

# Functional Enrichment Analysis

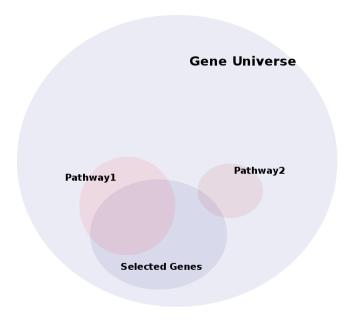
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## **Explorative Functional Analysis**

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- Challenge
- Functional databases
- Functional enrichment
  - Overrepresentation analysis
  - Gene set enrichment analysis
- Tools



# Data Interpretation - Challenge

### **Differential Gene Expression**

- Hundreds/thousands of candidate genes
- Gene-by-gene interpretation difficult
- Linking different experiments
- Filter results (Venn)
- Identify:
  - key molecules (TFs, miRNAs, common effectors, master regulators)
  - Enriched Biological Processes/Pathways
  - Networks (links across candidates)

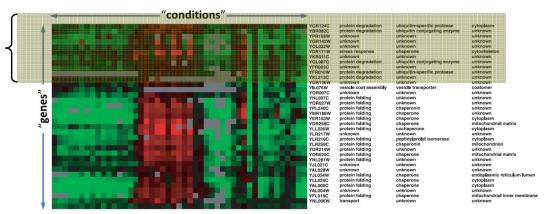




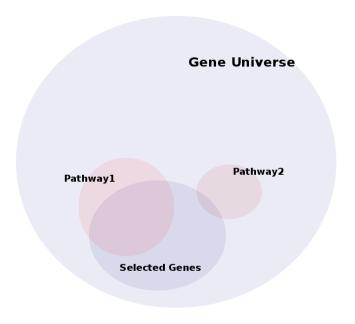
# Filtering list of DEGs

- FDR cutoff  $< 0.05 0.1 \rightarrow \text{p-value cutoff} < 0.01$
- Fold change cutoff (e.g., > 2 fold change)
- Rank list (top 10% based on p-value / fold change)
- Clustered genes with similar expression patterns
- Candidate gene list

Filtered/selected gene set







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# Which functional databases can be interrogated?

Gene Ontology



http://amigo1.geneontology.org/cgi-bin/amigo/go.cgi

Pathways









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Protein class





These databases are typically constructed based on protein-protein interaction experiments, signaling pathway disruption experiment, literature screening (and combinations of the above)



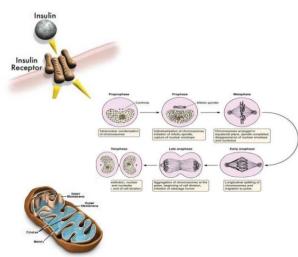
# Gene Ontology (GO) terms

- Three ontology domains:
  - 1. Molecular function (MF): basic activity or task e.g. catalytic activity, calcium ion binding
  - **2.** Biological process (BP): broad objective orgoal e.g. signal transduction, immune response
  - **3.** Cellular component (CC): location or complex *e.g. nucleus, mitochondrion*
- Genes can have multiple annotations:

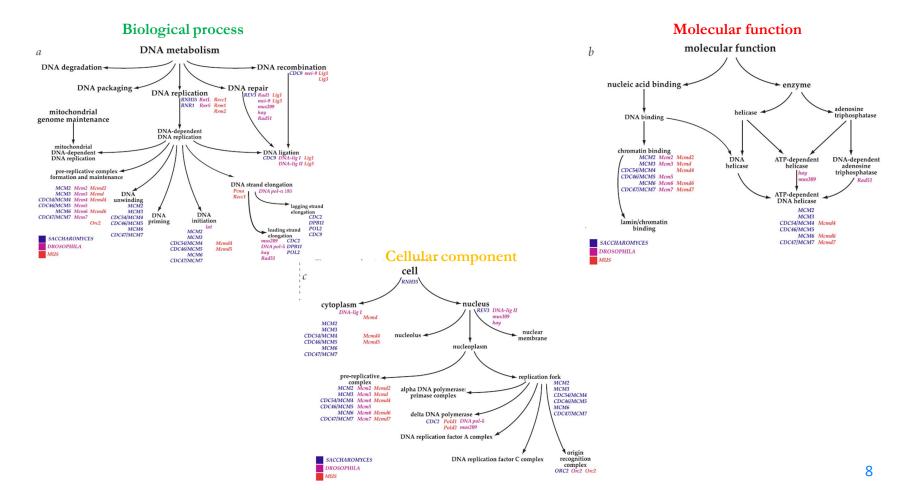
#### Cytochrome gene

- Molecular function (MF): oxidoreductase activity
- Biological process (BP): oxidative phosphorylation, induction of cell death
- Cellular component (CC): mitochondrial matrix, mitochondrial inner membrane





### GO terms are hierarchical

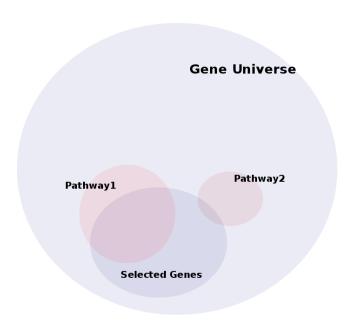




### Functional databases: limitations

- Accurate functional annotation is only available for model organisms
- Functional terms are abundant and highly correlated
  - Multiple testing of interdependent terms (FDR)





### **Explorative Functional Analysis**

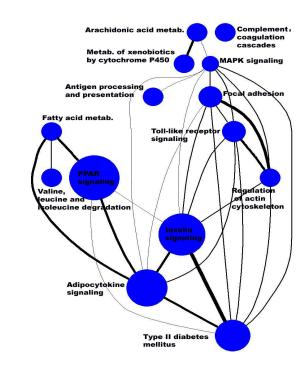
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# Overrepresentation analysis

### Case study:

- With a meta analysis Rasche et al. identified 213 genes associated with Type II Diabetes
- For a given KEGG pathway, they counted how many of these 213 genes are members of this pathway
- If this number is more than expected by chance, then this pathway could potentially be relevant to Type II Diabetes
  - Fisher's exact test



BMC Genomics. 2008 Jun 30;9:310. doi: 10.1186/1471-2164-9-310.

Meta-analysis approach identifies candidate genes and associated molecular networks for type-2 diabetes mellitus.

Rasche A<sup>1</sup>, Al-Hasani H, Herwig R.



### Fisher's exact test

- Statistical test used to determine if there are non-random associations between two categorical variables.
- Fisher devised the test following a comment from a colleague, who claimed to be able to detect whether the tea or the milk was added first to her cup.



Contingency table:	Present in seens (Not present in 15th		
	7. 18. 18. 18. 18. 18. 18. 18. 18. 18. 18	Not present in	, ota/
Member of KEGG pathway	107	637	780
Not member of KEGG pathway	452	8673	9125
Total	559	9346	9905

- Free tools
  - WebGestalt, R-packages, Cytoscape
- Commercial tools
  - MetaCore/GeneGo, Ingenuity (IPA)

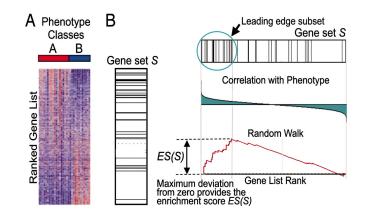
Observed: 107

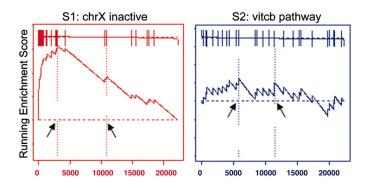
Expected: 
$$\frac{780 * 559}{9905} = 44$$

Enrichment: **2.43x**, p-val < 0.00001



- Input: Unfiltered genes ordered in a ranked list L, according to their fold change or p-value
- **Goal**: Determine whether members of a gene set S are randomly distributed throughout the list L or tend to occur toward the top (or bottom) of L, as measured by Enrichment score (ES)
- **Enrichment score (ES):** The degree to which a set S is overrepresented at the extremes (top or bottom) of the entire ranked list L
- Random walk down the list L, increasing the running-sum when encounter a gene in S and decreasing it when encounter a gene not in S.
- ES is the max deviation from zero encountered in the random walk (Kolmogorov–Smirnov test)

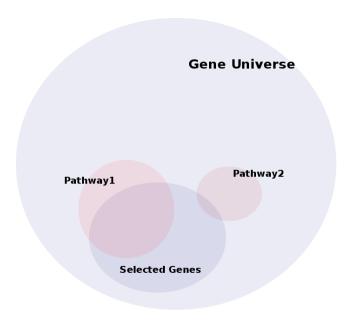




# Gene set enrichment analysis (GSEA): advantages

- Does not require a priori selection (e.g., p-value thresholds, clusters)
  - Not risking to generate too short lists
  - Meta-analyses are not limited by overlapping factors
- Aggregated small effects better captured
  - (would potentially miss with ORA)





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# Tools for functional enrichment analysis

- Free tools
  - WebGestalt (http://www.webgestalt.org/)
  - Panther (http://pantherdb.org/)
  - David (https://david.ncifcrf.gov/)
  - R Packages (topGO, GSEABase, clusterProfiler, ...)
  - GSEA (http://software.broadinstitute.org/gsea/index.jsp)
  - Cytoscape
- Commercial tools
  - MetaCore/GeneGO (https://portal.genego.com/)
  - Ingenuity Pathway Analysis (https://www.qiagenbioinformatics.com/products/ingenuity-pathway-analysis/)

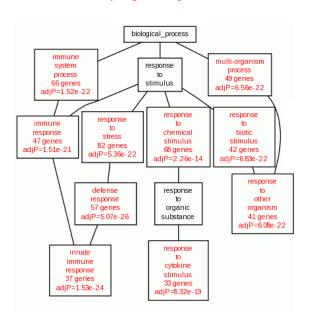
# GO Enrichment - Example from FGCZ Reports

GO-Term	P-Value	Ratio	
immune response	2.77e-14	50/289	289 Genes in the Gene Universe a annotated with 'Immune Respons
T-helper 17 cell differentiati	5.45e-05	3/3	50 Genes in Candidates List are
positive regulation of natural	4.24e-06	7/16	belonging to 'Immune Response'
regulation of immune response.	7.26e-07	16/81	
inflammatory response	3.11e-09	39/252	
cellular defense response	6.71e-07	13/49	
G-protein coupled receptor sig	7.12e-07	28/161	
cytokine-mediated signaling pa	2.39e-06	23/196	
.chemokine-mediated signaling p	1.66e-05	8/22	
negative regulation of viral g	3.33e-06	8/30	
positive regulation of natural	2.58e-05	4/5	
response to virus	5.88e-05	13/96	
defense response to virus	4.86e-08	20/120	
cytolysis	8.07e-05	5/13	
positive regulation of cell ad	9.08e-05	8/29	

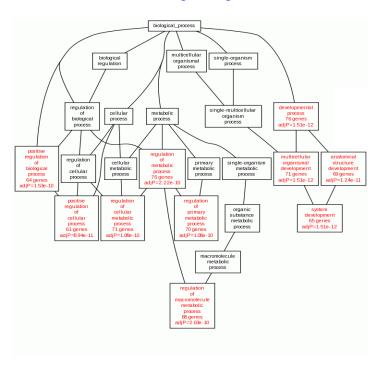
## GO Enrichment - Example with WebGestalt

http://webgestalt.org

#### Up regulated genes



#### **Down regulated genes**





### Pathway Enrichment - Example with Metacore

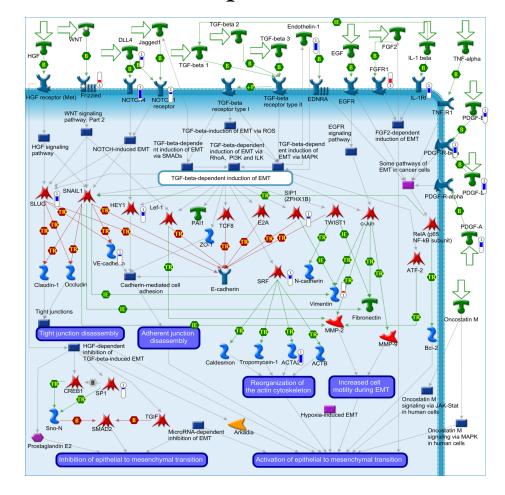
- https://portal.genego.com, commercial (FGCZ has a license)
- Upload a file with list of selected genes
- One click enrichment analysis (eg. pathway enrichment analysis)

Maps	0	2	4	6	8	-log(pValue)	pValue +	FDR	Ratio
Cell adhesion_Endothelial cell contacts by junctional mechanisms		:	:	:			4.742e-10	3.841e-7	13/26
Cell adhesion_Chemokines and adhesion		:	:	:			2.305e-9	6.466e-7	24/100
Development_Regulation of epithelial-to-mesenchymal transition (EMT)		-:					2.395e-9	6.466e-7	19/64
$\underline{\text{Main pathways of Schwann cells transformation in neurofibromatosis type 1}}$		-:	-:				1.307e-7	1.825e-5	19/80
Muscle contraction_Regulation of eNOS activity in endothelial cells		-:	-:				1.333e-7	1.825e-5	17/65
Development_Oligodendrocyte differentiation from adult stem cells		-:	-:				1.352e-7	1.825e-5	15/51
Development_Regulation of endothelial progenitor cell differentiation from adult stem cells		-:	-:				2.332e-7	2.699e-5	16/60
Cytoskeleton remodeling Cytoskeleton remodeling		-:	-:				3.887e-7	3.936e-5	21/102
Cell adhesion_Endothelial cell contacts by non-junctional mechanisms		-:-	-:				4.404e-7	3.964e-5	10/24
Role of red blood cell adhesion to endothelium in vaso-occlusion in Sickle cell disease			:				7.603e-7	5.174e-5	12/37

• By clicking on the pathway name, one can get a full picture of the genes involved in that pathway, with genes from the uploaded list specifically marked (example on the next slide: Development regulation of EMT)



# Pathway Enrichment - Example with Metacore



# **Summary**

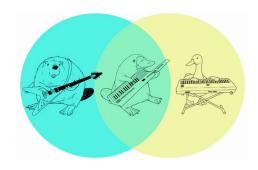
- Besides p value and/or fold change thresholds, clustering and venn diagram analysis can be used to filter DEGs
- GO terms are highly correlated, distributed across > 1,000 categories
- Functional analysis is only reliable for organisms with good annotation
  - GSEA of the full set of genes is more robust and versatile than functional overrepresentation analysis in selected/filtered set of genes
- Input for overrepresentation analysis
  - Filtered gene list based on p-value and/or fold change threshold
- Input for gene set enrichment analysis
  - Unfiltered gene list ranked based on p-value /or fold change



# **Additional Slides**

# Filtering list of DEGs: Venn diagram

- Idea
  - consider gene lists as sets and calculate intersection  $(A \cap B)$ , union  $(A \cup B)$  and complement  $(A \setminus B, B \setminus A)$
- Typical cases
  - Compare gene list to
    - data with similar context
    - public data
    - data from a different platform



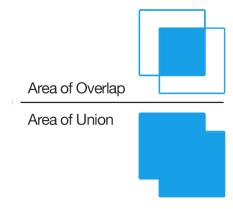
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# Jaccard index

- Jaccard index is a measure of the similarity between sets
- Given set A and set B, the Jaccard index is given by:

$$\frac{|A\cap B|}{|A\cup B|}$$

• Takes the values of 1 when sets are identical, and 0 when there is no overlap



# **FGCZ - Interactive Shiny Apps**

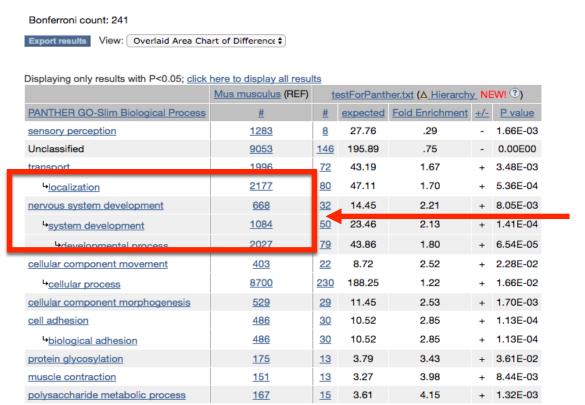
### http://fgcz-shiny.uzh.ch

ShinyApp	Description
Explore RNA-seq counts	Perform clustering and MDS plots; identify effect sizes and potential outliers
Explore RNA-seq differential expression	Filter and visualize your differential expression result; inspect individual genes; identify functional categories associated with gene lists
<u>pcaExplorer</u>	Visualization of RNA-seq data based on Pricipal Component analysis using pcaExplorer package
<u>Heatmap</u>	create an interactive gene expression heatmap based on a differential expression result generated by SUSHI
2-way-Venn	compare two sets of differential expression results to create a VennDiagram
<u>3-way-Venn</u>	compare three sets of differential expression results to create a VennDiagram
Correlation	interactive scatterplot for two sets of differential expression results

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# Which functional categories are really enriched for my set of genes?



Note hierarchy to avoid redundancy

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f. g. c. z. 10 0

# **Promoter Analysis**

### • Idea

- Identification and quantification of TFBS in promoter region of candidate genes
- Compare results with background model (typically 200-500 housekeeping or randomly selected genes)
- Compute ranking of TFs based on TFBS enrichment

### Typical case

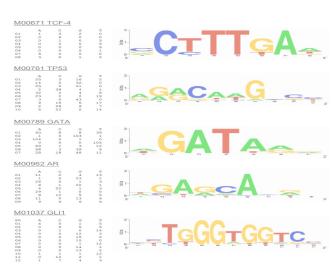
 Identify master regulatory transcription factors (potential marker discovery...)

#### Software tool

free tools (eg. Clover)
GeneXplain (uses TRANSFAC®-,
TRANSPATH®-DB from BIOBASE)

### Analog type of analysis

Compute a ranking of microRNAs (based upon target gene expression; e.g. R-package 'MiRAGE')



2. Key Molecule Identification