Population differentiation determined from putative adaptive SNP markers in Eulachon

Evelyn Takyi May 8, 2017

Requried packages

treemap, readr, devtools, gstudio, pegas, ape, ade4, adegenet, diveRsity, hierfstat, poppr, readxl

Importing data into R.

```
mon <- read.genepop("EulachonSig.gen")

##
## Converting data from a Genepop .gen file to a genind object...
##
##
## File description: Eulachon significant loci
## Warning in read.genepop("EulachonSig.gen"): Duplicate individual names
## detected. Coercing them to be unique.
##
## ...done.</pre>
```

summary of imported data.

Names of the 12 populations of samples used in analysis

```
popNames(mon)
## [1] "SS08" "BELCOLO3" "COW02" "CR12" "FRAS09" "KC02"
## [7] "KEM01" "KEN04" "KLK02" "SKE10" "STIK06" "TMR01"
```

convert dataset into genind objects for each population

```
pop_label <- c(rep("SS08",41),rep("BELCOL03",33),</pre>
               rep("COW02",37),rep("CR12",22),rep("FRAS09",40),rep("KC02",36),
               rep("KEM01",42),rep("KEN04",71),rep("KLK02",41),rep("SKE10",33),rep("STIK06",66),rep("TM
#. Creates a list of genind objects for each population
pops separat <- seppop(mon,pop=pop label)</pre>
pops_separat$COW02
## /// GENIND OBJECT ///////
##
   // 37 individuals; 193 loci; 386 alleles; size: 155.5 Kb
##
##
  // Basic content
##
##
      Otab: 37 x 386 matrix of allele counts
##
      @loc.n.all: number of alleles per locus (range: 2-2)
      @loc.fac: locus factor for the 386 columns of @tab
##
      @all.names: list of allele names for each locus
##
      Oploidy: ploidy of each individual (range: 2-2)
##
      @type: codom
##
##
      @call: .local(x = x, i = i, j = j, treatOther = ...1, quiet = ...2, drop = drop)
##
## // Optional content
      @pop: population of each individual (group size range: 37-37)
##
```

The mean observed and expected heterozygosity

```
#. Compute the mean observed and expected heterzygosity for each population over all loci
summaryCOW02 <- summary(pops_separat$COW02)
mean(summaryCOW02$Hobs)

## [1] 0.2239624
mean(summaryCOW02$Hexp)

## [1] 0.2246354
```

compute allelic richness for each population

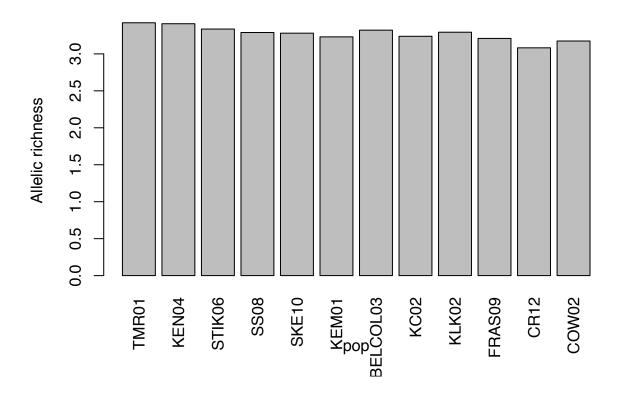
```
## 2
         KEN04
                         0.3921
                                       3.408
## 3
        STIK06
                         0.4058
                                       3.336
                         0.4056
## 4
          SS08
                                       3.289
         SKE10
## 5
                         0.4103
                                       3.280
## 6
         KEM01
                         0.4064
                                       3.230
## 7
      BELCOL03
                         0.4101
                                       3.321
## 8
          KC02
                         0.4132
                                       3.238
         KLK02
                         0.4078
                                       3.293
## 9
## 10
        FRAS09
                         0.4197
                                       3.209
                         0.4251
                                       3.082
## 11
          CR12
## 12
         COW02
                         0.4181
                                       3.174
```

\$FIS

[1] 0.01277384

#. The mean number of alleles (Ar) were plotted to determine the genetic diversity in each population. barplot(heteroz\$Allelicrich, main="plot of the mean number of alleles vs population", ylab="Allelic rich"

plot of the mean number of alleles vs population



Test for genetic differentiation between pairs of population

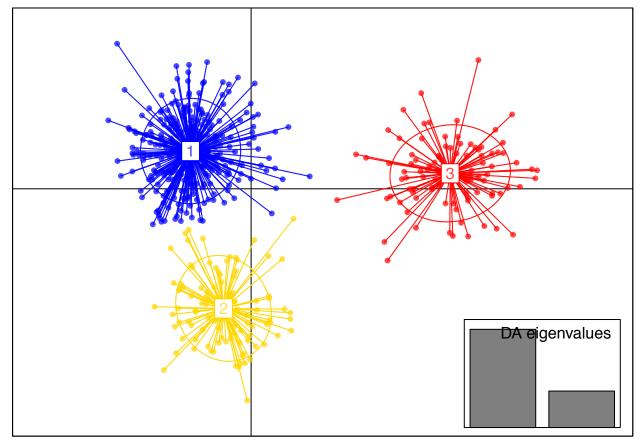
```
#. Population structure was determined using the test for genetic differentiation between pairs of population
wc(mon)
## $FST
## [1] 0.05218927
##
```

```
GF <- genet.dist(mon, method = "WC84")
##
                   SS08
                           BELCOL03
                                           COW02
                                                        CR12
                                                                  FRAS09
## BELCOLO3 0.010964926
## COW02
            0.050284725 0.051959708
## CR12
            0.076474711 0.084113123 0.031979660
## FRASO9
            0.051107361 0.048257454 0.013656907 0.048652160
            0.020033081\ 0.009531729\ 0.049303419\ 0.089151531\ 0.040137025
## KC02
## KEMO1
            0.003651866 0.011597369 0.053200231 0.081269978 0.048218065
## KENO4
            0.080730771 0.078461326 0.083510072 0.105055990 0.095159708
## KLK02
            0.012910011 0.007515463 0.049278960 0.080704024 0.043720618
            0.012045033 0.006881454 0.051292325 0.084330945 0.048515859
## SKE10
            0.011920935 0.008753050 0.047096889 0.081670619 0.052005081
## STIK06
## TMRO1
            0.095934381 0.091324233 0.096674264 0.123364057 0.103815008
                   KC02
##
                              KEM01
                                           KEN04
                                                       KLK02
                                                                   SKE10
## BELCOLO3
## COW02
## CR12
## FRAS09
## KC02
## KEMO1
            0.014783901
## KEN04
            0.091851764 0.074487167
## KLK02
            0.004208213 0.008749633 0.086386533
## SKE10
            0.015758806 0.008320082 0.076885941 0.011709553
## STIK06
            0.018175434\ 0.009495093\ 0.074535505\ 0.010840276\ 0.005994568
## TMRO1
            0.105964773 0.091485898 0.005083938 0.101830392 0.091415594
                 STIK06
##
## BELCOLO3
## COW02
## CR12
## FRAS09
## KC02
## KEMO1
## KEN04
## KLK02
## SKE10
## STIK06
## TMRO1
            0.088565356
```

Discriminant Analysis of Principal Components(DAPC)

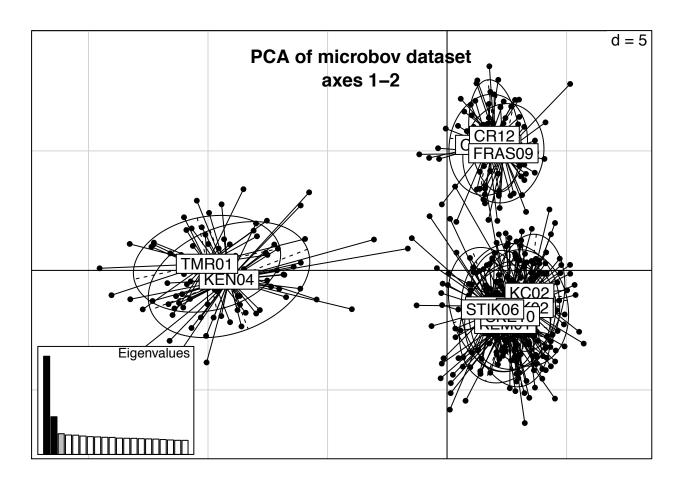
```
#. using Kmeans and DAPC in adegenet
#. Region membership for each group of sample was determined by DAPC conducted in R with the package A
set.seed(20160308) # Setting a seed for a consistent result
grp <- find.clusters(mon, max.n.clust = 10, n.pca = 20, choose.n.clust = FALSE)
## The "ward" method has been renamed to "ward.D"; note new "ward.D2"
names(grp)
## [1] "Kstat" "stat" "grp" "size"</pre>
```

```
## 469 470 471 472 473 474 475 476 477 478 479 480 481 482 483 484 485 486
                           3
                                   3
                                     3
                                           3
                                                  3
                                                      3
       3
           3 3
                  3
                        3
                               3
                                              3
                                                              3
                                                                  3
## 487 488 489 490 491 492 493 494
##
        3
           3
                3
                    3
                        3
## Levels: 1 2 3
dapc1 <- dapc(mon, grp$grp, n.pca = 20, n.da = 6)</pre>
scatter(dapc1) # plot of the group
```



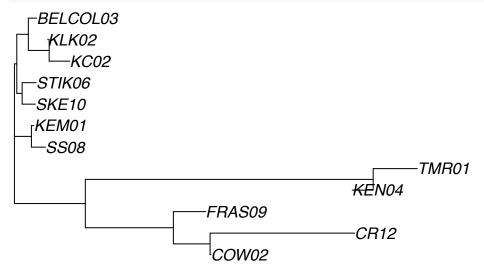
structure of the various groups with population sample names

```
#This plane is to show the structure of the various groups of population
s.class(pca1$li, pop(mon))
title("PCA of microbov dataset\naxes 1-2")
add.scatter.eig(pca1$eig[1:20], 3,1,2)
```



${\bf Neighbour joining\ Tree}$

#A neighbour-joining tree ploted to visualize the distances for each of these loci sets based on the ditree <- nj(GF)
plot.phylo(tree)



Reference

publication

Candy JR, Campbell NR, Grinnell MH, Beacham TD, Larson WA, Narum SR (2015) Population differentiation determined from putative neutral and divergent adaptive genetic markers in Eulachon (Thaleichthys pacificus, Osmeridae), an anadromous Pacific smelt. Molecular Ecology Resources 15(6): 1421-1434. http://dx.doi.org/10.1111/1755-0998.12400

Data obtained from Dryad

Candy JR, Campbell NR, Beacham TD, Grinnell MH, Narum SR, Larson WA (2015) Data from: Population differentiation determined from putative neutral and divergent adaptive genetic markers in Eulachon (Thaleichthys pacificus, Osmeridae), an anadromous Pacific smelt. Dryad Digital Repository. http://dx.doi.org/10.5061/dryad.1797v