                                             : 93
```

```

## Not coded : 53
## Commercial grade : 27
## Analytical grade : 15
## (Other) : 24
## Chemical.Analysis.Method
## Measured : 230
## Not coded : 51
## Not reported : 5
## Unmeasured :4321
## Unmeasured values (some measured values reported in article): 16
##
##
## Chemical.Purity Species.Scientific.Name
## NR :2502 Apis mellifera : 667
## 25 : 244 Bombus terrestris : 183
## 50 : 200 Apis mellifera ssp. carnica : 152
## 20 : 189 Bombus impatiens : 140
## 70 : 112 Apis mellifera ssp. ligustica: 113
## 75 : 89 Popillia japonica : 94
## (Other):1287 (Other) :3274
## Species.Common.Name
## Honey Bee : 667
## Parasitic Wasp : 285
## Buff Tailed Bumblebee: 183
## Carniolan Honey Bee : 152
## Bumble Bee : 140
## Italian Honeybee : 113
## (Other) :3083
## Species.Group
## Insects/Spiders :3569
## Insects/Spiders; Standard Test Species : 27
## Insects/Spiders; Standard Test Species; U.S. Invasive Species: 667
## Insects/Spiders; U.S. Invasive Species : 360
##
##
## Organism.Lifestage Organism.Age Organism.Age.Units
## Not reported:2271 NR :3851 Not reported :3515
## Adult :1222 2 : 111 Day(s) : 327
## Larva : 437 3 : 105 Instar : 255
## Multiple : 285 <24 : 81 Hour(s) : 241
## Egg : 128 4 : 81 Hours post-emergence: 99
## Pupa : 69 1 : 59 Year(s) : 64
## (Other) : 211 (Other): 335 (Other) : 122
## Exposure.Type Media.Type
## Environmental, unspecified:1599 No substrate:2934
## Food :1124 Not reported: 663
## Spray : 393 Natural soil: 393
## Topical, general : 254 Litter : 264
## Ground granular : 249 Filter paper: 230
## Hand spray : 210 Not coded : 51
## (Other) : 794 (Other) : 88
## Test.Location Number.of.Doses Conc.1.Type..Author.
## Field artificial : 96 2 :2441 Active ingredient:3161

```

```

## Field natural      :1663  3      : 499  Formulation      :1420
## Field undeterminable:  4  5      : 314  Not coded        :  42
## Lab                :2860  6      : 230
##                   4      : 221
##                   NR      : 217
##                   (Other): 701
## Conc.1..Author. Conc.1.Units..Author.          Effect
## 0.37/ : 208  AI kg/ha : 575  Population      :1803
## 10/   : 127  AI mg/L  : 298  Mortality       :1493
## NR/   : 108  AI lb/acre: 277  Behavior        : 360
## NR    :  94  AI g/ha  : 241  Feeding behavior: 255
## 1     :  82  ng/org   : 231  Reproduction    : 197
## 1023  :  80  ppm      : 180  Development     : 136
## (Other):3924 (Other)  :2821  (Other)         : 379
## Effect.Measurement Endpoint Response.Site
## Abundance      :1699  NOEL :1816  Not reported    :4349
## Mortality      :1294  LOEL :1664  Midgut or midgut gland: 63
## Survival       : 133  LC50 : 327  Not coded       : 51
## Progeny counts/numbers: 120 LD50 : 274  Whole organism  : 41
## Food consumption : 103  NR   : 167  Hypopharyngeal gland : 27
## Emergence      :  98  NR-LETH: 86  Head           : 23
## (Other)        :1176  (Other): 289 (Other)        : 69
## Observed.Duration..Days. Observed.Duration.Units..Days.
## 1 : 713  Day(s) :4394
## 2 : 383  Emergence : 70
## NR : 355  Growing season : 48
## 7 : 207  Day(s) post-hatch : 20
## 3 : 183  Day(s) post-emergence: 17
## 0.0417 : 133  Tiller stage : 15
## (Other):2649 (Other) : 59
##
## Author
## Peck,D.C. : 208
## Frank,S.D. : 100
## El Hassani,A.K., M. Dacher, V. Gary, M. Lambin, M. Gauthier, and C. Armengaud: 96
## Williamson,S.M., S.J. Willis, and G.A. Wright : 93
## Laurino,D., A. Manino, A. Patetta, and M. Porporato : 88
## Scholer,J., and V. Krischik : 82
## (Other) :3956
## Reference.Number
## Min. : 344
## 1st Qu.:108459
## Median :165559
## Mean :142189
## 3rd Qu.:168998
## Max. :180410
##
##
## Long-Term Effects of Imidacloprid on the Abundance of Surface- and Soil-Active Nontarget Fauna in T
## Reduced Risk Insecticides to Control Scale Insects and Protect Natural Enemies in the Production and
## Effects of Sublethal Doses of Acetamiprid and Thiamethoxam on the Behavior of the Honeybee (Apis me
## Exposure to Neonicotinoids Influences the Motor Function of Adult Worker Honeybees
## Toxicity of Neonicotinoid Insecticides on Different Honey Bee Genotypes
## Chronic Exposure of Imidacloprid and Clothianidin Reduce Queen Survival, Foraging, and Nectar Stori
## (Other)

```

```
## Source Publication.Year
## Agric. For. Entomol.11(4): 405-419 : 200 Min. :1982
## Environ. Entomol.41(2): 377-386 : 100 1st Qu.:2005
## Arch. Environ. Contam. Toxicol.54(4): 653-661: 96 Median :2010
## Ecotoxicology23:1409-1418 : 93 Mean :2008
## Bull. Insectol.66(1): 119-126 : 88 3rd Qu.:2013
## PLoS One9(3): 14 p. : 82 Max. :2019
## (Other) :3964
## Summary.of.Additional.Parameters
## Purity: \xca NR - NR | Organism Age: \xca NR - NR Not reported | Conc 1 (Author): \xca Active ingred
## Purity: \xca NR - NR | Organism Age: \xca NR - NR Not reported | Conc 1 (Author): \xca Active ingred
## Purity: \xca NR - NR | Organism Age: \xca NR - NR Not reported | Conc 1 (Author): \xca Active ingred
## Purity: \xca NR - NR | Organism Age: \xca NR - NR Not reported | Conc 1 (Author): \xca Active ingred
## Purity: \xca NR - NR | Organism Age: \xca NR - NR Not reported | Conc 1 (Author): \xca Active ingred
## Purity: \xca NR - NR | Organism Age: \xca NR - NR Not reported | Conc 1 (Author): \xca Formulation I
## (Other)
```

```
##finding number of rows and columns
nrow(Neonics)
```

```
## [1] 4623
```

```
ncol(Neonics)
```

```
## [1] 30
```

```
##finding the class of the dataset
class(Neonics)
```

```
## [1] "data.frame"
```

```
#5.
```

```
##The "Neonics" dataframe has 30 columns and 4623 rows/observations
```

6. Using the `summary` function on the “Effect” column, determine the most common effects that are studied. Why might these effects specifically be of interest?

```
#6.
##obtaining summary statistics of effects studied
summary(Neonics$Effect)
```

```
## Accumulation Avoidance Behavior Biochemistry
## 12 102 360 11
## Cell(s) Development Enzyme(s) Feeding behavior
## 9 136 62 255
## Genetics Growth Histology Hormone(s)
## 82 38 5 1
## Immunological Intoxication Morphology Mortality
## 16 12 22 1493
## Physiology Population Reproduction
## 7 1803 197
```

Answer: Most common effects studied: 1. Population (1803) 2. Mortality (1493) 3. Behavior (360)
The researchers are interested in these effects because they are measures of how the insecticides impact insects. Population abundance and mortality demonstrate how insecticides impact insect viability (yes??) while insect behavior could include important life history activities like feeding and mating.

7. Using the `summary` function, determine the six most commonly studied species in the dataset (common

name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed.

#7.

##obtaining summary statistics of species studied

summary(Neonics\$Species.Common.Name)

##	Honey Bee	Parasitic Wasp
##	667	285
##	Buff Tailed Bumblebee	Carniolan Honey Bee
##	183	152
##	Bumble Bee	Italian Honeybee
##	140	113
##	Japanese Beetle	Asian Lady Beetle
##	94	76
##	Euonymus Scale	Wireworm
##	75	69
##	European Dark Bee	Minute Pirate Bug
##	66	62
##	Asian Citrus Psyllid	Parastic Wasp
##	60	58
##	Colorado Potato Beetle	Parasitoid Wasp
##	57	51
##	Erythrina Gall Wasp	Beetle Order
##	49	47
##	Snout Beetle Family, Weevil	Sevenspotted Lady Beetle
##	47	46
##	True Bug Order	Buff-tailed Bumblebee
##	45	39
##	Aphid Family	Cabbage Looper
##	38	38
##	Sweetpotato Whitefly	Braconid Wasp
##	37	33
##	Cotton Aphid	Predatory Mite
##	33	33
##	Ladybird Beetle Family	Parasitoid
##	30	30
##	Scarab Beetle	Spring Tiphia
##	29	29
##	Thrip Order	Ground Beetle Family
##	29	27
##	Rove Beetle Family	Tobacco Aphid
##	27	27
##	Chalcid Wasp	Convergent Lady Beetle
##	25	25
##	Stingless Bee	Spider/Mite Class
##	25	24
##	Tobacco Flea Beetle	Citrus Leafminer
##	24	23
##	Ladybird Beetle	Mason Bee
##	23	22
##	Mosquito	Argentine Ant
##	22	21
##	Beetle	Flatheaded Appletree Borer
##	21	20

##	Horned Oak Gall Wasp	Leaf Beetle Family
##	20	20
##	Potato Leafhopper	Tooth-necked Fungus Beetle
##	20	20
##	Codling Moth	Black-spotted Lady Beetle
##	19	18
##	Calico Scale	Fairyfly Parasitoid
##	18	18
##	Lady Beetle	Minute Parasitic Wasps
##	18	18
##	Mirid Bug	Mulberry Pyralid
##	18	18
##	Silkworm	Vedalia Beetle
##	18	18
##	Araneoid Spider Order	Bee Order
##	17	17
##	Egg Parasitoid	Insect Class
##	17	17
##	Moth And Butterfly Order	Oystershell Scale Parasitoid
##	17	17
##	Hemlock Woolly Adelgid Lady Beetle	Hemlock Woolly Adelgid
##	16	16
##	Mite	Onion Thrip
##	16	16
##	Western Flower Thrips	Corn Earworm
##	15	14
##	Green Peach Aphid	House Fly
##	14	14
##	Ox Beetle	Red Scale Parasite
##	14	14
##	Spined Soldier Bug	Armoured Scale Family
##	14	13
##	Diamondback Moth	Eulophid Wasp
##	13	13
##	Monarch Butterfly	Predatory Bug
##	13	13
##	Yellow Fever Mosquito	Braconid Parasitoid
##	13	12
##	Common Thrip	Eastern Subterranean Termite
##	12	12
##	Jassid	Mite Order
##	12	12
##	Pea Aphid	Pond Wolf Spider
##	12	12
##	Spotless Ladybird Beetle	Glasshouse Potato Wasp
##	11	10
##	Lacewing	Southern House Mosquito
##	10	10
##	Two Spotted Lady Beetle	Ant Family
##	10	9
##	Apple Maggot	(Other)
##	9	670

Answer: Most common species studied: 1. Honey Bee (667) 2. Parasitic Wasp (285) 3. Buff

Tailed Bumblebee (183) [interest over other insects????????????]

8. Concentrations are always a numeric value. What is the class of `Conc.1..Author.` in the dataset, and why is it not numeric?

```
#8.  
##determining class of "Conc.1..Author" column  
class(Neonics$Conc.1..Author.)
```

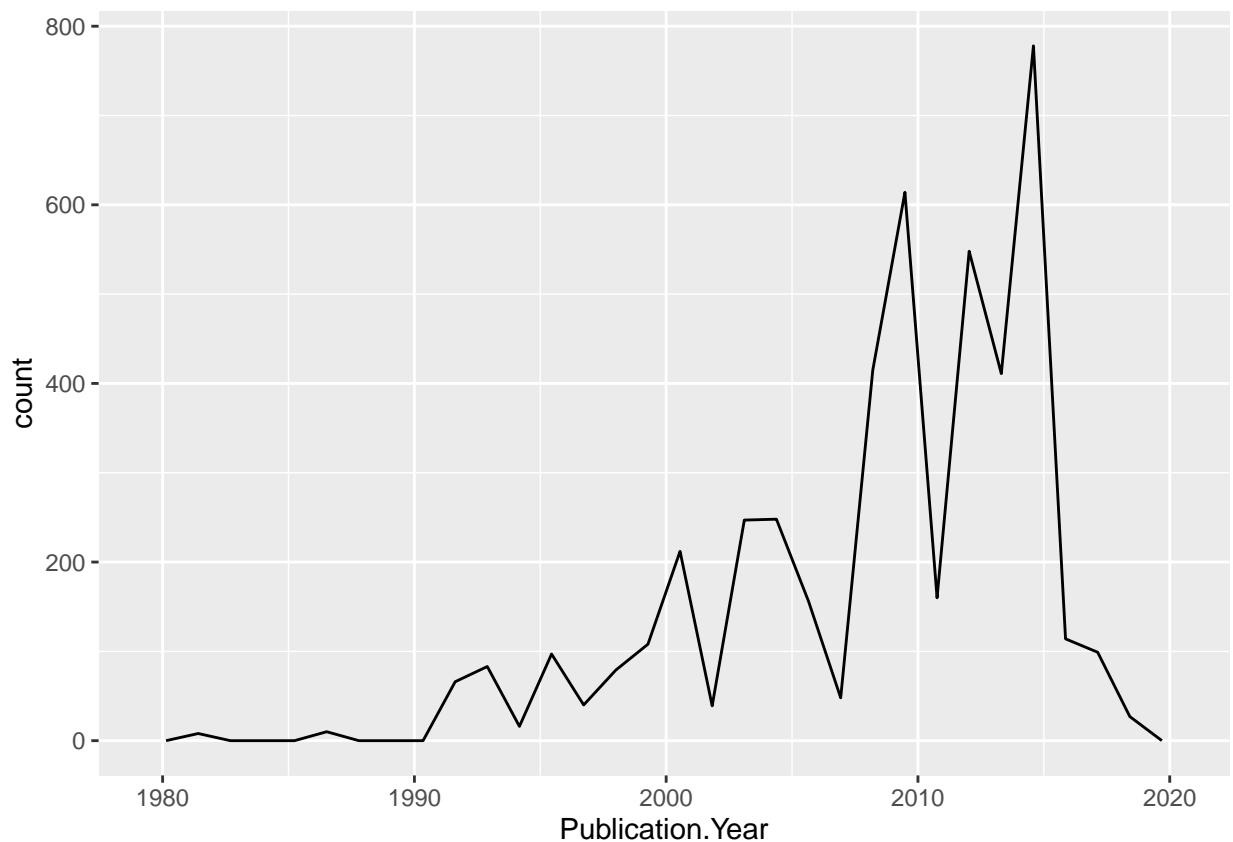
```
## [1] "factor"
```

Answer: The class of “Conc.1..Author” is “factor.” [why not numeric? not sure what column means??]

Explore your data graphically (Neonics)

9. Using `geom_freqpoly`, generate a plot of the number of studies conducted by publication year.

```
#9  
##creating plot of studies by publication year  
studies_by_year <- ggplot(data=Neonics, aes(x=Publication.Year))+  
  geom_freqpoly()  
studies_by_year  
  
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```



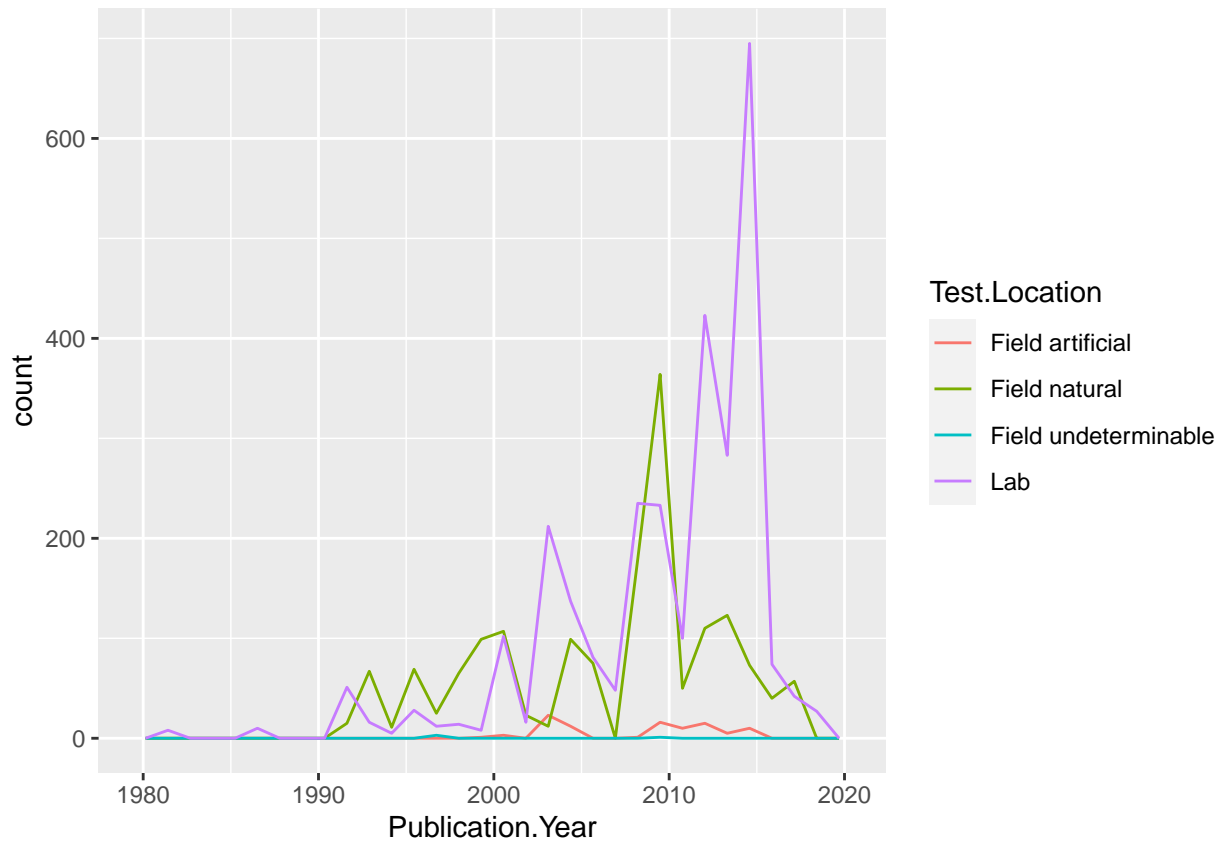
```
##change bin width???????
```

10. Reproduce the same graph but now add a color aesthetic so that different `Test.Location` are displayed as different colors.


```
#10
##creating plot of studies by publication year and test location
studies_by_year2 <- ggplot(data=Neonics, aes(x=Publication.Year, color=Test.Location))+
  geom_freqpoly()

studies_by_year2

## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```



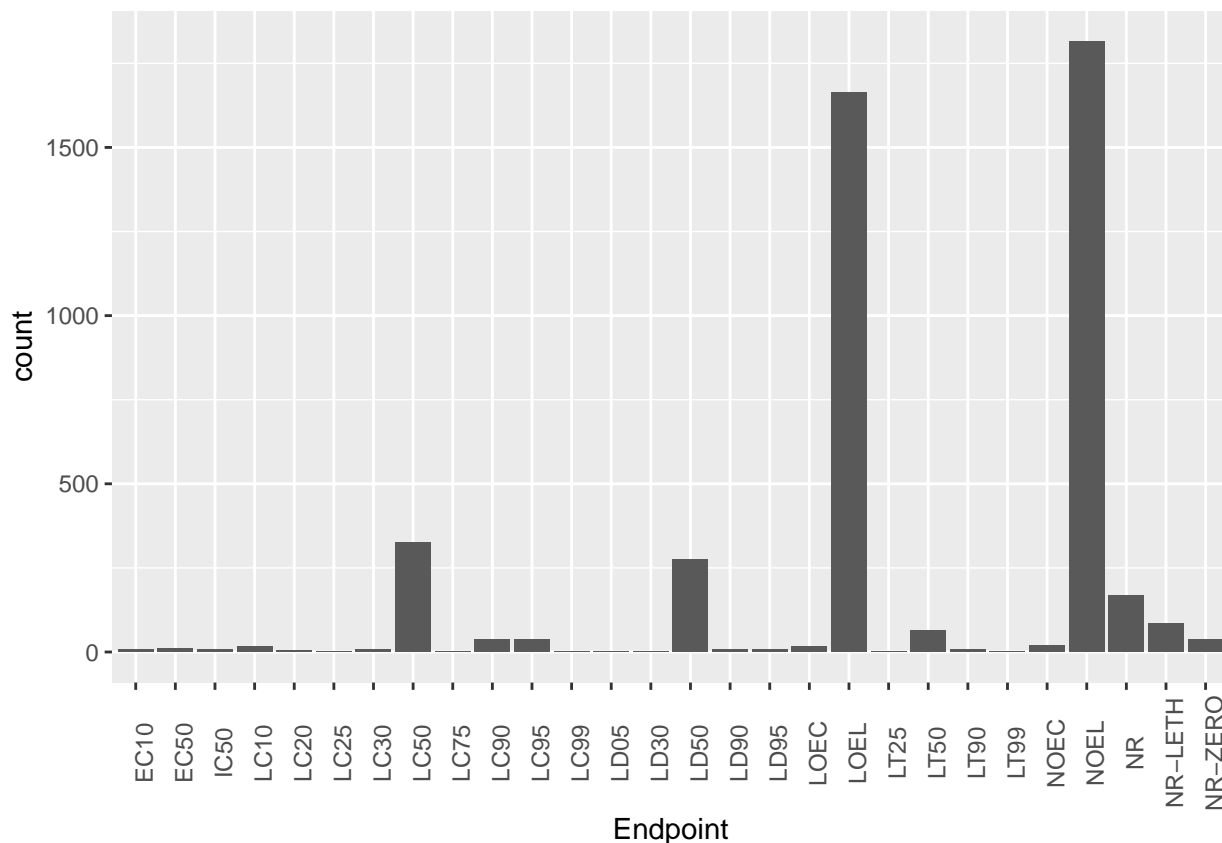
Interpret this graph. What are the most common test locations, and do they differ over time?

Answer: The most common test locations are labs and natural fields. The number of lab studies generally seems to have increased over time (before 2020) while the number of natural field studies peaked at around 2010 and has declined since then (perhaps due to increasing popularity of lab tests).

11. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX_CodeAppendix for more information.

```
#11
##creating bar graph for endpoint counts
endpoint_counts_graph <- ggplot(data=Neonics, aes(x=Endpoint))+
  geom_bar()+
  theme(axis.text.x=element_text(angle=90))

##output for endpoint counts graph
endpoint_counts_graph
```



Answer: The two most common endpoints are NOEL (no-observable-effect-level) and LOEL (lowest-observable-effect-level). The NOEL is defined as the greatest concentration of chemical that does not cause an effect significantly different than the control. The LOEL refers to the lowest chemical concentration that causes an effect that varies significantly from the control.

Explore your data (Litter)

- Determine the class of collectDate. Is it a date? If not, change to a date and confirm the new class of the variable. Using the unique function, determine which dates litter was sampled in August 2018.

```
#12
##determining the class of the litter collect date
class(Litter$collectDate) ##class = "factor"

## [1] "factor"

Litter$collectDate_date <- as.Date(Litter$collectDate) ##get NAs when specifying format
class(Litter$collectDate_date)

## [1] "Date"
```

- Using the unique function, determine how many plots were sampled at Niwot Ridge. How is the information obtained from unique different from that obtained from summary?

```
#13.
##determining number of plots sampled at Niwot Ridge
length(unique(Litter$plotID)) ###is this okay??????????
```

```
## [1] 12
```

```
##running summary command on plots
```

```
summary(Litter$plotID)
```

```
## NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 NIWO_058 NIWO_061
##      20      19      18      15      14       8      16      17
## NIWO_062 NIWO_063 NIWO_064 NIWO_067
##      14      14      16      17
```

Answer: There were 12 plots sampled at Niwot Ridge. A summary of Litter\$plotID gives you the number of observations at each plot while the unique function provides the number of plots studied.

14. Create a bar graph of functionalGroup counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

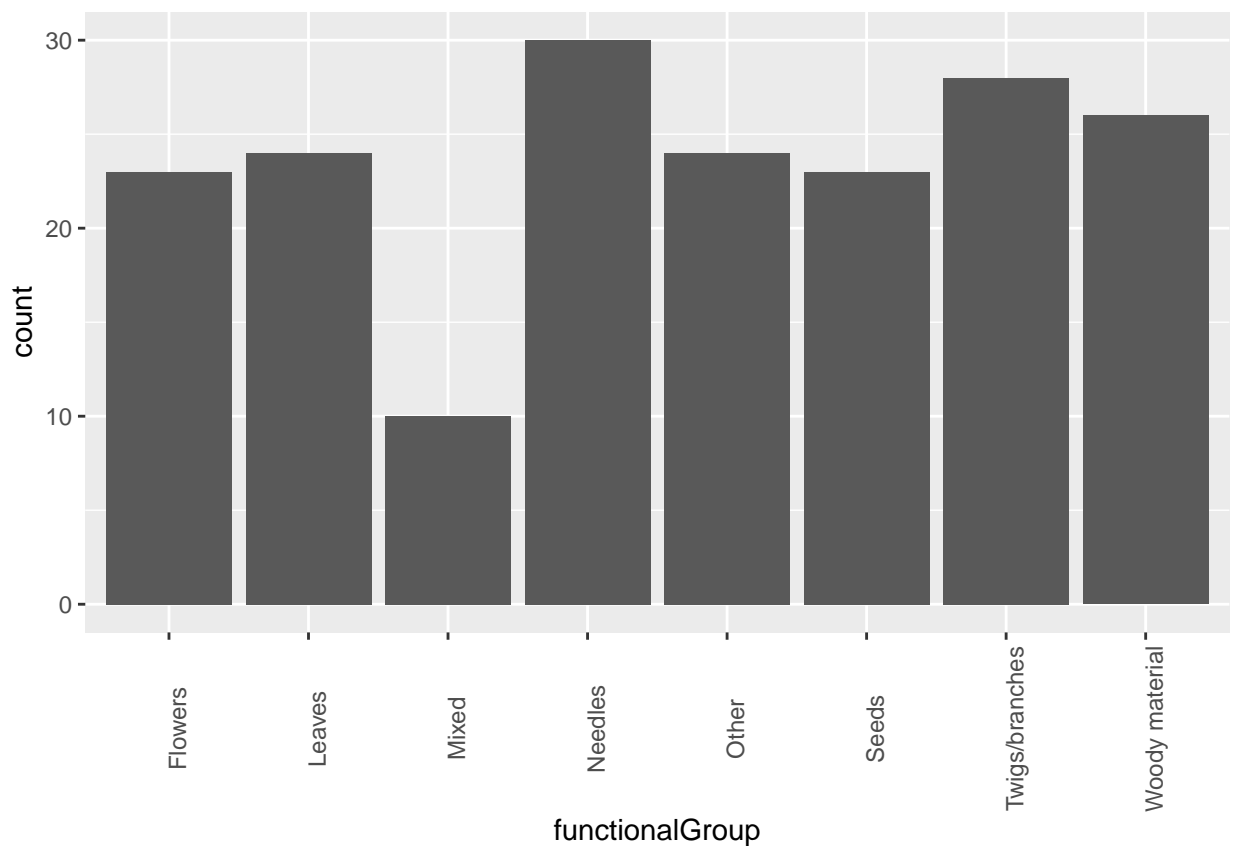
```
#14
```

```
##creating functionalGroup counts bar graph
```

```
functionalGroup_bar_graph <- ggplot(data=Litter, aes(x=functionalGroup))+
  geom_bar()+
  theme(axis.text.x=element_text(angle=90))
```

```
##output for functionalGroup counts bar graph
```

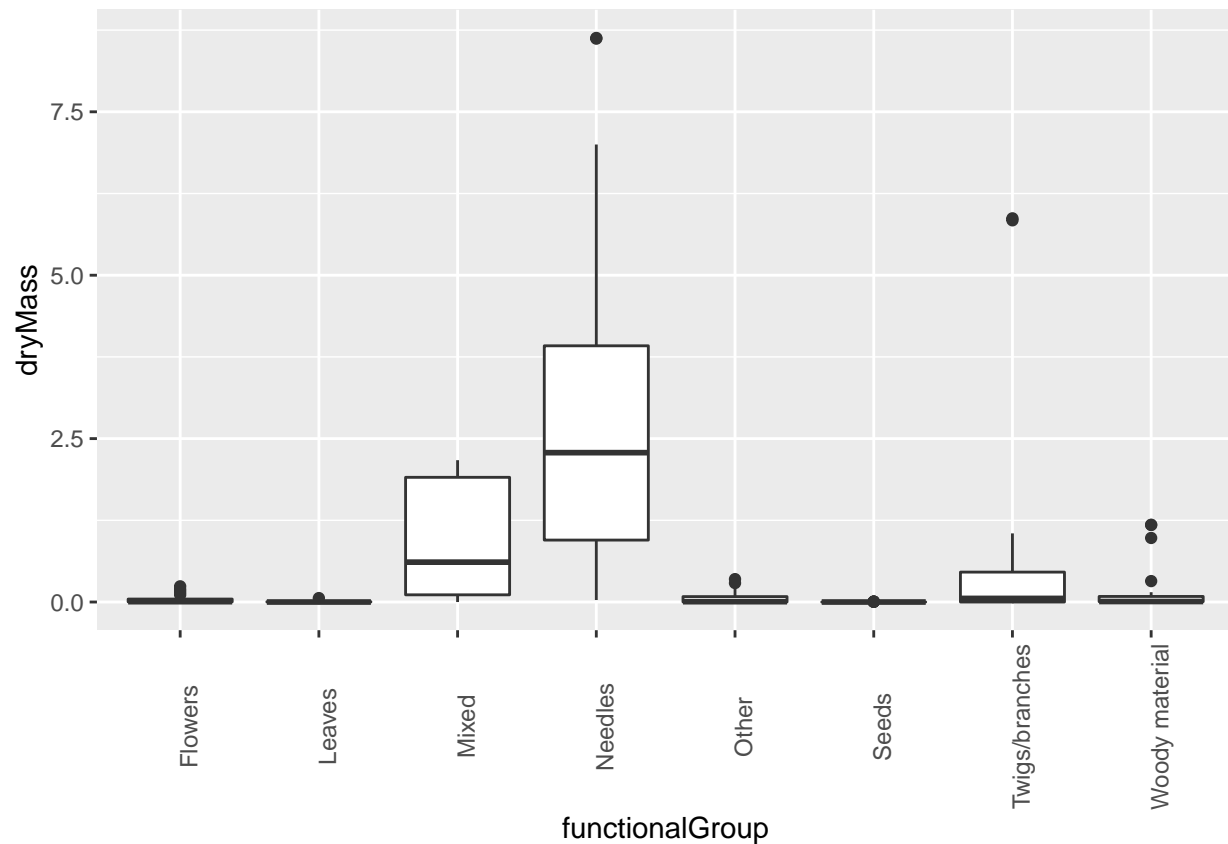
```
functionalGroup_bar_graph
```



15. Using geom_boxplot and geom_violin, create a boxplot and a violin plot of dryMass by functional-Group.

```
#15
##functionalGroup boxplot
functionalGroup_boxplot <- ggplot(data=Litter, aes(x=functionalGroup, y=dryMass))+
  geom_boxplot()+
  theme(axis.text.x=element_text(angle=90))

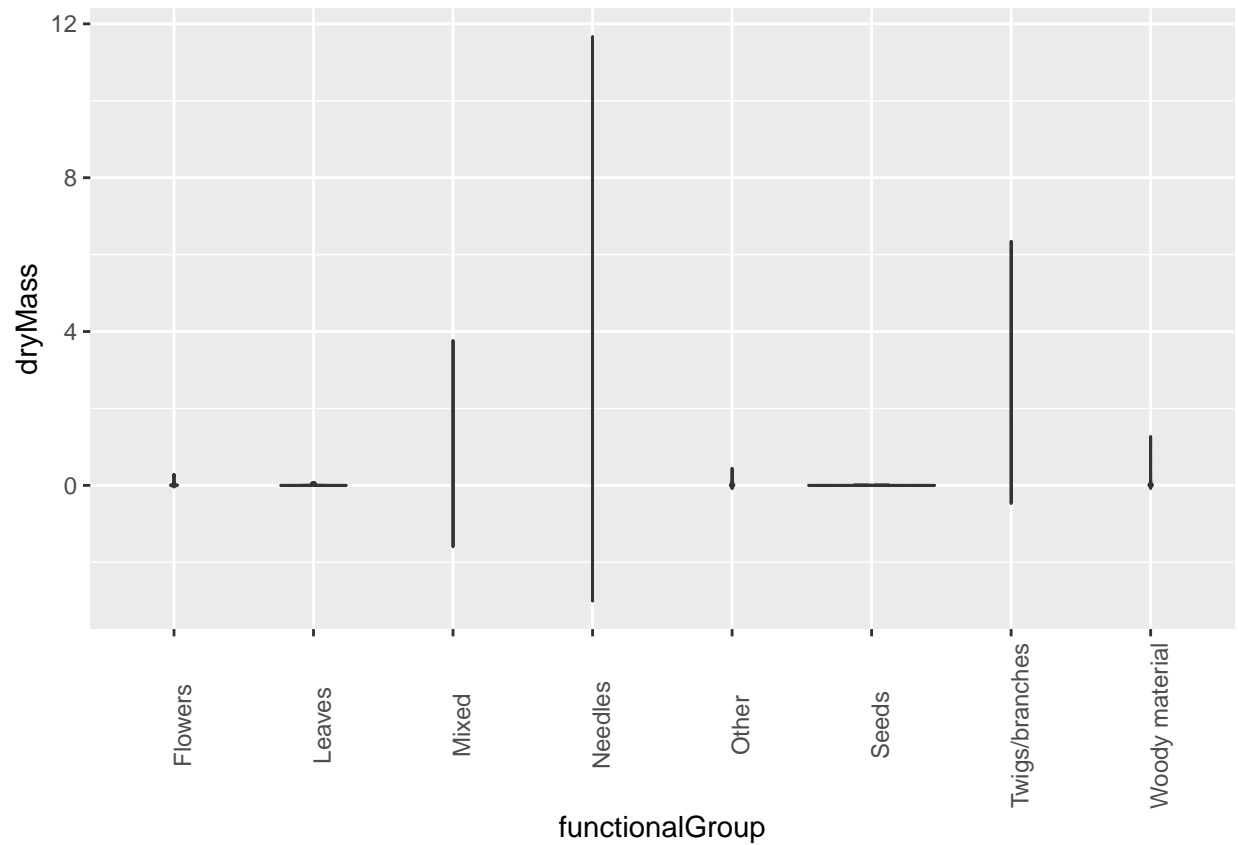
##boxplot output
functionalGroup_boxplot
```



```
##functionalGroup violin plot
functionalGroup_violin <- ggplot(data=Litter, aes(x=functionalGroup, y=dryMass))+
  geom_violin(pt.size = 10, trim=F)+
  theme(axis.text.x=element_text(angle=90))

## Warning: Ignoring unknown parameters: pt.size

##violin plot output
functionalGroup_violin ##violin plot just coming out as vertical lines??????????????????
```



Why is the boxplot a more effective visualization option than the violin plot in this case?

Answer:

What type(s) of litter tend to have the highest biomass at these sites?

Answer: The type of litter with the highest biomass is needles, followed by mixed litter and twigs and branches.