Single-Shot Magnetic Resonance Spectroscopic Imaging with Partial Parallel Imaging

Stefan Posse,^{1,2*} Ricardo Otazo,² Shang-Yueh Tsai,³ Akio Ernesto Yoshimoto,² and Fa-Hsuan Lin^{4,5,6}

A magnetic resonance spectroscopic imaging (MRSI) pulse sequence based on proton-echo-planar-spectroscopic-imaging (PEPSI) is introduced that measures two-dimensional metabolite maps in a single excitation. Echo-planar spatial-spectral encoding was combined with interleaved phase encoding and parallel imaging using SENSE to reconstruct absorption mode spectra. The symmetrical k-space trajectory compensates phase errors due to convolution of spatial and spectral encoding. Single-shot MRSI at short TE was evaluated in phantoms and in vivo on a 3-T whole-body scanner equipped with a 12-channel array coil. Four-step interleaved phase encoding and fourfold SENSE acceleration were used to encode a 16 × 16 spatial matrix with a 390-Hz spectral width. Comparison with conventional PEPSI and PEPSI with fourfold SENSE acceleration demonstrated comparable sensitivity per unit time when taking into account g-factor-related noise increases and differences in sampling efficiency. LCModel fitting enabled quantification of inositol, choline, creatine, and N-acetyl-aspartate (NAA) in vivo with concentration values in the ranges measured with conventional PEPSI and SENSE-accelerated PEPSI. Cramer-Rao lower bounds were comparable to those obtained with conventional SENSE-accelerated PEPSI at the same voxel size and measurement time. This single-shot MRSI method is therefore suitable for applications that require high temporal resolution to monitor temporal dynamics or to reduce sensitivity to tissue movement. Magn Reson Med 61:541-547, 2009. © 2008 Wiley-Liss, Inc.

DOI 10.1002/mrm.21855

Published online 18 December 2008 in Wiley InterScience (www.interscience.wiley.com).

Key words: magnetic resonance spectroscopic imaging; singleshot encoding; proton-echo-planar-spectroscopic-imaging; parallel imaging; SENSE; spectral quantification

Ultra—high-speed magnetic resonance spectroscopic imaging (MRSI) with acquisition times in the range of a few seconds will enable novel research applications, such as the measurement of metabolic dynamics in hyperpolarized MRI for tumor diagnosis and treatment monitoring (1–3), as well as assessment of cardiac function (4). Another possible application, which is complementary to diffusion-sensitive MRI (5–8), is spatial mapping of metabolite diffusion to study intracellular mobility (9,10). Ultra—high-speed MRSI is also applicable to biochemical imaging in moving organs, such as the heart, liver, and breast (11,12).

A number of laboratories have developed high-speed MRSI methods, mostly using echo-planar and spiral readout modules, which provide considerable acceleration as compared with conventional phase-encoded MRSI (13-22). Recently, Mayer et al. described ultra-high-speed metabolic imaging of systems with sparse spectra for applications in hyperpolarized ¹³C imaging (23). The imaging method in their study was based on spiral readout encoding and enabled single-shot mapping of ¹³C spectra in seconds, albeit at narrow spectral width, which required separate reconstruction of each spectral peak. In a follow-up publication, Levin et al. described a least-squares reconstruction to optimize fast spiral spectroscopic imaging by taking into account chemical shift evolution during the readout period (24). Both methods are most suitable for sparse spectra with well-separated peaks.

Recent advances in parallel MRI employing information from different channels in a radio-frequency (RF) coil array enable considerable acceleration of MRSI. Dydak et al. demonstrated acceleration of conventional phase-encoded MRSI with acquisition times of just a few minutes using sensitivity encoding (SENSE) (25,26).

Recently, we have developed even faster spatial–spectral encoding by combining proton–echo-planar-spectroscopic-imaging (PEPSI) with parallel imaging to obtain minimum encoding times of 1 min for three-dimensional (3D) encoding of a 32 \times 32 \times 8 matrix (27) and two-dimensional (2D) mapping with encoding times as short as 16 sec for a 32 \times 32 matrix (28,29).

In the present study, we extend this approach by interleaving partial phase encoding into the PEPSI readout trajectory (30,31) and using SENSE reconstruction (27) to achieve 2D spatial and spectral encoding in a single-shot with spectral width large enough to encompass the major singlet resonances in ¹H spectra in the human brain. The readout module was designed with symmetric forward

¹Department of Neurology, University of New Mexico School of Medicine, Albuquerque, New Mexico, USA.

²Department of Electrical & Computer Engineering, University of New Mexico, Albuquerque, New Mexico, USA.

³Department of Electrical Engineering, Chang Gung University, Taipei, Taiwan.

⁴Massachusetts General Hospital-Harvard Medical School-Massachusetts Institute of Technology (MGH-HMS-MIT) Athinoula A. Martinos Center for Biomedical Imaging, Charlestown, Massachusetts, USA.

⁵Department of Radiology, Massachusetts General Hospital, Boston, Massachusetts, USA.

⁶Institute of Biomedical Engineering, National Taiwan University, Taipei, Taiwan.

Presented in part at the 15th annual meeting of ISMRM, Berlin, Germany, 2007.

Grant sponsor: National Institutes of Health; Grant numbers: R01 HD040712, R01 NS037462, R01 EB000790-04, P41 RR14075, R21 EB007298, R01 DA14178-01; Grant sponsor: Mental Illness and Neuroscience Discovery Institute (MIND); Grant sponsor: Taiwan National Science Council; Grant numbers: NSC-95-2221-E-002-179, NSC 96-2320-B-002-085; Grant sponsor: National Health Research Institute Taiwan: Grant number: E29C97N.

^{*}Correspondence to: Stephan Posse, Department of Neurology, University of New Mexico School of Medicine, University of New Mexico, MSC10 5620, Albuquerque, NM 87131. E-mail: sposse@unm.edu

Received 29 January 2008; revised 22 August 2008; accepted 18 September 2008.

542 Posse et al.

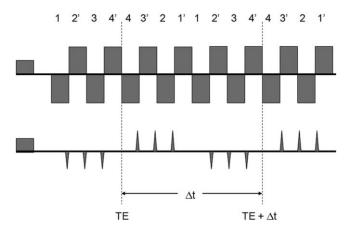


FIG. 1. Gradient switching scheme for single-shot spatial–spectral encoding with negative and positive phase-encoding gradient blips. A set of eight readout gradients with duration t forms a readout module with negative readout gradients labeled 1, 2, 3, and 4, and positive readout gradients labeled 1′, 2′, 3′, and 4′. This module encodes an imaging matrix with 16 pixels in the phase-encoding direction using fourfold acceleration with partial parallel imaging. The readout module is repeated N times to encode the spectral dimension (61 \times 41 mm; 300 \times 300 DPI).

and backward k-space trajectories to enable reconstruction of pure absorption mode spectra. We show quantitative mapping of major spectral peaks of inositol, choline, creatine, and N-acetyl-aspartate (NAA) in vivo on a 3-T whole-body MR scanner equipped with a 12-channel receive-only coil array. We also compare single-shot PEPSI with our recently published PEPSI methodology using conventional phase encoding (conventional PEPSI) (22) and fourfold SENSE acceleration (SENSE-PEPSI) (28), and with data from a pulse sequence that combines single-shot encoding and additional conventional phase encoding to characterize the point spread function (PSF-PEPSI). Conventional PEPSI data with spectral width identical to that of single-shot encoding (low bandwidth PEPSI-LBW-PEPSI) were extracted from PSF-PEPSI to characterize the effects of narrow spectral width on sensitivity and spectral quantification. Preliminary accounts of this work were presented at a recent meeting (32).

METHODS

Theory

The single-shot method is based on the PEPSI-encoding scheme described in previous publications (21,22) and requires a phased-array RF receive coil that generates a sufficient number of spatially distinct sensitivity patterns to enable acceleration with parallel imaging. Spatial–spectral encoding is performed by interleaving phase-encoding gradient blips into the alternating readout gradient train between pairs of positive and negative readout gradient pulses to form a series of 2D spatial-encoding modules with duration Δt (Fig. 1). Repetition of this spatial-encoding module encodes spectral information with reconstructed spectral width $1/\Delta t$. The k-space trajectory is designed to: (a) maximize signal-to-noise ratio (SNR) by minimizing analog-to-digital converter (ADC) dead time using ramp sampling; (b) minimize eddy current effects using

uniform readout gradient inversion periodicity; and (c) obtain symmetrical chemical shift—related phase evolution in the even and odd echo data by positioning the center of the first readout module at the spin—echo time TE. For each spatial encoding module a series of positive phase-encoding gradients blips is followed by a series of negative phase-encoding gradient blips, resulting in a zig-zag trajectory in k_x – k_y –t-space (Fig. 2).

Each readout module in k_x – k_y space is traversed symmetrically forward and backward. The end of the forward k-space trajectory of the first readout module is positioned at the spin–echo time TE to minimize first-order phase error upon combination of even and odd echo data. Accelerated phase-encoding steps through k_y -space by skipping R k_y -space lines.

Single-shot encoding requires interleaving of phase-encoding gradients, which reduces the sampling efficiency of the ADC as compared with conventional PEPSI encoding. With adaptation according to the theory described by Pohmann et al. (33), the normalized SNR per unit time and unit volume is given by:

$$SNR = \frac{SNR_{ref}V\sqrt{1 - t_d/(t_{ADC} + t_d)}\sqrt{TR \times N}}{g}$$
 [1]

where SNR_{ref} is the reference SNR per unit time and unit volume ($\sec^{-1/2}$ cm⁻¹) measured in data acquired with conventional PEPSI encoding, V is voxel size (cm³), t_d is ADC dead time during readout module (sec), t_{ADC} is ADC on time during readout module (sec), TR is repetition time (sec), N is number of scan repetitions number (taking into account R-fold acceleration with parallel imaging), and g is the g-factor. The sampling efficiency, σ , of the ADC is defined as:

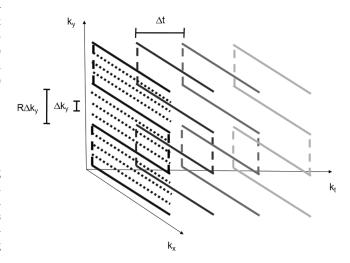


FIG. 2. The k_x – k_y –t-space trajectory for single-shot spatial–spectral encoding. Phase-encoding lines are measured with k_y -spacing $R\Delta k$, where R(=4) denotes the acceleration factor. Target k_y -spacing after SENSE reconstruction is Δk . Each readout module, which consists of four forward and four reverse k_x – k_y -space-encoding segments, is traversed forward and backward, and repeated N-times with spacing Δt to encode N spectral points. Data are only collected during the horizontal segments of the trajectory (69 \times 50 mm; 300 \times 300 DPI).

Single-Shot MRSI 543

$$\sigma = \sqrt{1 - t_d / (t_{ADC} + t_d)}$$
 [2]

Pulse Sequence Implementation

Single-shot PEPSI was implemented on a 3-T Siemens Tim Trio scanner equipped with 12-channel head array coils. Spatial-spectral encoding was performed with 2048 trapezoidal readout gradients (240-µs duration), 100-µs phaseencoding blips, and fourfold uniform acceleration of phase encoding to encode a 16 \times 16 spatial matrix with a 192-mm minimum field of view (FOV). Four readout segments for the forward trajectory and four readout segments for the backward trajectory, respectively, resulted in a 2.56-msec readout module. Data were collected with twofold oversampling, regridded to correct the k-space trajectory distortion due to linear ramp sampling, and decimated two-fold to remove oversampling, as described previously (21,22). The reconstructed spectral width was 390 Hz and the digital spectral resolution was 1.5 Hz. The pulse sequence included three-pulse water suppression and eight-slice outer volume suppression, as described previously (21,22). In addition, a modified pulse sequence was used to collect data to characterize the point spread function (PSF) of the single-shot encoding sequence (PSF-PEPSI) and to obtain conventionally phase-encoded PEPSI data with spectral width identical to that of single-shot encoding (low-bandwidth PEPSI-LBW-PEPSI). To collect these data the single-shot PEPSI sequence was repeated 16 times with a stepwise incremented conventional phaseencoding gradient along the y-direction to fill an additional k-space dimension, referred to here as the PSF dimension.

Data Acquisition, Reconstruction, and Analysis

Measurements were performed on a spherical phantom filled with a metabolite solution with concentration values in the millimolar range and in healthy volunteers after obtaining institutionally approved consent. Data were acquired with single-shot PEPSI, PSF-PEPSI, and with conventional PEPSI using TE = 11 msec, TR = 2 sec, and voxel size = $14 \times 14 \times 15$ mm³. Single-shot PEPSI was performed with a FOV of 230 mm and 16 averages resulting in a 32-sec scan time. PSF-PEPSI was performed with a FOV of 230 mm and 1 average resulting in a 32-sec scan time. Conventional PEPSI was performed with a 1087-Hz spectral width, a 32 \times 16 (readout \times phase encode) image matrix, a FOV of 460 mm imes 230 mm, and four averages resulting in 128-sec scan time. Fourfold accelerated PEPSI data (SENSE-PEPSI) were obtained from data measured with conventional PEPSI using fourfold undersampling along the k_v -dimension, resulting in an equivalent measurement time of 32 sec.

Data obtained with PSF-PEPSI were edited to obtain conventional phase-encoded low-bandwidth PEPSI (LBW-PEPSI) data by extracting for each repetition of the readout module the two gradient echoes that correspond to the center of the k_y -space encoded by the blipped phase-encoding gradients (labeled 2 and 2' in Fig. 1).

Coil sensitivity maps were estimated using spectral water images from an extra fully phase-encoded non-water-suppressed (NWS) acquisition, as described previously

(27,28). Array geometry-related noise amplification in the reconstruction was computed using the g-factor (27,28). Even and odd echo data were reconstructed separately using SENSE reconstruction with regularization, as described previously (27). Coil-by-coil SENSE reconstruction was generated by multiplying the combined SENSE reconstruction with the coil sensitivities in order to allow for application of spectral phase correction and frequency alignment to each channel separately. A sine bell filter was applied across the two k-space dimensions and Fourier transformation in the temporal domain was applied. The reconstructed NWS data were used to automatically correct zero-order phase error and frequency offset on a voxelby-voxel basis. First-order phase modulation in the even and the odd echo data sets canceled during combination of the two data sets, resulting in pure absorption mode spectra. Finally, the reconstructed multicoil data were combined using sensitivity-weighted combination.

Spectra were quantified using LCModel fitting with analytically modeled basis sets, as described previously (22). Errors in metabolite quantification in LCModel (% SD) are expressed in Cramer–Rao lower bound (CRLB, the lowest bound of the standard deviation of the estimated metabolite concentration expressed as percentage of this concentration), which, when multiplied by 2.0, represent 95% confidence intervals of the estimated concentration values (34). SNR values were taken from LCModel output.

Metabolite concentration images at 3 T were created using the following thresholds to accept voxels: (a) CRLB = 30% for creatine (Cr) and NAA, and CRLB = 50% for choline (Cho) and inositol (Ins); and (b) spectral line width (FWHM) = 0.2 ppm. Finally, the metabolite concentration maps were interpolated to a 128×128 matrix using zero-filling of the k-space data to improve visualization.

RESULTS

Data acquired with single-shot PEPSI encoding demonstrate adequate spectral width to encompass the major metabolites of interest in $^1\mathrm{H}$ spectroscopy of brain. PSF-PEPSI k-space data obtained at 3 T in a spherical phantom confirmed that blipped interleaved phase encoding was consistent with conventional phase encoding: When plotting the k-space signal intensity as a function of PSF k_y -dimension versus the single-shot k_y -dimension, a nearly linear relationship was found.

Fig. 3 shows metabolic images and water-suppressed spectra acquired on a phantom with conventional PEPSI, SENSE-PEPSI, LBW-PEPSI, and single-shot PEPSI. The average g-factor of SENSE reconstruction was 2.1 \pm 0.4 (Fig. 3b). The average SNR of the single-shot PEPSI data (7.8 \pm 6.8) was in a similar range as that in the SENSE-PEPSI data (7.5 ± 2.3) and the LBW-PEPSI data (8.0 ± 2.7) . The average spectral line width measured with all four methods was similar (conventional PEPSI 4.3 \pm 2.8 Hz, SENSE-PEPSI 5.0 \pm 3.2 Hz, LBW-PEPSI 4.0 \pm 2.4 Hz, single-shot PEPSI 4.5 \pm 2.6 Hz) (Fig. 3c'-f'). The baseline in the SENSE-PEPSI and single-shot PEPSI spectra was slightly distorted due to residual water aliased from distant regions with B_0 shifts and spatial aliasing of peripheral lipid signals due to imperfections in SENSE reconstruction. In the LBW-PEPSI and single-shot PEPSI data, the spectral alias544 Posse et al.

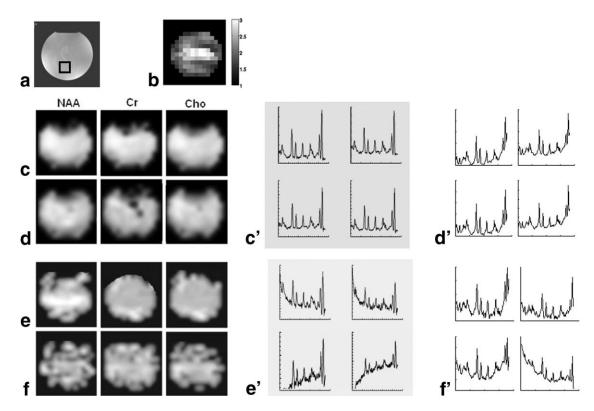


FIG. 3. Comparison of MRSI methods on the spherical phantom. (a) MRI of with volume location of spectral arrays shown below. (b) *g*-factor map for SENSE reconstruction. (c,d) Spectroscopic images of choline (Cho), creatine (Cr), and *N*-acetyl-aspartate (NAA) (TR/TE 2 sec/11 msec, voxel size 3 cm³) PEPSI (c) using conventional, (d) SENSE-PEPSI, (e) LBW-PEPSI, and (f) single-shot PEPSI. Corresponding spectral arrays (c'-f') display the spectral range from 1.8 to 4 ppm (80 × 51 mm; 300 × 300 DPI).

ing of NAA onto the low-field side of the residual water peak resulted in increased baseline distortion, which increased fitting errors and, as a result, introduced inhomogeneity in the corresponding metabolite maps (Fig. 3e and f).

Fig. 4 shows metabolic images and water-suppressed spectra acquired in an axial supraventricular slice in the human brain with PEPSI, SENSE-PEPSI, LBW-PEPSI, and single-shot PEPSI. The placement of the outer volume suppression slices along the periphery of the brain is shown in Fig. 4a. The mean g-factor (2.32 \pm 0.45) was comparable in magnitude to that in the phantom data (Fig. 4b). The single-shot PEPSI metabolite maps (Fig. 4f) show relatively uniform spatial distributions of Ins, Cho, Cr, and NAA, consistent with the maps obtained with conventional PEPSI and LBW-PEPSI (Fig. 4c and e). However, larger spatial nonuniformity was measured in the singleshot PEPSI data, which is in part due to g-factor-related noise enhancement and spatial aliasing of residual water and peripheral lipid peaks as a result of SENSE reconstruction errors. The single-shot spectrum in Fig. 4f shows clearly identifiable spectral peaks from Ins, Cho, Cr, NAA, as well as multiplet peaks between Ins and Cr at 3.9 ppm, consistent with the data obtained with conventional PEPSI, SENSE-PEPSI, and LBW-PEPSI (Fig. 4c-e). The LBW-PEPSI and single-shot PEPSI spectra show baseline distortion in the vicinity of NAA, which was aliased onto the low-field shoulder of the residual water peak. The slice-averaged spectral line width of all four methods was in a similar range as well (conventional PEPSI 8.7 \pm 3.8,

SENSE-PEPSI 8.1 \pm 3.8, LBW-PEPSI 8.0 \pm 3.9, single-shot PEPSI 10.2 \pm 5.3). Metabolite concentration values for the four methods (Table 1) were in the range reported in our previous study (22). Concentration values measured with SENSE-PEPSI, LBW-PEPSI, and single-shot PEPSI were slightly overestimated due to low SNR, consistent with our previous experience (22). The CRLBs of SENSE-PEPSI, LBW-PEPSI, and single-shot PEPSI were in similar ranges (Table 2).

DISCUSSION

The results of this study demonstrate the feasibility of single-shot 2D MRSI with spectral width large enough to encompass the major singlet resonances in ¹H spectra in the human brain. The methodology uses partial phase encoding interleaved into an echo-planar readout trajectory (30,31) and partial parallel imaging with SENSE reconstruction (25) to strongly accelerate spatial encoding. The achievable spectral width depends on gradient switching performance for interleaving phase encoding and on the spatial-encoding capabilities of the RF coil array for acceleration with parallel imaging, which determines the number of required phase-encoding blips. Large acceleration factors are required for achieving adequate spectral width and spatial resolution, which favors the use of large-scale array coils and high magnetic field strength (35). In this study we have used commercially available array coils, which exhibit relatively large g-factors at the Single-Shot MRSI 545

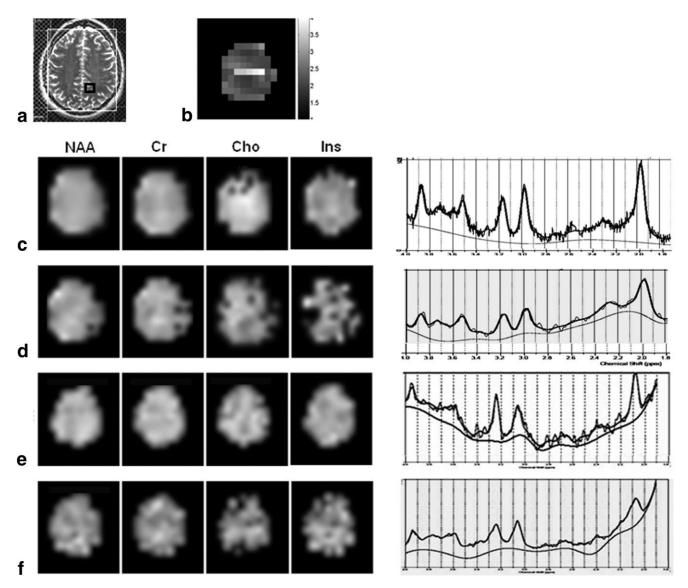


FIG. 4. Comparison of MRSI methods in vivo. (a) MRI of an axial supraventricular slice with positioning of outer volume suppression slices and voxel location of spectra shown below, and (b) *g*-factor map for SENSE reconstruction. (c-f) Spectroscopic images of inositol (lns), choline (Cho), creatine (Cr), and *N*-acetyl-aspartate (NAA) (TR/TE 2 sec/ 11 msec, voxel size 3 cm³) using conventional PEPSI (c), SENSE-PEPSI (d), LBW-PEPSI (e), and single-shot PEPSI (f). The selected spectra on the right display the superimposed LCModel fit and the estimated baseline in the spectral range from 1.8 to 4 ppm (69 × 56 mm; 300 × 300 DPI).

fourfold acceleration rate necessary for single-shot encoding. Alternate reconstruction schemes that avoid separation of even and odd echo data, such as the interlaced Fourier transform, could be used to double the spectral width (36,37). Further acceleration of spectral encoding to increase spectral width can be achieved by generating

multiple spin echoes that are individually phase encoded, as originally described by Duyn et al. (38).

Eq. [1] predicts that the sensitivity per unit time and unit volume of single-shot PEPSI is only slightly smaller than that of SENSE-PEPSI, which is consistent with our experimental data. Interleaving phase-encoding gradient blips

Table 1 Slice-Averaged Metabolite Concentration Values (mmol) in Human Brain (±SD)

	Ins	Cho	Cr	NAA
Conventional PEPSI	5.5 ± 2.2	2.0 ± 0.7	8.6 ± 3.7	11.7 ± 3.8
SENSE-PEPSI	7.2 ± 4.6	2.1 ± 0.8	9.0 ± 2.3	11.8 ± 4.3
LBW-PEPSI	5.5 ± 2.3	2.3 ± 1.5	9.8 ± 3.7	9.9 ± 3.3
Single-shot PEPSI	6.8 ± 4.6	2.4 ± 1.7	14.1 ± 10.9	13.6 ± 8.2

546 Posse et al.

Table 2 Slice-Averaged Cramer-Reo Lower Bound Values (%) in Human Brain

	Ins	Cho	Cr	NAA
Conventional PEPSI	20.4 ± 9.4	11.1 ± 7.2	8.1 ± 4.8	6.0 ± 4.8
SENSE-PEPSI	27.7 ± 8.6	20.0 ± 7.9	14.4 ± 4.7	11.2 ± 6.4
LBW-PEPSI	19.4 ± 12.0	28.1 ± 31.7	12.5 ± 9.2	12.2 ± 6.8
Single-shot PEPSI	22.2 ± 19.2	25.6 ± 28.2	14.4 ± 17.3	14.3 ± 18.6

in single-shot PEPSI results in slightly lower ADC data sampling efficiency (s=77%) as compared with conventional PEPSI and SENSE-PEPSI (s=89%). Theory also predicts that the sensitivity of single-shot PEPSI is similar to that of LBW-PEPSI when taking into account reduced sampling efficiency in the LBW-PEPSI data and g-factor-related noise increases in the single-shot PEPSI data, which can be seen as follows: The extraction of the two echoes at $k_y=0$ in the PSF-PEPSI data to generate the LBW-PEPSI data results in a 2.6-fold decrease in sensitivity per unit time as compared with single-shot PEPSI data, which is largely compensated by the g-factor-related noise enhancement in the single-shot PEPSI data. This is confirmed by our data.

The narrow spectral width at 3-T leads to spectral aliasing of the NAA peak onto the low-field shoulder of the residual water peak. This makes baseline estimation and spectral quantification challenging, particularly for NAA. The presence of overlapping multiplet resonances at the short TE used in this study further complicates spectral fitting. The simulated spectral basis sets in this study did not take into account spectral aliasing, which prevents accurate fitting of multiplet resonances and impairs estimation of the baseline. The LCModel fitting results in vivo in this study thus represent rough estimates of the actual metabolite concentration values. An approximately 10% increase in spectral width could be achieved using triangular instead of rectangular readout gradients at the expense of increased ramp sampling. This would increase the separation of NAA and residual water and thus improve the quantification of NAA. In future studies we will use numerical optimization of the spectral width to minimize information loss due to overlapping spectral peaks and explicitly model basis sets for narrow spectral width to enable quantification of multiplet peaks. Increasing spectral width using array coils with a larger number of coil elements and larger SENSE acceleration factors will further increase the robustness of spectral fitting.

Part of the baseline distortion in the single-shot PEPSI spectra arises from spatial aliasing of residual peripheral lipids signals as a result of imperfections in SENSE reconstruction due to errors in coil sensitivity estimation and intravoxel coil sensitivity variation due to the low spatial resolution of the acquisition. The use of short TE, which we chose to maximize SNR and to compare with our previous short TE studies, increases this baseline distortion. The use the minimum-norm SENSE reconstruction technique would minimize spatial bleeding due to intravoxel coil sensitivity variation (39).

Off-resonance effects in our 3-T data blur the spatial response function of aliased signals originating from outside of the encoded spectral width, as described by Mayer et al. (23). Centering the spectrum at 2.6 ppm using transmitter and receiver offsets (not yet implemented in this study) will reduce spatial blurring for the major singlet resonances, but not for aliased macromolecular and lipid signals. Additional complexity arises from the reordering of the readout trajectories to form even and odd echo data sets: spectral phase accumulation between individual readout segments, such as $1'\ 2',\ 3',\ 4'$, is slightly discontinuous, leading to additional phase modulation in k-space, resulting in broadening of the point spread function. Phase correction in k-space and the use of a first-order phase correction in the spectral domain, as described by Mayer (23) to correct off-resonance effects for different resonances or least-squares reconstruction (24), will result in narrower point spread function.

In conclusion, we have developed an MRSI method that allows collecting a complete spectroscopic image with two spatial dimensions and one spectral dimension in a single signal excitation with sensitivity per unit time and unit volume that is comparable to conventional SENSE-accelerated MRSI. Combining echo-planar readout, interleaved phase encoding, and partial parallel imaging using SENSE enables flexible trade-off between gradient and RF encoding to maximize spectral width and spatial resolution. The method is suitable for applications that require high temporal resolution, such as hyperpolarized MRI, and for reducing motion sensitivity in vivo.

ACKNOWLEDGMENTS

The authors thank Dr. Lawrence Wald for his support and encouragement during the development of parallel high-speed spectroscopic imaging.

REFERENCES

- Ardenkjaer-Larsen JH, Fridlund B, Gram A, Hansson G, Hansson L, Lerche MH, Servin R, Thaning M, Golman K. Increase in signal-to-noise ratio of >10,000 times in liquid-state NMR. Proc Natl Acad Sci USA 2003;100:10158-10163.
- Golman K, in't Zandt R, Thaning M. Real-time metabolic imaging. Proc Natl Acad Sci USA 2006;103:11270–11275.
- Golman K, Zandt RI, Lerche M, Pehrson R, Ardenkjaer-Larsen JH. Metabolic imaging by hyperpolarized ¹³C magnetic resonance imaging for in vivo tumor diagnosis. Cancer Res 2006;66:10855–10860.
- Golman K, Petersson JS. Metabolic imaging and other applications of hyperpolarized ¹³C1. Acad Radiol 2006;13:932–942.
- Posse S, Cuenod CA, Le Bihan D, Human brain: proton diffusion MR spectroscopy. Radiology 1993;188:719–725.
- Pfeuffer J, Tkac I, Gruetter R. Extracellular–intracellular distribution of glucose and lactate in the rat brain assessed noninvasively by diffusionweighted ¹H nuclear magnetic resonance spectroscopy in vivo. J Cereb Blood Flow Metab 2000;20:736–746.
- Valette J, Guillermier M, Besret L, Hantraye P, Bloch G, Lebon V. Isoflurane strongly affects the diffusion of intracellular metabolites, as shown by ¹H nuclear magnetic resonance spectroscopy of the monkey brain. J Cereb Blood Flow Metab 2007;27:588–596. Epub 2006 June 21.

- Le Bihan D, editor. Diffusion and perfusion: magnetic resonance imaging. New York: Raven Press; 1995.
- Posse S, Cuenod, CA, Le Bihan D. Motion artifact compensation in ¹H spectroscopic imaging. J Magn Reson 1993;102:222–227.
- Bito Y, Hirata S, Nabeshima T, Yamamoto E. Echo-planar diffusion spectroscopic imaging. Magn Reson Med 1995;33:69-73.
- Jacobs MA, Barker PB, Bottomley PA, Bhujwalla Z, Bluemke DA. Proton magnetic resonance spectroscopic imaging of human breast cancer: a preliminary study. J Magn Reson Imaging 2004;19:68-75.
- Hu J, Vartanian SA, Xuan Y, Latif Z, Soulen RL. An improved 1H magnetic resonance spectroscopic imaging technique for the human breast: preliminary results. Magn Reson Imaging 2005;23:5716.
- Mansfield P. Multi-planar image formation using NMR spin echoes. J Physics 1977;C10:L55–L58.
- 14. Norris DG, Dreher W. Fast proton spectroscopic imaging using the sliced k-space method. Magn Reson Med 1993;30:641–645.
- Posse S, DeCarli CS, Le Bihan D. Three-dimensional echo-planar MR spectroscopic imaging at short echo times in human brain. Radiology 1994;192:733–738.
- Posse S, Tedeschi G, Risinger R, Ogg R, Bihan DL. High speed 1H spectroscopic imaging in human brain by echo planar spatial-spectral encoding. Magn Reson Med 1995;33:34-40.
- Adalsteinsson E, Irarrazabal P, Spielman DM, Macovski A. Threedimensional spectroscopic imaging with time-varying gradients. Magn Reson Med 1995;33:461–466.
- Guimaraes AR, Baker JR, Jenkins BG, Lee PL, Weisskoff RM, Rosen BR, González RG. Echoplanar chemical shift imaging. Magn Reson Med 1999:41:877–882.
- Ebel A, Soher BJ, Maudsley AA. Assessment of 3D proton MR echoplanar spectroscopic imaging using automated spectral analysis. Magn Reson Med 2001;46:1072–1078.
- Schuster C, Dreher W, Geppert C, Leibfritz D. Fast 3D ¹H spectroscopic imaging at 3 Tesla using spectroscopic missing-pulse SSFP with 3D spatial preselection. Magn Reson Med 2007;57:82–89.
- Otazo R, Mueller B, Ugurbil K, Wald LL, Posse S. Signal-to-noise ratio and spectral line width improvements between 1.5 and 7 Tesla in proton-echo-planar spectroscopic imaging (PEPSI). Magn Reson Med 2006;56:1200-1210.
- Posse S, Otazo R, Caprihan A, Bustillo J, Chen H, Henry P-G, Marjanska M, Gasparovic C, Zuo C, Magnotta V, Mueller B, Mullins P, Renshaw P, Ugurbil K, Lim KO, Alger JR. Proton echo planar spectroscopic imaging of J-coupled resonances in human brain at 3 and 4 Tesla. Magn Reson Med 2007;58:236–244.
- Mayer D, Levin YS, Hurd RE, Glover GH, Spielman DM. Fast metabolic imaging of systems with sparse spectra: application for hyperpolarized ¹³C imaging. Magn Reson Med 2006;56:932–937.

- Levin YS, Mayer D, Yen YF, Hurd RE, Spielman DM. Optimization of fast spiral chemical shift imaging using least squares reconstruction: application for hyperpolarized (13)C metabolic imaging. Magn Reson Med 2007;58:245–252.
- Dydak U, Weiger M, Pruessmann KP, Meier D, Boesiger P. Sensitivityencoded spectroscopic imaging. Magn Reson Med 2001;46:713–722.
- Dydak U, Pruessmann KP, Weiger M, Tsao J, Meier D, Boesiger P. Parallel spectroscopic imaging with spin-echo trains. Magn Reson Med 2003:50:196–200.
- Otazo R, Tsai SY, Lin FH, Posse S. Accelerated short-TE 3D proton echo-planar spectroscopic imaging using 2D-SENSE with a 32-channel array coil. Magn Reson Med 2007;58:1107–1116.
- Lin F-H, Tsai S-Y, Otazo R, Caprihan A, Wald LL, Belliveau JW, Posse S, Sensitivity-encoded (SENSE) proton-echo-planar-spectroscopic imaging (PEPSI) in human brain. Magn Reson Med 2007;57:249–257.
- Tsai, S-Y, Posse S, Otazo R, Lin Y-R, Chung H-W, Lin F-H. Accelerated proton–echo-planar spectroscopy imaging (PEPSI) using GRAPPA with large-N phased-array coils. Magn Reson Med 2008;59:989–998.
- Webb P, Spielman D, Macovski A. A fast spectroscopic imaging method using a blipped phase encode gradient. Magn Reson Med 1989;12:306– 215
- Ebel A, Schuff N. Accelerated 3D echo-planar spectroscopic imaging at 4 tesla using modified blipped phase-encoding. Magn Reson Med 2007; 58:1061–1066.
- Posse S, Otazo R, Tsai S-Y, Lin F-H, Single-shot MR spectroscopic imaging with partial parallel imaging. In: Proceedings of the 15th annual meeting of the ISMRM, Berlin, Germany; 2007. Abstract 49.
- Pohmann R, von Kienlin M, Haase A. Theoretical evaluation and comparison of fast chemical shift imaging methods. J Magn Reson 1997; 129:145–160.
- Provencher SW. Estimation of metabolite concentrations from localized in vivo proton NMR spectra. Magn Reson Med 1993;30:672–679.
- 35. Wiesinger F, Van de Moortele PF, Adriany G, De Zanche N, Ugurbil K, Pruessmann KP. Parallel imaging performance as a function of field strength—an experimental investigation using electrodynamic scaling. Magn Reson Med 2004;52:953–964.
- Metzger G, Hu X. Application of interlaced Fourier transform to echoplanar spectroscopic imaging. J Magn Reson 1997;125:166–170.
- Ebel A, Maudsley AA, Weiner MW, Schuff N. Achieving sufficient spectral bandwidth for volumetric 1H echo-planar spectroscopic imaging at 4 Tesla. Magn Reson Med 2005;54:697–701.
- 38. Duyn JH, Moonen CT. Fast proton spectroscopic imaging of human brain using multiple spin-echoes. Magn Reson Med 1993;30:409-414.
- Sanchez-Gonzalez J, Tsao J, Dydak U, Desco M, Boesiger P, Pruessmann KP. Minimum-norm reconstruction for sensitivity-encoded magnetic resonance spectroscopic imaging. Magn Reson Med 2006;55:287–295.