

Proving the absence of unbounded polymers in rule-based models

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Abstract

Rule-based languages, such as Kappa and BNGL, allow for the description of very combinatorial models of interactions between proteins. A huge (when not infinite) number of different kinds of bio-molecular compounds may arise due to proteins with multiple binding and phosphorylation sites. Knowing beforehand whether a model may involve an infinite number of different kinds of bio-molecular compounds is crucial for the modeller. On the first hand, it is sometimes a hint for modelling flaws: forgetting to specify the conflicts among binding rules is a common mistake. On the second hand, it impacts the choice of the semantics for the models (among stochastic, differential, hybrid).

In this paper, we introduce a data-structure to abstract the potential unbounded polymers that may be formed in a rule-based model. This data-structure is a graph, the nodes of which are labelled with patterns while edges are labelled with overlaps between these patterns. By construction, every potentially unbounded polymer is associated to at least one cycle in that graph. This data-structure has two main advantages. Firstly, as opposed to site-graphs, one can reason about cycles without enumerating them, by the means of Tarjan's algorithm for detecting strongly connected components. Secondly, this data-structures may be combined easily with information coming from additional reachability analysis: the edges that are labelled with an overlap that is proved unreachable in the model may be safely discarded.

Keywords: Rule-based modelling, Polymers, Static analysis, Strongly connected components

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1 Introduction

Rule-based languages, such as Kappa [9] and BNGL [2], propose a transparent way to encode models of interactions between proteins. Systems involving races for shared resources, different time- and concentration-scales, non linear feedback loops may be described by the means of rewrite rules. This allows for the description of very combinatorial models. A huge (when not infinite) number of different kinds of bio-molecular compounds may arise due to the presence of scaffold and/or proteins with multiple binding and phosphorylation sites. The long term goal is then to understand how the collective behaviour of these proteins emerges from the mechanistic interactions between proteins.

Detecting whether such a model involve an infinite number of different kinds of bio-molecular compounds, is important. Often, the models come from a higher level of description [15] or from automatic mining of the literature [14]. The presence of an infinite number of bio-molecular compounds is often a hint for a lack of specification. Namely, conflicts between potential bindings have not be specified enough and there is a need to refine the model. Sometimes the assembling of giant molecules is involved. In that later case, it is important to confirm that the model implements properly what the modeller has in mind. The presence of an infinite number of distinct kinds of bio-molecular compounds also matters when choosing the most appropriate semantics for the models (among stochastic, differential, hybrid).

In this paper, we introduce some graph structures to abstract the potential presence of unbounded polymers in a rule-based model. These graphs either cope for the potential succession of sites along chains of proteins in reachable bio-molecular compounds, or for the succession of bonds in these chains. They provide a sound and complete (with respect to the information provided by the contact map of the model) description of the potential binding between the sites of proteins. The contact map encodes only non relational information: it cannot establish relationships about the different binding states of pairs of sites. Then, we show how to refine the graph of the potential successive links in bio-molecular compounds in order to refine it with the result of external relational static analyses [7,13,3]. Such static analyses provide a list of patterns that are known unreachable. As a result, we get a sound, but not complete approach (the detection of unreachable patterns in a rule-base language is undecidable anyway [17]) that may detect and prove that the set of non-isomorphic bio-molecular compounds of a model is finite, without executing the model.

The rest of the paper is organised as follows. Sec. 2 introduces some case study to provide intuitions about the property we want to infer, and to highlight pitfalls we will have to avoid. Sec. 3 gives some reminders about Kappa. In Sec. 4, we introduce two families of graph to decide whether or not the set of bio-molecular compounds that are compatible with a contact map is finite. We refine our approach to deal with black-listed patterns in Sec. 5.

2 Case studies

In this section, we introduce some examples to explain intuitively why there may be an unbounded number of bio-molecular compounds in a rule-based model. We also

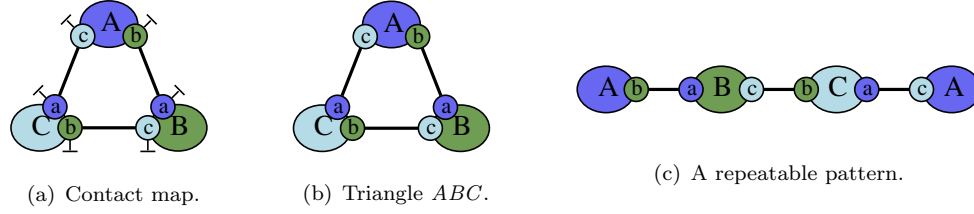


Fig. 1. The ABC example. The contact map (Fig.1(a)) specifies a typing discipline. It displays every kind of protein and specifies their interfaces. The contact map also provides the potential states for each site: either free \neg , or bound to another site (which is encoded as a link between pair of sites in the contact map). In Fig. 1(b) is described a bio-molecular compound that is compatible with the contact map. Every instance of proteins belongs to the contact map. Their interfaces are the same as in the contact map. Also any bond between two sites complies with one link explicitly written in the contact map. Fig. 1(c) describes a repeatable pattern. This pattern is compatible with the contact map and can be repeated in order to form arbitrarily large bio-molecular compounds.

explain why naive approaches may fail in proving that the number of bio-molecular compounds is finite in a given model when it is the case, while identifying the pitfalls that shall be avoided to achieve this goal.

2.1 Elementary cycles

Let us start with a simple example. We consider a model involving three kinds of protein A , B , C . Each protein has two binding sites: the protein A has the binding sites b and c , the protein B has the binding sites a and c , and the protein C has the binding sites a and b . Each binding site may be free, or bound to another site. Only three kinds of bond are possible: the site b of an instance of the protein A may be bound to the site a of an instance of the protein B ; the site c of an instance of the protein B may be bound to the site b of an instance of the protein C ; and the site a of an instance of the protein C may be bound to the site c of a protein A .

These assumptions are summarised in a graph in Fig. 1(a). This graph is called the contact map of the model. It describes every kind of protein and every site in their interfaces. The potential state of each site is also indicated. In our model, every site may be free: they are all tagged with the symbol \neg . Potential bonds are indicated by the means of non oriented edges between pairs of sites. The contact map provides a typing discipline. Every bio-molecular compound in our models shall satisfy the constraints the contact map is encoding about the interface of agents, the potential states of sites, and their potential bindings. An example of bio-molecular compound that is compatible with the contact map is drawn in Fig. 1(b). This bio-molecular compound is made of three proteins A , B , and C that are bound pair-wise so as to form a triangular shape. In a bio-molecular compound, every site shall be exclusively either free, or bound to at most one other site. In general, a bio-molecular compound does not have to contain an instance of each kind of protein. Also it may contain several instances of some of them.

The contact map that is given in Fig. 1(a) is compatible with an infinite number of different (i.e. *non isomorphic*) molecular compounds. Indeed we show in Fig. 1(c), a pattern that may be repeated an unbounded number of times in order to form arbitrary many different bio-molecular compounds. This is tempting to relate the potential presence of an arbitrary number of different bio-molecular compounds to the one of a cycle in the contact map. However we shall see in the next examples

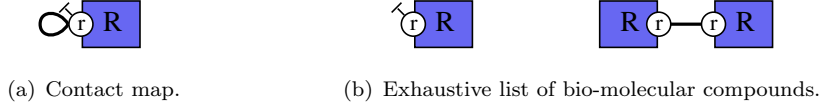


Fig. 2. The example of a protein that may form monomers and dimers. The contact map (e.g. see Fig. 2(a)) contains a cycle, since the unique site of an instance of a protein may be linked to the unique site of another instance of another protein. However, only once instance of this cycle may occur in a given bio-molecular compound and the number of bio-molecular compound remains bounded despite this cycle (e.g. see Fig. 2(b)).



Fig. 3. An example of a protein with two sites a and b such that the site a of a protein may be bound to the site a of another protein and the site b may be bound to the site b of another protein. The contact map (Fig. 3(a)) contains two self-loops. The pattern that is made of three proteins, the first two bound via their respective sites a and the last two bound via their respective sites b is a repeatable patterns. Thus, an infinite number of bio-molecular compounds is compatible with the contact map.

that this intuition is misleading.

2.2 Self loops

In this example we consider a model with only one kind of protein. This protein has a single site which may be either free, or bound to the unique site of another protein of the same kind. Roughly speaking proteins may form monomers and dimers. These assumptions are encoded in the contact map that is given in Fig. 2(a). We notice a cycle in this contact map (from the unique site of the protein to itself). Yet only the two bio-molecular compounds that are depicted in Fig. 2(b) are compatible with this contact map: there is a finite number of kinds of bio-molecular compound them despite the presence of a cycle in the contact map.

One could think that self-loops should not be considered as cycles when trying to prove the finiteness of the set of bio-molecular compounds of a model. Indeed whenever a molecular compound contains a bond that corresponds to a self-loop in the contact map, then both sites are necessarily bound together and they are no longer available to form links with other sites. Yet the contact map that is given in Fig. 3(a) shows that it is unsafe in general to discard self-loops from the contact map. In this example, we consider only one kind of protein with two sites. Each site may be either free, or bound to the same site of another instance of the protein. It is then possible to form a chain of three proteins (see Fig. 3(b)) that may be repeated an arbitrary number of times in a bio-molecular compound.

2.3 Conflicting bindings

In this example, we consider three kinds of protein G , R , and S . The proteins of kind G have a single site; the proteins of kind R have two sites g and s ; and the proteins of kind S have two sites g and r . Proteins R and S may bind to each-other via their respective sites s and r . The unique site of proteins G may bind either to the site g of an instance of the protein R , or to the site g of an instance of the protein S . Thus, there is a competition, or a conflict, on the site of the protein G .

The contact map for this example is provided in Fig. 4(a). We notice that the

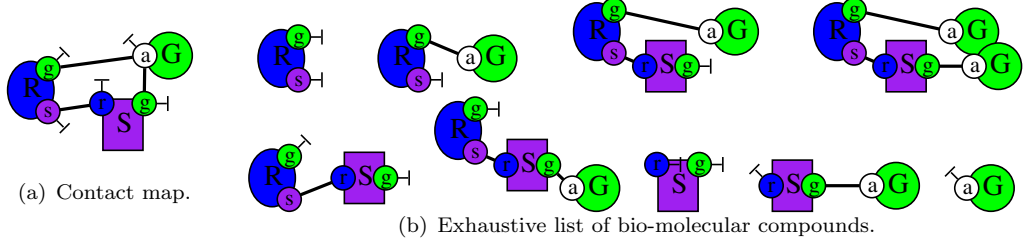


Fig. 4. An example of a protein with a site that may be bound to two different kinds of site. As drawn in the contact map (e.g. see Fig. 4(a)), the site of the protein G may be either free, bound to the site g of the protein R , or bound to the site g of the protein S . The cycle in the contact map does not induce an infinite number of different bio-molecular compounds (e.g. see Fig. 4(b)).

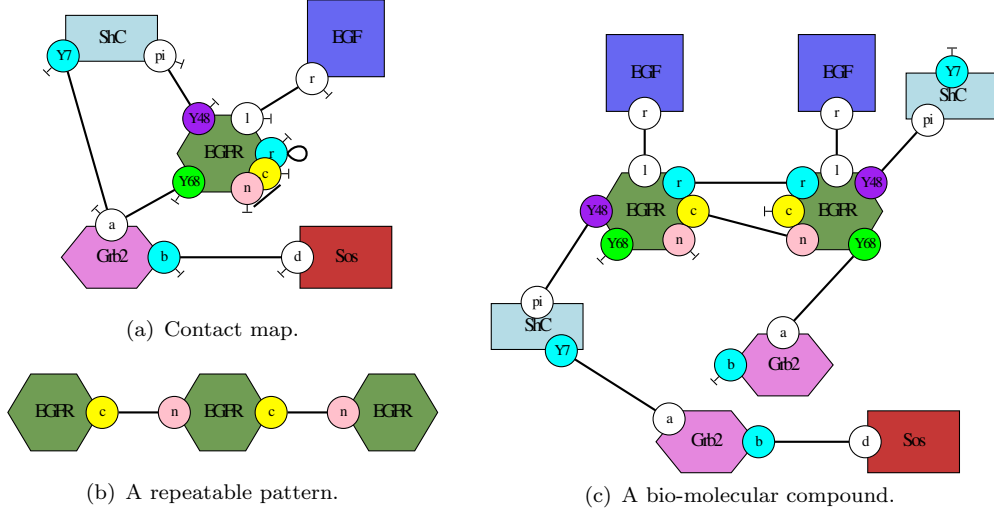


Fig. 5. The example of the early events in the epidermic growth factor [1]. In Fig. 5(a) is drawn the contact map. Compared to the original model in BNGL, we have omitted phosphorylation states, since they have no impact on the binding topology. We have also added two sites in the receptor to model the asymmetric bond between receptors $EGFR$ in dimers. The model is constrained by the following property: whenever the site c of a receptor $EGFR$ is bound, then its site r is bound as well, and both sites are bound to the same instance of protein. The contact map is compatible with the repeatable pattern that is given in Fig. 5(b). Yet this pattern does not satisfy the additional constraint. Indeed the model has only a finite set of different bio-molecular compounds. In Fig. 5(c) is given an example of a typical bio-molecular compound.

competition on the site of the protein G belongs to a cycle in this contact map. Yet, in a given bio-molecular compound, the site of each instance of G is either free, or bound to at most one site. Thus the cycle of the contact map is not realisable in a concrete bio-molecular compound. In Fig. 4(b), we enumerate all the bio-molecular compounds that are compatible with the constraints encoded in the contact map. There is a finite amount of them, despite the presence of a cycle in the contact map.

2.4 Early events in the epidermic growth factor pathway

So far, we have considered only toy examples, since we tried to understand which conditions on a contact map are necessary to induce only a finite number of bio-molecular compounds. In Fig. 5, we consider a model for the early events in the integration of the epidermic growth factor (EGF) [1]. In this model, the acquisition of the protein Sos by the membrane of the cell is made in several steps. Firstly a pair of receptors $EGFR$ on the membrane of the cell shall be activated by the ligands EGF . Once bound to their respective ligands, they can form a dimer thanks

to a symmetric bond via their respective sites r . Compared to the BNGL model of [1], asymmetric bonds between receptors are also considered. To stabilise dimers, pairs of receptors that are bound via their sites r form an asymmetric binding by connecting the site c of one receptor to the site n of the other receptor. The symmetric bond in a dimer cannot be released in the presence of an asymmetric one. As a consequence, whenever the site c of a receptor is bound to the site n of another receptor, these receptors are also connected by a symmetric bond. This property can be inferred by the static analysis that is described in [13,3]. Each receptor in a dimer may activate the sites $Y48$ and $Y68$ of the other receptor (since we focus only on the binding topology, we omit the details about these activations which are performed by the means of phosphorylation). The site $Y68$ may bind to the protein *Grb2*, which may be, or not, bound to the protein *Sos*. The site $Y48$ connects to the protein *Grb2* indirectly, thanks to the adapter protein *Shc*.

It is worth noticing that the contact map, that is depicted in Fig. 5(a) does not provide all the information about the model. The constraints on the sites c , n , and r emerge from some mechanisms that are described by the means of rules. Rules are omitted here so as to focus on the topology of the potential bindings between the sites of proteins. Yet some additional constraints may be provided as a list of forbidden patterns. This way, we assume that the bio-molecular compounds of our model are the ones that are compatible with the contact map and that does not contain black-listed patterns.

Interestingly, the contact map of the EGF model (e.g. see 5(a)) contains both issues that we have pointed out in Sec. 2.2 and in Sect. 2.3. Indeed, the site r of a receptor may be bound to the site r of another receptor. Moreover there is a conflict on the site a of the protein *Grb2* which may be bound to the receptor directly or via an adapter protein. Another issue is raised by this model. The constraints provided by the contact map are not enough to ensure the finiteness of the set of the different bio-molecular compounds. Indeed, the pattern that is provided in Fig. 5(b) is compatible with the contact map, and could be repeated an unbounded number of times to form an infinite number of different bio-molecular compounds. Nevertheless, this pattern is not compatible with the additional constraints about symmetric and asymmetric bindings in dimers: there is only a finite number of different bio-molecular compounds that satisfies both the constraints from the contact map and the additional constraint. In Fig. 5(c), we provide a typical example of bio-molecular compound in the EGF model. This example is made of a dimer, with one site $Y68$ free, one site $Y68$ connected to a *Grb2* not connected to a *Sos*, one site $Y48$ connected to an adapter not connected to a *Grb2*, and a site $Y48$ connected to a *Sos*. In total, a dimer may be connected to up to four instances of *Sos*.

On such a rather small model, it is possible to enumerate the different bio-molecular compounds thanks to reaction enumeration engines [2,5]. This model is made of 253 kinds of bio-molecular compounds. Taking into account phosphorylation states would lead to a model with 932 kinds of bio-molecular compounds. Nevertheless, enumeration engines do not scale to large combinatorial networks such as the longer version of the EGF model (including the interactions with the proteins Ras, Erk, and Mapk) that is described in [6] and that involves about 10^{19} different kinds of bio-molecular compounds [7] or as the model of the interactions found in

the cytoplasmic portion of the Structural Interaction Network (cSIN) [10,16] that involves an infinite number of bio-molecular compounds.

We will design a well-suited data-structure to abstract the elementary repeatable patterns that are compatible with a contact map and with additional constraints.

2.5 Clique

In large combinatorial models, the set of elementary repeatable patterns may not be represented explicitly. It is important to abstract it.

Let us consider the example of a clique of n proteins. We call a clique of n proteins any n kinds of protein such that each protein has exactly $n - 1$ sites and that every pair of proteins of distinct kinds may be connected by exactly one pair of sites. The number of elementary repeatable patterns in a clique of n proteins is exponential with respect to n (there is indeed $\frac{n!}{k!}$ elementary repeatable patterns with exactly $k + 1$ proteins, for any k such that $2 \leq k \leq n$). Thus they cannot be all enumerated. In this paper, we will instead compute exactly the set of bonds that may occur in repeatable patterns. Our approach is based on the use of some graphs that are derived from the contact map, and for which edges correspond to the potential bonds in elementary repeatable patterns. We use Tarjan’s algorithm [19] to compute the strongly connected components of these graphs. Our analysis is sound and complete with respect to the constraints that are encoded in the contact map: a bond may occur in a repeatable pattern that is compatible with a given contact map if and only if it corresponds to an edge in a non trivial strongly connected component of the graph that is associated to this contact map. Moreover, it is possible to take into account additional constraints about the patterns that are proved to be unreachable by traditional static analysis [13,3].

Outline. The rest of the paper is organised as follows. In Sec. 3, we give some reminders about Kappa. We focus only on static reasoning about graphs. We do not introduce the notion of rules. We assume that additional constraints about reachable patterns come from a black box that we do not describe in this paper. In Sec. 4, we introduce two notions of graphs: the graph of the sites and the graph of the potential links. Both notions can be used to reason about the finiteness of the set of bio-molecular compounds in a Kappa model. Yet we will see in Sec. 5, that the graph of the potential links may be refined to take into account the patterns that may be proved unreachable by an external tool.

3 Kappa

In this section, we give some reminders about Kappa. We do not introduce the full semantics of Kappa. Instead, we introduce only the notions of site-graphs and of embeddings among them. We omit the notions of rules and of rule applications. We also omit internal states, since we focus on the topology of the potential bindings between proteins. We refer to [9,12] for a more complete description of Kappa.

3.1 Signature

Firstly we define the signature of a model.

Definition 3.1 (signature) A signature is a triple $\Sigma \triangleq (\Sigma_{ag}, \Sigma_{site}, \Sigma_{ag-st})$ where:

- (i) Σ_{ag} is a finite set of agent types,
- (ii) Σ_{site} is a finite set of site identifiers;
- (iii) $\Sigma_{ag-st} : \Sigma_{ag} \rightarrow \wp(\Sigma_{site})$ is a site map.

Agent types in Σ_{ag} denote agents of interest, as kinds of protein for instance. Site identifiers in Σ_{site} represent identified loci for capabilities of interactions. Agent types $A \in \Sigma_{ag}$ are associated with sets of sites $\Sigma_{ag-st}(A)$ which may be linked.

Example 3.2 (signature (model of the triangle)) We define the signature for the model of the triangle (e.g. see Sec. 2.1):

$$\Sigma \triangleq (\Sigma_{ag}, \Sigma_{site}, \Sigma_{ag-st})$$

where:

- (i) $\Sigma_{ag} \triangleq \{A, B, C\}$;
- (ii) $\Sigma_{site} \triangleq \{a, b, c\}$;
- (iii) $\Sigma_{ag-st} \triangleq [A \mapsto \{b, c\}, B \mapsto \{a, c\}, C \mapsto \{a, b\}]$.

Example 3.3 (signature) We define the signature for the model of the early events in the epidermic growth factor (e.g. see Sec. 2.4):

$$\Sigma \triangleq (\Sigma_{ag}, \Sigma_{site}, \Sigma_{ag-st})$$

where:

- (i) $\Sigma_{ag} \triangleq \{EGF, EGFR, Grb2, ShC, Sos\}$;
- (ii) $\Sigma_{site} \triangleq \{a, b, c, d, n, l, pi, r, Y7, Y48, Y68\}$;
- (iii) $\Sigma_{ag-st} \triangleq \left[\begin{array}{l} EGF \mapsto \{r\}, EGFR \mapsto \{c, n, l, r, Y48, Y68\}, \\ Grb2 \mapsto \{a, b\}, ShC \mapsto \{pi, Y7\}, Sos \mapsto \{d\} \end{array} \right]$

3.2 Σ -graphs and morphisms among Σ -graphs

Σ -graphs are graphs. Their nodes are typed agents with some sites which may bear sets of binding states. Contact maps, patterns and bio-molecular compounds are specific kinds of Σ -graph.

Definition 3.4 (Σ -graphs) A Σ -graph is a tuple $G \triangleq (\mathcal{A}_G, type_G, \mathcal{S}_G, \mathcal{L}_G)$ where:

- (i) $\mathcal{A}_G \subseteq \mathbb{N}$ is a finite set of agents,
- (ii) $type_G : \mathcal{A}_G \rightarrow \Sigma_{ag}$ is a function mapping each agent to its type,
- (iii) \mathcal{S}_G is a subset of the set $\{(n, i) \mid n \in \mathcal{A}_G, i \in \Sigma_{ag-st}(type_G(n))\}$,
- (iv) \mathcal{L}_G is a function between the set \mathcal{S}_G and the set $\wp(\mathcal{S}_G \cup \{\perp\})$ such that for any two sites $(n, i), (n', i') \in \mathcal{S}_G$, we have $(n', i') \in \mathcal{L}_G(n, i)$ if and only if $(n, i) \in \mathcal{L}_G(n', i')$.

The set \mathcal{S}_G denotes the set of binding sites. Whenever $\perp \in \mathcal{L}_G(n, i)$, the site (n, i) may be free. Whenever $(n', i') \in \mathcal{L}_G(n, i)$ (and hence $(n, i) \in \mathcal{L}_G(n', i')$), the

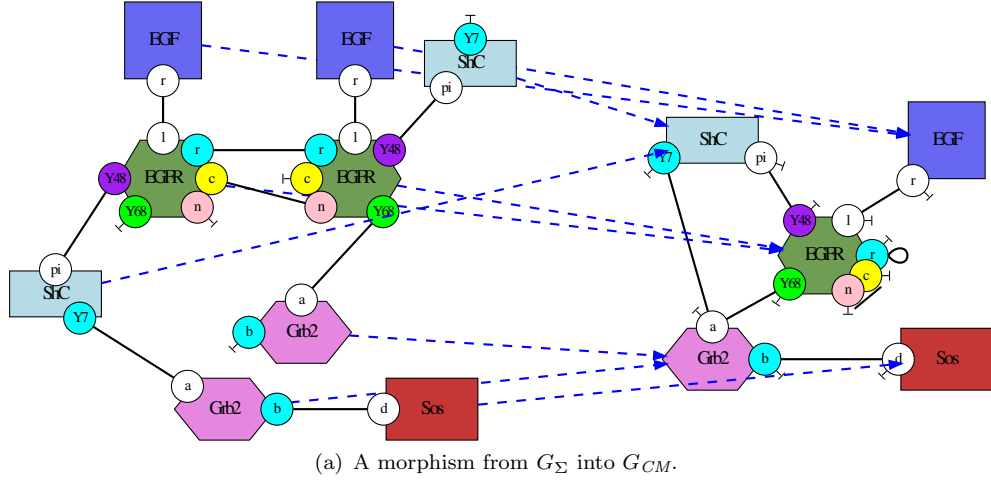


Fig. 6. Two Σ -graphs G_{CM} and G_{SP} , and a morphism from G_{CM} to G_Σ . The Σ -graph G_{CM} is a contact map. It provides context-insensitive information about the potential state of each binding site. The Σ -graph G_{SP} is a bio-molecular compounds. It contains several instances of some proteins. Every site is documented in each protein instance and each site is either free, or bound to another site. The morphism between G_{CM} and G_{SP} smashes all the proteins of the Σ -graph G_{SP} according to their type. This is the unique morphism from the site graph G_{CM} into the site-graph G_{SP} .

sites (n, i) and (n', i') may be bound together.

For a Σ -graph G , we write as \mathcal{A}_G its set of agents, $type_G$ its typing function, \mathcal{S}_G its set of sites, and \mathcal{L}_G its set of links.

Example 3.5 (Σ -graphs (model of the triangle)) We give two examples of Σ -graph for the model of the triangle (eg. see Fig. 1).

The graph that is depicted in Fig. 1(a) is the Σ -graph \mathcal{T}_{CM} defined as follows:

- (i) $\mathcal{A}_{\mathcal{T}_{CM}} \triangleq \{1, 2, 3\}$;
- (ii) $type_{\mathcal{T}_{CM}} \triangleq [1 \mapsto A, 2 \mapsto B, 3 \mapsto C]$;
- (iii) $\mathcal{S}_{\mathcal{T}_{CM}} \triangleq \bigcup \{(n, i) \mid n \in \mathcal{A}_{\mathcal{T}_{CM}}, i \in \Sigma_{ag-st}(type_{\mathcal{T}_{CM}})\}$;
- (iv) $\mathcal{L}_{\mathcal{T}_{CM}} \triangleq \left[\begin{array}{l} (1, b) \mapsto \{\neg, (2, a)\}, (1, c) \mapsto \{\neg, (3, a)\}, (2, a) \mapsto \{\neg, (1, b)\}, \\ (2, c) \mapsto \{\neg, (3, b)\}, (3, a) \mapsto \{\neg, (1, c)\}, (3, b) \mapsto \{\neg, (2, c)\} \end{array} \right]$.

and the bio-molecular compound that is drawn in Fig. 1(b), is the Σ -graph \mathcal{T}_Σ that is defined as follows:

- (i) $\mathcal{A}_{\mathcal{T}_\Sigma} \triangleq \{1, 2, 3\}$;
- (ii) $type_{\mathcal{T}_\Sigma} \triangleq [1 \mapsto A, 2 \mapsto B, 3 \mapsto C]$;
- (iii) $\mathcal{S}_{\mathcal{T}_\Sigma} \triangleq \bigcup \{(n, i) \mid n \in \mathcal{A}_{\mathcal{T}_\Sigma}, i \in \Sigma_{ag-st}(type_{\mathcal{T}_\Sigma})\}$;
- (iv) $\mathcal{L}_{\mathcal{T}_\Sigma} \triangleq \left[\begin{array}{l} (1, b) \mapsto \{(2, a)\}, (1, c) \mapsto \{(3, a)\}, (2, a) \mapsto \{(1, b)\}, \\ (2, c) \mapsto \{(3, b)\}, (3, a) \mapsto \{(1, c)\}, (3, b) \mapsto \{(2, c)\} \end{array} \right]$.

Example 3.6 (Σ -graph (EGF model)) We give two examples of Σ -graph for the model of the early events of the integration of the epidermic growth factor (eg. see Fig. 2.4).

The graph that is depicted in Fig. 5(a) is the Σ -graph G_{CM} defined as follows:

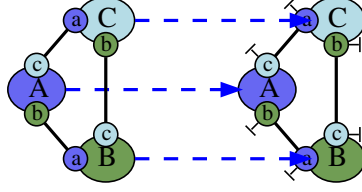
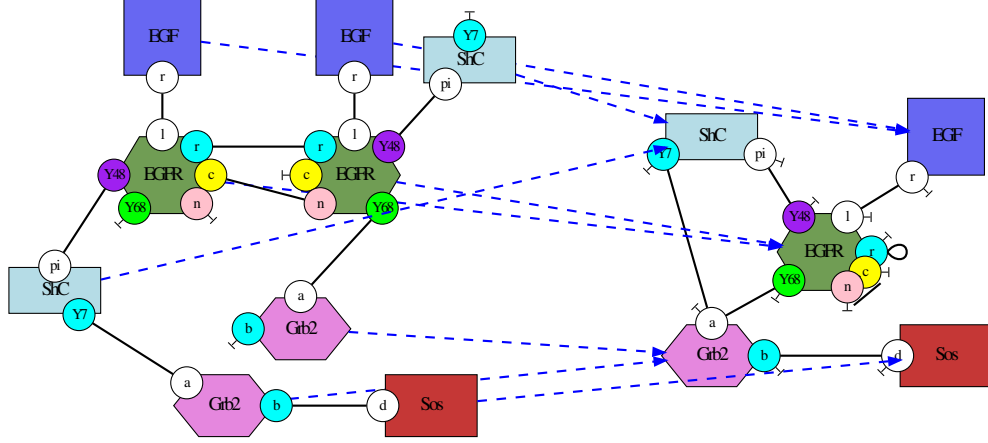
- (i) $\mathcal{A}_{G_{CM}} \triangleq \{1, 2, 3, 4, 5\}$;
- (ii) $type_{G_{CM}} \triangleq [1 \mapsto EGF, 2 \mapsto EGFR, 3 \mapsto Grb2, 4 \mapsto ShC, 5 \mapsto Sos]$;
- (iii) $\mathcal{S}_{G_{CM}} \triangleq \bigcup \{(n, i) \mid n \in \mathcal{A}_{G_{CM}}, i \in \Sigma_{ag-st}(type_{G_{CM}})\}$;
- (iv) $\mathcal{L}_{G_{CM}} \triangleq \left[\begin{array}{l} (1, r) \mapsto \{\neg, (2, l)\}, \\ (2, l) \mapsto \{\neg, (1, r)\}, (2, r) \mapsto \{\neg, (2, r)\}, (2, c) \mapsto \{\neg, (2, n)\}, \\ (2, n) \mapsto \{\neg, (2, c)\}, (2, Y48) \mapsto \{\neg, (4, pi)\}, (2, Y68) \mapsto \{\neg, (3, a)\}, \\ (3, a) \mapsto \{\neg, (2, Y68), (4, Y7)\}, (3, b) \mapsto \{\neg, (5, d)\}, \\ (4, pi) \mapsto \{\neg, (2, Y48)\}, (4, Y7) \mapsto \{\neg, (3, a)\}, \\ (5, d) \mapsto \{\neg, (3, b)\}, \end{array} \right].$

and the Σ -graph G_{Σ} that is defined as follows:

- (i) $\mathcal{A}_{G_{\Sigma}} \triangleq \{1, 2, 3, 4\}$;
- (ii) $type_{G_{\Sigma}} \triangleq \left[\begin{array}{l} 1 \mapsto EGF, 2 \mapsto EGF, 3 \mapsto EGFR, 4 \mapsto EGFR, \\ 5 \mapsto Grb2, 6 \mapsto Grb2, 7 \mapsto ShC, 8 \mapsto ShC, 9 \mapsto Sos \end{array} \right];$
- (iii) $\mathcal{S}_{G_{\Sigma}} \triangleq \bigcup \{(n, i) \mid n \in \mathcal{A}_{G_{\Sigma}}, i \in \Sigma_{ag-st}(type_{G_{\Sigma}})\}$;
- (iv) $\mathcal{L}_{G_{\Sigma}} \triangleq \left[\begin{array}{l} (1, r) \mapsto \{(3, l)\}, (2, r) \mapsto \{(4, l)\}, \\ (3, l) \mapsto \{(1, r)\}, (3, r) \mapsto \{(4, r)\}, (3, c) \mapsto \{(4, n)\}, \\ (3, n) \mapsto \{\neg\}, (3, Y48) \mapsto \{(7, pi)\}, (3, Y68) \mapsto \{\neg\}, \\ (4, l) \mapsto \{(2, r)\}, (4, r) \mapsto \{(3, r)\}, (4, c) \mapsto \{\neg\}, \\ (4, n) \mapsto \{(3, c)\}, (4, Y48) \mapsto \{(8, pi)\}, (4, Y68) \mapsto \{(6, a)\}, \\ (5, a) \mapsto \{(7, Y7)\}, (5, b) \mapsto \{(9, d)\}, \\ (6, a) \mapsto \{(4, Y68)\}, (6, b) \mapsto \{\neg\}, \\ (7, pi) \mapsto \{(3, Y48)\}, (7, Y7) \mapsto \{(5, a)\}, \\ (8, pi) \mapsto \{(4, Y48)\}, (8, Y7) \mapsto \{\neg\}, \\ (9, d) \mapsto \{(5, b)\} \end{array} \right].$

The Σ -graphs \mathcal{T}_{CM} and G_{CM} play a specific role: we call them the contact maps of their respective models. In a contact map each agent type occurs exactly once and each agent documents its full set of sites. Moreover every sites may be free, but may also be bound to some other sites as specified in the corresponding Σ -graph. Contact maps encode some specific typing disciplines [8]: they summarise the potential bonds and provide contextual conditions over them [4].

Σ -graphs may be related by structure-preserving maps of agents, called morphisms. The definition of a morphism between two Σ -graphs is given as follows:


 Fig. 7. The unique morphism from the Σ -graph \mathcal{T}_Σ and the Σ -graph \mathcal{T}_{CM} .

 Fig. 8. The unique morphism from the Σ -graph G_Σ and the Σ -graph G_{CM} .

Definition 3.7 (morphisms) A morphism $h : G \rightarrow H$ from the Σ -graph G into the Σ -graph H is a function of agents $h : \mathcal{A}_G \rightarrow \mathcal{A}_H$ satisfying, for all agent identifiers $n, n' \in \mathcal{A}_G$, for all site identifiers $i \in \Sigma_{ag-st}(\text{type}_G(n))$, $i' \in \Sigma_{ag-st}(\text{type}_G(n'))$:

- (i) $\text{type}_G(n) = \text{type}_H(h(n))$;
- (ii) if $(n, i) \in \mathcal{S}_G$, then $(h(n), i) \in \mathcal{S}_H$;
- (iii) if $(n', i') \in \mathcal{L}_G(n, i)$, then $(h(n'), i') \in \mathcal{L}_H(h(n), i)$;
- (iv) if $\neg \in \mathcal{L}_G(n, i)$, then $\neg \in \mathcal{L}_H(h(n), i)$.

Morphisms preserve the type of agents. They also preserve each agent set of sites, but more sites may be documented in the image of the morphism. A site that may be free shall be mapped to a site that may be free. Two sites that may be bound together shall be mapped to two sites that may be bound together.

Example 3.8 (morphisms (model of the triangle)) A morphism between the Σ -graph \mathcal{T}_Σ and the Σ -graph \mathcal{T}_{CM} is depicted in Fig. 7. This morphism maps any agent of the Σ -graph \mathcal{T}_Σ to the unique agent of the Σ -graph \mathcal{T}_{CM} having the same type. This is indeed the unique morphism from the Σ -graph \mathcal{T}_Σ to the Σ -graph \mathcal{T}_{CM} .

Example 3.9 (morphisms (EGF model)) A morphism between the Σ -graph G_Σ and the Σ -graph G_{CM} is depicted in Fig. 8. This morphism maps any agent of the Σ -graph G_Σ to the unique agent of the Σ -graph G_{CM} having the same type. This is indeed the unique morphism from the Σ -graph G_Σ to the Σ -graph G_{CM} .

Two morphisms from a Σ -graph E to a Σ -graph F , and from the Σ -graph F to a Σ -graph G respectively, compose in the usual way (and form a morphism from the Σ -graph E into the Σ -graph G).

3.3 Patterns and embeddings

Now we restrict the definition of Σ -graphs so as to focus on the ones that may express parts of the state of the system. These Σ -graphs, that we call patterns, are defined as follows:

Definition 3.10 (patterns) *A pattern is a Σ -graph P such that, for every site $s \in \mathcal{S}_P$ both following conditions are satisfied:*

- (i) *the set $\mathcal{L}_P(s)$ contains at most one element;*
- (ii) *the set $\mathcal{L}_P(s)$ does not contain the element s .*

The first condition ensures that the state of every site is either unspecified, or free, or bound to a single specific site. The second condition ensures that a site is never bound to itself.

A bio-molecular compound is a connected pattern in which the state of each site is documented (no further information may be added).

Patterns may be related by embeddings. Besides preserving the structure of patterns, embeddings map agents to agents injectively.

Definition 3.11 (embeddings) *An embedding is a morphism from a pattern into another one, that is induced by an injective agent function.*

As opposed to classical notions of embeddings between graphs, embeddings between patterns preserve free sites. When there exists an embedding from a pattern E into a pattern F , we often write that the pattern E embeds in the pattern F , or that E occurs in the pattern F . The composition of two embeddings is an embedding. Two patterns E and F are isomorphic whenever there exist an embedding from the pattern E to the pattern F and an embedding from the pattern F to the pattern E , which is denoted as $E \approx F$. We also denote as $[E]_{\approx}$ the \approx -equivalence class of the pattern E . The \approx -equivalence class $[E]_{\approx}$ of the pattern E is made of all the patterns that are isomorphic to the pattern E .

4 Reasoning on repeatable patterns

In this section, we formalise the problem of deciding whether or not a contact map is compatible with an infinite set of bio-molecular compounds. Then we introduce two kinds of graph to reason about this problem.

4.1 Interpretation of a contact map

Intuitively, a contact map may be interpreted as the set of the bio-molecular compounds which may be projected into that contact map by the means of a morphism. However this notion is not relevant to reason about the finiteness of the set of the bio-molecular compounds in a given model. Indeed with such a definition, each model admitting at least one bio-molecular compound would admit an infinite number of bio-molecular compounds due to isomorphisms. Thus we consider \approx -equivalence classes of bio-molecular compounds instead.

Definition 4.1 (interpretation of a contact map) *The interpretation $\llbracket G_{CM} \rrbracket$ of a contact map G_{CM} is defined as the set of all the \approx -equivalence classes of bio-*

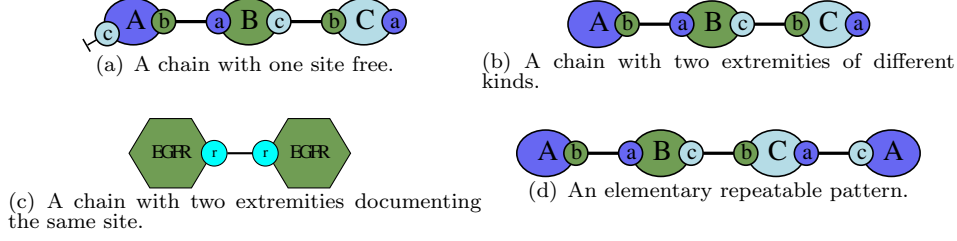


Fig. 9. Four patterns. Each of them is a chain. But only the last one is repeatable.

molecular compound $[G]_{\approx}$ such that there exists a morphism from the site graph G into the contact map G_{CM} .

We can now state properly the problem we want to solve:

Problem 4.2 *Let G_{CM} be a contact map. We are looking for an automatic procedure to decide whether the set $\llbracket G_{CM} \rrbracket$ is finite, or not.*

4.2 Chains

In this section, we introduce a kind of pumping lemma in order to reduce Problem 4.2 to the one of detecting a repeatable pattern.

Firstly, we define properly a repeatable pattern as a chain of agents which may be iterated to form arbitrarily long patterns.

Definition 4.3 (chain) *A pattern is called a chain if and only if it satisfies the following properties:*

- (i) *every agent documents at most two sites;*
- (ii) *there is an agent with a site free or that documents at most one site (or both);*
- (iii) *at most two agents do not have two sites bound.*

In particular, every chain is connected. A chain is formed either of a single agent with at most two sites all free, or of a linear chain of agents with exactly two extremities. In the latter case, every agent not in the extremities has two sites and these sites are bound. The agents on the extremity either have exactly one site that is bound. Additionally, it may have at most one other site (which is free).

A chain is a repeatable patterns whenever it contains at least two agents and its extremities may be replug to each other. This is formalised as follows.

Definition 4.4 (repeatable pattern) *A chain is called a repeatable pattern if and only if the following conditions are satisfied:*

- (i) *it has two distinct extremities;*
- (ii) *it has no free sites;*
- (iii) *both agents at the extremities are of the same kind;*
- (iv) *both sites documented at the extremities are different.*

A repeatable pattern is said elementary if and only if it contains no occurrence of repeatable patterns (besides itself).

Example 4.5 *We consider four patterns in Fig. 9. All these patterns are chains.*

The pattern in Fig. 9(a) is not repeatable because one of its extremity has a free site. The pattern in Fig. 9(b) is not repeatable because its extremities are not of the same kind. The pattern in Fig. 9(c) is not repeatable because its extremities document the same site. The pattern in Fig. 9(d) is repeatable (and elementary).

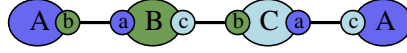
We can now establish our pumping lemma.

Lemma 4.6 (pumping lemma) *Let G_{CM} be a contact map. Both following assertions are equivalent:*

- (i) *The set $\llbracket G_{CM} \rrbracket$ is infinite;*
- (ii) *There exist an elementary repeatable pattern P and a morphism between the pattern P and the contact map G_{CM} .*

4.3 Graph of the sites

It is tempting to interpret the following repeatable pattern:



as the sequence of sites b of A , a of B , c of B , b of C , a of C , and c of A . Yet in this sequence, sites are polarised. Each site on a odd position and the next one always belong to the same kind of protein. While there always exists a link between each site on an even position and the next one. Due to this polarisation, it is tempting to consider the sub-sequence of each other site in that sequence of sites.

Next we define a graph that stands for all the potential sequences of sites that may occur on even occurrences in the repeatable patterns that are compatible with a given contact map. This is the graph of the sites of this contact map.

Definition 4.7 (graph of the sites) *Let G_{CM} be a contact map.*

The contact map G_{CM} is associated with a classical graph $(\mathcal{V}, \mathcal{E})$, called the graph of the sites of the contact map G_{CM} , which is defined as follows:

- \mathcal{V} is the set $\mathcal{S}_{G_{CM}}$ of the sites of the Σ -graph G_{CM} .
- \mathcal{E} is the subset of $V \times V$ such that $((n, i), (n', i')) \in E$ if and only if there exists a site $i'' \in \Sigma_{ag-st}(\text{type}_{G_{CM}}(n'))$ such that: $i'' \neq i'$ and $(n', i'') \in \mathcal{L}_{G_{CM}}(n, i)$.

In the edges of the graph of the sites, the sites via which we enter the target agent is kept implicit.

The following theorem relates the cycles in the graph of the sites to the existence of repeatable patterns.

Theorem 4.8 *Let G_{CM} be a contact map.*

Let A and B be two kinds of agent and i and i' be two site names.

Both following properties are equivalent:

- (i) *There exists a repeatable pattern with an agent of kind A connected via its site i to one site of an agent of kind B itself connected to another agent on site i' .*
- (ii) *There exist two agents n and n' respectively of kinds A and B , and a cycle in the graph of the sites of the contact map G_{CM} that passes by the edge $((n, i), (n', i'))$.*

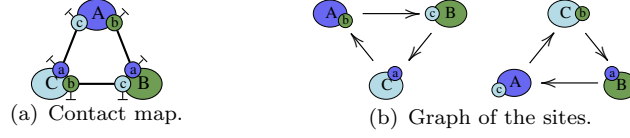


Fig. 10. ABC model. In 10(a), we recall the contact map. In Fig. 10(b), we give the graph of the sites that is associated with this contact map. The nodes of these graphs are the sites of the contact map. There is an oriented edge between a node s and a node t if and only if there is a site connected in the contact map to the site s , in the same kind of protein as the site t but distinct from t .

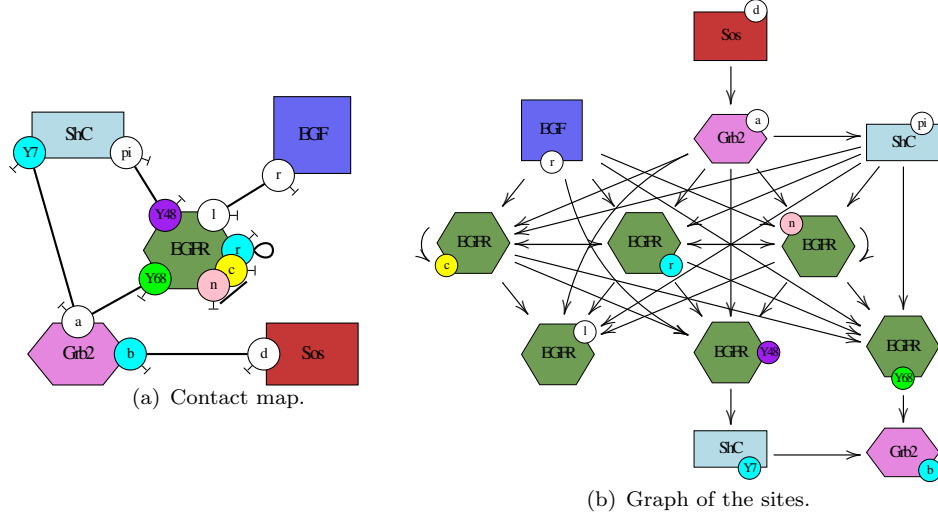


Fig. 11. EGF model. In 11(a), we recall the contact map. In Fig. 11(b), we give the graph of the sites that is associated with this contact map.

Thus, Thm. 4.8 reduces the problem of deciding whether a contact map is compatible with an infinite number of non-isomorphic bio-molecular compounds to the one of computing the strongly connected components of the graph of the sites of this contact map.

Example 4.9 (graph of the sites (ABC model)) In Fig. 10, we compute the graph of the sites for the contact map of the model with three proteins that may form a triangle. It is worth noticing that this graph is made of exactly two non trivial strongly connected components. Each one corresponds to the triangle ABC depending whether it is scanned clockwise or counter-clockwise. Further constraints would be required on the bio-molecular compounds of the models to prove that there is a finite amount of them (the contact map of the model is compatible with an infinite number of them).

Example 4.10 (graph of the sites (EGF model)) In Fig. 11, we compute the graph of the sites for the contact map of the model of the early events in the integration of the epidermic growth factor. It is worth noticing that this graph has only the following non trivial strongly connected component:

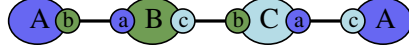


Further constraints are required on the bio-molecular compounds of the models to prove that there is a finite amount of them (the contact map of the model is compatible with an infinite number of different bio-molecular compounds).

4.4 Graph of the potential links

We do not know how to refine the graph of the sites of a given contact map to take into account further constraints about the reachable bio-molecular compounds. We consider in this section another kind of graphs which focuses on the different links in the contact map and that will be easier to refine.

Now we interpret the following repeatable pattern:



as the sequence of (oriented) links from the site b of A to the site a of B , from the site c of B to the site b of C , and from the site a of C to the site c of A .

In the following, we define a graph that stands for all the potential sequences of links that may occur on repeatable patterns that are compatible with a given contact map. We call this graph the graph of the potential links.

Definition 4.11 (graph of the potential links) *Let G_{CM} be a contact map.*

The contact map G_{CM} is associated with a classical graph $(\mathcal{V}, \mathcal{E})$, called the graph of the sites that is defined as follows:

- \mathcal{V} is the subset of the pairs of elements (s, s') of the set $\mathcal{S}_{G_{CM}}$ of the sites of the Σ -graph G_{CM} such that $s' = \mathcal{L}_{G_{CM}}(s)$.
- \mathcal{E} is the subset of the pairs $((s, s'), (s'', s'''))$ of pairs of sites in $\mathcal{V} \times \mathcal{V}$ for which there exists an agent $n \in \mathcal{A}_{G_{CM}}$ and two different site names i and $i' \in \Sigma_{ag-st}(\text{type}_{G_{CM}})$ such that $s' = (n, i)$ and $s'' = (n, i')$.

The condition on the edges of the graph of the potential links ensures that both links may be consecutive in a repeatable pattern.

The following theorem relates the cycles in the graph of the potential links to the existence of repeatable patterns.

Theorem 4.12 *Let G_{CM} be a contact map.*

Let A and B be two kinds of agent and i and i' be two site names.

Both following properties are equivalent:

- (i) *There exists a repeatable pattern with an agent of kind A connected via its site i to the site i' of an agent of kind B ;*
- (ii) *There exist two agents n and n' respectively of kinds A and B , and a cycle in the graph of the potential links of the contact map G_{CM} that passes by the vertex $((n, i), (n', i'))$.*

Thus, Thm. 4.12 reduces the problem of deciding whether a contact map is compatible with an infinite number of non-isomorphic bio-molecular compounds to the one of computing the strongly connected components of the graph of its potential links.

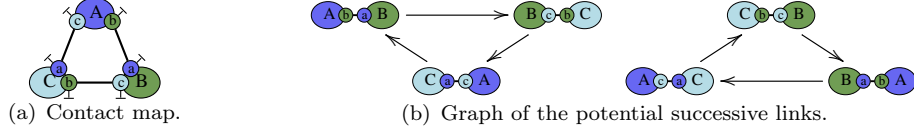


Fig. 12. ABC model. In 12(a), we recall the contact map. In Fig. 12(b), we give the graph of the links that is associated with this contact map. The nodes of these graphs are obtained by orienting the links of the contact map (hence there are two nodes per links).

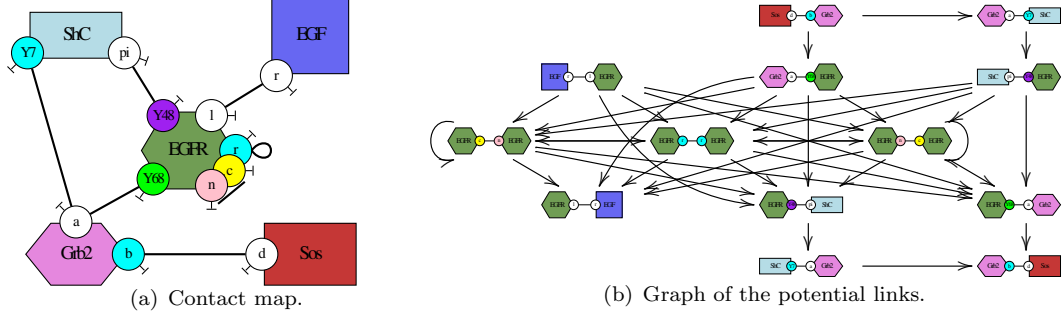
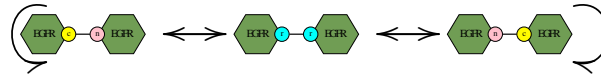


Fig. 13. EGF model. In 13(a), we recall the contact map. In Fig. 13(b), we give the graph of the links that is associated with this contact map. There are two nodes per links, except for the link between the site r of $EGFR$ and itself, for which there is a unique node.

Example 4.13 (graph of the potential links (ABC model)) In Fig. 12, we compute the graph of the potential links for the contact map of the model with three proteins that may form a triangle. It is worth noticing that this graph is made of exactly two non trivial strongly connected components. Each one corresponds to the triangle ABC depending whether it is scanned clockwise or counter-clockwise. Further constraints would be required on the bio-molecular compounds of the models to prove that there is a finite amount of them (the contact map of the model is compatible with an infinite number of them).

Example 4.14 (graph of the potential links (EGF model)) In Fig. 13, we compute the graph of the potential links for the contact map of the model of the early events in the integration of the epidermic growth factor. It is worth noticing that this graph has only one non trivial strongly connected component:



Further constraints are required on the bio-molecular compounds of the models to prove that there is a finite amount of them (the contact map of the model is compatible with an infinite number of them).

5 Taking into account the result of a static analysis

In this section, we explain how to refine the graph of the potential links of a given contact map, in order to take into some additional constraints about the potentially reachable bio-molecular compounds. These constraints may come from a static analysis [13,3] taken as a black box and they may take the form of a set of patterns that shall occur in no reachable bio-molecular compounds. These constraints cannot

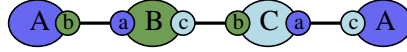
be written in the contact map which can cope only with non relational information about the potential state of sites.

In the case of the model of the early events of the integration of the epidermic growth factor, the analysis that is described in [13] can infer automatically, from the set of rules and the initial state, that none of the following patterns:



is reachable. That is to say that a receptor cannot be bound to two different other instances of receptors.

The analysis that is described in [11] generalises this approach to arbitrary cycles of proteins. In the example of the triangle, it infers (providing that it is a consequence of the rules and of the initial state), that no two *As* may occur in a given connected compound, by proving that the following pattern:



is unreachable.

We refine the statement of Problem 4.2 so as take into account the constraints potentially coming from an external static analysis.

Definition 5.1 (interpretation with a set of forbidden patterns) *The interpretation $\llbracket G_{CM}, \mathcal{P} \rrbracket$ of a contact map G_{CM} with a set of forbidden patterns \mathcal{P} is defined as the set of the \approx -equivalence classes of bio-molecular compound $[G]_{\approx}$ such that there exists a morphism from the site graph G into the contact map G_{CM} and that G contains no occurrence of patterns from the set \mathcal{P} .*

Problem 5.2 *Let G_{CM} be a contact map and \mathcal{P} be a set of patterns.*

We are looking for an automatic procedure to decide whether the set $\llbracket G_{CM}, \mathcal{P} \rrbracket$ is finite, or not.

In the following, we propose a graph structure to answer to Problem 5.2. Our approach is sound but not complete. It can detect and prove that the set of bio-molecular compounds is finite. But when it warns about potential repeatable patterns, it may be a false positive. We do not look for a complete procedure because on the first hand detecting whether or not a pattern is reachable is not decidable in Kappa [17], and on the second hand detecting whether a pattern may occur in a set of bio-molecular compounds that do not contain patterns from a given set is not so easy due to potential overlaps between patterns. Thus we rely on a sound but not complete procedure.

We perform in two steps.

Firstly we label every edge of the graph of the link by a pattern. More precisely, the label of an edge is a chain that is obtained by fusing the second agent of the source node and the first agent of the target node. The chain is oriented. That is to say that two links are identified as respectively the source and the target of the chain of agents.

Example 5.3 (labelled edge) We give as follows an example of a labelled edge in the graph of the links for the EGF model:



The edge is labelled with a chain of three agents. The source of the chain is the link the site c and the site n whereas the target of the chain is the link between the two sites r .

We notice that in our model this chain is indeed unreachable.

The label of an edge must be understood as an explanation about how the link of the source of this edge and of the target of this edge may be connected in reachable bio-molecular compounds. When an edge is labelled with a chain that is unreachable then this edge may be safely discarded.

The longer a chain of agents is, the more constraints it imposes. The second step consists in combining consecutive edges, in order to extend their labels into longer chains. It is worth noticing that given a node in the graph of the links, the target of the label of every incoming edge and the source of the label of every outgoing edge is indeed the pattern that is labelling the node. We will keep this property as a structural invariant of the graph. This structural invariant will be preserved when combining the edges of the graph. Given a node in the graph of the links, an incoming edge and an outgoing edge may be composed to form an edge from the source of the incoming edge to the target of the outgoing edge. The label of this new edge is obtained by using the label of the outgoing edge and the label of the incoming edge by identifying the target of the former with the source of the later one. This way, we form a chain of agents from the source of the label of the incoming edge to the target of the label of the outgoing edge, which ensures that the structural invariant of the graph of the links is preserved.

Now we can define precisely the second step: the second step consists in selecting a node and replacing a given incoming edge with the set of the edges that is obtained by combining this edge with every outgoing edge. This transformation preserves the structural invariant and increases the length of the chains of agent that are used as labels of the edges in the graph.

Example 5.4 (graph refinement (the model of the triangle))

The following theorem states the soundness of our approach.

Theorem 5.5 Let G_{CM} be a contact map. Let \mathcal{P} be a set of patterns. Let G be a refinement of the graph of the potential links of the contact map, according to the set of patterns \mathcal{P} . We assume that there exists a bio-molecular compound S such that $[S]_{\approx} \in \llbracket G_{CM}, \mathcal{P} \rrbracket$ that contains a repeatable pattern P such that no iteration of the pattern P contains an occurrence of the pattern in \mathcal{P} .

Then, for every repetition Q of the pattern P , for every two agent identifiers n, n' and every two site names i, i' such that $\mathcal{L}_Q(n, i) = (n', i')$, there exists two agent identifiers n'', n''' such that $\text{type}_P(n) = \text{type}_{G_{CM}}(n'')$, $\text{type}_P(n') = \text{type}_{G_{CM}}(n''')$ and there exists a cycle in the graph G passing by the vertex $((n'', i), (n''', i'))$.

Intuitively, if an iteration of a pattern P contains a forbidden pattern, then, the

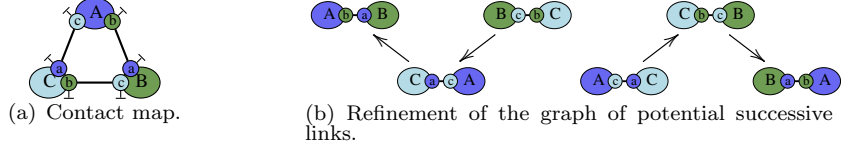


Fig. 14. ABC model. In 14(a), we recall the contact map. In Fig. 14(b), we refine the graph of the potential successive links to take into account the constraints that two instances of A may not occur in a same connected component.

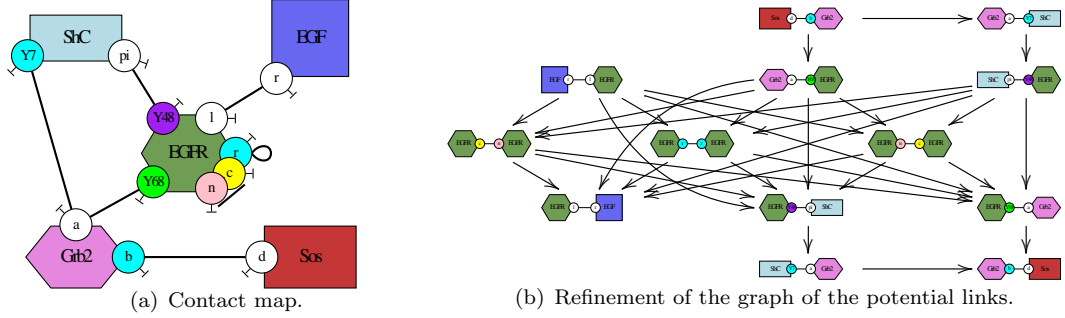


Fig. 15. EGF model. In 15(a), we recall the contact map. In Fig. 15(b), we refine the graph of the links that is associated with this contact map, by taking into account that a given receptor cannot be bound simultaneously to two different other receptors.

pattern P cannot be repeated an unbounded number of times in a reachable bio-molecular compound. The theorem states that vertices that belong to non trivial connected components in a refined graph is a super-set of the bonds that may occur in an iteration of the patterns which are compatible with the contact map and with the black-listed patterns. If the refined graph is acyclic, then the set of reachable bio-molecular compound is necessarily finite.

Example 5.6 (refined graph of the links (model with the triangle)) In Fig. 14, we refine the graph of the potential links for the contact map of the model ABC by taking into account that any pattern with several instances of the protein A is unreachable. This is achieved by refining the label of the edge between the site b of A and the site a of B until reaching a black-listed pattern.

The graph (see Fig. 14(b)) is acyclic which proves that the set of bio-molecular compounds is finite in this model.

Example 5.7 (refined graph of the potential links (EGF model)) In Fig. 15, we refine the graph of the potential links for the contact map of the model of the early events in the integration of the epidermic growth factor, by taking into account the fact that a given receptor cannot be bound simultaneously to several other receptors. Indeed every edge of the strongly connected component is initially labelled with a black-listed pattern, thus they can be discarded without iteratively refining the graph. The graph that is obtained (see Fig. 15(b)) is acyclic, which proves that the model involves only a finite set of bio-molecular compounds.

6 Conclusion

In this paper, we have provided some decision procedures to detect whether or not the set of bio-molecular compounds of rule-based models, such as the ones that are written in Kappa [9] or in BNGL [2], is finite or not. Our approach is mainly based on top of the contact map, a Σ -graph which summarizes the potential links between the binding sites of proteins. The contact map is translated into a classical graph which encodes either the potential succession of sites, or the potential succession of links in bio-molecular compounds. Non trivial connected components in this graph correspond to patterns that may be repeated an arbitrary number of times in reachable bio-molecular compounds. They can be detected using classical depth-first exploration without having to enumerate every elementary cycle [19]. The graph that stands for the potential succession of links in bio-molecular compounds can be refined in order to handle with some additional constraints computed by reachability analysis [7,13,3].

Our approach has been partially integrated in the static analyzer KaSa [3]. More precisely, the construction of the graph of the potentially successive links has been implemented as well as the reduction with the static analysis that is described in [13]. This way, the analyzer can cope accurately with the constraints involving potential cycles of two proteins. We plan to implement the generalisation that has been proposed in [11], that can handle precisely with models that can generate cyclic structures without creating arbitrary long bio-molecular compounds.

As future works, we plan to use weakly relational domains [18] to abstract more precisely the chains of proteins that may embed in the reachable bio-molecular compounds. This analysis will allow to analyse accurately the rules that behave differently when applied in a uni-molecular or in a bi-molecular context.

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