

HMMancestry

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Type Package

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Description to be added

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est_maxLnL	<i>Numerically Estimating the Maximum LnL parameter values from the data</i>
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Description

Infers the genome-wide recombination rate (Morgans / bp) and the assignment probability directly from the data using maximum likelihood estimates.

Usage

```
est_maxLnL(dat, ploidy = "diploid", initial_p_assign = "NULL",
  initial_scale = "NULL", tolerance = 0.001, n_coarse_steps = 5,
  n_iterations = 30, plot = FALSE)
```

Arguments

dat	a data.frame with five columns: <ul style="list-style-type: none"> • Ind The individual ID • Chr The chromosome ID • Snp The (sorted) snp location (in bps) • p0 The number of reads that mapped to parent 0 • p1 The number of reads that mapped to parent 1 Note that the names of the columns can be different and there can be additional columns to the right (est_maxLnL only uses the first 5 columns of the data.frame).
ploidy	The ploidy to analyze. If ploidy="diploid" (default) then fb_diploid is used to infer states and estimate likelihood. If ploidy="haploid" then fb_haploid will be used.
initial_p_assign	The initial assignment probability (see details). If "NULL" (default), then a coarse search will be performed with n_coarse_steps equally spaced from 0.95 to 0.999.
initial_scale	The initial genome-wide recombination rate (Morgans / bp). If "NULL" (default), then a coarse search will be performed n_coarse_steps equally spaced from 1e-06 to 1e-04.
tolerance	The tolerance used to stop the search.
n_coarse_steps	The size of the 1D (if either initial_p_assign or initial_scale is "NULL") or 2D grid (if both are "NULL"). Increasing n_coarse_steps can greatly increase computation time.
n_iterations	The number of iterations during the fine scale parameter estimate search (see details).

Details

est_maxLnL has a coarse and fine scale method to estimate the genome-wide recombination rate, \hat{c} , and assignment probability, \hat{p} . The coarse scale uses either [fb_haploid](#) or [fb_diploid](#) output and estimates the natural log-likelihood, LnL, of the data across a coarse (in parameter space) grid of varying p or c values and the fine scale uses a two-variable Newton-Raphson method to hone in on a closer estimate. For further details see Hether et al. (in prep).

Value

A data.frame containing the following columns:

- `p_assign_hat` An estimate of the assignment probability
- `scale_hat` An estimate of mean recombination rate
- `n_iterations` The number of iterations carried out

Author(s)

Tyler D. Hether

References

Hether, T.D., C. G. Wiench1, and P.A. Hohenlohe (in review). 2015. Novel molecular and analytical tools for efficient estimation of rates of meiotic crossover, non-crossover and gene conversion

P. Deuffhard, Newton Methods for Nonlinear Problems. Affine Invariance and Adaptive Algorithms. Springer Series in Computational Mathematics, Vol. 35. Springer, Berlin, 2004.

See Also

[fb_haploid](#), [fb_haploid](#)

Examples

```
# Simulating 30 haploid recombinants
set.seed(1234567)      # For reproducibility
n_spores <- 30         # number of recombinants
l <- 1000              # number of snps to simulate
c <- 1e-06             # recombination rate between snps (Morgan/bp)
snps <- c(1:l)*1e2     # snps are evenly spaced 100bps apart
p_a <- 0.985           # assignment probability
coverage <- 1          # mean coverage
# Now simulate
sim <- sim_en_masse(n.spores=n_spores, scale=c, snps=snps,
  p.assign=p_a, mu.rate=0, f.cross=1, f.convert=0,
  length.conversion=0, coverage=coverage)
# Now estimate params
res <- est_maxLnL(sim, ploidy="haploid", plot=TRUE)
res
```

fb_diploid

Inferring hidden ancestry states from diploids

Description

Uses the forward-backward algorithm to estimate ancestry along a given chromosome for a given genotyped diploid.

Usage

```
fb_diploid(snp_locations, p0, p1, p_assign, scale)
```

Arguments

snp_locations	a numeric vector specifying the locations of each snp (in bps). This vector is assumed to be ordered (sorted from smallest to largest snp).
p0	a vector specifying the number of reads that mapped to parent 0. p0 is assumed to be in the same order as snp_locations.
p1	a vector specifying the number of reads that mapped to parent 1. p1 is assumed to be in the same order as snp_locations.
p_assign	a value specifying the assignment probability (see details).
scale	a numeric specifying the genome-wide recombination rate (Morgans / bp). scale is assumed to be between 0 and 1 but in practice it is usually quite small.

Details

This is an extension of [fb_haploid](#) with three hidden states: two homozygous states and a heterozygous state.

Value

a dataframe with the following columns:

- snp_locations
- p0
- p1
- The emission matrix (3 columns) specifying the emission probability of belong to the only parent 0 (emiss.1), both parents (emiss.2), and only parent 2 (emiss.3)
- the forward probability matrix (3 columns) giving the scaled forward probabilities of belong to the only parent 0 (forward.1), both parents (forward.2), and only parent 2 (forward.3)
- the backward probability matrix (3 columns) giving the scaled backward probabilities of belong to the only parent 0 (backward.1), both parents (backward.2), and only parent 2 (backward.3)
- Fscale The forward scaling factor for each snp
- Bscale The backward scaling factor for each snp
- posterior probability matrix (3 columns) posterior probabilities for belong to each parental state
- states_inferred a vector of length snp_locations giving the state with the highest posterior.
- lnL a numeric giving the total likelihood of the data (see [fb_haploid](#)).

Author(s)

Tyler D. Hether

References

- Hether, T.D., C. G. Wiench, and P.A. Hohenlohe (in review). 2015. Novel molecular and analytical tools for efficient estimation of rates of meiotic crossover, non-crossover and gene conversion
- Drubin, R. S. Eddy, A. Krogh, and G. Mitchison. 1998. Biological Sequence Analysis: Probabilistic Models of proteins and nucleic acids. Cambridge University Press, Cambridge CB2 8RU, UK.

See Also[fb_haploid](#)**Examples**

```

set.seed(1234567)      # For reproducibility
n_spores <- 1          # number of spores
l <- 75                # number of snps to simulate
c <- 3.5e-05           # recombination rate between snps (Morgan/bp)
snps <- c(1:l)*1.3e4    # snps are evenly spaced 20kbp apart
p_a <- 0.95            # assignment probability
coverage <- 2.1        # mean coverage
# Now simulate two haploids
sim1 <- sim_en_masse(n.spores=n_spores, scale=c, snps=snps,
  p.assign=p_a, mu.rate=0, f.cross=0.8, f.convert=0.3,
  length.conversion=2e3, coverage=coverage)
sim2 <- sim_en_masse(n.spores=n_spores, scale=c, snps=snps,
  p.assign=p_a, mu.rate=0, f.cross=0.8, f.convert=0.3,
  length.conversion=2e3, coverage=coverage)
# Now merge the two haploids to make a diploid
p0 <- sim1$p0+sim2$p0
p1 <- sim1$p1+sim2$p1
res <- fb_diploid(snp_locations=sim1$Snp, p0=p0, p1=p1, p_assign=p_a, scale=c)
res

```

fb_haploid

*Inferring hidden ancestry states from haploids***Description**

Uses the forward-backward algorithm to estimate ancestry along a given chromosome for a given genotyped tetrad or simulated data.

Usage

```
fb_haploid(snp_locations, p0, p1, p_assign, scale)
```

Arguments

snp_locations	a numeric vector specifying the locations of each snp (in bps). This vector is assumed to be ordered (sorted from smallest to largest snp).
p0	a vector specifying the number of reads that mapped to parent 0. p0 is assumed to be in the same order as snp_locations.
p1	a vector specifying the number of reads that mapped to parent 1. p1 is assumed to be in the same order as snp_locations.
p_assign	a value specifying the assignment probability (see details).
scale	a numeric specifying the the genome-wide recombination rate (Morgans / bp). scale is assumed to be between 0 and 1 but in practice it is usually quite small.

Details

The function `fb_haploid` attempts to estimate parental genotypic 'states' along a chromosome given empirical or simulated F2 cross data. Next-generation data inherits both sequencing error and missing data – especially when sequencing coverage is low. Also, parental populations can be polymorphic with respect to gene frequencies before admixture. In these cases the parental state is ambiguous or unknown. `fb_haploid` takes these uncertainties into account and estimates the most likely sequence of ancestry.

The three steps taken in `fb_haploid` are:

1. Calculate forward probabilities for each state (5' to 3')
2. Calculate backward probabilities for each state (3' to 5')
3. From these two probabilities, calculate the posterior probability that a snp location is of a given parental state.

The two element vector of forward probabilities, f_i , for each parental state at each position i is calculated as:

$$f_i = \mathbf{e}_i \mathbf{T}_i \mathbf{f}_{i-1}$$

where e_i is the emission probabilities for each state (see below), T_i is a 2-by-2 matrix describing the transition (recombination) probabilities between two states, and f_{i-1} is the forward probability at the previous position along the chromosome (5' of position i). It is assumed that each state is equally likely to occur at the first position.

The emission probabilities are calculated independently for each snp and depends on the sequence reads assigned to parent "0" and parent "1" and `p_assign`. The emission probabilities are calculated using the binomial equation. For example, the emission probability for parental state "0" at snp position i is:

$$e_i^{(0)} = \binom{n}{k_0} p^{k_0} (1-p)^{n-k_0}$$

where n is the sum of the number of reads from both parents at snp i and p is the assignment probability, `p_assign`.

Like the emission probabilities, the transition probabilities are also calculated for each snp position. The transition probabilities in matrix \mathbf{T}_i are calculated from the genome-wide recombination rate, scale, and the physical distance (in bps) between the two snps. Specifically, Haldane's function is used to estimate the probability of getting an odd number of crossovers between snp i and snp $i-1$. For more details, see Hether et al. (in prep).

To avoid underflow, we rescale the forward (and backward) probabilities each iteration to that they sum to unity.

The backward probabilities are calculated similarly but in the 3' to 5' direction. It is assumed that the backward probability is equally likely in each state. Following Durbin et al. (1998) the backward probability at snp position i is:

$$b_i = \mathbf{T}_i \mathbf{e}_{i+1} \mathbf{b}_{i+1}$$

Again, these probabilities are rescaled to avoid underflow.

The posterior probability that the state at position i is k given the observed sequence read counts for each parent at position i , x_i is calculated by:

$$P(\pi_i = k \mid x_i) = \frac{f_i b_i}{\sum_{k=0}^1 f_i^{(k)} b_i^{(k)}}$$

Ties in posterior probabilities are rare with sufficient coverage and/or signal. However, in the event of a tie state 1 is picked.

Finally, we can determine the log likelihood of the whole sequence of observations by summing up the log of all the scale factors in the forward probability calculation.

Value

a dataframe with the following columns:

- snp_locations
- p0
- p1
- The emission matrix (2 columns) specifying the emission probability of belong to the parent 0 (emiss.1) and parent 2 (emiss.2)
- the forward probability matrix (2 columns) giving the scaled forward probabilities of belong to the parent 0 (forward.1) and parent 2 (forward.2)
- the backward probability matrix (2 columns) giving the scaled backward probabilities of belong to the parent 0 (backward.1) and parent 2 (backward.2)
- Fscale The forward scaling factor for each snp
- Bscale The backward scaling factor for each snp
- posterior probability matrix (2 columns) posterior probabilities for belong to each parental state
- states_inferred a vector of length snp_locations giving the state with the highest posterior.
- lnL a numeric giving the total likelihood of the data (see details).

Author(s)

Tyler D. Hether

References

- Hether, T.D., C. G. Wiench1, and P.A. Hohenlohe (in review). 2015. Novel molecular and analytical tools for efficient estimation of rates of meiotic crossover, non-crossover and gene conversion
- Drubin, R. S. Eddy, A. Krogh, and G. Mitchison. 1998. Biological Sequence Analysis: Probabilistic Models of proteins and nucleic acids. Cambridge University Press, Cambridge CB2 8RU, UK.

See Also

[fb_diploid](#)

Examples

```

set.seed(1234567)      # For reproducibility
n_spores <- 1          # number of spores
l <- 75                # number of snps to simulate
c <- 3.5e-05           # recombination rate between snps (Morgan/bp)
snps <- c(1:l)*1.3e4   # snps are evenly spaced 20kbp apart
p_a <- 0.95            # assignment probability
coverage <- 2.1        # mean coverage
# Now simulate
sim1 <- sim_en_masse(n.spores=n_spores, scale=c, snps=snps,
  p.assign=p_a, mu.rate=0, f.cross=0.8, f.convert=0.3,
  length.conversion=2e3, coverage=coverage)
fb_haploid(snp_locations=sim1$Snp, p0=sim1$p0, p1=sim1$p1, p_assign=p_a, scale=c)

```

id_recombination_events

Identify crossover points from state sequences and snp locations

Description

This is a simple function that takes a vector of parental states along a chromosome and identifies where states have changed.

Usage

```
id_recombination_events(snps_genotypes_df)
```

Arguments

snps_genotypes_df

a data.frame with two columns:

- snps a vector of ordered snp locations (in bps)
- states a vector of corresponding inferred states, either haploid (2 states = 0 or 1) or diploid with three states possible (0,1,2)

Value

a data.frame containing the midpoint between focal snp i and snp $i - 1$ and whether their states were the same (0) or different (1).

Author(s)

Tyler D. Hether

See Also

[fb_haploid](#), [fb_diploid](#)

Examples

```

df <- data.frame(snps=100*(1:10),
  states=c(rep(0,4), rep(1,6)))
id_recombination_events(df)

```


infer_tracts

*Identify crossover, non-crossover, and gene conversion tracts***Description**

Infers different types of tracts (crossover, non-crossover, and gene conversion) along a chromosome

Usage

```
infer_tracts(dat, threshold_size = 2500)
```

Arguments

data A data.frame with 7 columns:

1. c("Tetrad", "Chr", "Snp", "one", "two", "three", "four")
2. Tetrad specifying the tetrad ID
3. Chr giving the chromosome name
4. Snp a vector of snp locations (in bps)
5. one the inferred states for spore 1
6. two the inferred states for spore 2
7. three the inferred states for spore 3
8. four the inferred states for spore 4

threshold_size The size (in bps) of the threshold (see details)

Details

Uses the 3 step classification scheme described in Hether et al. (in review) to identify the location of these specific CO, NCO, and telomeric gene conversion tracts. Specifically, infer_tracts attempts to identify the following tracts:

- 2_2 a tract with 2:2 segregation
- COyesGC a region of gene conversion that was associated with a crossover event
- COnoGC a region between a crossover event that did not have a detectable gene conversion
- NCO a non-crossover; a gene conversion tract without crossing
- GC_tel a gene conversion tract located at the chromosome end

Value

A data.frame containing the following columns:

1. type The type of inferred tract
2. start_snp The starting snp position in base pairs
3. end_snp The ending snp position in base pairs
4. extent The size of the tract. For COnoGC, this extent is the s spanning region between flanking CO events.

Author(s)

Tyler D. Hether

References

Hether, T.D., C. G. Wiench1, and P.A. Hohenlohe (in review). 2015. Novel molecular and analytical tools for efficient estimation of rates of meiotic crossover, non-crossover and gene conversion

Examples

```
set.seed(1234567)      # For reproducibility
n_tetrads <- 3          # number of tetrads
l <- 1000              # number of snps to simulate
c <- 3e-05             # recombination rate between snps (Morgan/bp)
snps <- c(1:l)*250     # snps are evenly spaced 250 bp apart
p_a <- 0.95            # assignment probability
coverage <- 1          # mean coverage
# simulate tetrads
tetrad <- sim_tetrad(n.tetrads=n_tetrads, scale=c, snps=snps,
  p.assign=p_a, mu.rate=1e-03, f.cross=0.6, f.convert=0.8,
  length.conversion=2e3, coverage=coverage)
#' # Example 1 -- infer tracts directly from simulated data
inf_tracts_sim <- infer_tracts(tetrad)
inf_tracts_sim
#' # Example 2 -- infer tracts from inferred data
inf_states <- ddply(tetrad, .(Tetrad, Spore, Chr),
  function(x){
    return(fb_haploid(snp_locations=x$Snp, p0=x$p0,
      p1=x$p1, p_assign=p_a, scale=c)))
inf_tracts_inf_states <- infer_tracts(inf_states)
inf_tracts_inf_states
```

make_parents

Simulate haploid parental genotypes along a chromosome

Description

This function creates divergent genotypes between two parents. The first parent's genotypes are denoted by a vector of zeros of length L while the second parent's genotypes are a vector of ones.

Usage

```
make_parents(snps)
```

Arguments

snps a vector giving the locations of snps along a chromosome

Value

an object of class `parent.genomes` giving the parental ancestry at each snp.

Author(s)

Tyler D. Hether

References

none.

See Also

[recombine_index](#), [recobmine](#)

Examples

```
make_parents(snps=c(1:10)*10)
```

recombine	<i>Simulate recombination (crossovers or non-crossovers) along a chromosome</i>
-----------	---

Description

This function simulates recombination between two parent chromosomes generated from class `parent.genomes`.

Usage

```
recombine(parents, r.index, mu.rate = 0, f.cross = 0.5, f.convert = 0,
  length.conversion = 20)
```

Arguments

<code>parents</code>	an object of class <code>parent.genomes</code> specifying the parental ancestry at each simulated snp.
<code>r.index</code>	a vector of length <code>snps-1</code> specifying whether a recombination is to be simulated (1) or not (0) in between two adjacent snps.
<code>mu.rate</code>	a numeric between 0 and 1 (inclusive) specifying the per snp mutation rate.
<code>f.cross</code>	a numeric between 0 and 1 (inclusive) giving the frequency of recombination events that result in crossing over. This is same as 1 minus the frequency of non-crossovers.
<code>f.convert</code>	a numeric between 0 and 1 (inclusive) that gives the frequency of gene conversion during recombination.
<code>length.conversion</code>	an integer specifying the mean (and variance) of a given gene conversion tract (in bps).

Details

This function simulates crossover events (with or without gene conversions) and non-crossover events given the parental snps and location of recombination events. First, parental chromosomes are duplicated into 2 pairs of sister chromatids. During this step mutations occur at rate `mu.rate` that switch parental alleles. Second, recombination is simulated by systematically going down the length of the chromosome. At recombination points, identified in `r.index`, two non-sister chromatids are picked at random. Third, a CO or NCO is simulated; this is done at a frequency of `f.cross` or `1-f.cross`, respectively. If a CO occurs, the genotypes of the 3' end of the non-sister chromatids are

simply switched. If a NCO occurs, no such switch takes place. Note that it is impossible to detect NCO events without an associated gene conversion tract. Following every CO or NCO event gene conversion is simulated at a rate of `f.convert`. During a gene conversion, one of the two unpicked chromatids' genotypes are switched. The length of the gene conversion tract is sampled from a poisson distribution with mean (variance) of `length.conversion`. Note that the gene conversion tract will stop at the end of the chromosome if necessary.

Value

an object of class `recombine` that contains the following data:

- `parents` input data
- `r.index` input data
- `m.rate` input data
- `f.cross` input data
- `f.convert` input data
- `length.conversion` input data
- `chromatids.mutated_snps` a list giving the genotypes of 4 chromatids (two pairs of sister chromatids) following mutation
- `chromatids.recombined` a list giving the genotypes of 4 chromatids following recombination

Author(s)

Tyler D. Hether

See Also

[recombine_index](#)

Examples

```
set.seed(1234567)      # For reproducibility
l <- 50                # number of snps to simulate
c <- 3e-05             # recombination rate between snps (Morgan/bp)
snps <- c(1:l)*1e4     # snps are evenly spaced 10kbp apart
p <- make_parents(snps) # make the parents
#
# The recombination indeces are:
r_points <- recombine_index(scale=c, snps=snps)
#
# Now recombine the parents:
recomb_sim <- recombine(parents=p, r.index=r_points, mu.rate=1e-05,
  f.cross=0.6, f.convert=0.2, length.conversion=15000)
recomb_sim
```

recombine_index	<i>Simulate recombination points along a chromosome.</i>
-----------------	--

Description

Simulate recombination events given the vector of snp locations and recombination rate(s). Haldane's function is used to simulate the probability of getting an odd number of crossover events between any two snps given their physical distance and the specified recombination rate.

Usage

```
recombine_index(scale, snps)
```

Arguments

scale	either vector of length 1 specifying the genome-wide recombination rate (Morgans/bp) or a vector of length snps-1 specifying the recombination rate between all snps. In either case rates must be positive.
snps	a vector giving the locations of snps along a chromosome

Value

a vector of length snps-1 specifying the location of recombination points.

Author(s)

Tyler D. Hether

See Also

[make_parents](#)

Examples

```
set.seed(1234567)
# Simulating recombination across 1000 loci randomly spaced
# along a 200kbp chromosome
chromSize <- 2e5 # Chromosome size
l <- 1e3 # number of loci to simulate
c <- 1e-04 # Morgans/bp
snps <- sort(sample(size=l, 1:chromSize, replace=FALSE))
# Find recombination points between all the snps
recomb_points <- recombine_index(scale=c, snps=snps)
recomb_points
```

simulate_coverage	<i>Simulate sequencing across simulated data:</i>
-------------------	---

Description

This function populates simulated data of class `recombine` with read counts given a specified coverage and assignment probability.

Usage

```
simulate_coverage(simdata, p.assign, coverage)
```

Arguments

<code>simdata</code>	an object of class <code>recombine</code>
<code>p.assign</code>	a numeric between 0 and 1 (inclusive) that gives the probability of correct sequencing assignment. A value of 1 means that no sequencing error or ancestral polymorphism is to be simulated.
<code>coverage</code>	the mean (and variance) of sequencing read coverage to be simulated. coverage is sampled from a poisson distribution.

Value

an object of class `simulated.coverage` providing the a list of simulated sequencing reads for each parental type for each of the four haploid recombinants. There are four lists in the output – one for each recombinant. Within each list the read counts for parent 0 and parent 1 are given as well as the snp locations and given (simulated) states.

Author(s)

Tyler D. Hether

See Also

[recombine](#)

Examples

```
set.seed(1234567)      # For reproducibility
l <- 50                 # number of snps to simulate
c <- 3e-05              # recombination rate between snps (Morgan/bp)
snps <- c(1:l)*1e4      # snps are evenly spaced 10kbp apart
p_a <- 0.97             # assignment probability
coverage <- 1           # mean coverage
p <- make_parents(snps) # make the parents
#
# The recombination indeces are:
r_points <- recombine_index(scale=c, snps=snps)
#
# Now recombine the parents:
recomb_sim <- recombine(parents=p, r.index=r_points, mu.rate=1e-05,
f.cross=0.6, f.convert=0.2, length.conversion=15000)
```

```
# And finally simulated read coverage for each haploid recombinant
sim_cov <- simulate_coverage(simdata=recomb_sim,
  p.assign=p_a, coverage=coverage)
sim_cov
```

sim_en_masse	<i>Simulate random spores en masse</i>
--------------	--

Description

This is a wrapper function of many other functions that simulates a given number of haploids each recombinant between two parents. As the name implies, information of the all four products are lost since spores are random. See `\line{sim_tetrad}` if all four haploid recombinants are desired.

Usage

```
sim_en_masse(n.spores, scale, snps, p.assign, mu.rate, f.cross, f.convert,
  length.conversion, coverage, chr.name = "I")
```

Arguments

n.spores	An integer specifying the number of spores to simulate.
p.assign	a numeric between 0 and 1 (inclusive) that gives the probability of correct sequencing assignment. A value of 1 means that no sequencing error or ancestral polymorphism is to be simulated.
mu.rate	a numeric between 0 and 1 (inclusive) specifying the per snp mutation rate.
f.cross	a numeric between 0 and 1 (inclusive) giving the frequency of recombination events that result in crossing over. This is same as 1 minus the frequency of non-crossovers.
f.convert	a numeric between 0 and 1 (inclusive) that gives the frequency of gene conversion during recombination.
length.conversion	an integer specifying the mean (and variance) of a given gene conversion tract (in bps).
chr.name	a numeric or character value specifying the chromosome name (default to "I")
scale	either vector of length 1 specifying the genome-wide recombination rate (Morgans/bp) or a vector of length snps-1 specifying the recombination rate between all snps. In either case rates must be positive.
snps	a vector giving the locations of snps along a chromosome
r.index	a vector of length snps-1 specifying whether a recombination is to be simulated (1) or not (0) in between two adjacent snps.

Value

A data.frame of class `en.masse`. Each row contains the following information: which contains three elements:

- Spore The spore ID (1:length(n.spores))
- Chr The chromosome name

- Snp The snp location (in bps)
- p0 The number of reads that mapped to parent 0
- p1 The number of reads that mapped to parent 1
- states_given The simulated states (0 or 1)

Author(s)

Tyler D. Hether

See Also

[recombine](#), [make_parents](#), [simulate_coverage](#), [recombine_index](#), [id_hotspots](#)

Examples

```
# Simulating 5 haploid recombinants
set.seed(1234567)          # For reproducibility
n_spores <- 5              # number of tetrads (meiosis events)
l <- 75                    # number of snps to simulate
c <- 3.5e-05               # recombination rate between snps (Morgan/bp)
snps <- c(1:l)*1.3e4       # snps are evenly spaced 20kbp apart
p_a <- 0.95                # assignment probability
coverage <- 2.1            # mean coverage
# Now simulate
sim5 <- sim_en_masse(n.spores=n_spores, scale=c, snps=snps,
  p.assign=p_a, mu.rate=0, f.cross=0.8, f.convert=0.3,
  length.conversion=2e3, coverage=coverage)
sim5
```

sim_tetrad

Simulate recombination across a given number of meiosis events

Description

This is a wrapper function of many other functions that simulates a given number of tetrads each with four haploid spores that are recombinants between two parents.

Usage

```
sim_tetrad(n.tetrads, scale, snps, p.assign, mu.rate, f.cross, f.convert,
  length.conversion, coverage, chr.name = "I")
```

Arguments

n.tetrads	An integer specifying the number of tetrads to simulate.
p.assign	a numeric between 0 and 1 (inclusive) that gives the probability of correct sequencing assignment. A value of 1 means that no sequencing error or ancestral polymorphism is to be simulated.
mu.rate	a numeric between 0 and 1 (inclusive) specifying the per snp mutation rate.
f.cross	a numeric between 0 and 1 (inclusive) giving the frequency of recombination events that result in crossing over. This is same as 1 minus the frequency of non-crossovers.

f.convert	a numeric between 0 and 1 (inclusive) that gives the frequency of gene conversion during recombination.
length.conversion	an integer specifying the mean (and variance) of a given gene conversion tract (in bps).
chr.name	a numeric or character value specifying the chromosome name (default to "I")
scale	either vector of length 1 specifying the genome-wide recombination rate (Morgans/bp) or a vector of length snps-1 specifying the recombination rate between all snps. In either case rates must be positive.
snps	a vector giving the locations of snps along a chromosome
r.index	a vector of length snps-1 specifying whether a recombination is to be simulated (1) or not (0) in between two adjacent snps.

Value

A data.frame of class tetrad. Each row contains the following information:

- Tetrad The tetrad ID
- Spore The spore ID (1:4)
- Chr The chromosome name
- Snp The snp location (in bps)
- p0 The number of reads that mapped to parent 0
- p1 The number of reads that mapped to parent 1
- states_given The simulated states (0 or 1)

Author(s)

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See Also

[recombine](#), [make_parents](#), [simulate_coverage](#), [recombine_index](#), [id_hotspots](#)

Examples

```
# Simulating 100 recombination events
set.seed(1234567)          # For reproducibility
n_tetrads <- 100           # number of tetrads (meiosis events)
l <- 50                    # number of snps to simulate
c <- 3e-05                 # recombination rate between snps (Morgan/bp)
snps <- c(1:l)*2e4         # snps are evenly spaced 20kbp apart
p_a <- 0.97                # assignment probability
coverage <- 1              # mean coverage
# Now simulate
sim100 <- sim_tetrad(n.tetrads=n_tetrads, scale=c, snps=snps,
p.assign=p_a, mu.rate=0, f.cross=0.8, f.convert=0.3,
length.conversion=2e3, coverage=coverage)
sim100
```

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