

1. Analysis

Fay

2022-09-15

GAPDH HKG

B-actin HKG

Ppia HKG

Ppip HKG

CDC42 HKG susceptible to DNA contamination

Housekeeping genes selected: GAPDH and PPIB

Relm-b mucosal defense factor (goblet cells)

Muc2 the major secretory mucin within the gastrointestinal tract

TFF3 mucosal defense factor (goblet cells)

Muc5ac similar to MUC2, produced by surface goblet cells

NKp46 NK marker

F4/80 macrophage marker (distinguish by immune response trend)

Mpo myeloperoxidase in Neutrophils

MyD88 TLR protein, NF-kB IRAK protein, inflammation marker by TLR MyD88-Dependent Pathway
caspase-1 inflammasome marker (IL-1b and IL-18 production)

IL-1Ra natural IL-1b antagonist for infection control (if not increase in Tregs is seen)

CXCL9, immune cell migration marker + Th1 activator (confirm FACS)

CXCR3, CXCL9 and CXCL11 receptor

IL-6 TNF inhibitor,

IL-12ra T-cell marker Th1

IFN-γ compare with IFN-γ producing cells and IFN-γ ELISAs, should correlate with PRF1, NKp46 and F4/80.

One of these cell types just have to be doing the job!

IRG6A autonomous cell defense (opsonization)

TNF-α upregulated in eimeria but not well explained. Could be present and driving infection where IFN-γ isn't

IL-17 in case IFN-γ isn't coming up but pathogenicity is

TRIF Type I IFN production TRIF Dependent Pathway

Socs1 JAK/STAT signaling pathway, proinflammatory regulating + T-cell differentiation, could explain severity

IDO1 DC, monocyte and MC protein regulating T-cell activity

Prf1 perforin, should be dominant in primary infections, but must be correlated between T-cell and NK cell expressions

CD56 CD56bright = more cytokine producing NKs, CD56dim = more direct cytotoxic killing

IL-4

IL-13

IL-10

Load libraries

Import data

Challenge = experimental challenge infection data

SOTA = State of the Art, of wild data

```
Challenge <- read.csv("https://raw.githubusercontent.com/derele/Eimeria_Lab/master/data_products/Challenge")
```

```
SOTA <- read.csv("https://raw.githubusercontent.com/derele/Mouse_Eimeria_Field/master/data_products/SOTA")
```

```
# Vectors for selecting genes
```

```
#Lab genes
```

```
# The measurements of IL.12 and IRG6 are done with an other assay and will
```

```
#ignore for now
```

```
Gene_lab <- c("IFNy", "CXCR3", "IL.6", "IL.10", "IL.13", "IL.10", "IL.13",  
             "IL1RN", "CXCR3", "CASP1", "CXCL9", "IDO1", "IRGM1", "MPO",  
             "MUC2", "MUC5AC", "MYD88", "NCR1", "PRF1", "RETNLB", "SOCS1",  
             "TICAM1", "TNF") #"IL.12", "IRG6")
```

```
Genes_wild <- c("IFNy", "IL.12", "IRG6", "CXCR3", "IL.6", "GBP2",  
              "IL.10", "IL.13", "IL.10", "IL.13", "IL1RN",  
              "CXCR3", "CASP1", "CXCL9",  
              "IDO1", "IRGM1", "MPO", "MUC2", "MUC5AC", "MYD88",  
              "NCR1", "PRF1", "RETNLB", "SOCS1", "TICAM1", "TNF")
```

```
Facs_lab <- c("Position", "CD4", "Treg", "Div_Treg", "Treg17", "Th1",  
            "Div_Th1", "Th17", "Div_Th17", "CD8", "Act_CD8",  
            "Div_Act_CD8", "IFNy_CD4", "IFNy_CD8", "Treg_prop",  
            "IL17A_CD4")
```

```
Facs_wild <- c("Treg", "CD4", "Treg17", "Th1", "Th17", "CD8",  
             "Act_CD8", "IFNy_CD4", "IL17A_CD4", "IFNy_CD8")
```

Cleaning

Challenge

```
## Adding missing grouping variables: `infection`
```

```
## Joining, by = c("infection", "EH_ID")
```

Join wild and lab data

```
## [1] "CASP1"      "CXCL9"      "CXCR3"      "IDO1"      "IFNy"      "IL.10"  
## [7] "IL1RN"     "IL.6"      "IRGM1"     "MPO"      "MUC2"      "MUC5AC"  
## [13] "MYD88"     "NCR1"      "PPIB"      "PRF1"     "RETNLB"    "SOCS1"  
## [19] "TICAM1"    "TNF"      "GAPDH"     "IL.13"    "CD4"      "Treg"  
## [25] "Treg17"    "Th1"      "Th17"     "CD8"      "Act_CD8"   "IFNy_CD4"  
## [31] "IFNy_CD8"  "IL17A_CD4"
```

```
## [1] "Address"      "Aspiculuris_sp"  
## [3] "batch"        "Body_Length"
```

##	[5]	"Body_Weight"	"Btk"
##	[7]	"Caecum"	"CASP1_N"
##	[9]	"Catenotaenia_pusilla"	"challenge_infection"
##	[11]	"counter"	"Ct.Eimeria"
##	[13]	"Ct.Mus"	"CXCL9_N"
##	[15]	"CXCR3_N"	"Date_count"
##	[17]	"death"	"delta"
##	[19]	"delta_ct_cewe_MminusE"	"dilution"
##	[21]	"Dissection_Date"	"Div_Act_CD8"
##	[23]	"Div_Th1"	"Div_Th17"
##	[25]	"Div_Treg"	"dpi"
##	[27]	"dpi_max"	"Ectoparasites_Logical"
##	[29]	"EH_ID"	"Eim_MC"
##	[31]	"eimeriaSpecies"	"end_rel_weight"
##	[33]	"Es1"	"Es1C"
##	[35]	"experiment"	"FEC_Eim_Ct"
##	[37]	"feces_weight"	"Feces_Weight"
##	[39]	"Fleas"	"Gpd1"
##	[41]	"Gpd1C"	"Heligmosomoides_polygurus"
##	[43]	"Heterakis_sp"	"HI"
##	[45]	"HI_NLoci"	"Host"
##	[47]	"hybrid_status"	"Hymenolepis_sp"
##	[49]	"Idh1"	"Idh1C"
##	[51]	"ID01_N"	"IFNy_CEW"
##	[53]	"IFNy_FEC"	"IFNy_MES"
##	[55]	"IFNy_N"	"IL.10_N"
##	[57]	"IL.12"	"IL.12A"
##	[59]	"IL.12A_N"	"IL.13_N"
##	[61]	"IL.17A"	"IL.17A_N"
##	[63]	"IL.6_N"	"IL1RN_N"
##	[65]	"ILWE_Crypto_Ct"	"ILWE_DNA_Content_ng.microliter"
##	[67]	"infection"	"infection_history"
##	[69]	"IRG6"	"IRGM1_N"
##	[71]	"labels"	"Latitude"
##	[73]	"Left_Embryo"	"Left_Epididymis"
##	[75]	"Left_Ovarium_Weight"	"Left_Testis"
##	[77]	"Liver"	"Longitude"
##	[79]	"Mastophorus_muris"	"max_dpi"
##	[81]	"max_OOC"	"max_WL"
##	[83]	"MC.Eimeria"	"MC.Eimeria.FEC"
##	[85]	"MCs"	"mean_neubauer"
##	[87]	"Mouse_ID"	"mouse_strain"
##	[89]	"Mpi"	"MpiC"
##	[91]	"MPO_N"	"mtBamH"
##	[93]	"MUC2_N"	"MUC5AC_N"
##	[95]	"MYD88_N"	"N_oocysts_sq1"
##	[97]	"N_oocysts_sq2"	"N_oocysts_sq3"
##	[99]	"N_oocysts_sq4"	"N_oocysts_sq5"
##	[101]	"N_oocysts_sq6"	"N_oocysts_sq7"
##	[103]	"N_oocysts_sq8"	"Ncells"
##	[105]	"NCR1_N"	"Np"
##	[107]	"NpC"	"O04sq"
##	[109]	"OOC"	"Oocyst_Predict_Crypto"
##	[111]	"oocyst_sq1"	"oocyst_sq2"

```
## [113] "oocyst_sq3"           "oocyst_sq4"
## [115] "OPG"                  "OPG_0"
## [117] "Parasite_challenge"   "Parasite_primary"
## [119] "PBS_dil_in_mL"        "Position"
## [121] "PRF1_N"               "primary_infection"
## [123] "Region"               "relative_weight"
## [125] "RETNLB_N"             "Right_Embryo"
## [127] "Right_Ovary_Weight"    "Right_Testis"
## [129] "Seminal_Vesicles_Weight" "Sex"
## [131] "SOCS1_N"              "Sod1"
## [133] "Sod1C"                "Sperm"
## [135] "Spleen"               "Status"
## [137] "Syap1"                "Syphacia_sp"
## [139] "Taenia_sp"            "Tail_Length"
## [141] "TICAM1_N"             "Ticks"
## [143] "TNF_N"                "Trap_Date"
## [145] "Treg_prop"            "Trichuris_muris"
## [147] "Tsx"                  "weight"
## [149] "weight_dpi0"          "Worms_presence"
## [151] "X332"                 "X347"
## [153] "X65"                  "Y"
## [155] "Year"                 "YNPAR"
## [157] "Zfy2"
```

Lab

Heatmap on lab gene expression

```
gene <- as_tibble(data) %>%
  dplyr::filter(origin == "Lab", infection == "challenge", dpi == dpi_max) %>%
  dplyr::group_by(Mouse_ID) %>%
  dplyr::select(c(Mouse_ID, MC.Eimeria, all_of(Gene_lab)))

gene <- unique(gene)

# turn the data frame into a matrix and transpose it. We want to have each cell
# type as a row name
gene <- t(as.matrix(gene %>%
  dplyr::select(c(Mouse_ID, all_of(Gene_lab))))))

#switch the matrix back to a data frame format
gene <- as.data.frame(gene)

# turn the first row into column names
gene %>%
  row_to_names(row_number = 1) -> heatmap_data

table(rowSums(is.na(heatmap_data)) == nrow(heatmap_data))

##
## FALSE TRUE
##    17    3
```

```

# turn the columns to numeric other wise the heatmap function will not work
heatmap_data[] <- lapply(heatmap_data, function(x) as.numeric(as.character(x)))

# remove columns with only NAs
heatmap_data <- Filter(function(x)!all(is.na(x)), heatmap_data)

#remove rows with only NAs
heatmap_data <- heatmap_data[, colSums(is.na(heatmap_data)) !=
                                nrow(heatmap_data)]

#Prepare the annotation data frame
annotation_df <- as_tibble(data) %>%
  dplyr::filter(origin == "Lab", infection == "challenge", dpi == dpi_max)%>%
  dplyr::group_by(Mouse_ID) %>%
  dplyr::select(c("Mouse_ID", "Parasite_challenge", "infection_history",
                  "mouse_strain", "max_WL"))

annotation_df <- unique(annotation_df) %>%
  dplyr::filter(Mouse_ID %in% colnames(heatmap_data))

annotation_df <- as.data.frame(annotation_df)

### Prepare the annotation columns for the heatmap
rownames(annotation_df) <- annotation_df$Mouse_ID

# Match the row names to the heatmap data frame
rownames(annotation_df) <- colnames(heatmap_data)

#remove the unnecessary column
annotation_df <- annotation_df %>% dplyr::select(-Mouse_ID, )

```

Heatmap on lab gene expression data:

