

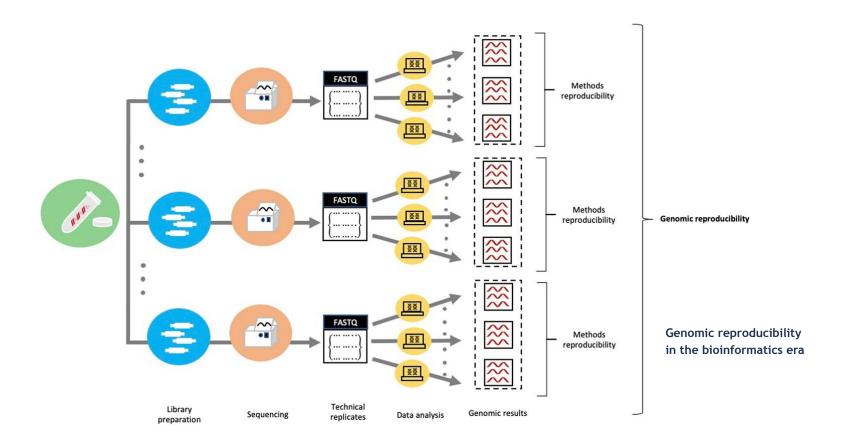
Netflow y automatización de pipeline de análisis, uso de pipeline y reproducibilidad.

Evelin González F. evelyn.gonzalez@uoh.cl

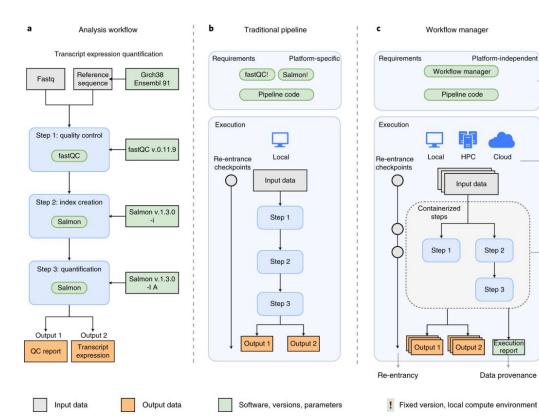
Organización de las clases: Parte 1

- Clase 1: Preprocesamiento de los datos y reportes de calidad.
- Clase 2: Llamado de variantes y visualización con IGV
- Clase 3: Anotación de variantes (uso de bases de datos y clasificación de variantes patogénicas).
- Clase 4: Netflow y automatización de pipeline de análisis, uso de pipeline y reproducibilidad. Taller Práctico.

Reproducibilidad genómica en bioinformática



Flujos de trabajos





Un motor de flujos de trabajo para la automatización y escalabilidad de análisis bioinformáticos.

Principales características:

Portability

Scalability

Automatic resource

management

Integración con **contenedores** (Docker, Singularity).

Escalabilidad en **HPC** y la nube (AWS, Azure, Google Cloud).

Reproducibilidad y portabilidad de pipelines.

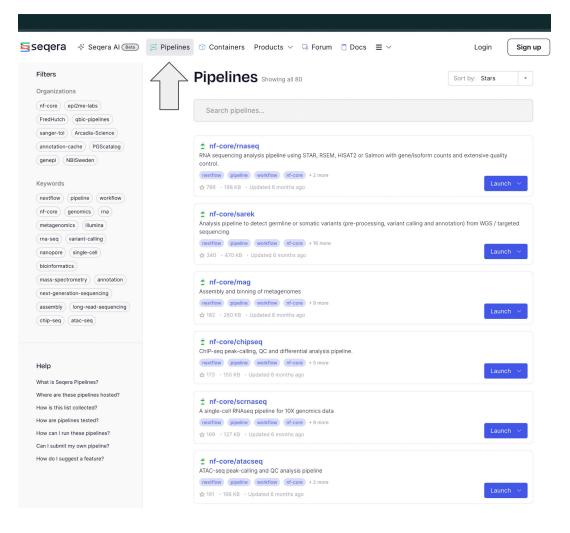
Beneficios en bioinformática:

- Optimización de análisis complejos.
- Trazabilidad y manejo de datos masivos.
- Reducción del tiempo de ejecución mediante paralelización.

Gestores de flujos de trabajo

Tool	Class	Ease of use ^a	Expressiveness ^b	Portability ^c	Scalability ^d	Learning resources ^e	Pipeline initiatives ^f
Galaxy	Graphical	•••	●00	•••	•••	•••	••0
KNIME	Graphical	•••	●00	000	••0	•••	••0
Nextflow	DSL	••0	•••	•••	•••	•••	•••
Snakemake	DSL	••0	•••	•••	•••	••0	•••
GenPipes	DSL	••0	•••	••0	••0	••0	••0
bPipe	DSL	••0	•••	••0	••0	••0	●00
Pachyderm	DSL	••0	•••	●00	••0	•••	000
SciPipe	Library	••0	•••	000	000	••0	000
Luigi	Library	••0	•••	●00	••0	••0	000
Cromwell + WDL	Execution + workflow specification	●00	••0	•••	•••	••0	••0
cwltool + CWL	Execution + workflow specification	●00	••0	•••	000	•••	••0
Toil + CWL/ WDL/Python	Execution + workflow specification	●00	•••	•••	•••	••0	••0

Please refer to Supplementary Table 1 for details. This information is based on online documentation and manuscripts and may not be reflective of the current state of the projects. Scores for Galaxy are based on the graphical user interface. "Ease of use: graphical interface with execution environment (score of 3), programming interface with in-built execution environment (score of 2), separated development and execution environment (score of 1). "Expressiveness: based on an existing programming language (3) or a new language or restricted vocabulary (2), primary interaction with graphical user interface (1). "Portability: integration with three or more container and package manager platforms (3), two platforms are supported (2), one platform is supported (1). "Scalability: considers cloud support, scheduler and orchestration tool integration, and executor support. Please refer to Supplementary Table 1. "Learning resources: official tutorials, forums, and events (3), tutorials and forums (2), tutorials or forums (1). "Pipelines initiatives: community and curated (3), community or curated (1).



Pipelines de análisis

Seqera es una compañía que desarrolla soluciones para la gestión y escalabilidad de flujos de trabajo bioinformáticos, siendo la creadora de **Nextflow**.

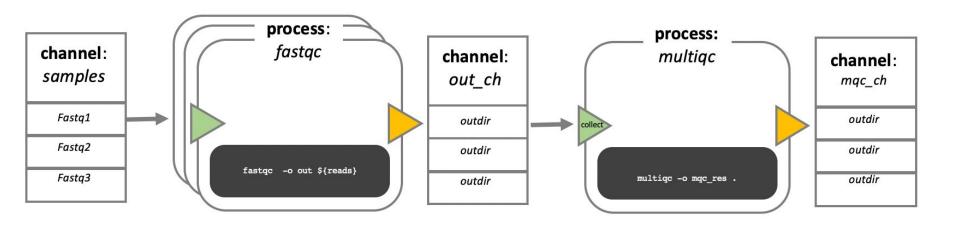
https://segera.io/pipelines/

¿Qué es Nextflow?

- Herramienta para automatizar flujos de trabajo en bioinformática.
- Diseñada para procesar grandes volúmenes de datos.
- Compatible con múltiples entornos: local, clústeres HPC, nube.
- Lenguaje declarativo sencillo, basado en scripts.
- Facilita la reproducibilidad y el control de versiones.
- Ejecuta tareas en paralelo de manera eficiente.

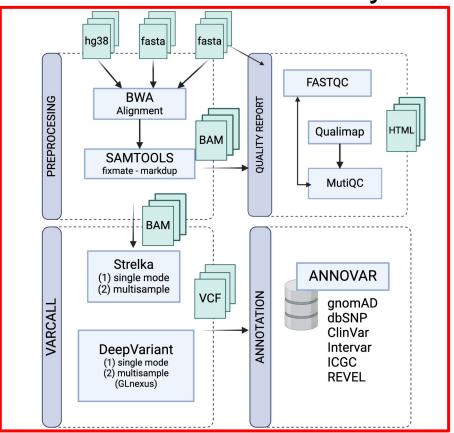


Procesos en Nextflow



Un **proceso** en Nextflow es la **unidad básica de ejecución** dentro de un pipeline. Representa un paso específico de análisis que se ejecuta de manera independiente, con entradas definidas, un script que realiza la tarea y salidas generadas.

Flujo de trabajo bionformático para la detección de variantes en *BRCA1* y *BRCA2*



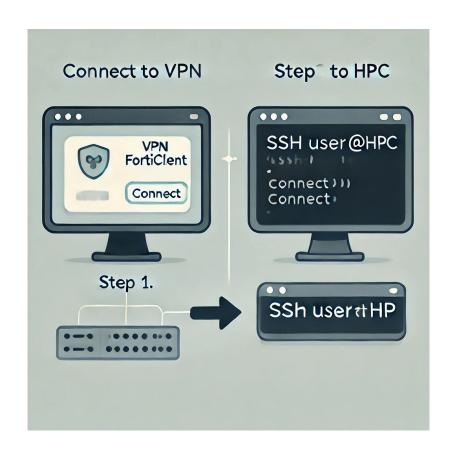
Ejecución de todo el flujo de trabajo de manera automática en un solo paso.

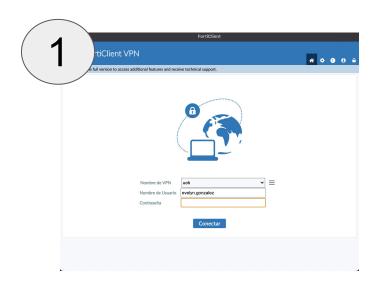
Interpretación de resultados y procesamiento de datos

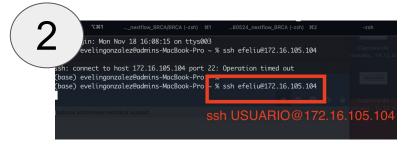
The workflow, including the configurations and tools, is publicly available on the GitHub repository:

https://github.com/digenoma-lab/BRCA.

Primer Paso: Conexión a cluster HPC







```
nextflow run main.nf \
-c nextflow.config \
-profile kutral \
-params-file ../params-brca.yml \
--csv ../readsHRR_1-4.csv \
-resume
```

Ejecutar pipeline en Nextflow de BRCA1

```
(brca12) [efeliu@host04 BRCA]$ nextflow run main.nf -c nextflow.config -profile kutral -params-file ../params-brca.yml --csv ../readsHRR_1-4.csv -resum
curl: (6) Could not resolve host: www.nextflow.io
NEXTFLOW \sim version 23.10.1
Launching `main.nf` [cranky_euclid] DSL2 - revision: 4b9c1c4390
[a4/2c84e0] process > PRINT_VERSIONS
                                       [100%] 1 of 1, cached: 1 🕏
                                       [ 0%] 0 of 3
          ] process > FASTQC
          ] process > BWAMEM
                                        [ 0%] 0 of 3
          1 process > MERGEB
          ] process > SAMTOOLS
          ] process > QUALIMAP
          ] process > B2C
          1 process > STRELKA_ONESAMPLE -
          ] process > STRELKA_POOL
          ] process > BCFT00LS_FILTER
          ] process > BF
          1 process > ANNOVAR_SP
          7 process > ANNOVAR SS
```

```
nextflow run main.nf \
-c nextflow.config \
-profile kutral \
-params-file ../params-brca.yml \
--csv ../readsHRR_1-4.csv \
-resume
```

```
workflow {
   // merge bam files
                                 BODY:
  MERGEB(greads)
                                 Inputs
   //ONT aligment
  MINIMAP2_ALIGN(MERGEB.out.mbam
                  ch ref fasta,
                  'bai',
  NANOPLOT(MINIMAP2_ALIGN.out.bam)
   //germline varcall to log reads, ONT
   CLAIR3(MINIMAP2_ALIGN.out.bam,
          MINIMAP2 ALIGN.out.index,
          ch ref fasta,
           ch ref index)
   LONGPHASE(CLAIR3.out.vcf.
            MINIMAP2 ALIGN.out.bam.
            MINIMAP2 ALIGN.out.index,
            ch_ref_fasta,
            ch_ref_index)
  METHYLARTIST_SEG(LONGPHASE.out.phasebam,
                  LONGPHASE.out.phaseindex,
                  params.bed,
                  ch_ref_fasta,
                  ch_ref_index)
   METHYLARTIST_WGMETH(LONGPHASE.out.phasebam,
                  LONGPHASE.out.phaseindex,
                  ch_ref_fasta,
                  ch_ref_index)
```

nextfow run main.nf

```
#!/usr/bin/env nextflow
nextflow.enable.dsl = 2
                                                 HEAD:
//loading scripts from modules
                                                 MÓDULOS
include {MERGEB} from './modules/mergeb'
include {MINIMAP2 ALIGN} from './modules/minimap2'
include {CLAIR3} from './modules/clair3'
include {NANOPLOT} from './modules/nanoplot'
include {LONGPHASE} from './modules/longphase'
include {METHYLARTIST SEG} from './modules/methylartist'
include {METHYLARTIST_WGMETH} from './modules/methylartist_wgmeth'
process PRINT_VERSIONS {
   publishDir "$params.outdir/software", mode: "copy"
   output:
   path("versions.txt")
   echo "samtools: 1.19" > versions.txt
   echo "minimap2: 2.28-r1209" >> versions.txt
   echo "clair3: 1.0.10" >> versions.txt
   echo "nanoplot: 1.41.6" > versions.txt
   echo "longphase: 1.5.1" >> versions.txt
   echo "methylartist: 1.3.0" >> versions.txt
```

```
nextflow run main.nf \
-c nextflow.config \
-profile kutral \
-params-file ../params-brca.yml \
--csv ../readsHRR_1-4.csv \
-resume
```

-c nextflow.config -profile kutral

```
process {
    withName: 'PRINT_VERSIONS()'{
    cpus = 1
    memory = '1 GB'
}
    withName: 'MERGEB' {
        cpus = 10
        memory = '20 GB'
}
    withName: 'MINIMAP2_ALIGN' {
        cpus = 20
    memory = '40 GB'
}
```

```
kutral {
    singularity.enabled = true
    singularity.autoMounts = true
    docker.enabled = false
    podman.enabled = false
    shifter.enabled = false
    charliecloud.enabled = false
    executor.queueSize = 8
    process.executor = 'slurm'
    process.queue = 'uohhm'
    }
}
```

```
params {
  debug = false
}
```

-c nextflow.config:

Especifica un archivo de configuración adicional, en este caso nextflow.config. Este archivo puede contener configuraciones como:

- Recursos (CPU, memoria).
- Parámetros globales del pipeline.
- Configuraciones específicas para perfiles y entornos.

```
nextflow run main.nf \
-c nextflow.config \
-profile kutral \
-params-file ../params-brca.yml \
--csv ../readsHRR_1-4.csv \
-resume
```

-params-file [ARCHIVO_PARAMETROS.YML]

```
1 ref: /mnt/beegfs/labs/DiGenomaLab/Prunus_dulcis/references/rosaceae/GCA_021292205.2_OSU_Pdul_2.5_genomic.fa
2 minimap2: "-ax map-ont"
3 outdir: ont_result
4 aln_only: true
5 bed: /mnt/beegfs/home/efeliu/work2024/120924_Nextflow_meth/protein_coding_prunusDulcis.bed
6
```

-params-file:

Permite cargar un archivo de parámetros en formato YAML para configurar el pipeline. Este archivo contiene valores específicos para las variables que se utilizan en el pipeline, facilitando la personalización sin modificar el código.

```
nextflow run main.nf \
-c nextflow.config \
-profile kutral \
-params-file ../params-brca.yml \
--csv ../readsHRR_1-4.csv \
-resume
```

-csv [ARCHIVO_ENTRADA]

```
Una linea por muestra (n=3)

sampleId,part,read1,read2

AL 0,/mnt/beegfs/labs/DiGenomaLab/HRR/reads/AL_S9.R1.fastq.gz /mnt/beegfs/labs/DiGenomaLab/HRR/reads/DC_S14.R1.fastq.gz,/mnt/beegfs/labs/DiGenomaLab/HRR/reads/DC_S14.R2.fastq.gz

JC,0,/mnt/beegfs/labs/DiGenomaLab/HRR/reads/JC_S13.R1.fastq.gz,/mnt/beegfs/labs/DiGenomaLab/HRR/reads/JC_S13.R2.fastq.gz

JC,0,/mnt/beegfs/labs/DiGenomaLab/HRR/reads/JC_S13.R1.fastq.gz,/mnt/beegfs/labs/DiGenomaLab/HRR/reads/JC_S13.R2.fastq.gz
```

Archivo de entrada

```
nextflow run main.nf \
-c nextflow.config \
-profile kutral \
-params-file ../params-brca.yml \
--csv ../readsHRR_1-4.csv \
```

-resume

-resume

```
NEXTFLOW
                     version 24.04.4
Launching `main.nf` [condescending_heisenberg] DSL2 - revision: 5080d20c78
  7/871c81] process > PRINT_VERSIONS
                                          [100%] 1 of 1, cached: 1 ✓
   /871c81] process > PRINT_VERSIONS
                                                  [100%] 1 of 1, cached: 1 ✓
         17 process > PRINT_VERSIONS
                                                  [100%] 1 of 1, cached: 1 ✓
          process > MERGEB (CHI01-merge)
                                                  [100%] 2 of 2, cached: 2 ✓
          process > MINIMAP2_ALIGN (CHI01)
                                                  [100%] 2 of 2, cached: 2 ✓
  /977a7a7 process > NANOPLOT (B0872)
                                                  [100%] 2 of 2, cached: 2 ✓
  /92f393] process > CLAIR3 (B0872)
                                                  [100%] 2 of 2, cached: 2 /
  /39c3d7] process > LONGPHASE (CHI01)
                                                  [100%] 2 of 2, cached: 2 /
 3/aae843] process > METHYLARTIST_WGMETH (B0872) [100%] 2 of 2, cached: 2 ✓
[B0872, [1, 2], [/home/adigenova/methylation-Chilean/bams/B0872.PAQ20471.fail.mod.bam, /home/adigenova/methylation-Chilean/bams/B0872.PAQ20471.pass.mod.bam]]
[CHI01, [1, 2, 3], [/home/adigenova/methylation-Chilean/bams/CHI01.PAQ20767.fail.mod.bam, /home/adigenova/methylation-Chilean/bams/CHI01.PAQ20767.pass.mod.bam, /home/adigenova/methylation
HI01.PA020767.rep.pass_fail.mod.bam]]
```

-resume:

Indica que Nextflow debe **reutilizar los resultados previos** de ejecuciones anteriores. Esto es útil si la ejecución se interrumpe o si se desean realizar pequeños ajustes sin reanalizar todos los pasos.Nextflow verifica los pasos ya completados y reanuda desde donde se quedó, ahorrando tiempo y recursos.

Archivos de salida: BRCA pipeline

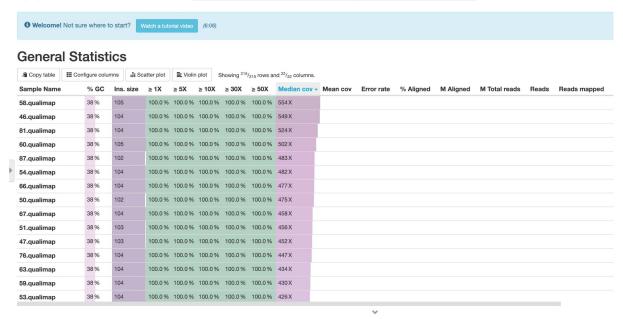
- Reporte de calidad: archivo HTML con parámetros de calidad. MultiQC.
- Archivos BAM (uno para cada muestra)
- Archivos VCF (uno para cada muestra)
- Anotación de variantes ANNOVAR (uno para cada muestra)

QC, datos de calidad



A modular tool to aggregate results from bioinformatics analyses across many samples into a single report.

Report generated on 2024-11-11, 11:30 -03 based on data in: /mnt/beegfs/home/efeliu/work2024/080524_nextflow_BRCA/Runs_nextflow_BRCA/HHR_parte2



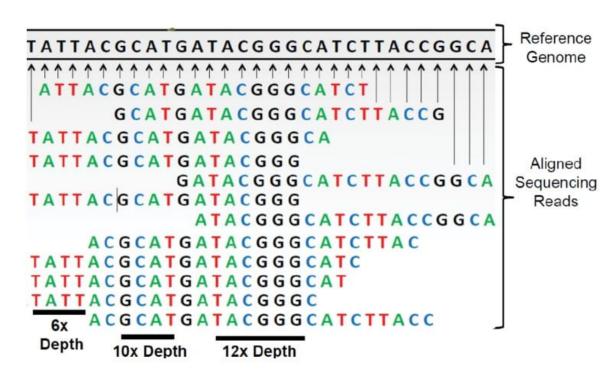
FastQC Qualimap MultiQC

QualiMap

QualiMap is a platform-independent application to facilitate the quality control of alignment sequencing data and its derivatives like feature counts. DOI: 10.1093/bioinformatics/btv566; 10.1093/bioinformatics/bts503.

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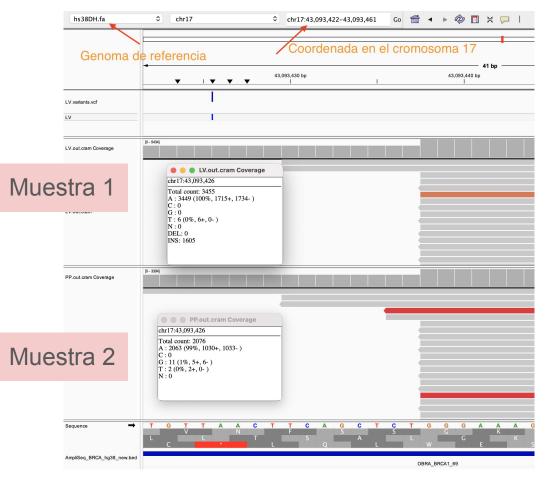
Archivos BAM: Lecturas alineadas al genoma



BAM: Formato binario para el almacenamiento de datos de secuenciación.

La extensión de archivo (.bam) contiene información sobre lecturas de secuencias después de haber sido estas alineadas contra un genoma de referencia.

Utilizado para almacenar datos de secuenciación masiva.



Ejemplo: Variante frameshift insertion p.L655Ffs*10 BRCA1

Inspección visual de las lecturas alineadas al genoma de referencia de dos pacientes.

Variant Call Format: VCF

```
VCF example
 ##fileformat=VCFv4.1
 ##fileDate=20110413
  ##source=VCFtools
  ##reference=file:///refs/human NCBI36.fasta
 ##contig=<ID=1,length=249250621,md5=1b22b98cdeb4a9304cb5d48026a85128,species="Homo Sapiens">
  ##contig=<ID=X,length=155270560,md5=7e0e2e580297b7764e31dbc80c2540dd,species="Homo Sapiens">
  ##INFO=<ID=AA, Number=1, Type=String, Description="Ancestral Allele">
  ##INFO=<ID=H2, Number=0, Type=Flag, Description="HapMap2 membership">
 ##FORMAT=<ID=GT, Number=1, Type=String, Description="Genotype">
 ##FORMAT=<ID=G0.Number=1.Type=Integer.Description="Genotype Quality">
  ##FORMAT=<ID=DP, Number=1, Type=Integer, Description="Read Depth">
  ##ALT=<ID=DEL,Description="Deletion">
  ##INFO=<ID=SVTYPE, Number=1, Type=String, Description="Type of structural variant">
  ##INFO=<ID=END, Number=1, Type=Integer, Description="End position of the variant">
 #CHROM POS ID
                    REF
                                                                               SAMPLE1
                                QUAL FILTER
                                             INFO
                                                                    FORMAT
                                                                                        SAMPLE2
                         A.AT
                                  40 PASS
                                                                    GT:DP
                                                                               1/1:13
                                                                                        2/2:29
                         T,CT
                                      PASS
                                              H2:AA=T
                                                                    GT
                                                                               0|1
                                                                                        2/2
                                                                    GT:DP
                                                                                        2/2:20
                                     PASS
                                                                               1|0:16
                         <DEL>
                                      PASS
                                              SVTYPE=DEL; END=299
                                                                               1:12:.
                                                                                        0/0:20:36
         100
                                                                    GT:GQ:DP
```

B SNP			C Insert	D Dele	tion		E Replacement							
Alignment 1234 ACGT ATGT	100000000000000000000000000000000000000		sentation ALT T	12345 AC-GT ACTGT	POS 2	C	ALT CT	1234 ACGT AT	POS 1	REF	1234 ACGT A-TT	POS 1	REF ACG	

100 results per page

back to HOME

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Chr	Start	End	Ref	Alt	Func	Gene	GeneDetail	ExonicFunc	AAChange
-112	22222242	22222242				DDCAO		CANV	DDCA2-NH 0000F0 Assessment ACCFCs at N200H
chr13	32332343	32332343	Α	С	exonic	BRCA2		nonsynonymous SNV	BRCA2:NM_000059.4:exon10:c.A865C:p.N289H
chr13	32332592	32332592	Α	С	exonic	BRCA2		nonsynonymous SNV	BRCA2:NM_000059.4:exon10:c.A1114C:p.N372H
chr13	32332843	32332843	Α	G	exonic	BRCA2		synonymous SNV	BRCA2:NM_000059.4:exon10:c.A1365G:p.S455S
chr13	32336584	32336584	T	С	exonic	BRCA2		synonymous SNV	BRCA2:NM_000059.4:exon11:c.T2229C:p.H743H
chr13	32336744	32336744	Α	С	exonic	BRCA2		nonsynonymous SNV	BRCA2:NM_000059.4:exon11:c.A2389C:p.K797Q
chr13	32337326	32337326	Α	G	exonic	BRCA2		nonsynonymous SNV	BRCA2:NM_000059.4:exon11:c.A2971G:p.N991D
chr13	32337751	32337751	Α	G	exonic	BRCA2		synonymous SNV	BRCA2:NM_000059.4:exon11:c.A3396G:p.K1132K
chr13	32338162	32338162	T	С	exonic	BRCA2		synonymous SNV	BRCA2:NM_000059.4:exon11:c.T3807C:p.V1269V
chr13	32338918	32338918	Α	G	exonic	BRCA2		synonymous SNV	BRCA2:NM_000059.4:exon11:c.A4563G:p.L1521L
chr13	32340099	32340099	С	Т	exonic	BRCA2		nonsynonymous SNV	BRCA2:NM_000059.4:exon11:c.C5744T:p.T1915M
chr13	32340678	32340678	G	Α	exonic	BRCA2		nonsynonymous SNV	BRCA2:NM_000059.4:exon11:c.G6323A:p.R2108H
chr13	32340868	32340868	G	С	exonic	BRCA2		synonymous SNV	BRCA2:NM_000059.4:exon11:c.G6513C:p.V2171V
chr13	32355095	32355095	Α	G	exonic	BRCA2		synonymous SNV	BRCA2:NM_000059.4:exon14:c.A7242G:p.S2414S
chr13	32355250	32355250	Т	С	exonic	BRCA2		nonsynonymous SNV	BRCA2:NM_000059.4:exon14:c.T7397C:p.V2466A
chr13	32356461	32356461	Т	С	exonic	BRCA2		nonsynonymous SNV	BRCA2:NM_000059.4:exon15:c.T7469C:p.I2490T
chr13	32379413	32379413	G	Α	exonic	BRCA2		nonsynonymous SNV	BRCA2:NM_000059.4:exon22:c.G8851A:p.A2951T
chr17	43071077	43071077	T	С	exonic	BRCA1		nonsynonymous SNV	BRCA1:NM_007297.4:exon14:c.A4696G:p.S1566G,E
chr17	43082453	43082453	Α	G	exonic	BRCA1		synonymous SNV	BRCA1:NM_007297.4:exon11:c.T4167C:p.S1389S,E
chr17	43091983	43091983	T	С	exonic	BRCA1		nonsynonymous SNV	BRCA1:NM_007297.4:exon9:c.A3407G:p.K1136R,BF
chr17	43092412	43092412	С	Т	exonic	BRCA1		nonsynonymous SNV	BRCA1:NM_007297.4:exon9:c.G2978A:p.S993N,BRC
chr17	43092418	43092418	Т	С	exonic	BRCA1		nonsynonymous SNV	BRCA1:NM_007297.4:exon9:c.A2972G:p.E991G,BRC
chr17	43092919	43092919	G	Α	exonic	BRCA1		nonsynonymous SNV	BRCA1:NM_007297.4:exon9:c.C2471T:p.P824L,BRC
chr17	43093220	43093220	Α	G	exonic	BRCA1		synonymous SNV	BRCA1:NM_007297.4:exon9:c.T2170C:p.L724L,BRC
chr17	43093425	43093425	-	Α	exonic	BRCA1		frameshift insertion	BRCA1:NM_007297.4:exon9:c.1964dupT:p.L655Ffs
chr17	43093449	43093449	G	Α	exonic	BRCA1		synonymous SNV	BRCA1:NM_007297.4:exon9:c.C1941T:p.S647S,BRC
chr17	43093454	43093454	С	Т	exonic	BRCA1		nonsynonymous SNV	BRCA1:NM_007297.4:exon9:c.G1936A:p.D646N,BRC

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rage.						
Sort by:	~	Buscar variantes "Pathogenic" en CLINVAR				
Filter by:						
CLNALLELEID:	CLNDN: = V	CLNSIG: = vathogenic				
SIFT_pred:	SIFT4G_pred:	Polyphen2_HVAR_score: V				
LRT_pred:	MutationTaster_pred:	FATHMM_score:				
PROVEAN_pred: V	MetaSVM_score:	MetaLR_score:				
MetaLR_pred:	Reliability_index:					
Chr:						
Start: V						
End:						

VCF anotados con ANNOVAR

Filtrado de Variantes para la Identificación de:

- Variantes Patogénicas
- VUS (Variantes de Significado Incierto)
- Variantes Novel
- Variantes Benignas

Repositorio del pipeline BRCA en GitHub



