Week 8: Spatial Point Pattern Analysis II

Species distribution modeling

Poisson process

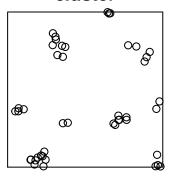
- Homogeneous Poisson process with intensity λ have two properties:
 - -N(A) is Poisson distributed with mean $\lambda |A|$, for all A
 - Condition on N(A), the n points are independent and uniformly distributed in A
- model for complete spatial randomness and null model in a statistical analysis.

Non-Poisson Process

- Poisson cluster processes
 - Hierarchical processes: Parent Poisson processes with offspring point processes
 - Example, Matern cluster process, each parent has $Poisson(\mu)$ number of offsprings uniformly distributed around the parent.
- · Cox processes
 - A Poisson process with a random intensity function
 - Example, log-Gaussian Cox processes (LGCP) in which intensity $\lambda(u)$ is a Gaussian random field.

```
# Materm cluster proccess
cluster <- rMatClust(20, 0.05, 4)
plot(cluster)</pre>
```

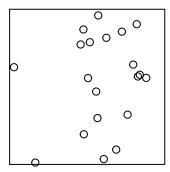
cluster



```
# log-Gaussian Cox procces

lgcp <- rLGCP("exp", 3, var=0.2, scale=.1)
plot(lgcp)</pre>
```

Igcp

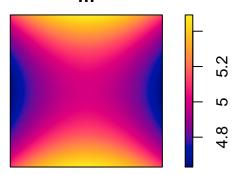


```
# inhomogeneous LGCP with Gaussian covariance function

m \leftarrow as.im(function(x, y){5 - 1.5 * (x - 0.5)^2 + 2 * (y - 0.5)^2}, W=owin())

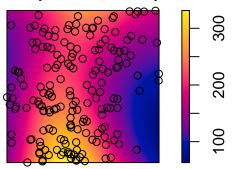
plot(m)
```

m



```
X <- rLGCP("gauss", m, var=0.15, scale =0.5)
plot(attr(X, "Lambda"))
points(X)</pre>
```

attr(X, "Lambda")



Species distribution modeling

Species distribution models (SDMs) estimate the relationship between species records at sites and the environmental and/or spatial characteristics of those sites.

Background data

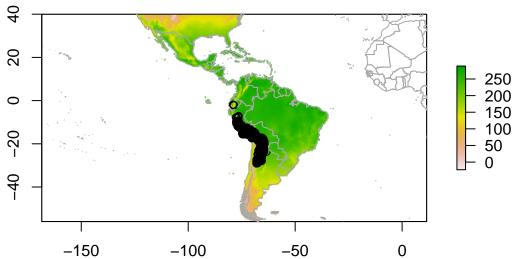
- Background data are not attempting to guess at absence locations, but rather to characterize environments in the study region.
- Background data establishes the environmental domain of the study or regions where a specie should habitat but hasn't, whilst presence data should establish under which conditions a species is more likely to be present than on average.
- A closely related concept is "pseudo-absences", which is also used for generating the non-presence class. In this case, researchers try to guess where absences might occur they may sample the whole region except at presence locations, or they might sample at places unlikely to be suitable for the species.

```
# Random background data
files <- list.files(path=paste(system.file(package="dismo"), '/ex', sep= ''), pattern= 'grd', full.nam
mask <- raster(files[1])</pre>
set.seed(1963)
bg <- randomPoints(mask, 500 )</pre>
# set up the plotting area for two maps
par(mfrow=c(1,2))
plot(!is.na(mask), legend=FALSE)
points(bg, cex=0.5)
# now we repeat the sampling, but limit
# the area of sampling using a spatial extent
e \leftarrow extent(-80, -53, -39, -22)
bg2 <- randomPoints(mask, 50, ext=e)
plot(!is.na(mask), legend=FALSE)
plot(e, add=TRUE, col= ' red ')
points(bg2, cex=0.5)
4
                                            2
                                            0
0
                                            9
   -120
              -80
                        -40
                                               -120
                                                          -80
                                                                    -40
# Pseudo-abasense
file <- paste(system.file(package="dismo"), '/ex/acaule.csv' , sep='')</pre>
ac <- read.csv(file)
coordinates(ac) <- ~lon+lat</pre>
projection(ac)<-CRS('+proj=longlat +datum=WGS84')</pre>
```

```
data(wrld_simpl)
plot(mask)
points(ac)
plot(wrld_simpl, add=TRUE, border='dark grey')

# Generate a cicule centered at each acaule
x <- circles(ac, d=50000, lonlat=TRUE)
pol <- polygons(x)

# Now sample 250 random 'pseudo' location around each acaule sample locations
samp1 <- spsample(pol, 250, type= 'random', iter=25)
points(samp1)</pre>
```



Regression models

```
# Regression models
library(maptools)

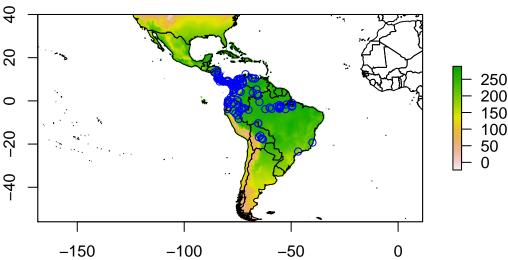
# sample locations
file <- paste(system.file(package="dismo"), "/ex/bradypus.csv", sep="")
bradypus <- read.table(file, header=TRUE, sep= ',')
bradypus <- bradypus[,-1]

# environment conditions
files <- list.files(path=paste(system.file(package="dismo"), '/ex', sep= ''), pattern= 'grd' , full.nam
predictors <- stack(files)

# first layer of the RasterStack
plot(predictors, 1)
#plot(predictors, 1)
# plot(predictors)
# note the "add=TRUE" argument with plot
plot(wrld_simpl, add=TRUE)
# with the points function, "add" is implicit</pre>
```

```
points(bradypus, col= ' blue ')
```

bio1



```
# 'biome' is a categorical data that needs special treatment in
# regression case, let's simply drop it from the predicator stacks.
pred_nf <- dropLayer(predictors, 'biome')</pre>
# Generate background data. To speed up processing, let's restrict the
# predictions to a more restricted area:
ext = extent(-90, -32, -33, 23)
backg <- randomPoints(pred_nf, n=1000, ext=ext, extf = 1.25)</pre>
colnames(backg) = c('lon', 'lat')
backg=as.data.frame(backg)
# Randomly seperate the bradypus observations into 5 groups and use
# four of them as training data and the rest one for validation
group <- kfold(bradypus, 5)</pre>
pres_train <- bradypus[group != 1, ]</pre>
pres_test <- bradypus[group == 1, ]</pre>
group <- kfold(backg, 5)</pre>
backg_train <- backg[group != 1, ]</pre>
backg_test <- backg[group == 1, ]</pre>
# Display the training and validation test data
r = raster(pred_nf, 1)
plot(!is.na(r), col=c(' white ' , ' light grey '), legend=FALSE)
plot(ext, add=TRUE, col= ' red ' , lwd=2)
points(backg_train, pch= ' + ' , cex=1.5, col= ' yellow ')
points(backg_test, pch= ' + ' , cex=1.5, col= ' red ')
points(pres_train, pch= ' + ' , col= ' green ')
points(pres_test, pch= '+' , col= 'blue')
```

```
-150 -100 -50 0
```

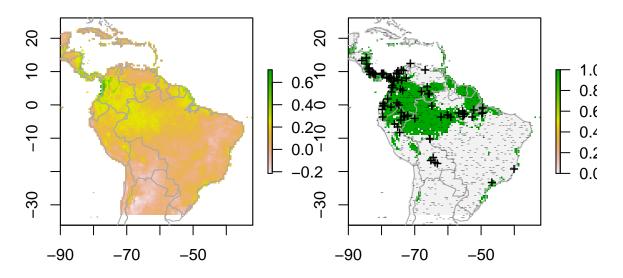
```
# Combine sample locations and environment conditions into one data frame
train <- rbind(pres_train, backg_train)</pre>
pb_train <- c(rep(1, nrow(pres_train)), rep(0, nrow(backg_train)))</pre>
envtrain <- extract(predictors, train)</pre>
envtrain <- data.frame( cbind(pa=pb_train, envtrain) )</pre>
envtrain[,'biome'] = factor(envtrain[,'biome'], levels=1:14)
head(envtrain)
testpres <- data.frame( extract(predictors, pres_test) )</pre>
testbackg <- data.frame( extract(predictors, backg_test) )</pre>
testpres[ ,'biome'] = factor(testpres[ ,'biome'], levels=1:14)
testbackg[ ,'biome'] = factor(testbackg[ ,'biome'], levels=1:14)
\# In\ R , GLM is implemented in the 'glm' function, and the link function and
#error distribution are specified with the 'family' argument. Examples are:
#family = binomial(link = "logit")
#family = qaussian(link = "identity")
#family = poisson(link = "log")
# logistic regression:
gm1 <- glm(pa \sim bio1 + bio5 + bio6 + bio7 + bio8 + bio12 + bio16 + bio17,
family = binomial(link = "logit"), data=envtrain)
summary(gm1)
gm2 \leftarrow glm(pa \sim bio1+bio5 + bio6 + bio7 + bio8 + bio12 + bio16 + bio17,
family = gaussian(link = "identity"), data=envtrain)
# Model evaluation
# ROC curve (reveiver operating characterstic curve) and related AUC
# (area under the curve) are commonly used metrics for classification
# performance. The ROC curve is based on confusion matrix. X is false
# positive rate (FP/(FP+TN)) and Y is true positive rate (TP/(TP+FN)).
# Applying these two rate to different sets of classification
# threshold leads to the ROC curve
ge1=evaluate(testpres, testbackg, gm1)
```

```
ge2=evaluate(testpres, testbackg, gm2)

pg <- predict(predictors, gm2, ext=ext)
par(mfrow=c(1,2))
plot(pg, main='GLM/gaussian, raw values')
plot(wrld_simpl, add=TRUE, border='dark grey')
tr <- threshold(ge2, 'spec_sens')
plot(pg > tr, main='presence/absence')
plot(wrld_simpl, add=TRUE, border='dark grey')
points(pres_train, pch='+')
points(backg_train, pch='-', cex=0.25)
```

GLM/gaussian, raw values

presence/absence



MaxEnt

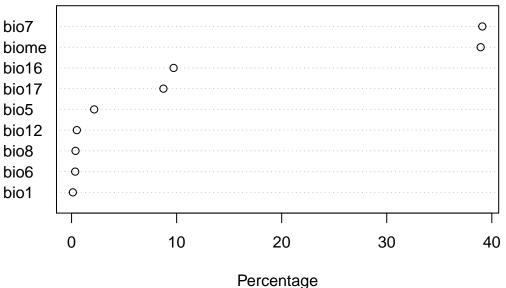
• One of the most widely used SDM framework with consistently competitive predictive performance

```
# MaxEnt, is a seperate package written in JAVA. Package 'dismo'
# provide the interface to Maxent

jar <- paste(system.file(package="dismo"), "/java/maxent.jar", sep= '')

xm <- maxent(predictors, p=pres_train, a= backg_train,factors= 'biome')
plot(xm)</pre>
```

Variable contribution

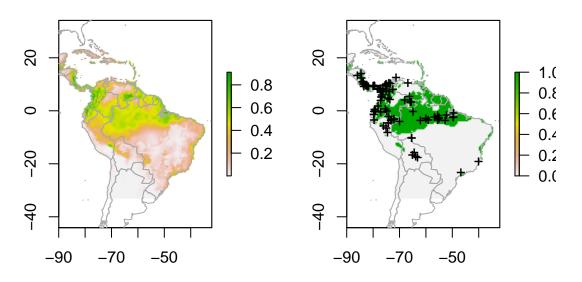


Check the response curve of each environmental conditions response(xm) predicted value predicted value predicted value 0.0 0.0 0.0 0 100 200 300 0 2000 6000 0 500 1500 2500 bio1 bio12 bio16 predicted value predicted value predicted value 0.0 500 1000 1500 100 200 300 -100 0 100 200 0 bio17 bio6 bio5 predicted value predicted value predicted value 0.0 200 300 250 12 100 -50 50 150 6 8 bio7 bio8 biome e <- evaluate(pres_test, backg_test, xm, predictors)</pre> px <- predict(predictors, xm, ext=ext, progress='')</pre> par(mfrow=c(1,2)) plot(px, main='Maxent, raw values') plot(wrld_simpl, add=TRUE, border='dark grey') tr <- threshold(e, 'spec_sens')</pre>

```
plot(px > tr, main='presence/absence')
plot(wrld_simpl, add=TRUE, border='dark grey')
points(pres_train, pch='+')
```

Maxent, raw values

presence/absence



Geographic models

Distance

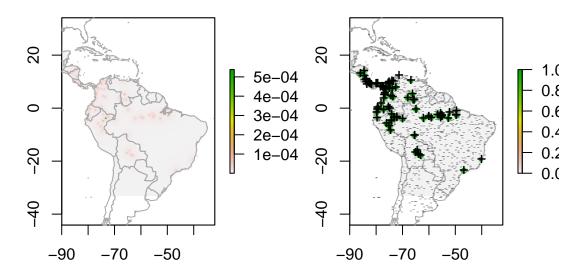
```
# first create a mask to predict to, and to use as a mask
# to only predict to land areas
seamask <- crop(predictors[[1]], ext)
distm <- geoDist(pres_train, lonlat=TRUE)
ds <- predict(seamask, distm, mask=TRUE)
e <- evaluate(distm, p=pres_test, a=backg_test)
e

par(mfrow=c(1,2))
plot(ds, main='Geographic Distance')

plot(wrld_simpl, add=TRUE, border='dark grey')
tr <- threshold(e, 'spec_sens')
plot(ds > tr, main='presence/absence')
plot(wrld_simpl, add=TRUE, border='dark grey')
points(pres_train, pch='+')
points(backg_train, pch='-', cex=0.25)
```

Geographic Distance

presence/absence

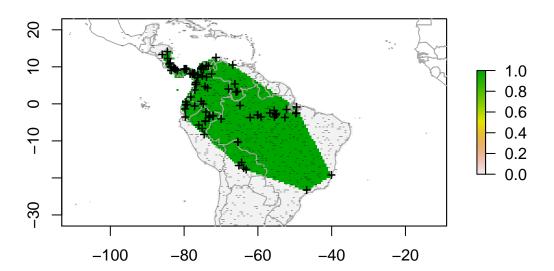


Convex Hull

```
hull <- convHull(pres_train, lonlat=TRUE)
e <- evaluate(hull, p=pres_test, a=backg_test)
e

h <- predict(seamask, hull, mask=TRUE)
plot(h, main='Convex Hull')
plot(wrld_simpl, add=TRUE, border='dark grey')
points(pres_train, pch='+')
points(backg_train, pch='-', cex=0.25)</pre>
```

Convex Hull

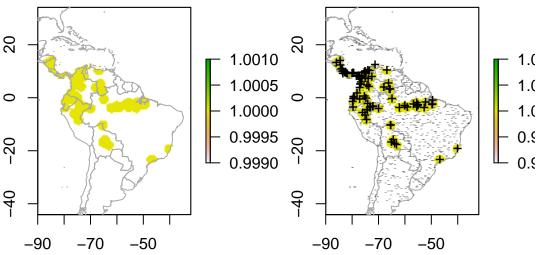


Geographic circles

```
circ <- circles(pres_train, lonlat=TRUE)
pc <- predict(seamask, circ, mask=TRUE)
e <- evaluate(circ, p=pres_test, a=backg_test)
e
par(mfrow=c(1,2))
plot(pc, main='Circles')
plot(wrld_simpl, add=TRUE, border='dark grey')
tr <- threshold(e, 'spec_sens')
plot(pc > tr, main='presence/absence')
plot(wrld_simpl, add=TRUE, border='dark grey')
points(pres_train, pch='+')
points(backg_train, pch='-', cex=0.25)
```

Circles

presence/absence

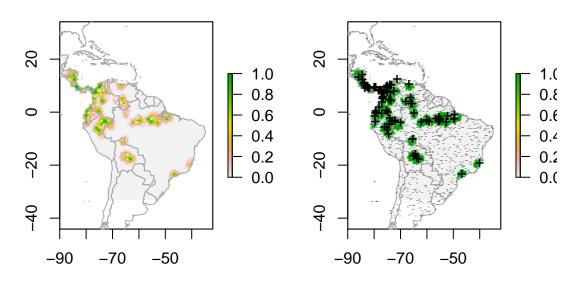


```
idwm <- geoIDW(p=pres_train, a=data.frame(backg_train))
e <- evaluate(idwm, p=pres_test, a=backg_test)
e

iw <- predict(seamask, idwm, mask=TRUE)
par(mfrow=c(1,2))
plot(iw, main='Inv. Dist. Weighted')
plot(wrld_simpl, add=TRUE, border='dark grey')
tr <- threshold(e, 'spec_sens')
pa <- mask(iw > tr, seamask)
plot(pa, main='presence/absence')
plot(wrld_simpl, add=TRUE, border='dark grey')
points(pres_train, pch='+')
points(backg_train, pch='-', cex=0.25)
```

Inv. Dist. Weighted

presence/absence



Voronoi diagram

```
# take a smallish sample of the background training data
va <- data.frame(backg_train[sample(nrow(backg_train), 100), ])
vorm <- voronoiHull(p=pres_train, a=va)
e <- evaluate(vorm, p=pres_test, a=backg_test)
e
vo <- predict(seamask, vorm, mask=T)
plot(vo, main='Voronoi Hull')
plot(wrld_simpl, add=TRUE, border='dark grey')
points(pres_train, pch='+')
points(backg_train, pch='-', cex=0.25)</pre>
```

Voronoi Hull

