

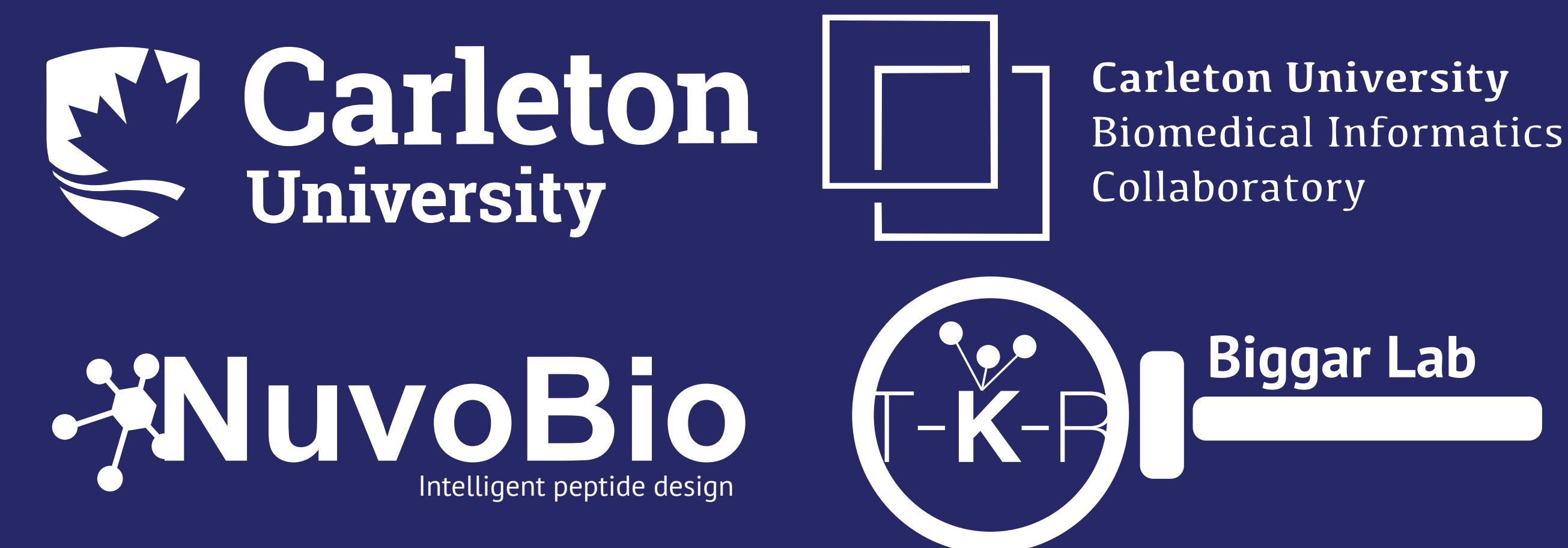
In silico design of a novel SMYD3 inhibitor with Darwin

François Charih^{1,2,3}, Skye Wang², Kyle K. Biggar^{2,3}, James R. Green^{1,3}

¹Department of Systems and Computer Engineering, Carleton University, Ottawa, ON, Canada

²Institute of Biochemistry, Carleton University, Ottawa, ON, Canada

³NuvoBio, Ottawa, ON, Canada



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Background

- Lysine methylation is an important post-translational modification of proteins carried out by lysine methyltransferases (KMTs).
- The methylation status of a protein has consequences on its subcellular localization, stability, interactions with other proteins, among others.
- Overexpression of the KMT **SMYD3** is observed in several cancers, including pancreatic, lung, gastric, breast, ovarian, and esophageal cancer [1]. As such, SMYD3 constitutes a prime target for inhibition.
- Few peptide drugs have been approved for use in the clinic, but they are becoming increasingly promising, as they can be designed to disrupt protein-protein interactions or inhibit enzymes with high specificity.
- Designing peptide inhibitors is challenging due to the vastness of the synthesizable peptide space. Moreover, only an infinitesimal fraction of that space corresponds to chemically relevant peptides, *i.e.* that can bind and/or inhibit a specific protein target.
- The *In Silico Peptide Synthesizer* (InSiPS) [2] is a genetic algorithm-based program that optimizes a peptide *binder* sequence, using the PIPE interaction score of the sequence with the target as a measure of fitness.
- InSiPS was shown to have the ability to generate peptide binders for yeast and viral targets [2,3], but the computational requirements (*i.e.* thousands of CPUs) make InSiPS difficult to deploy at larger scales and in more complex organisms such as humans.

Objectives

- Implement a more *computationally efficient* sequence-based peptide binder design algorithm, using InSiPS as a starting point.
- Design an effective peptide inhibitor against SMYD3 to demonstrate Darwin's ability to design a bioactive inhibitor for a human protein target *in silico*, using sequence information only.

Darwin for *in silico* peptide design

- Similar to InSiPS, our implementation, named **Darwin**, uses a genetic algorithm-based strategy to optimize the binder sequence.
- We designed a new, proprietary fitness function that favours high SPRINT interaction score [4] for the peptide-target pair and penalizes high off-target interaction scores.

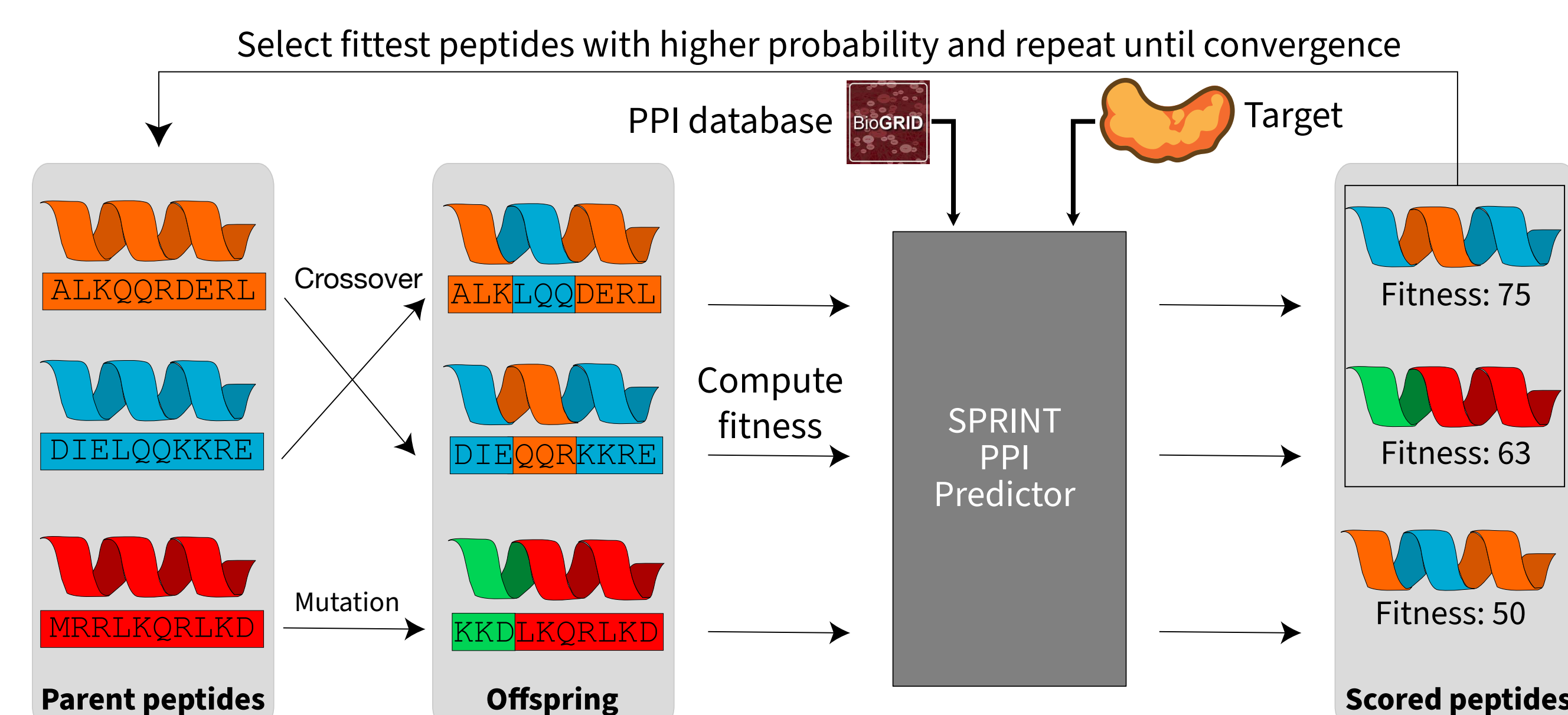


Figure 1. Overview of the Darwin algorithm.

The Darwin algorithm is strongly inspired by InSiPS, and uses a genetic algorithm optimization strategy to produce peptide binders using a protein-protein interaction score as a measure of peptide fitness.

Design of anti-SMYD3 inhibitors with Darwin

We ran Darwin to design short peptides (20-mers) against the SMYD3 methyltransferase with a population size of 2000. We considered 20 iterations without improvement in maximum fitness to indicate convergence.

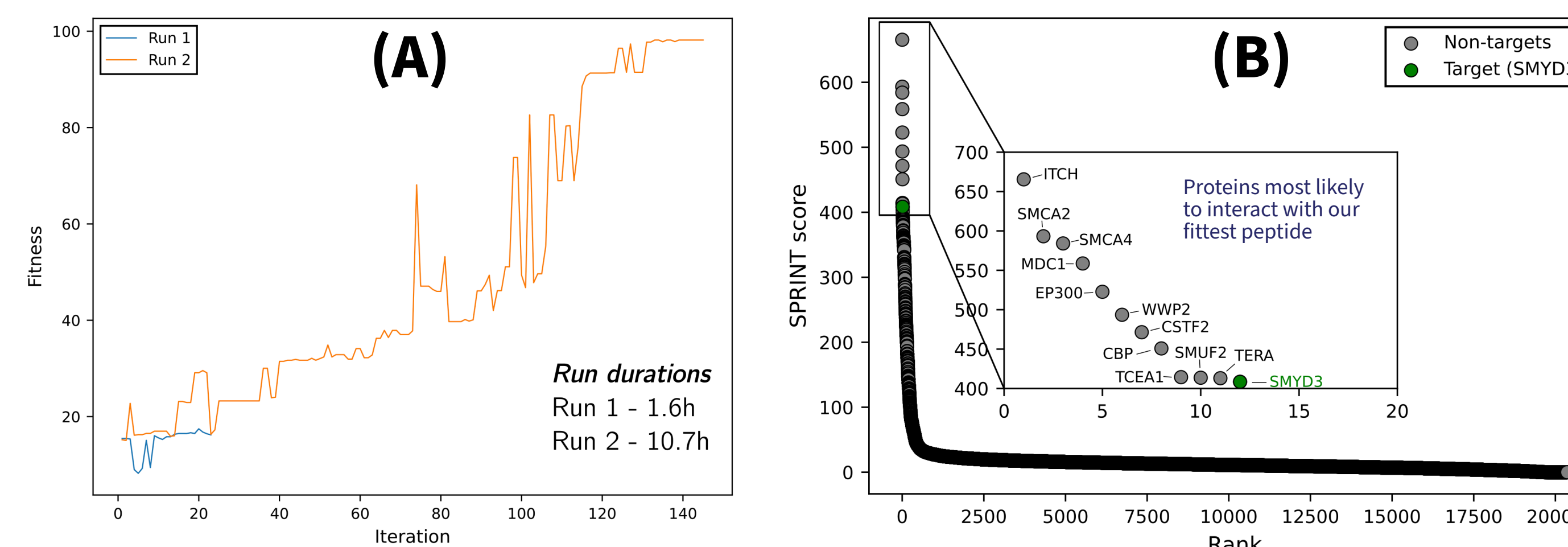


Figure 2. *In silico* design of anti-SMYD3 peptide inhibitors.

(A) Evolution curve for 2 Darwin runs against SMYD3 on a 11-node computer cluster with 440 Intel Xeon 2.40GHz CPUs. (B) One-to-all curve showing the SPRINT interaction scores between the fittest peptide and all proteins exposed to it.

Assessment of our anti-SMYD3 peptides binding profiles

We investigated whether the peptides generated with Darwin actually interact with SMYD3 *in vitro*. To this end, we printed peptides on SPOT arrays, and incubated the peptide with the target protein and a non-target protein to assess interaction specificity.

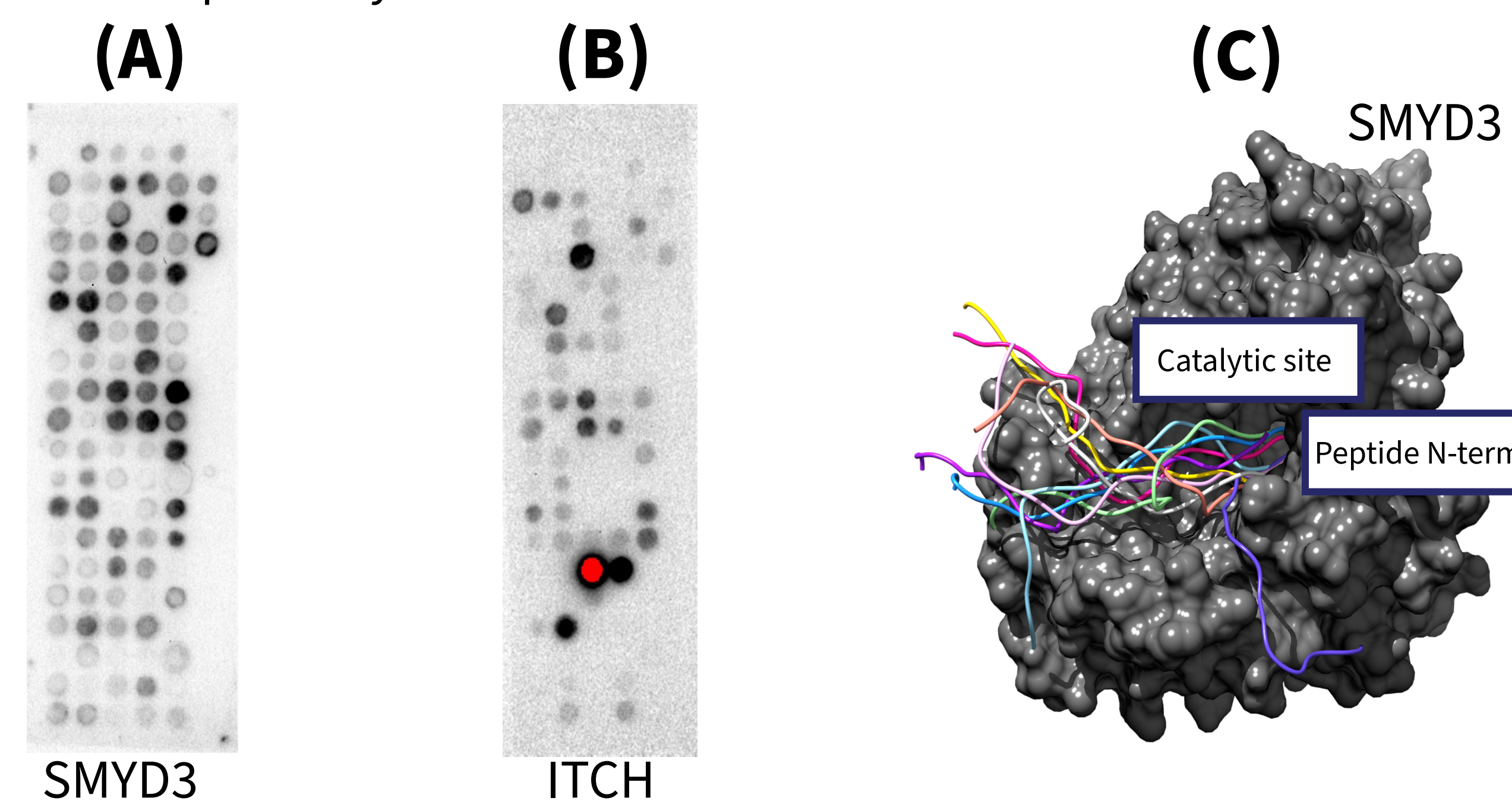


Figure 3. Assessment of the binding activity of Darwin-generated peptides.

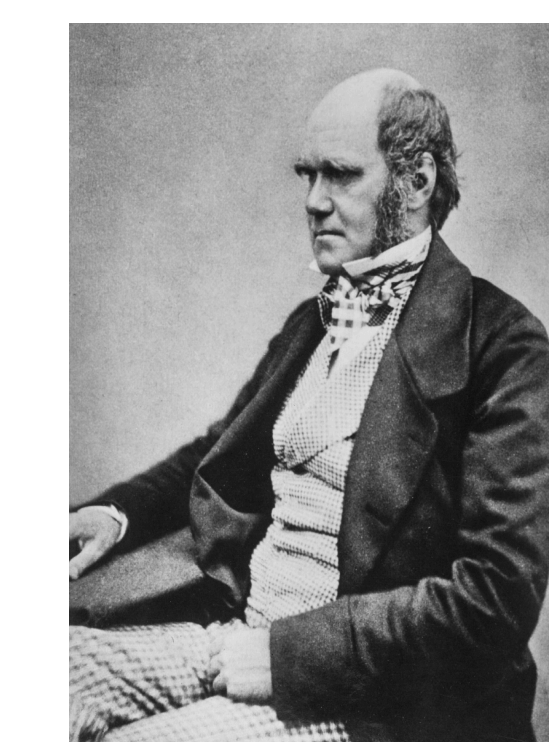
(A) We printed 96 peptides on SPOT arrays, and incubated the peptides with our target SMYD3 to identify actual binders. (B) We repeated the experiment with ITCH (ubiquitin protein ligase), a high-ranking off-target protein to assess off-target interactions. (C) Using HPEPDOCK [5], we docked our lead peptide in SMYD3 (PDB: 7O2C; ligand removed). Shown are the the 10 most thermodynamically-favoured poses predicted.

Darwin: behind the name

You may wonder why we named our algorithm *Darwin*...

Genetic algorithms, the optimization strategy that underpins our work relies heavily on a core concept described by Charles Darwin's theory of evolution: *survival of the fittest*.

Our algorithm iteratively *evolves* peptides. The "fittest" (most likely to specifically bind the target protein) peptides in a given generation are more likely to be selected for the next generation, and to subsequently give rise to fitter peptides when subjected to mutation and/or crossover operations.



Charles Darwin
(1809-1882)

Evaluation of inhibition activity

Given that Darwin only aims to produce peptides that bind to a selected target, we sought to determine whether the observed binding is associated with a reduction in the enzymatic activity of the target.

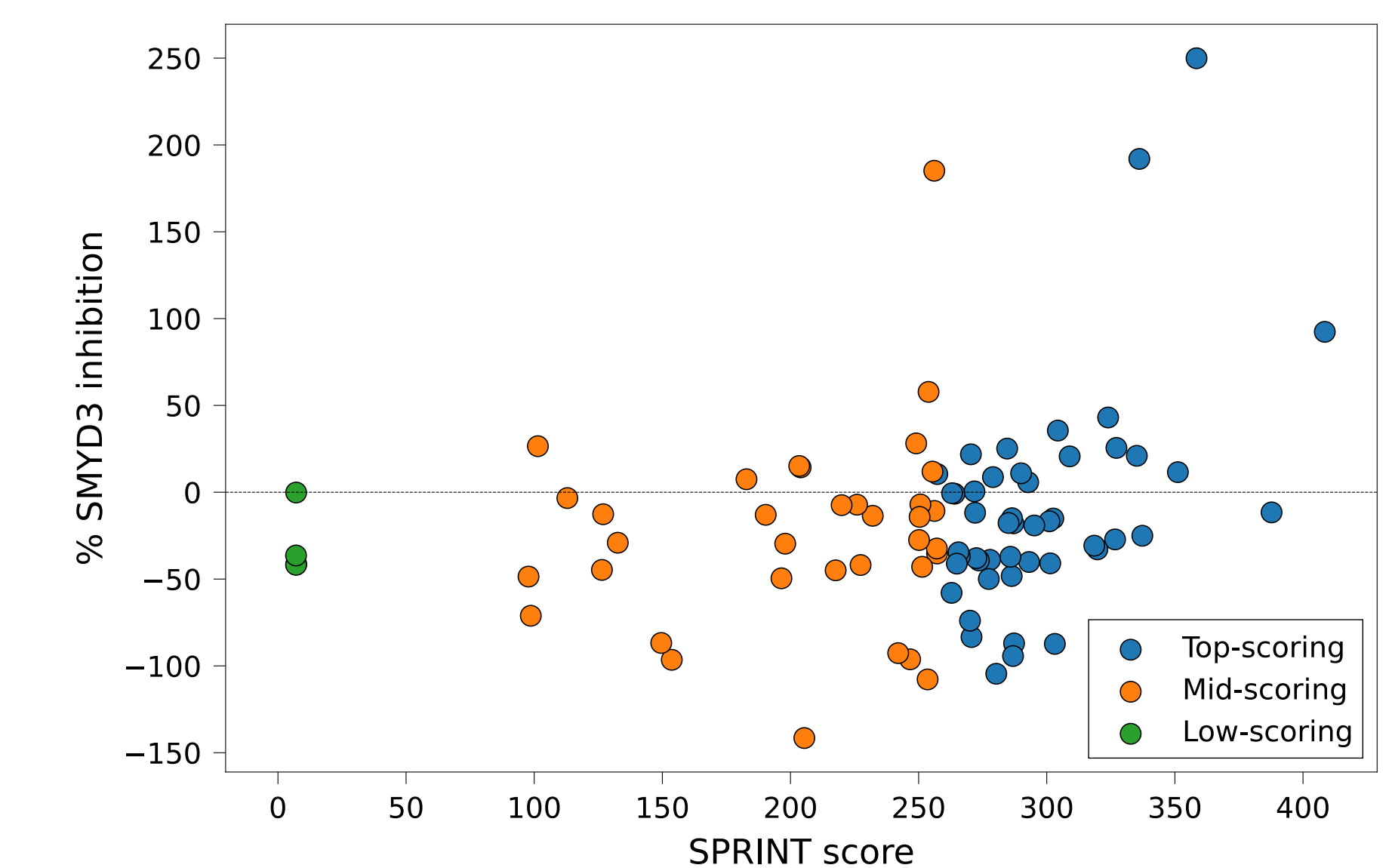


Figure 4. *In vitro* assessment of enzyme inhibition for 96 peptides with a luminescence-based methyltransferase assay.

Using a MAP3K2 fragment known to be a SMYD3 substrate, we measured the reduction in methylation signal upon incubation of SMYD3 with the substrate and our 96 Darwin-generated peptide binders.

References

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- [4] Li, Y. & Ilie, L. SPRINT: ultrafast protein-protein interaction prediction of the entire human interactome. *BMC Bioinformatics* 18, 485 (2017).
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Acknowledgements

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Highlights

- We have shown that Darwin can generate peptide binders for human enzyme targets *in silico*, using sequence only, and with little computational resources.
- Some of the peptide binders generated with Darwin for the SMYD3 enzyme are bioactive, *i.e.* achieve enzyme inhibition.
- For inquiries or collaboration opportunities, please visit <https://nuvobio.com>.