Gene normalization

Fay

2022-11-04

Aim:

Normalize gene expression

I have tried the delta delta ct to get the fold gene expression and the delta ct method.

As we are having data from wild infections with unknown status we can't do the delta delta ct method. For that we would have needed control samples - for example uninfected vs infected. I tried to correct for that by getting the mean of every sample for each target gene and using it as a control but that didn't work. The values became non-comparable and then my data became very bad at predicting. By using the delta ct method, I increased the accuracy of my predictions.

Since the data sets are "mixed", wild and laboratory data, plus differnt runs, the delta ct should be enough to get the difference between the housekeeping gene and each sample (representing each run of the sample).

Another problem is that we have measurements for the house keeping genes PPIB and GAPDH. For the field samples, more than 50 % of PPIB is missing, whereas for the laboratory samples 80 % of GAPDH is missing.

I would have liked to use the geometric mean of two housekeeping genes. In this case I am limited to use one for each data set.

https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3489534/#:~:text=We%20suggest%20that%20normalization%20be,imputati In publication: We suggest that normalization be done first followed by missing value imputations. Software tools such as DanteR as well as stand-alone functions in R and Matlab may be used to perform normalization, imputation, significance analysis and visualization.

Load libraries

```
##
## Attaching package: 'mice'
## The following object is masked from 'package:stats':
##
## filter
## The following objects are masked from 'package:base':
##
## cbind, rbind
library(tidyr)
library(tidyverse)
```

```
## -- Attaching packages ------ tidyverse 1.3.2 --
## v ggplot2 3.4.0
                      v dplyr 1.0.10
                   v stringr 1.5.0
v forcats 0.5.2
## v tibble 3.1.8
           2.1.3
## v readr
## v purrr
           0.3.5
## -- Conflicts ----- tidyverse conflicts() --
## x dplyr::filter() masks mice::filter(), stats::filter()
## x dplyr::lag()
                   masks stats::lag()
library(VIM)
## Loading required package: colorspace
## Loading required package: grid
## VIM is ready to use.
## Suggestions and bug-reports can be submitted at: https://github.com/statistikat/VIM/issues
## Attaching package: 'VIM'
## The following object is masked from 'package:datasets':
##
##
      sleep
library(fitdistrplus)
## Loading required package: MASS
## Attaching package: 'MASS'
## The following object is masked from 'package:dplyr':
##
##
      select
##
## Loading required package: survival
library(fitur)
##
## Attaching package: 'fitur'
## The following object is masked from 'package:purrr':
##
##
      rdunif
library(visdat)
library(DESeq2)
## Loading required package: S4Vectors
## Loading required package: stats4
## Loading required package: BiocGenerics
## Attaching package: 'BiocGenerics'
## The following objects are masked from 'package:dplyr':
##
      combine, intersect, setdiff, union
##
```

```
##
## The following objects are masked from 'package:mice':
##
##
       cbind, rbind
##
## The following objects are masked from 'package:stats':
##
##
       IQR, mad, sd, var, xtabs
##
## The following objects are masked from 'package:base':
##
##
       anyDuplicated, append, as.data.frame, basename, cbind, colnames,
##
       dirname, do.call, duplicated, eval, evalq, Filter, Find, get, grep,
##
       grepl, intersect, is.unsorted, lapply, Map, mapply, match, mget,
##
       order, paste, pmax, pmax.int, pmin, pmin.int, Position, rank,
##
       rbind, Reduce, rownames, sapply, setdiff, sort, table, tapply,
##
       union, unique, unsplit, which.max, which.min
##
##
## Attaching package: 'S4Vectors'
##
## The following objects are masked from 'package:dplyr':
##
##
       first, rename
##
## The following object is masked from 'package:tidyr':
##
##
       expand
##
## The following objects are masked from 'package:base':
##
##
       expand.grid, I, unname
##
## Loading required package: IRanges
## Attaching package: 'IRanges'
##
## The following objects are masked from 'package:dplyr':
##
##
       collapse, desc, slice
##
## The following object is masked from 'package:purrr':
##
##
       reduce
## Loading required package: GenomicRanges
## Loading required package: GenomeInfoDb
## Loading required package: SummarizedExperiment
## Loading required package: MatrixGenerics
## Loading required package: matrixStats
##
## Attaching package: 'matrixStats'
##
## The following object is masked from 'package:dplyr':
```

```
##
##
       count
##
##
##
  Attaching package: 'MatrixGenerics'
##
## The following objects are masked from 'package:matrixStats':
##
##
       colAlls, colAnyNAs, colAnys, colAvgsPerRowSet, colCollapse,
##
       colCounts, colCummaxs, colCummins, colCumprods, colCumsums,
##
       colDiffs, colIQRDiffs, colIQRs, colLogSumExps, colMadDiffs,
##
       colMads, colMaxs, colMeans2, colMedians, colMins, colOrderStats,
       colProds, colQuantiles, colRanges, colRanks, colSdDiffs, colSds,
##
##
       colSums2, colTabulates, colVarDiffs, colVars, colWeightedMads,
##
       colWeightedMeans, colWeightedMedians, colWeightedSds,
##
       colWeightedVars, rowAlls, rowAnyNAs, rowAnys, rowAvgsPerColSet,
##
       rowCollapse, rowCounts, rowCummaxs, rowCummins, rowCumprods,
##
       rowCumsums, rowDiffs, rowIQRDiffs, rowIQRs, rowLogSumExps,
##
       rowMadDiffs, rowMads, rowMaxs, rowMeans2, rowMedians, rowMins,
##
       rowOrderStats, rowProds, rowQuantiles, rowRanges, rowRanks,
##
       rowSdDiffs, rowSds, rowSums2, rowTabulates, rowVarDiffs, rowVars,
##
       rowWeightedMads, rowWeightedMeans, rowWeightedMedians,
##
       rowWeightedSds, rowWeightedVars
##
## Loading required package: Biobase
  Welcome to Bioconductor
##
       Vignettes contain introductory material; view with
##
##
       'browseVignettes()'. To cite Bioconductor, see
       'citation("Biobase")', and for packages 'citation("pkgname")'.
##
##
##
  Attaching package: 'Biobase'
##
## The following object is masked from 'package:MatrixGenerics':
##
##
       rowMedians
##
## The following objects are masked from 'package:matrixStats':
##
##
       anyMissing, rowMedians
```

Load data

Import data

```
hm <- read.csv("output_data/1.MICE_cleaned_data.csv")</pre>
```

I only include GAPDH as a housekeeping gene, as PPIB is missing in a large number

```
# Vectors for selecting genes
#Lab genes
# The measurements of IL.12 and IRG6 are done with an other assay and will
```

```
#ignore for now
         <- c("IFNy", "CXCR3", "IL.6", "IL.13", "IL.10",
Gene_lab
                "IL1RN", "CASP1", "CXCL9", "ID01", "IRGM1", "MP0",
                "MUC2", "MUC5AC", "MYD88", "NCR1", "PRF1", "RETNLB", "SOCS1",
                "TICAM1", "TNF") #"IL.12", "IRG6")
Genes wild
           <- c("IFNy", "CXCR3", "IL.6", "IL.13", "IL.10",
                  "IL1RN", "CASP1", "CXCL9", "ID01", "IRGM1", "MP0",
                  "MUC2", "MUC5AC", "MYD88", "NCR1", "PRF1", "RETNLB", "SOCS1",
                  "TICAM1", "TNF") #, "IL.12", "IRG6")
Facs_lab <- c("CD4", "Treg", "Div_Treg", "Treg17", "Th1",</pre>
                    "Div_Th1", "Th17", "Div_Th17", "CD8", "Act_CD8",
                    "Div_Act_CD8", "IFNy_CD4", "IFNy_CD8", "Treg_prop",
                    "IL17A CD4")
Facs_wild <- c( "Treg", "CD4", "Treg17", "Th1", "Th17", "CD8",
                     "Act_CD8", "IFNy_CD4", "IL17A_CD4", "IFNy_CD8")
```

Genes

```
hm$Mouse_ID <- str_replace(hm$Mouse_ID, "_", "")</pre>
field <- hm %>%
 dplyr::filter(origin == "Field")
field <- unique(field)</pre>
genes mouse field <- field %>%
 dplyr::select(c(Mouse_ID, all_of(Genes_wild), GAPDH))
genes field <- genes mouse field %>%
 dplyr::select(-Mouse ID)
#remove rows with only nas
genes_field <- genes_field[,colSums(is.na(genes_field)) < nrow(genes_field)]</pre>
#remove colums with only nas
genes_field <- genes_field[rowSums(is.na(genes_field)) != ncol(genes_field), ]</pre>
genes_mouse_field <- genes_mouse_field[row.names(genes_field), ]</pre>
##select same rows in the first table
field <- field[row.names(genes_field), ]</pre>
#########ab
#select the genes and lab muce
lab <- hm %>%
 dplyr::filter(origin == "Lab", Position == "mLN") #selecting for mln to avoid
# duplicates
lab <- unique(lab)</pre>
gene lab mouse <- lab %>%
 dplyr::select(c(Mouse_ID, "IFNy", "CXCR3", "IL.6", "IL.13", "IL.10",
                                                                                            "IL1RN", "CASP1",
```

```
genes_lab <- gene_lab_mouse[, -1]</pre>
#remove rows with only nas
genes_lab <- genes_lab[,colSums(is.na(genes_lab)) < nrow(genes_lab)]</pre>
#remove colums with only nas
genes_lab <- genes_lab[rowSums(is.na(genes_lab)) != ncol(genes_lab), ]</pre>
genes_lab <- unique(genes_lab)</pre>
#select same rows in the first table
gene_lab_mouse <- gene_lab_mouse[row.names(genes_lab), ]</pre>
##select same rows in the first table
lab <- lab[row.names(genes_lab), ]</pre>
Do we have all the measurements of our housekeeping genes?
#glimpse(hm_selection_g)
#dplyr::select(-Mouse_ID)
# looking at patterns of nas)
#pattern_na <-as.data.frame(md.pattern(field_genes))</pre>
sapply(field %>%
         dplyr::select(c(all_of(Genes_wild), "PPIB", "GAPDH")),
                       function(x) sum(is.na(x)))
```

```
IRGM1
                                                                                     MPO
                                                   CASP1
                                                           CXCL9
                                                                    ID01
##
     IFNy
            CXCR3
                     IL.6
                           IL.13 IL.10
                                          IL1RN
##
       35
              110
                       99
                               28
                                      217
                                               31
                                                     128
                                                              42
                                                                      29
                                                                              11
                                                                                      39
##
     MUC2 MUC5AC
                   MYD88
                             NCR1
                                    PRF1 RETNLB
                                                   SOCS1 TICAM1
                                                                     TNF
                                                                            PPIB
                                                                                  GAPDH
##
       14
               30
                       20
                              126
                                      135
                                             108
                                                      11
                                                             118
                                                                      40
                                                                             212
                                                                                      10
```

```
##
            CXCR3
                     IL.6
                            IL.13
                                    IL.10
                                            IL1RN
                                                    CASP1
                                                           CXCL9
                                                                     ID01
                                                                            IRGM1
                                                                                      MPO
     IFNy
##
        27
                 0
                        12
                                96
                                       13
                                                0
                                                        3
                                                                        0
                                                                                 0
                                                                                       15
     MUC2 MUC5AC
                    MYD88
                                     PRF1 RETNLB
                                                    SOCS1 TICAM1
                                                                      TNF
                                                                                    GAPDH
##
                             NCR1
                                                                             PPIB
##
                 0
                                13
                                        23
                                                0
                                                         0
                                                                 3
                                                                                 1
                                                                                       96
```

Gene normalization

For the field samples the missing values in the house-keeping genes are: PPIB: 212 GAPDH: 10

For the lab samples the missing values in the house-keeping genes are: PPIB: 1 GAPDH: 96

Downlad and install the package for Gene normalization

gene_lab_mouse <- unique(gene_lab_mouse)</pre>

Step 1. Normalize to (REF): DCq = Cq (TAR) – Cq (REF) Step 2. Exponential expression transform: DCq Expression = 2–DCq Step 3. Average replicates and calculate standard deviation Step 4. Normalize to treatment control Step 5. % $KD = (1 - DDCq) \times 100$

```
df <- genes_mouse_field</pre>
######### IL.13
# dct
df <- df %>%
 mutate(IL.13_dct = IL.13 - GAPDH)
# mean of dct
dct_mean <- mean(df$IL.13_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(IL.13_N = 2^- - (IL.13_dct - dct_mean)) \%
  mutate(IL.13_N = round(IL.13_N, digits = 2))
######## I tried writing a nice function but I failed so now I am
 # going t repeat this many many times
######### IFNy
 # dct
df <- df %>%
 mutate(IFNy_dct = IFNy - GAPDH)
# mean of dct
dct_mean <- mean(df$IFNy_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(IFNy_N = 2^ - (IFNy_dct - dct_mean)) %>%
  mutate(IFNy_N = round(IFNy_N, digits = 2))
 ######### CXCR3
df <- df %>%
 mutate(CXCR3_dct = CXCR3 - GAPDH)
# mean of dct
dct_mean <- mean(df$CXCR3_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(CXCR3_N = 2^ - (CXCR3_dct - dct_mean)) %>%
  mutate(CXCR3_N = round(CXCR3_N, digits = 2))
 ######## IL.6
df <- df %>%
 mutate(IL.6_dct = IL.6 - GAPDH)
# mean of dct
dct_mean <- mean(df$IL.6_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(IL.6_N = 2^- - (IL.6_dct - dct_mean)) \%
  mutate(IL.6_N = round(IL.6_N, digits = 2))
```

```
########## IL1RN
df <- df %>%
 mutate(IL1RN_dct = IL1RN - GAPDH)
# mean of dct
dct_mean <- mean(df$IL1RN_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(IL1RN_N = 2^ - (IL1RN_dct - dct_mean)) %>%
  mutate(IL1RN_N = round(IL1RN_N, digits = 2))
 ######### CASP1
df <- df %>%
 mutate(CASP1_dct = CASP1 - GAPDH)
# mean of dct
dct_mean <- mean(df$CASP1_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(CASP1_N = 2^ - (CASP1_dct - dct_mean)) %>%
  mutate(CASP1_N = round(CASP1_N, digits = 2))
 ######### CXCL9
df <- df %>%
 mutate(CXCL9_dct = CXCL9 - GAPDH)
# mean of dct
dct_mean <- mean(df$CXCL9_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(CXCL9_N = 2^ - (CXCL9_dct - dct_mean)) %>%
  mutate(CXCL9_N = round(CXCL9_N, digits = 2))
 ######## ID01
df <- df %>%
 mutate(ID01_dct = ID01 - GAPDH)
# mean of dct
dct_mean <- mean(df$ID01_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(ID01_N = 2^- - (ID01_dct - dct_mean)) \%
  mutate(ID01_N = round(ID01_N, digits = 2))
 ######### IRGM1
df <- df %>%
 mutate(IRGM1_dct = IRGM1 - GAPDH)
# mean of dct
dct_mean <- mean(df$IRGM1_dct, na.rm = TRUE)</pre>
```

```
#fold gene expression
df <- df %>%
  mutate(IRGM1_N = 2^ - (IRGM1_dct - dct_mean)) %>%
  mutate(IRGM1_N = round(IRGM1_N, digits = 2))
  ######## MPO
df <- df %>%
 mutate(MPO_dct = MPO - GAPDH)
# mean of dct
dct_mean <- mean(df$MPO_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(MPO_N = 2^- - (MPO_dct - dct_mean)) \%
  mutate(MPO_N = round(MPO_N, digits = 2))
  ######## MUC2
df <- df %>%
 mutate(MUC2_dct = MUC2 - GAPDH)
# mean of dct
dct_mean <- mean(df$MUC2_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(MUC2_N = 2^ - (MUC2_dct - dct_mean)) %>%
  mutate(MUC2_N = round(MUC2_N, digits = 2))
  ######### MUC5AC
df <- df %>%
 mutate(MUC5AC_dct = MUC5AC - GAPDH)
# mean of dct
dct_mean <- mean(df$MUC5AC_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(MUC5AC_N = 2^ - (MUC5AC_dct - dct_mean)) %>%
  mutate(MUC5AC_N = round(MUC5AC_N, digits = 2))
  ######## MYD88
df <- df %>%
 mutate(MYD88_dct = MYD88 - GAPDH)
# mean of dct
dct_mean <- mean(df$MYD88_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(MYD88_N = 2^- - (MYD88_dct - dct_mean)) \%
  mutate(MYD88_N = round(MYD88_N, digits = 2))
 ######### NCR1
df <- df %>%
 mutate(NCR1_dct = NCR1 - GAPDH)
# mean of dct
dct_mean <- mean(df$NCR1_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
```

```
mutate(NCR1_N = 2^- - (NCR1_dct - dct_mean)) \%
  mutate(NCR1_N = round(NCR1_N, digits = 2))
  ######## PRF1
df <- df %>%
 mutate(PRF1_dct = PRF1 - GAPDH)
# mean of dct
dct_mean <- mean(df$PRF1_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(PRF1_N = 2^ - (PRF1_dct - dct_mean)) %>%
  mutate(PRF1_N = round(PRF1_N, digits = 2))
  ######## RETNLB
df <- df %>%
 mutate(RETNLB_dct = RETNLB - GAPDH)
# mean of dct
dct_mean <- mean(df$RETNLB_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(RETNLB_N = 2^ - (RETNLB_dct - dct_mean)) %>%
  mutate(RETNLB_N = round(RETNLB_N, digits = 2))
  ######### SOCS1
df <- df %>%
 mutate(SOCS1_dct = SOCS1 - GAPDH)
# mean of dct
dct_mean <- mean(df$SOCS1_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(SOCS1_N = 2^ - (SOCS1_dct - dct_mean)) %>%
  mutate(SOCS1_N = round(SOCS1_N, digits = 2))
 ######### TICAM1
df <- df %>%
 mutate(TICAM1_dct = TICAM1 - GAPDH)
# mean of dct
dct_mean <- mean(df$TICAM1_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
  mutate(TICAM1_N = 2^ - (TICAM1_dct - dct_mean)) %>%
  mutate(TICAM1_N = round(TICAM1_N, digits = 2))
 ######### TNF
df <- df %>%
 mutate(TNF_dct = TNF - GAPDH)
# mean of dct
dct_mean <- mean(df$TNF_dct, na.rm = TRUE)</pre>
#fold gene expression
df <- df %>%
```

```
mutate(TNF_N = 2^ - (TNF_dct - dct_mean)) %>%
mutate(TNF_N = round(TNF_N, digits = 2))

df -> df_field
```

Genes Lab

```
############ lab
 # select first the field samples
df lab <- gene lab mouse
  ######### IFNy
 df_lab <- df_lab %>%
 mutate(IFNy_dct = IFNy - PPIB)
# mean of dct
dct_mean <- mean(df_lab$IFNy_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(IFNy_N = 2^ - (IFNy_dct - dct_mean)) %>%
  mutate(IFNy_N = round(IFNy_N, digits = 2))
 ######### CXCR3
 df_lab <- df_lab %>%
 mutate(CXCR3_dct = CXCR3 - PPIB)
# mean of dct
dct_mean <- mean(df_lab$CXCR3_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(CXCR3_N = 2^ - (CXCR3_dct - dct_mean)) %>%
  mutate(CXCR3_N = round(CXCR3_N, digits = 2))
 ######### IL.6
  df_lab <- df_lab %>%
 mutate(IL.6_dct = IL.6 - PPIB)
# mean of dct
dct_mean <- mean(df_lab$IL.6_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(IL.6_N = 2^- - (IL.6_dct - dct_mean)) \%
  mutate(IL.6_N = round(IL.6_N, digits = 2))
 ######## IL.13
  df lab <- df lab %>%
 mutate(IL.13_dct = IL.13 - PPIB)
# mean of dct
dct_mean <- mean(df_lab$IL.13_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
```

```
mutate(IL.13_N = 2^- - (IL.13_dct - dct_mean)) \%
   mutate(IL.13_N = round(IL.13_N, digits = 2))
 ######### IL1RN
  df_{ab} \leftarrow df_{ab} \%
 mutate(IL1RN_dct = IL1RN - PPIB)
# mean of dct
dct mean <- mean(df lab$IL1RN dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(IL1RN_N = 2^ - (IL1RN_dct - dct_mean)) %>%
  mutate(IL1RN_N = round(IL1RN_N, digits = 2))
 ######### CASP1
  df_lab <- df_lab %>%
 mutate(CASP1_dct = CASP1 - PPIB)
# mean of dct
dct_mean <- mean(df_lab$CASP1_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(CASP1_N = 2^ - (CASP1_dct - dct_mean)) %>%
  mutate(CASP1_N = round(CASP1_N, digits = 2))
 ######### CXCL9
  df_lab <- df_lab %>%
 mutate(CXCL9_dct = CXCL9 - PPIB)
# mean of dct
dct_mean <- mean(df_lab$CXCL9_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(CXCL9_N = 2^ - (CXCL9_dct - dct_mean)) %>%
  mutate(CXCL9_N = round(CXCL9_N, digits = 2))
  ######## ID01
  df_lab <- df_lab %>%
 mutate(ID01_dct = ID01 - PPIB)
# mean of dct
dct_mean <- mean(df_lab$ID01_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(ID01_N = 2^- - (ID01_dct - dct_mean)) \%
  mutate(ID01_N = round(ID01_N, digits = 2))
  ######### IRGM1
  df_lab <- df_lab %>%
 mutate(IRGM1_dct = IRGM1 - PPIB)
# mean of dct
dct_mean <- mean(df_lab$IRGM1_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(IRGM1_N = 2^ - (IRGM1_dct - dct_mean)) %>%
```

```
mutate(IRGM1_N = round(IRGM1_N, digits = 2))
  ######## MPO
  df_lab <- df_lab %>%
  mutate(MPO_dct = MPO - PPIB)
# mean of dct
dct_mean <- mean(df_lab$MPO_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(MPO_N = 2^- - (MPO_dct - dct_mean)) %>%
  mutate(MPO_N = round(MPO_N, digits = 2))
  ######## MUC2
  df_lab <- df_lab %>%
 mutate(MUC2_dct = MUC2 - PPIB)
# mean of dct
dct_mean <- mean(df_lab$MUC2_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(MUC2_N = 2^ - (MUC2_dct - dct_mean)) %>%
  mutate(MUC2_N = round(MUC2_N, digits = 2))
  ######### MUC5AC
  df_lab <- df_lab %>%
 mutate(MUC5AC dct = MUC5AC - PPIB)
# mean of dct
dct_mean <- mean(df_lab$MUC5AC_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(MUC5AC_N = 2^ - (MUC5AC_dct - dct_mean)) %>%
  mutate(MUC5AC_N = round(MUC5AC_N, digits = 2))
  ######### MYD88
  df_lab <- df_lab %>%
 mutate(MYD88_dct = MYD88 - PPIB)
# mean of dct
dct_mean <- mean(df_lab$MYD88_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(MYD88_N = 2^- - (MYD88_dct - dct_mean)) %>%
  mutate(MYD88_N = round(MYD88_N, digits = 2))
  ######## NCR1
  df_lab <- df_lab %>%
 mutate(NCR1_dct = NCR1 - PPIB)
# mean of dct
dct_mean <- mean(df_lab$NCR1_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(NCR1_N = 2^ - (NCR1_dct - dct_mean)) %>%
  mutate(NCR1_N = round(NCR1_N, digits = 2))
```

```
######### PRF1
  df_lab <- df_lab %>%
 mutate(PRF1 dct = PRF1 - PPIB)
# mean of dct
dct_mean <- mean(df_lab$PRF1_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(PRF1 \ N = 2^- - (PRF1 \ dct - dct \ mean)) \%
  mutate(PRF1_N = round(PRF1_N, digits = 2))
  ######## RETNLB
  df_lab <- df_lab %>%
 mutate(RETNLB_dct = RETNLB - PPIB)
# mean of dct
dct_mean <- mean(df_lab$RETNLB_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(RETNLB_N = 2^ - (RETNLB_dct - dct_mean)) %>%
  mutate(RETNLB_N = round(RETNLB_N, digits = 2))
  ######### SOCS1
  df_lab <- df_lab %>%
 mutate(SOCS1_dct = SOCS1 - PPIB)
# mean of dct
dct_mean <- mean(df_lab$SOCS1_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(SOCS1_N = 2^- - (SOCS1_dct - dct_mean)) \%
  mutate(SOCS1_N = round(SOCS1_N, digits = 2))
  ######## TICAM1
  df_lab <- df_lab %>%
 mutate(TICAM1_dct = TICAM1 - PPIB)
# mean of dct
dct_mean <- mean(df_lab$TICAM1_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
  mutate(TICAM1 N = 2^{-} - (TICAM1 dct - dct mean)) \%
  mutate(TICAM1_N = round(TICAM1_N, digits = 2))
 ######### TNF
  df_lab <- df_lab %>%
 mutate(TNF_dct = TNF - PPIB)
# mean of dct
dct_mean <- mean(df_lab$TNF_dct, na.rm = TRUE)</pre>
#fold gene expression
df_lab <- df_lab %>%
   mutate(TNF_N = 2^ - (TNF_dct - dct_mean)) %>%
  mutate(TNF_N = round(TNF_N, digits = 2))
```

#now join the data again

without the housekeeping genes

First clean and remove the non normalizes genes

```
#df_lab <- df_lab %>%
# dplyr::select(-c("IFNy", "CXCR3", "IL.6", "IL.13", "IL.10",
#"IL1RN","CASP1", "CXCL9", "ID01", "IRGM1", "MP0", "MUC2", "MUC5AC", "MYD88",
#"NCR1", "PRF1", "RETNLB", "SOCS1", "TICAM1", "TNF", PPIB, contains("_dct")))
# remove ending _N
#df lab <- df lab %>%
# rename with(~str remove(.x, " N"))
#df field <- df field %>%
# dplyr::select(-c(all_of(Genes_wild), GAPDH, contains("_dct")))
# remove ending N
#df field <- df field %>%
\# rename_with(~str_remove(.x, "_N"))
######################## let's try just using DCT
df_lab <- df_lab %>%
dplyr::select(-c("IFNy", "CXCR3", "IL.6", "IL.13", "IL.10", "IL1RN", "CASP1",
                  "CXCL9", "IDO1", "IRGM1", "MPO", "MUC2", "MUC5AC", "MYD88",
                  "NCR1", "PRF1", "RETNLB", "SOCS1", "TICAM1", "TNF", PPIB,
                  contains("_N")))
# remove ending _dct
df lab <- df lab %>%
 rename_with(~str_remove(.x, "_dct"))
df field <- df field %>%
  dplyr::select(-c("IFNy", "CXCR3", "IL.6", "IL.13", "IL.10", "IL1RN", "CASP1",
                  "CXCL9", "IDO1", "IRGM1", "MPO", "MUC2", "MUC5AC", "MYD88",
                  "NCR1", "PRF1", "RETNLB", "SOCS1", "TICAM1", "TNF", GAPDH,
                  contains("_N")))
# remove ending
df_field <- df_field %>%
 rename_with(~str_remove(.x, "_dct"))
# add the new genes to the complete data sets
lab <- lab %>%
dplyr::select(-c("IFNy", "CXCR3", "IL.6", "IL.13", "IL.10", "IL1RN", "CASP1",
               "CXCL9", "IDO1", "IRGM1", "MPO", "MUC2", "MUC5AC", "MYD88",
               "NCR1", "PRF1", "RETNLB", "SOCS1", "TICAM1", "TNF")) %>%
 left_join(df_lab, by = "Mouse_ID")
field <- field %>%
dplyr::select(-c("IFNy", "CXCR3", "IL.6", "IL.13", "IL.10", "IL1RN", "CASP1",
               "CXCL9", "IDO1", "IRGM1", "MPO", "MUC2", "MUC5AC", "MYD88",
```

```
"NCR1", "PRF1", "RETNLB", "SOCS1", "TICAM1", "TNF")) %>%
left_join(df_field, by = "Mouse_ID")

hm_norm <- rbind(lab, field)

##save the imputed data
write.csv(hm_norm, "output_data/2.1.norm_MICE_data_set.csv", row.names = FALSE)</pre>
```