

Reduced Nucleotide Diversity on a Plant Y Chromosome Following Recent Recombination Suppression

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ABSTRACT X and Y chromosomes differ in effective population size (N_e), rates of recombination, and exposure to natural selection, all of which can significantly affect levels of genetic variability within the genome. On Y chromosomes with suppressed recombination, both positive and negative selection can reduce the frequencies of genetically linked neutral variants, resulting in a reduction in N_e and the level of neutral polymorphism on Y compared to X chromosomes or autosomes. However, non-selective factors including biased sex ratios and high variance in male fitness can also reduce Y-linked N_e , making it difficult to infer the causes of low Y-chromosome diversity. Here, we investigate the factors affecting X- and Y-linked polymorphism during the early stages of plant sex chromosome evolution in *Rumex hastatulus* (Polygonaceae). Strikingly, our analysis revealed that nucleotide diversity on the Y is approximately 12.5 fold lower than predicted from models of neutral evolution. We demonstrate that the magnitude of this reduction is unlikely to be explained by the observed occurrence of female biased sex ratios in this species, or by reduced male N_e caused by high variance in male fitness. Rather, using forward simulations, we show that the diversity reduction on the Y is consistent with the predicted effects of background selection arising from suppressed recombination. Given the recent origin of *R. hastatulus* sex chromosomes, our results indicate that the low level of sequence diversity commonly observed on ancient Y chromosomes might evolve soon after sex chromosomes originate, with background selection playing an important role during the early stages of their evolution.

KEYWORDS Sex Chromosome Evolution; Nucleotide Diversity; Recombination; Deleterious Mutations

Introduction

Morphologically distinct sex chromosomes have evolved multiple times independently in both plant and animal kingdoms (Westergaard 1958; Ohno 1967; Bull 1983; Charlesworth 1991). Despite numerous biological differences between the kingdoms, X and Y chromosomes in both lineages have undergone similar genetic changes, suggesting that general evolutionary mechanisms might drive sex chromosome divergence (Charlesworth 1978, 1996b; Charlesworth and Charlesworth 2000). Investigating the shared features of plant and animal sex chromosomes, including the loss of recombination (Bergero and Charlesworth 2009), the degeneration of

Y chromosomes (Hough *et al.* 2014; Bergero *et al.* 2015), and the evolution of dosage compensation (Muyle *et al.* 2012; Papadopulos *et al.* 2015), thus provides an informative context for investigating the evolutionary forces driving sex chromosome evolution.

One fundamental difference between sex chromosomes and autosomes is their respective copy number in a population; whereas autosomes in diploid species are present in two copies in each sex, the X chromosome is present in two copies in females and only one copy in males. As a consequence, the effective population size (N_e) of the X chromosome is predicted to be 3/4 that of autosomes, whereas the N_e of the male-limited Y chromosome is expected to equal 1/4 that of autosomes (assuming an equal number of reproducing females and males). Such differences in N_e are predicted to directly affect relative levels of neutral polymorphism maintained on these chromosomes, which is proportional to the product of N_e and the neutral mutation rate, μ

(Kimura 1984). This variation in neutral polymorphism is in turn expected to have important consequences on patterns of DNA sequence evolution, including the effectiveness of both positive and negative selection (Charlesworth *et al.* 1987).

Several demographic factors are expected to modulate or accentuate differences in polymorphism levels maintained on sex chromosomes (Ellegren 2009). These include population subdivision and sex-biased dispersal, deviations from a 1:1 breeding sex ratio, and high variance in reproductive success (Caballero 1995; Charlesworth 2001; Laporte and Charlesworth 2002; Pool and Nielsen 2007). For example, studies comparing levels of variability on the X chromosome and autosomes in humans have found evidence that sex-biased dispersal has reduced levels of X-to-autosome diversity below the neutrally expected level of 0.75 (Keinan *et al.* 2009), though there is considerable variation in current estimates of the X/A ratio in humans, and other demographic processes have also been suggested (Hammer *et al.* 2010; Bustamante and Ramachandran 2009). Theoretical work also indicates that population bottlenecks can lead to disproportionate decreases in sex-linked compared to autosomal variation (Pool and Nielsen 2007), a prediction that has been supported in a diverse array of taxa, including chimpanzees and orangutans (Kaessmann *et al.* 2001; Fischer *et al.* 2006), the house mouse *Mus musculus* (Baines and Harr 2007), *Drosophila* (Andolfatto 2001), and humans (Keinan *et al.* 2009).

In addition to demographic factors, evolutionary theory predicts that genetic variability on sex chromosomes should also be affected by the evolution of suppressed X-Y recombination, which lowers the coalescent N_e for Y-linked genes and increases their vulnerability to the effects of selective sweeps (Maynard Smith and Haigh 1974; Aquadro *et al.* 1994) and background selection (Charlesworth 1996a, 1994). These processes can be viewed as examples of the Hill–Robertson (HR) effect, in which the effective population size for a given chromosomal region depends strongly on the rate of recombination, such that selection at a focal site is not independent of selection at nearby sites (Hill and Robertson 1966). Moreover, both the background selection and selective sweep models predict a reduction in N_e and, as a result, reduced levels of neutral polymorphism, selection efficacy, and rate of adaptation (Comeron *et al.* 2008). Although the HR effect can occur throughout the genome (McVean and Charlesworth 2000), large genomic regions in which recombination is suppressed, such as the Y chromosome, are expected to experience the most severe reductions in neutral diversity due to chromosome-wide linkage (Charlesworth 1996b; Charlesworth and Charlesworth 2000; Bachtrog 2013). Consistent with this, studies examining Y chromosome variability in mammals (Hellborg and Ellegren 2004; Wilson Sayres *et al.* 2014) and *Drosophila* (McAllister and Charlesworth 1999; Bachtrog and Charlesworth 2000) have found that Y-linked diversity is considerably lower than predicted under models of neutral evolution, suggesting that the HR effect plays a key role determining patterns of nucleotide diversity during sex chromosome evolution across a broad range of animal taxa.

Despite widespread interest in determining the effects of recombination on patterns of genetic diversity and the forces contributing to sex chromosome divergence (Ellegren 2011; Otto *et al.* 2011; Bachtrog 2013), we still know little about the influence of the HR effect on patterns nucleotide polymorphism in young sex chromosome systems, where *de novo* recombination suppression has evolved relatively recently (Charlesworth 2016). The timescales over which selection affects neutral polymor-

phism on sex chromosomes is therefore not well understood. Recent studies of young sex chromosome system in plants have revealed that several canonical features of X and Y chromosome evolution are shared between plants and animals, including the genetic deterioration of the Y chromosome (Bergero *et al.* 2015; Hough *et al.* 2014), the occurrence of ‘evolutionary strata’ (Bergero and Charlesworth 2009), and the evolution of dosage compensation (Muyle *et al.* 2012; Papadopoulos *et al.* 2015). Nonetheless, except for a small number of genes in *Silene latifolia* (Filatov *et al.* 2001; Qiu *et al.* 2010), the associated changes in genetic diversity that are predicted from evolutionary models of X- and Y-chromosome evolution have not been widely studied in young plant sex chromosomes. Given that both background selection and selective sweeps are predicted to have the strongest effects during the earliest stages of Y-chromosome evolution, before Y-chromosomes have lost a large proportion of their genes (Bachtrog 2008), studying recently evolved plant sex chromosomes provides an important test of the temporal dynamics of the HR effect.

To investigate the factors affecting nucleotide diversity in the early stages of sex chromosome evolution, we analyzed single nucleotide polymorphisms (SNPs) on sex chromosomes and autosomes in the plant *R. hastatulus* (Polygonaceae). This species is a dioecious annual with highly heteromorphic X and Y chromosomes that are estimated to have evolved approximately 15 MYA (Quesada del Bosque *et al.* 2011; Grabowska-Joachimciak *et al.* 2015; Navajas-Pérez *et al.* 2005), making sex chromosomes in this species over 100 million years younger than those in mammals (Lahn and Page 1999; Ross *et al.* 2005). *Rumex hastatulus* has received particular attention because of the unique occurrence of an intraspecific polymorphism for sex chromosome system, in which both XY and XY₁XY₂ males occur in geographically distinct chromosomal races (Smith 1963). The XY₁XY₂ sex chromosome system (the North Carolina race) is thought to have originated through an X-autosome fusion, with the XY system (the Texas race) maintaining the ancestral chromosome complement (Smith 1964). Despite the recent origin of the sex chromosome systems in this species, there is evidence that both the ancestral and neo-Y chromosomes have undergone significant genetic degeneration, though less degeneration was inferred for younger sex-linked genes unique to the North Carolina race (Hough *et al.* 2014).

Of particular relevance to the present study is that *R. hastatulus* populations throughout their geographical range (southern USA) consistently exhibit female-biased sex ratios, with a mean sex ratio of $N_m/N_f = 0.6$ (Pickup and Barrett 2013). This is important because female-biased sex ratios are predicted to affect levels of sex chromosome diversity (Ellegren 2009), though few studies have attempted to empirically distinguish between the effects of biased sex ratios and selective interference. Here, focusing on the Y₁ chromosome that is shared between the chromosomal races of *R. hastatulus*, we tested whether levels of nucleotide diversity on X, Y, and autosomal chromosomes were jointly consistent with predictions from models of sex ratio bias and variance in reproductive success. In addition, we use population genetic parameters estimated from our data to conduct forward-time simulations of both purifying and positive selection to determine the relative support for models of background selection and selective sweeps in explaining the levels of diversity we observed.

Materials and Methods

Population Samples and Sex-Linked Genes

We analyzed sex-linked and autosomal genes identified from Illumina RNA sequence data from 24 individuals (12 males and 12 females, with 1 male and 1 female from each of 12 populations, 6 populations of each sex chromosome race). Samples of both sex chromosomal races were collected in 2010 from throughout the native range of *R. hastatulus* (locations in Table S1), and plants were grown in the glasshouse from seeds collected from open-pollinated females. We extracted RNA from leaf tissue using Spectrum Plant Total RNA kits (Sigma-Aldrich). The isolation of mRNA and cDNA synthesis was conducted according to standard Illumina RNAseq procedures, with sequencing conducted on two Illumina HiSeq lanes with 150-bp end reads at the Genome Quebec Innovation Center. Reads from these 24 samples were mapped to the *R. hastatulus* reference transcriptome (Hough *et al.* 2014), and raw sequences are available from the Gen-Bank Short Read Archive under accession no. SRP041588. Reads were mapped using the Burrows–Wheeler Aligner (release 0.6.2-r126 (Li and Durbin 2010)), followed by Stampy (release 1.0.20; (Lunter and Goodson 2011)). We used Picard tools (release 1.78, <http://picard.sourceforge.net>) to modify mapping output into the format required for the Genome Analysis Toolkit (GATK version 2.1-11; (McKenna *et al.* 2010)) variant calling software, and subsequently removed genes with low coverage (<10x) and Phred Quality Scores (<20). The population samples analyzed here were previously reported in Hough *et al.* (2014), where they were used to validate the ascertainment of sex-linked contigs identified through segregation analysis. From the list of sex-linked genes identified in Hough *et al.* (2014) that were shared between the Texas (XY) and North Carolina (XY1Y2) sex chromosome races, we identified a total of 460 sex-linked genes for analysis.

Autosomal Genes

In evaluating evidence for nucleotide diversity differences between X and Y chromosomes, it is important to distinguish between reduced Y-linked diversity, and the possibility that X-linked diversity is elevated above the level predicted from a neutral model. To do this, we normalized sex-linked diversity estimates by autosomal diversity, and compared empirical X/A and Y/A nucleotide diversity ratios to those predicted from a neutral model (described below). Because the criteria for identifying autosomal loci in Hough *et al.* (2014) were based on the occurrence of four segregating SNPs per locus, and this set is therefore probably higher in diversity than the average autosome, here we incorporated a broader set of all non-sex linked (punitively autosomal) genes from our transcriptome data. We filtered this set of non-sex linked genes to remove those that may have been sex-linked but were not identified as such by the ascertainment criteria in Hough *et al.* (2014). In particular, we removed: (i) any genes in which there was evidence for at least one SNP with a sex-linked segregation pattern, (ii) any genes with SNPs showing fixed heterozygosity in males and fixed homozygosity in females, (iii) any genes with less than 10X coverage or greater than 100X coverage from independently obtained genomic coverage data (to filter out duplicate genes or those with highly repetitive sequences), and (iv) any genes containing SNPs with large (>0.4) allele frequency differences between males and females. Finally, we removed genes with fewer than 100 synonymous sites following filtering to avoid biasing results toward genes that may have been particularly

short due to assembly problems. This filtering resulted in a set of 12,356 and 11,350 autosomal genes in the Texas and North Carolina races, respectively.

Phasing X and Y alleles

To estimate polymorphism for the X and Y sequences separately, it is necessary to infer the phase of SNPs in sex-linked transcripts in males. In previous work, phasing alleles on *R. hastatulus* sex chromosomes was achieved using segregation analysis from a genetic cross. Here, to phase SNPs from population samples where such segregation data was unavailable, we used HAPCUT (Bansal and Bafna 2008), a maximum-cut based algorithm that reconstructs haplotypes using sequenced fragments (Illumina read data) from the two homologous chromosomes in a diploid individual to output a list of phased haplotype blocks containing the SNP variants on each chromosome. Because the resulting haplotype blocks produced by HAPCUT contained SNPs that were phased relative to each other, but not designated to either the X or Y chromosome, we assigned individual variants to X or Y by independently identifying fixed X-Y differences with each haplotype block (i.e., sites in which all females were homozygous, and all males were heterozygous). Identifying such fixed differences within phased haplotype blocks enabled us to then infer the correct phase (X or Y) of the polymorphisms from HAPCUT's output. This was done by matching the phase of fixed X-Y differences with their neighboring polymorphic sites - i.e., when a fixed X-Y difference occurred in the same phased haplotype block as a polymorphic site, the polymorphic variants in that block were assigned to either X or Y based on the known phase of the fixed difference with which they were matched. SNPs that were identified outside of phased blocks, or in blocks without fixed X-Y differences, were recorded as missing data. Finally, we filtered SNPs for coverage > 60, QUAL score > 60, and those within a distance of 10bp or less from indels. This procedure was conducted using a combination of Perl and Bash and this resulted in fasta-formatted alignments of X and Y sequences for 372 sex-linked genes from the 24 individuals in our study.

We further validated the results of HAPCUT's allele phasing by comparing the accuracy of this method with the phasing-by-segregation method that was conducted using data from a genetic cross in Hough *et al.* (2014). To do this, we first phased the sequence data from parents and their progeny using HAPCUT's algorithm (using the same parameters as for the population data). In particular, we looked for cases where SNPs were inferred on the Y chromosome by HAPCUT, but where the 'true' level polymorphism was zero, since Y-linked sequences from a single family should all be identical, as we had confirmed through phasing by segregation. We identified 7 percent of sex-linked genes that had phasing errors of this kind, or that we determined to have genotyping errors resulting in a false SNP calls on the Y chromosome, leading to a SNP error rate estimate of 1.7×10^{-4} . This rate is very low relative to the population-based estimates of polymorphism on the X and autosomes (Table 1), and therefore should have minimal effects on our estimation of X/A polymorphism. However, because this rate is high relative to the expected level of true polymorphism on the Y-chromosome, we further filtered genes in which we found evidence for false-positive Y-polymorphisms arising from: (i) phasing errors caused by gene duplicates (more than two haplotypes), (ii) polymorphisms around indels, and (iii) genotyping errors caused by low Y-expression. This filtering was done by manually inspecting sequences in IGV (Robinson *et al.* 2011) and

identifying each individual putative polymorphism on the Y chromosome.

Estimating neutral polymorphism on sex chromosomes and autosomes

For each locus in our analysis, we calculated Watterson's (1975) estimator of the population parameter $\theta = 4N_e\mu$, where N_e is the effective population size, and μ is the mutation rate (Watterson 1975), using a modified version of the Perl program Polymorphura (Bachtrog and Andolfatto 2006). To compare sex-linked and autosomal loci, we calculated the average value of θ , weighted by the number of synonymous sites in each gene (Figure 2; Table 1). We obtained 95 percent confidence intervals for our estimates of the X/A and Y/A diversity ratios by bootstrapping by gene using the BCa method (Efron and Tibshirani 1994) implemented in the Boot package in R (Canty and Ripley 2012), and calculating X/A and Y/A diversity on each iteration for 20000 replicates each. Bootstrapping was conducted on the final filtered set of 173 sex-linked, and 12355 autosomal genes from the Texas race, and separately for the 176 sex-linked and 11349 autosomal genes from the North Carolina race. Note that the lack of recombination on the Y chromosome implies that assumptions of independence are likely violated, since Y-linked loci are expected to be non-independent, and therefore the true uncertainty in our Y/A diversity estimate obtained by this method is likely an underestimate. To address this issue, we used a maximum likelihood approach implemented in the software (Wright and Charlesworth 2004) to obtain independent estimates of the Y/A ratio. Here, [explain more]. Finally, we also tested whether estimates of diversity on the X chromosome calculated from females were consistent with estimates from phased X sequences from males. As no significant difference was observed, we report only results from females.

Neutral predictions and the effect of sex ratio bias on diversity

Wright's (1931) formula $N_e = 4N_fN_m/(N_f + N_m)$ gives the effective population size of a randomly mating population with separate sexes and a Poisson distribution of offspring number

$$S_n = \frac{X_1 + X_2 + \dots + X_n}{n} = \frac{1}{n} \sum_{i=1}^n X_i \quad (1)$$

Simulations of Background Selection and Selective Sweeps

Results and Discussion

Evolutionary relationships of sex chromosome races in *R. hastulatus*

Y-chromosome diversity is significantly lower than predicted by neutrality

Male-to-female sex ratio of N_e

Background Selection and Selective Sweeps

Conclusions

Acknowledgments

Literature Cited

Andolfatto, P., 2001 Contrasting patterns of x-linked and autosomal nucleotide variation in drosophila melanogaster and drosophila simulans. *Molecular Biology and Evolution* **18**: 279–290.

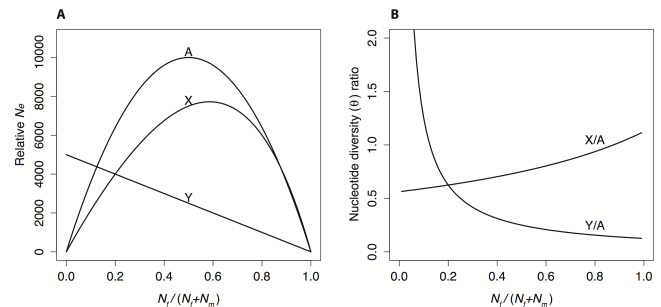


Figure 1 Example figure from 10.1534/genetics.114.173807. Please include your figures in the manuscript for the review process. You can upload figures to Overleaf via the Project menu. Upon acceptance, we'll ask for your figure files to be uploaded in any of the following formats: TIFF (.tiff), JPEG (.jpg), Microsoft PowerPoint (.ppt), EPS (.eps), or Adobe Illustrator (.ai). Images should be a minimum of 300 dpi in resolution and 500 dpi minimum if line art images. RGB, CMYK, and Grayscale are all acceptable. Halftones should be high contrast with sharp detail, because some loss of detail and contrast is inevitable in the production process. Figures should be 10-20 cm in width and 1-25 cm in height. Graph axes must be exactly perpendicular and all lines of equal density. Label multiple figure parts with A, B, etc. in bolded type, and use Arrows and numbers to draw attention to areas you want to highlight. Legends should start with a brief title and should be a self-contained description of the content of the figure that provides enough detail to fully understand the data presented. All conventional symbols used to indicate figure data points are available for typesetting; unconventional symbols should not be used. Italicize all mathematical variables (both in the figure legend and figure), genotypes, and additional symbols that are normally italicized.

Table 1 Students and their grades

Student	Grade ^a	Rank	Notes
Alice	82%	1	Performed very well.
Bob	65%	3	Not up to his usual standard.
Charlie	73%	2	A good attempt.

^a This is an example of a footnote in a table. Lowercase, superscript italic letters (a, b, c, etc.) are used by default. You can also use *, **, and *** to indicate conventional levels of statistical significance, explained below the table.

- Aquadro, C. F., D. J. Begun, and E. C. Kindahl, 1994 Selection, recombination, and dna polymorphism in drosophila. In *Non-neutral evolution*, pp. 46–56, Springer.
- Bachtrog, D., 2008 The temporal dynamics of processes underlying y chromosome degeneration. *Genetics* **179**: 1513–1525.
- Bachtrog, D., 2013 Y-chromosome evolution: emerging insights into processes of y-chromosome degeneration. *Nature Reviews Genetics* **14**: 113–124.
- Bachtrog, D. and P. Andolfatto, 2006 Selection, recombination and demographic history in drosophila miranda. *Genetics* **174**: 2045–2059.
- Bachtrog, D. and B. Charlesworth, 2000 Reduced levels of microsatellite variability on the neo-y chromosome of drosophila miranda. *Current Biology* **10**: 1025–1031.
- Baines, J. F. and B. Harr, 2007 Reduced x-linked diversity in derived populations of house mice. *Genetics* **175**: 1911–1921.
- Bansal, V. and V. Bafna, 2008 Hapcut: an efficient and accurate algorithm for the haplotype assembly problem. *Bioinformatics* **24**: i153–i159.
- Bergero, R. and D. Charlesworth, 2009 The evolution of restricted recombination in sex chromosomes. *Trends in Ecology & Evolution* **24**: 94–102.
- Bergero, R., S. Qiu, and D. Charlesworth, 2015 Gene loss from a plant sex chromosome system. *Current Biology* **25**: 1234–1240.
- Bull, J., 1983 *Evolution of sex determining mechanisms..* The Benjamin/Cummings Publishing Company, Inc.
- Bustamante, C. D. and S. Ramachandran, 2009 Evaluating signatures of sex-specific processes in the human genome. *Nature genetics* **41**: 8–10.
- Caballero, A., 1995 On the effective size of populations with separate sexes, with particular reference to sex-linked genes. *Genetics* **139**: 1007–1011.
- Canty, A. and B. Ripley, 2012 boot: Bootstrap r (s-plus) functions. R package version 1.
- Charlesworth, B., 1978 Model for evolution of y chromosomes and dosage compensation. *Proceedings of the National Academy of Sciences* **75**: 5618–5622.
- Charlesworth, B., 1991 The evolution of sex chromosomes. *Science* **251**: 1030–1033.
- Charlesworth, B., 1994 The effect of background selection against deleterious mutations on weakly selected, linked variants. *Genetical research* **63**: 213–227.
- Charlesworth, B., 1996a Background selection and patterns of genetic diversity in drosophila melanogaster. *Genetical research* **68**: 131–149.
- Charlesworth, B., 1996b The evolution of chromosomal sex determination and dosage compensation. *Current Biology* **6**: 149–162.
- Charlesworth, B., 2001 The effect of life-history and mode of inheritance on neutral genetic variability. *Genetical research* **77**: 153–166.
- Charlesworth, B. and D. Charlesworth, 2000 The degeneration of y chromosomes. *Philosophical Transactions of the Royal Society of London B: Biological Sciences* **355**: 1563–1572.
- Charlesworth, B., J. Coyne, and N. Barton, 1987 The relative rates of evolution of sex chromosomes and autosomes. *American Naturalist* pp. 113–146.
- Charlesworth, D., 2016 Plant sex chromosomes. *Annual review of plant biology* **67**: 397–420.
- Comeron, J. M., A. Williford, and R. Kliman, 2008 The hill-robertson effect: evolutionary consequences of weak selection and linkage in finite populations. *Heredity* **100**: 19–31.
- Efron, B. and R. J. Tibshirani, 1994 *An introduction to the bootstrap*. CRC press.
- Ellegren, H., 2009 The different levels of genetic diversity in sex chromosomes and autosomes. *Trends in Genetics* **25**: 278–284.
- Ellegren, H., 2011 Sex-chromosome evolution: recent progress and the influence of male and female heterogamety. *Nature Reviews Genetics* **12**: 157–166.
- Filatov, D. A., V. Laporte, C. Vitte, and D. Charlesworth, 2001 Dna diversity in sex-linked and autosomal genes of the plant species silene latifolia and silene dioica. *Molecular Biology and Evolution* **18**: 1442–1454.
- Fischer, A., J. Pollack, O. Thalmann, B. Nickel, and S. Pääbo, 2006 Demographic history and genetic differentiation in apes. *Current Biology* **16**: 1133–1138.
- Grabowska-Joachim, A., A. Kula, T. Książczyk, J. Chojnicka, E. Sliwinska, and A. J. Joachim, 2015 Chromosome landmarks and autosome-sex chromosome translocations in rumex hastatulus, a plant with xx/xy1y2 sex chromosome system. *Chromosome Research* **23**: 187–197.
- Hammer, M. F., A. E. Woerner, F. L. Mendez, J. C. Watkins, M. P. Cox, and J. D. Wall, 2010 The ratio of human x chromosome to autosome diversity is positively correlated with genetic distance from genes. *Nature genetics* **42**: 830–831.
- Hellborg, L. and H. Ellegren, 2004 Low levels of nucleotide diversity in mammalian y chromosomes. *Molecular Biology and Evolution* **21**: 158–163.
- Hill, W. G. and A. Robertson, 1966 The effect of linkage on limits to artificial selection. *Genetical research* **8**: 269–294.
- Hough, J., J. D. Hollister, W. Wang, S. C. Barrett, and S. I. Wright, 2014 Genetic degeneration of old and young y chromosomes in the flowering plant rumex hastatulus. *Proceedings of the National Academy of Sciences* **111**: 7713–7718.
- Kaessmann, H., V. Wiebe, G. Weiss, and S. Pääbo, 2001 Great ape dna sequences reveal a reduced diversity and an expansion in humans. *Nature genetics* **27**: 155–156.
- Keinan, A., J. C. Mullikin, N. Patterson, and D. Reich, 2009 Accelerated genetic drift on chromosome x during the human dispersal out of africa. *Nature genetics* **41**: 66–70.
- Kimura, M., 1984 *The neutral theory of molecular evolution*. Cambridge University Press.

- Lahn, B. T. and D. C. Page, 1999 Four evolutionary strata on the human x chromosome. *Science* **286**: 964–967.
- Laporte, V. and B. Charlesworth, 2002 Effective population size and population subdivision in demographically structured populations. *Genetics* **162**: 501–519.
- Li, H. and R. Durbin, 2010 Fast and accurate long-read alignment with burrows–wheeler transform. *Bioinformatics* **26**: 589–595.
- Lunter, G. and M. Goodson, 2011 Stampy: a statistical algorithm for sensitive and fast mapping of illumina sequence reads. *Genome research* **21**: 936–939.
- Maynard Smith, J. and J. Haigh, 1974 The hitch-hiking effect of a favourable gene. *Genetical research* **23**: 23–35.
- McAllister, B. F. and B. Charlesworth, 1999 Reduced sequence variability on the neoy chromosome of *drosophila americana americana*. *Genetics* **153**: 221–233.
- McKenna, A., M. Hanna, E. Banks, A. Sivachenko, K. Cibulskis, A. Kernytsky, K. Garimella, D. Altshuler, S. Gabriel, M. Daly, *et al.*, 2010 The genome analysis toolkit: a mapreduce framework for analyzing next-generation dna sequencing data. *Genome research* **20**: 1297–1303.
- McVean, G. A. and B. Charlesworth, 2000 The effects of hill-robertson interference between weakly selected mutations on patterns of molecular evolution and variation. *Genetics* **155**: 929–944.
- Muyle, A., N. Zemp, C. Deschamps, S. Mousset, A. Widmer, and G. A. Marais, 2012 Rapid de novo evolution of x chromosome dosage compensation in *silene latifolia*, a plant with young sex chromosomes. *PLoS Biol* **10**: e1001308.
- Navajas-Pérez, R., R. de la Herrán, G. L. González, M. Jamilena, R. Lozano, C. R. Rejón, M. R. Rejón, and M. A. Garrido-Ramos, 2005 The evolution of reproductive systems and sex-determining mechanisms within *rumex* (polygonaceae) inferred from nuclear and chloroplastidial sequence data. *Molecular Biology and Evolution* **22**: 1929–1939.
- Ohno, S., 1967 *Sex chromosomes and sex-linked genes*, volume 1. Springer-Verlag.
- Otto, S. P., J. R. Parnell, C. L. Peichel, T.-L. Ashman, D. Charlesworth, A. K. Chippindale, L. F. Delph, R. F. Guerrero, S. V. Scarpino, and B. F. McAllister, 2011 About par: the distinct evolutionary dynamics of the pseudoautosomal region. *Trends in Genetics* **27**: 358–367.
- Papadopoulos, A. S., M. Chester, K. Ridout, and D. A. Filatov, 2015 Rapid y degeneration and dosage compensation in plant sex chromosomes. *Proceedings of the National Academy of Sciences* **112**: 13021–13026.
- Pickup, M. and S. C. Barrett, 2013 The influence of demography and local mating environment on sex ratios in a wind-pollinated dioecious plant. *Ecology and evolution* **3**: 629–639.
- Pool, J. E. and R. Nielsen, 2007 Population size changes reshape genomic patterns of diversity. *Evolution* **61**: 3001–3006.
- Qiu, S., R. Bergero, A. Forrest, V. B. Kaiser, and D. Charlesworth, 2010 Nucleotide diversity in *silene latifolia* autosomal and sex-linked genes. *Proceedings of the Royal Society of London B: Biological Sciences* p. rspb20100606.
- Quesada del Bosque, M., R. Navajas-Pérez, J. Panero, A. Fernández-González, and M. Garrido-Ramos, 2011 A satellite dna evolutionary analysis in the north american endemic dioecious plant *rumex hastatulus* (polygonaceae). *Genome* **54**: 253–260.
- Robinson, J. T., H. Thorvaldsdóttir, W. Winckler, M. Guttman, E. S. Lander, G. Getz, and J. P. Mesirov, 2011 Integrative genomics viewer. *Nature biotechnology* **29**: 24–26.
- Ross, M. T., D. V. Grafham, A. J. Coffey, S. Scherer, K. McLay, D. Muzny, M. Platzer, G. R. Howell, C. Burrows, C. P. Bird, *et al.*, 2005 The dna sequence of the human x chromosome. *Nature* **434**: 325–337.
- Smith, B. W., 1963 The mechanism of sex determination in *rumex hastatulus*. *Genetics* **48**: 1265.
- Smith, B. W., 1964 The evolving karyotype of *rumex hastatulus*. *Evolution* pp. 93–104.
- Watterson, G., 1975 On the number of segregating sites in genetical models without recombination. *Theoretical population biology* **7**: 256–276.
- Westergaard, M., 1958 The mechanism of sex determination in dioecious flowering plants. *Advances in genetics* **9**: 217–281.
- Wilson Sayres, M. A., K. E. Lohmueller, and R. Nielsen, 2014 Natural selection reduced diversity on human y chromosomes. *PLoS Genet* **10**: e1004064.
- Wright, S. I. and B. Charlesworth, 2004 The hka test revisited. *Genetics* **168**: 1071–1076.