COVID-19 subject PQ-Seq3

2021-05-05

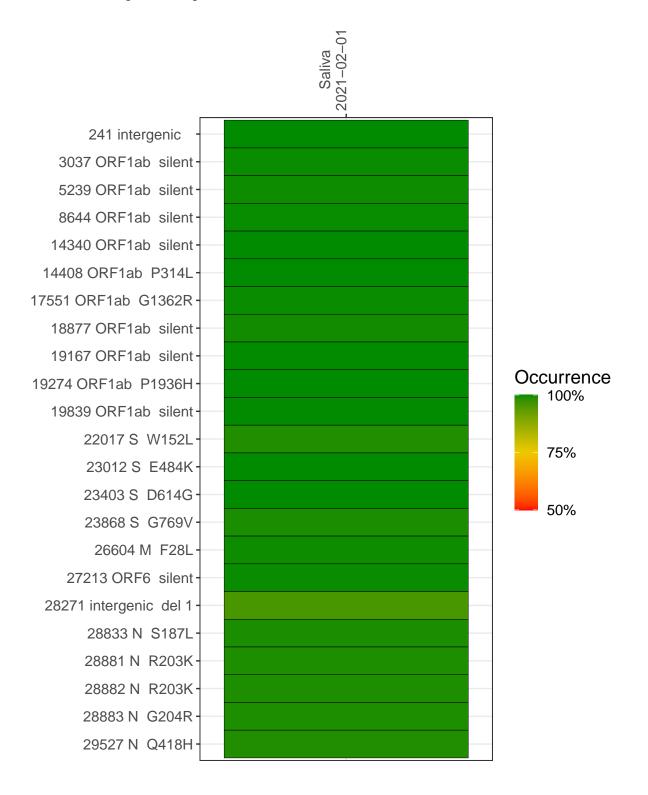
The table below provides a summary of subject samples for which sequencing data is available. The experiments column shows the number of sequencing experiments performed for each specimen. Experiment specific analyses are shown at the end of this report. Lineages are called with the Pangolin software tool (Rambaut et al 2020) for genomes with > 90% sequence coverage.

Table 1. Sample summary.

Experiment	Туре	Genomes	Sample type	Sample date	Largest contig (KD)	Lineage	Reference read coverage	Reference read coverage (>= 5 reads)
VSP0772	composite	NA	Saliva	2021-02-01	21.71	R.1	99.5%	99.1%
VSP0772-1	single experiment	NA	Saliva	2021-02-01	21.71	R.1	99.4%	99.1%
VSP0772-2	single experiment	NA	Saliva	2021-02-01	3.69	NA	93.7%	71.6%

Variants shared across samples

The heat map below shows how variants (reference genome /home/everett/projects/SARS-CoV-2-Philadelphia/Wuhan-Hu-1) are shared across subject samples where the percent variance is colored. Variants are called if a variant position is covered by 5 or more reads, the alternative base is found in > 50% of read pairs and the variant yields a PHRED score > 20. Gray tiles denote positions where the variant was not the major variant or no variants were found. The relative base compositions of each experiment used to calculate tiles are shown in the following plot where the total number of position reads are shown atop of each plot.



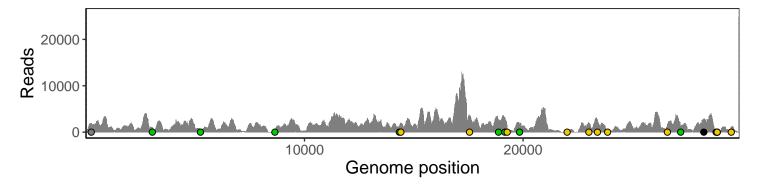
Saliva 2021-02-01

	2021-		
241 intergenic	1862	7	
3037 ORF1ab silent	955	4	
5239 ORF1ab silent	546	12	
8644 ORF1ab silent	536	3	
14340 ORF1ab silent	1406	2	
14408 ORF1ab P314L	1989	2	
17551 ORF1ab G1362R	3576	12	
18877 ORF1ab silent	2622	24	
19167 ORF1ab silent	3083	17	
19274 ORF1ab P1936H	1491	43	Base change Expected
19839 ORF1ab silent	1801	5	A T
22017 S W152L	166	4	С
23012 S E484K	1903	20	G N
23403 S D614G	2743	32	Ins/Del No data
23868 S G769V	243	2	
26604 M F28L	1624	11	
27213 ORF6 silent	1318	15	
28271 intergenic del 1	2871	26	
28833 N S187L	242	13	
28881 N R203K	200	10	
28882 N R203K	200	10	
28883 N G204R	200	10	
29527 N Q418H	1077	6	
	VSP0772-1	VSP0772-2	
	-	-	

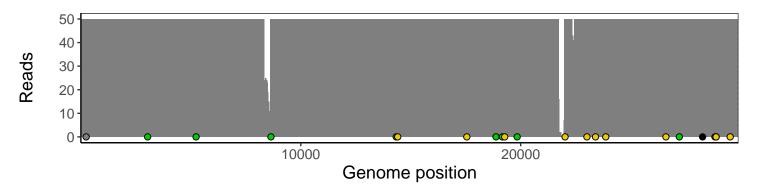
Analyses of individual experiments and composite results

$VSP0772 \mid 2021-02-01 \mid Saliva \mid PQ-Seq3 \mid composite result$

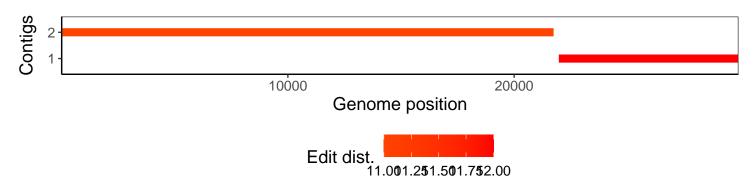
The plot below shows the number of reads covering each nucleotide position in the reference genome. Variants are shown as colored dots along the bottom of the plot and are color coded according by variant types: gray - transgenic, green - silent, gold - missense, red - nonsense, black - indel.



Excerpt from plot above focusing on reads coverage from 0 to 50 NT.

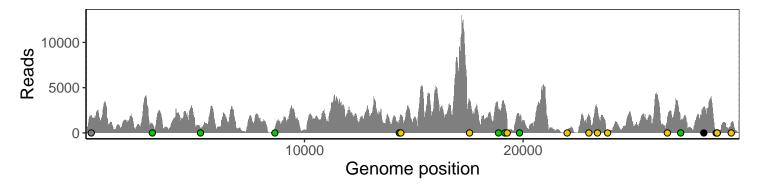


The longest five assembled contigs are shown below colored by their edit distance to the reference genome.

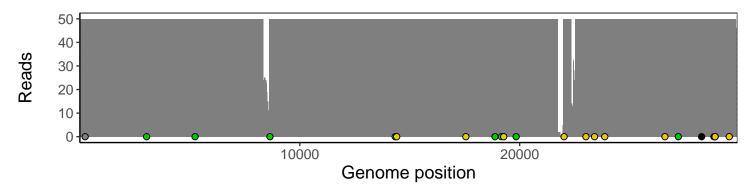


VSP0772-1 | 2021-02-01 | Saliva | PQ-Seq3 | genomes | single experiment

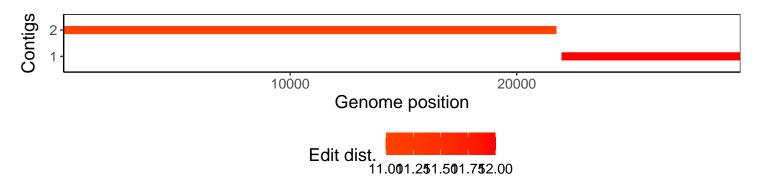
The plot below shows the number of reads covering each nucleotide position in the reference genome. Variants are shown as colored dots along the bottom of the plot and are color coded according by variant types: gray - transgenic, green - silent, gold - missense, red - nonsense, black - indel.



Excerpt from plot above focusing on reads coverage from 0 to 50 NT.

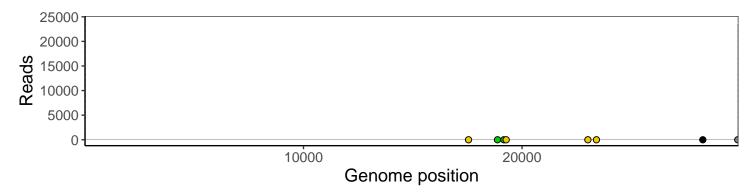


The longest five assembled contigs are shown below colored by their edit distance to the reference genome.

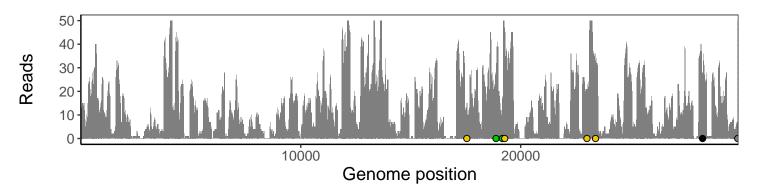


VSP0772-2 | 2021-02-01 | Saliva | PQ-Seq3 | genomes | single experiment

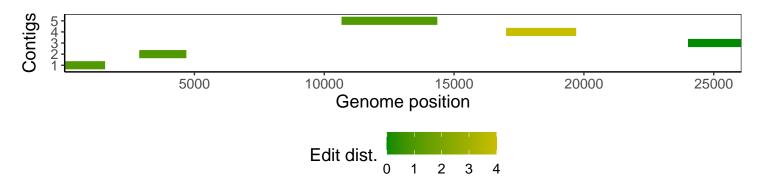
The plot below shows the number of reads covering each nucleotide position in the reference genome. Variants are shown as colored dots along the bottom of the plot and are color coded according by variant types: gray - transgenic, green - silent, gold - missense, red - nonsense, black - indel.



Excerpt from plot above focusing on reads coverage from 0 to 50 NT.



The longest five assembled contigs are shown below colored by their edit distance to the reference genome.



Software environment

Software/R package	Version
R	3.4.0
bwa	0.7.17-r1198-dirty
samtools	1.10 Using htslib 1.10
bcftools	1.10.2-34-g1a12af0-dirty Using htslib 1.10.2-57-gf58a6f3
pangolin	2.3.8
genbankr	1.4.0
optparse	1.6.0
forcats	0.3.0
stringr	1.4.0
dplyr	0.8.1
purrr	0.2.5
readr	1.1.1
tidyr	0.8.1
tibble	2.1.2
ggplot2	3.0.0
tidyverse	1.2.1
ShortRead	1.34.2
${\it Genomic Alignments}$	1.12.2
SummarizedExperiment	1.6.5
DelayedArray	0.2.7
matrixStats	0.54.0
Biobase	2.36.2
Rsamtools	1.28.0
GenomicRanges	1.28.6
$\operatorname{GenomeInfoDb}$	1.12.3
Biostrings	2.44.2
XVector	0.16.0
IRanges	2.10.5
S4Vectors	0.14.7
BiocParallel	1.10.1
BiocGenerics	0.22.1