

R for bioinformatics, data wrangler part 2

HUST Bioinformatics course series

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section 1: TOC

前情提要

tidyverse

- gather()
- spread()
- 没有仔细讲的
- unite()
- separate()

dplyr

- select()
- filter()
- mutate()
- summarise()
- arrange()
- group_by() ...

factors (部分)

本次提要

- ① factors 的更多应用 (forcats)
- ② 3 个生信任务的 R 解决方案
- ③ pipe

注不按上述顺序讲 ...

section 2: contents

生信任务 1 : network analysis and visualisation (dplyr & some plot packages)

protein-protein interaction data

why PPI is important?

- ① most of the time, protein functions together with other proteins (interactions)
- ② interacting partners tend to have similar functions (guilty by association, can be used in gene annotation)

STRING database

- ① contains >2 billion interactions for 24.5 million proteins in 5090 organisms (as of Aug 2019)
- ② contains:
 - physical interaction
 - genetic interactions
 - gene co-occurrence (text-mining)
 - transfers through orthologous relationships

a typical STRING plot

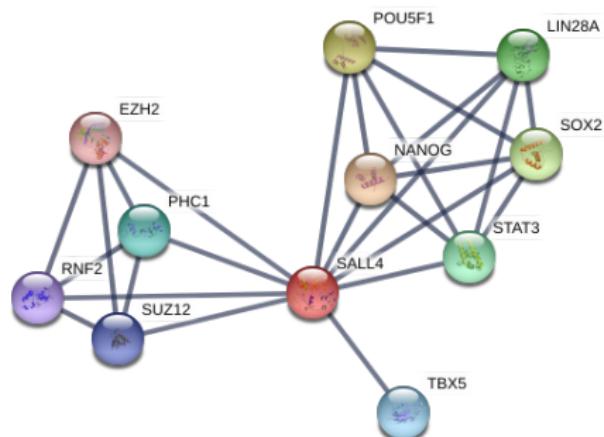


Figure 1: interacting partners of SALL4

note to previous plot

note this plot was generated using the following parameters:

- use SALL4 in human as query
- show only the top ten connectivity partners
- only connectivity score ≥ 900 (or 0.9) were shown

tasks of task 1

- ① get human PPI data
- ② limit the interactions to those with scores ≥ 900 (or 0.9)
- ③ find SALL4 and its top ten interaction partners
- ④ visualize the PPI network
- ⑤ calculate connectivities of sall4 and its top ten partners in the dataset

问题

生物学意义何在??

download human ppi data from STRING

go to : <https://string-db.org/cgi/download.pl>

The screenshot shows the STRING download interface. At the top, a red arrow points to a search bar containing "Homo sapiens". Another red arrow points to the "download" link above the table. The table lists five protein network data files for Homo sapiens, each with a CC-BY license link.

File	Description	Access
9606.protein.links.v11.0.txt.gz (71.2 Mb)	protein network data (scored links between proteins)	(cc) BY
9606.protein.links.detailed.v11.0.txt.gz (110.1 Mb)	protein network data (incl. subscores per channel)	(cc) BY
9606.protein.links.full.v11.0.txt.gz (127.6 Mb)	protein network data (incl. distinction: direct vs. interologs)	(cc) BY
9606.protein.actions.v11.0.txt.gz (14.4 Mb)	interaction types for protein links	(cc) BY

Figure 2: Download human PPI data from STRING

load and load the human PPI data

```
library(tidyverse);

## read_csv 也能处理压缩文件!!!
ppi <- read_delim( file = "data/talk06/ppi900.txt.gz", col_names = T,
                   delim = "\t", quote = "" );

## 查看一下数据 --
ppi %>% filter( gene1 == "SALL4" ) %>% do( head(., n = 10) );

## # A tibble: 10 x 3
##   gene1 gene2   score
##   <chr> <chr>   <dbl>
## 1 SALL4 POLR2E     900
## 2 SALL4 POLR2C     900
## 3 SALL4 POLR2I     900
## 4 SALL4 NANOG     992
## 5 SALL4 SALL1     923
## 6 SALL4 ZSCAN10    912
## 7 SALL4 LIN28A     957
## 8 SALL4 POU5F1     986
## 9 SALL4 SMAD2     906
## 10 SALL4 EPAS1     900
```

start to process the data

```
## get top 10 interacting partners of SALL4 by interaction score ...
toppart <- ppi %>% filter( gene1 == "SALL4" ) %>%
  arrange( desc( score ) ) %>% slice( 1:10 );

## get the interaction network consisting the top genes --
genes <- unique( c( "SALL4", toppart$gene2 ) );
netdata <- ppi %>% filter( gene1 %in% genes & gene2 %in% genes );
nrow(netdata);

## [1] 80
```

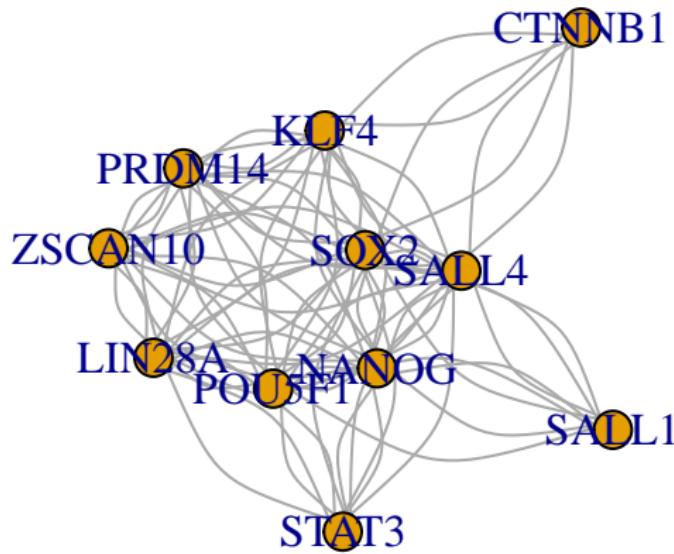
load the igraph package

```
## -- to make sure the installation will only run once ...
if (!require("igraph")){
  chooseCRANmirror();
  install.packages("igraph");
}

library( igraph );
```

visualise the network ...

```
netnet <- graph_from_data_frame( netdata, directed = FALSE );
plot(netnet);
```



图的问题

- very ugly
- two lines (redundancy) between every two nodes

redundancy among data

```
netdata %>% filter( gene1 %in% c("SALL4", "NANOG") & gene2 %in% c("SALL4", "NANOG") );  
  
## # A tibble: 2 x 3  
##   gene1 gene2 score  
##   <chr>  <chr> <dbl>  
## 1 NANOG  SALL4    992  
## 2 SALL4  NANOG    992
```

how to remove redundancy?

```
## create a new column, sort the two gene names, and concatenate them ...
testdata <-
  netdata %>% filter( gene1 %in% c("SALL4", "NANOG") & gene2 %in% c("SALL4", "NANOG") ) %>%
  mutate( group =
    if_else( gene1 > gene2,
            str_c( gene1, gene2, sep = "-" ),
            str_c( gene2, gene1, sep = "-" ) ) );

testdata;

## # A tibble: 2 x 4
##   gene1 gene2 score group
##   <chr> <chr> <dbl> <chr>
## 1 NANOG SALL4     992 SALL4-NANOG
## 2 SALL4 NANOG     992 SALL4-NANOG
```

Note `str_c` is from the `stringr` package!!

remove redundancy!

```
testdata %>% group_by( group ) %>% slice( 1 );
```

```
## # A tibble: 1 x 4
## # Groups:   group [1]
##   gene1 gene2 score group
##   <chr>  <chr>  <dbl> <chr>
## 1 NANOG SALL4    992 SALL4-NANOG
```

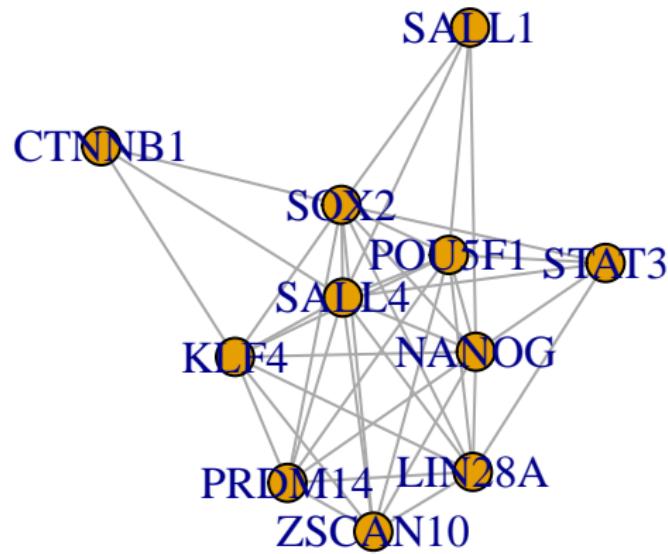
remove redundancy, cont.

```
netdata.nr <-  
  netdata %>%  
  mutate( group =  
    if_else( gene1 > gene2,  
            str_c( gene1, gene2, sep = "-"),  
            str_c( gene2, gene1, sep = "-" ) ) ) %>%  
  group_by( group ) %>% slice( 1 );  
  
nrow(netdata.nr);
```

```
## [1] 40
```

plot the non-redundant data

```
netnet.nr <- graph_from_data_frame( netdata.nr, directed = FALSE );  
plot(netnet.nr);
```



redundant 的数据可以用来计算 degree

```
net.stats <-  
  netdata %>% group_by( gene1 ) %>% summarise( degree = n() ) %>%  
  arrange( desc (degree) );  
net.stats;  
  
## # A tibble: 11 x 2  
##   gene1     degree  
##   <chr>     <int>  
## 1 SALL4      10  
## 2 SOX2       10  
## 3 NANOG       9  
## 4 POU5F1      9  
## 5 KLF4        8  
## 6 LIN28A      8  
## 7 PRDM14      7  
## 8 ZSCAN10      7  
## 9 STAT3        5  
## 10 SALL1       4  
## 11 CTNNB1       3
```

继续美化 network

```

## node 大小由 degree 决定
## 查看网络中基因名存储的顺序
V(netnet.nr)$name;

## [1] "KLF4"      "LIN28A"     "POU5F1"     "PRDM14"    "SALL1"     "CTNNB1"     "NANOG"
## [8] "SOX2"       "STAT3"      "ZSCAN10"    "SALL4"

## 获取它们相对应的 degree ...
net.stats[match( V(netnet.nr)$name , net.stats$gene1 ), ];

```

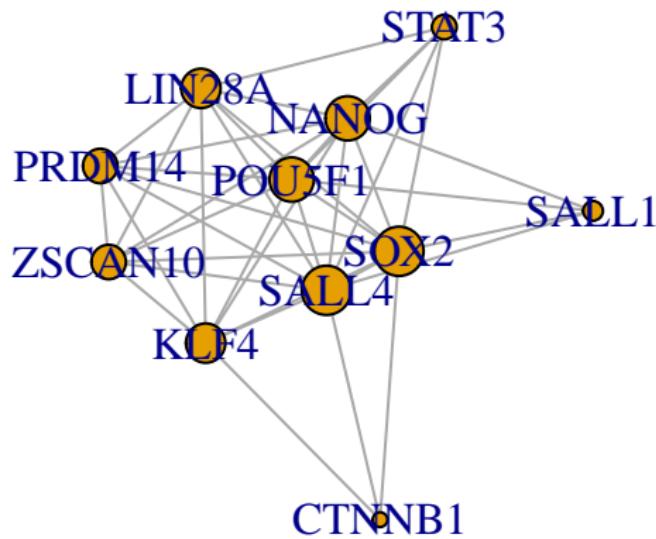
```

## # A tibble: 11 x 2
##   gene1   degree
##   <chr>     <int>
## 1 KLF4        8
## 2 LIN28A      8
## 3 POU5F1      9
## 4 PRDM14      7
## 5 SALL1       4
## 6 CTNNB1      3
## 7 NANOG       9
## 8 SOX2        10
## 9 STAT3       5
## 10 ZSCAN10     7
## 11 SALL4      10

```

继续美化 network, cont.

```
vertex_attr(netnet.nr, "size") <-  
  net.stats$degree[match( V(netnet.nr)$name , net.stats$gene1 ) ] * 2;  
plot( netnet.nr );
```



更多美化

详见：

- ① [igraph introduction](#)
- ② [a comprehensive network visualisation tutorial in R: igraph and beyond](#)

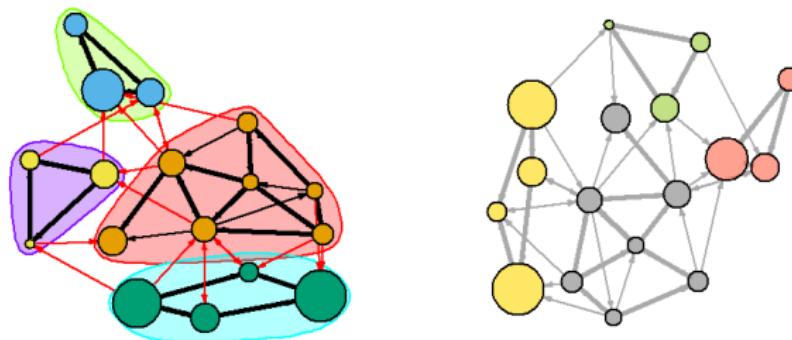


Figure 3: Example final outcomes

other network visualisation packages

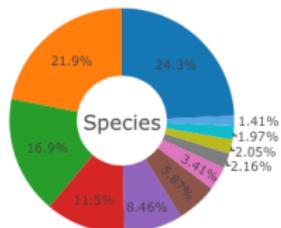
- ① ggnet
- ② interactive networkD3 R package
- ③ plotly.js
- ④ D3

生信任务 2：宏基因基因数据展示的小应用 (forcats)

Relative abundances

Species

Relative abundance



Relative abundance

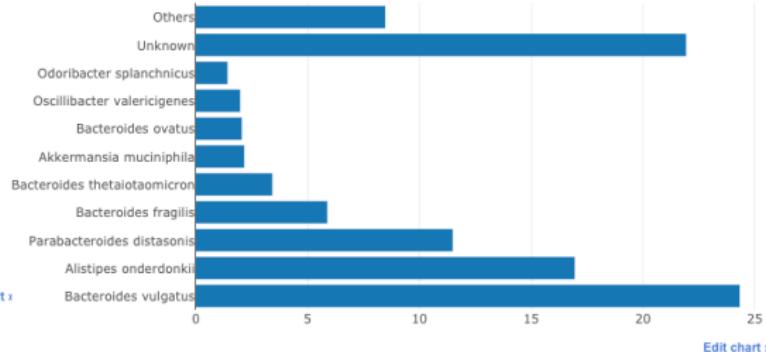


Figure 4: 肠道物种丰度示意图

Find more at the GMrepo database.

什么是宏基因组？

Metagenomics is the study of genetic material recovered directly from environmental samples.

research contents

- mostly prokaryotes
- unicellular eukaryotes
- viruses

techniques

- 16S (universially conserved gene in prokaryotes)
- whole genome sequencing (WGS or metagenomics)

what can metagenomics do?

- identify new species
- reveal species kinds and abundances
- relate species changes to human health and disease

biomes



Figure 5: Biomes

Screenshot taken from the EBI metagenomics database on Aug 8, 2019.

environmental microbiome

- soil
- ocean



Figure 6: The tara oceans expedition

host associated microbiomes

- human body sites
- animals & plants



Figure 7: NIH human microbiome project

why human gut microbiota is important?

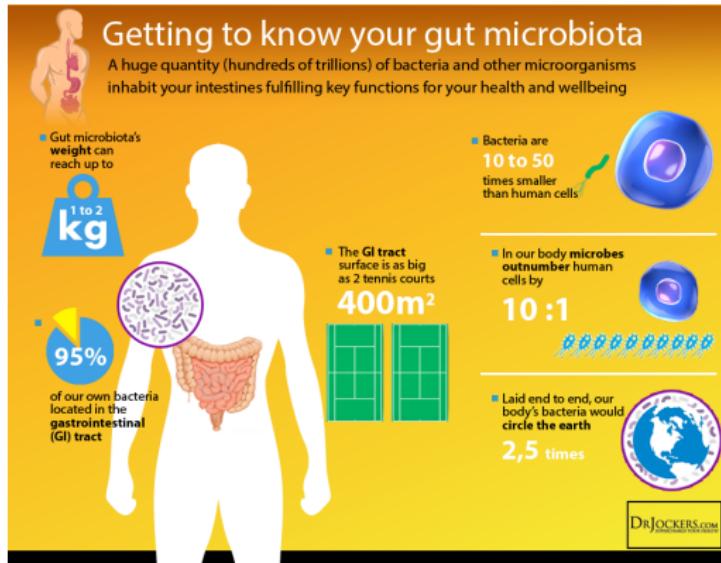


Figure 8: human gut microbiota

tasks of human gut microbiota analysis

- identify new species
- find good, bad and commensal microbes
- link microbial variations to human health
- mechanisms
- modulation, intervention and regulation

typical human gut microbiome data

Species abundances

```
abu <- read_delim(file = "data/talk06/relative_abundance_for_RUN_ERR1072629_taxonlevel_species.csv",
                   delim = "\t", quote = "", comment = "#");

## Parsed with column specification:
## cols(
##   ncbi_taxon_id = col_double(),
##   relative_abundance = col_double(),
##   scientific_name = col_character()
## )

nrow(abu);

## [1] 122
```

Species abundances, cont.

```
abu %>% arrange( desc( relative_abundance ) ) %>% do( head(., n = 10) );
```

```
## # A tibble: 10 x 3
##   ncbi_taxon_id relative_abundance scientific_name
##       <dbl>            <dbl> <chr>
## 1         821            24.3  Bacteroides vulgatus
## 2           -1            21.9  Unknown
## 3        328813           16.9  Alistipes onderdonkii
## 4         823             11.5  Parabacteroides distasonis
## 5         817              5.87  Bacteroides fragilis
## 6         818              3.41  Bacteroides thetaiotaomicron
## 7        239935             2.16  Akkermansia muciniphila
## 8        28116              2.05  Bacteroides ovatus
## 9        351091             1.97  Oscillibacter valericigenes
## 10        28118             1.41  Odoribacter splanchnicus
```

相对丰度作图要求

- ① 按丰度从高到低排序
- ② 只取前 10 个 species (保留 10 行)
- ③ 将后面的丰度累加在一起，汇总为“Others”分类

数据处理

```

library( tidytidbits );
abu.dat <-
  abu %>% arrange( desc( relative_abundance ) ) %>%
    lump_rows( scientific_name, relative_abundance, n = 10, other_level = "Others" );

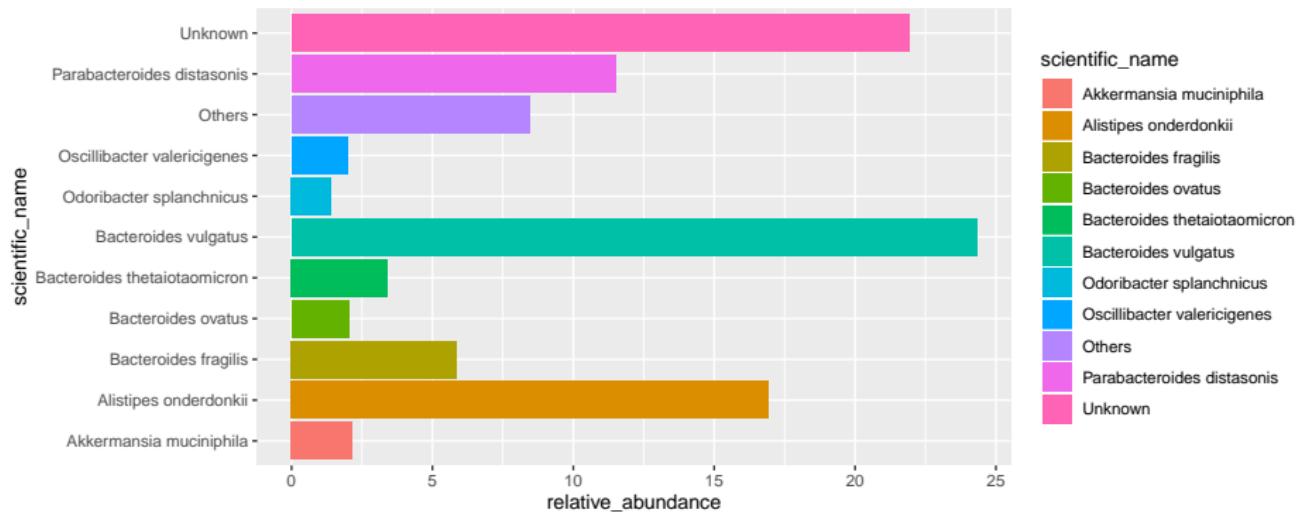
head(abu.dat, n = 11);

## # A tibble: 11 x 3
##   ncbi_taxon_id relative_abundance scientific_name
##   <dbl>           <dbl> <chr>
## 1 821            24.3  Bacteroides vulgatus
## 2 -1              21.9  Unknown
## 3 328813          16.9  Alistipes onderdonkii
## 4 823             11.5  Parabacteroides distasonis
## 5 817             5.87   Bacteroides fragilis
## 6 818             3.41   Bacteroides thetaiotaomicron
## 7 239935          2.16   Akkermansia muciniphila
## 8 28116            2.05   Bacteroides ovatus
## 9 351091          1.97   Oscillibacter valericigenes
## 10 28118           1.41   Odoribacter splanchnicus
## 11 31199992        8.46   Others

```

尝试作图

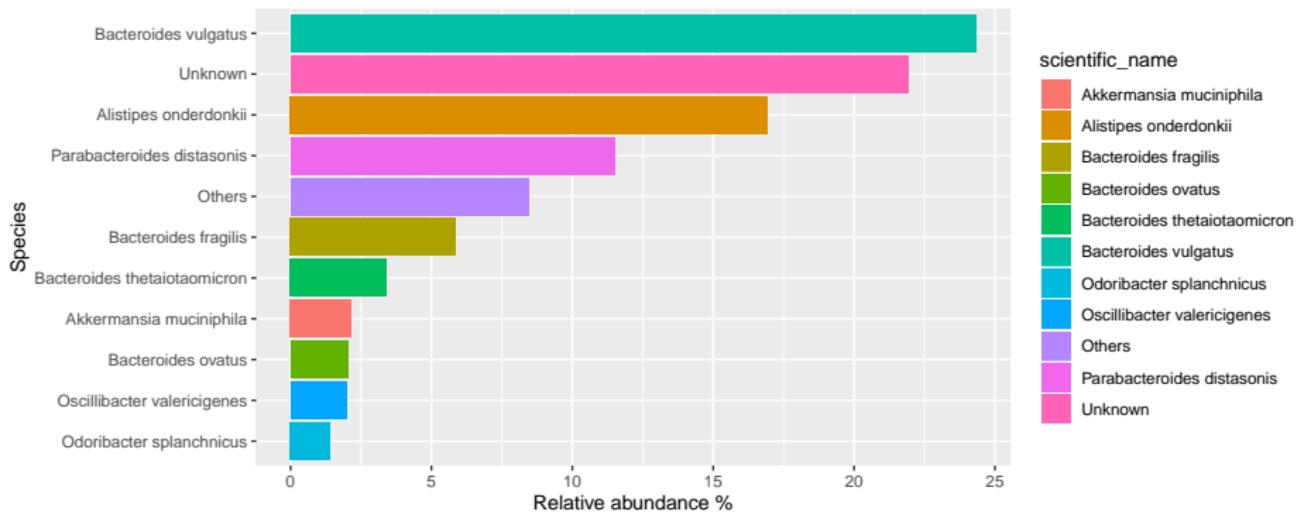
```
ggplot(abu.dat, aes(x = scientific_name, y = relative_abundance, fill = scientific_name ) ) +  
  geom_bar( stat = "identity" ) +  
  coord_flip()
```



调整排列顺序

用 `forcats` 包的 `fct_reorder` 函数；注意它 3 个参数的意义!!!

```
ggplot(abu.dat, aes(x = fct_reorder( scientific_name, relative_abundance, .desc = F),
                     y = relative_abundance, fill = scientific_name ) ) +
  geom_bar( stat = "identity" ) +
  coord_flip() + xlab("Species") + ylab( "Relative abundance %" )
```



更多forcats的应用 ...

see here: <https://cran.r-project.org/web/packages/forcats/vignettes/forcats.html>

更多应用将会在作图（ggplot2）时讲到。

生信任务 3：整合基因表达、甲基化、突变数据 (dplyr::join)

在生信分析中，常需要将多个来源的数据整合在一个表格中，以方便后续分析。

```
meth <- read_delim( file = "data/talk06/methylation_data.txt.gz",
                     delim = "\t", quote = "", col_names = T);

## Parsed with column specification:
## cols(
##   gene = col_character(),
##   site = col_character(),
##   methylation_score = col_double()
## )

head(meth, n = 3);

## # A tibble: 3 x 3
##   gene    site  methylation_score
##   <chr>   <chr>          <dbl>
## 1 A1BG   1stExon        0.799
## 2 A1BG   5UTR           0.799
## 3 A1BG   Body            0.811
```

问题什么是甲基化 ??

表达数据

```
expr <- read_delim( file = "data/talk06/expression_data.txt.gz",
                     delim = "\t", quote = "", col_names = T );
```

```
## Parsed with column specification:
## cols(
##   gene = col_character(),
##   rkpm = col_double()
## )
```

```
head( expr, n = 5);
```

```
## # A tibble: 5 x 2
##   gene      rkpm
##   <chr>    <dbl>
## 1 5S_RRNA     0
## 2 7SK        0
## 3 A1BG       1
## 4 A1BGAS1    4
## 5 A1CF       0
```

整合后的结果应该是什么 ??

gene	TSS200	TSS1500	UTR	body	expression
gene1	0.1	0.2	NA	0.8	100
gene2	0.12	0.32	NA	0.9	18

...

第一种方法，使用 spread

先合并，再 spread 用 bind_rows 合并两个 tibble 时，列名需要一致

```
meth2 <- meth %>% select( gene, group=site, value=methylation_score );
head(meth2, n=2);
```

```
## # A tibble: 2 x 3
##   gene   group   value
##   <chr>  <chr>   <dbl>
## 1 A1BG   1stExon 0.799
## 2 A1BG   5UTR     0.799
```

```
expr2 <- expr %>% mutate( group = "rkpm" ) %>%
  select( gene, group, value=rkpm ) %>%
  group_by( gene ) %>% slice( 1 );
head(expr2, n=2);
```

```
## # A tibble: 2 x 3
## # Groups:   gene [2]
##   gene   group value
##   <chr>  <chr> <dbl>
## 1 5S_RRNA rkpm     0
## 2 7SK      rkpm     0
```

注： gene name 与 group 组合必须是唯一的。即：基因 A 只能有一个表达量值。

合并 & spread

```
comb <- bind_rows( meth2, expr2 );
comb.wide <- comb %>% spread( group, value );

head(comb.wide);

## # A tibble: 6 x 8
##   gene    `1stExon` `3UTR` `5UTR`     Body    rkpm TSS1500 TSS200
##   <chr>    <dbl>   <dbl>   <dbl>    <dbl>   <dbl>   <dbl>   <dbl>
## 1 5S_RRNA    NA      NA      NA        0     NA     NA
## 2 7SK        NA      NA      NA        0     NA     NA
## 3 A1BG       0.799   NA      0.799    0.811    1    0.852  0.850
## 4 A1BGAS1    NA      NA      NA        4     NA     NA
## 5 A1CF       0.0946  NA      0.118    0.00798   0    0.110  NA
## 6 A2M        0.899   0.776  NA      0.716     1    0.654  NA
```

方法二：使用 join ...

首先对 methylation 数据进行处理

```
meth3 <-  
  meth %>% spread( site, methylation_score );  
  
head(meth3);  
  
## # A tibble: 6 x 7  
##   gene   `1stExon` `3UTR` `5UTR`     Body TSS1500 TSS200  
##   <chr>    <dbl>   <dbl>   <dbl>    <dbl>   <dbl>   <dbl>  
## 1 A1BG      0.799    NA     0.799  0.811    0.852  0.850  
## 2 A1CF      0.0946   NA     0.118  0.00798   0.110  NA  
## 3 A2M       0.899    0.776  NA     0.716    0.654  NA  
## 4 A2ML1     0.859    0.469  0.859  0.678    0.441  NA  
## 5 A4GALT    NA       0.814  0.844  0.821    0.890  0.513  
## 6 A4GNT     0.680    NA     0.680  0.538    0.402  NA
```

dplyr::join

```
comb2 <- left_join( meth3, expr2, by = "gene" ) %>% select( -group );  
head(comb2);
```

```
## # A tibble: 6 x 8  
##   gene   `1stExon` `3UTR` `5UTR`     Body TSS1500 TSS200 value  
##   <chr>     <dbl>   <dbl>   <dbl>     <dbl>   <dbl>   <dbl> <dbl>  
## 1 A1BG      0.799    NA     0.799  0.811     0.852   0.850    1  
## 2 A1CF      0.0946   NA     0.118  0.00798    0.110   NA       0  
## 3 A2M       0.899    0.776  NA     0.716     0.654   NA       1  
## 4 A2ML1     0.859    0.469  0.859  0.678     0.441   NA       0  
## 5 A4GALT    NA       0.814  0.844  0.821     0.890   0.513    0  
## 6 A4GNT     0.680    NA     0.680  0.538     0.402   NA       0
```

注意 left_join 的语法

join 详解

- `left_join()`: return all rows from x, and all columns from x and y.
Rows in x with no match in y will have NA values in the new columns. If there are multiple matches between x and y, all combinations of the matches are returned.
- `inner_join()`
- `right_join()`
- `full_join()`

更多请见: <https://dplyr.tidyverse.org/reference/join.html>

section 3: pipe

什么是 pipe ?

- pipe 就是 `%>%`
- it comes from the `magrittr` package by **Stefan Milton Bache**
- Packages in the tidyverse load `%>%` for you automatically, so you don't usually load `magrittr` explicitly.
- 实质是中间值的传递

示例：

是否所有函数都支持 pipe ?

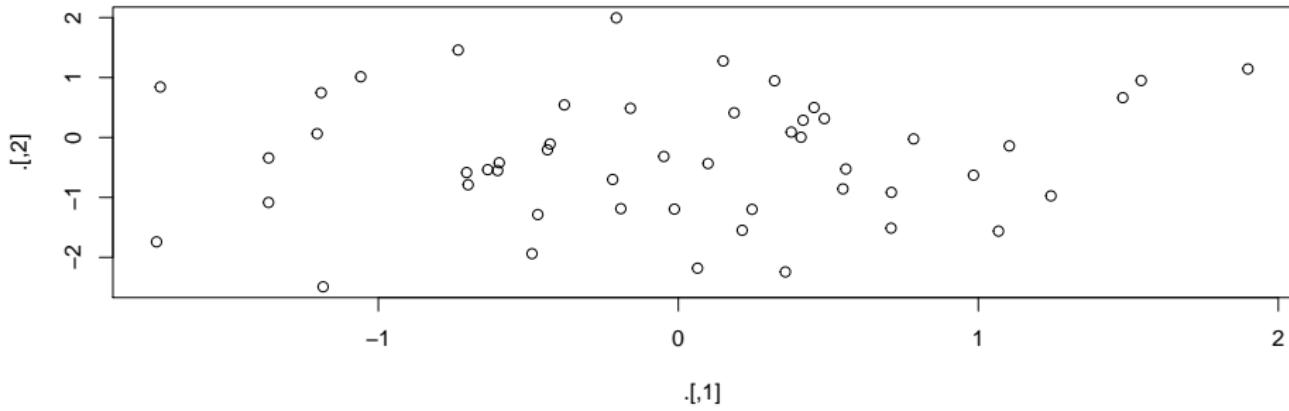
是的。

通常需要用 . 指代传递来的数据，并以参数的形式赋予下游函数：

其它形式的 pipe

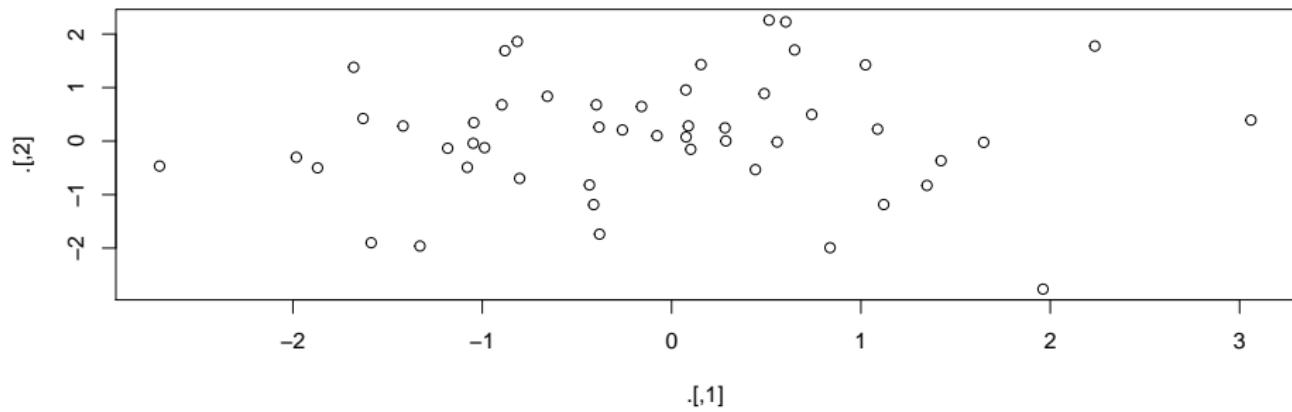
%T>% : 返回上游的值 (???)

```
library(magrittr)
## 示例: res1 是空值 ...
res1 <-
  rnorm(100) %>%
  matrix(ncol = 2) %>%
  plot();
```



%T>%: 返回上游值 (left-side values)

```
## 示例: res2 是 matrix() 内容 ...
res2 <-
  rnorm(100) %>%
  matrix(ncol = 2) %T>%
  plot();
```



%T>%: 返回上游值 (left-side values), cont.

```
head(res2);  
  
##           [,1]      [,2]  
## [1,] -0.1585816  0.64605052  
## [2,]  0.4426997 -0.53331789  
## [3,] -1.5866564 -1.89942995  
## [4,] -1.0480959 -0.04084271  
## [5,] -1.0448633  0.34545853  
## [6,] -0.2598246  0.20875275
```

%\$% : attach ???

```
attach( mtcars ); ## note the warning message ...
```

```
## The following object is masked from package:ggplot2:  
##  
##     mpg
```

```
cor.test( cyl, mpg ); ## 汽缸数与燃油效率
```

```
##  
## Pearson's product-moment correlation  
##  
## data: cyl and mpg  
## t = -8.9197, df = 30, p-value = 6.113e-10  
## alternative hypothesis: true correlation is not equal to 0  
## 95 percent confidence interval:  
## -0.9257694 -0.7163171  
## sample estimates:  
## cor  
## -0.852162
```

%\$% : attach ??? , cont.

```
detach( mtcars );
with( mtcars, cor.test( cyl, mpg ) );

##
## Pearson's product-moment correlation
##
## data: cyl and mpg
## t = -8.9197, df = 30, p-value = 6.113e-10
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
## -0.9257694 -0.7163171
## sample estimates:
##       cor
## -0.852162
```

%\$% : attach ??? , cont.

```
mtcars %$%
  cor.test( cyl, mpg );
```

```
##
##  Pearson's product-moment correlation
##
## data: cyl and mpg
## t = -8.9197, df = 30, p-value = 6.113e-10
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
## -0.9257694 -0.7163171
## sample estimates:
##       cor
## -0.852162
```

其它 pipe 及注意事项

```
## 双向 pipe  
mtcars %<>% transform(cyl = cyl * 2);
```

注

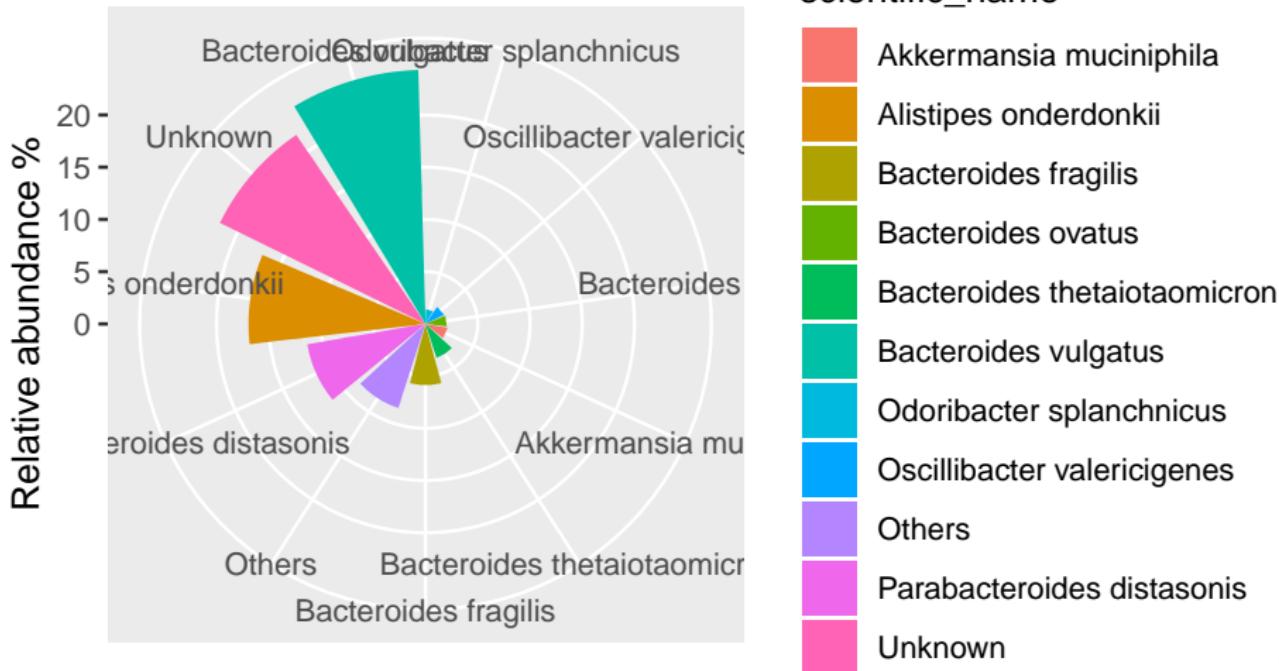
- pipe 的使用可以使思路更清晰
- 因此，尽量使用 %>% (方向明确)，而不使用其它方向不明确的 pipe

section 4: Exercise & home work

练习

练习本堂讲到的三个示例

用示例 2 的数据画饼图



作业: pie chart

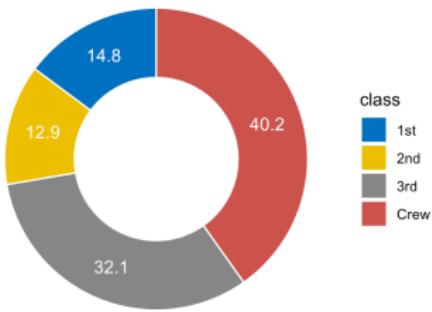


Figure 9: make a pie chart like this using the meteagenomics data

详见: [https://www.datanovia.com/en/blog/
how-to-create-a-pie-chart-in-r-using-ggplot2/](https://www.datanovia.com/en/blog/how-to-create-a-pie-chart-in-r-using-ggplot2/)

更多练习

- join: <https://r4ds.had.co.nz/relational-data.html>
- forcats: <https://cran.r-project.org/web/packages/forcats/vignettes/forcats.html>

下次预告

Strings and regular expression