

# Spectral Characterization of Four Kinds of Biodegradable Plastics: Poly (Lactic Acid), Poly (Butylenes Adipate-Co-Terephthalate), Poly (Hydroxybutyrate-Co-Hydroxyvalerate) and Poly (Butylenes Succinate) with FTIR and Raman Spectroscopy

Yanming Cai · Jungang Lv · Jimin Feng

Published online: 18 September 2012  
© Springer Science+Business Media, LLC 2012

**Abstract** Fourier transform infrared microscope and confocal Raman spectroscopy were employed in this study to investigate four kinds of biodegradable plastics: poly(lactic acid), poly(butylenes adipate-co-terephthalate), poly(butylenes succinate) and poly(hydroxybutyrate-co-hydroxyvalerate), which are used more and more popularly in everyday life but can not be identified easily with other instruments. Infrared and Raman spectra of the plastics were tentatively interpreted. The indicative peaks to characterize the four polymers were also summarized. The result in this study can help the forensic scientists discriminate the plastics accurately when they occurred as trace evidences in cases, it also offers the producer and environment scientists an effective, non-invasive and fast method to characterize and identify these four polymers.

**Keywords** Fourier transform infrared spectroscopy · Confocal Raman spectroscopy · Biodegradable plastics

## Introduction

Discarded synthetic plastics products induced serious problem of solid waste pollution, so many studies has been carried out on the biodegradable polymers which could offer an effective way to replace the commodity synthetic

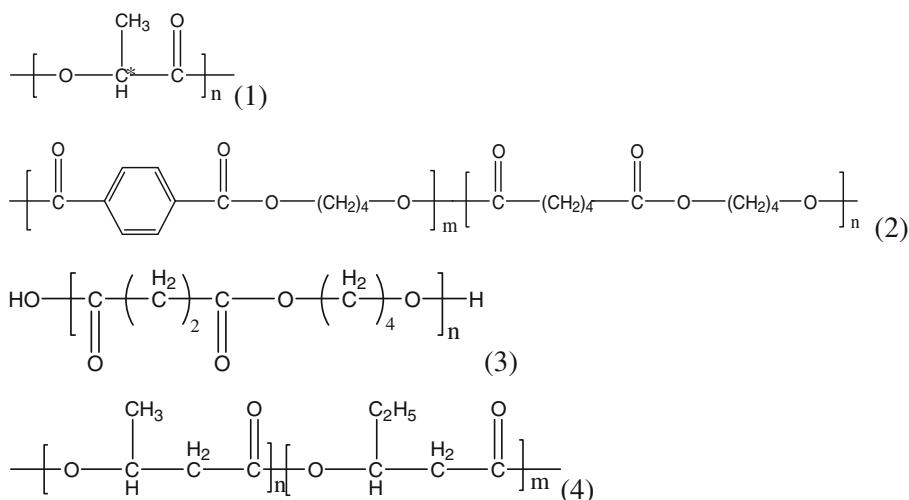
polymers and solve the solid waste pollution in recent years [1, 2]. Poly(lactic acid) (PLA, Fig. 1(1)), poly(butylenes adipate-co-terephthalate) (PBAT, Fig. 1(2)), poly(butylenes succinate) (PBS, Fig. 1(3)) and poly(hydroxybutyrate-co-hydroxyvalerate) (PHBV, Fig. 1(4)) are four kinds of biodegradable plastics used most popularly in people's everyday lives.

Poly(lactic acid) is a thermoplastic aliphatic polyester derived from renewable resources which can easily be biodegraded under the presence of oxygen. Poly-L-lactide (PLLA) is the product used most popularly in everyday life among several distinct forms of PLAs. Poly-L-lactide has a crystallinity of around 37 %, a glass transition temperature between 60 and 65 °C, a melting temperature between 173 and 178 °C and a tensile modulus between 2.7–16 GPa [3]. PLA has good mechanical, thermal and biodegradable properties and therefore is a good polymer for various end-use applications. However, high brittleness and cost of PLA is the major problem for their commercialization and applications [4, 5]. PBAT is also a commercially available biodegradable plastics now widely used even worldwide. PBAT is an aliphatic-co-aromatic co-polyester synthesized by esterification of 1, 4-butanediol with aromatic dicarboxylic acid and then polycondensation with succinic acid. PBAT has many advantages among all the biodegradable materials, including high flexibility, excellent impact strength and melt processibility. In particular PBAT has very fast biodegradation rate. It is reported that whether in the form of films or molded objects, PBAT has exhibited significant biodegradation within 1 year in soil, water with activated sludge, and seawater [6–9]. PBS is another biodegradable aliphatic polyester produced by the polycondensation reaction of 1,4-butanediol with succinic acid [10, 11]. It has high flexibility, excellent impact

Y. Cai  
Agro-Environment Protection Institute, Ministry of Agriculture,  
Tianjin 300191, China

J. Lv (✉) · J. Feng  
Procuratorial Technology and Information Research Center,  
Supreme People's Procuratorate, Lugu East Street 5, Beijing  
100040, China  
e-mail: sdta1jg@163.com

**Fig. 1** Chemical structures of PLA (1), PBAT (2), PBS (3) and PHBV (4)



strength, and thermal and chemical resistance [12]. PBS can be processed easily. Many studies on PBS have indicated that PBS can easily be biodegraded within several months in the environment [5]. PHBV is another promising biopolymers known as polyhydroxy alkanates (PHAs) that can be produced via bacterial fermentation of plant starches. PHBV has gained special importance because of favorable mechanical characteristics, biological properties, highly adaptable structure, non-toxic degradation products, and minimal inflammatory response when used as scaffold, PHBV is also found with fast degradation rate in the environment in the previous studies [13].

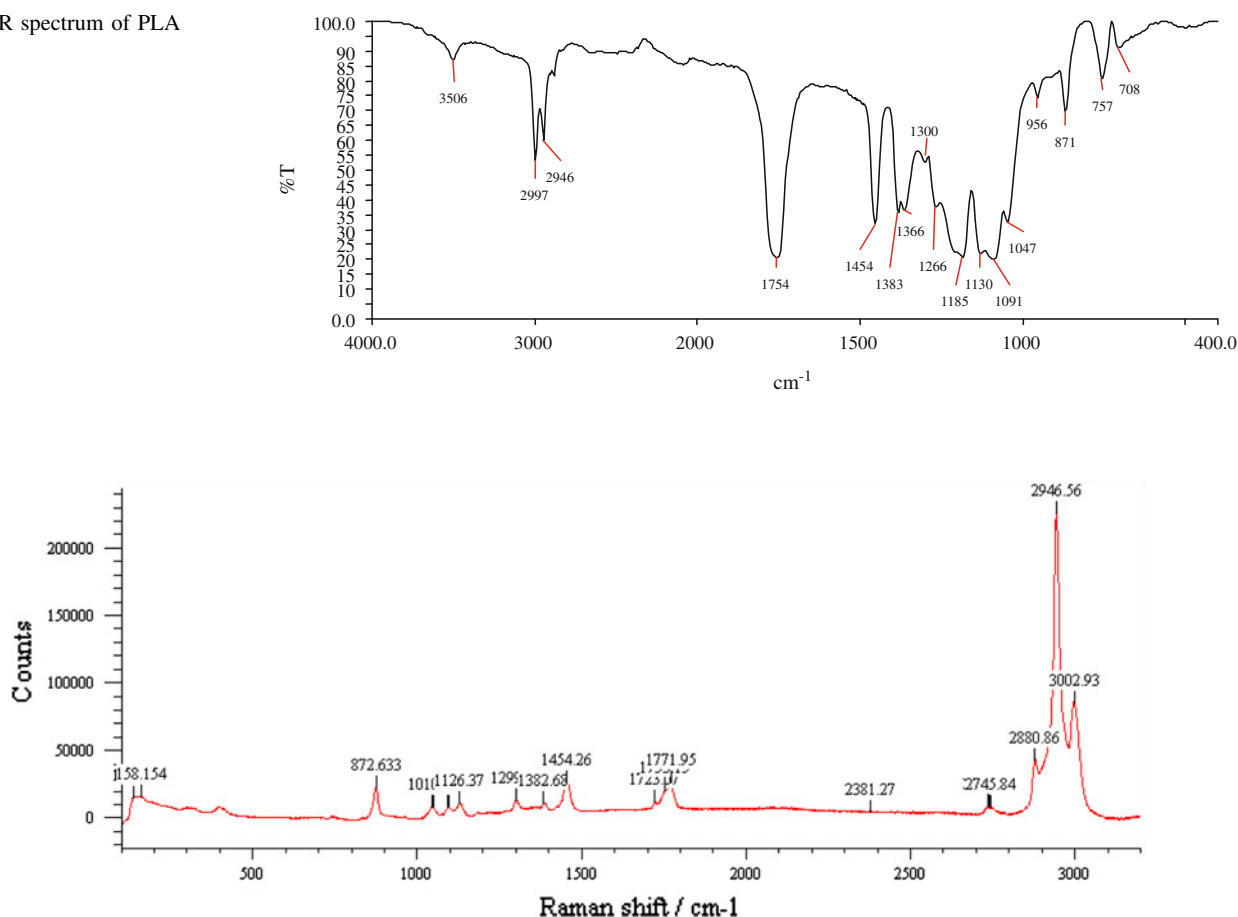
It is important to characterize the plastics in relative studies, especially in the research on synthesis and environmental degradation progress. Thermal analysis, X-ray diffraction, Nuclear magnetic resonance (NMR) and spectroscopy such as scanning electronic microscopy-energy disperse spectroscopy are all ideal methods to characterize these biodegradable plastics. However, previous studies only focused on the characterization of mechanical (tensile and impact) properties, thermal properties, rheological properties, morphology and biodegradability in the synthesis and production research [14–18]. IR characterization was also involved in some previous studies, but peak assignment of the functional groups was not systemic and detailed enough. To our best of knowledge, few studies on fast and noninvasive characterization of these four kinds of plastics with Raman, IR and Raman in combination were found. FTIR spectroscopy had been proved to be an effective tool with many advantages in the characterization of plastics [19, 20]. Very few sample was needed for analysis and high-quality spectra could be obtained only in a few seconds. Raman spectroscopy also requires only small sample volumes and minimal sample

preparation, the analysis is fast and accurate. Raman spectroscopy can cover the shortage of FTIR spectroscopy (such as serious water interference) and give extra structure information. More structure information would be obtained if these two methods used in combination, and optical observation with these two methods were performed in the study to investigate the spectral characteristics.

## Materials and Methods

Four kinds of biodegradable plastics samples were kindly presented by the National Technical Committee on Plastic Products of Standardization Administration of China. A Spectrum GX 2000 system from Perkin-Elmer Company with diamond anvil cell, DTGS (Deuterated triglycine sulfate) detector and Spectra 5.01 workstation was employed for IR investigation. Background was subtracted for every measurement. Triplicate tests were performed at different sites for every sample.

A Renishaw inVia confocal Raman microscope system with two lasers emitting at 532 nm/633 nm, charge-coupled device (CCD) detector was employed collecting the Raman spectrum. 532 nm laser was chosen for emission in 10–100 % power depending on whether the laser would damage the sample or not. Before every sample measurement, crystal silicon with fixed peak position at 520 cm<sup>-1</sup> was used to calibrate the Raman shifts. The spectra were collected in WIRE3 workstation in extensive mode ranging from 100 to 3,200 cm<sup>-1</sup>. For some samples, most peaks appeared in the bands of 150–2,000 cm<sup>-1</sup>, so 2,000–3,200 cm<sup>-1</sup> were not shown in this study and the spectrum in 150–2,000 cm<sup>-1</sup> was magnified to get better

**Fig. 2** IR spectrum of PLA**Fig. 3** Raman spectrum of PLA

spectrum. Every spectrum was obtained by 3–15 accumulations to enhance the signal/noise ratio.

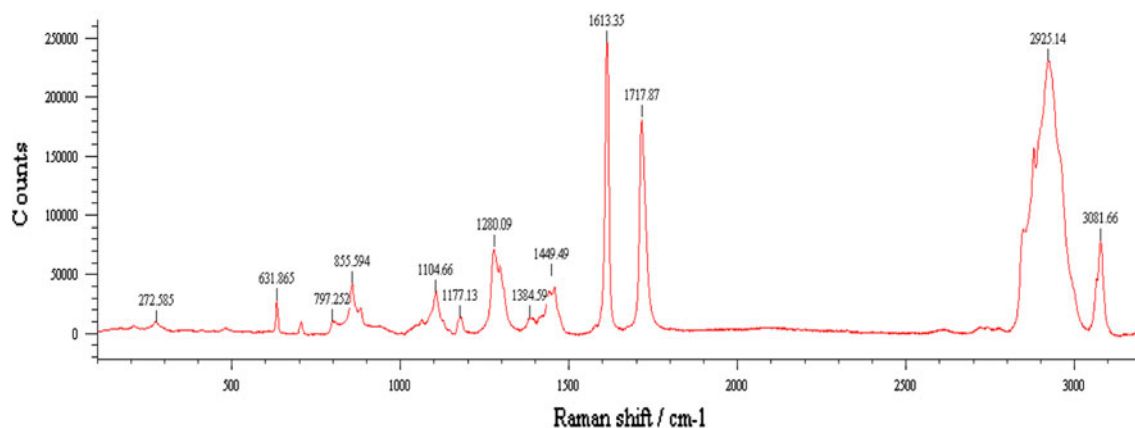
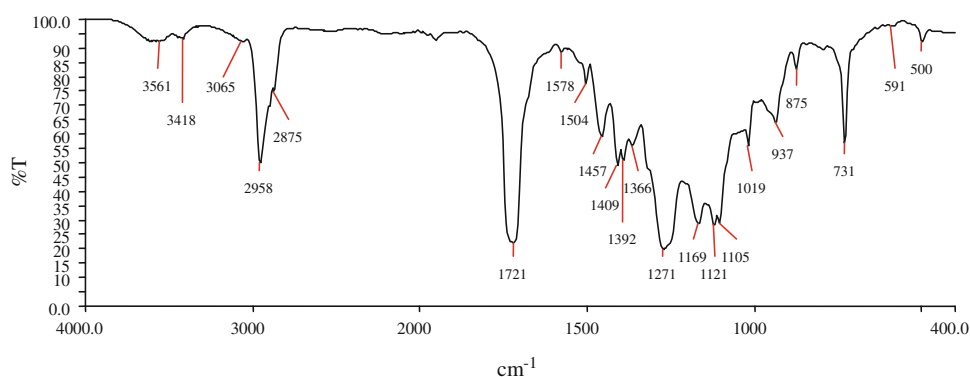
## Results and Discussion

### Spectrum of PLA

Figures 2 and 3 showed the IR and Raman spectra of PLA separately. Sixteen peaks: 3,506, 2,997, 2,946, 1,754, 1,454, 1,383, 1,366, 1,300, 1,185, 1,130, 1,091, 1,047, 956, 871, 757, 708  $\text{cm}^{-1}$  were identified in the IR spectra of PLA. More specifically, 3,065  $\text{cm}^{-1}$  should be assigned as the OH absorption in stretching mode; 2,997, 2,946  $\text{cm}^{-1}$   $\text{CH}_3$  stretching mode;  $\text{CH}_3$  stretching band was increased by the inductive effect of the adjoining C = O or O atom. 1,754  $\text{cm}^{-1}$  was the absorption of C = O stretching which was increased from about 1,730  $\text{cm}^{-1}$  by the hyper conjugation of  $\alpha$ -H and the C = O. 1,454  $\text{cm}^{-1}$  was the asymmetric bending absorption of  $\text{CH}_3$ ; 1,383, 1,365  $\text{cm}^{-1}$  should be assigned as the symmetric bending absorption of  $\text{CH}_3$ ; CH; 1,266, 1,185, 1,130, 1,091, 1,047  $\text{cm}^{-1}$  were all

stretching absorption of C–O–C which were complex because of the different atoms of function groups adjoining to C–O–C; 870  $\text{cm}^{-1}$  should be assigned as the absorption of O–CH– $\text{CH}_3$  and 757  $\text{cm}^{-1}$  as the wagging absorption of  $\alpha$ - $\text{CH}_3$ . The OH absorption at 3,506  $\text{cm}^{-1}$ , the shift of C = O stretching mode from 1,730 to 1,754  $\text{cm}^{-1}$  and the wagging absorption of  $\alpha$ - $\text{CH}_3$  at 757  $\text{cm}^{-1}$  were all characteristic and indicative in the identification of PLA.

IR and Raman spectroscopy can give more information by using these two instruments in combination. Some C–C or C = C absorption, especially strong absorption of C = C in benzene ring (1,620  $\text{cm}^{-1}$ ), appeared in Raman spectrum but disappeared in IR spectrum; while some absorption of polar function groups appeared in IR spectrum but disappeared in Raman spectrum. It indicated that Raman spectrum has fewer and less overlapped bands than in the infrared spectrum. Many peaks appeared at similar positions in IR and Raman, so they were not further interpreted in Raman spectrum. From Figs. 2 and 3, C = O stretching was observed at both IR (1,754  $\text{cm}^{-1}$ ) and Raman (1,772  $\text{cm}^{-1}$ ) spectrum. 315, 401, 873, 1,010, 1,096, 1,126, 1,299, 1,383, 1,454, 1,723, 1,772, 2,746,

**Fig. 4** IR spectrum of PBAT**Fig. 5** Raman spectrum of PBAT

2,881, 2,947 and 3,003  $\text{cm}^{-1}$  were the main peaks in the Raman spectrum of PLA.

#### Spectrum of PBAT

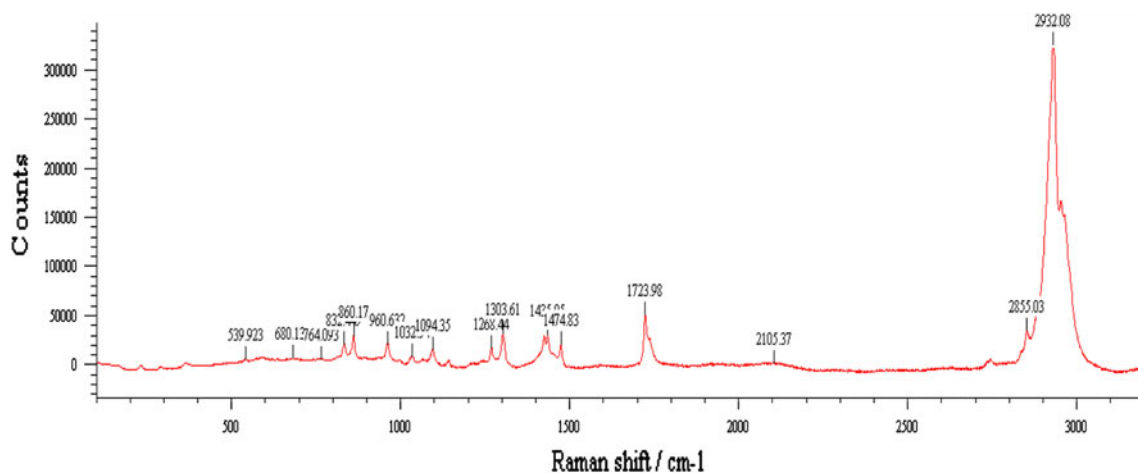
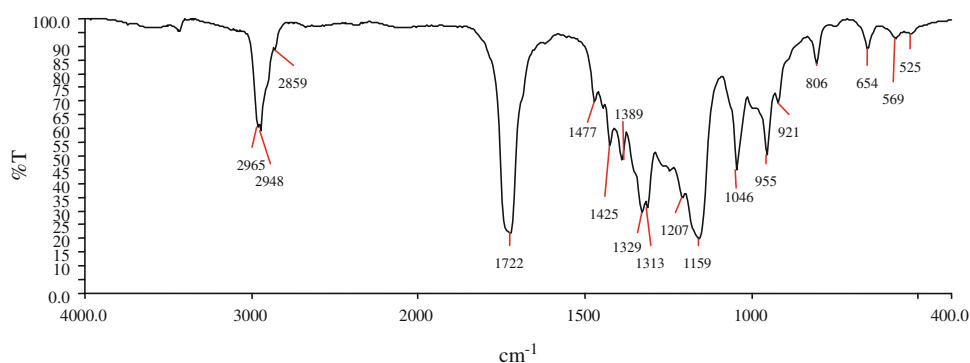
Main IR peaks of PBAT were 3,065, 2,958, 2,875, 1,392, 1,366, 1,169, 1,121 and 937  $\text{cm}^{-1}$  from Fig. 4. 3,065  $\text{cm}^{-1}$  in PBAT should be assigned as the =C–H absorption in stretching mode; 2,958, 2,875  $\text{cm}^{-1}$   $\text{CH}_3$ ,  $\text{CH}_2$  stretching mode; 1,578, 1,504  $\text{cm}^{-1}$  skeleton vibration of benzene ring; 1,457  $\text{cm}^{-1}$   $\text{CH}_2$  in-plane bending mode; 1,409  $\text{cm}^{-1}$  bending mode of O– $\text{CH}_2$ ; 1,366  $\text{cm}^{-1}$  out-plane bending mode of  $\text{CH}_2$ ; 1,271, 1,105  $\text{cm}^{-1}$  C–O stretching mode; 1,019  $\text{cm}^{-1}$  in-plane bending mode of =C–H in benzene ring; 875, 731  $\text{cm}^{-1}$  should be assigned as out-plane bending mode of =C–H in benzene ring and 850  $\text{cm}^{-1}$  as in-plane bending mode of  $\text{CH}_2$ . The out-plane bending mode of =C–H in benzene ring generally appeared in 830  $\text{cm}^{-1}$ , but influenced by the conjugation of C=O and the benzene ring, the absorption occurred in 731  $\text{cm}^{-1}$  was characteristic in the spectrum and were found to be indicative in the identification of PBAT. From the difference in the structures and spectra of PBAT and PET, 1,121,

1,169  $\text{cm}^{-1}$  should be assigned as the stretching mode of the C–O in aliphatic acid. Peaks at 731, 1,121 and 1,169  $\text{cm}^{-1}$  were characteristic and can be used as indicative peaks in the identification of PBAT.

From Fig. 5, the main peaks in the Raman spectrum of PBAT included: 119, 206, 273, 632, 704, 797, 856, 878, 1,024, 1,043, 1,105, 1,177, 1,280, 1,300, 1,396, 1,449, 1,577, 1,613, 1,673 and 1,718  $\text{cm}^{-1}$ . The peaks at 2,003, 2,760, 2,780, 2,782, 2,875, 3,045 and 3,067  $\text{cm}^{-1}$  were also found in the Raman spectrum of PBAT in the magnified spectrum, considering these peaks were not characteristic enough, they were not labeled in Fig. 5.

#### Spectrum of PBS

Figure 6 showed the IR spectrum of PBS. From Figs. 1(3) and 6, peaks in the IR spectrum of PBS should be assigned as following: 2,965, 2,948, 2,859  $\text{cm}^{-1}$ ,  $\text{CH}_3$ ,  $\text{CH}_2$  stretching mode; 1,722  $\text{cm}^{-1}$ , C=O stretching mode; 1,159  $\text{cm}^{-1}$ , C–O stretching mode; 1,046  $\text{cm}^{-1}$ , O( $\text{CH}_2$ )<sub>4</sub> O vibration; 955  $\text{cm}^{-1}$ , C–O symmetric stretching mode; 806  $\text{cm}^{-1}$ ,  $\text{CH}_2$  in OC( $\text{CH}_2$ )<sub>2</sub>CO in-plane bending mode and 654  $\text{cm}^{-1}$ , COO bending mode. Peaks at 955, 806 and

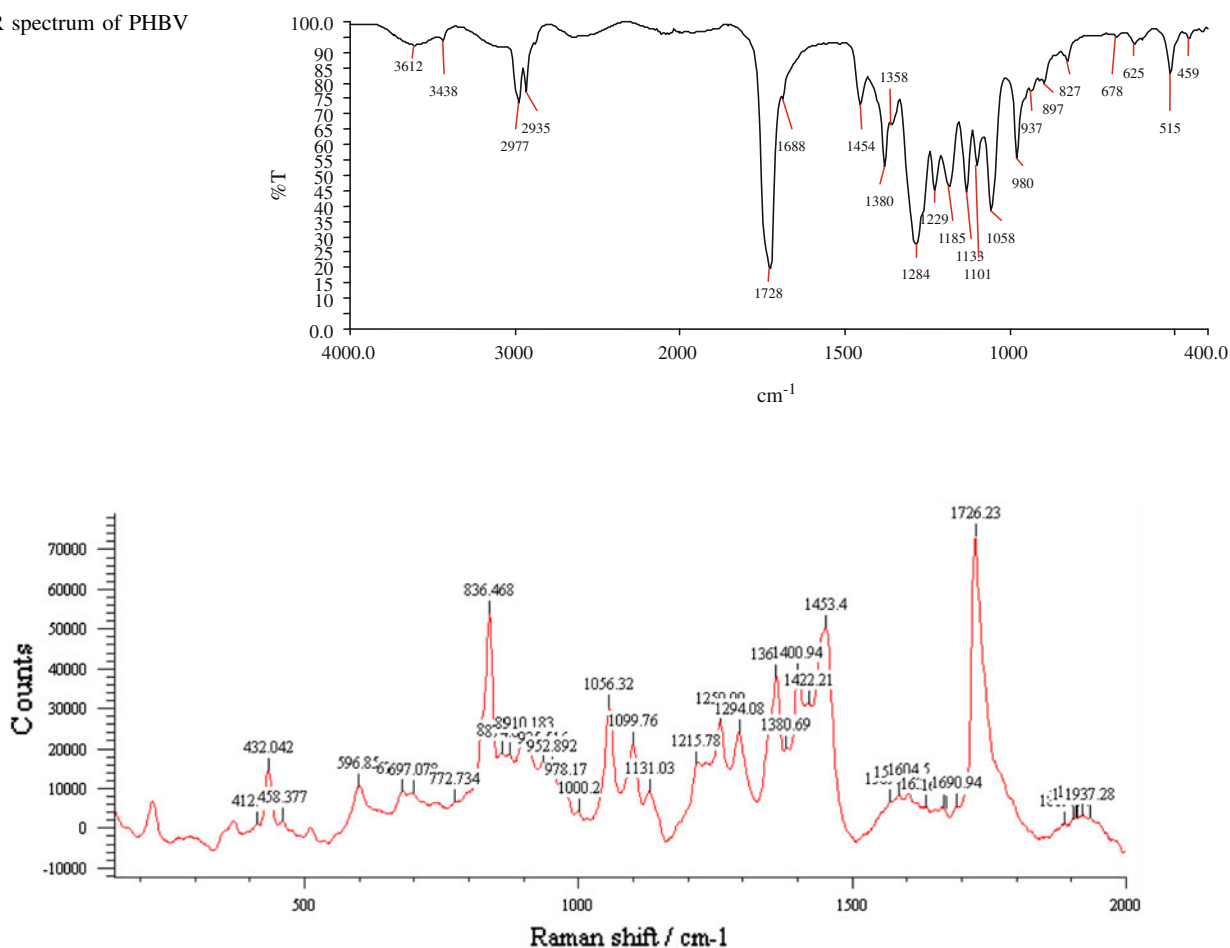
**Fig. 6** IR spectrum of PBS**Fig. 7** Raman spectrum of PBS

$654\text{ cm}^{-1}$  can be used as indicative peaks to discriminate PBS to other kinds of plastics. From Fig. 7, the main peaks in the Raman spectrum of PBS were 838, 860, 961, 1,032, 1,094, 1,268, 1,304, 1,425, 1,475, 1,724, 2,855 and  $2,932\text{ cm}^{-1}$ .

#### Spectrum of PHBV

IR spectrum of PHBV (3HB-co-3H) was shown in Fig. 8. From the chemical structure of PHBV in Figs. 1(4) and 8, Peaks in IR spectrum of PHBV should be assigned as:  $3,612\text{ cm}^{-1}$ , OH stretching mode; 2,977, 2,935,  $2,879\text{ cm}^{-1}$ ,  $\text{CH}_2$ ,  $\text{CH}_3$  stretching mode;  $1,728\text{ cm}^{-1}$ , C = O stretching mode;  $1,454\text{ cm}^{-1}$ ,  $\text{CH}_2$  bending mode; 1,380 and  $1,358\text{ cm}^{-1}$ ,  $\text{CH}_3$  bending mode;  $1,300\text{--}1,000\text{ cm}^{-1}$ , C–O–C stretching mode and  $980\text{ cm}^{-1}$ ,  $\text{CH}_2$  in-plane bending mode. Peaks at  $1,000\text{--}1,300\text{ cm}^{-1}$  in fingerprint bands and  $980\text{ cm}^{-1}$  will be helpful in PHBV identification. From Fig. 9, the main peaks in the Raman spectrum of PHBV in  $100\text{--}2,000\text{ cm}^{-1}$  were: 217, 289, 366, 432, 597, 697, 836, 910, 953, 1,056, 1,100, 1,131, 1,216, 1,250, 1,294, 1,361, 1,401, 1,453, 1,604 and  $1,726\text{ cm}^{-1}$ .

It was easy to discriminate the Raman spectrum of PBAT from PLA, PBS and PHBV. From Fig. 1, there were C = O and benzene rings structures in the PBAT chains while there were only C = O structures in the other three polymers. So, the Raman shifts relative to the benzene rings only appeared in the spectrum of PBAT. Specifically, the peak at  $3,082\text{ cm}^{-1}$  was the absorption of the unsaturated = C–H;  $1,613\text{ cm}^{-1}$  might be assigned as the absorption of the C–C stretching in the benzene ring;  $1,280\text{ cm}^{-1}$  as the C–C stretching and the –C–CH in-plane bending in the benzene ring;  $1,105\text{ cm}^{-1}$  as the –C–CH in-plane bending and 856,  $797\text{ cm}^{-1}$  as the –C–CH out-plane bending. The intense peaks at 1,613 and  $3,082\text{ cm}^{-1}$  were helpful in the identification of PBAT. The super conjugation effect of the  $\alpha$ -H and the C = O in the PLA shift the stretching vibration of C = O to  $1,772\text{ cm}^{-1}$ . It was significantly higher than those in the other compounds with C = O including PBS ( $1,724\text{ cm}^{-1}$ ) and PHBV ( $1,726\text{ cm}^{-1}$ ), so it could be used as an indicative peak of PLA. Intense peaks at 860 and  $961\text{ cm}^{-1}$  appeared in the spectrum of PBS but did not appear in that of PHBV, so PBS and PHBV could be discriminated with the two peaks effectively.

**Fig. 8** IR spectrum of PHBV**Fig. 9** Raman spectrum of PHBV

In conclusion, four kinds of biodegradable plastics: PLA, PBAT, PBS and PHBV were collected and examined for spectral characterization in this study. Peak positions were investigated. Main peaks in the spectra were also tentatively assigned to corresponding function groups. Peaks of the OH absorption at  $3,506\text{ cm}^{-1}$ , the shift of  $\text{C}=\text{O}$  stretching mode from  $1,730$  to  $1,754\text{ cm}^{-1}$  and the wagging absorption of  $\alpha\text{-CH}_3$  at  $757\text{ cm}^{-1}$ ; peaks at  $731$ ,  $1,121$  and  $1,169\text{ cm}^{-1}$ ; peaks at  $955$ ,  $806$  and  $654\text{ cm}^{-1}$  and peaks at  $1,000$ – $1,300\text{ cm}^{-1}$  in fingerprint bands and  $980\text{ cm}^{-1}$  were found to be indicative in the identification of PLA, PBAT, PBS and PHBV in IR spectrum separately. The characterization would be more accurate with the complementary information from IR and Raman spectroscopy and the method proved to be an effective, non-invasive and fast method to characterize and identify these four polymers.

**Acknowledgments** This study was financially supported by the State Key Laboratory of Environmental Chemistry and Ecotoxicology, Research Center for Eco-Environmental Sciences, Chinese Academy of Sciences (KF 2011-09).

## References

1. Lee Y, Chang JB, Kim HK, Park TG (2006) *Macromol Res* 14:359
2. Sato H, Murakami R, Zhang J, Ozaki Y (2006) *Macromol Res* 14:408
3. Anders S, Stolt M (2002) *Prog Polym Sci* 27:1123
4. Qi F, Hanna MA (1999) *Ind Crop Prod* 10:47
5. Bhatia A, Gupta RK, Bhattacharya SN, Choi HJ (2007) *Korea-Aust Rheol J* 19:125
6. Rantze E, Kleeberg I, Witt U, Müller RJ, Deckwer WD (1998) *Macromol Symp* 130:319
7. Witt U, Müller RJ, Deckwer WD (1996) *Macromol Chem Phys* 197:1525
8. Uwe W, Rolf-Joachim M, Wolf-Dieter D (1995) *J Polym Environ* 3:215
9. Witt U, Müller RJ, Deckwer WD (1997) *J Polym Environ* 5:81
10. Bhari K, Mitomo H, Enjoji T, Yoshii F, Makuuchi K (1998) *Polym Degrad Stab* 62:551
11. Doi Y, Kasuya K, Abe H, Koyama N, Ishiwatara S, Takagi K (1996) *Polym Degrad Stab* 51:281
12. Doi Y, Fukuda K (1994) *Studies in Polymer Science* 12. Elsevier, Amsterdam
13. Liu H (2008) Tailoring of poly (3-hydroxybutyrate-co-3-hydroxyvalerate) (PHBV) copolymers for bone tissue applications. *Howard Univ* 134:3335311

14. Gu SY, Zhang K, Ren J, Zhan H (2008) *Carbohydr Polym* 74:79
15. Jiang L, Wolcott MP, Zhang JW (2006) *Biomacromolecules* 7:199
16. Shi XQ, Ito H, Kikutani T (2006) *Polymer* 47:611
17. Chivrac F, Kadlecová Z, Pollet E, Avérous L (2006) *J Polym Environ* 14:393
18. Lee S, Lee Y, Lee JW (2007) *Macromol Res* 15:44
19. Feng JM (2009) *Application of infrared spectrometry in the trace evidence*. Chemistry industry Press, Beijing
20. Gelder JD, Willemse-Erix D, Scholtes MJ, Sanchez JI, Maquelin K, Vandenabeele P, Boever PD, Puppels GJ, Moens L, Vos PD (2008) *Anal Chem* 80:2155