

# Canonical Correlation Analysis III

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# Canonical Coefficients

- ❖ Two sets of canonical coefficients (weights)
  - One set to combine the  $X$ s
  - The other set to combine the  $Y$ s
  - Interpreted similarly to regression coefficients

$$F_1 = E1\$Eigenvector[1,1] * X_1 + E1\$Eigenvector[1,1] * X_2$$

$$F_2 = E1\$Eigenvector[1,2] * X_1 + E1\$Eigenvector[2,2] * X_2$$

$$G_1 = E2\$Eigenvector[1,1] * Y_1 + E2\$Eigenvector[1,1] * Y_2$$

$$G_2 = E2\$Eigenvector[1,2] * Y_1 + E2\$Eigenvector[2,2] * Y_2$$

# Canonical Coefficients

- ❖ Two sets of canonical coefficients (weights)
  - One set to combine the Xs
  - The other set to combine the Ys
  - Interpreted similarly to regression coefficients
- ❖ Test statistical significance of canonical correlations
  - The maximum number of canonical correlations equals to the number of variables in the smaller set
  - Not all will be statistically significant
  - Not all statistical significant CCs will be meaningful
  - This can be done using **CCP** package in R by **p.asym()**, with options for test statistics, commonly use “Wilks” stand for Wilk’s Lambda to be approximated by Chi-square distribution – Bartlett’s Chi Square test

# Example: head lengths of 1<sup>st</sup> and 2<sup>nd</sup> sons (IAMA 3.13.1)

```
> headsize
```

	head1	breadth1	head2	breadth2
[1,]	191	155	179	145
[2,]	195	149	201	152
[3,]	181	148	185	149
[4,]	183	153	188	149
[5,]	176	144	171	142
[6,]	208	157	192	152
[7,]	189	150	190	149
[8,]	197	159	189	152
[9,]	188	152	197	159
[10,]	192	150	187	151
[11,]	179	158	186	148
[12,]	183	147	174	147
[13,]	174	150	185	152
[14,]	190	159	195	157
[15,]	188	151	187	158
[16,]	163	137	161	130
[17,]	195	155	183	158
[18,]	186	153	173	148
[19,]	181	145	182	146
[20,]	175	140	165	137
[21,]	192	154	185	152
[22,]	174	143	178	147
[23,]	176	139	176	143
[24,]	197	167	200	158
[25,]	190	163	187	150

```
headsize.std <- sweep(headsize, 2, apply(headsize, 2, sd), FUN="/")
R <- cor(headsize.std)
R11 <- R[1:2,1:2]
R12 <- R[3:4,1:2]
R21 <- R[1:2,3:4]
R22 <- R[3:4,3:4]
R11.inv <- solve(R11)
R22.inv <- solve(R22)
```

```
# compute E1 and E2
```

```
E1 <- R11.inv %*% R12 %*% R22.inv %*% R21
E2 <- R22.inv %*% R21 %*% R11.inv %*% R12
```

```
# compute eigenvalues and eigenvectors of E1 and E2:
```

```
eigen(E1)
```

```
eigen(E2)
```

$$E_1 = R_{11}^{-1} R_{12} R_{22}^{-1} R_{21}, \quad E_2 = R_{22}^{-1} R_{21} R_{11}^{-1} R_{12}$$

$E_1$  is p-by-p;  $E_2$  is q-by-q.

p is the dimension of the predictors set

q is the dimension of the predictants set.

```
> eigen(E1)
```

```
$values
```

```
[1] 0.621781555 0.002887785
```

```
$vectors [,1] [,2]
```

```
[1,] -0.6947269 -0.7089828
```

```
[2,] -0.7192736 0.7052258
```

```
> eigen(E2)
```

```
$values
```

```
[1] 0.621781555 0.002887785
```

```
$vectors [,1] [,2]
```

```
[1,] 0.7424369 -0.7039264
```

```
[2,] 0.6699160 0.7102729
```

The canonical correlations express the association between the x and y variables after removal of the within-set correlation.

$$F_1 = -0.69 \cdot \text{head1} - 0.72 \cdot \text{breadth1};$$

$$F_2 = -0.70 \cdot \text{head1} + 0.71 \cdot \text{breadth1};$$

$$G_1 = +0.74 \cdot \text{head2} + 0.70 \cdot \text{breadth2};$$

$$G_2 = -0.70 \cdot \text{head2} + 0.71 \cdot \text{breadth2}.$$

# Since we work with standardized data from the beginning , so  
# no correction of scaling of CC vectors is needed:

# OPTIONAL:

# compute the canonical correlation vectors:

```
(a1 <- eigen(E1)$vectors[,1])
```

```
(a2 <- eigen(E1)$vectors[,2])
```

```
(b1 <- eigen(E2)$vectors[,1])
```

```
(b2 <- eigen(E2)$vectors[,2])
```

# correct the scaling of the canonical correlation vectors:

```
(a1 <- -1 * a1 / sqrt(t(a1) %*% R11 %*% a1))
```

```
(a2 <- -1 * a2 / sqrt(t(a2) %*% R11 %*% a2))
```

```
(b1 <- b1 / sqrt(t(b1) %*% R22 %*% b1))
```

```
(b2 <- -1 * b2 / sqrt(t(b2) %*% R22 %*% b2))
```

# check scaling:

```
t(a1) %*% R11 %*% a1
```

```
t(a2) %*% R11 %*% a2
```

```
t(b1) %*% R22 %*% b1
```

```
t(b2) %*% R22 %*% b2
```

```
# compute canonical correlation variables:
```

```
F1 <- headsize.std[,1:2] %%% a1    # size first son  
F2 <- headsize.std[,1:2] %%% a2    # shape first son  
G1 <- headsize.std[,3:4] %%% b1    # size second son  
G2 <- headsize.std[,3:4] %%% b2    # shape second son
```

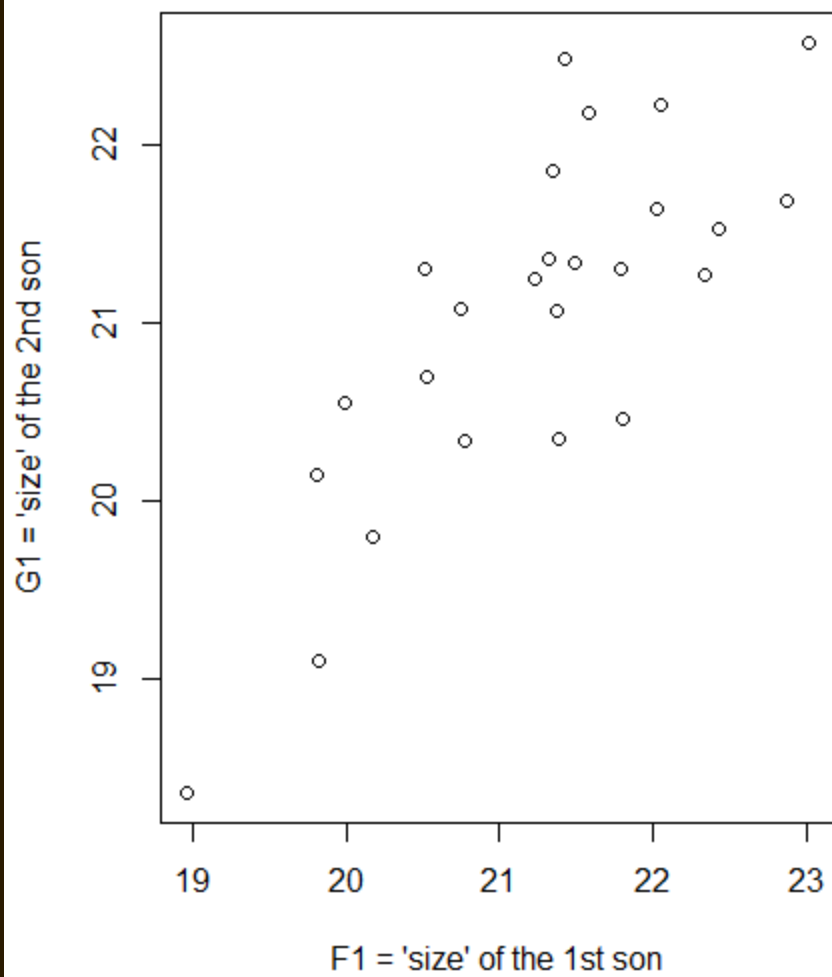
```
# check covariance matrix:
```

```
round(var(cbind(F1,F2,G1,G2)),3)
```

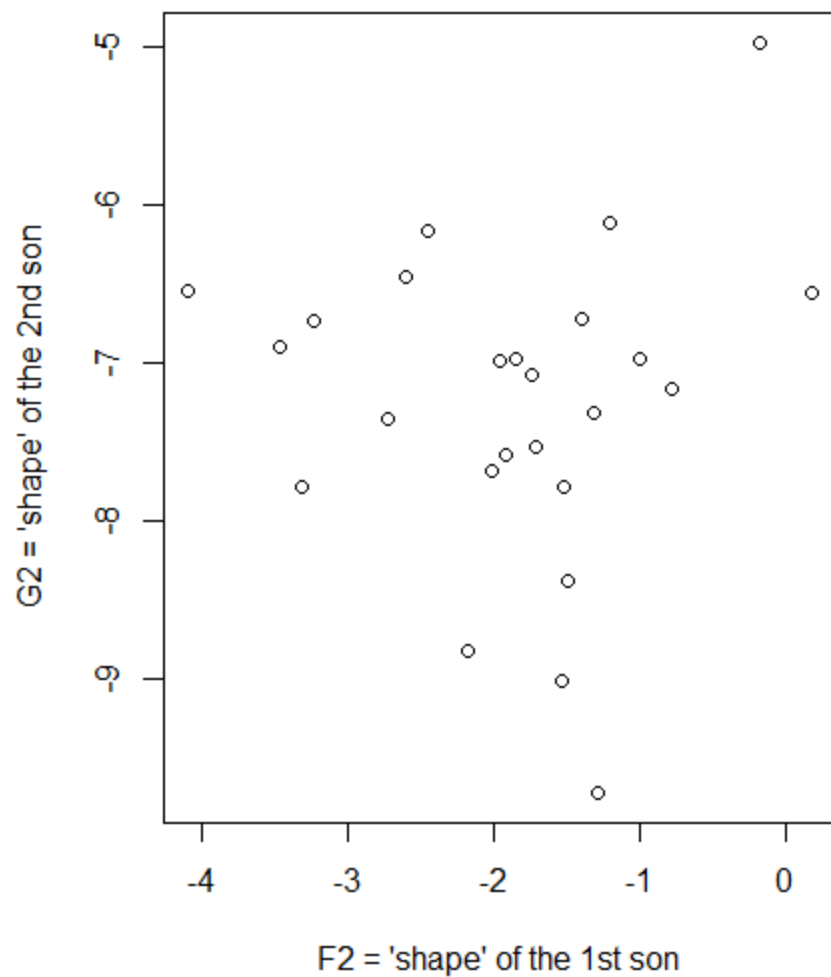
```
# plot canonical correlation variables:
```

```
par(mfrow=c(1,2))  
plot(F1,G1,main="Head size, correlation=0.788", xlab="F1 =  
'size' of the 1st son", ylab="G1 = 'size' of the 2nd son")  
plot(F2,G2, main="Head shape, correlation=.053", xlab="u2 =  
'shape' of the 1st son", ylab="G2 = 'shape' of the 2nd son")
```

Head size, correlation=0.788



Head shape, correlation=.053





# Loadings (structure coefficients)

- Questions to assist your interpretations:
  1. How well do the variates on either side relate to their own original variables?
  2. What is the associations between a variable and its respective canonical variate? – correlations
- Recall that: the pair of canonical variates with best correlation might not exactly interpretable
- Coefficients are for the computation of the variates, loadings are more for expressing the relationships of the variables to the construct (canonical variates)
  - Canonical communality coefficient
  - Canonical variate adequacy coefficient

# Coefficients

## ☐ Canonical communality coefficient

- Sum of the squared structure coefficients (loadings) across selected\* canonical variates for a given variable
- Measures how much variance of that original variable is explained or reproduced by the canonical variates
- \* If looking at all variates, it shall equal one, typically we often want to check those retained for interpretation or prediction.

## ☐ Canonical variate adequacy coefficient

- Average of all the squared structure coefficients (loadings) for one set of variables with respect to their canonical variate
- Measures how well a resulted canonical variate represents the variance in that set of original variables.

# Redundancy

- Questions to assist checking redundancy:
  1. How strongly do the original variables in one set relate to the canonical variates on the other side?
  2. How much of the average proportion of variance of the original variables in one set may be predicted from the variables in the other set – High redundancy suggests potentially high ability to predict
    - Product of the mean squared structure coefficient, i.e., the canonical adequacy coefficient for a canonical variate times the squared canonical correlation coefficient.
    - Canonical correlation reflects the % of variance in the predictant canonical variate explained by the predictor canonical variate
- Redundancy has to do with assessing the effectiveness of the canonical analysis in capturing the variance of the original variables

# CCA in R

**cancor()**

`data(LifeCycleSavings)`: savings ratio from 1960-1970

sr	numeric	aggregate personal savings
pop15	numeric	% of population under 15
pop75	numeric	% of population over 75
dpi	numeric	real per-capita disposable income
ddpi	numeric	% growth rate of dpi

```
pop = LifeCycleSavings[, 2:3] # select pop15 and pop75
oec = LifeCycleSavings[, -(2:3)] # select all the others

cancor(pop, oec)
```

# CCA in R

```
cancor(pop, oec)
$cor
[1] 0.8247966 0.3652762
```

**cancor()**

```
$xcoef
[,1]      [,2]
pop15 -0.009110856 -0.03622206
pop75  0.048647514 -0.26031158
```

```
$ycoef
[,1]      [,2]      [,3]
sr  0.0084710221  3.337936e-02 -5.157130e-03
dpi  0.0001307398 -7.588232e-05  4.543705e-06
ddpi 0.0041706000 -1.226790e-02  5.188324e-02
```

```
$xcenter 0.08(sr)+0.0001(dpi)+0.004(ddpi) = -0.009(pop15)+0.048(pop75)
pop15    pop75
35.0896  2.2930
```

**1<sup>st</sup> Canonical variate function**

```
$ycenter
sr      dpi      ddpi
9.6710 1106.7584  3.7576
```

# CCA in R

**matcor()&cc() in package(CCA)**

package(CCA) provides extension to the cancor()

- cancor() has limited outputs, package(CCA) provides a complete list of outputs in terms of both numerical and graphical
- package(CCA) enhances handling of missing values
- package (CCA) can perform regularized CCA when  $n < \max(p, q)$ .

data(*nutrimouse*): from the Toxicology & Pharmacology Laboratory

A list containing the following components:

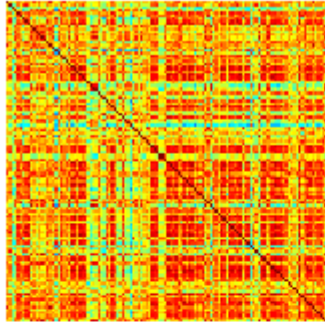
- gene: data frame (40 \* 120) with numerical variables
- lipid: data frame (40 \* 21) with numerical variables
- diet: factor vector (40)
- genotype: factor vector (40)

# CCA in R

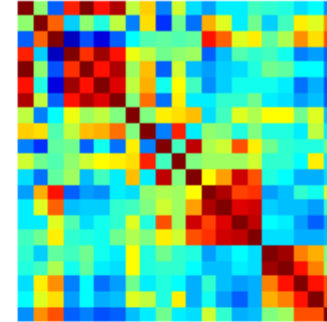
**matcor()&cc()**

```
X = as.matrix(nutrimouse$gene)
Y = as.matrix(nutrimouse$lipid)
correl = matcor(X, Y)
img.matcor(correl, type = 2)
```

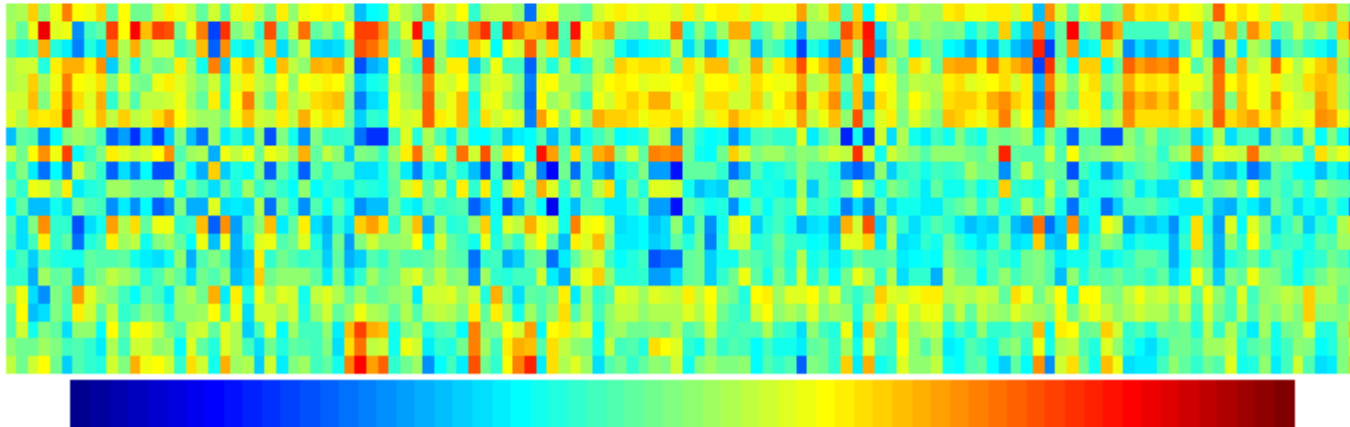
X correlation



Y correlation



Cross-correlation



Correlation matrices for: X variables (upper-left), Y variables (upper-right), cross-correlation  $X \times Y$  (bottom). Increasing values are translated into colors from blue (negative correlation) to red (positive correlation).

# CCA in R

Classical CCA:  $n > p+q$

**matcor()&cc()**

```
Xr = as.matrix(nutrimouse$gene[, sample(1:120, size = 10)])  
res.cc = cc(Xr, Y)  
par(mar=c(4,4,4,4))  
barplot(res.cc$cor, xlab = "Dimension", ylab = "Canonical  
correlations", names.arg = 1:10, ylim = c(0,1))  
plt.cc(res.cc)
```

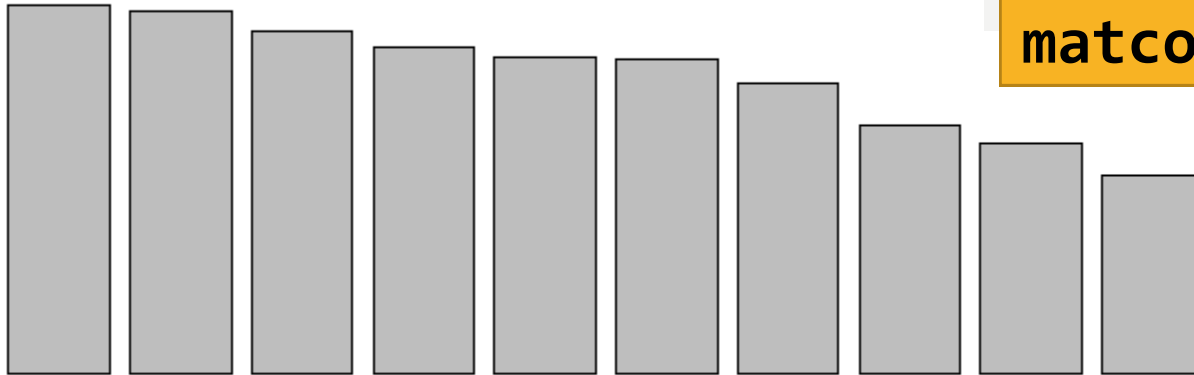
1. Randomly choose 10 genes among the 120  $\rightarrow$  Xr;
2. CCA on Xr (40-by-10) and Y (40-by-21)
3. Barplot of the canonical correlations (remember scree plot?)
4. Original variables and units are plotted on the first two canonical variates (remember similar presentations in PCA, MDS and EFA etc. ?)



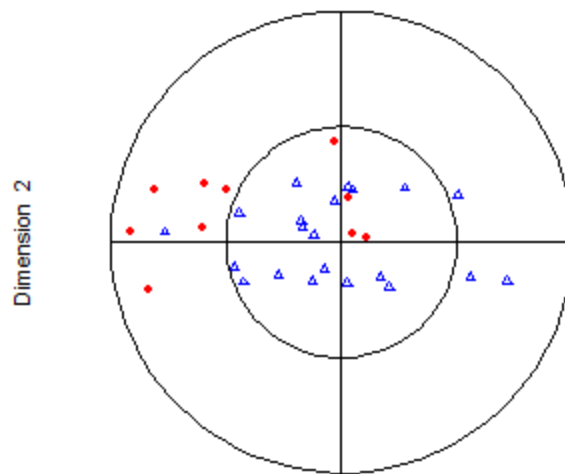
# CCA in R

`matcor()` & `cc()`

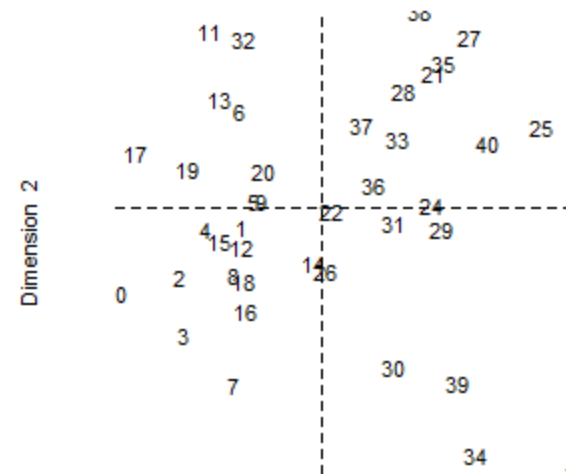
Canonical correlations



Dimension

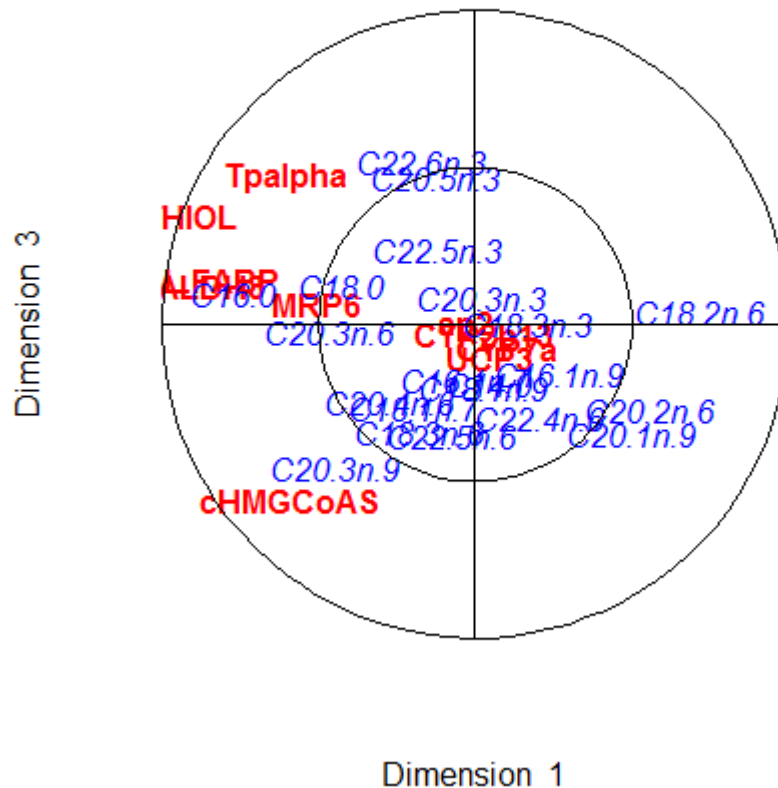


Dimension 1



Dimension 1

## matcor() & cc()

[illegible]

## Regularized CCA

Gene (X): 40-by-120; Lipid (Y): 40-by-21

Correlation matrix:  $S_{XX}$  and  $S_{YY}$

RCCA:  $\Sigma_{XX}(\lambda_1) = S_{XX} + \lambda_1 I_p$ ;  $\Sigma_{YY}(\lambda_2) = S_{YY} + \lambda_2 I_q$

How to set 'good' values for the regularization parameters?

- Leave-one-out cross validation by `estim.regul()`
- Default grid is 5 equally-spaced points [0.001,1]

```
res.regul = estim.regul(X, Y, plt = TRUE, grid1 =  
seq(0.0001, 0.2, l=51), grid2 = seq(0, 0.2, l=51))
```

```
#  $\lambda_1 = 0.008096$ ;
```

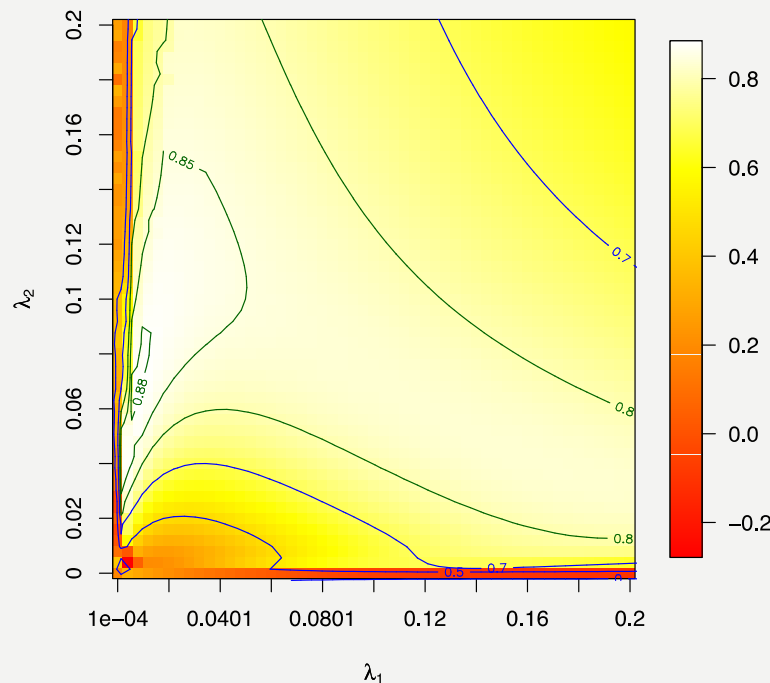
```
#  $\lambda_2 = 0.064$ ;
```

```
# CV-score = 0.8852923
```

```
contour(res.regul$grid1, res.regul$grid2, res.regul$mat, add = TRUE,  
levels = c(0,0.5,0.7), col = "blue")
```

```
contour(res.regul$grid1, res.regul$grid2, res.regul$mat, add = TRUE,  
levels = c(0.8,0.85,0.88), col = "darkgreen")
```

```
estim.regul()  
rcc()
```



CV-score for  $\lambda_1$  and  $\lambda_2$  on a  $51 \times 51$  grid defined by equally-spaced discretization points on the region:  $0.0001 \leq \lambda_1 \leq 0.2$  and  $0 \leq \lambda_2 \leq 0.2$ . Two kinds of contour plots are also displayed for values equal to  $\{0-0.5-0.7\}$  (in blue) and to  $\{0.8-0.85-0.88\}$  (in green).

### # Regularized CCA

```
res.rcc = rcc(X, Y, 0.008096, 0.064)
barplot(res.rcc$cor, xlab = "Dimension", ylab = "Canonical
correlations", names.arg = 1:21, ylim = c(0,1))
plt.cc(res.rcc, var.label = TRUE, ind.names =
paste(nutrimouse$genotype, nutrimouse$diet, sep = "-"))
```



# “CCA” in R

`cca()` in package(vegan)

- package(vegan) developed for community ecologists; thus a good collection of ordination methods for the field;
  - PCA, MDS, FA and more...
- But it has a lot of functions & associated visualization tools for us to use for other data.

data(*dune*): Vegetation and Environment in Dutch Dune Meadows

- The *dune* meadow vegetation data has cover class values of 30 species on 20 sites. The corresponding environmental data frame *dune.env* is a data frame of 20 observations on 5 variables

**CCA in package(vegan) stands for Canonical Correspondence Analysis**

## MDS in package(vegan)

- Non-metric MDS

- Dissimilarity: *vegdist*(,...)

- "manhattan", "euclidean", "canberra", "bray", "kulczynski", "jaccard", "gower", "altGower", "morisita", "horn", "mountford", "raup", "binomial", "chao", "cao" or "mahalanobis"

```
data(dune)
dis = vegdist(dune)
m = monoMDS(dis, model = "loc")
```

loc: non-metric MDS with separate regressions for each point

MDS model: "global" is normal non-metric MDS with a monotone regression, "local" is non-metric MDS with separate regressions for each point, "linear" uses linear regression, and "hybrid" uses linear regression for dissimilarities below a threshold in addition to monotone regression.

## MDS in package(vegan)

```
> m
```

Call:

```
monoMDS(dist = dis, model = "loc")
```

Local non-metric Multidimensional Scaling

20 points, dissimilarity 'bray', call 'vegdist(x = dune)'

Dimensions: 2

Stress: 0.1232248

Stress type 1, weak ties

Scores scaled to unit root mean square, rotated to principal components

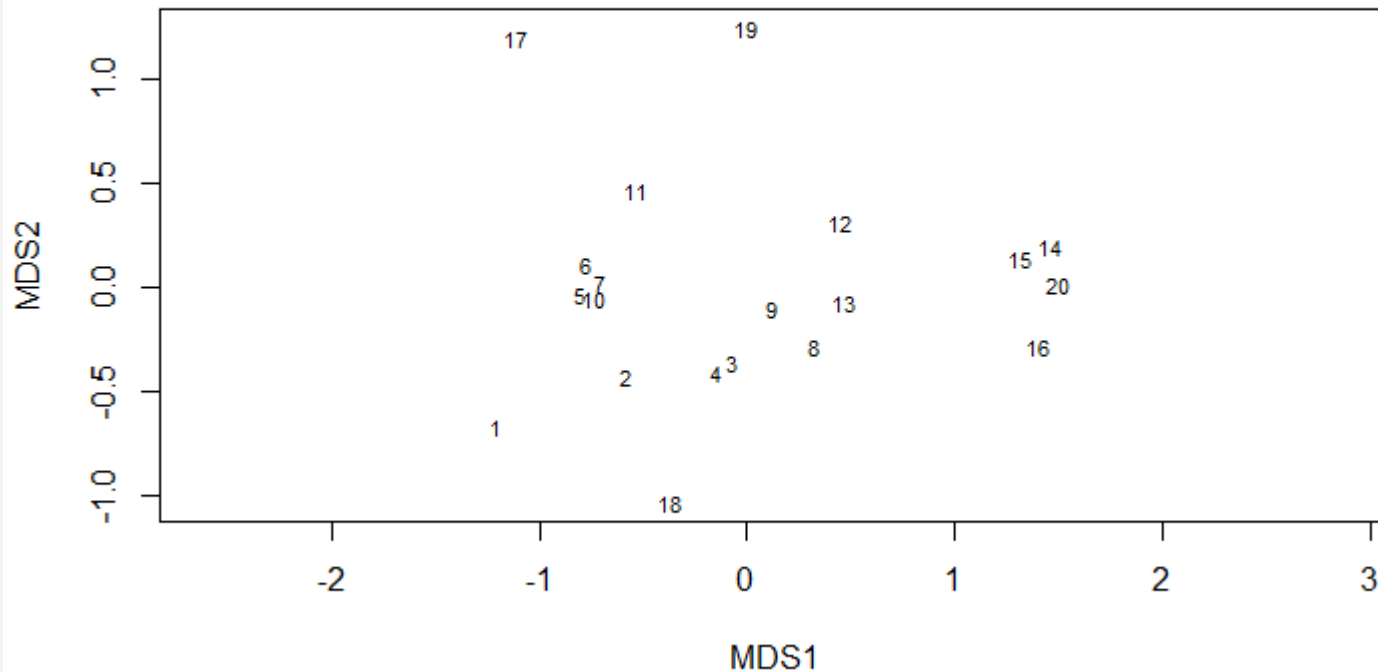
Stopped after 71 iterations: Stress nearly unchanged (ratio > sratmax)

```
> plot(m)
```

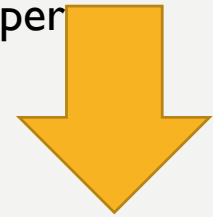
monoMDS has some rotation options to assist your initial diagnosis



## MDS in package(vegan)



Let's see another better wrapper  
for non-metric MDS in *vegan*



## metaMDS() in package(vegan)

### *metaMDS()* – THE POWERFUL WRAPPER

1. Use adequate dissimilarity measures (function *vegdist*),
2. Run NMDS several times with random starting configurations – avoid local minimum,
3. Compares results, w.r.t. minimizing sum of squared differences (function *procrustes*),
4. Stop after finding twice a similar minimum stress solution,
5. Scale & rotate the solution,
6. Add species scores to the configuration as weighted averages (function *wascores*): **before we only see the 20 sites**

A COMPLETE  
ANALYSIS

## metaMDS() in package(vegan)

```
> ord <- metaMDS(dune)
Run 0 stress 0.1192678
Run 1 stress 0.1812935
Run 2 stress 0.330578
Run 3 stress 0.1183186
... New best solution
... procrustes: rmse 0.02026999  max resid 0.06495494
Run 4 stress 0.1922247
Run 5 stress 0.2361935
Run 6 stress 0.119269
Run 7 stress 0.1808916
Run 8 stress 0.1192679
Run 9 stress 0.1192679
Run 10 stress 0.1192685
Run 11 stress 0.1183186
... procrustes: rmse 2.444379e-06  max resid
7.278529e-06
*** Solution reached
```

## metaMDS() in package(vegan)

```
> ord
```

```
Call:
```

```
metaMDS(comm = dune)
```

```
global Multidimensional Scaling using monoMDS
```

```
Data:      dune
```

```
Distance: bray
```

```
Dimensions: 2
```

```
Stress:      0.1183186
```

```
Stress type 1, weak ties
```

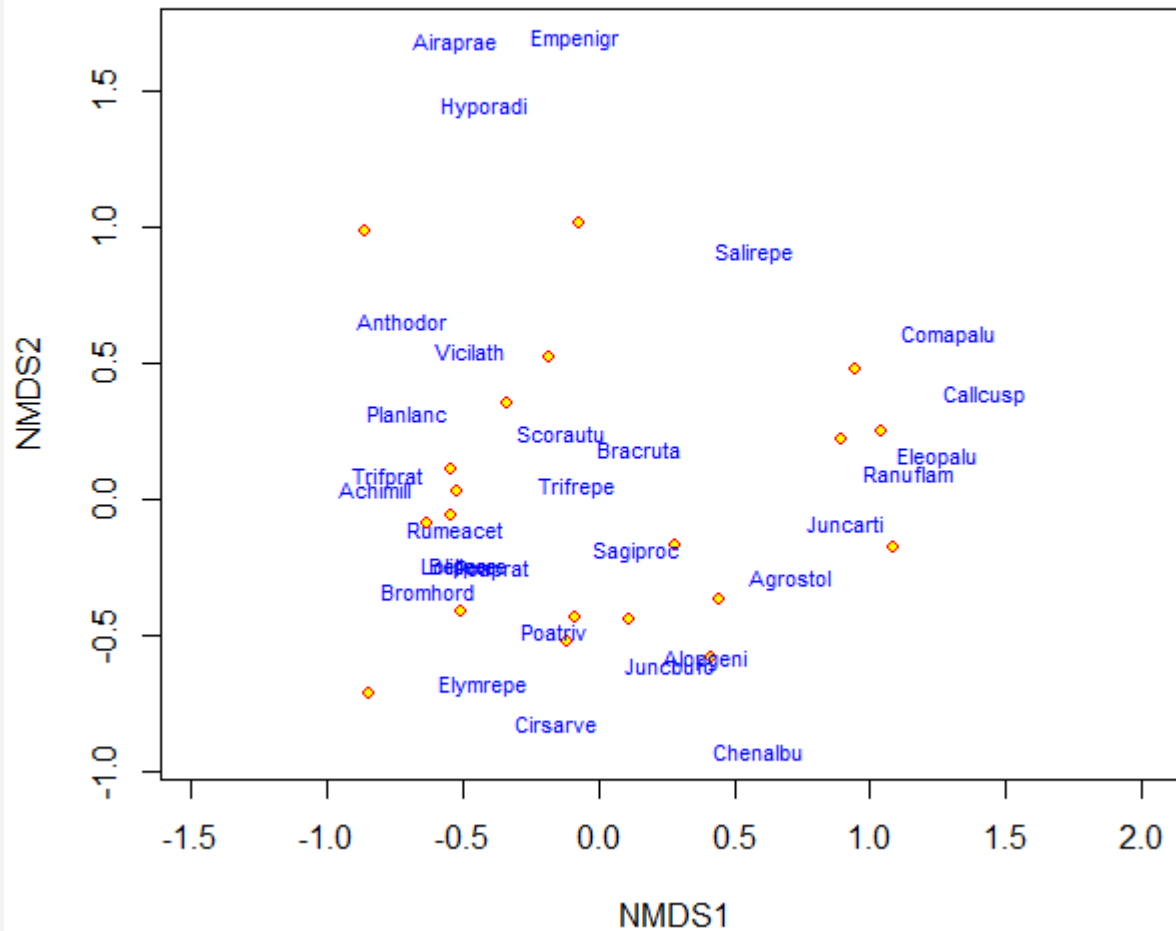
```
Two convergent solutions found after 11 tries
```

```
Scaling: centring, PC rotation, halfchange scaling
```

```
Species: expanded scores based on 'dune'
```

# CCA in R

`metaMDS()` in package(vegan)



# “CCA” in R

`cca()` in package(vegan)

`data(dune)`: Vegetation and Environment in Dutch Dune Meadows

- The *dune* meadow vegetation data has cover class values of 30 species on 20 sites. The corresponding environmental data frame *dune.env* is a data frame of 20 observations on the following 5 variables:

A1: a numeric vector of thickness of soil A1 horizon.

Moisture: an ordered factor with levels: 1 < 2 < 4 < 5.

Management: a factor with levels: BF (Biological farming), HF (Hobby farming), NM (Nature Conservation Management), and SF (Standard Farming).

Use: an ordered factor of land-use with levels: Hayfield < Haypastu < Pasture.

Manure: an ordered factor with levels: 0 < 1 < 2 < 3 < 4.

# “CCA” in R

`cca()` in package(vegan)

```
> cca(dune ~ ., data=dune.env)
```

```
Call: cca(formula = dune ~ A1 + Moisture + Management + Use +  
        Manure, data = dune.env)
```

Inertia Proportion Rank

Total	2.1153	1.0000
-------	--------	--------

Constrained	1.5032	0.7106	12
-------------	--------	--------	----

Unconstrained	0.6121	0.2894	7
---------------	--------	--------	---

Inertia is mean squared contingency coefficient

Some constraints were aliased because they were collinear (redundant)

Eigenvalues for constrained axes:

CCA1	CCA2	CCA3	CCA4	CCA5	CCA6	CCA7	CCA8	CCA9	CCA10
0.4671	0.3410	0.1761	0.1532	0.0953	0.0703	0.0589	0.0499	0.0318	0.0260
CCA11	CCA12								
0.0228	0.0108								

Eigenvalues for unconstrained axes:

CA1	CA2	CA3	CA4	CA5	CA6	CA7
0.27237	0.10876	0.08975	0.06305	0.03489	0.02529	0.01798

# Canonical Correspondence Analysis (Optional topic)

- References provided on CourseWorks



# SUMMARY

## Things to check

### ☐ Number of canonical variate pairs

- How many prove to be statistically/practically significant?

### ☐ Interpreting the canonical variates

- Where is the meaning in the combinations of variables created?

### ☐ Importance of canonical variates

- How strong is the correlation between the canonical variates?
- What is the nature of the variate's relation to the individual variables in its own set? The other set?

### ☐ Canonical variate scores

- If one did directly measure the variate, what would the subjects' scores be? (similar to the factor scores)

# Limitations

1. By maximizing the correlation between the linear combinations of X and Y, there is a possibility of having the combination unexplainable.
2. Nonlinearity still causes problems: Classical  $\rightarrow$  Regularized CCA
3. CCA is very sensitive to data involved
4. Correlation does not imply causality, necessary but not sufficient condition for causality.
5. It is more of a descriptive procedure, as being a simple correlation in the end.