

FACS_analysis

Fay

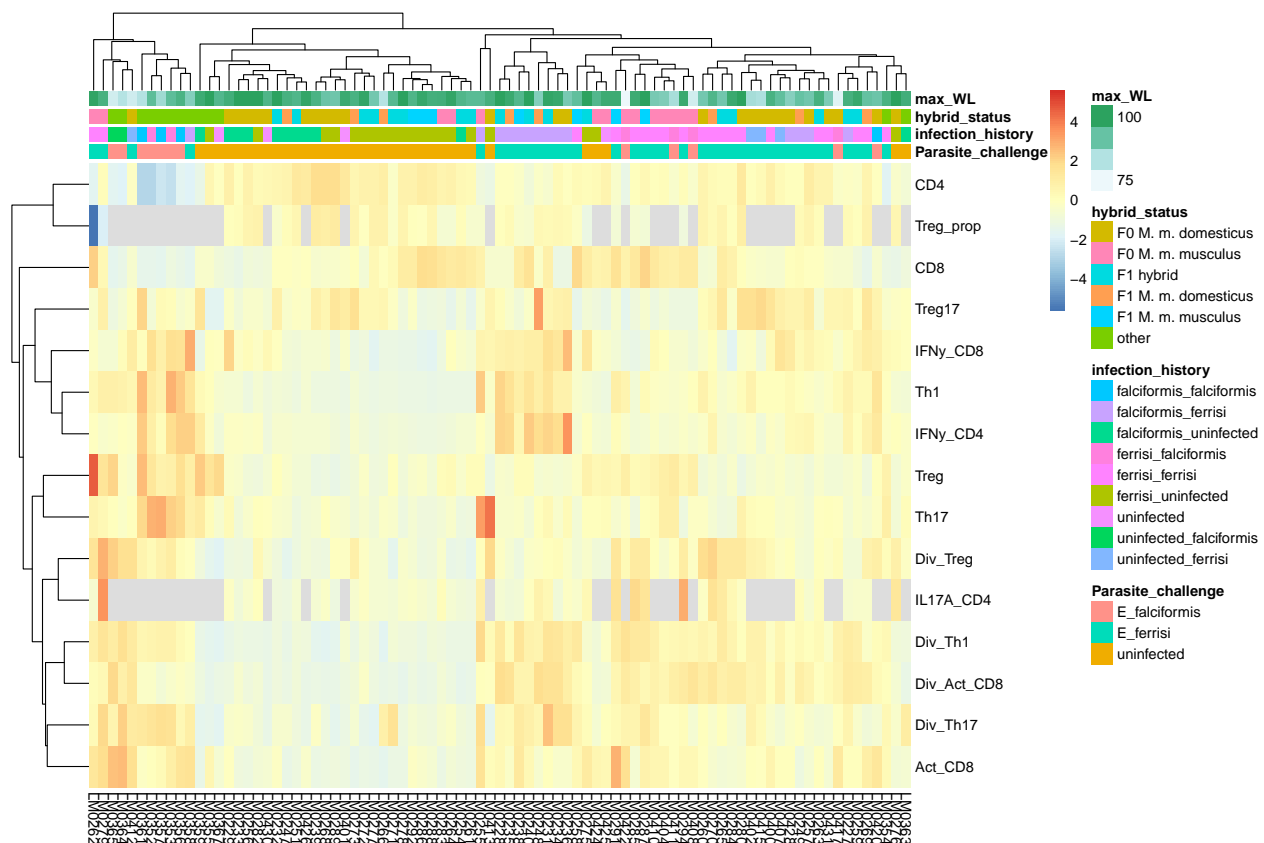
2022-05-18

1. facs expression in the laboratory infections - Heatmap

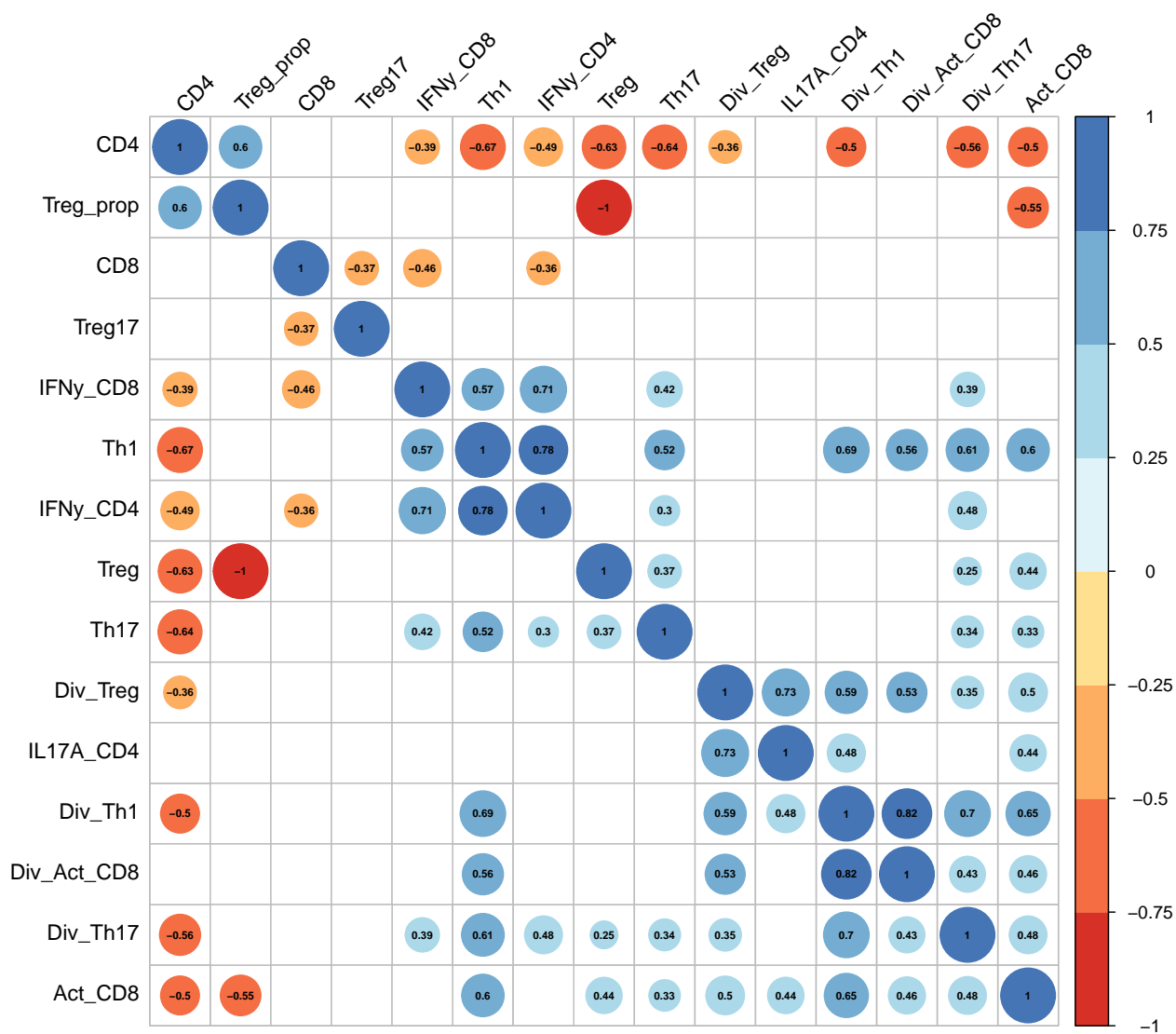
FACS

```
heatmap_data %>%
```

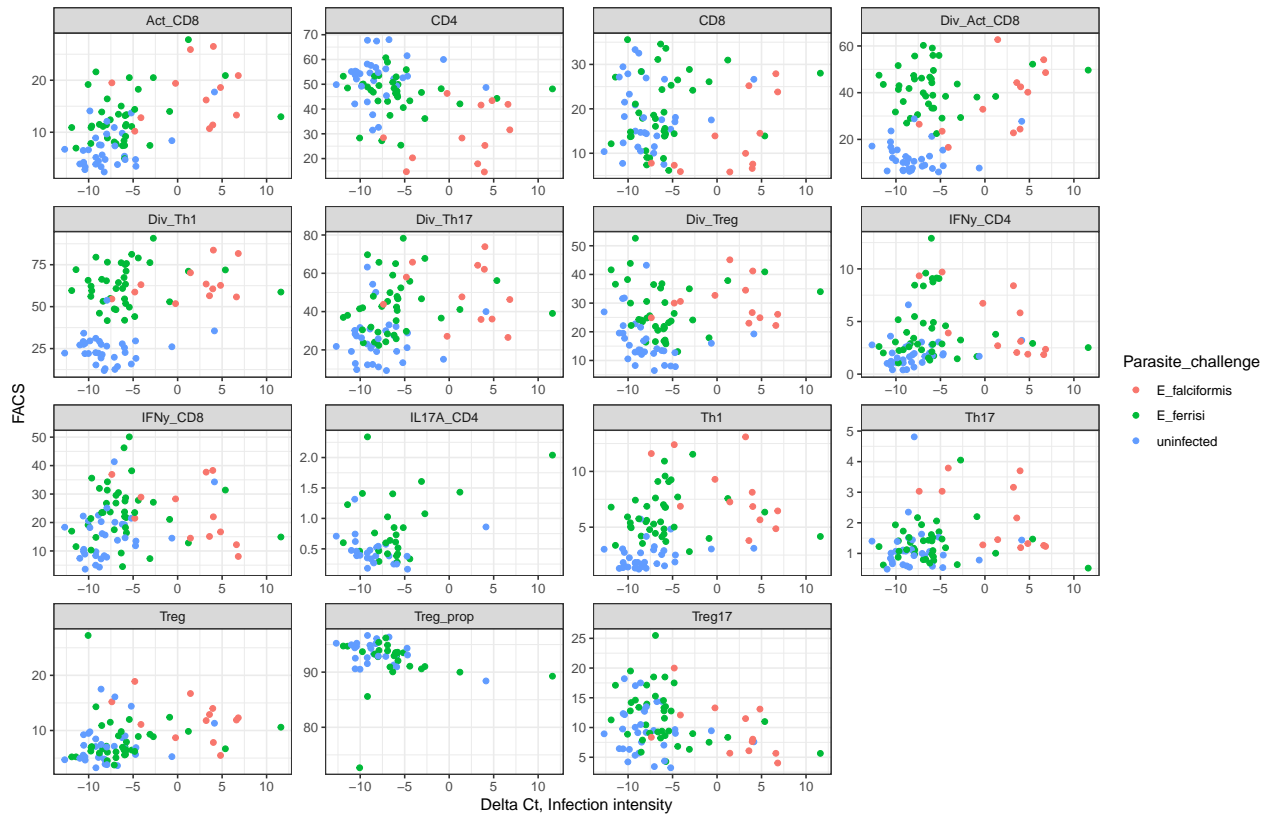
```
phemap(annotation_col = annotation_df, scale = "row")
```



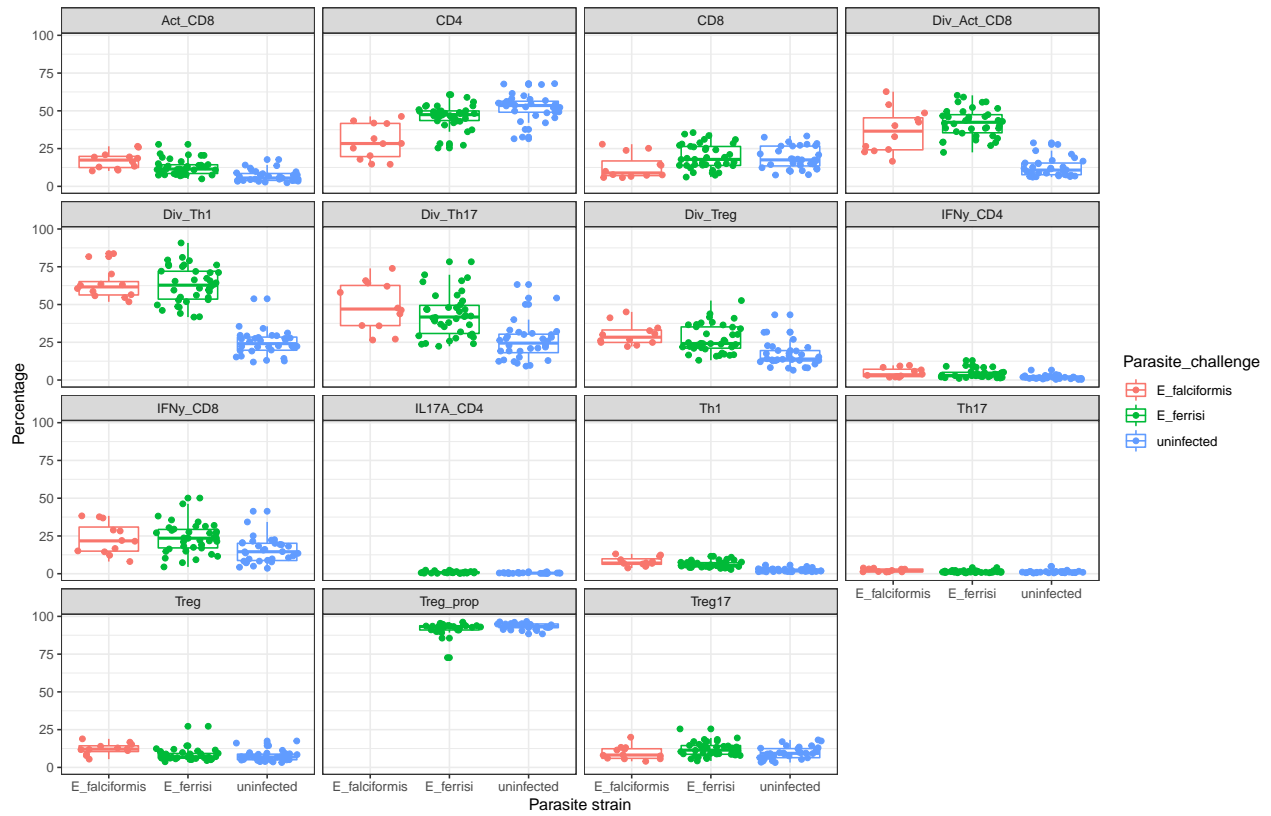
2. Correlations between the cells

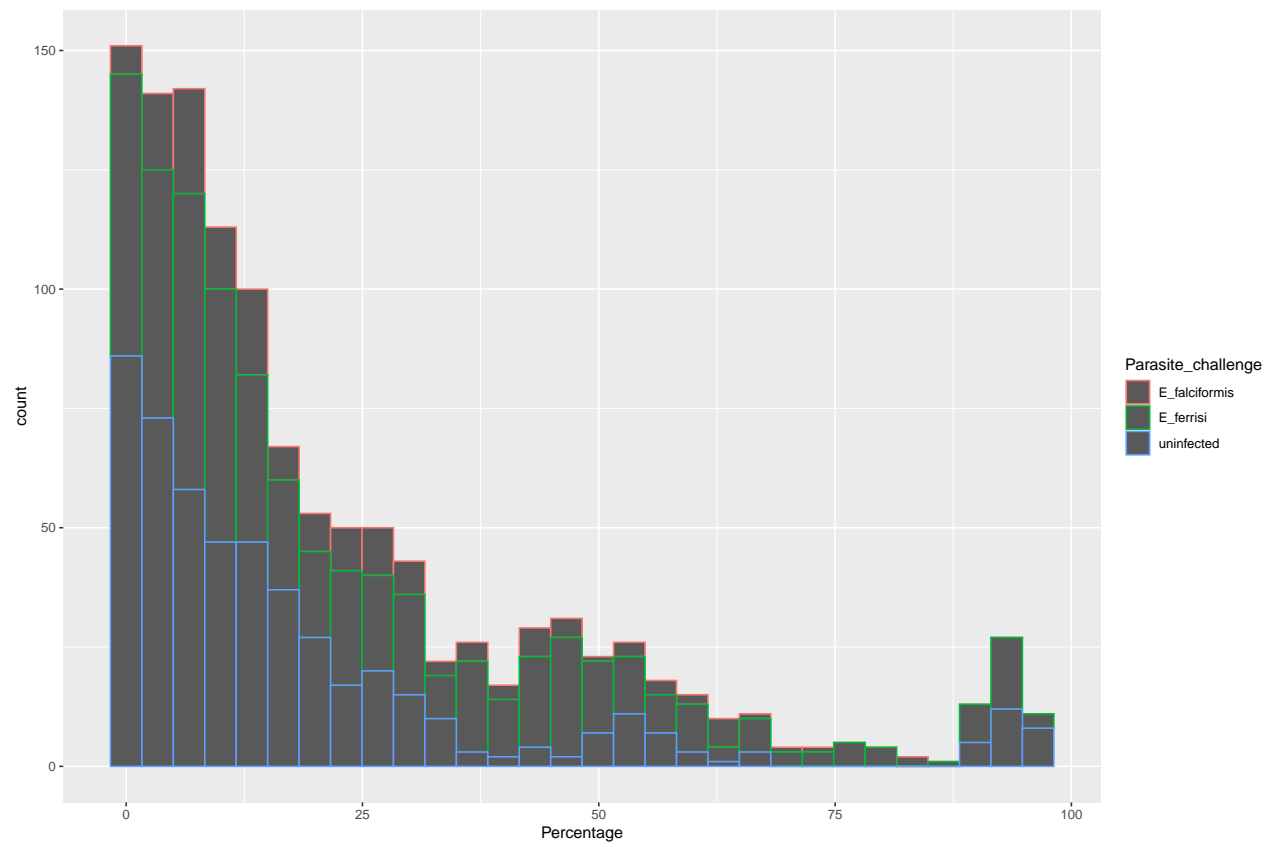


Cell counts in response to infection intensity

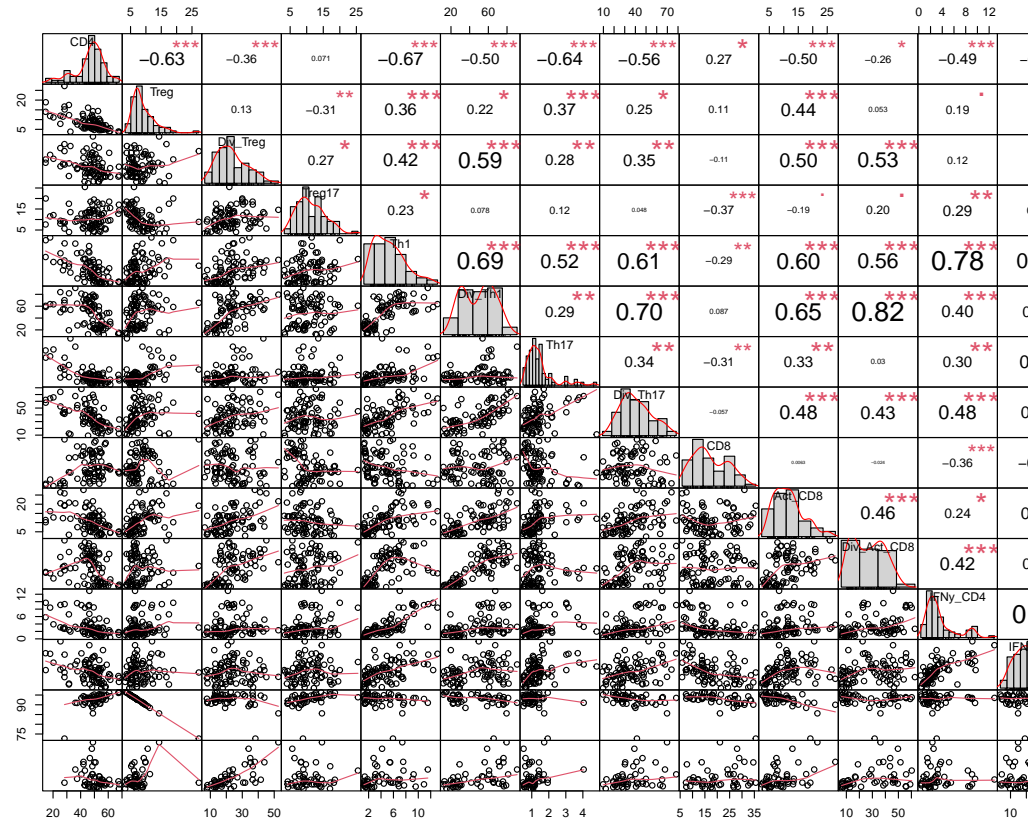


Cells in response to parasite strain





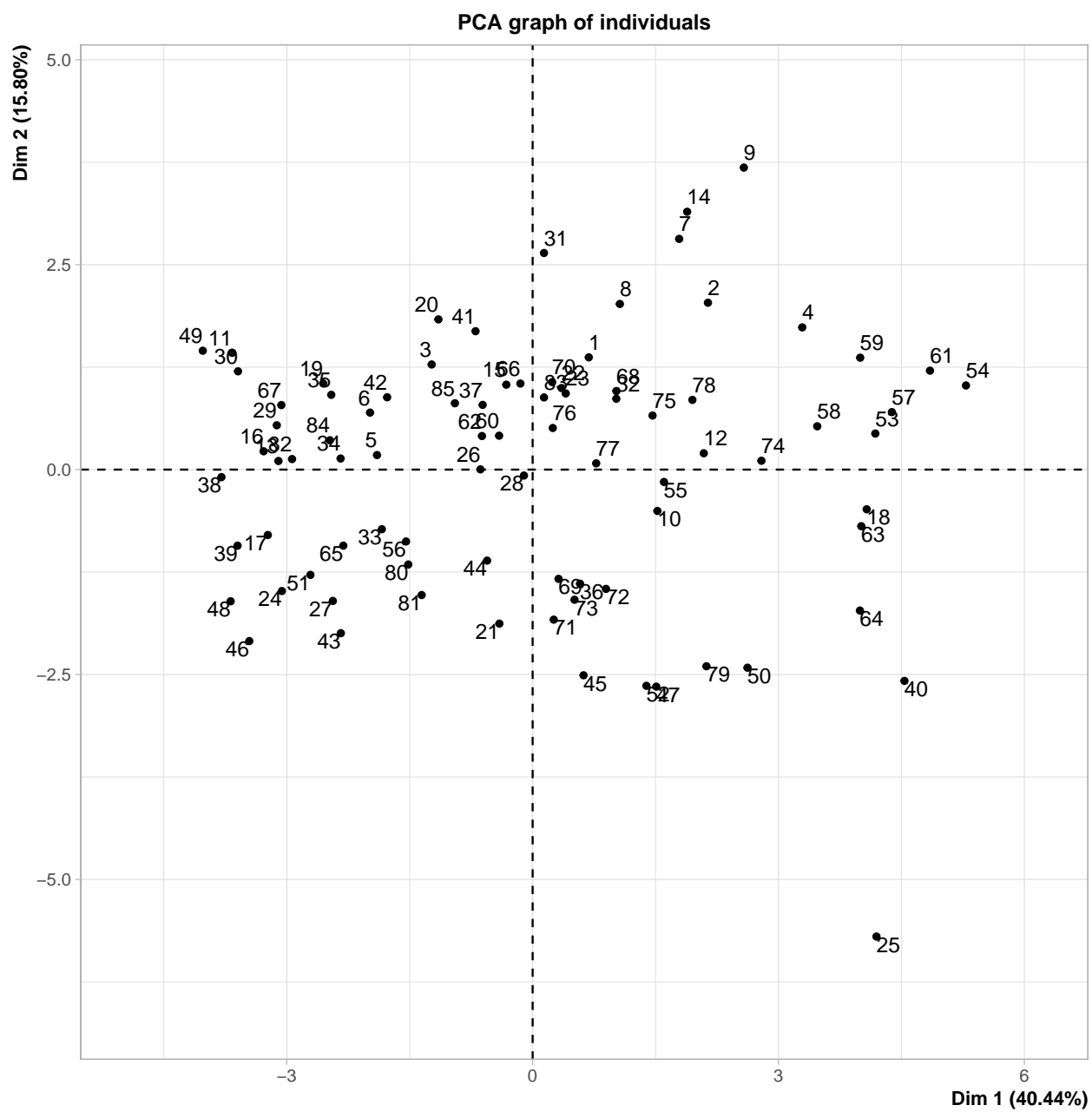
3. PCA

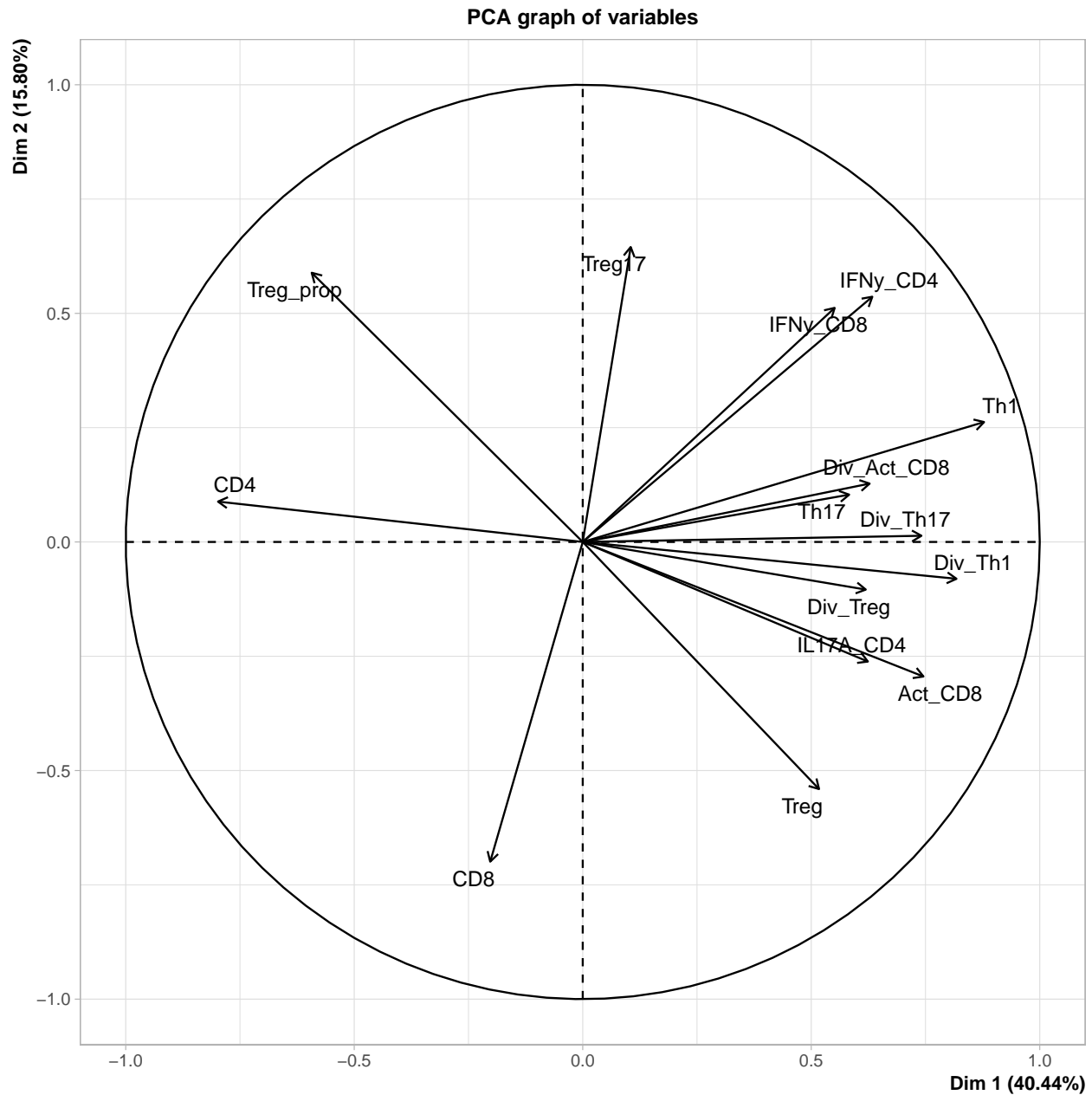


Handling missing data in a pca:

We will now continue by using an iterative pca to impute missing data A. Initialization: impute using the mean B. Step lampda: # a. do pca on imputed data table S dimensions retained # b. missing data imputed using pca # c. means (and standard deviations) updated C. Iterate the estimation and imputation steps (until convergence) (convergence: the act of converging and especially moving toward union or uniformity)

Overfitting is a common problem due to believing too much in links between variables. -> regularized iterative PCA (This version is what is being implmented in missMDA) This is a way of taking less risk when imputing the missing data. The algorithm estimates the missing data values with values that have no influence on the PCA results, i.e., no influence on the coordinates of the individuals or variables.





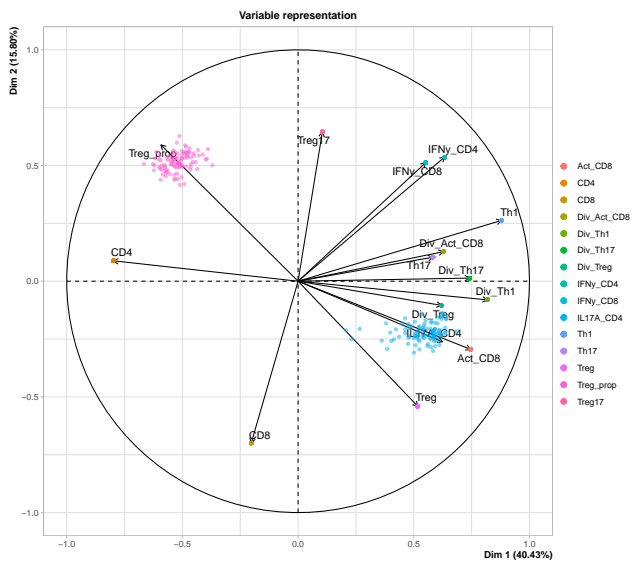
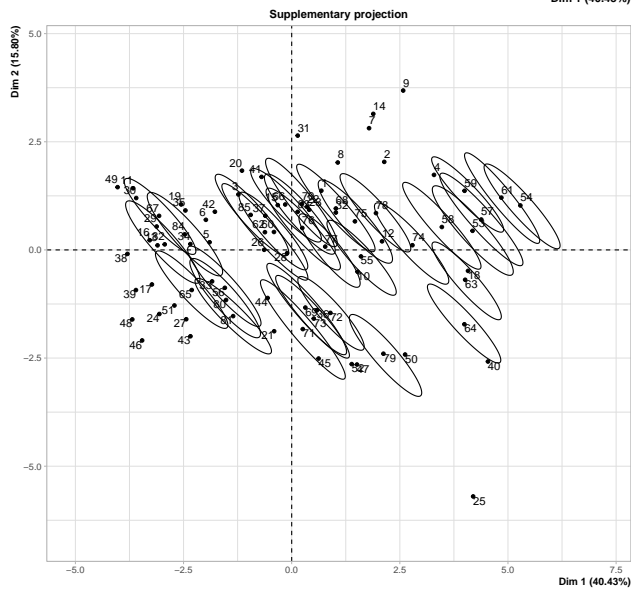
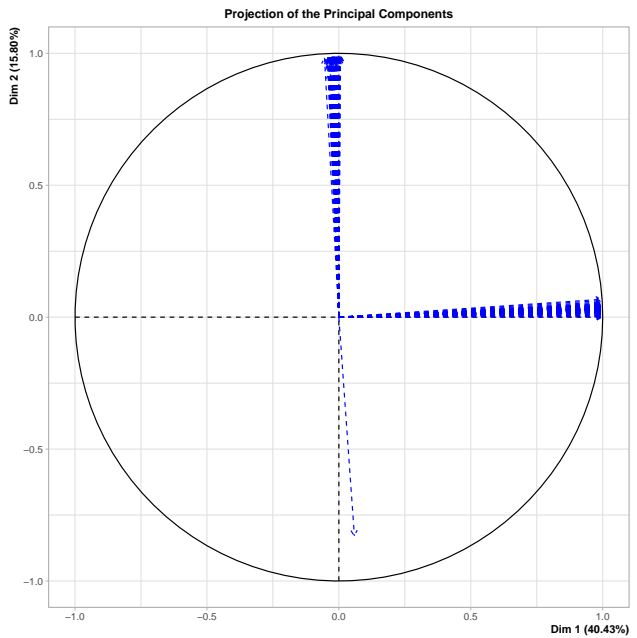
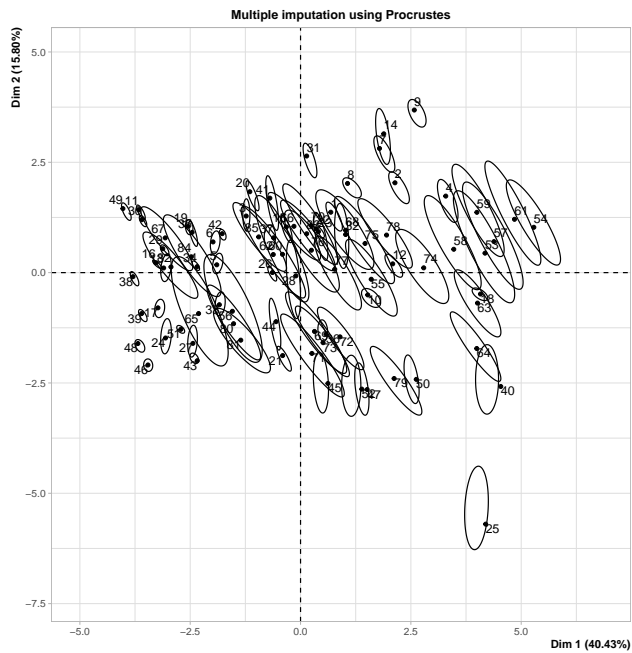
Caution: When imputing data, the percentages of inertia associated with the first dimensions will be overestimated.

Another problem: the imputed data are, when the pca is performed considered like real observations. But they are estimations!!

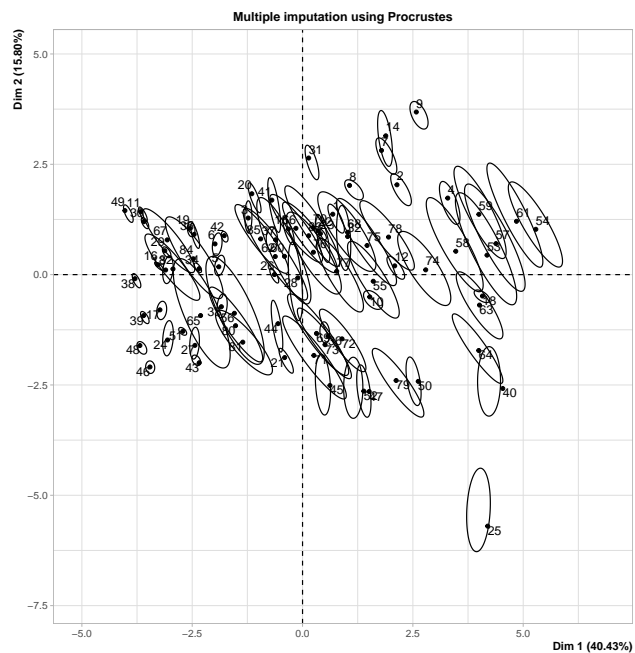
Visualizing uncertainty due to missing data:

→ multiple imputation: facstrate several plausible values for each missing data point

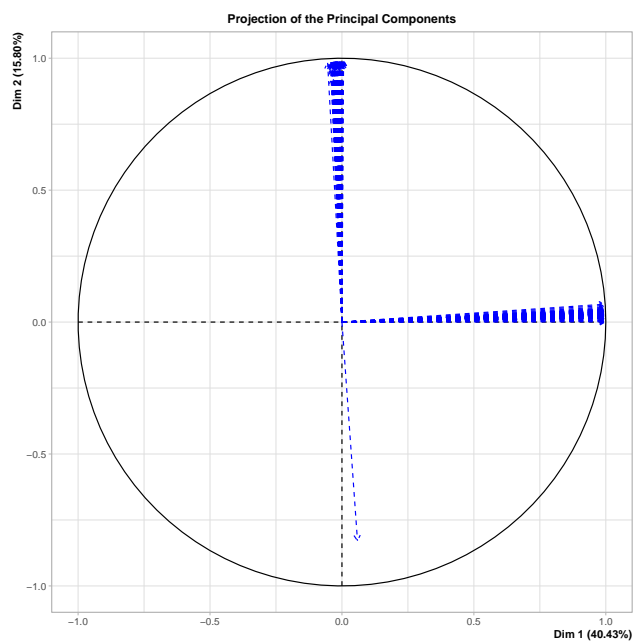
We here visualize the variability, that is uncertainty on the plane defined by two pca axes.



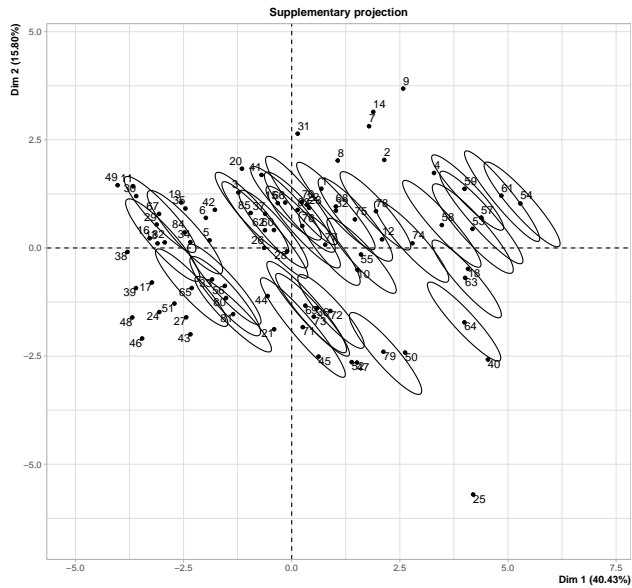
\$PlotIndProc



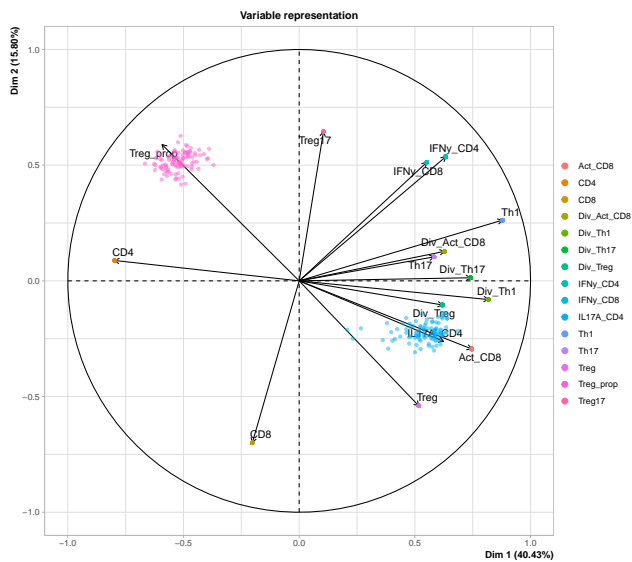
```
##
## $PlotDim
```



```
##
## $PlotIndSupp
```



```
##
## $PlotVar
```



Individuals lying on the axis have no missing data, but individuals that far away have many missing data.
big ellipse = big uncertainty tight ellipse (line) = low uncertainty

Variable representation: Points tight together)look like one) - have no missing variables -> low uncertainty
Points spread - > higher variability - > higher uncertainty

High uncertainty-> we should interpret the result with care

The individuals with many missing data values make the axes move, and thus the positions of all individuals

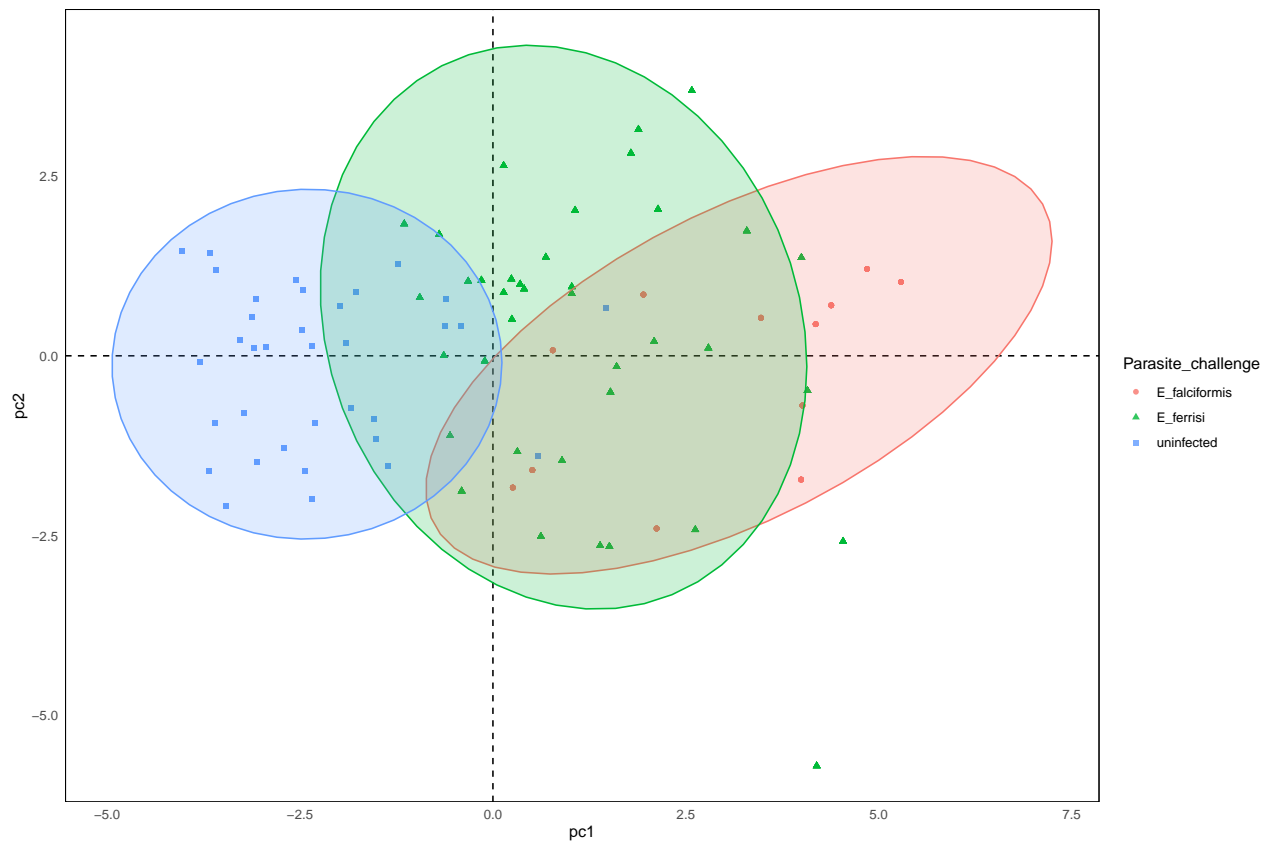
Therefore in the last plots every individual is getting an eclipse as they are as well influenced by the missing data of the others.

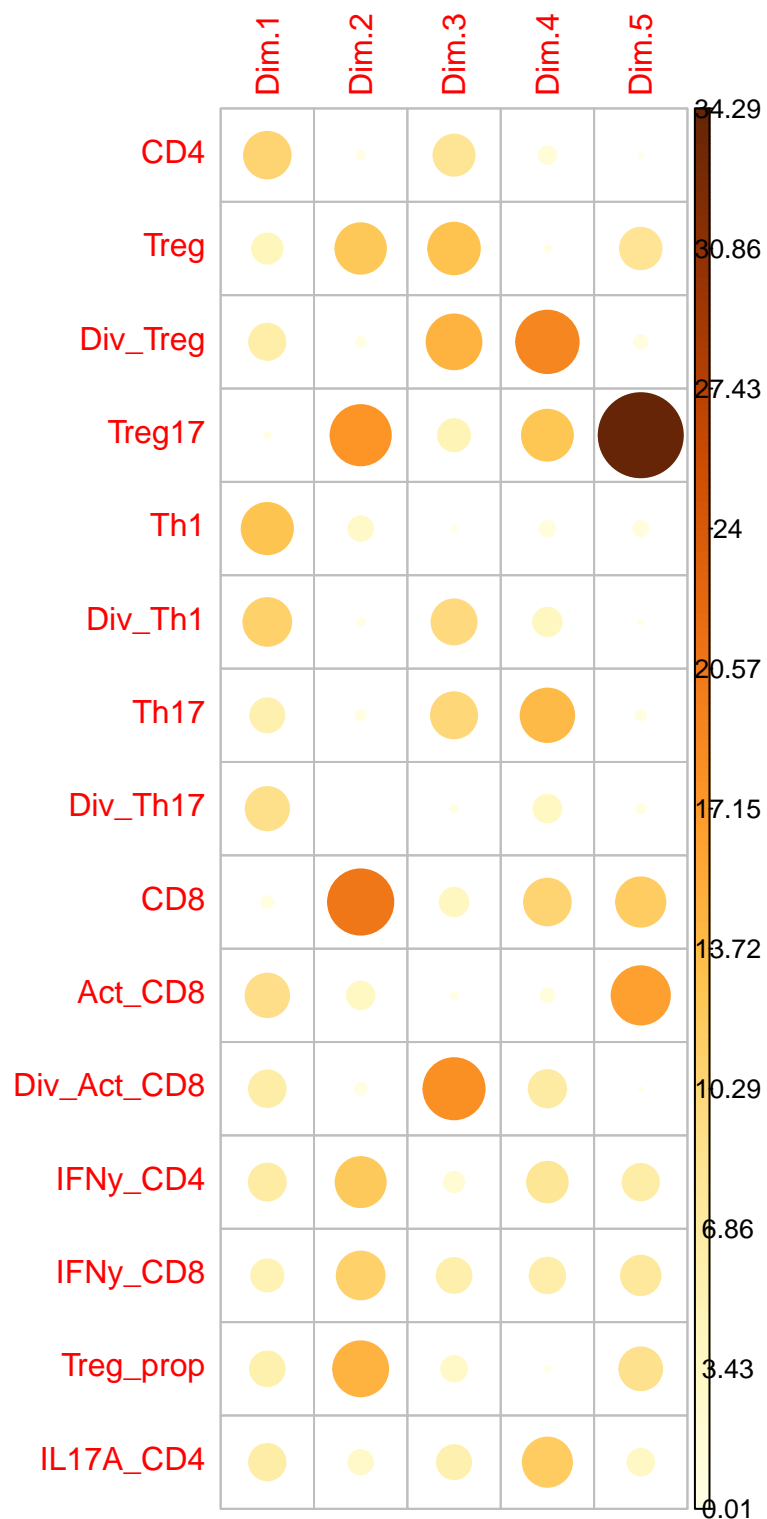
The plot with the dimensions shows the projections of the pca dimensions of each imputed table on the pca plane obtained using the original imputed data table

As all of the arrows are close to either the first or second axes, this means that the axes are stable with respect to the set of imputed tables -> we don't have evidence of instability here.

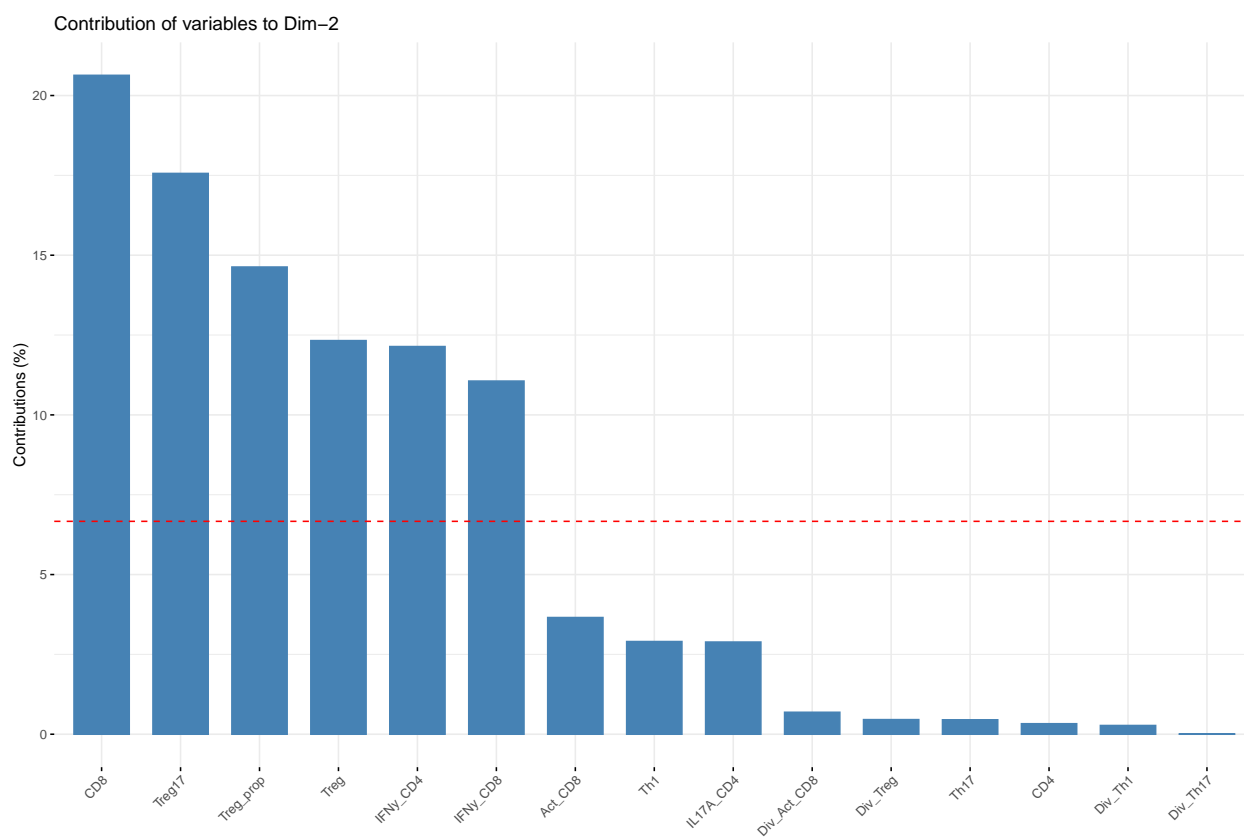
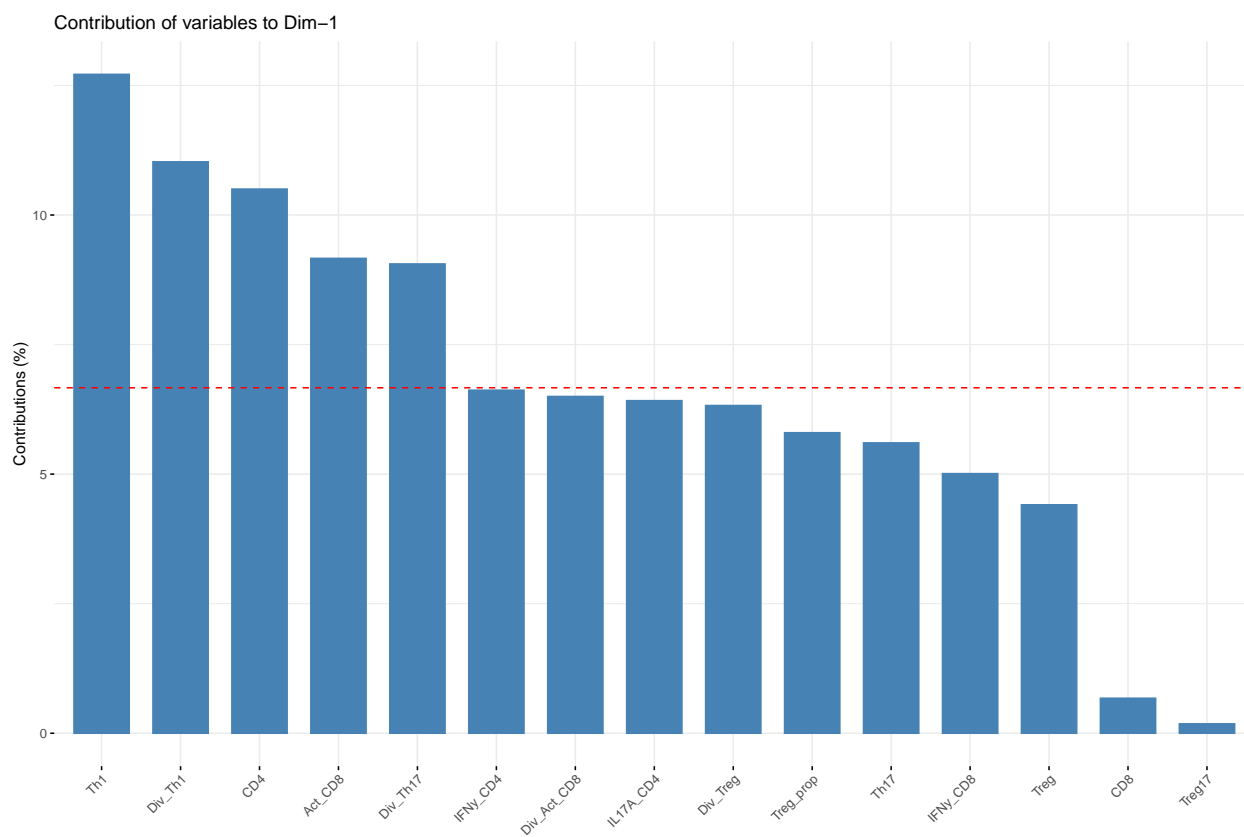
#Now we can make our initial plot of the PCA.

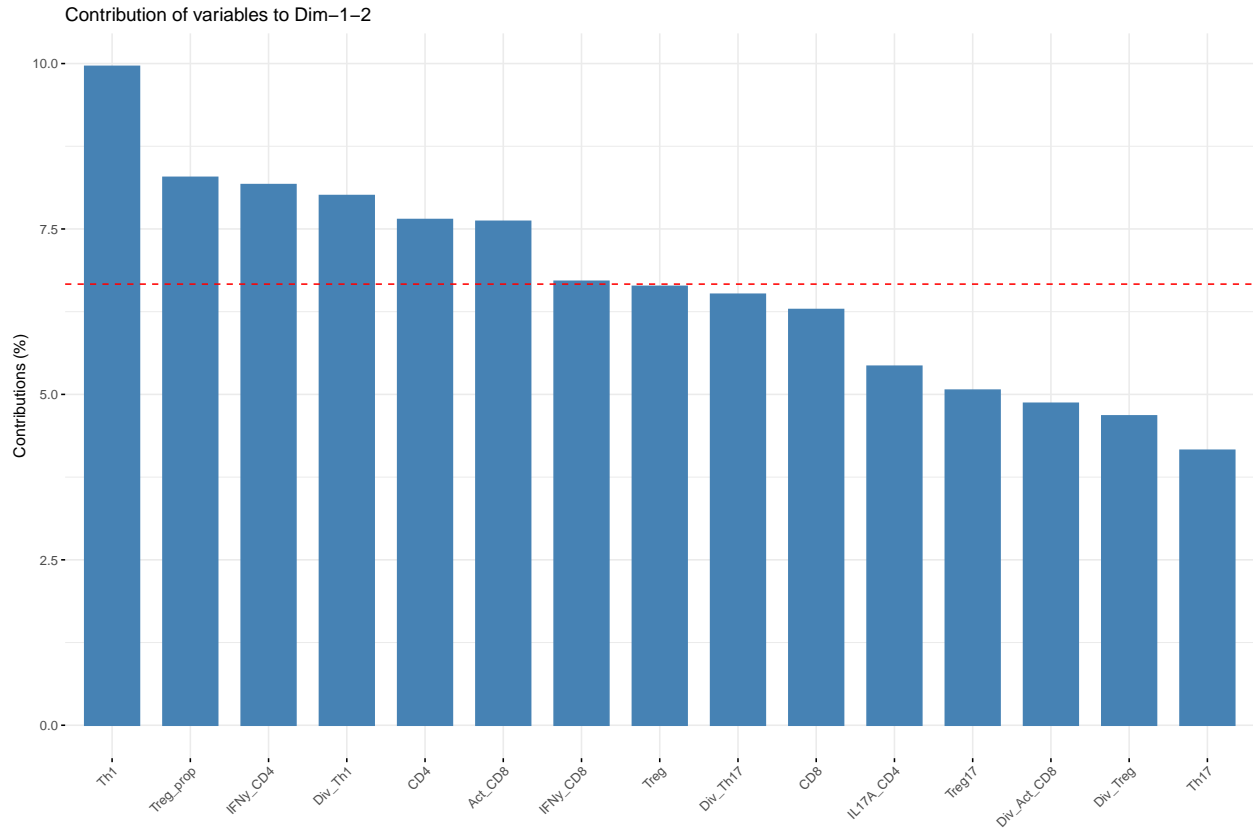
```
imputed_facs %>%
  pivot_longer(10:24, names_to = "Cells", values_to = "Proportion") %>%
  ggplot(aes(x = pc1, y = pc2, color = Parasite_challenge, shape = Parasite_challenge)) +
  geom_hline(yintercept = 0, lty = 2) +
  geom_vline(xintercept = 0, lty = 2) +
  geom_point(alpha = 0.8) +
  stat_ellipse(geom="polygon", aes(fill = Parasite_challenge), alpha = 0.2, show.legend = FALSE,
              level = 0.95) +
  theme_minimal() +
  theme(panel.grid = element_blank(), panel.border = element_rect(fill= "transparent"))
```





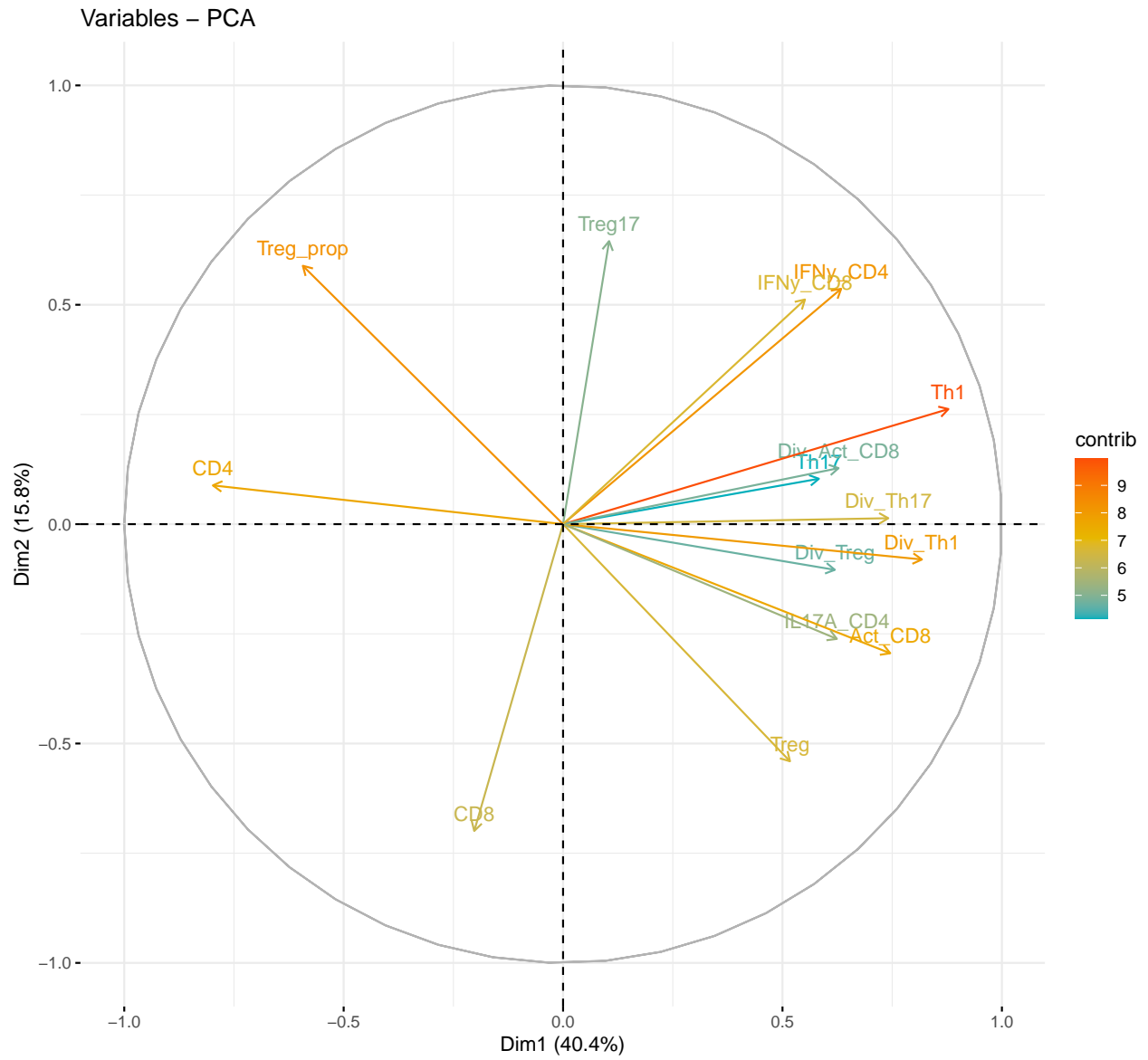
The function `fviz_contrib()` [factoextra package] can be used to draw a bar plot of variable contributions. If your data contains many variables, you can decide to show only the top contributing variables. The R code below shows the top 10 variables contributing to the principal components:



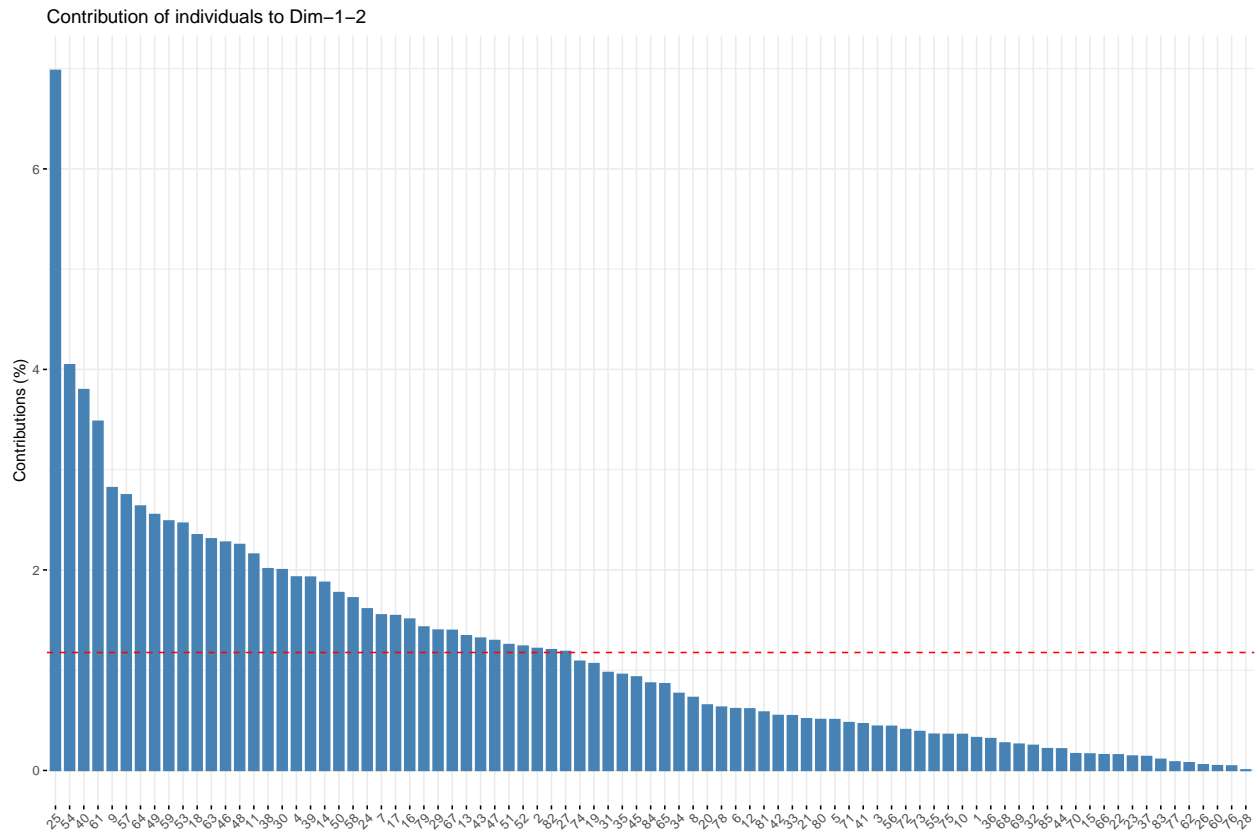


The red dashed line on the graph above indicates the expected average contribution. If the contribution of the variables were uniform, the expected value would be $1/\text{length}(\text{variables}) = 1/10 = 10\%$. For a given component, a variable with a contribution larger than this cutoff could be considered as important in contributing to the component.

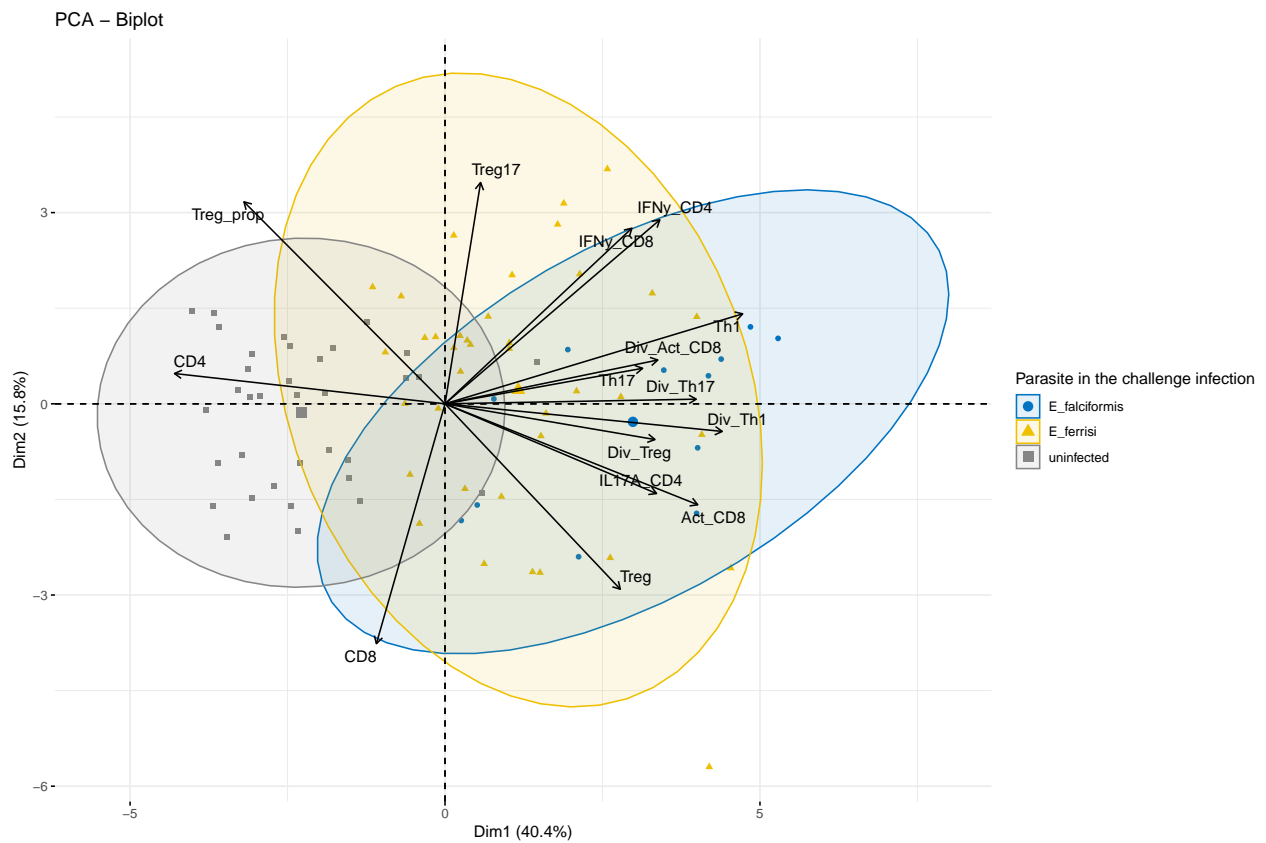
Note that, the total contribution of a given variable, on explaining the variations retained by two principal components, say PC1 and PC2, is calculated as $\text{contrib} = [(C1 * \text{Eig1}) + (C2 * \text{Eig2})]/(\text{Eig1} + \text{Eig2})$, where C1 and C2 are the contributions of the variable on PC1 and PC2, respectively Eig1 and Eig2 are the eigenvalues of PC1 and PC2, respectively. Recall that eigenvalues measure the amount of variation retained by each PC. In this case, the expected average contribution (cutoff) is calculated as follow: As mentioned above, if the contributions of the 10 variables were uniform, the expected average contribution on a given PC would be $1/10 = 10\%$. The expected average contribution of a variable for PC1 and PC2 is : $[(10 * \text{Eig1}) + (10 * \text{Eig2})]/(\text{Eig1} + \text{Eig2})$



To visualize the contribution of individuals to the first two principal components:



PCA + Biplot combination



In the following example, we want to color both individuals and variables by groups. The trick is to use `pointshape = 21` for individual points. This particular point shape can be filled by a color using the argument `fill.ind`. The border line color of individual points is set to “black” using `col.ind`. To color variable by groups, the argument `col.var` will be used.

Linear models:

```
##
## Call:
## lm(formula = max_WL ~ pc1 + pc2 + Parasite_challenge, data = imputed_facs)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -16.5521  -2.9263   0.1456   3.6345   9.9540
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)      80.9739      1.8237  44.401 < 2e-16 ***
## pc1              0.9508      0.3772   2.521  0.0137 *
## pc2             -0.1475      0.3542  -0.416  0.6782
## Parasite_challengeE_ferrisi 11.0501      1.7898   6.174 2.60e-08 ***
## Parasite_challengeuninfected 17.1202      2.5903   6.609 3.95e-09 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 4.975 on 80 degrees of freedom
## Multiple R-squared:  0.4265, Adjusted R-squared:  0.3978
## F-statistic: 14.87 on 4 and 80 DF,  p-value: 3.976e-09
## [1] 520.8357
##
## Call:
## lm(formula = max_WL ~ pc1 + pc2 + Parasite_challenge + hybrid_status,
##     data = imputed_facs)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -14.9842  -3.1816   0.5616   3.5633   9.4894
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)      79.8812      2.1017  38.007 < 2e-16 ***
## pc1              0.9103      0.4255   2.140  0.0356 *
## pc2              0.3741      0.5915   0.633  0.5290
## Parasite_challengeE_ferrisi 10.6313      1.9281   5.514 4.76e-07 ***
## Parasite_challengeuninfected 16.6312      2.5658   6.482 8.55e-09 ***
## hybrid_statusF0 M. m. musculus 1.5746      2.5368   0.621  0.5367
## hybrid_statusF1 hybrid      3.5399      1.6592   2.134  0.0362 *
## hybrid_statusF1 M. m. domesticus -0.9093      2.0039  -0.454  0.6513
## hybrid_statusF1 M. m. musculus  4.2002      2.7063   1.552  0.1249
## hybrid_statusother      1.6583      2.1713   0.764  0.4474
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 4.91 on 75 degrees of freedom
```

```

## Multiple R-squared:  0.4763, Adjusted R-squared:  0.4134
## F-statistic: 7.578 on 9 and 75 DF,  p-value: 8.217e-08

## [1] 523.1115

##
## Call:
## lm(formula = max_WL ~ pc1 + pc2 + hybrid_status, data = imputed_facs)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -17.0089  -3.8001   0.7976   4.3336  11.4052
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)      91.3120      1.4723  62.020  <2e-16 ***
## pc1             -0.6637      0.3125  -2.124  0.0369 *
## pc2              0.4622      0.7284   0.635  0.5276
## hybrid_statusF0 M. m. musculus  1.6599      3.1596   0.525  0.6009
## hybrid_statusF1 hybrid          4.5068      2.0539   2.194  0.0312 *
## hybrid_statusF1 M. m. domesticus -0.5451      2.4849  -0.219  0.8270
## hybrid_statusF1 M. m. musculus   5.0932      3.3323   1.528  0.1305
## hybrid_statusother              0.7101      2.4448   0.290  0.7723
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 6.118 on 77 degrees of freedom
## Multiple R-squared:  0.1654, Adjusted R-squared:  0.08956
## F-statistic:  2.18 on 7 and 77 DF,  p-value: 0.04505

## [1] 558.7177

##
## Call:
## lm(formula = max_WL ~ pc1 + pc2 + infection_history, data = imputed_facs)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -12.5834  -2.8158  -0.4039   3.2253   8.7078
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)      84.39648      2.65405  31.799  < 2e-16
## pc1              0.98773      0.36148   2.732  0.007858
## pc2             -0.09311      0.38599  -0.241  0.810049
## infection_historyfalciformis_ferrisi  7.62628      2.63621   2.893  0.005011
## infection_historyfalciformis_uninfected 15.00027      3.40217   4.409  3.46e-05
## infection_historyferrisi_falciformis  -4.18836      3.02105  -1.386  0.169791
## infection_historyferrisi_ferrisi       9.29476      2.78012   3.343  0.001301
## infection_historyferrisi_uninfected    13.15007      3.36444   3.909  0.000204
## infection_historyuninfected            13.65829      3.90798   3.495  0.000805
## infection_historyuninfected_falciformis -8.53966      3.95493  -2.159  0.034073
## infection_historyuninfected_ferrisi    -1.22495      3.30747  -0.370  0.712174
##
## (Intercept)      ***
## pc1              **

```

```
## pc2
## infection_historyfalciiformis_ferrisi **
## infection_historyfalciiformis_uninfected ***
## infection_historyferrisi_falciiformis
## infection_historyferrisi_ferrisi **
## infection_historyferrisi_uninfected ***
## infection_historyuninfected ***
## infection_historyuninfected_falciiformis *
## infection_historyuninfected_ferrisi
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 4.48 on 74 degrees of freedom
## Multiple R-squared:  0.5699, Adjusted R-squared:  0.5117
## F-statistic: 9.804 on 10 and 74 DF,  p-value: 3.216e-10
## [1] 508.3793
##
## Call:
## lm(formula = max_WL ~ pc1 + pc2, data = imputed_facs)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -18.168  -3.267   1.382   4.218   8.878
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  92.9634     0.6727 138.191 < 2e-16 ***
## pc1         -0.7582     0.2732  -2.776  0.00682 **
## pc2         -0.1078     0.4370  -0.247  0.80584
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 6.202 on 82 degrees of freedom
## Multiple R-squared:  0.08651, Adjusted R-squared:  0.06423
## F-statistic: 3.883 on 2 and 82 DF,  p-value: 0.02448
##
##              df      AIC
## weight_lm      6 520.8357
## weight_lm_exp_only 4 556.3975
```

repeating the heatmap on the now imputed data

```
facs <- imputed_facs %>%
  dplyr::select(c(EH_ID, all_of(CellCount.cols))) %>%
  dplyr::select(-Position)

# turn the data frame into a matrix and transpose it. We want to have each cell
# type as a row name
facs <- t(as.matrix(facs))

#switch the matrix back to a data frame format
facs <- as.data.frame(facs)
```

```

# turn the first row into column names
facs %>%
  row_to_names(row_number = 1) -> heatmap_data

table(rowSums(is.na(heatmap_data)) == nrow(heatmap_data))

##
## FALSE
## 15

# turn the columns to numeric other wise the heatmap function will not work
heatmap_data[] <- lapply(heatmap_data, function(x) as.numeric(as.character(x)))
# remove columns with only NAs
heatmap_data <- Filter(function(x)!all(is.na(x)), heatmap_data)

#remove rows with only NAs
heatmap_data <- heatmap_data[, colSums(is.na(heatmap_data)) != nrow(heatmap_data)]
rownames(annotation_df) <- colnames(heatmap_data)

```

Heatmap on facs expression data:

