# COVID-19 subject UPHS-1173

2021-06-23

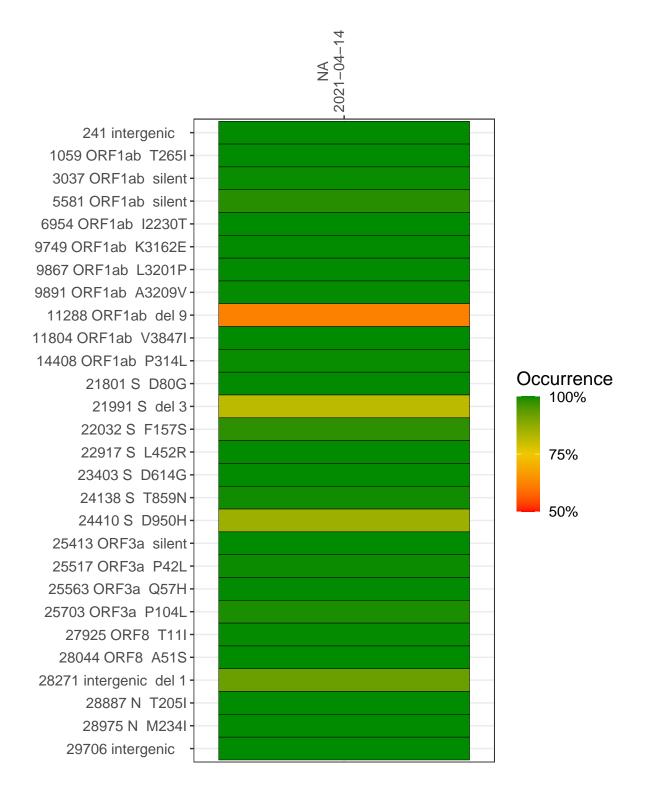
The table below provides a summary of subject samples for which sequencing data is available. The experiments column shows the number of sequencing experiments performed for each specimen. Experiment specific analyses are shown at the end of this report. Lineages are called with the Pangolin software tool (Rambaut et al 2020) for genomes with > 90% sequence coverage.

Table 1. Sample summary.

| Experiment | Туре              | Genomes | Sample type | Sample date | Largest contig<br>(KD) | Lineage | Reference read<br>coverage | Reference read coverage (>= 5 reads) |
|------------|-------------------|---------|-------------|-------------|------------------------|---------|----------------------------|--------------------------------------|
| VSP2430-1  | single experiment | NA      | NA          | 2021-04-14  | 29.88                  | B.1.526 | 99.8%                      | 99.7%                                |

#### Variants shared across samples

The heat map below shows how variants (reference genome /home/common/SARS-CoV-2-Philadelphia/Wuhan-Hu-1) are shared across subject samples where the percent variance is colored. Variants are called if a variant position is covered by 5 or more reads, the alternative base is found in > 50% of read pairs and the variant yields a PHRED score > 20. Gray tiles denote positions where the variant was not the major variant or no variants were found. The relative base compositions of each experiment used to calculate tiles are shown in the following plot where the total number of position reads are shown atop of each plot.



#### NA 2021-04-14

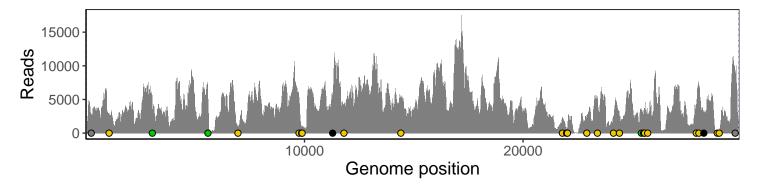
|                        | 2021-04-14  |
|------------------------|-------------|
| 241 intergenic         | 2400        |
| 1059 ORF1ab T265I      | 2725        |
| 3037 ORF1ab silent     | 3691        |
| 5581 ORF1ab silent     | 5042        |
| 6954 ORF1ab I2230T     | 740         |
| 9749 ORF1ab K3162E     | 5348        |
| 9867 ORF1ab L3201P     | 725         |
| 9891 ORF1ab A3209V     | 897         |
| 11288 ORF1ab del 9     | 3203        |
| 11804 ORF1ab V3847I    | 4070        |
| 14408 ORF1ab P314L     | 4520        |
| 21801 S D80G           | 2293        |
| 21991 S del 3          | 525         |
| 22032 S F157S          | <b>76</b> 9 |
| 22917 S L452R          | 2548        |
| 23403 S D614G          | 5191        |
| 24138 S T859N          | 2507        |
| 24410 S D950H          | 2715        |
| 25413 ORF3a silent     | 3553        |
| 25517 ORF3a P42L       | 2596        |
| 25563 ORF3a Q57H       | 3545        |
| 25703 ORF3a P104L      | 2683        |
| 27925 ORF8 T11I        | 4281        |
| 28044 ORF8 A51S        | 6983        |
| 28271 intergenic del 1 | 3093        |
| 28887 N T205I          | 374         |
| 28975 N M234I          | 338         |
| 29706 intergenic       | 8784        |
|                        | VSP2430-1   |
|                        |             |



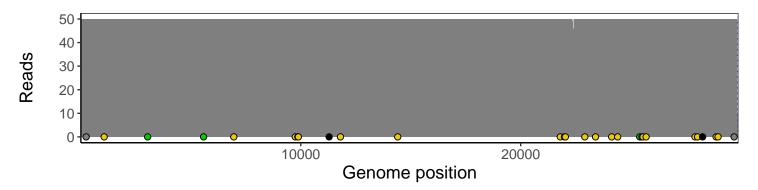
### Analyses of individual experiments and composite results

#### VSP2430-1 | 2021-04-14 | NA | UPHS-1173 | genomes | single experiment

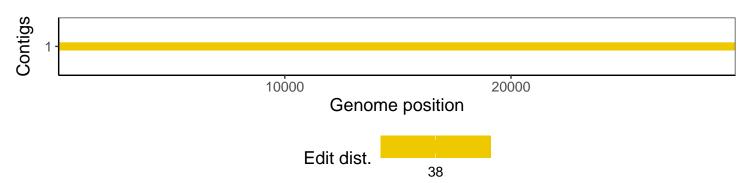
The plot below shows the number of reads covering each nucleotide position in the reference genome. Variants are shown as colored dots along the bottom of the plot and are color coded according by variant types: gray - transgenic, green - silent, gold - missense, red - nonsense, black - indel.



Excerpt from plot above focusing on reads coverage from 0 to 50 NT.



The longest five assembled contigs are shown below colored by their edit distance to the reference genome.



## Software environment

| Software/R package            | Version  |  |  |  |  |
|-------------------------------|--|--|--|--|--|
| R                             | 3.4.0  |  |  |  |  |
| bwa                           | 0.7.17-r1198-dirty                                       |  |  |  |  |
| samtools                      | 1.10 Using htslib 1.10                                   |  |  |  |  |
| bcftools                      | 1.10.2-34-g1a12af0-dirty Using htslib 1.10.2-57-gf58a6f3 |  |  |  |  |
| pangolin                      | 3.1.3  |  |  |  |  |
| genbankr                      | 1.4.0  |  |  |  |  |
| optparse                      | 1.6.0  |  |  |  |  |
| forcats                       | 0.3.0  |  |  |  |  |
| stringr                       | 1.4.0  |  |  |  |  |
| dplyr                         | 0.8.1  |  |  |  |  |
| purrr                         | 0.2.5  |  |  |  |  |
| readr                         | 1.1.1  |  |  |  |  |
| tidyr                         | 0.8.1  |  |  |  |  |
| tibble                        | 2.1.2  |  |  |  |  |
| ggplot2                       | 3.3.3  |  |  |  |  |
| tidyverse                     | 1.2.1  |  |  |  |  |
| ShortRead                     | 1.34.2   |  |  |  |  |
| GenomicAlignments             | 1.12.2   |  |  |  |  |
| ${\bf Summarized Experiment}$ | 1.6.5  |  |  |  |  |
| DelayedArray                  | 0.2.7  |  |  |  |  |
| matrixStats                   | 0.54.0   |  |  |  |  |
| Biobase                       | 2.36.2   |  |  |  |  |
| Rsamtools                     | 1.28.0   |  |  |  |  |
| GenomicRanges                 | 1.28.6   |  |  |  |  |
| GenomeInfoDb                  | 1.12.3   |  |  |  |  |
| Biostrings                    | 2.44.2   |  |  |  |  |
| XVector                       | 0.16.0   |  |  |  |  |
| IRanges                       | 2.10.5   |  |  |  |  |
| S4Vectors                     | 0.14.7   |  |  |  |  |
| BiocParallel                  | 1.10.1   |  |  |  |  |
| BiocGenerics                  | 0.22.1   |  |  |  |  |