



Linear Regression Models

STATS 762 – Lecture Slides 6

March 25, 2019

Interactions

Interactions may be useful to model in situations where the impact of one factor may depend on the level of another factor.

- ▶ Interactions can involve just factors, just numeric regressors or both factors and numeric regressors.
- ▶ Useful when the impact on the response of one regressor depends on the level of another regressor.

ANOVA

Classically, ANOVA was used to analyse data from a designed experiment where:

- ▶ All of the explanatory variables were factors,
- ▶ The set of covariate patterns consisted of all possible combinations of levels for these factors (this was called a factorial design),
- ▶ The same number of observations were taken for each covariate pattern.

This structure simplified both the mathematical calculations and the interpretation of results. It also allowed interactions between the factors to be estimated.

Example: Weight Gain of Rats

An experiment investigated the effect of diet on the weight gained by rats.

- ▶ Two treatment factors:
 1. source of protein: beef, pork, or cereal
 2. level: high or low
- ▶ Ten rats were assigned to each combination of “source” and “level”
 - ▶ this is called a balanced design – each combination of factors was used for the same number of observations

The Data Frame

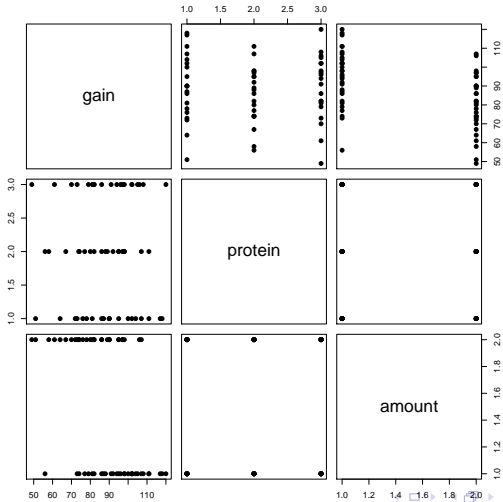
```
> rats.df
```

	gain	protein	amount
1	73	Beef	High
2	98	Cereal	High
3	94	Pork	High
4	90	Beef	Low
5	107	Cereal	Low
6	49	Pork	Low

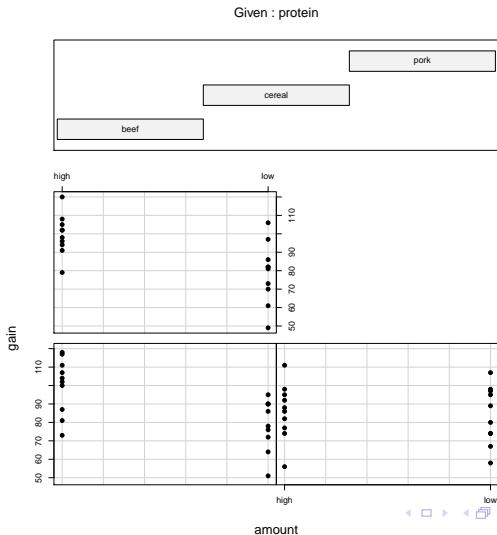
=====

56	92	Cereal	High
57	105	Pork	High
58	78	Beef	Low
59	58	Cereal	Low
60	82	Pork	Low

Pairs Plot



Conditional Plot



Two Factors

Both of the regressors (protein and amount) are factors. In *R* the factor function is used to designate factors:

```
> attach(rats.df)
> proteinA<-factor(protein)
> attributes(proteinA)
$levels
[1] "beef"    "cereal"  "pork"
$class
[1] "factor"

> amountA<-factor(amount)
> attributes(amountA)
$levels
[1] "high" "low"
$class
[1] "factor"
```


Indicator Variables

A set of indicator variables is needed for each factor:

- ▶ The indicator variables allow the mean response to be adjusted for each different category of the factor individually.
- ▶ To accomplish this the number of indicator variables needs to be one fewer than the number of levels for the factor.
- ▶ By default *R* creates indicator variables for a “baseline” model – each factor has a baseline level and all of the indicator variables represent comparisons of the other levels to the baseline level.

Indicator Variables (cont.)

In *R* we can view the indicator variables that have been set up for a factor using the `contrasts` command:

```
> contrasts(proteinA)
```

	cereal	pork
beef	0	0
cereal	1	0
pork	0	1

```
> contrasts(amountA)
```

	low
high	0
low	1

- Each column defines an indicator variable (a contrast).

The Model Defined by these Indicator Variables

Our model matrix will have four columns consisting of the intercept the two indicator variables for proteinA and the indicator for amountA. Let the coefficients for these be β_0 , β_C , β_P and β_L respectively. Then the model defines the means for the six different combinations of protein source and amount as:

	<u>Protein Source</u>		
amount	beef	cereal	pork
high	β_0	$\beta_0 + \beta_C$	$\beta_0 + \beta_P$
low	$\beta_0 + \beta_L$	$\beta_0 + \beta_C + \beta_L$	$\beta_0 + \beta_P + \beta_L$

Output for Fitted Model

```
> ratsA.lm=lm(gain~proteinA+amountA)
> summary(ratsA.lm)
```

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)	
(Intercept)	96.867	3.898	24.850	< 2e-16	***
proteinAcereal	-4.700	4.774	-0.984	0.329126	
proteinApork	-0.500	4.774	-0.105	0.916965	
amountAlow	-14.533	3.898	-3.728	0.000451	***

Residual standard error: 15.1 on 56 degrees of freedom
Multiple R-squared: 0.212, Adjusted R-squared: 0.1698
F-statistic: 5.023 on 3 and 56 DF, p-value: 0.003739

ANOVA for Fitted Model

```
> anova(ratsA.lm)
```

Response: gain

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
proteinA	2	266.5	133.3	0.5847	0.5606496
amountA	1	3168.3	3168.3	13.9001	0.0004511 ***
Residuals	56	12764.1	227.9		

- ▶ The line for proteinA tests whether both indicator variables for protein are needed in the model.
- ▶ No evidence of a difference between in mean gain between the three different sources of protein (but we'll revisit this).

ANOVA for Fitted Model (cont.)

What if we try adding the variables in the opposite order:

```
> ratsA1.lm=lm(gain~amountA+proteinA)
> anova(ratsA1.lm)
```

Analysis of Variance Table

Response: gain

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
amountA	1	3168.3	3168.3	13.9001	0.0004511 ***
proteinA	2	266.5	133.3	0.5847	0.5606496
Residuals	56	12764.1	227.9		

- ▶ Exactly the same results as before – this is a consequence of using a balanced orthogonal design.

Different Indicator Variables

We can adjust the indicator variables that *R* uses for factors.

- ▶ We may decide that we would rather have cereal be the baseline for protein and low be the baseline for amount.

```
> proteinB<-factor(protein,levels=c("cereal","pork","beef"))
```

```
> contrasts(proteinB)
```

	pork	beef
cereal	0	0
pork	1	0
beef	0	1

```
> amountB<-factor(amount,levels=c("low","high"))
```

```
> contrasts(amountB)
```

	high
low	0
high	1

New Version of the Model

The new version of the model defines the means for the combinations of protein source and amount:

	<u>Protein Source</u>		
amount	beef	cereal	pork
high	$\beta_0 + \beta_B + \beta_H$	$\beta_0 + \beta_H$	$\beta_0 + \beta_P + \beta_H$
low	$\beta_0 + \beta_B$	β_0	$\beta_0 + \beta_P$

Fitted Model

The coefficients for this new version of the model are:

```
> ratsB.lm=lm(gain~proteinB+amountB)
```

```
> summary(rats2.lm)
```

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	77.633	3.898	19.916	< 2e-16 ***
proteinBpork	4.200	4.774	0.880	0.382767
proteinBbeef	4.700	4.774	0.984	0.329126
amountBhigh	14.533	3.898	3.728	0.000451 ***

Residual standard error: 15.1 on 56 degrees of freedom

Multiple R-squared: 0.212, Adjusted R-squared: 0.1698

F-statistic: 5.023 on 3 and 56 DF, p-value: 0.003739

ANOVA

The ANOVA table for the new version of the model is exactly the same as before (changing the way we define indicator variables does not affect this analysis).

```
> anova(rats2.lm)
```

Analysis of Variance Table

Response: gain

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
proteinB	2	266.5	133.3	0.5847	0.5606496
amountB	1	3168.3	3168.3	13.9001	0.0004511 ***
Residuals	56	12764.1	227.9		

A Third Version

There are many different ways of defining indicator variables for factors. *R* has a built-in method of generating indicator variables that produces the “Helmert contrasts”.

```
> proteinC<-factor(protein,levels=c("pork","beef","cereal"))
> contrasts(proteinC)<-contr.helmert(3)
> contrasts(proteinC)
      [,1] [,2]
pork    -1  -1
beef     1  -1
cereal   0   2
>
> amountC<-factor(amount,levels=c("low","high"))
> contrasts(amountC)<-contr.helmert(2)
> contrasts(amountC)
      [,1]
low     -1
high     1
```

Helmert Contrasts

The first Helmert contrast compares the mean response for the second level of a factor to that for the first level. For each subsequent level, the mean response is compared to average mean response for all preceding levels.

For our example:

- ▶ The first contrast generated for `protein` compares the mean response for beef to that for pork.
- ▶ The second contrast for `protein` compares the mean response for cereal to the average mean response for beef and pork.
- ▶ The contrast for `amount` compares the mean response for high to that for low.

Helmert Version of the Model

Using the Helmert contrasts as our indicator variables defines the means for the combinations of protein source and amount quite differently from the baseline models. Let β_0 be intercept coefficient, β_{P1} and β_{P2} the coefficients for the protein contrasts and β_{A1} be the coefficients for the amount contrast.

amount	<u>Protein Source</u>		
	beef	cereal	pork
high	$\beta_0 + \beta_{A1} + \beta_{P1} - \beta_{P2}$	$\beta_0 + \beta_{A1} + 2\beta_{P2}$	$\beta_0 + \beta_{A1} - \beta_{P1} - \beta_{P2}$
low	$\beta_0 - \beta_{A1} + \beta_{P1} - \beta_{P2}$	$\beta_0 - \beta_{A1} + 2\beta_{P2}$	$\beta_0 - \beta_{A1} - \beta_{P1} - \beta_{P2}$

Fitted Model: Version 3

```
> ratsC.lm=lm(gain~proteinC+amountC)
```

```
> summary(ratsC.lm)
```

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	87.867	1.949	45.081	< 2e-16 ***
proteinC1	0.250	2.387	0.105	0.916965
proteinC2	-1.483	1.378	-1.076	0.286414
amountC1	7.267	1.949	3.728	0.000451 ***

Residual standard error: 15.1 on 56 degrees of freedom

Multiple R-squared: 0.212, Adjusted R-squared: 0.1698

F-statistic: 5.023 on 3 and 56 DF, p-value: 0.003739

ANOVA: Version 3

The ANOVA table is the same for this version of our model as it was for the first two versions.

```
> anova(ratsC.lm)
```

Analysis of Variance Table

Response: gain

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
proteinC	2	266.5	133.3	0.5847	0.5606496
amountC	1	3168.3	3168.3	13.9001	0.0004511 ***
Residuals	56	12764.1	227.9		

Equivalent Models

Changing the way we designate the indicator variables for factors doesn't change the model. If we look at the table of fitted values for the six combinations of protein and amount we get the same estimates for all three versions:

amount	<u>Protein Source</u>		
	beef	cereal	pork
high	96.87	92.17	96.37
low	82.33	77.63	81.83

Interaction?

Our model assumes that the impact of protein doesn't depend on the level of amount and vice versa.

- ▶ The difference between the two levels of amount is the same for each level of protein.
- ▶ The difference between any two levels of protein is the same for each level of amount.

However, if we examine our conditional plot (slide 7) this doesn't seem to be the case.

The Interaction Model

If we include the interaction between `amount` and `protein`, then the impact of `amount` can be different for each level of `protein` (and vice versa).

- ▶ The interaction contrasts for the `amount:protein` interaction are obtained by multiplying each `amount` contrast by each `protein` contrast.
- ▶ It doesn't matter which version of our model we start with, adding the `amount:protein` interaction will have the same impact although the coefficients will be different and have different interpretations.

The Interaction Contrasts

Adding the interaction to the first version of our model (baseline model with beef and high as the baseline levels) gives the following table of indicator variables.

protein	amount	<u>Contrasts</u>				
		Main Effects			Interactions	
		cereal	pork	low	cereal:low	pork:low
beef	high	0	0	0	0	0
beef	low	0	0	1	0	0
cereal	high	1	0	0	0	0
cereal	low	1	0	1	1	0
pork	high	0	1	0	0	0
pork	low	0	1	1	0	1

The Model Means

The table of model means (see slide 11) becomes:

amount	<u>Protein Source</u>		
	beef	cereal	pork
high	β_0	$\beta_0 + \beta_C$	$\beta_0 + \beta_P$
low	$\beta_0 + \beta_L$	$\beta_0 + \beta_C + \beta_L + \beta_{C:L}$	$\beta_0 + \beta_P + \beta_L + \beta_{P:L}$

- ▶ The difference between the two levels of amount is not necessarily the same for each level of protein.
- ▶ The difference between any two levels of protein is not necessarily the same for each level of amount.

The Fitted Interaction Model

```
> ratsAI.lm=lm(gain~proteinA*amountA)
> summary(ratsAI.lm)
```

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)	
(Intercept)	1.000e+02	4.632e+00	21.589	< 2e-16	***
proteinAcereal	-1.410e+01	6.551e+00	-2.152	0.03585	*
proteinApork	-5.000e-01	6.551e+00	-0.076	0.93944	
amountAllow	-2.080e+01	6.551e+00	-3.175	0.00247	**
proteinAcereal:amountAllow	1.880e+01	9.264e+00	2.029	0.04736	*
proteinApork:amountAllow	2.247e-15	9.264e+00	0.000	1.00000	

Residual standard error: 14.65 on 54 degrees of freedom

Multiple R-squared: 0.2848, Adjusted R-squared: 0.2185

F-statistic: 4.3 on 5 and 54 DF, p-value: 0.002299

ANOVA Table

```
> anova(ratsAI.lm)
```

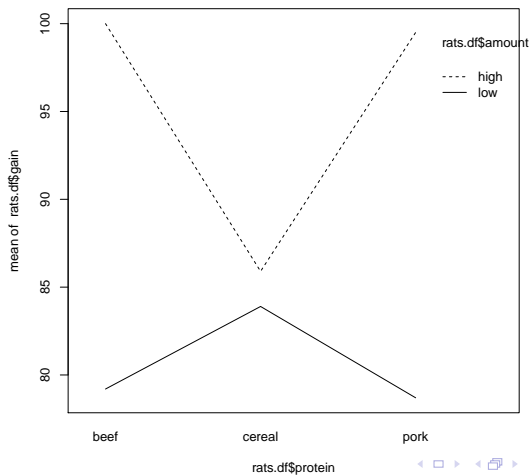
Analysis of Variance Table

Response: gain

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
proteinA	2	266.5	133.3	0.6211	0.5411319
amountA	1	3168.3	3168.3	14.7666	0.0003224 ***
proteinA:amountA	2	1178.1	589.1	2.7455	0.0731879 .
Residuals	54	11586.0	214.6		

- ▶ Moderate evidence of a protein:amount interaction.
- ▶ If an interaction is significant it implies that both factors have an impact on the response (keep main effects in the model regardless of whether they show up as significant or not).

Interaction Plot



Binary ANOVA: Plum Root Stock Example

A study was conducted on the reproduction of plum trees by taking cuttings from older trees.

- ▶ Half the cuttings were planted immediately while the other half were bedded in sand until spring when they were planted.
- ▶ Two lengths of cuttings were used: long (12 cm) and short (6cm).
- ▶ A total of 240 cuttings were taken for each of the 4 combinations of planting time and cutting length
- ▶ Interested in the survival rate of the cuttings.

Plum Root Stock Data

Length of cutting	Time of planting	Number surviving (out of 240)
short	at once	107
	in spring	31
long	at once	156
	in spring	84

- Note: the model that contains both main effects and their interaction is the maximal model.

Plum Root Stock Analysis

```
> length<-rep(c("short","long"),c(2,2))
> time<-c("at once","spring","at once","spring")
> survive<-c(107,31,156,84)
> time<-C(factor(time),treatment)
> length<-C(factor(length),treatment)
> plum.glm<-glm(cbind(survive,240-survive)~time*length,
                 family=binomial)
> anova(plum.glm,test="Chisq")
```

Analysis of Deviance Table

	Df	Deviance	Resid. Df	Resid. Dev	Pr(>Chi)
NULL			3	151.019	
time	1	97.579	2	53.440	< 2.2e-16 ***
length	1	51.147	1	2.294	8.572e-13 ***
time:length	1	2.294	0	0.000	0.1299

Plum Root Stock Analysis (cont.)

The previous analysis suggests that the interaction can be dropped (only a hint of evidence that it is needed). The output for the no interaction model is:

```
> plum2.glm<-glm(cbind(survive,240-survive)~time+length,  
                  family=binomial)
```

```
> summary(plum2.glm)
```

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)	
(Intercept)	0.7138	0.1217	5.867	4.45e-09	***
timespring	-1.4275	0.1465	-9.747	< 2e-16	***
lengthshort	-1.0177	0.1455	-6.995	2.64e-12	***

Null deviance: 151.0193 on 3 degrees of freedom
Residual deviance: 2.2938 on 1 degrees of freedom

Fitted Values

The fitted values for the main effects only model are:

```
> plum2.glm$fitted.values  
      1      2      3      4  
0.4245994 0.1504006 0.6712339 0.3287661
```

Which are very similar to those for the maximal model:

```
> plum.glm$fitted.values  
      1      2      3      4  
0.4458333 0.1291667 0.6500000 0.3500000
```

Fitted Values (cont.)

To summarize:

Length of cutting	Time of planting	Survival probability	
		observed	fitted
short	at once	0.446	0.425
	in spring	0.129	0.150
long	at once	0.650	0.671
	in spring	0.350	0.329

Confidence Intervals

To calculate 95% confidence intervals for the fitted values:

```
> preds<-predict(plum2.glm,se=TRUE)
> endpts<-cbind(preds$fit -1.96*preds$se.fit,
                preds$fit +1.96*preds$se.fit)
> round(exp(endpts)/(1+exp(endpts)),3)
      [,1] [,2]
1 0.370 0.481
2 0.118 0.190
3 0.617 0.722
4 0.278 0.383
```

Chick Weight Gain Example

An experiment was conducted to compare different diets for feeding chickens. The diets depended on three variables:

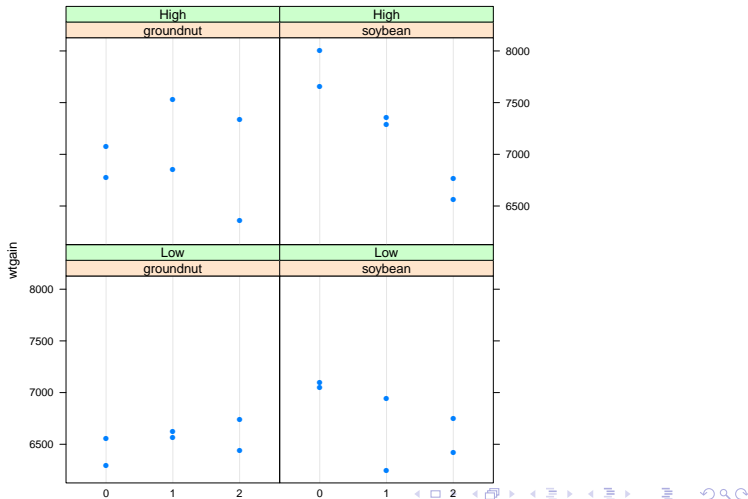
protein source of protein – either groundnut or soybean;

protlev level of protein – either 0, 1 or 2;

fish level of fish solubles – either low or high.

Response variable is **wtgain** and measures the amount of weight gained.

Dotpot of the Data



Three-Way ANOVA

In this example we have three factors. As a result we have the following possible terms for our model:

- ▶ The three main effect terms.
- ▶ Three two-factor interactions.
- ▶ The three-factor interaction.

The interpretation of a k -factor interaction is that the effect of any one of the factors depends on the combination of levels used for the remaining $k - 1$ factors.

Specifying Factors

First, we want to designate all the regressors in our data frame as factors and take the opportunity to adjust the order of levels for fish.

```
> chickwts.df<-read.table("chickwts.txt",header=TRUE)
> summary(chickwts.df)

  wtgain      protein      protlev      fish
Min.   :6249  groundnut:12  Min.     :0  High:12
1st Qu.:6560  soybean   :12  1st Qu.:0  Low :12
Median :6772                      Median :1
Mean   :6887                      Mean   :1
3rd Qu.:7144                      3rd Qu.:2
Max.   :8005                      Max.   :2
> chickwts.df$protein<-factor(chickwts.df$protein)
> chickwts.df$protlev<-factor(chickwts.df$protlev)
> chickwts.df$fish<-factor(chickwts.df$fish,
                           levels=c("Low","High"))
```

Fitting the Full Model

First we'll fit the model that contains all the possible interactions and look at the anova output.

```
> model1 <- lm(wtgain~protein*protlev*fish,data=chickwts.df)
```

```
> anova(model1)
```

Response: wtgain

	Df	Sum Sq	Mean Sq	F value	Pr(>F)	
protein	1	373751	373751	3.7346	0.077244	.
protlev	2	636283	318141	3.1789	0.078011	.
fish	1	1421553	1421553	14.2044	0.002677	**
protein:protlev	2	858158	429079	4.2874	0.039361	*
protein:fish	1	7176	7176	0.0717	0.793418	
protlev:fish	2	308888	154444	1.5432	0.253258	
protein:protlev:fish	2	50128	25064	0.2504	0.782426	
Residuals	12	1200938	100078			

Interpreting the ANOVA Table

To interpret this ANOVA table at the bottom (with the highest order interaction) and work your way up.

- ▶ For an ordinary regression with a balanced design the order that terms are added is not going to affect the hypothesis tests.
- ▶ For this table we see that there is no evidence that `protein:protlev:fish`, `protlev:fish` or `protein:fish` are needed in the model.
- ▶ Empirical evidence suggests that as the order of an interaction increases it becomes less likely to be important. If you do see a lot of significant high order interactions, try transforming the response.

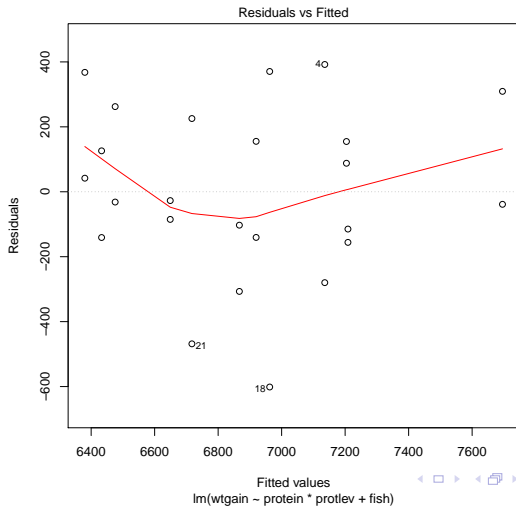
The Reduced Model

```
> model2 <- lm(wtgain~protein*protlev+fish,data=chickwts.df)
> summary(model2)
```

	Estimate	Std. Error	t value	Pr(> t)	
(Intercept)	6432.88	163.97	39.231	< 2e-16	***
proteinsoybean	776.00	214.69	3.615	0.00214	**
protlev1	216.25	214.69	1.007	0.32793	
protlev2	42.75	214.69	0.199	0.84453	
fishHigh	486.75	123.95	3.927	0.00109	**
proteinsoybean:protlev1	-707.75	303.62	-2.331	0.03232	*
proteinsoybean:protlev2	-871.50	303.62	-2.870	0.01061	*

Residual standard error: 303.6 on 17 degrees of freedom
Multiple R-squared: 0.6773, Adjusted R-squared: 0.5635
F-statistic: 5.948 on 6 and 17 DF, p-value: 0.001687

Diagnostic Plots



Interpreting the Model

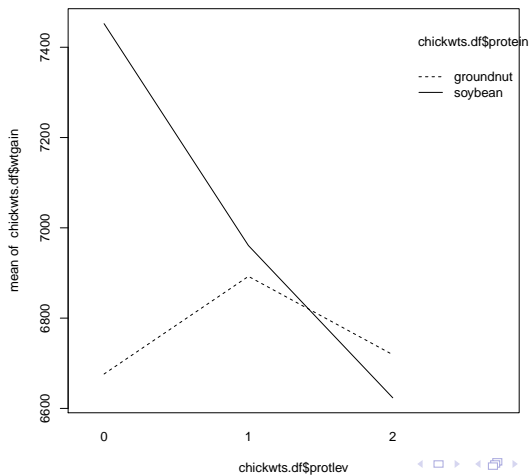
As `fish` doesn't interact with either of the other regressors we can interpret its impact separately.

- ▶ A chicken raised on a high fish solubles diet will gain on average 486.75 g(?) more than one on a low fish solubles diet for any combination of the other two factors.

As the `protein:protlev` interaction is significant we should evaluate the impact of these two factors together.

- ▶ An interaction plot is very useful in this regard.

Interaction Plot



Conclusions?

Interactions between Factors and Numeric Regressors

Interactions can exist between factors and quantitative regressors.

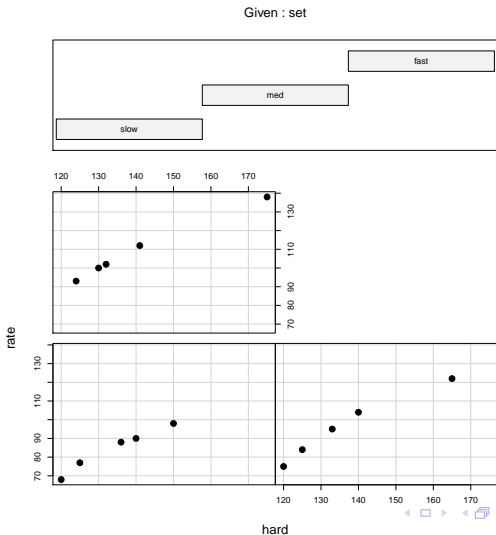
- ▶ The coefficient for a factor represents the impact on the response if we change from one level to another. If the factor interacts with a quantitative regressors, this size of the impact depends on the level of this regressor.
- ▶ The coefficient of a numeric regressor represents the change in the response per unit change in that regressor (i.e. a slope). If this regressor interacts with a factor, it means that this slope is different for different levels of the factor.

Metal Lathe Example

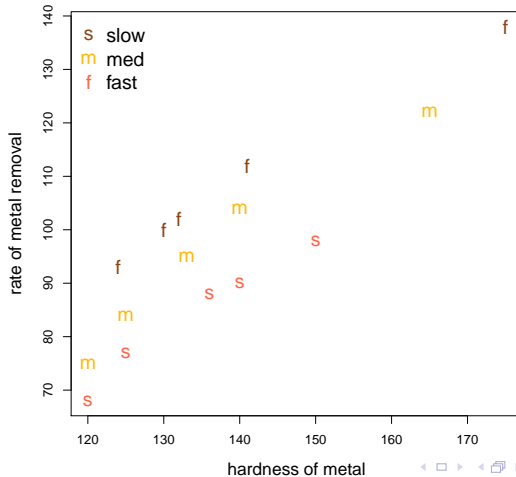
Consider an experiment to measure the rate of metal removal in a machining process on a lathe. The rate depends on:

- ▶ the speed setting of the lathe (fast, med or slow): a categorical measurement;
- ▶ the hardness of the material being machined: a continuous measurement.

Conditional Plot



Scatter Plot



Same Slopes?

Does the relationship between `rate` and `hardness` have the same slope for each level of `setting`?

- ▶ If the slopes are different then we should have the `rate:hardness` interaction in the model.
- ▶ If the slopes are the same then this interaction isn't needed.

The No Interaction Model

The baseline contrasts were used to create the indicator variables for setting:

```
> contrasts(metal.df$set)
```

	med	fast
slow	0	0
med	1	0
fast	0	1

The Model Matrix

```
> metal1.lm <- lm(rate~hard+set,data=metal.df)
> model.matrix(metal1.lm)
```

	(Intercept)	hard	setmed	setfast
1	1	120	0	0
2	1	140	0	0
3	1	150	0	0
4	1	125	0	0
5	1	136	0	0
6	1	165	1	0
7	1	140	1	0
8	1	120	1	0
9	1	125	1	0
10	1	133	1	0
11	1	175	0	1
12	1	132	0	1
13	1	124	0	1
14	1	141	0	1
15	1	130	0	1

The Model Coefficients

So for the baseline model, the relationship between rate and hard for each level of set has been parameterized as:

$$\text{set} = \text{slow} \quad \text{rate} = \beta_0 + \beta_H \text{ hard}$$

$$\text{set} = \text{med} \quad \text{rate} = \beta_0 + \beta_M + \beta_H \text{ hard}$$

$$\text{set} = \text{fast} \quad \text{rate} = \beta_0 + \beta_F + \beta_H \text{ hard}$$

The fitted lines are parallel (all have slope β_H) but different intercepts.

Model Matrix for the Full Model

```
> metal2.lm <- lm(rate~hard*set,data=metal.df)
```

```
> model.matrix(metal2.lm)
```

	(Intercept)	hard	setmed	setfast	hard:setmed	hard:setfast
1	1	120	0	0	0	0
2	1	140	0	0	0	0
3	1	150	0	0	0	0
4	1	125	0	0	0	0
5	1	136	0	0	0	0
6	1	165	1	0	165	0
7	1	140	1	0	140	0
8	1	120	1	0	120	0
9	1	125	1	0	125	0
10	1	133	1	0	133	0
11	1	175	0	1	0	175
12	1	132	0	1	0	132
13	1	124	0	1	0	124
14	1	141	0	1	0	141
15	1	130	0	1	0	130

Coefficients for the Full Model

Now the relationship between rate and hard for each level of set has been parameterized as:

$$\text{set} = \text{slow} \quad \text{rate} = \beta_0 + \beta_H \text{ hard}$$

$$\text{set} = \text{med} \quad \text{rate} = \beta_0 + \beta_M + (\beta_H + \beta_{H:M}) \text{ hard}$$

$$\text{set} = \text{fast} \quad \text{rate} = \beta_0 + \beta_F + (\beta_H + \beta_{H:F}) \text{ hard}$$

The fitted lines have different slopes and different intercepts.

Fitted No Interaction Model

```
> summary(metal1.lm)
```

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)	
(Intercept)	-41.17799	6.84927	-6.012	8.77e-05	***
hard	0.93426	0.05008	18.654	1.13e-09	***
setmed	9.55777	1.86692	5.120	0.000334	***
setfast	19.00757	1.88875	10.064	6.94e-07	***

Residual standard error: 2.946 on 11 degrees of freedom

Multiple R-squared: 0.9795, Adjusted R-squared: 0.9739

F-statistic: 175.1 on 3 and 11 DF, p-value: 1.455e-09

Fitted Full Model

```
> summary(metal2.lm)
```

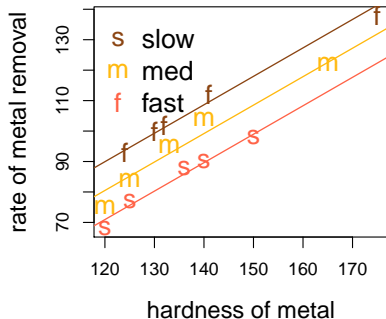
Coefficients:

	Estimate	Std. Error	t value	Pr(> t)	
(Intercept)	-45.78282	16.64522	-2.751	0.0225	*
hard	0.96858	0.12364	7.834	2.62e-05	***
setmed	3.44396	20.25964	0.170	0.8688	
setfast	33.60120	19.58902	1.715	0.1204	
hard:setmed	0.04415	0.14947	0.295	0.7744	
hard:setfast	-0.10546	0.14356	-0.735	0.4813	

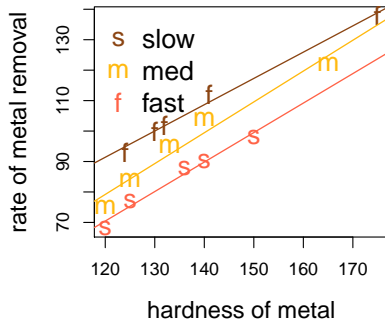
Residual standard error: 2.959 on 9 degrees of freedom
Multiple R-squared: 0.9831, Adjusted R-squared: 0.9737
F-statistic: 104.5 on 5 and 9 DF, p-value: 1.085e-07

Which Fit?

No Interaction Model



Interaction Model



ANOVA Table

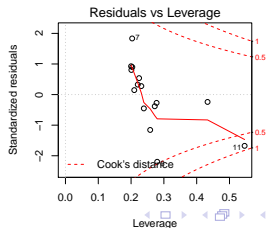
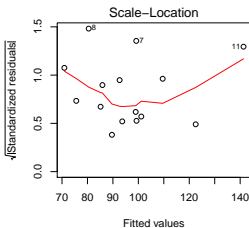
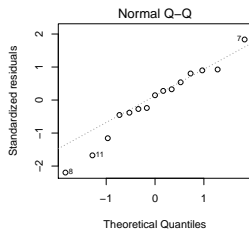
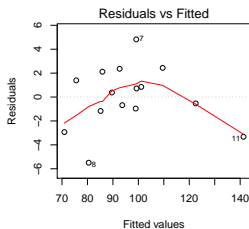
To decide we look at the output of anova for the full model.

```
> anova(metal2.lm)
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)	
hard	1	3679.3	3679.3	420.1866	7.307e-09	***
set	2	878.8	439.4	50.1834	1.316e-05	***
hard:set	2	16.6	8.3	0.9504	0.4222	
Residuals	9	78.8	8.8			

- ▶ No evidence that the interaction is need – the parallel lines model is fine.

Diagnostic Plots



Conclusions?

Interactions Between Numerical Regressors

Interaction terms between numerical regressors can also be useful in a regression model.

- ▶ A bit harder to understand but work much the same way: an interaction allows the impact of one regressor to change with the level of another.
- ▶ A two-factor interaction allows the surface to twist in the direction of one regressor as the value of another regressor changes.

Scottish Hill Races

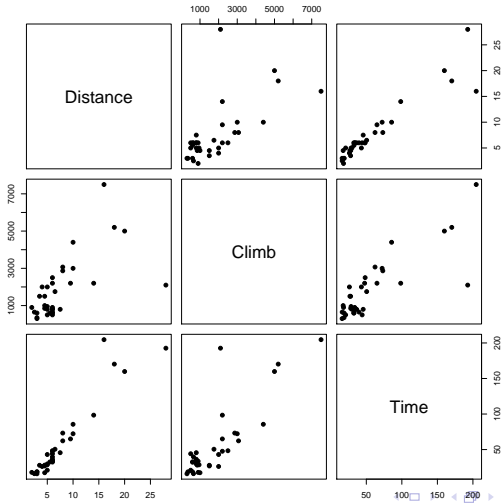
This data consists of the record times for 35 “hill” races in Scotland.

Distance the length of the race in miles.

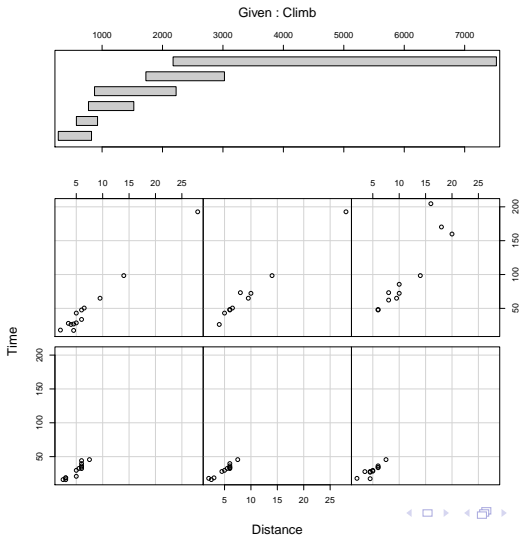
Climb the change in elevation in feet.

Time the record winning time in minutes.

Pairs Plot



Conditional Plot



Interaction Model

The conditional plot suggests that an interaction between Distance and Climb may be useful.

```
> hills1.lm<-lm(Time~Climb*Distance,data=hills.df)
```

```
> anova(hills1.lm)
```

Response: Time

	Df	Sum Sq	Mean Sq	F value	Pr(>F)	
Climb	1	59667	59667	1104.372	< 2.2e-16	***
Distance	1	23997	23997	444.163	< 2.2e-16	***
Climb:Distance	1	803	803	14.865	0.0005449	***
Residuals	31	1675	54			

The Model Matrix

```
> model.matrix(hills1.lm)
      (Intercept)  Climb Distance Climb:Distance
1              1    650       2.5          1625
2              1   2500       6.0         15000
3              1    900       6.0          5400
4              1    800       7.5          6000
5              1   3070       8.0         24560
6              1   2866       8.0         22928
7              1   7500      16.0        120000
8              1    800       6.0          4800
9              1    800       5.0          4000
=====
31             1   4400      10.0         44000
32             1    600       6.0          3600
33             1   5200      18.0         93600
34             1    850       4.5          3825
35             1   5000      20.0        100000
```

The Fitted Model

```
> summary(hills1.lm)
```

Coefficients:

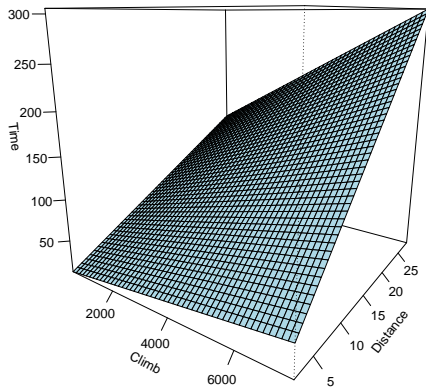
	Estimate	Std. Error	t value	Pr(> t)	
(Intercept)	-0.3532285	3.9121887	-0.090	0.928638	
Climb	0.0035217	0.0023686	1.487	0.147156	
Distance	4.9290166	0.4750168	10.377	1.32e-11	***
Climb:Distance	0.0006731	0.0001746	3.856	0.000545	***

Residual standard error: 7.35 on 31 degrees of freedom

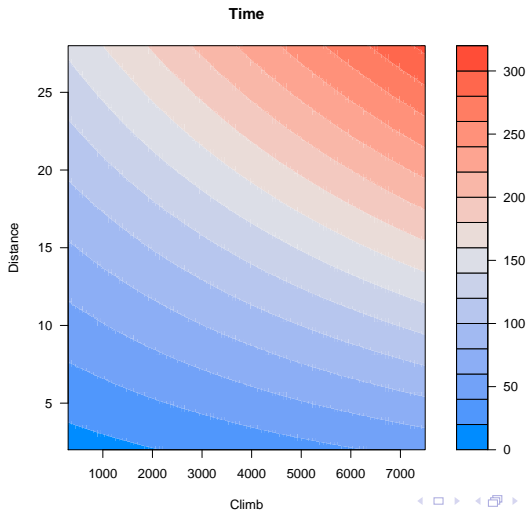
Multiple R-squared: 0.9806, Adjusted R-squared: 0.9787

F-statistic: 521.1 on 3 and 31 DF, p-value: < 2.2e-16

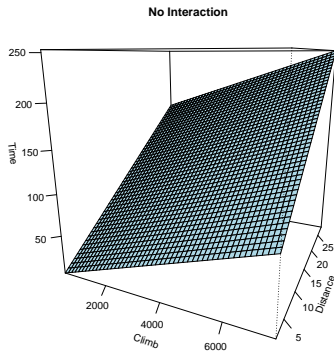
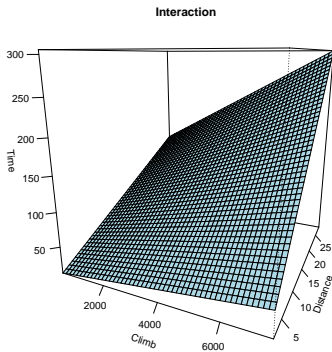
Wireframe Plot



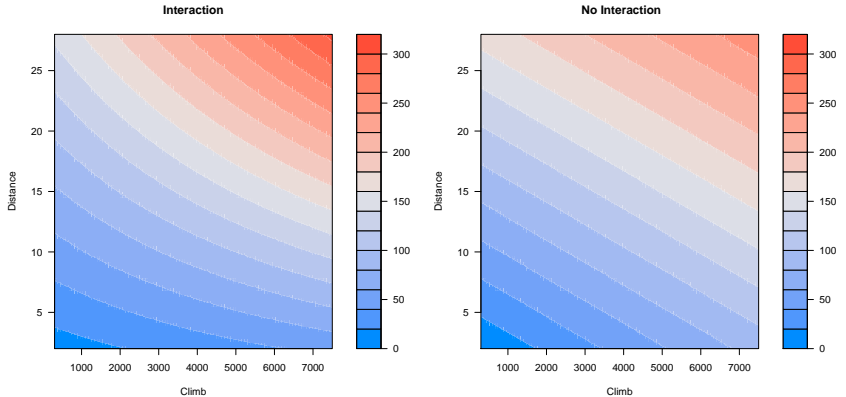
Filled Contour Plot



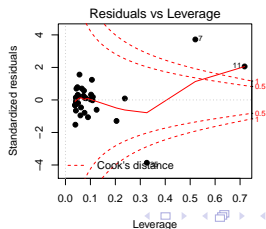
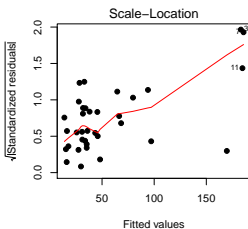
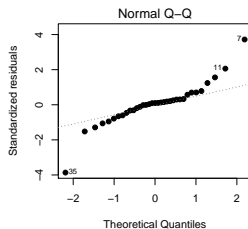
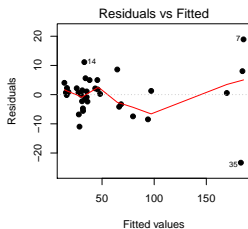
Comparison of Surfaces



Comparison of Surfaces



Diagnostic Plots



Log the Response?

Might try using the log of the response – expect variability to increase as the expected time increases.

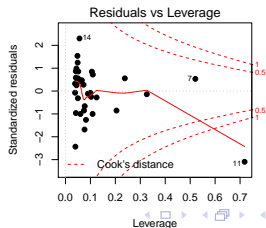
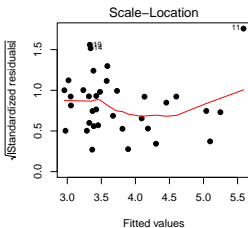
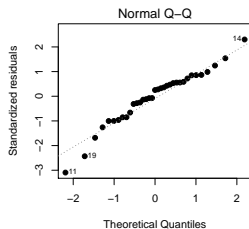
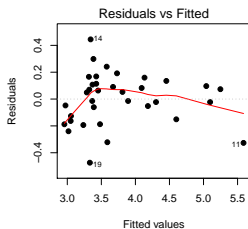
```
> hills3.lm<-lm(log(Time)~Climb*Distance,data=hills.df)
> anova(hills3.lm)
```

Analysis of Variance Table

Response: log(Time)

	Df	Sum Sq	Mean Sq	F value	Pr(>F)	
Climb	1	11.5876	11.5876	293.087	< 2.2e-16	***
Distance	1	4.0030	4.0030	101.250	2.762e-11	***
Climb:Distance	1	0.4301	0.4301	10.879	0.002447	**
Residuals	31	1.2256	0.0395			

Diagnostic Plots



Fitted Model

```
> summary(hills3.lm)
```

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)	
(Intercept)	2.517e+00	1.058e-01	23.780	< 2e-16	***
Climb	3.686e-04	6.407e-05	5.753	2.49e-06	***
Distance	1.147e-01	1.285e-02	8.929	4.46e-10	***
Climb:Distance	-1.558e-05	4.722e-06	-3.298	0.00245	**

Residual standard error: 0.1988 on 31 degrees of freedom

Multiple R-squared: 0.9289, Adjusted R-squared: 0.9221

F-statistic: 135.1 on 3 and 31 DF, p-value: < 2.2e-16