Supplementary Note

GameTag: A New Sequence Tag Generation Algorithm Based on Cooperative Game Theory

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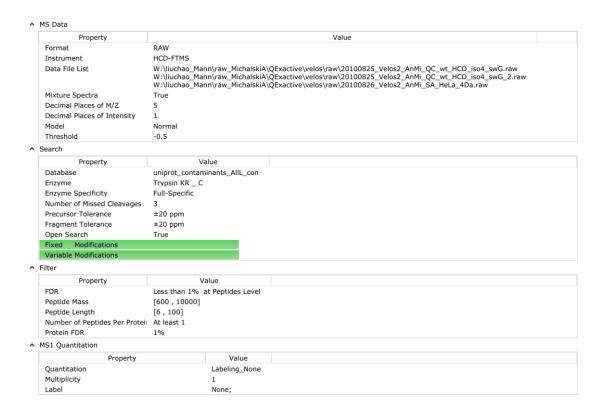
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1. Dataset Annotation

In this section, we provide the detailed parameter settings of Open-pFind, PEAKS, and MSFragger, for dataset pre-processing and annotating. The intersection of identification results from the three can be considered as ground-truth PSMs. In practice, we employed data label union.py for intersection operation and result statistics.

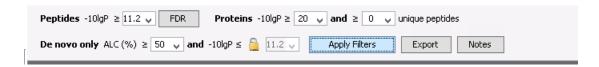
1.1 Open-pFind

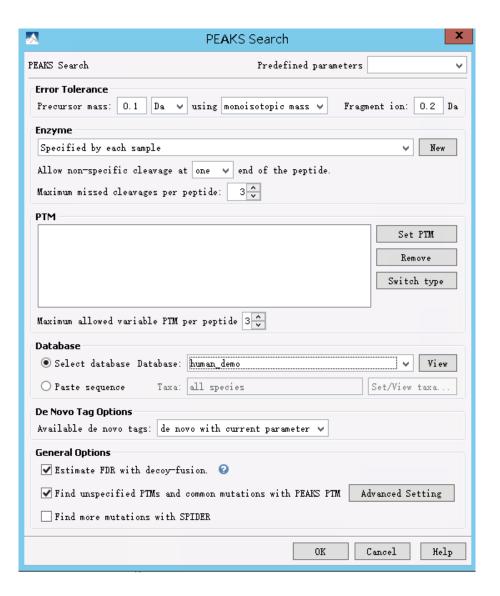
- 1) Tool download: http://pfind.ict.ac.cn/software/pFind3/index.html
- 2) Parameter setting interface:(Please note that no screenshots are all set by default)



1.2 PEAKS

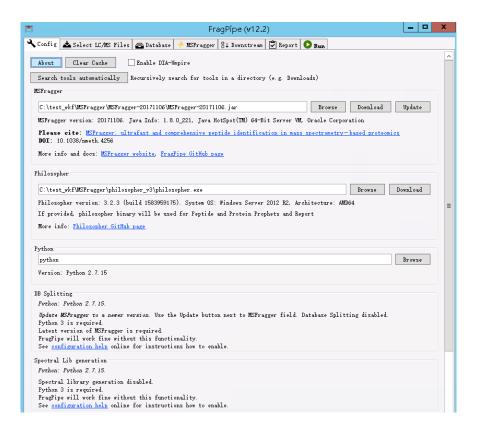
- 1) Tool download: https://www.bioinfor.com/download-peaks-studio/
- 2) Parameter setting interface:

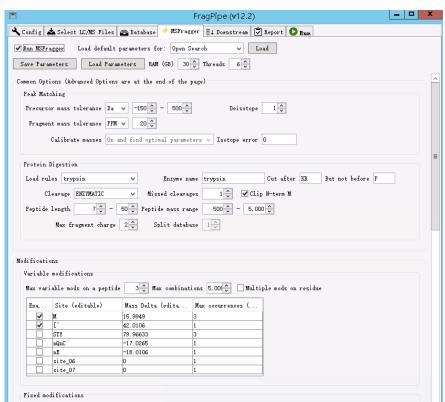




1.3 MSFragger

- 1) Tool download: http://msfragger.nesvilab.org/
- 2) Parameter setting interface:





2. Baseline Setting

In this section, we give the parameter setting for the baselines employed in the experiments for tagging performance comparison.

2.1 InsPecT

- 1) Tool download: http://proteomics.ucsd.edu/Software/Inspect/
- 2) Dataset format conversion (we use .mgf as spectrum file): http://tools.proteomecenter.org/wiki/index.php?title=Main Page
- 3) Database Setup: run CMD as belows> python PrepDB.py FASTA [myDB.fasta]
- 4) Parameter InputFile:

```
speactra, [FILENAME.mgf]
protease, Trypsin
mod, 57, C, fix
TagCount, 100
TagLength, 5
```

5) Run the InsPect:

```
> InsPecT.exe -i InputFile.text -o OutputFile.txt
```

(In order to output the extracted tags, we can adopt the "DEBUG" module, more detail can be refer to document: <a href="http://proteomics.ucsd.edu/Software/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/Inspect/In

2.2 PepNovo+

- 1) Tool download: https://github.com/jmchilton/pepnovo
- 2) Run the pepnovo+ with the below command:

```
>PepNovo.exe -file [mgfPath] -model CID_IT_TRYP
C+57:M+16 -digest NON_SPECIFIC -tag_length 5 -
num_solutions 100 -fragment_tolerance 0.01 >
[OutputFilePath]
```

(Note that we utilize the high-resolution data version for tag extraction. In practice, If the MS/MS spectra come from high-resolution instruments, the sequencing performance can be improved by manipulating the tolerances. For instance if the spectra have fragment tolerances of 0.01, this can be set with the flag: - fragment tolerance 0.01.)

2.3 SVM:

- 1) Package download: https://www.csie.ntu.edu.tw/~cjlin/libsvm/
- 2) Parameter setting:

Input features	Peak intensity
	Edge mass error
	Node relevance degree
Feature Normalization	Yes
Kernel	RBF
С	512
Gamma	0.03125
Validation	5-fold cross-validation

(For convenience, the tag features produced by tag generator are used directly.)

3) Train the SVM model as:

>svm-train.exe -t 2 -g 0.03125 -c 512 -v 5 -s 1 [DatasetName] [ModelName]

4) Predict the results using the trained SVM model:

>svm-predict.exe [DatasetName] [ModelName] [OutputFile]

3. Additional Experimental Results

Due to the limitation of paper space, we will present some addition results here.

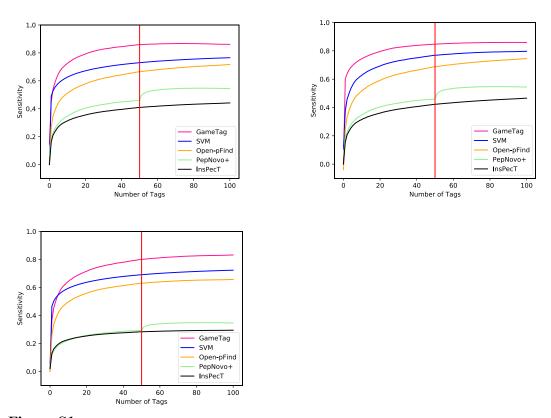


Figure S1: Sensitivity of different methods as a function of the number of tags considered for each spectrum on Gygi-Human-QE, Dong-Ecoli-QE, and Xu-Yeast-QEHF, respectively.