Choosing Genomics Tools

February, 2023

Table of Contents

# About this Course

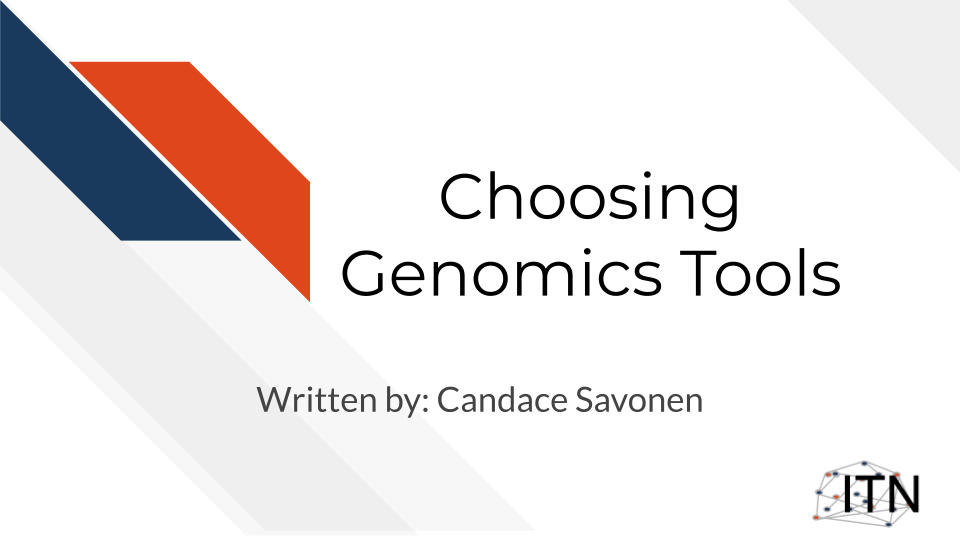
This course is part of a series of courses for the [Informatics Technology for Cancer Research (ITCR)](https://itcr.cancer.gov/) called the Informatics Technology for Cancer Research Education Resource. This material was created by the ITCR Training Network (ITN) which is a collaborative effort of researchers around the United States to support cancer informatics and data science training through resources, technology, and events. This initiative is funded by the following grant: [National Cancer Institute (NCI)](https://www.cancer.gov/) UE5 CA254170. Our courses feature tools developed by ITCR Investigators and make it easier for principal investigators, scientists, and analysts to integrate cancer informatics into their workflows. Please see our website at <www.itcrtraining.org> for more information.

## 0.1 Available course formats

This course is available in multiple formats which allows you to take it in the way that best suites your needs. You can take it for certificate which can be for free or fee.

* The material for this course can be viewed without login requirement on this [Bookdown website](https://hutchdatascience.org/Choosing_Genomics_Tools/). This format might be most appropriate for you if you rely on screen-reader technology.
* This course can be taken for free certification through Leanpub.
* This course can be taken on [Coursera for certification here](https://www.coursera.org/learn/) (but it is not available for free on Coursera).
* Our courses are open source, you can find the [source material for this course on GitHub](https://github.com/jhudsl/Choosing_Genomics_Tools).

# 1 Introduction

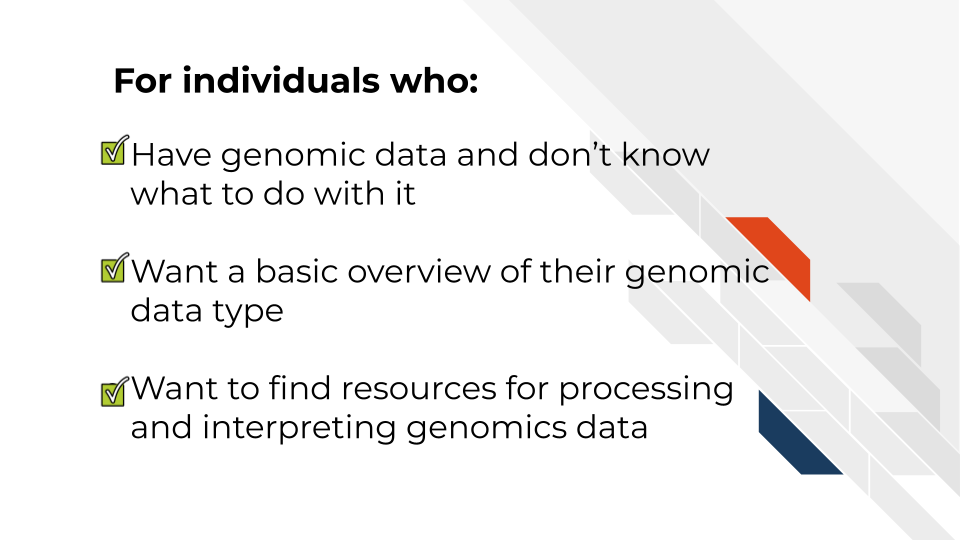


## 1.1 Target Audience

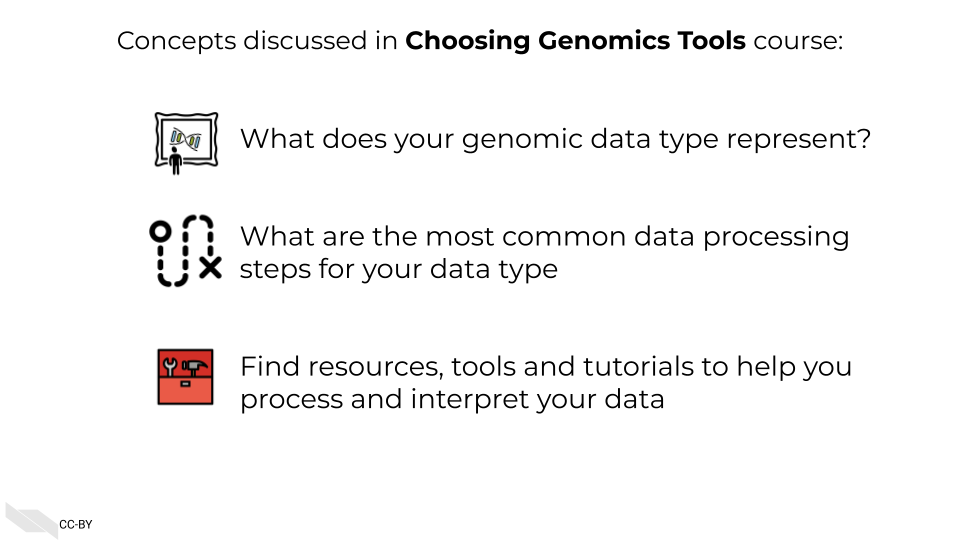
The course is intended for students in the biomedical sciences and researchers who have been given data and don’t know what to do with it or would like an overview of the different genomic data types that are out there.

*This course is written for individuals who:*

* Have genomic data and don’t know what to do with it.
* Want a basic overview of genomic data types.
* Want to find resources for processing and interpreting genomics data.



## 1.2 Topics covered:

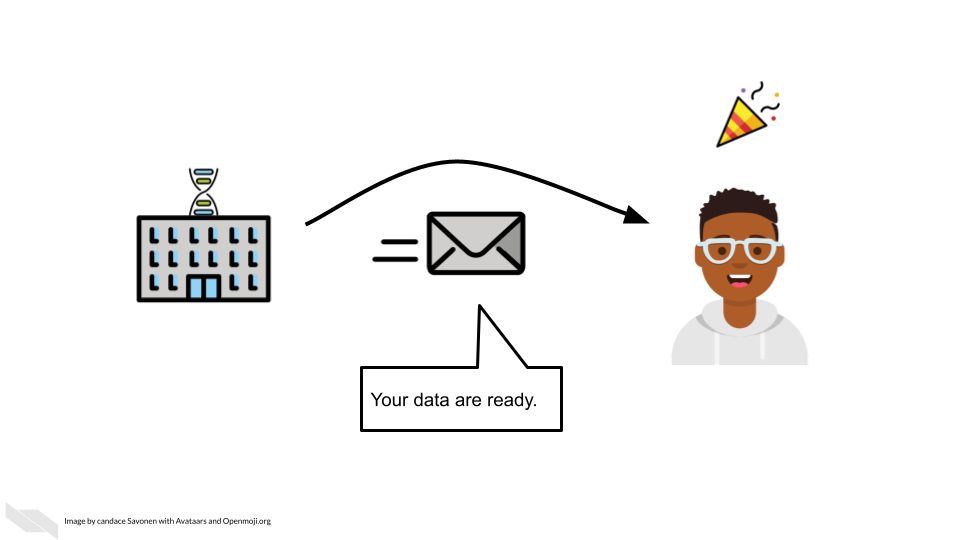


## 1.3 Motivation

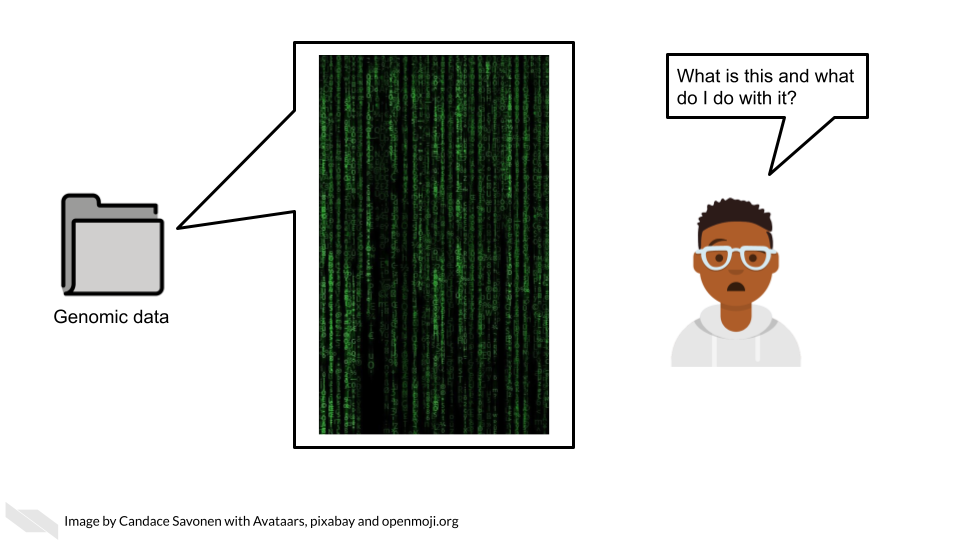
Cancer datasets are plentiful, complicated, and hold untold amounts of information regarding cancer biology. Cancer researchers are working to apply their expertise to the analysis of these vast amounts of data but training opportunities to properly equip them in these efforts can be sparse. This includes training in reproducible data analysis methods.

Often students and researchers need to utilize genomic data to reach the next steps of their research but may not have formal training in computational methods or the basics of the genomic data they are attempting to utilize.

Often researchers receive their genomic data processed from another lab or institution, and although they are excited to gain insights from it to inform the next steps of their research, they may not have a practical understanding of how the data they have received came to be or what needs to be done with it.

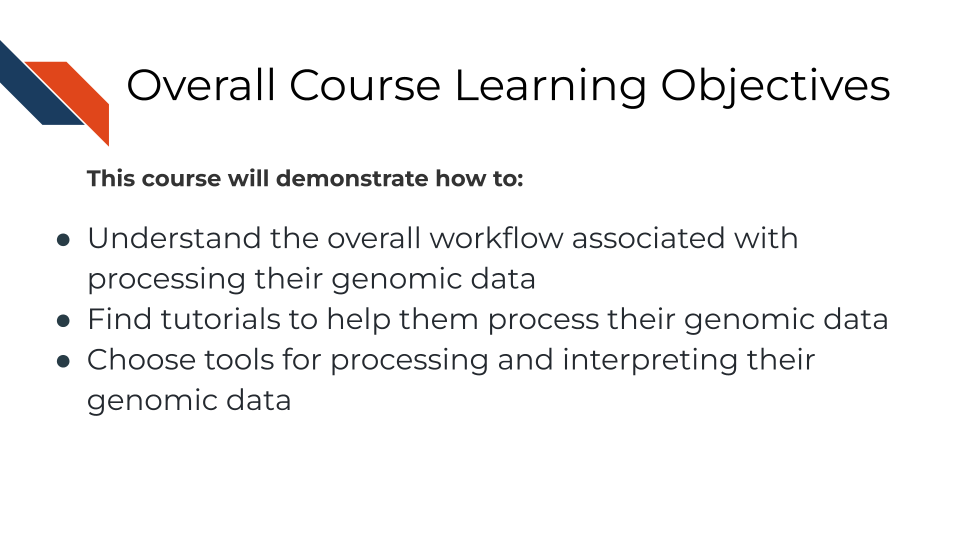


As an example, data file formats may not have been covered in their training, and the data they received seems unintelligible and not as straightforward as they hoped.



This course attempts to give this researcher the basic bearings and resources regarding their data, in hopes that they will be equipped and informed about how to obtain the insights for their researcher they originally aimed to find.

## 1.4 Curriculum



**Goal of this course:**  
Equip learners with tutorials and resources so they can understand and interpret their genomic data in a way that helps them meet their goals and handle the data properly. This includes helping learners formulate questions they will need to ask others about their data

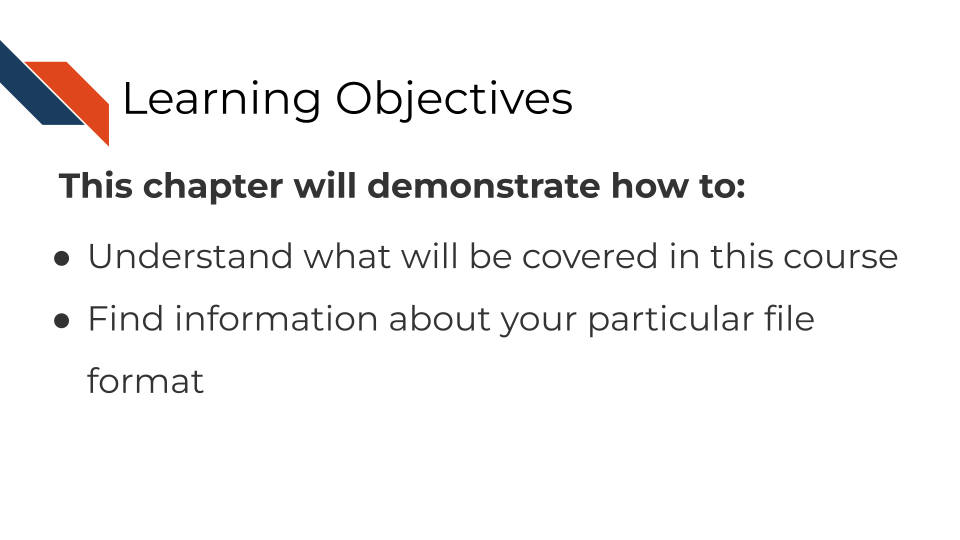
**What is not the goal**  
Teach learners about choosing parameters or about the ins and outs of every genomic tool they might be interested in. This course is meant to connect people to other resources that will help them with the specifics of their genomic data and help learners have more efficient and fruitful discussions about their data with bioinformatic experts.

## 1.5 How to use the course

This course is designed to be a jumping off point to more specific resources based on a genomic data type the learner has in mind (or currently on their computer). We encourage learners to follow links to resources we provide and feel free to jump around to chapters that are most useful for them.

# 2 A Very General Genomics Overview

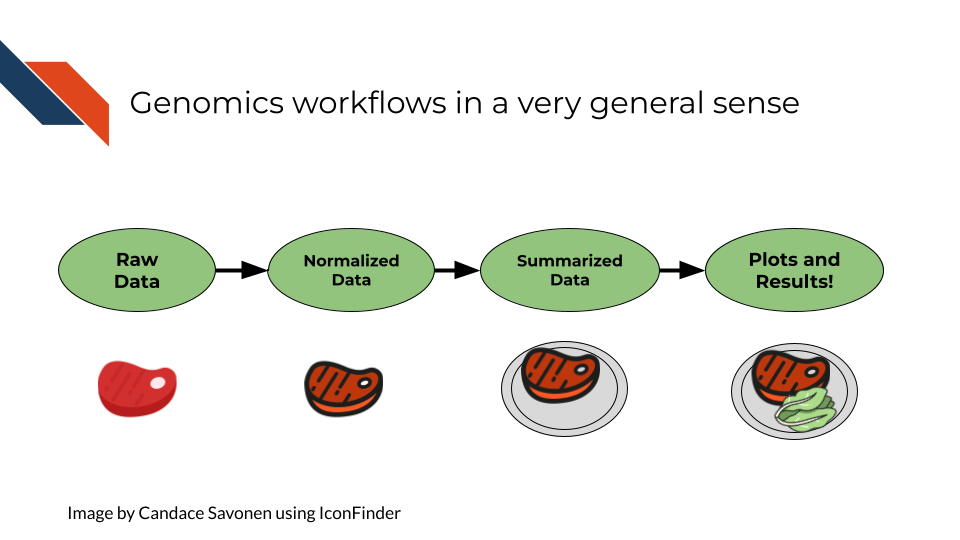
## 2.1 Learning Objectives



In this chapter we are going to cover sequencing and microarray workflows at a very general high level overview to give you a first orientation. As we dive into specific data types and experiments, we will get into more specifics. Here we will cover the most common file formats. If you have a file format you are dealing with that you don’t see listed here, it may be specific to your data type and we will discuss that more in that data type’s respective chapter. We still suggest you go through this chapter to give you a basic understanding of commonalities of all genomic data types and workflows

### 2.1.1 What do genomics workflows look like?

In the most general sense, all genomics data when originally collected is raw, it needs to undergo processing to be normalized and ready to use. Then normalized data is generally summarized in a way that is ready for it to be further consumed. Lastly, this summarized data is what can be used to make inferences and create plots and results tables.



### 2.1.2 Basic file formats

Before we get into bioinformatic file types, we should establish some general file types that you likely have already worked with on your computer. These file types are used in all kinds of applications and not specific to bioinformatics.

#### 2.1.2.1 TXT - Text

A text file is a very basic file format that contains text!

#### 2.1.2.2 TSV - Tab Separated Values

Tab separated values file is a text file is good for storing a data table. It has rows and columns where each value is separated by (you guessed it), *tabs*. Most commonly, if your genomics data has been provided to you in a TSV or CSV file, it has been processed and summarized! It will be your job to know *how* it was processed and summarized

Here the literal ⇥ represents tabs which often may show up invisible in your text editor’s preference settings.

gene\_id⇥sample\_1⇥sample\_2  
gene\_a⇥12⇥15,  
gene\_b⇥13⇥14

#### 2.1.2.3 CSV - Comma Separated Values

A comma separated values file is list just like a TSV file but instead of values being separated by tabs it is separated by… (you guessed it), *commas*!

In its raw form, a CSV file might look like our example below (but if you open it with a program for spreadsheets, like Excel or Googlesheets, it will look like a table)

gene\_id, sample\_1, sample\_2,  
gene\_a, 12, 15,  
gene\_b, 13, 14

### 2.1.3 Sequencing file formats

#### 2.1.3.1 SAM - Sequence Alignment Map

SAM Files are text based files that have sequence information. It generally has not been quantified or mapped. It is the reads in their raw form. [For more about SAM files](https://samtools.github.io/hts-specs/SAMv1.pdf).

#### 2.1.3.2 BAM - Binary Alignment Map

BAM files are like SAM files but are compressed (made to take up less space on your computer). This means if you double click on a BAM file to look at it, it will look jumbled and unintelligible. You will need to convert it to a SAM file if you want to see it yourself (but this isn’t necessary necessarily).

#### 2.1.3.3 FASTA - “fast A”

Fasta files are sequence files that can be either nucleotide or amino acid sequences. They look something like this (the example below illustrating an amino acid sequence):

>SEQ\_ID  
GATTTGGGGTTCAAAGCAGTATCGATCAAATAGTAAATCCATTTGTTCAACTCACAGTTT

For [more about fasta files](https://en.wikipedia.org/wiki/FASTA_format).

#### 2.1.3.4 FASTQ - “Fast q”

A Fastq file is like a Fasta file except that it also contains information about the **Q**uality of the read. By quality, we mean, how sure was the sequencing machine that the nucleotide or amino acid called was indeed called correctly?

@SEQ\_ID  
GATTTGGGGTTCAAAGCAGTATCGATCAAATAGTAAATCCATTTGTTCAACTCACAGTTT  
+  
!''\*((((\*\*\*+))%%%++)(%%%%).1\*\*\*-+\*''))\*\*55CCF>>>>>>CCCCCCC65

For [more about fastq files](https://en.wikipedia.org/wiki/FASTQ_format).

Later in this course we will discuss the importance of examining the quality of your sequencing data and how to do that. If you received your data from a bioinformatics core it is possible that they’ve already done this quality analysis for you.

*Sequencing data that is not of high enough quality should not be trusted!* It may need to be re-run entirely or may need extra processing (trimming) in order to make it more trustworthy. We will discuss this more in later chapters.

#### 2.1.3.5 BCL - binary base call (BCL) sequence file format

This type of sequence file is specific to Illumina data. In most cases, you will simply want to convert it to Fastq files for use with non-Illumina programs.

[More about BCL to Fastq conversion](https://medium.com/@marija190396/bcl-to-fastq-conversion-e289852823d0).

#### 2.1.3.6 VCF - Variant Call Format

VCF files are further processed form of data than the sequence files we discussed above. VCF files are specially for storing only where a particular sample’s sequences differ or are *variant* from the reference genome or each other.

This will only be pertinent to you if you care about DNA variants. We will discuss this in the DNA seq chapter.

For [more on VCF files](https://en.wikipedia.org/wiki/Variant_Call_Format).

#### 2.1.3.7 MAF - Mutation Annotation Format

MAF files are aggregated versions of VCF files. So for a group of samples for which each has a VCF file, your entire group of samples’ variants will be summarized in the form of a MAF file.

For [more on MAF files](https://docs.gdc.cancer.gov/Data/File_Formats/MAF_Format/#:~:text=Mutation%20Annotation%20Format%20(MAF)%20is,(or%20open%2Daccess).).

### 2.1.4 Microarray file formats

#### 2.1.4.1 IDAT - intensity data file

This is an Illumina microarray specific file that contains the chip image intensity information for each location on the microarray. It is a binary file, which means it will not be readable by double clicking and attempting to open the file directly.

Currently, Illumina appears to suggest directly converting IDAT files into a GTC format. We advise looking into [this package to help you do that](https://github.com/freeseek/gtc2vcf).

[For more on IDAT files](https://www.illumina.com/content/dam/illumina-marketing/documents/products/technotes/technote_array_analysis_workflows.pdf).

#### 2.1.4.2 DAT - data file

This is an Affymetrix’ microarray specific file parallel to the IDAT file in that it contains the image intensity information for each location on the microarray. It’s stored as pixels.

[For more on DAT files](https://www.affymetrix.com/support/developer/powertools/changelog/gcos-agcc/dat.html).

#### 2.1.4.3 CEL

This is an Affymetrix microarray specific file that is made from a DAT file but translated into numeric values. It is not normalized yet but can be normalized into a CHP file.

[For more on CEL files](https://www.affymetrix.com/support/developer/powertools/changelog/gcos-agcc/cel.html)

#### 2.1.4.4 CHP

CHP files contain the gene-level and normalized data from an Affymetrix array chip. CHP files are obtained by normalizing and processing CEL files.

[For more about CHP files](https://www.affymetrix.com/support/developer/powertools/changelog/gcos-agcc/chp-xda.html).

## 2.2 General informatics files

At various points in your genomics workflows, you may need to use other types of files to help you annotate your data. We’ll also discuss some of these common files that you may encounter:

#### 2.2.0.1 BED - Browser Extensible Data

A BED file is a text file that has coordinates to genomic regions. THe other columns that accompany the genomic coordinates are variable depending on the context. But every BED file contains the chrom, chromStart and chromEnd columns to start.

A BED file might look like this:

chrom chromStart chromEnd other\_optional\_columns  
chr1 0 1000 good  
chr2 100 3000 bad

For [more on BED files](https://en.wikipedia.org/wiki/BED_(file_format)).

#### 2.2.0.2 GFF/GTF General Feature Format/Gene Transfer Format

A GFF file is a tab delimited file that contains information about genomic features. These types of files are available from databases and what you can use to annotate your data.

You may see there are GFF2, GFF3, and GTF files. These only refer to different versions and variations. They generally have the same information. In general, GFF2 is being phased out so using GFF3 is generally a better bet unless the program or package you are using specifies it needs an older GFF2 version.

A GFF file may look like this (borrowed example from Ensembl):

1 transcribed\_unprocessed\_pseudogene gene 11869 14409 . + . gene\_id "ENSG00000223972"; gene\_name "DDX11L1"; gene\_source "havana"; gene\_biotype "transcribed\_unprocessed\_pseudogene";

Note that it will be useful for annotating genes and what we know about them.

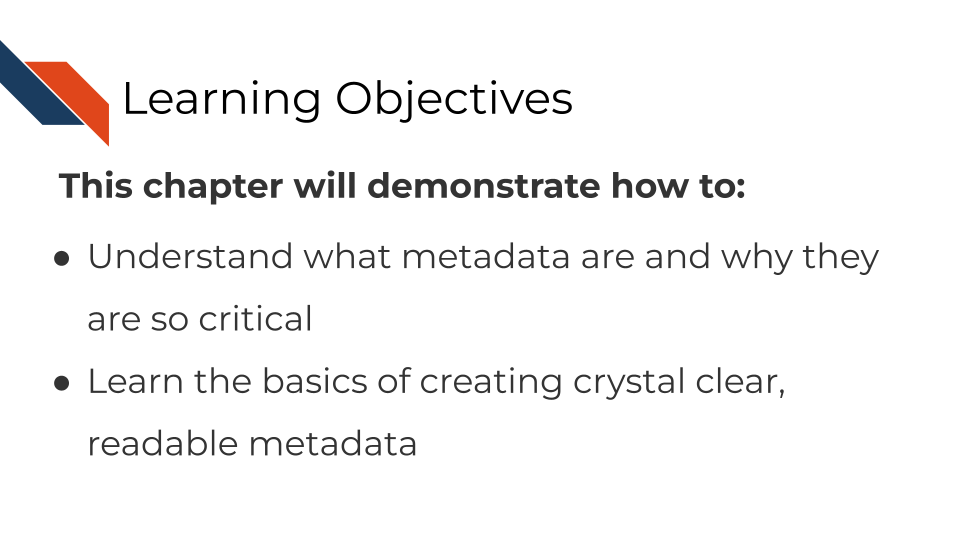
For [more about GTF and GFF files](https://useast.ensembl.org/info/website/upload/gff.html).

### 2.2.1 Other files

\* If you didn’t see a file type listed you are looking for, take a look at this [list by the BROAD](https://software.broadinstitute.org/cancer/software/gsea/wiki/index.php/Data_formats). Or, it may be covered in the data type specific chapters.

# 3 What are Metadata?

## 3.1 Learning Objectives

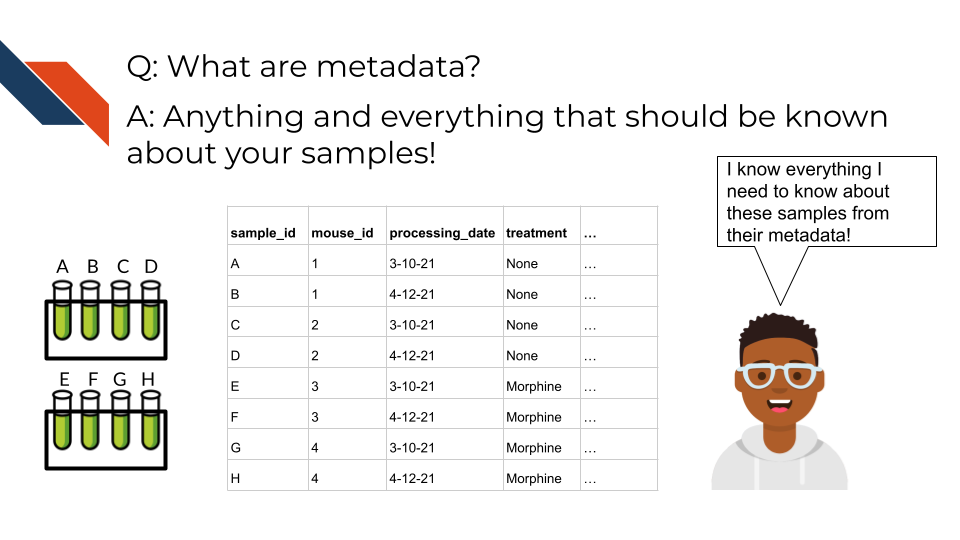


## 3.2 What are metadata?

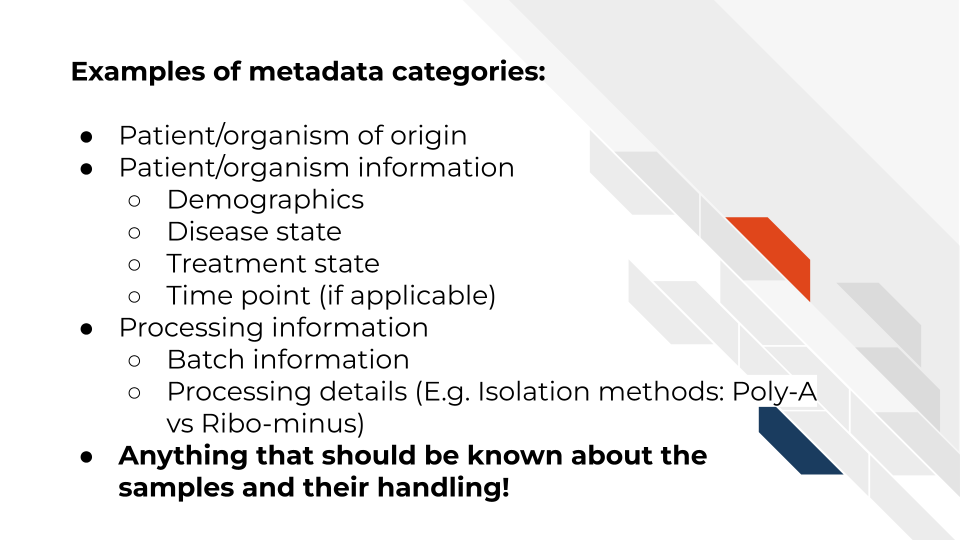
Metadata are critically important descriptive information about your data.

**Without metadata, the data themselves are useless or at best vastly limited.**

Metadata describe how your data came to be, what organism or patient the data are from and include any and every relevant piece of information about the samples in your data set.



Metadata includes but isn’t limited to, the following example categories:



At this time it’s important to note that if you work with human data or samples, your metadata will likely contain personal identifiable information (PII) and protected health information (PHI). It’s critical that you protect this information! For more details on this, we encourage you to see our [course about data management](https://jhudatascience.org/Ethical_Data_Handling_for_Cancer_Research/data-privacy.html).

## 3.3 How to create metadata?

Where do these metadata come from? The notes and experimental design from anyone who played a part in collecting or processing the data and its original samples. If this includes you (meaning you have collected data and need to create metadata) let’s discuss how metadata can be made in the most useful and reproducible manner.

### 3.3.1 The goals in creating your metadata:

#### 3.3.1.1 Goal A: Make it *crystal clear* and *easily readable* by both humans and computers!

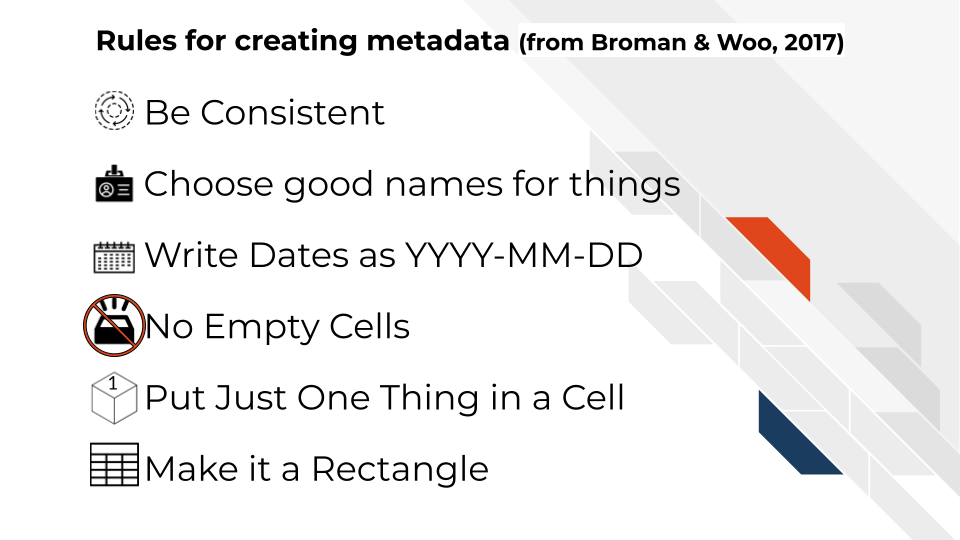
#### 3.3.1.2 Goal B: Avoid introducing errors into your metadata in the future!

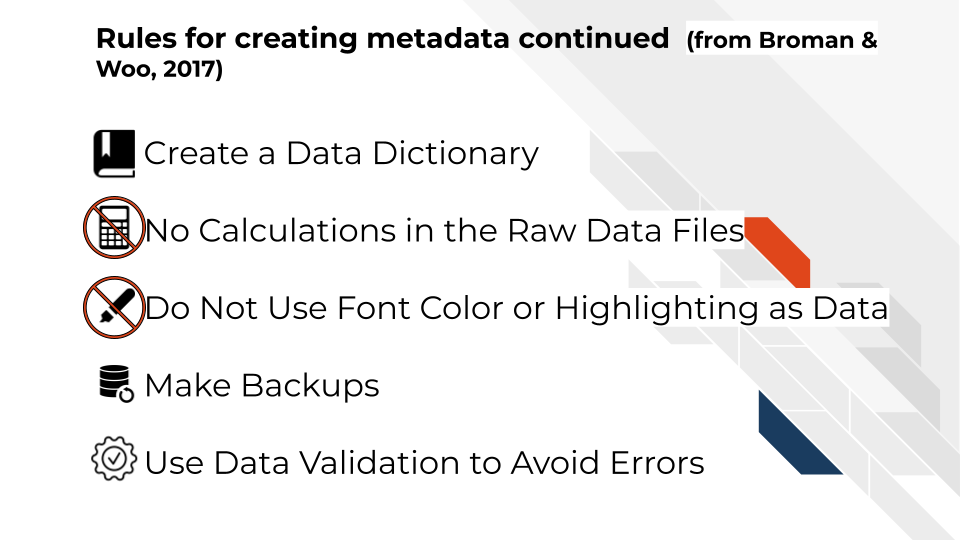
Toward these two goals, [this excellent article](https://www.tandfonline.com/doi/full/10.1080/00031305.2017.1375989) by Broman & Woo discusses metadata design rules. We will very briefly cover the major points here but highly suggest you read the original article.

1. *Be Consistent* - Whatever labels and systems you choose, use it universally. This not only means in your metadata spreadsheet but also anywhere you are discussing your metadata variables.
2. *Choose good names for things* - avoid spaces, special characters, or within the lab jargon.
3. *Write Dates as YYYY-MM-DD* - this is a global standard and less likely to be messed up by Microsoft Excel.
4. *No Empty Cells* - If a particular field is not applicable to a sample, you can put NA but empty cells can lead to formatting errors or just general confusion.
5. *Put Just One Thing in a Cell* - resist the urge to combine variables into one, you have no limit on the number of metadata variables you can make!
6. *Make it a Rectangle* - This is the easiest way to read data, for a computer and a human. Have your samples be the rows and variables be columns.
7. *Create a Data Dictionary* - Have somewhere that you describe what your metadata mean in detailed paragraphs.
8. *No Calculations in the Raw Data Files* - To avoid mishaps, you should always keep a clean, original, raw version of your metadata that you do not add extra calculations or notes to.
9. *Do Not Use Font Color or Highlighting as Data* - This only adds to confusion to others if they don’t understand your color coding scheme. Instead create a new variable for anything you might be tempted to color code.
10. *Make Backups* - Metadata are critical, you never want to lose them because of spilled coffee on a computer. Keep the original backed up in a multiple places. We recommend keeping writing your metadata in something like GoogleSheets because it is both free and also saved online so that it is safe from computer crashes.
11. *Use Data Validation to Avoid Errors* - set data types to have googlesheets or excel check that the data in the columns is the type of data it expects for a given variable.

Note that it is very dangerous to open gene data with Excel. According to Ziemann, Eren, and El-Osta ([2016](#ref-Ziemann2016)), approximately one-fifth of papers with Excel gene lists have errors. This happens because Excel wants to interpret everything as a date. We strongly caution against opening (and saving afterward) gene data in Excel. 

### 3.3.2 To recap:

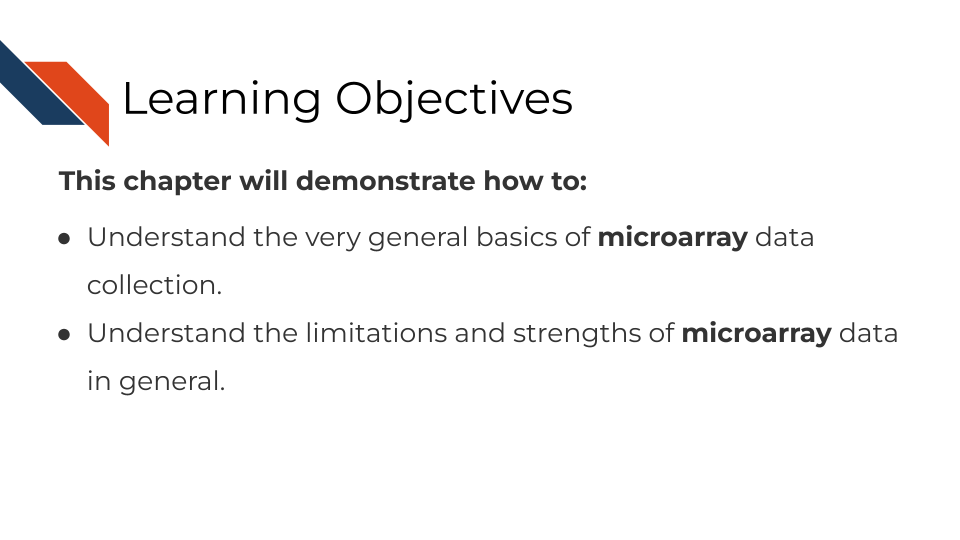




If you are not the person who has the information needed to create metadata, or you believe that another individual already has this information, make sure you get ahold of the metadata that correspond to your data. It will be critical for you to have to do any sort of meaningful analysis!

# 4 Microarray Data

## 4.1 Learning Objectives

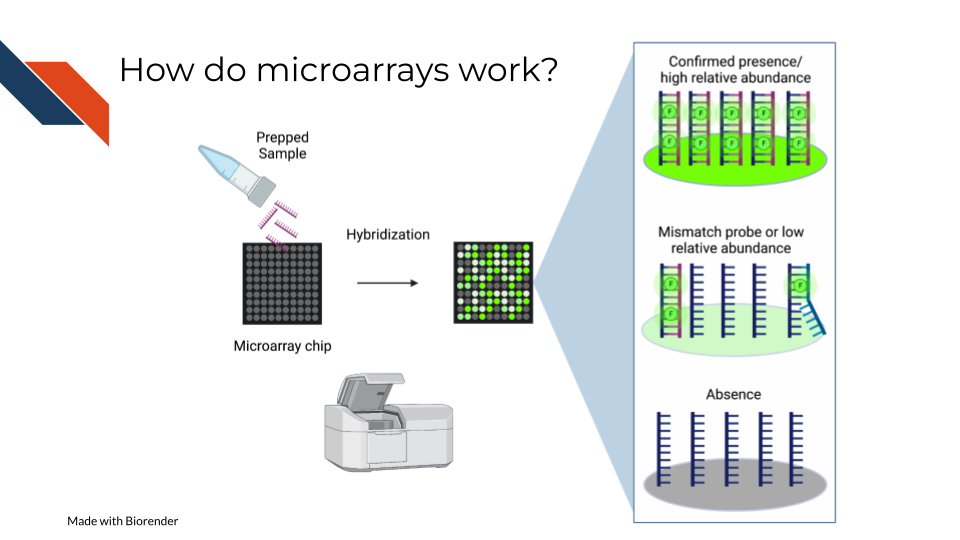


## 4.2 Summary of microarrays

Microarrays have been in use since before high throughput sequencing methods became more affordable and widespread, but they still can be a effective and affordable tool for genomic assays. Depending on your goals, microarray may be a suitable choice for your genomic study.

## 4.3 How do microarrays work?

All microarrays work on hybridization to sets of oligonucleotides on a chip. However, the preparation of the samples, and the oligonucleotides’ hybridization targets vary depending on the assay and goals.



On a basic principle, oligonucleotide probes are designed for different targets sets designed for the same targets are put together. On the whole chip, these probes are arranged in a grid like design so that after a sample is hybridized to them, you can detect how much of the target is detected by taking an image and knowing what target each location is designed to.

### 4.3.1 Pros:

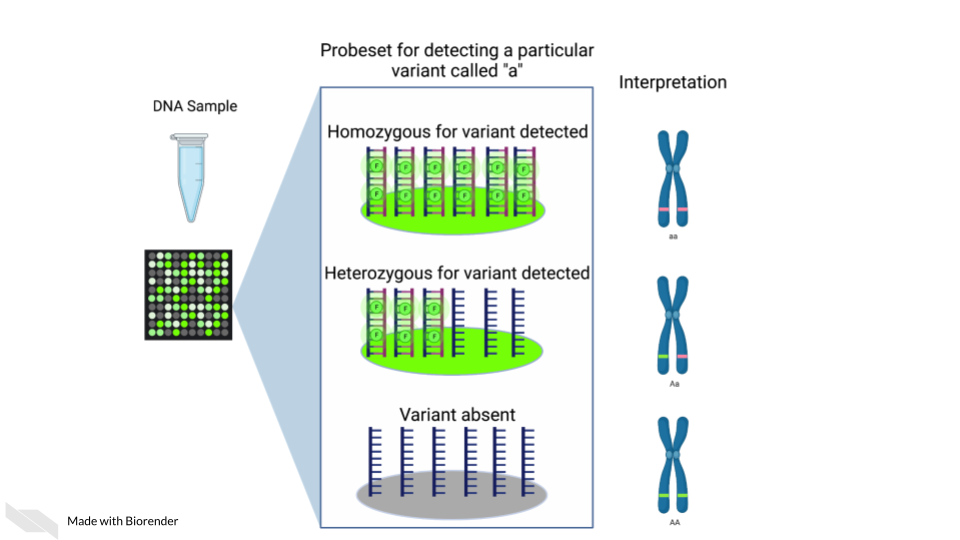
* Microarrays are much more affordable than high throughput sequencing which can allow you to run more samples and have more statistical power ([Tarca, Romero, and Draghici 2006](#ref-Tarca2006); [ALSF 2019](#ref-refinebioexamples2019)).
* Microarrays take less time to process than most high throughput sequencing methods([Tarca, Romero, and Draghici 2006](#ref-Tarca2006); [ALSF 2019](#ref-refinebioexamples2019)).
* Microarrays are generally less computationally intensive to process and you can get your results more quickly([Tarca, Romero, and Draghici 2006](#ref-Tarca2006); [ALSF 2019](#ref-refinebioexamples2019)).
* Microarrays are generally as good as sequencing methods for detecting clinical endpoints ([Zhang et al. 2015](#ref-Zhang2015)).

### 4.3.2 Cons:

* Microarray chips can only measure the targets they are designed for, and cannot be used for exploratory purposes ([Zhang et al. 2015](#ref-Zhang2015)).
* Microarrays’ probe designs can only be as up to date as the genome they were designed against at the time ([Mantione et al. 2014](#ref-Mantione2014); [**refinebioexamples?**](#ref-refinebioexamples)).
* Microarray does not escape oligonucleotide biases like GC content and sequence composition biases([ALSF 2019](#ref-refinebioexamples2019)).

## 4.4 What types of arrays are there?

### 4.4.1 SNP arrays

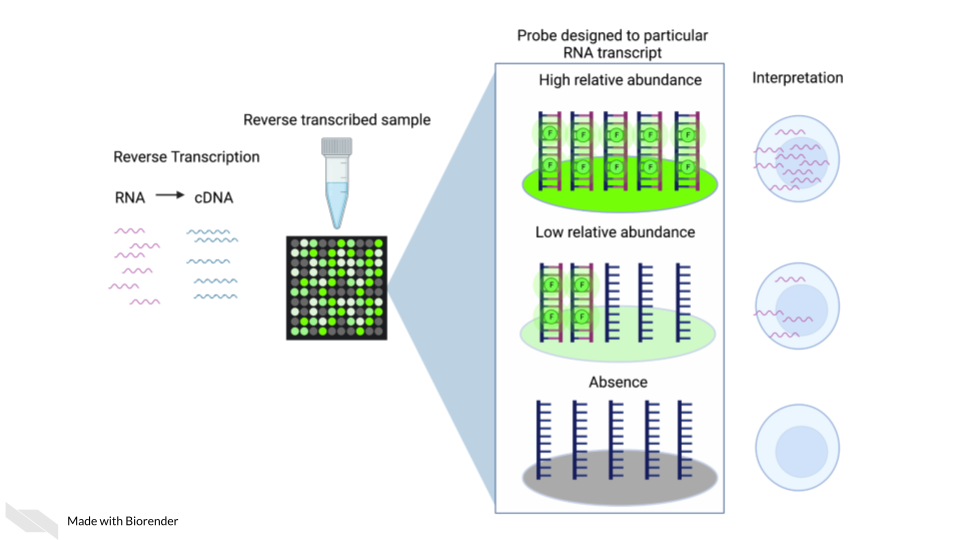


Single nucleotide polymorphism arrays are designed to measure DNA variants. They are designed to target DNA variants. When the sample is hybridized, the amount of fluorescence detected can be interpreted to indicate the presence of the variant and whether the variant is homogeneous or heterogenous. The samples prepped for SNP arrays then need to be DNA samples.

#### 4.4.1.1 Examples:

* [The 1000 genomes project](https://www.internationalgenome.org/) is a large collection of SNP data array from many populations around the world and is available for download.

### 4.4.2 Gene expression arrays

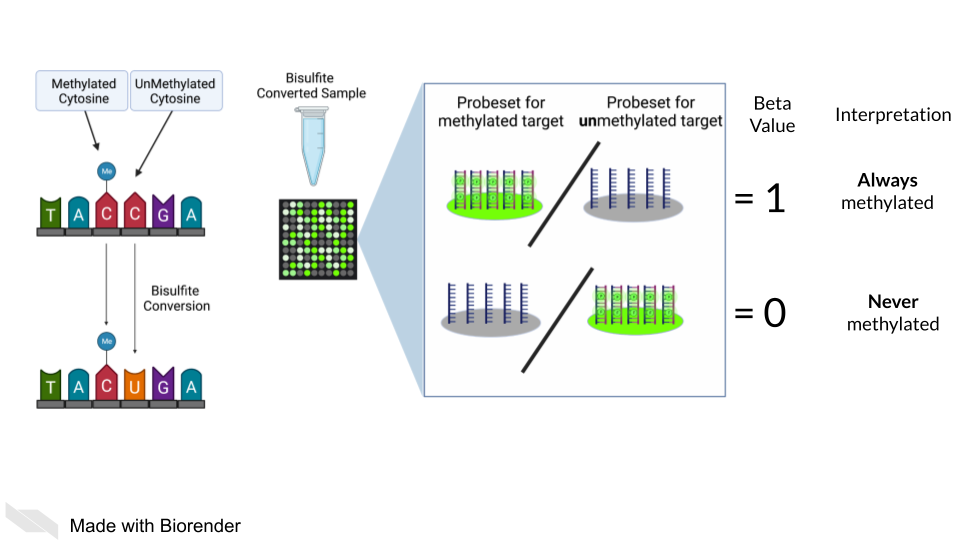


Gene expression arrays are designed to measure gene expression. They are designed to target and measure relative transcript abundance level.

#### 4.4.2.1 Examples:

* [refine.bio](https://www.refine.bio/) is the largest collection of publicly available, already normalized gene expression data (including gene expression microarrays).
* [Getting started in gene expression microarray analysis](https://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1000543) ([**Slonim2009?**](#ref-Slonim2009)).
* [Microarray and its applications](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3467903/) ([**Govindarajan2012?**](#ref-Govindarajan2012)).
* [Analysis of microarray experiments of gene expression profiling](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2435252/) ([Tarca, Romero, and Draghici 2006](#ref-Tarca2006)).

### 4.4.3 DNA methylation arrays

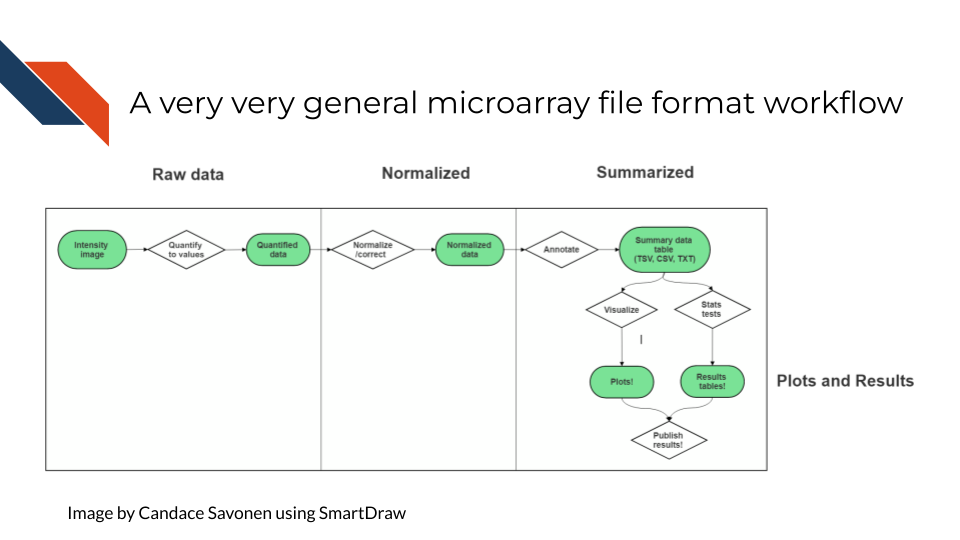


DNA methylation can also be measured by microarray. To detect methylated cytosines (5mC), DNA samples are prepped using bisulfite conversion. This converts unmethylated cytosines into uracils and leaves methylated cytosines untouched. Probes are then designed to bind to either the uracil or the cytosine, representing the unmethylated and methylated cytosines respectively.

A ratio of the fluorescence signal can be used to identify the relative abundance of the methylated and unmethylated versions of the sequence.

Additionally, 5-hydroxymethylated cytosines (5hmC) can also be detected by oxidative bisulfite bisulfite sequencing ([Booth et al. 2013](#ref-Booth2013)). Note that bisulfite conversion alone will not distinguish between 5mC and 5hmC though these often may indicate different biological mechanics.

## 4.5 General processing of microarray data



After scanning, microarray data starts as an image that needs to be quantified, normalized and further corrected and edited based on the most current genome and probe annotation.

As noted above, microarrays do not escape the base sequence biases that accompany most all genomic assays. The normalization methods you use ideally will mitigate these sequence biases and also make sure to remove probes that may be outdated or bind to multiple places on the genome.

The tools and methods by which you normalize and correct the microarray data will be dependent not only on the type of microarray assay you are performing (gene expression, SNP, methylation), but most of all what kind of microarray chip design/platform you are using.

### 4.5.1 Examples

* [Refine.bio describes their processing methods](https://docs.refine.bio/en/latest/main_text.html#processing-information).
* [Brainarray keeps up to date microarray annotation for all kinds of platforms](http://brainarray.mbni.med.umich.edu/Brainarray/Database/CustomCDF/genomic_curated_CDF.asp)

### 4.5.2 Microarray Platforms

There are so many microarray chip designs out there designed to target different things. Three of the largest commercial manufacturers have ready to use microarrays you can purchase. You can also design microarrays to hit your own targets of interest.

Here are full lists of platforms that have been published on Gene Expression Omnibus.

* [Affymetrix platforms](https://www.ncbi.nlm.nih.gov/geo/browse/?view=platforms&submitter=5&zsort=series&display=20)
* [Agilent platforms](https://www.ncbi.nlm.nih.gov/geo/browse/?view=platforms&submitter=63&display=20&zsort=series).
* [Illumina platforms](https://www.ncbi.nlm.nih.gov/geo/browse/?view=platforms&submitter=1242&zsort=series&display=20).

## 4.6 Very General Microarray Workflow

In the data type specific chapters, we will cover the microarray workflow and file formats in more detail. But in the most general sense, microarray workflows look like this, note that the exact file formats are specific to the chip brand and type you use (e.g. Illumina, Affymetrix, Agilent, etc.):

### 4.6.1 Microarray file formats

#### 4.6.1.1 IDAT - intensity data file

This is an Illumina microarray specific file that contains the chip image intensity information for each location on the microarray. It is a binary file, which means it will not be readable by double clicking and attempting to open the file directly.

Currently, Illumina appears to suggest directly converting IDAT files into a GTC format. We advise looking into [this package to help you do that](https://github.com/freeseek/gtc2vcf).

[For more on IDAT files](https://www.illumina.com/content/dam/illumina-marketing/documents/products/technotes/technote_array_analysis_workflows.pdf).

#### 4.6.1.2 DAT - data file

This is an Affymetrix’ microarray specific file parallel to the IDAT file in that it contains the image intensity information for each location on the microarray. It’s stored as pixels.

[For more on DAT files](https://www.affymetrix.com/support/developer/powertools/changelog/gcos-agcc/dat.html).

#### 4.6.1.3 CEL

This is an Affymetrix microarray specific file that is made from a DAT file but translated into numeric values. It is not normalized yet but can be normalized into a CHP file.

[For more on CEL files](https://www.affymetrix.com/support/developer/powertools/changelog/gcos-agcc/cel.html)

#### 4.6.1.4 CHP

CHP files contain the gene-level and normalized data from an Affymetrix array chip. CHP files are obtained by normalizing and processing CEL files.

[For more about CHP files](https://www.affymetrix.com/support/developer/powertools/changelog/gcos-agcc/chp-xda.html).

## 4.7 General informatics files

At various points in your genomics workflows, you may need to use other types of files to help you annotate your data. We’ll also discuss some of these common files that you may encounter:

#### 4.7.0.1 BED - Browser Extensible Data

A BED file is a text file that has coordinates to genomic regions. THe other columns that accompany the genomic coordinates are variable depending on the context. But every BED file contains the chrom, chromStart and chromEnd columns to start.

A BED file might look like this:

chrom chromStart chromEnd other\_optional\_columns  
chr1 0 1000 good  
chr2 100 3000 bad

For [more on BED files](https://en.wikipedia.org/wiki/BED_(file_format)).

#### 4.7.0.2 GFF/GTF General Feature Format/Gene Transfer Format

A GFF file is a tab delimited file that contains information about genomic features. These types of files are available from databases and what you can use to annotate your data.

You may see there are GFF2, GFF3, and GTF files. These only refer to different versions and variations. They generally have the same information. In general, GFF2 is being phased out so using GFF3 is generally a better bet unless the program or package you are using specifies it needs an older GFF2 version.

A GFF file may look like this (borrowed example from Ensembl):

1 transcribed\_unprocessed\_pseudogene gene 11869 14409 . + . gene\_id "ENSG00000223972"; gene\_name "DDX11L1"; gene\_source "havana"; gene\_biotype "transcribed\_unprocessed\_pseudogene";

Note that it will be useful for annotating genes and what we know about them.

For [more about GTF and GFF files](https://useast.ensembl.org/info/website/upload/gff.html).

### 4.7.1 Other files

\* If you didn’t see a file type listed you are looking for, take a look at this [list by the BROAD](https://software.broadinstitute.org/cancer/software/gsea/wiki/index.php/Data_formats). Or, it may be covered in the data type specific chapters.

### 4.7.2 Microarray processing tutorials:

For the most common microarray platforms, you can see these examples for how to process the data:

#### 4.7.2.1 General arrays

* [Using Bioconductor for Microarray Analysis](https://www.bioconductor.org/packages/devel/workflows/vignettes/arrays/inst/doc/arrays.html).

#### 4.7.2.2 Gene Expression Arrays

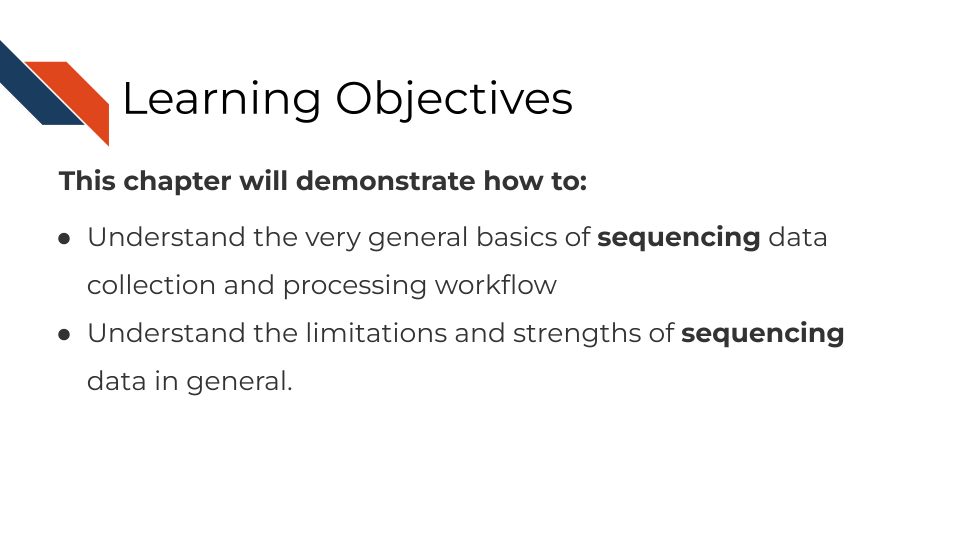
* [An end to end workflow for differential gene expression using Affymetrix microarrays](https://f1000research.com/articles/5-1384).

#### 4.7.2.3 DNA Methylation Arrays

* [DNA Methylation array workflow](https://nbis-workshop-epigenomics.readthedocs.io/en/latest/content/tutorials/methylationArray/Array_Tutorial.html).

# 5 Sequencing Data

## 5.1 Learning Objectives



In this section, we are going to discuss generalities that apply to all sequencing data. This is meant to be a “primer” for you which data-type specific chapters will build off of to give you more specific and practical steps and advice in regards to your data type.

## 5.2 How does sequencing work?



Sequencing methods, whether they are targeting DNA, transcriptomes, or some other target of the genome, have some commonalities in the steps as well as what types of biases and data generation artifacts to look out for.

All sequencing experiments start out with the extraction of the biological material of interest. This biological material will be processed in some way to isolate to the genomic target of interest (we will cover the various techniques for this in more detail in each respective data chapter since it is highly specific to the data type).

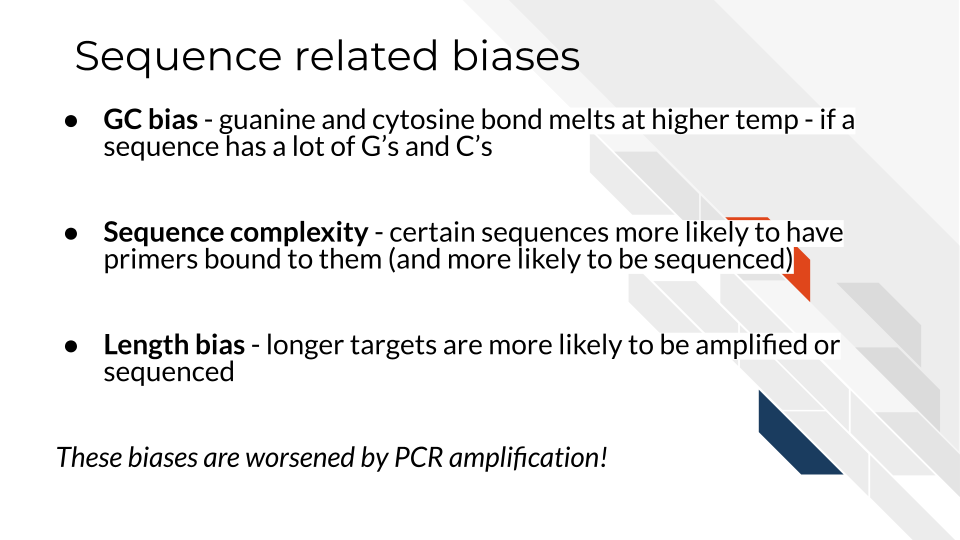
This set of processing steps will lead up to library generation – adding a way to catalog what molecules came from where. Sometimes for this library prep the sequences need to be fragmented before hand and an adapter bound to them.

The resulting sample material is often a very small quantity, which means Polymerase Chain Reaction (PCR) needs to be used to amplify the material to a quantity large enough to be reliably sequenced. We will talk about how this very common method not only amplifies the sequences we want to read but amplifies sequence method biases that we would like to avoid.

At the end of this process, base sequences are called for the samples (with varying degrees of confidence), creating huge amounts of data and what hopefully contains valuable research insights.

## 5.3 Sequencing concepts

### 5.3.1 Inherent biases



Sequences are not all sequenced or amplified at the same rate. In a perfect world, we could take a simple snapshot of the genome we are interested in and know exactly what and how many sequences were in a sample. But in reality, sequencing methods and the resulting data always have some biases we have to be aware of and hopefully use methods that attempt to mitigate the biases.

#### 5.3.1.1 GC bias

You may recall that with nucleotides: adenine binds with thymine and guanine binds with cytosine. But, the guanine-cytosine bond (GC) has 3 hydrogen bonds whereas the adenine-thymine bond (AT) has only 2 bonds. This means that the GC bond is stickier (to put it scientifically) and needs higher temperatures to unbind. The sequencing and PCR amplification process involves cycling through temperatures and binding and unbinding of sequences which means that if a sequence has a lot of G’s and C’s (high GC content) it will unbind at a different temperatures than a sequence of low GC content.

#### 5.3.1.2 Sequence complexity

Nonrepeating sequences are harder to sequence and amplify than repeating sequences. This means that the complexity of a target sequence influences the PCR amplification and detection.

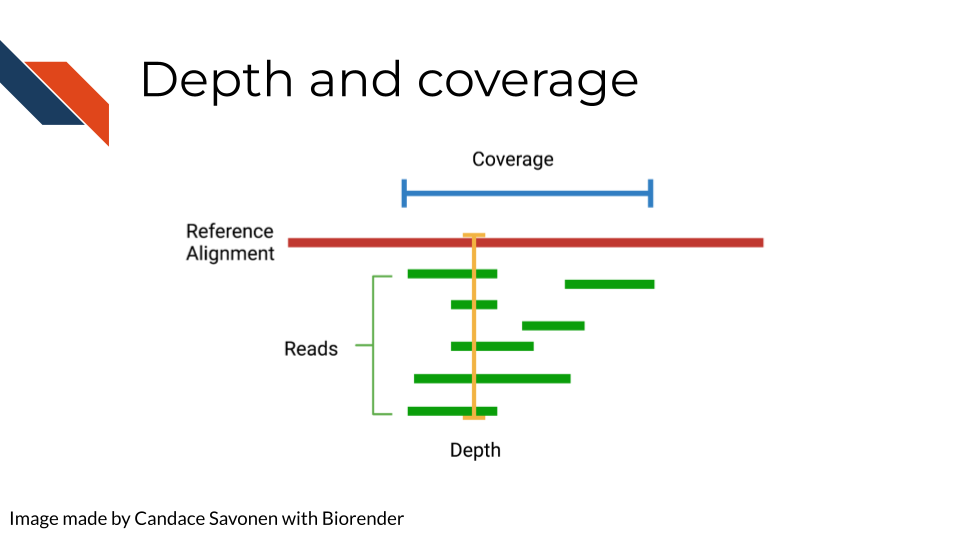
#### 5.3.1.3 Length bias

Longer sequences – whether they represent long sequence variants, long transcripts, or etc, are more likely to be identified than shorter ones! So if you are attempting to quantify the presence of a sequence, a longer sequence is much more likely to be counted more often.

### 5.3.2 PCR Amplification

All of the above biases are amplified when the sequences are being amplified! You can picture that if each of these biases have a certain effect for one copy, then as PCR steps copy the sequence exponentially, the error is also being multiplied! PCR amplification is generally a necessary part of the process. But there are tools that allow you to try to combat the biases of PCR amplification in your data analysis. These tools will be dependent on the type of sequencing methods you are using and will be something that is discussed in each data type chapter.

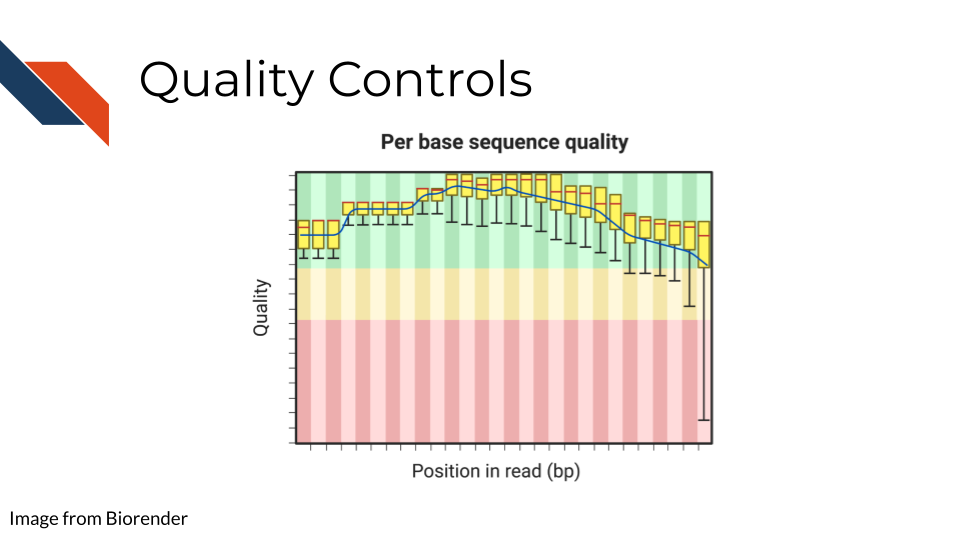
### 5.3.3 Depth of coverage



The depth of sequencing refers to how many times on average a particular base is sequenced. Obviously the more times something is sequenced, the more you can be confident that the base call is accurate. However, sequencing at greater depths also takes more time and money. Depending on your sequencing goals and methods there is an appropriate level of depth that is needed.

Coverage on the other hand has to do with how much of the target is covered. If you are doing Whole Genome Sequencing, what percentage of the whole genome were you able to sequence? You may realize how depth is related to coverage, in that the greater depth of sequencing you use the more likely you are to also cover more of the genome. As discussed in relation to the biases, some part of the genome are harder to reach than others, so by reading at greater depths some of those “hard to read” parts of the genome will be able to be covered.

### 5.3.4 Quality controls

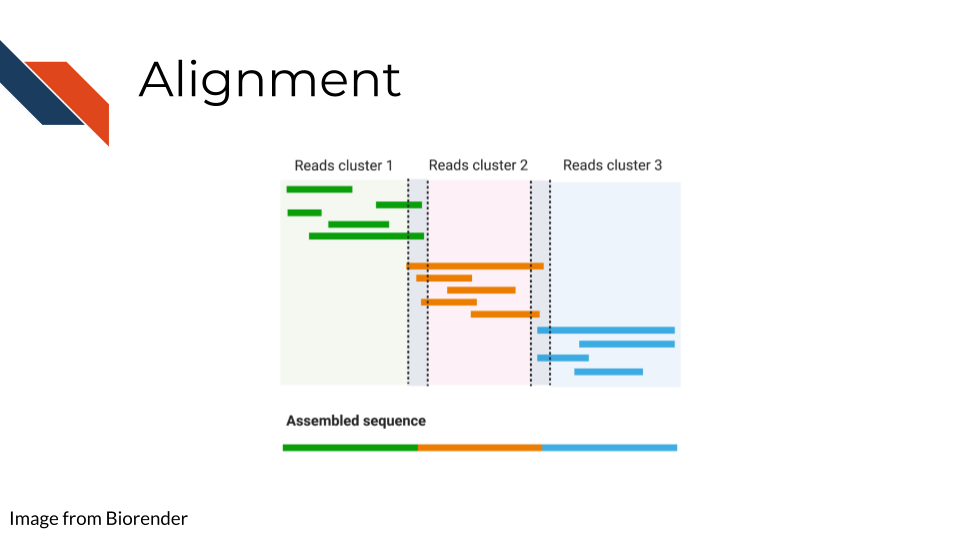


Sequencing bases involves some error/confidence rate. As mentioned, some parts of the genome are harder to read than others. Or, sometimes your sequencing can be influenced by poor quality sample that has degraded. Before you jump in to further analyzing your data, you will want to investigate the quality of the sequencing data you’ve collected.

The most common and well-known method for assessing sequencing quality controls is [FASTQC](https://www.bioinformatics.babraham.ac.uk/projects/fastqc/). FASTQC creates an abundance of sequencing quality control reports from fastq files. These reports need to be interpreted within the context of your sequencing methods, samples, and experimental goals. Often bioinformatics cores are good to contact about these reports (they may have already run FASTQC on your data if that is where you obtained your data initially). They can help you wade through the flood of quality control reports printed out by FASTQC.

FASTQC also has great documentation that can attempt to guide you through report interpretation. This also includes examples of good and bad FASTQC reports. But note that all FASTQC report interpretations must be done relative to the experiment that you have done. In other words, there is not a one size fits all quality control cutoffs for your FASTQC reports. The failure/success icons FASTQC reports back are based on defaults that may not be accurate or applicable to your data, so further investigation and consultation is warranted before you decided to trust or pitch your sequencing data.

### 5.3.5 Alignment

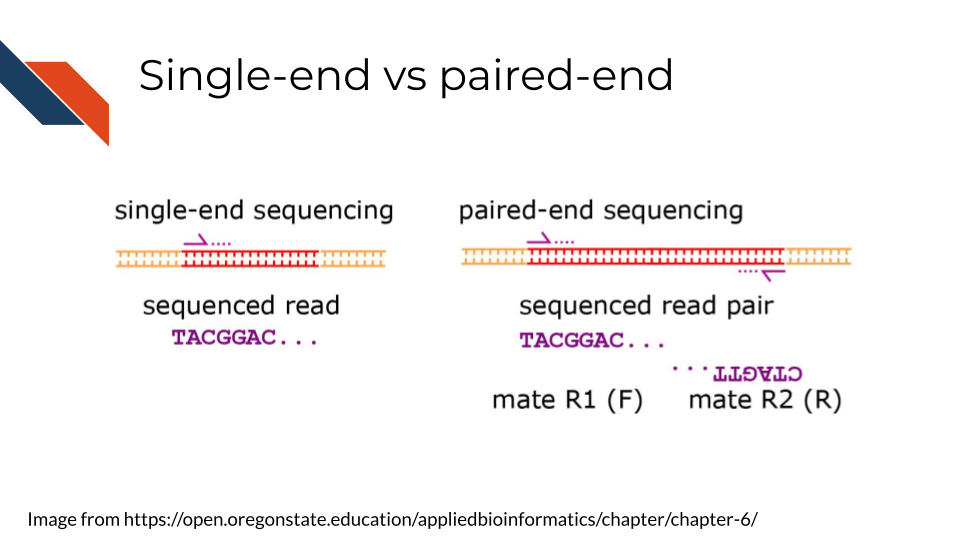


Once you have your reads and you find them reasonably trustworthy through quality control checks, you will want to align them to your reference. The reference you align your sequences to will depend on the data type you have: a reference genome, a reference transcriptome, something else?

* Traditional aligners - Align your data to a reference using standard alignment algorithms. Can be very computationally intensive.
* Pseudo aligners - much faster and the trade off for accuracy is often negligible (but again is dependent on the data you are using).

TODO: considerations for alignment.

### 5.3.6 Single End vs Paired End



Sequencing can be done single-end or paired-end. Paired end means the primers are going to bind to both sides of a sequence. This can help you avoid some 3’ bias and give you more complete coverage of the area you are sequencing. But, as you may guess, pair-end read sequencing is more expensive than single end.

You will want to determine whether your sequencing is paired end or single end. If it is paired end you will likely see file names that indicate this. You should have pairs of files that may or may not be labeled with \_1 and \_2 or \_F and \_R. We will discuss file nomenclature more specifically as it pertains to different data types in the upcoming chapters.

## 5.4 Very General Sequencing Workflow

In the data type specific chapters, we will cover the sequencing data workflows and file formats in more detail. But in the most general sense, sequencing workflows look like this:

### 5.4.1 Sequencing file formats

#### 5.4.1.1 SAM - Sequence Alignment Map

SAM Files are text based files that have sequence information. It generally has not been quantified or mapped. It is the reads in their raw form. [For more about SAM files](https://samtools.github.io/hts-specs/SAMv1.pdf).

#### 5.4.1.2 BAM - Binary Alignment Map

BAM files are like SAM files but are compressed (made to take up less space on your computer). This means if you double click on a BAM file to look at it, it will look jumbled and unintelligible. You will need to convert it to a SAM file if you want to see it yourself (but this isn’t necessary necessarily).

#### 5.4.1.3 FASTA - “fast A”

Fasta files are sequence files that can be either nucleotide or amino acid sequences. They look something like this (the example below illustrating an amino acid sequence):

>SEQ\_ID  
GATTTGGGGTTCAAAGCAGTATCGATCAAATAGTAAATCCATTTGTTCAACTCACAGTTT

For [more about fasta files](https://en.wikipedia.org/wiki/FASTA_format).

#### 5.4.1.4 FASTQ - “Fast q”

A Fastq file is like a Fasta file except that it also contains information about the **Q**uality of the read. By quality, we mean, how sure was the sequencing machine that the nucleotide or amino acid called was indeed called correctly?

@SEQ\_ID  
GATTTGGGGTTCAAAGCAGTATCGATCAAATAGTAAATCCATTTGTTCAACTCACAGTTT  
+  
!''\*((((\*\*\*+))%%%++)(%%%%).1\*\*\*-+\*''))\*\*55CCF>>>>>>CCCCCCC65

For [more about fastq files](https://en.wikipedia.org/wiki/FASTQ_format).

Later in this course we will discuss the importance of examining the quality of your sequencing data and how to do that. If you received your data from a bioinformatics core it is possible that they’ve already done this quality analysis for you.

*Sequencing data that is not of high enough quality should not be trusted!* It may need to be re-run entirely or may need extra processing (trimming) in order to make it more trustworthy. We will discuss this more in later chapters.

#### 5.4.1.5 BCL - binary base call (BCL) sequence file format

This type of sequence file is specific to Illumina data. In most cases, you will simply want to convert it to Fastq files for use with non-Illumina programs.

[More about BCL to Fastq conversion](https://medium.com/@marija190396/bcl-to-fastq-conversion-e289852823d0).

#### 5.4.1.6 VCF - Variant Call Format

VCF files are further processed form of data than the sequence files we discussed above. VCF files are specially for storing only where a particular sample’s sequences differ or are *variant* from the reference genome or each other.

This will only be pertinent to you if you care about DNA variants. We will discuss this in the DNA seq chapter.

For [more on VCF files](https://en.wikipedia.org/wiki/Variant_Call_Format).

#### 5.4.1.7 MAF - Mutation Annotation Format

MAF files are aggregated versions of VCF files. So for a group of samples for which each has a VCF file, your entire group of samples’ variants will be summarized in the form of a MAF file.

For [more on MAF files](https://docs.gdc.cancer.gov/Data/File_Formats/MAF_Format/#:~:text=Mutation%20Annotation%20Format%20(MAF)%20is,(or%20open%2Daccess).).

### 5.4.2 Other files

\* If you didn’t see a file type listed you are looking for, take a look at this [list by the BROAD](https://software.broadinstitute.org/cancer/software/gsea/wiki/index.php/Data_formats). Or, it may be covered in the data type specific chapters.

# 6 DNA Methods

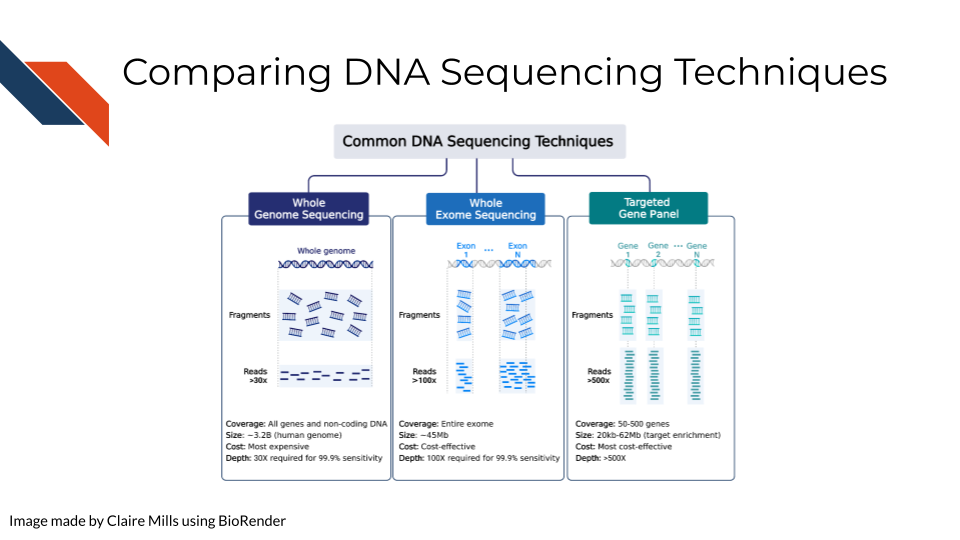
## 6.1 Learning Objectives



## 6.2 What are the goals of analyzing DNA sequences?



## 6.3 Comparison of DNA methods

 Compared to WXS and Targeted Gene Sequencing, WGS is the most expensive but requires the lowest depth of coverage to achieve 95% sensitivity. In other words, WGS requires sequencing each region of the genome (3.2 billion bases) 30 times in order to confidently be able to pick up all possible meaningful variants. ([Sims et al. 2014](#ref-Sims2014)) goes into more depth on how these depths are calculated.

Alternatively, WXS is a more cost effective way to study the genome, focusing places in the genome that have open reading frames – aka generally genes that are able to be expressed. This focuses on enriching for exons and not introns so splicing variants may be missed. In this case, each gene must be sequenced 80-100x for sufficient sensitivity to pick up meaningful variants.

In targeted gene sequencing, a panel of 50-500 regions of interest are selected. This technique is very applicable for studying a set of specific genes of interest at great depth to identify all varieties of mutations within those specific genes. These genes must be sequenced at much greater depth (>500x) to confidently identify all meaningful variants. [This page from Illumina](https://www.illumina.com/science/technology/next-generation-sequencing/plan-experiments/coverage.html) also provides information regarding sequencing depth considerations for different modalities.

Additional references: WGS: ([Bentley et al. 2008](#ref-Bentley2008)) WES: ([Clark et al. 2011](#ref-Clark2011)) Targeted: ([Bewicke-Copley et al. 2019](#ref-BewickeCopley2019))

## 6.4 How to choose a DNA sequencing method

Before starting any sequencing method, you likely have a research question or hypothesis in mind. In order to choose a DNA sequencing method, you will need to consider a few items in balance of each other:

### 6.4.1 1. What region(s) of the genome pertain to your research question?

Is this unknown? Can it be narrowed down to non-coding or coding regions? Is there an even more specific subset of interest?

### 6.4.2 2. What does your project budget allow for?

Some methods are much more costly than others. Cost is not only a factor for the reagents needed to sequence, but also the computing power needed to process and store the data and people’s compensation for their work on the data. All of these costs increase as the amounts of data that are collected increase. For more information on computing decisions see our [Computing in Cancer Informatics course](https://www.itcrtraining.org/courses#h.civy2cnri95t).

### 6.4.3 3. What is your detection power for these variants?

Detecting DNA variants is not simply a matter of yes or no, but a confidence level due to sequencing errors in data collection. Are the variants you are looking for very rare and/or small (single nucleotide or very few copy number differences)? If so you will need more samples and potentially more sequencing depth to detect these variants with confidence.

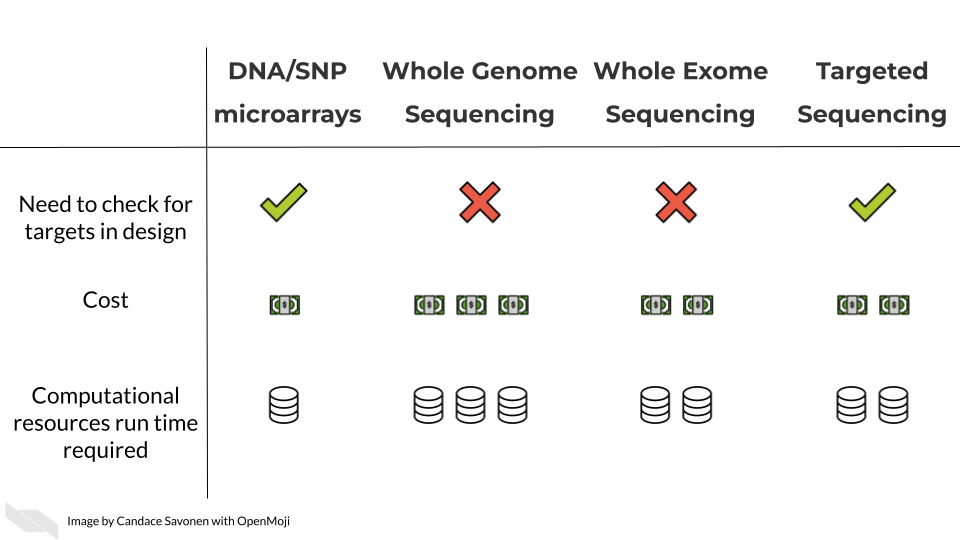
## 6.5 Strengths and Weaknesses of different methods

Is not much known about DNA variants in your organism or disease in question? In this instance you may want to cast a large net to explore more variants by using WGS.

If previous research has identified sections of the genome that are of interest to your research question, then it’s highly advisable to not sequence the entire genome with WGS methods. Not only will whole genome sequencing be more costly, but it will decrease your statistical power to discover true positive variants of interest and increase your chances of discovering false positive variants. This is because multiple testing correction needs to be applied in instances where many tests are being done currently. In this instance, the tests being performed are across the whole genome.

If your research question does not pertain to non-coding regions of the genome or splicing, then its advisable to use WXS. Recall that only about 1-2% of the genome is coding sequences meaning that if you are uninterested in noncoding regions but still use WGS then 98-99% of your data will be uninteresting to you and will only serve to increase your chances of finding false positives or cost you a lot of funding. Not only does sequencing more of the genome take more money and time but it will be more costly in time and resources in terms of the computing power needed to analyze it.

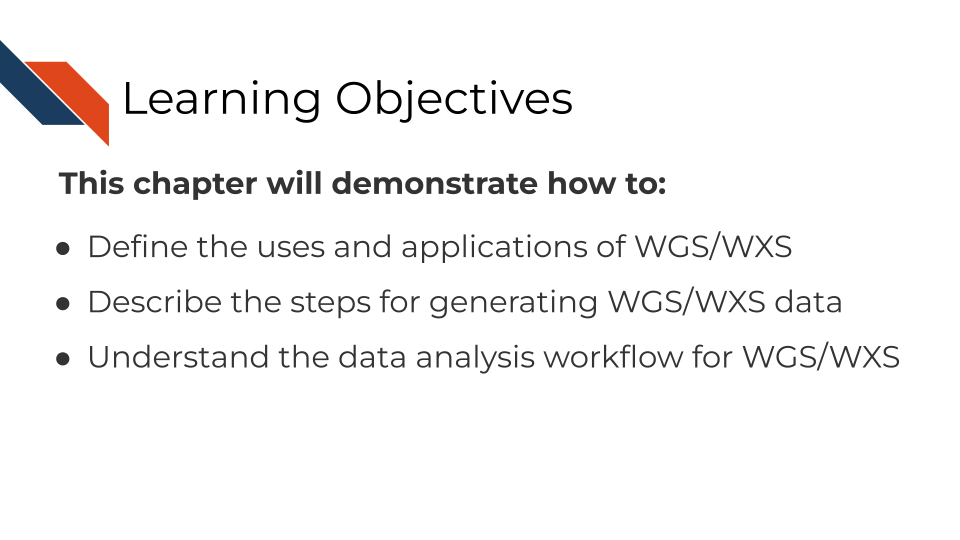
Furthermore, if you are able to narrow down even further what regions are of interest this would be better in terms of cost and detection abilities. A targeted sequencing panel or DNA microarray are ideal for assaying known groups of targets. DNA microarrays are the least costly of all the methods to identify DNA variants, but with both targeted sequencing and DNA microarray you will need to find or create a custom probe or primer set. Ideally a probe or primer set that hits your regions of interest already exists commercially but if not, then you will have to design your own – which also costs time and money.



In these upcoming chapters we will discuss in more detail each of these methods, what the data represent, what you need to consider, and what resources you can consult for analyzing your data.

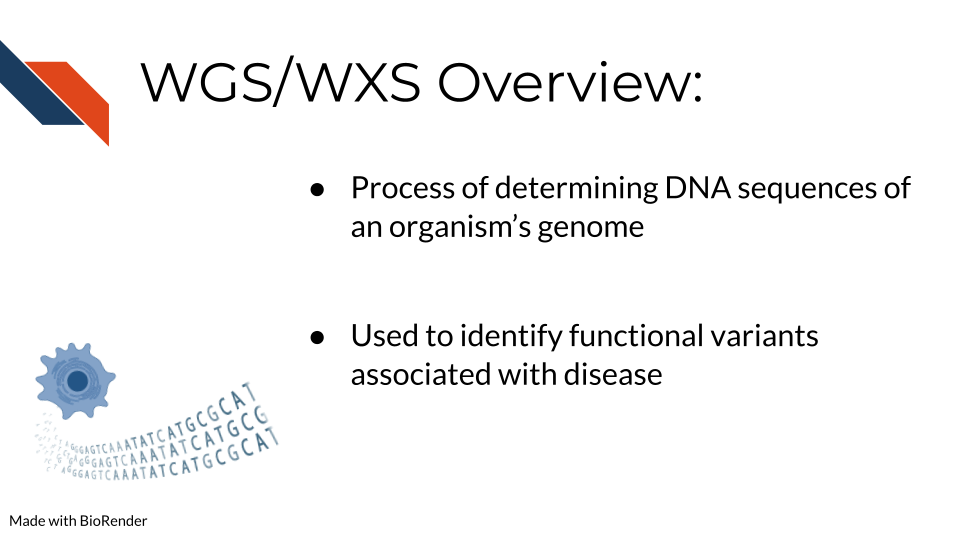
# 7 Whole Genome/Exome Sequencing

## 7.1 Learning Objectives

 The learning objectives for this course are to explain the use and application of Whole Genome Sequencing (WGS) and Whole Exome Sequencing (WES/WXS) for genomics studies, outline the technical steps in generating WGS/WXS data, and detail the processing steps for analyzing and interpreting WGS/WXS data.

**To familiarize yourself with sequencing methods as a whole, we recommend you read our** [**chapter on sequencing first**](http://hutchdatascience.org/Choosing_Genomics_Tools/sequencing-data.html)**.**

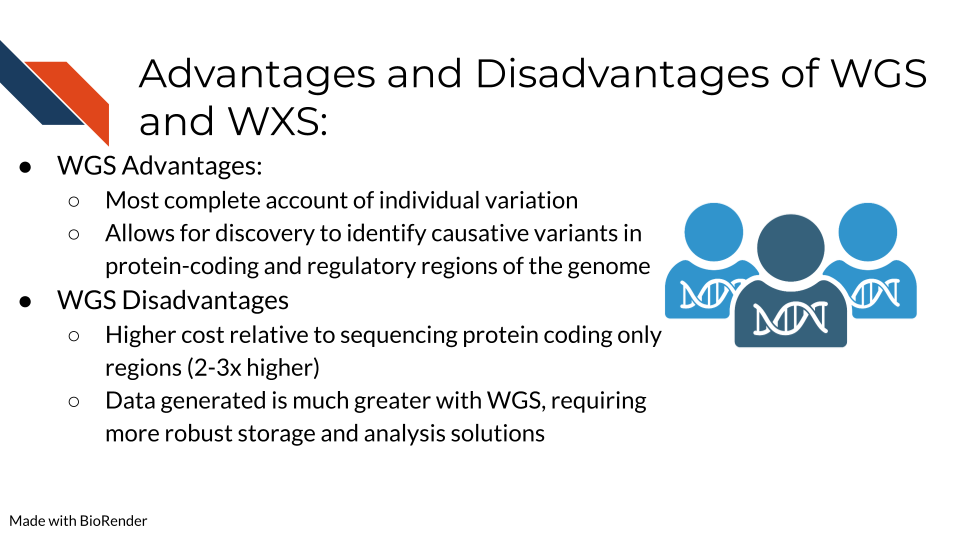
## 7.2 WGS and WGS Overview

 The difference between WGS and WXS sequencing is whether or not the open reading frames and thus coding regions are targeted in sequencing. WGS attempts to sequence the whole genome, while for WXS only exons with open reading frames are targeted for sequencing. Both of these methods can be massively beneficial for studying rare and complex diseases.

Thus, whole genome sequencing is a technique to thoroughly analyze the entire DNA sequence of an organism’s genome. This includes sequencing all genes both coding and non-coding and all mitochondrial DNA. WGS is beneficial for identifying new and previously established variants related to disease and the regulatory elements of the genome including promoters, enhancers, and silencers. Increasingly non-coding RNAs have also been identified to play a functional role in biological mechanisms and diseases. In order to learn more about the non-coding regions of the genome, WGS is necessary.

Alternatively whole exome sequencing is used to sequence the coding regions of an organism’s genome. Although non-coding regions can sometimes reveal valuable insights, coding regions can be a useful area of the genome to focus sequencing methods on, since changes in a protein coding sequence of the genome generally have more information known about them. Often protein coding sequences can have more clearly functional changes - like if a stop codon is introduced or a codon is changed to a predictable amino acid. This can more easily lead to downstream investigations on the functional implications of the protein affected.

## 7.3 Advantages and Disadvantages of WGS vs WXS



We more thoroughly discuss how to choose DNA sequencing methods [here in the previous chapter](http://hutchdatascience.org/Choosing_Genomics_Tools/dna-methods.html), but we will briefly cover this here. Alternatives to WGS include Whole Exome Sequencing (WES/WXS), which sequences the open reading frame areas of the genome or Targeted Gene Sequencing where probes have been designed to sequence only regions of interest. The main advantages of WGS include the ability to comprehensively analyze all regions of a genome, the ability to study structural rearrangements, gene copy number alterations, insertions and deletions, single nucleotide polymorphisms (SNPs), and sequencing repeats. Some disadvantages include higher sequencing costs and the necessity for more robust storage and analysis solutions to manage the much larger data output generated from WGS.

## 7.4 WGS/WXS Considerations

 Some important considerations for WGS/WXS include:

* What genome you are studying and the size of this genome. Included in this considerations is whether this genome has been sequenced before and you will have a “reference” genome to compare your data against or whether you will have to make a reference genome yourself. [This bioinformatics resource](https://eriqande.github.io/eca-bioinf-handbook/alignment-of-sequence-data-to-a-reference-genome-and-associated-steps.html) provides a great overview of genome alignment.
* The depth of coverage for sequencing is an important consideration. The typical recommendation for WGS coverage is 30x, but this is on the lower side and many researchers find it does not provide sufficient coverage compared to 50x. Illumina has an [infographic](https://www.illumina.com/content/dam/illumina-marketing/documents/techniques/wgs-wes-infographic-web.pdf) that explains this information
* The tissue source and whether genetic alterations were introduced during processing are important. Fixation for formalin-fixed paraffin embedded (FFPE) can introduce mutations/genetic changes that will need to be accounted for during data analysis. [This page from Beckman](https://www.beckman.com/resources/technologies/next-generation-sequencing/challenges-with-ffpe-tissue-samples) addresses many of the questions researchers often have about utilizing FFPE samples for their sequencing studies.
* The library preparation method of DNA amplification via PCR is very important as PCR can often introduce duplicates that interfere with interpreting whether a mutant gene is truly frequent or just over amplified during sequencing preparation. [Illumina](https://www.illumina.com/products/by-type/sequencing-kits/library-prep-kits/dna-pcr-free-prep.html) provides a comparison of using PCR and PCR-free library preparation methods on their website.

### 7.4.1 Target enrichment techniques

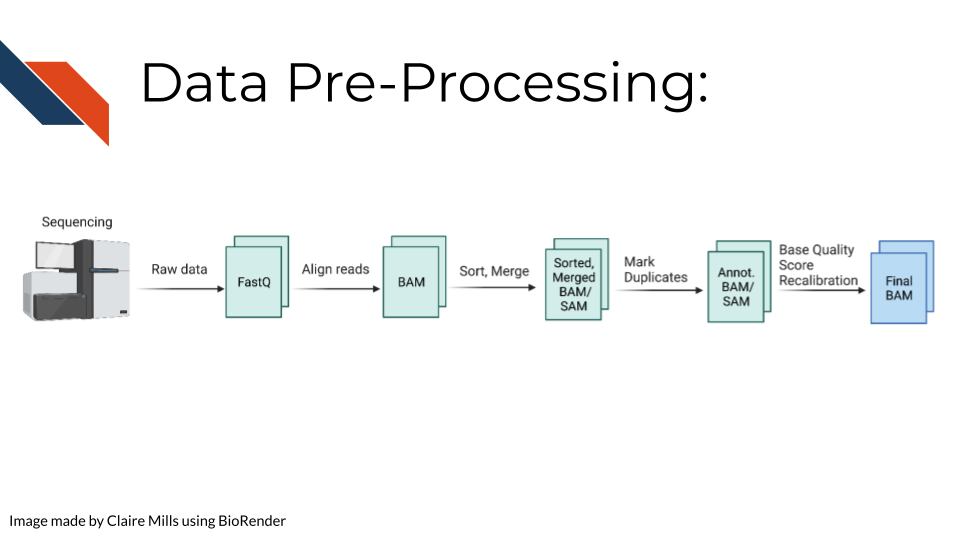
For WXS or other targeted sequencing specifically (so not relevant to WGS data), what methods were used to enrich for the targeted sequences? (Which is the entire exome in the case of general WXS) These methods are generally summarized into two major categories: Hybridization based and amplicon based enrichment.

- [Hybridization based enrichment](https://www.paragongenomics.com/target-enrichment/). This includes a variety of widely used methods that we will broadly categorize in two groups: Array-based and In-solution:  
 - [Array-based capture](https://en.wikipedia.org/wiki/Exome\_sequencing#:~:text=Target%2Denrichment%20strategies-,Array%2Dbased%20capture,-In%2Dsolution%20capture) uses microarrays that have probes designed to bind to known coding sequences. Fragments that do not bind to these probes are washed away, leaving the sample with known coding sequences bound and ready for PCR amplification [@Hodges2007; @Turner2009].  
 - [In-solution capture](https://en.wikipedia.org/wiki/Exome\_sequencing#In-solution\_capture) has become more popular in recent years because it [requires less sample DNA than array-base capture](https://sequencing.roche.com/global/en/article-listing/what-is-ngs-target-enrichment-and-why-is-it-important.html). To enrich for coding sequences, in-solution capture has a pool of custom probes that are designed to bind to the coding regions in the sample. Attached to these probes are beads which can be physically separated from DNA that is not bound to the probes (this should be the non-coding sequences) [@Mamanova2010].   
- [PCR/Amplicon based enrichment](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9318977/) requires even less sample than the other two strategies and so is ideal for when the amount of sample is limited or the DNA has been otherwise processed harshly (e.g. with paraffin embedding). Because the other two enrichment methods are done after PCR amplification has been done to the whole genomic DNA sample, its thought that this method of selective PCR amplification for enrichment can result in more uniformly amplified DNA in the resulting sample. However this is less suitable the more gene targets you have (like if you truly need to sequence all of the exome) since amplicons need to be designed for each target. Overall it is much more affordable of a method. There are several variations of this method that are [discussed thoroughly by @Singh2022](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9318977/).

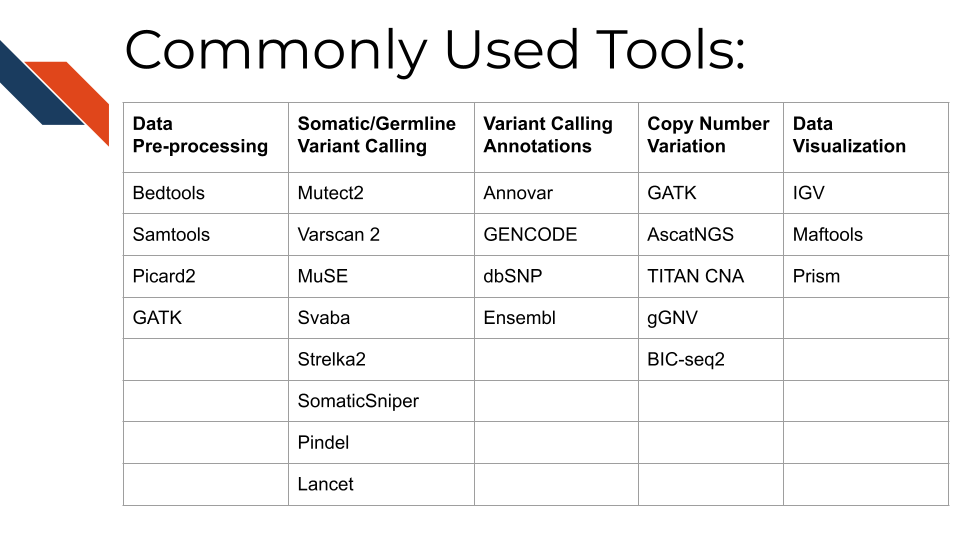
## 7.5 DNA Sequencing Pipeline Overview

 In order to create WGS/WXS data, DNA is first extracted from a specific sample type (tissue, blood samples, cells, FFPE blocks, etc.). Either traditional (involving phenol and chloroform) or commercial kits can be used for this first step. Next, the DNA sequencing libraries are prepared. This involves fragmenting the DNA, adding sequencing adapters, and DNA amplification if the input DNA is not of sufficient quantity. Recall that for WXS After sequencing, data is analyzed by converting and aligning reads to generate a BAM file. Many analysis tools will use the BAM file to identify variants, which then generates a VCF file. More information about sequencing and BAM and VCF file generation can be found [here](http://hutchdatascience.org/Choosing_Genomics_Tools/sequencing-data.html) in the sequencing data chapter.

## 7.6 Data Pre-processing

 Raw sequencing reads are first transformed into a fastq file (more information about fastq files can be found [here](http://hutchdatascience.org/Choosing_Genomics_Tools/sequencing-data.html) in the sequencing data chapter in the Quality Controls section. Then the sequencing reads are aligned to a reference genome to create a BAM file. This data is sorted and merged, and PCR duplicates are identified. The confidence that each read was sequenced correctly is reflected in the base quality score. This score must be recalibrated at this step before variants are called. A final BAM file is thus created. This can be used for future analysis steps include variant or mutation identification, which is outlined on the following slide.

## 7.7 Commonly Used Tools

 The following link provides the data analysis pipeline written by researchers in the NCI division of the NIH and provides a helpful overview of the typical steps necessary for [WGS analysis](https://docs.gdc.cancer.gov/Data/Bioinformatics_Pipelines/DNA_Seq_Variant_Calling_Pipeline/).

Here are many of the tools and resources used by researchers for analyzing WGS data.

## 7.8 Data pre-processing tools

In most cases, all of these tools will be used sequentially to prepare the data for downstream mutational and copy number variation (CNV) analysis.

* [Bedtools](https://bedtools.readthedocs.io/en/latest/) including the bamtofastq function, which is the first step in converting data off the sequencer to a usable format for downstream analysis
* [Samtools](http://www.htslib.org/) including tools for converting fastq to BAM files while mapping genes to the genome, duplicate read marking, and sorting reads
* [Picard2](https://broadinstitute.github.io/picard/) including tools to covert fastq to SAM files, filter files, create indices, mark read duplicates, sort files, and merge files
* [GATK](https://gatk.broadinstitute.org/hc/en-us) is a comprehensive set of tools from the Broad Institute for analyzing many types of sequencing data. For pre-processing, the print read function is very beneficial for writing the reads from a BAM or SAM file that pass specific criteria to a new file

## 7.9 Tools for somatic and germline variant identification

These tools are used to identify either somatic or germline mutations from a sequenced sample. Many researchers will often use a combination of these tools to narrow down only variants that are identified using a combination of these analysis algorithms. All of these mutation calling tools except SvABA can be used on both WGS and WXS data.

* [Mutect2](https://gatk.broadinstitute.org/hc/en-us/articles/9570422171291-Mutect2) This is a beneficial variant calling tool with functions including using a “panel of normals” (samples provided by the user of many normal controls) to better compare disease samples to normal and filtering functions for samples with orientation bias artifacts (FFPE samples) called F1R2, which is explained in the link above.
* [Varscan 2](https://varscan.sourceforge.net/) This is a helpful tool that utilizes a heuristic/statistic approach to variant calling. This means that it detects somatic CNAs (SCNAs) as deviations from the log-ratio of sequence coverage depth within a tumor–normal pair, and then quantify the deviations statistically. This approach is unique because it accounts for differences in read depth between the tumor and normal sample. Varscan 2 can also be used for identifying copy number alterations in tumor-normal pairs.
* [MuSE](https://bioinformatics.mdanderson.org/public-software/muse/) This is a beneficial mutation calling tool when you have both tumor and normal datasets. [The Markov Substitution Model for Evolution](https://genomebiology.biomedcentral.com/articles/10.1186/s13059-016-1029-6) utilized in this tool models the evolution of the reference allele to the allelic composition of the tumor and normal tissue at each genomic locus.
* [SvABA](https://github.com/walaj/svaba) This tool is especially useful for calling insertions and deletions (indels) because it assembles aberrantly aligned sequence reads that reflect indels or structural variants using a custom [String Graph Assembler](https://github.com/jts/sga). Indels can be difficult to detect with standard alignment-based variant callers.
* [Strelka2](https://github.com/Illumina/strelka) This is a small variant caller designed by Illumina. It is used for identifying germline variants in cohorts of samples and somatic variants in tumor/normal sample pairs.
* [SomaticSniper](https://gmt.genome.wustl.edu/packages/somatic-sniper/) SomaticSniper can be used to identify SNPs in tumor/normal pairs. It calculates the probability that the tumor and normal genotypes are different and reports this probability as a somatic score.
* [Pindel](https://www.sanger.ac.uk/tool/pindel/) Pindel is a tool that uses a pattern growth approach to detect breakpoints of large deletions, medium size insertion/inversion, tandem duplications.
* [Lancet](https://github.com/nygenome/lancet) This is a newer variant calling tool that uses colored de Bruijn graphs to jointly analyze tumor and normal pairs, offering strong indel detection. More information about the processes used in this variant calling tool can be found [here](https://www.nature.com/articles/s42003-018-0023-9)

Researchers may want to create a consensus file based on the mutation calls using multiple tools above. [OpenPBTA-analysis](https://github.com/AlexsLemonade/OpenPBTA-analysis/tree/master/analyses/snv-callers) shows an open source code example of how you might compare and contrast different SNV caller’s results.

## 7.10 Tools for variant calling annotation

These are beneficial for providing functional meaning to the mutational hits identified above.

* [Annovar](https://annovar.openbioinformatics.org/en/latest/) This is a helpful tool for annotating, filtering, and combining the output data from the above tools. It can be used for gene-based, region-based, or filter-based annotations.
* [GENCODE](https://www.gencodegenes.org/#) This tool can be used to identify and classify gene features in human and mouse genomes.
* [dbSNP](https://www.ncbi.nlm.nih.gov/snp/) This is a resource to look up specific human single nucleotide variations, microsatellites, and small-scale insertions and deletions.
* [Ensembl](https://uswest.ensembl.org/index.html) This resource is a genome browser for annotating genes from a wide variety of species.

## 7.11 Tools for copy number variation analysis

Similar to the mutation calling tools, many researchers will use several of these tools and investigate the overlapping hits seen with different copy number variant calling algorithms:

* [GATK](https://gatk.broadinstitute.org/hc/en-us/articles/360035531092--How-to-part-I-Sensitively-detect-copy-ratio-alterations-and-allelic-segments) GATK has a variety of tools that can be used to study changes in copy numbers of genes. This link provides a tutorial for how to use the tools.
* [AscatNGS](https://www.crick.ac.uk/research/labs/peter-van-loo/software) These tools (allele-specific copy number analysis of tumors) are specific for WGS copy number variation analysis. They can be used to dissect allele-specific copy numbers of tumors by estimating and adjusting for tumor ploidy and nonaberrant cell admixture.
* [TitanCNA](https://github.com/gavinha/TitanCNA) This tool is used to analyze copy number variation and loss of heterozygosity at the subclonal level for both WGS and WXS data in tumors compared to matched normals. It accounts for mixtures of cell populations and estimates the proportion of cells harboring each event. The Ha lab has developed a [snakemake pipeline](https://github.com/gavinha/TitanCNA/tree/master/scripts/snakemake) to more easily use this tool. [Ha et al. published a paper describing this tool in detail here](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4216928/)
* [gGNV](https://gatk.broadinstitute.org/hc/en-us/articles/4404604788891-GermlineCNVCaller) This is a germline CNV calling tool that can be used on both WGS and WXS data. This tool has booth COHORT and CASE modes. COHORT mode is used when providing a cohort of germline samples where CASE mode is used for individual samples. More details about these modes are described in the link above.
* [BIC-seq2](http://compbio.med.harvard.edu/BIC-seq/) This tool is used to detect CNVs with or without control samples. The steps involved in this data processing tool include normalization and CNV detection.

## 7.12 Tools for data visualization

These tools are often used in parallel to look at regions of the genome, develop plots, and create other relevant figures:

* [IGV](https://software.broadinstitute.org/software/igv/) IGV is an interactive tool used to easily visualize genomic data. It is available as a desktop application, web application, and JavaScript to embed in web pages. This application is very beneficial for visualizing both mutational and CNV data for WGS and WXS. IGV has many [tutorials on YouTube](https://www.youtube.com/channel/UCb5W5WqauDOwubZHb-IA_rA/featured) that are helpful for using the tool to its full potential.
* [Maftools](https://www.bioconductor.org/packages/devel/bioc/vignettes/maftools/inst/doc/maftools.html) Maftools is an R package that can be used to create informative plots from your WGS data output. It has tools to import both VCF files and ANNOVAR output for data analysis.
* [Prism](https://www.graphpad.com/scientific-software/prism/) Prism is a widely used tool in scientific research for organizing large datasets, generating plots, and creating readable figures. WGS or WXS data regarding mutations and CNV can be used as input for creating plots with this tool.

## 7.13 Resources for WGS

Online tutorials:

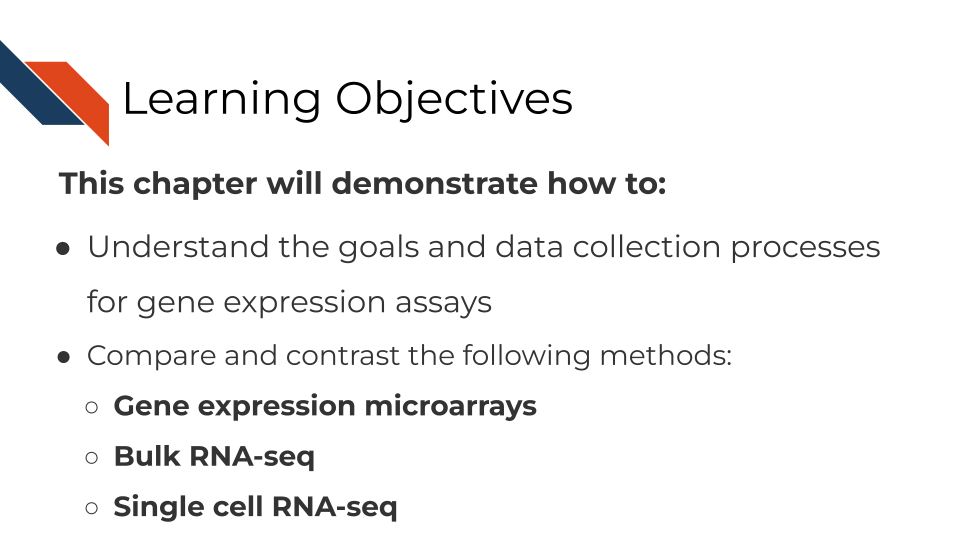
* [Galaxy tutorials](https://training.galaxyproject.org/training-material/topics/sequence-analysis/)
* [NCI resources](https://gdc.cancer.gov/about-data/gdc-data-processing/genomic-data-processing)
* [Bioinformaticsdotca tutorial](https://bioinformaticsdotca.github.io/CAN_2021_module4_lab)

Papers comparing analysis tools:

* ([Hwang et al. 2019](#ref-Hwang2019))
* ([Naj et al. 2019](#ref-Naj2019))
* ([He et al. 2020](#ref-He2020))

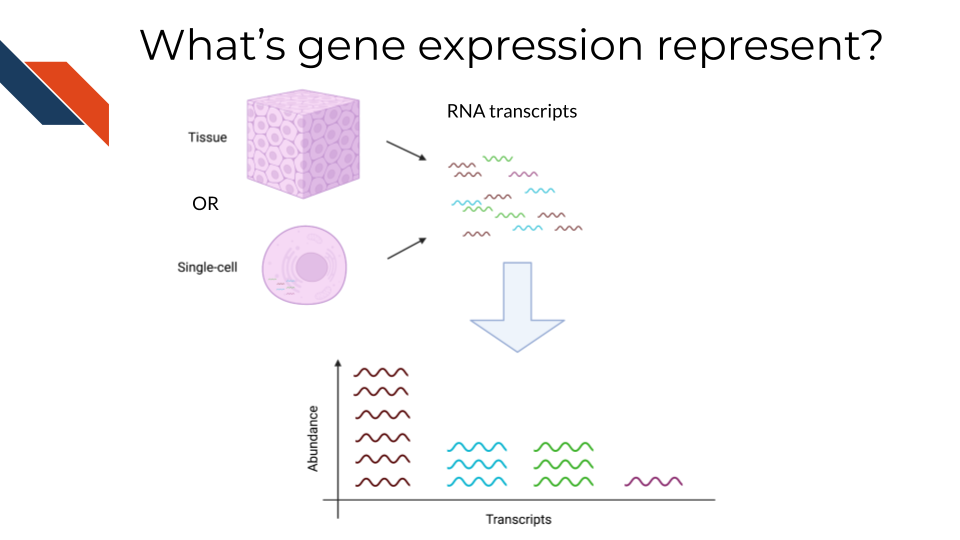
# 8 RNA Methods

## 8.1 Learning Objectives



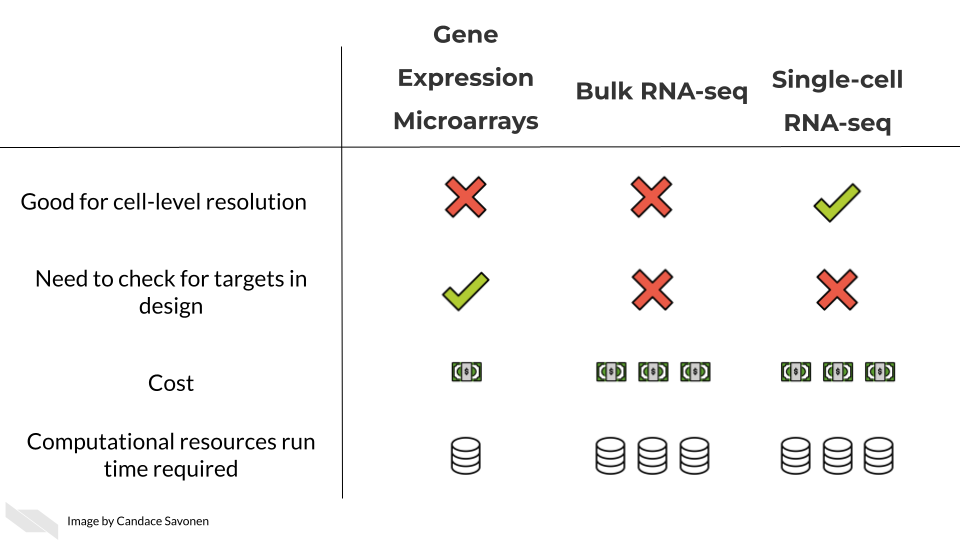
## 8.2 What are the goals of gene expression analysis?

The goal of gene expression analysis is to quantify RNAs across the genome. This can signify the extent to which various RNAs are being transcribed in a particular cell. This can be informative for what kinds of activity a cell is undergoing and responding to.



## 8.3 Comparison of RNA methods

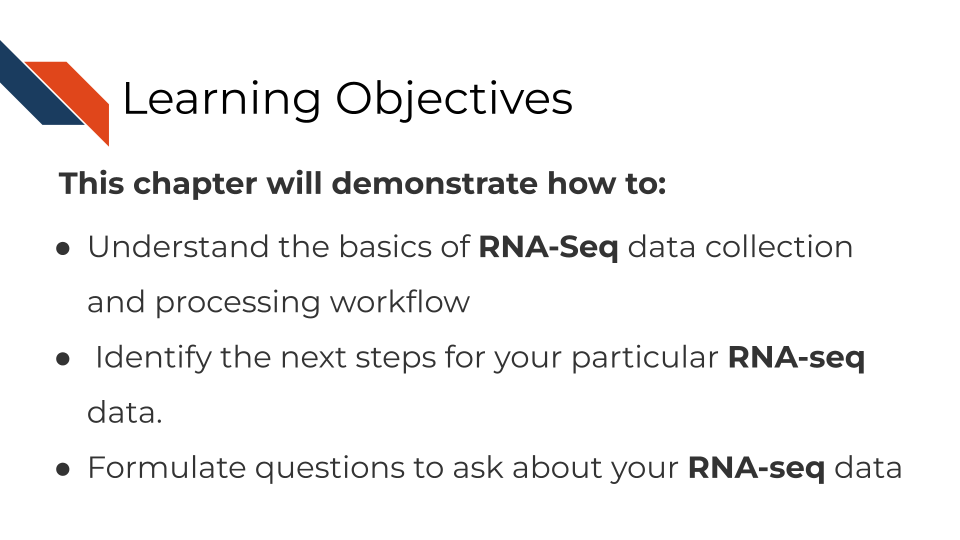
There are three general methods we will discuss for evaluating gene expression. RNA sequencing (whether bulk or single-cell) allows you to catch more targets than gene expression microarrays but is much more costly and computationally intensive. Gene expression microarrays have a lower dynamic range than RNA-seq generally but are much more cost effective.



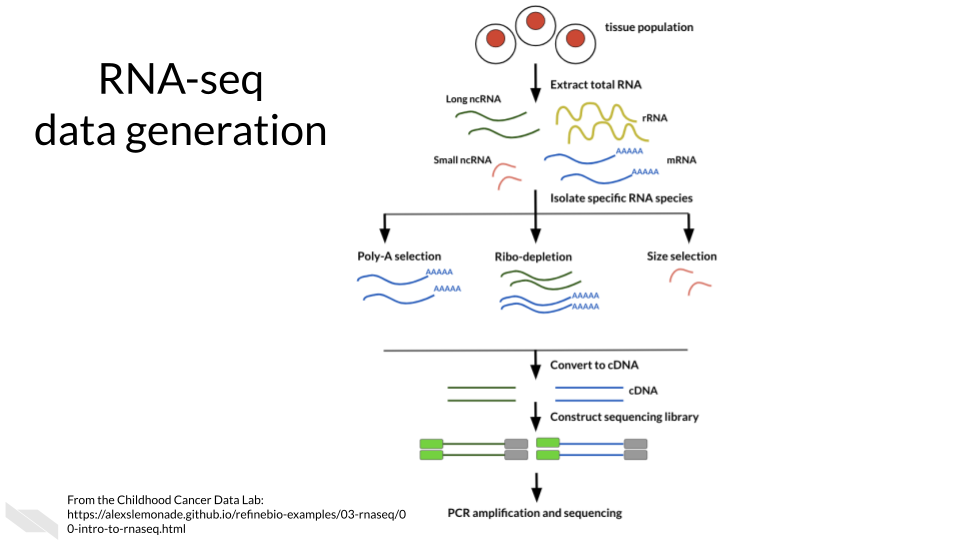
In these upcoming chapters we will discuss in more detail each of these methods, what the data represent, what you need to consider, and what resources you can consult for analyzing your data.

# 9 Bulk RNA-seq

## 9.1 Learning Objectives

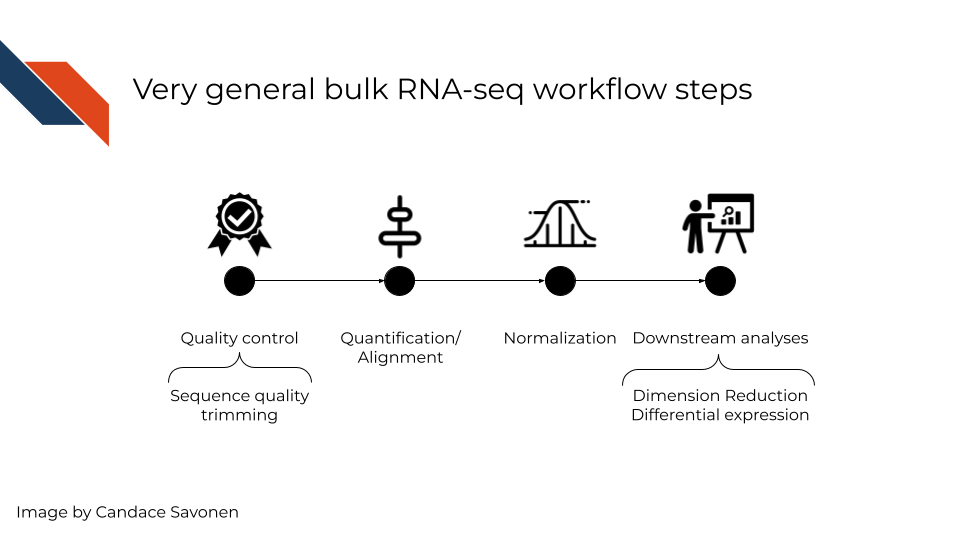


## 9.2 Where RNA-seq data comes from



## 9.3 RNA-seq workflow

In a very general sense, RNA-seq workflows involves first quantification/alignment. You will also need to conduct quality control steps that check the quality of the sequencing done. You may also want to trim and filter out data that is not trustworthy. After you have a set of reliable data, you need to normalize your data. After data has been normalized you are ready to conduct your downstream analyses. This will be highly dependent on the original goals and questions of your experiment. It may include dimension reduction, differential expression, or any number of other analyses.



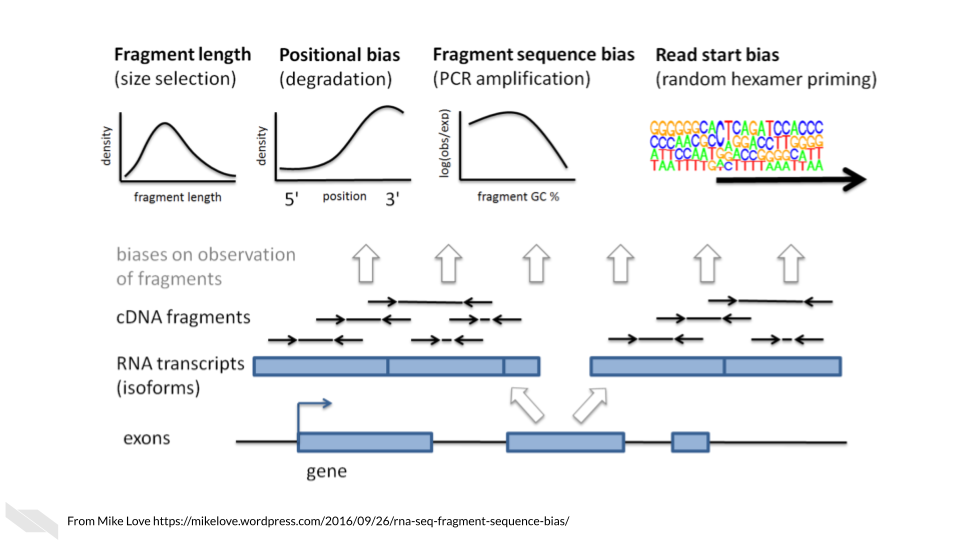
In this chapter we will highlight some of the more popular RNA-seq tools, that are generally suitable for most experiment data but there is no “one size fits all” for computational analysis of RNA-seq data ([Conesa et al. 2016](#ref-Conesa2016)). You may find tools out there that better suit your needs than the ones we discuss here.

## 9.4 RNA-seq data **strengths**

* RNA-seq can give you an idea of the transcriptional activity of a sample.
* RNA-seq has a more dynamic range of quantification than gene expression microarrays are able to measure.
* RNA-seq is able to be used for transcript discovery unlike gene expression microarrays.

## 9.5 RNA-seq data **limitations**

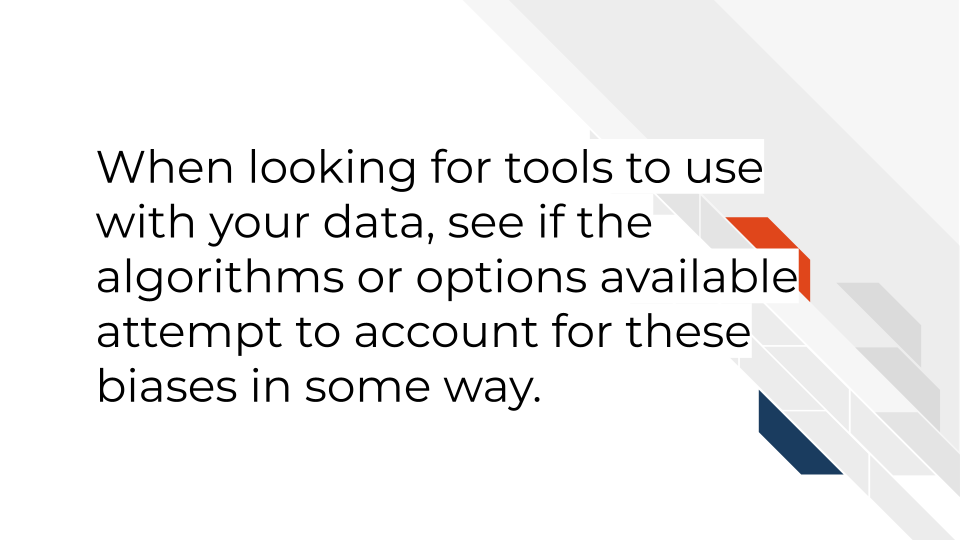
RNA-seq suffers from a lot of the common sequence biases which are further worsened by PCR amplification steps. We discussed some of the sequence biases in the previous sequencing chapter.



These biases are nicely covered in [this blog by Mike Love](https://mikelove.wordpress.com/2016/09/26/rna-seq-fragment-sequence-bias/) and we’ll summarize them here:

* **Fragment length**: Longer transcripts are more likely to be identified than shorter transcripts because there’s more material to pull from.
* **Positional bias**: 3’ ends of transcripts are more likely to be sequenced due to faster degradation of the 5’ end.
* **Fragment sequence bias**: The complexity and GC content of a sequence influences how often primers will bind to it (which influences PCR amplification steps as well as the sequencing itself).
* **Read start bias**: Certain reads are more likely to be bound by random hexamer primers than others.

*Main Takeaway*: When looking for tools, you will want to see if the algorithms or options available attempt to account for these biases in some way.



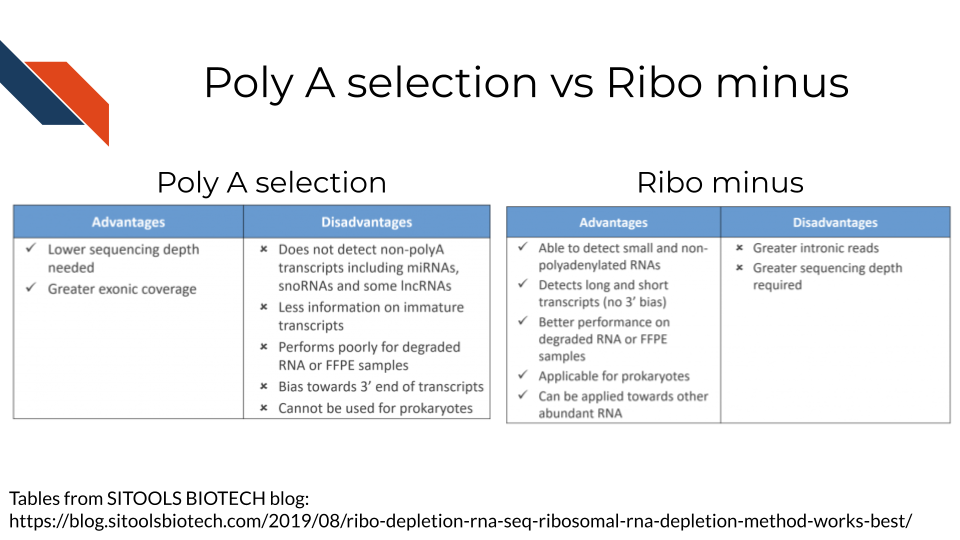
## 9.6 RNA-seq data considerations

### 9.6.1 Ribo minus vs poly A selection

Most of the RNA in the cell is not mRNA or noncoding RNAs of interest, but instead loads of ribosomal RNA a. So before you can prepare and sequence your data you need to isolate the RNAs to those you are interested in. There are two major methods to do this:

* **Poly A selection** - Keep only RNAs that have poly A tails – remember that mRNAs and some kinds of noncoding RNAs have poly A tails added to them after they are transcribed. A drawback of this method is that transcripts that are not generally polyadenylated: microRNAs, snoRNAs, certain long noncoding RNAs, or immature transcripts will be discarded. There is also generally a worse 3’ bias with this method since you are selecting based on poly A tails on the 3’ end.
* **Ribo-minus** - Subtract all the ribosomal RNA and be left with an RNA pool of interest. A drawback of this method is that you will need to use greater sequencing depths than you would with poly A selection (because there is more material in your resulting transcript pool).

[This blog by Sitools Biotech does a good summary](https://blog.sitoolsbiotech.com/2019/08/ribo-depletion-rna-seq-ribosomal-rna-depletion-method-works-best/) of the pros and cons of either selection method.

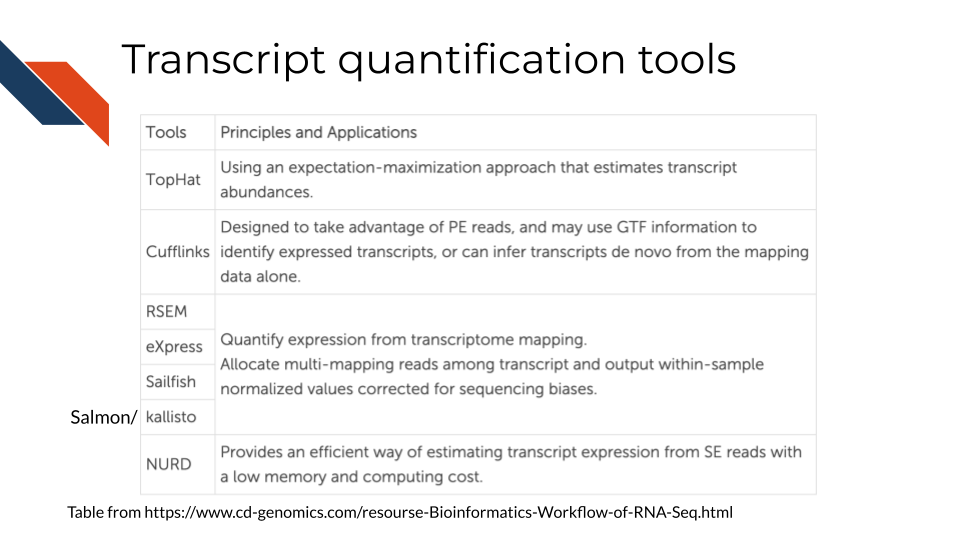


### 9.6.2 Transcriptome mapping

How do you know which read belongs to which transcript? This is where alignment comes into play for RNA-seq There are two major approaches we will discuss with examples of tools that employ them.

* **Traditional aligners** - Align your data to a reference using standard alignment algorithms. Can be very computationally intensive. Traditional alignment is the original approach to alignment which takes each read and finds where and how in the genome/transcriptome it aligns. If you are interested in identifying the intracacies of different splices and their boundaries, you may need to use one of these traditional alignment methods. But for common quantification purposes, you may want to look into pseudo alignment to save you time. *Examples of traditional aligners*:
  + [STAR](https://github.com/alexdobin/STAR/blob/master/doc/STARmanual.pdf)
  + [HISAT2](http://daehwankimlab.github.io/hisat2/manual/)
* This blog compares some of the traditional alignment tools
* [**Pseudo aligners**](https://tinyheero.github.io/2015/09/02/pseudoalignments-kallisto.html) - much faster and the trade off for accuracy is often negligible (but as always, this is likely dependent on the data you are using). The biggest drawback to pseudoaligners is that if you care about local alignment (e.g. perhaps where splice boundaries occur) instead of just transcript identification then a traditional alignment may be better for your purposes. These pseudo aligners often include a verification step where they compare a subset of the data to its performance to a traditional aligner (and for most purposes they usually perform well). Pseudo aligners can potentially save you hours/days/weeks of processing time as compared to traditional aligners so are worth looking into. *Examples of pseudo aligners*:
  + [Salmon](https://salmon.readthedocs.io/en/latest/salmon.html#using-salmon)
  + [Kallisto](https://pachterlab.github.io/kallisto/)
* **Reference free assembly** - The first two methods we’ve discussed employ aligning to a reference genome or transcriptome. But alternatively, if you are much more interested in transcript identification or you are working with a model organism that doesn’t have a well characterized reference genome/transcriptome, then de novo assembly is another approach to take. As you may suspect, this is the most computationally demanding approach and also requires deeper sequencing depth than alignment to a reference. But depending on your goals, this may be your preferred option.

These strategies are discussed at greater length [in this excellent manuscript by Conesa et al, 2016](https://genomebiology.biomedcentral.com/articles/10.1186/s13059-016-0881-8).



### 9.6.3 Abundance measures

If your RNA-seq data has already been processed, it may have abundance measure reported with it already. But there are various types of abundance measures used – what do they represent?

* **raw counts** - this is a raw number of how many times a transcript was counted in a sample.

Two considerations to think of:  
1. **Library sizes**: Raw counts does not account for differences between samples’ library sizes. In other words, how many reads were obtained from each sample? Because library sizes are not perfectly equal amongst samples and not necessarily biologically relevant, its important to account for this if you wish to compare different samples in your set.  
2. **Gene length**: Raw counts also do not account for differences in gene length (remember how we discussed longer transcripts are more likely to be counted).

Because of these items, some sort of transformation needs to be done on the raw counts before you can interpret your data.

These other abundance measures attempt to account for library sizes and gene length. [This blog and video by StatQuest](https://www.rna-seqblog.com/rpkm-fpkm-and-tpm-clearly-explained/) does an excellent job summarizing the differences between these quantifications and we will quote from them:

* **Reads per kilobase million (RPKM)**

1. Count up the total reads in a sample and divide that number by 1,000,000 – this is our “per million” scaling factor.
2. Divide the read counts by the “per million” scaling factor. This normalizes for sequencing depth, giving you reads per million (RPM)
3. Divide the RPM values by the length of the gene, in kilobases. This gives you RPKM.

* **Fragments per kilobase million (FPKM)**

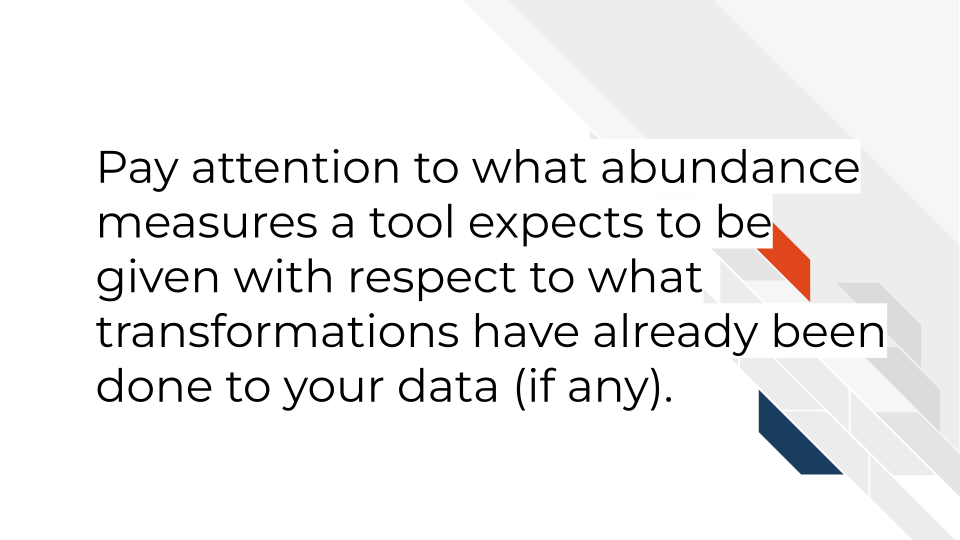
FPKM is very similar to RPKM. RPKM was made for single-end RNA-seq, where every read corresponded to a single fragment that was sequenced. FPKM was made for paired-end RNA-seq. With paired-end RNA-seq, two reads can correspond to a single fragment, or, if one read in the pair did not map, one read can correspond to a single fragment. The only difference between RPKM and FPKM is that FPKM takes into account that two reads can map to one fragment (and so it doesn’t count this fragment twice).

* **Transcripts per million (TPM)**

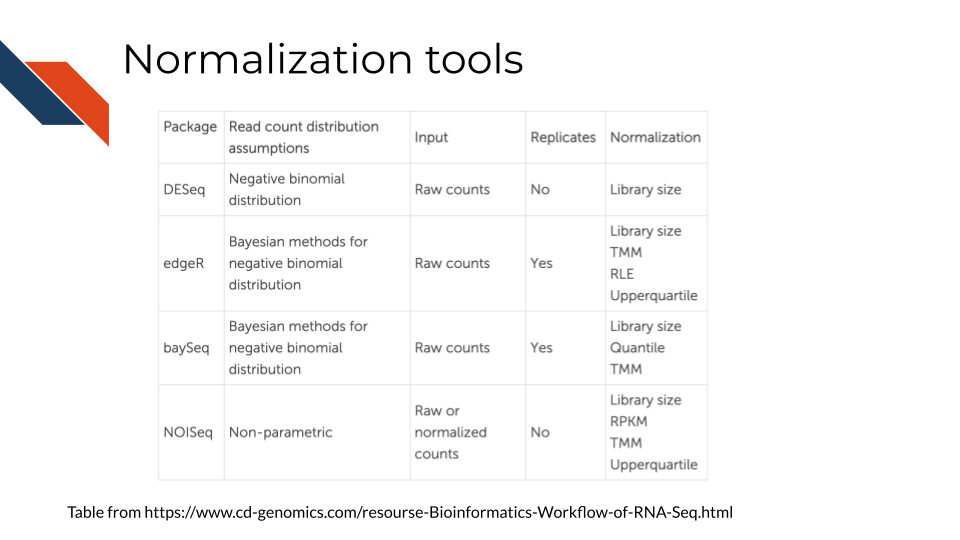
1. Divide the read counts by the length of each gene in kilobases. This gives you reads per kilobase (RPK).
2. Count up all the RPK values in a sample and divide this number by 1,000,000. This is your “per million” scaling factor.
3. Divide the RPK values by the “per million” scaling factor. This gives you TPM.

TPM has gained a popularity in recent years because it is more intuitive to understand:

When you use TPM, the sum of all TPMs in each sample are the same. This makes it easier to compare the proportion of reads that mapped to a gene in each sample. In contrast, with RPKM and FPKM, the sum of the normalized reads in each sample may be different, and this makes it harder to compare samples directly.



### 9.6.4 RNA-seq downstream analysis tools

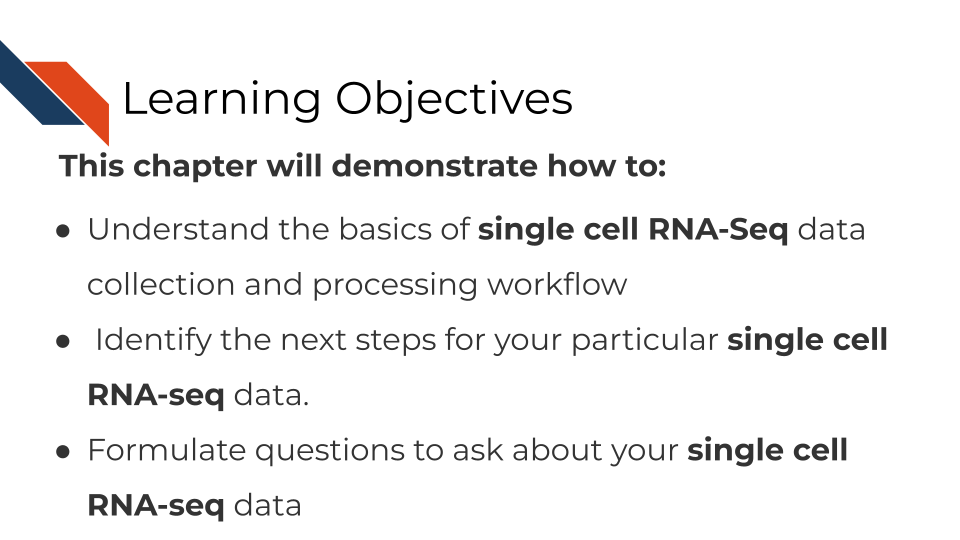


## 9.7 More reading about RNA-seq data

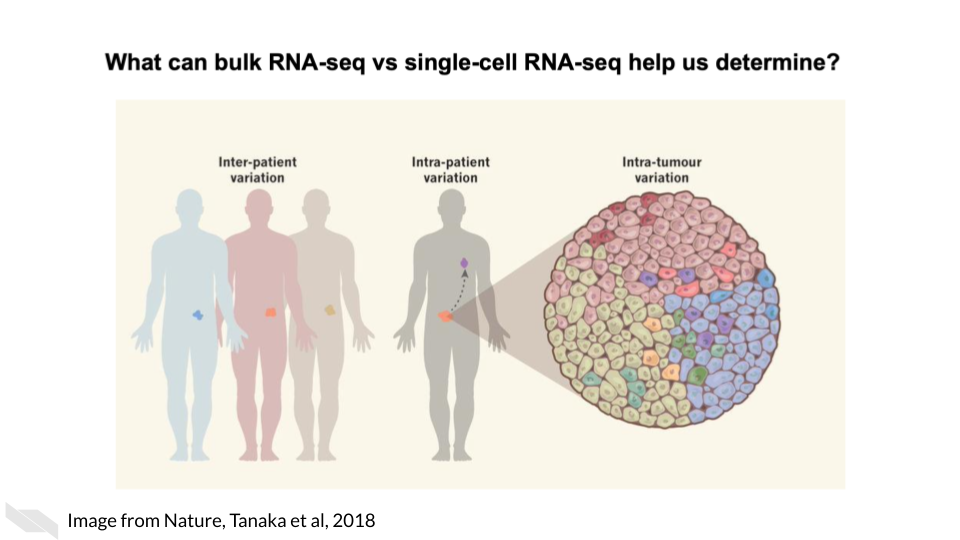
* [Refine.bio’s introduction to RNA-seq](https://alexslemonade.github.io/refinebio-examples/03-rnaseq/00-intro-to-rnaseq.html)
* [StatQuest: A gentle introduction to RNA-seq](https://www.youtube.com/watch?v=tlf6wYJrwKY) ([**Starmer2017-rnaseq?**](#ref-Starmer2017-rnaseq)).
* [A general background on the wet lab methods of RNA-seq](https://bitesizebio.com/13542/what-everyone-should-know-about-rna-seq/) ([**Hadfield2016?**](#ref-Hadfield2016)).
* [Modeling of RNA-seq fragment sequence bias reduces systematic errors in transcript abundance estimation](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5143225/) ([**Love2016?**](#ref-Love2016)).
* [Mike Love blog post about sequencing biases](https://mikelove.wordpress.com/2016/09/26/rna-seq-fragment-sequence-bias/) ([**bias-blog?**](#ref-bias-blog))
* [Biases in Illumina transcriptome sequencing caused by random hexamer priming](https://pdfs.semanticscholar.org/9d16/997f5de72d6c606fef3d673db70e5d1d8e1e.pdf?_ga=2.131436679.965169313.1600175795-124991789.1600175795) ([**Hansen2010?**](#ref-Hansen2010)).
* [Computation for RNA-seq and ChIP-seq studies](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4121056/) ([**Pepke2009?**](#ref-Pepke2009)).

# 10 Single-cell RNA-seq

## 10.1 Learning Objectives

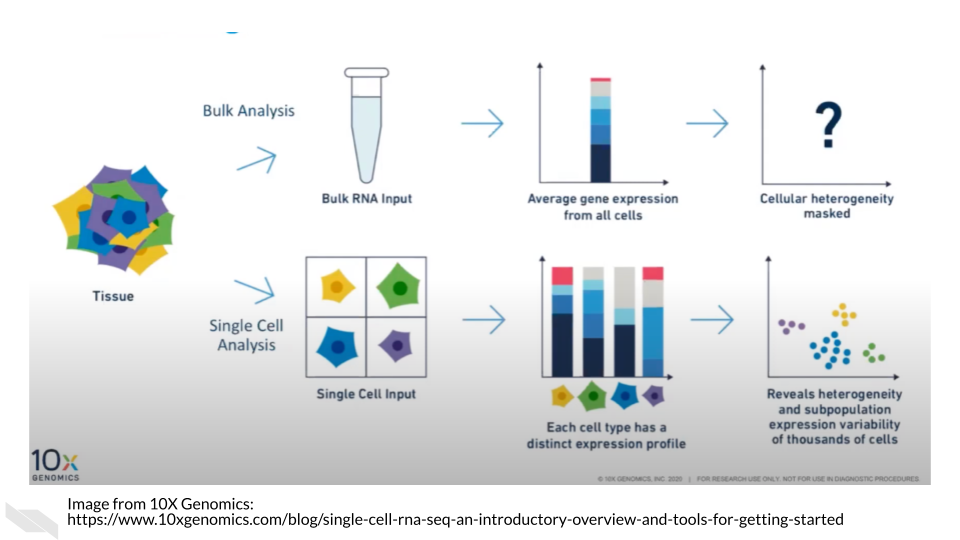


## 10.2 Where single-cell RNA-seq data comes from



As opposed to bulk RNA-seq which can only tell us about tissue level and within patient variation, single-cell RNA-seq is able to tell us cell to cell variation in transcriptomics including intra-tumor heterogeneity.

Single cell RNA-seq can give us cell level transcriptional profiles. Whereas bulk RNA-seq masks cell to cell heterogeneity. If your research questions require cell-level transcriptional information, single-cell RNA-seq will on interest to you.



## 10.3 Single-cell RNA-seq data types

There are broadly two categories of single-cell RNA-seq data methods we will discuss.

* **Full length RNA-seq**: Individual cells are physically separated and then sequenced.
* **Tag Based RNA-seq**: Individual cells are tagged with a barcode and their data is separated computationally.

Depending on your goals for your single cell RNA-seq analysis, you may want to choose one method over the other.

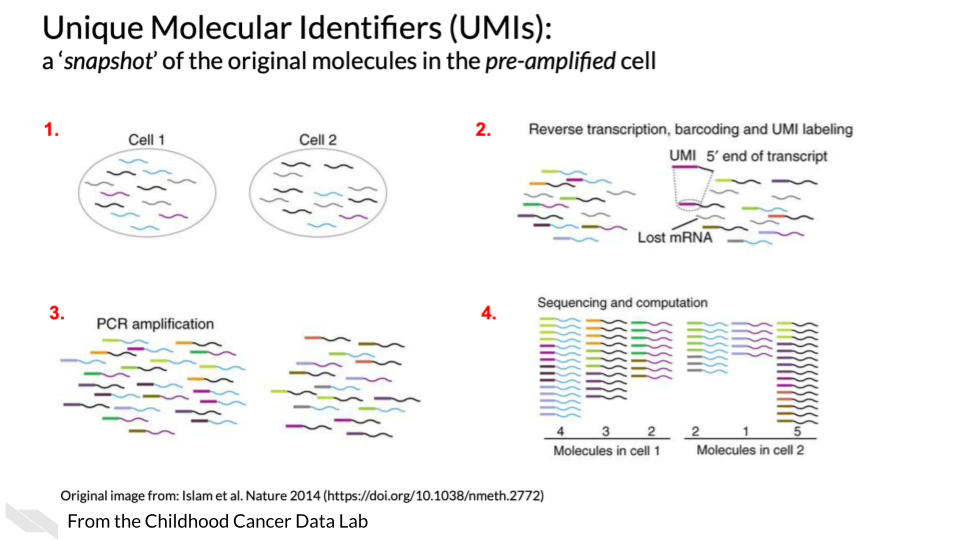




(Material borrowed from ([“Alex’s Lemonade Training Modules” 2022](#ref-AlexsLemonade2022))).

### 10.3.1 Unique Molecular identifiers

Often Tag based single cell RNA-seq methods will include not only a cell barcode for cell identification but will also have a unique molecular identifier (UMI) for original molecule identification. The idea behind the UMIs is it is a way to have insight into the original snapshot of the cell and potentially combat PCR amplification biases.



## 10.4 Single cell RNA-seq tools

There are a lot of scRNA-seq tools for various steps along the way.



In a very general sense, single cell RNA-seq workflows involves first quantification/alignment. You will also need to conduct quality control steps that may involve using UMIs to check for what’s detected, detecting duplets, and using this information to filter out data that is not trustworthy. After you have a set of reliable data, you need to normalize your data. Single cell data is highly skewed - a lot of genes barely or not detected and a few genes that are detected a lot. After data has been normalized you are ready to conduct your downstream analyses. This will be highly dependent on the original goals and questions of your experiment. It may include dimension reduction, cell classification, differential expression, detecting cell trajectories or any number of other analyses.

Each step of this very general representation of a workflow can be conducted by a variety of tools. We will highlight some of the more popular tools here. But, to look through a full list, you can consult the [scRNA-tools website](https://www.scrna-tools.org/table).

## 10.5 Quantification/Alignment

TODO: How to choose between different quantification tools

* [Alevin](https://salmon.readthedocs.io/en/latest/alevin.html)
* [CellRanger](https://support.10xgenomics.com/single-cell-gene-expression/software/pipelines/latest/what-is-cell-ranger)
* [Kallisto bustools](https://www.kallistobus.tools/)

## 10.6 Quality control

TODO: More about considerations for quality control

#### 10.6.0.1 Checking UMIs

* [AlevinQC](https://bioconductor.org/packages/release/bioc/html/alevinQC.html)

#### 10.6.0.2 Checking for doublets

* [scDblFinder](https://bioconductor.org/packages/release/bioc/html/scDblFinder.html)
* [DoubletDetection](https://doubletdetection.readthedocs.io/en/stable/)

## 10.7 Normalization and downstream analyses

TODO: More about considerations for downstream analysis and normalization

* [Seurat](https://satijalab.org/seurat/)
* [Single Cell Genome Viewer](https://github.com/KrasnitzLab/SCGV)
* For normalization [scater](https://bioconductor.org/packages/devel/bioc/vignettes/scater/inst/doc/overview.html)
* [scanpy](https://scanpy.readthedocs.io/en/stable/)

## 10.8 Useful tutorials

These tutorials cover explicit steps, code, tool recommendations and other considerations for analyzing RNA-seq data.

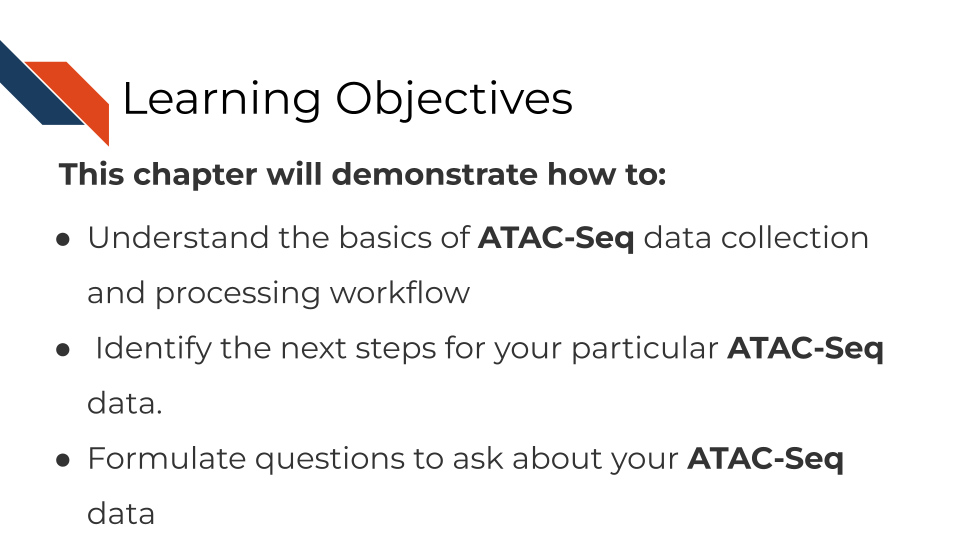
* [Orchestrating Single Cell Analysis with Bioconductor](http://bioconductor.org/books/3.15/OSCA.intro/) - An excellent tutorial for processing single cell data using Bioconductor.
* [Advanced Single Cell Analysis with Bioconductor](http://bioconductor.org/books/3.15/OSCA.advanced/) - a companion book to the intro version that contains code examples.
* [Alex’s Lemonade scRNA-seq Training module](https://alexslemonade.github.io/training-modules/scRNA-seq/) - A cancer based workshop module based in R, with exercise notebooks.
* [Sanger Single Cell Course](https://www.singlecellcourse.org/) - a general tutorial based on using R.
* [ASAP: Automated Single-cell Analysis Pipeline](https://asap.epfl.ch/) is a web server that allows you to process scRNA-seq data.
* [Processing raw 10X Genomics single-cell RNA-seq data (with cellranger)](https://swaruplab.bio.uci.edu/tutorial/cellranger/cellranger-rna.html) - a tutorial based on using CellRanger.

## 10.9 Useful readings

* [An Introduction to the Analysis of Single-Cell RNA-Sequencing Data](https://doi.org/10.1016/j.omtm.2018.07.003) ([**AlJanahi2018?**](#ref-AlJanahi2018)).
* [Orchestrating single-cell analysis with Bioconductor](https://www.nature.com/articles/s41592-019-0654-x) ([**Amezquita2019?**](#ref-Amezquita2019)).
* [UMIs the problem, the solution and the proof](https://cgatoxford.wordpress.com/2015/08/14/unique-molecular-identifiers-the-problem-the-solution-and-the-proof/) ([Smith 2015](#ref-Smith2015)).
* [Experimental design for single-cell RNA sequencing](https://doi.org/10.1093/bfgp/elx035) ([Baran-Gale, Chandra, and Kirschner 2018](#ref-BaranGale2018)).
* [Tutorial: guidelines for the experimental design of single-cell RNA sequencing studies](https://doi.org/10.1038/s41596-018-0073-y) ([**Lafzi2019?**](#ref-Lafzi2019)).
* [Comparative Analysis of Single-Cell RNA Sequencing Methods](http://dx.doi.org/10.1016/j.molcel.2017.01.023) ([**Ziegenhain2018?**](#ref-Ziegenhain2018)).
* [Comparative Analysis of Droplet-Based Ultra-High-Throughput Single-Cell RNA-Seq Systems](https://doi.org/10.1016/j.molcel.2018.10.020) ([**Zhang2018?**](#ref-Zhang2018)).
* [Single cells make big data: New challenges and opportunities in transcriptomics](http://dx.doi.org/10.1016/j.coisb.2017.07.004) ([Angerer et al. 2017](#ref-Angerer2017)).
* [Comparative Analysis of common alignment tools for single cell RNA sequencing](https://www.biorxiv.org/content/10.1101/2021.02.15.430948v2) ([Brüning et al. 2021](#ref-Bruning2021)).
* [Current best practices in single-cell RNA-seq analysis: a tutorial](https://doi.org/10.15252/msb.20188746) ([Luecken and Theis 2019](#ref-Luecken2019)).

# 11 ATAC-Seq

## 11.1 Learning Objectives



## 11.2 What are the goals of ATAC-Seq analysis?

The goals of ATAC-seq are to identify the accessible regions of the genome in a particular set of samples. These data allow us to understand the relationships between the chromatin accessibility patterns and cell states, and to understand the mechanistic causes and consequences of these chromatin accessibility patterns.

ATAC-seq data is generated by fragmenting the genome with the Tn5 endonuclease and sequencing the shorter DNA fragments. While most of the genome is associated with protein complexes that preclude the digestion of DNA by Tn5, some regions of the genome have accessible chromatin that can be cleaved by Tn5 resulting in short (<500bp) fragments. These regions of the genome are of biological interest as they are likely to harbor transcription factor binding sites and to constitute cis-regulatory elements, genomic regions that are involved in the regulation of gene expression.



## 11.3 ATAC-Seq general workflow overview

A basic ATAC-seq workflow involves mapping sequence reads to the genome, identifying peaks, assessing data quality, and identifying patterns of interest through clustering or identification of differentially accessible regions or other statistical means.

###Data quality metrics:

#### 11.3.0.1 Number of mapped reads

As for all DNA-sequencing based genomics technologies, a sufficient number of mapped reads is required to obtain meaningful results from a sample. You can read more about [general sequencing technologies in our previous chapter here](http://hutchdatascience.org/Choosing_Genomics_Tools/sequencing-data.html). For experiments on human samples this number should be greater than 20 million mapped unique reads.

#### 11.3.0.2 Fragment size distribution:

ATAC-seq data is often generated using paired end sequencing technologies, which allow for characterization of ATAC-seq fragments. Histograms of these distributions using single base pair resolution bins reveal patterns of enrichment relative to the nucleosome scale of 147bp and the DNA-helix scale ~10.5bp.

#### 11.3.0.3 Number of peaks:

Although the number of accessible chromatin regions can vary from one cell type to another, there are several regions that appear to be constitutively accessible across a most cell types. At least 20,000 peaks can be identified in a high quality experiment.

#### 11.3.0.4 FRiP score (fraction of reads in peaks)

In high quality ATAC-seq data a large fraction of reads overlap with peaks, while in low quality data there is a high level of fragments that map to background regions.

#### 11.3.0.5 Overlap with other chromatin accessibility data

Thousands of ATAC-seq samples have been produced in human and mouse. High quality ATAC-seq data will share a substantial proportion of peaks with many of these datasets.

#### 11.3.0.6 Overlap with promoters

The promoter regions of many genes are constitutively accessible. Examining peak overlap with regions close to known protein coding gene transcription start sites can be used as a check for data quality.

## 11.4 ATAC-Seq data **strengths**:

* ATAC-seq is broadly used by many laboratories to characterize accessible chromatin in cell lines or sorted cells derived from tissues.
* In principle, ATAC-seq can identify a large proportion of cis-regulatory elements.
* In contrast to ChIP-seq, ATAC-seq does not require specific antibodies, or knowledge of the important transcription factors in a system.

## 11.5 ATAC-Seq data **limitations**:

* ATAC-seq does not precisely identify the transcription factors or other chromatin associated factors that bind in or around chromatin accessible regions. This type of information needs to be inferred through analysis of transcription factor binding motif analysis or ChIP-seq data.
* Accessible regions are not necessarily cis-regulatory regions, although many of them are.
* The genes that are regulated by cis-regulatory elements cannot be identified conclusively by ATAC-seq alone.
* ATAC-seq data can be biased making direct comparisons difficult.
* ATAC-seq can be affected by batch effects.

## 11.6 ATAC-Seq data considerations

The nucleosome is the fundamental unit of chromatin packaging in the genome and nucleosomal DNA is far less likely to be cleaved by the Tn5 nuclease than linker DNA. When DNA is fragmented by Tn5 the positions of the endpoints relative to the nucleosomes is an important consideration. When the ends are less than 147bp apart it is likely that both ends originate from the same linker region. Longer fragments can result from cuts on opposite sides of the same nucleosome, or even opposite sides of a genomic interval that encompasses multiple nucleosomes. The short fragments are therefore most likely to be nucleosome free and provide stronger evidence for transcription factor binding sites.

As will other genomics protocols, ATAC-seq data is subject to biases introduced in the ATAC-seq protocol and in the sequencing itself. Comparison of ATAC-seq data generated in different batches, by different laboratories or using different protocols might not be directly comparable. In addition, the Tn5 endonuclease does have biases in the precise DNA sequences it can cut. This should be taken into consideration when carrying out base pair resolution analyses including footprinting analysis and analysis of the effects of sequence variants on chromatin accessibility.

Read depth will impact ATAC-seq signal, but enzyme strength and conditions can also alter the distribution of cuts.

When using ATAC-seq data to answer biological questions it is important to understand what types of bias could impact the results. To ensure valid results the analysis needs to use appropriate statistical methods, ensure enough high quality ATAC-seq data is available, including controls, and possibly reframing the questions.

## 11.7 ATAC-seq analysis tools

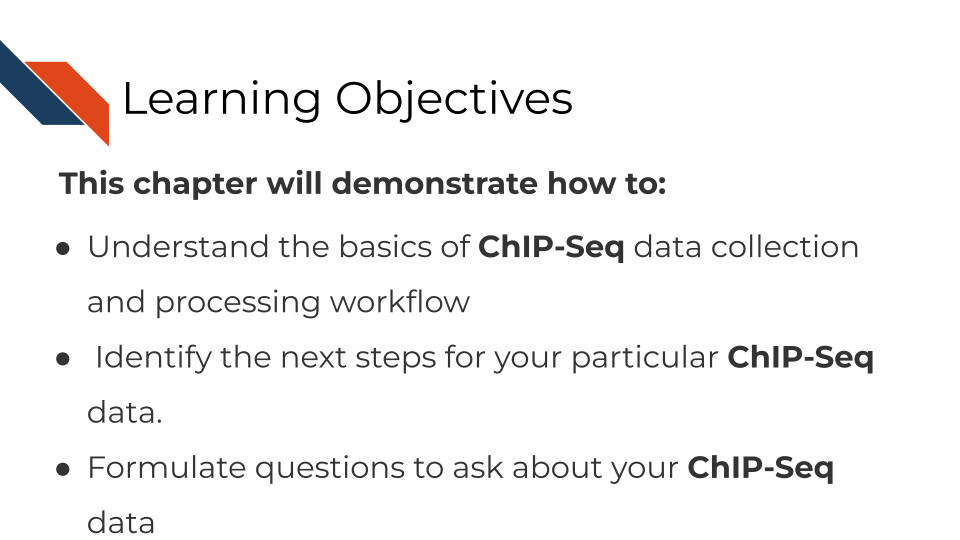
* [A Galaxy based tutorial for ATAC-seq](https://training.galaxyproject.org/training-material/topics/epigenetics/tutorials/atac-seq/tutorial.html) - Galaxy is a good recommendation for those new to informatics who would like a cloud-based GUI option to use for the analysis of their data.
* [MACS - Model-based analysis for ChIP-Seq](https://macs3-project.github.io/MACS/) - A command line tool for the identification of transcription factor binding sites. Can be used with ChIP-seq or ATAC-seq.
* [CHIPS](https://github.com/liulab-dfci/CHIPS) - A Snakemake pipeline for quality control and reproducible processing of chromatin profiling data. This tool will require some snakemake and coding knowledge. For more recommendations about coding see our later chapter about general data analysis tools.
* [Cistrome DB](http://cistrome.org/db/#/) - a visual tool to allow you to browse your ATAC-seq data.
* [SELMA - Simplex Encoded Linear Model for Accessible Chromatin](https://github.com/zang-lab/SELMA) - SELMA is a python based tool for the assessment of biases in Chromatin based data.

## 11.8 More resources about ATAC-seq data

* [ATAC-seq overview from Galaxy](https://training.galaxyproject.org/training-material/topics/epigenetics/tutorials/atac-seq/slides.html#1) - these slides explain the overarching concepts of ATAC-seq.
* [ATAC seq guidelines from Harvard](https://informatics.fas.harvard.edu/atac-seq-guidelines.html) - this workflow runs through step by step how to analysis ATAC-seq data and what different parameters mean.
* [ATAC-seq review](https://genomebiology.biomedcentral.com/articles/10.1186/s13059-020-1929-3) - this paper gives a great overview of ATAC-seq data and step by step what needs to be considered.
* [Identifying and mitigating bias in chromatin](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4473780/)
* [CHIP Snakemake pipeline for analyzing ChIP-seq and chromatin accessibility data](https://f1000research.com/articles/10-517)
* [Paper on bias in DNase-seq footprinting analysis and fragment size effects, similar comments apply to ATAC-seq](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4018771/)
* [SELMA Method for evaluating footprint bias in ATAC-seq](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9492688/)

# 12 ChIP-Seq

## 12.1 Learning Objectives



## 12.2 What are the goals of ChIP-Seq analysis?

The goal of ChIP-Seq is to….



## 12.3 ChIP-Seq general workflow overview

## 12.4 ChIP-Seq data **strengths**:

## 12.5 ChIP-Seq data **limitations**:

## 12.6 ChIP-Seq data considerations

## 12.7 ChiP-seq analysis tools

## 12.8 More resources about ChiP-seq data

# 13 DNA Methylation Sequencing

## 13.1 Learning Objectives



## 13.2 What are the goals of analyzing DNA methylation?

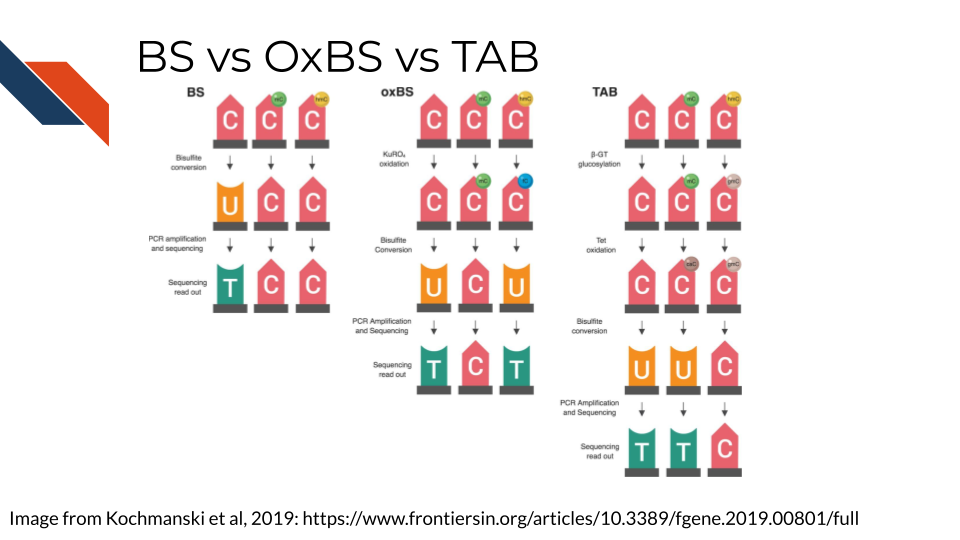


To detect methylated cytosines (5mC), DNA samples are prepped using bisulfite (BS) conversion. This converts unmethylated cytosines into uracils and leaves methylated cytosines untouched. Probes are then designed to bind to either the uracil or the cytosine, representing the unmethylated and methylated cytosines respectively.

For a given sample, you will obtain a fraction, known as the Beta value, that indicates the relative abundance of the methylated and unmethylated versions of the sequence. Beta values exist then on a scale of 0 to 1 where 0 indicates none of this particular base is methylated in the sample and 1 indicates all are methylated.

Note that bisulfite conversion alone will not distinguish between 5mC and 5hmC though these often may indicate different biological mechanics.

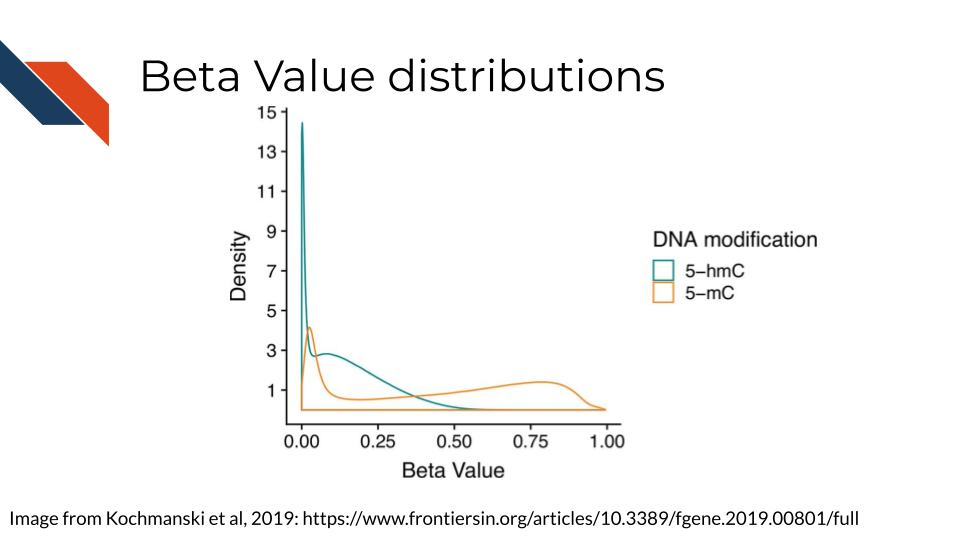
Additionally, 5-hydroxymethylated cytosines (5hmC) can also be detected by oxidative bisulfite sequencing (OxBS) [Booth et al. ([2013](#ref-Booth2013)). oxidative bisulfite conversion measures both 5mC and 5hmC. If you want to identify 5hmC bases you either have to pair oxBS data with BS data OR you have to use Tet-assisted bisulfite (TAB) sequencing which will exclusively tag 5hmC bases ([Yu et al. 2012](#ref-Yu2012)).



## 13.3 Methylation data considerations

### 13.3.1 Beta values binomially distributed

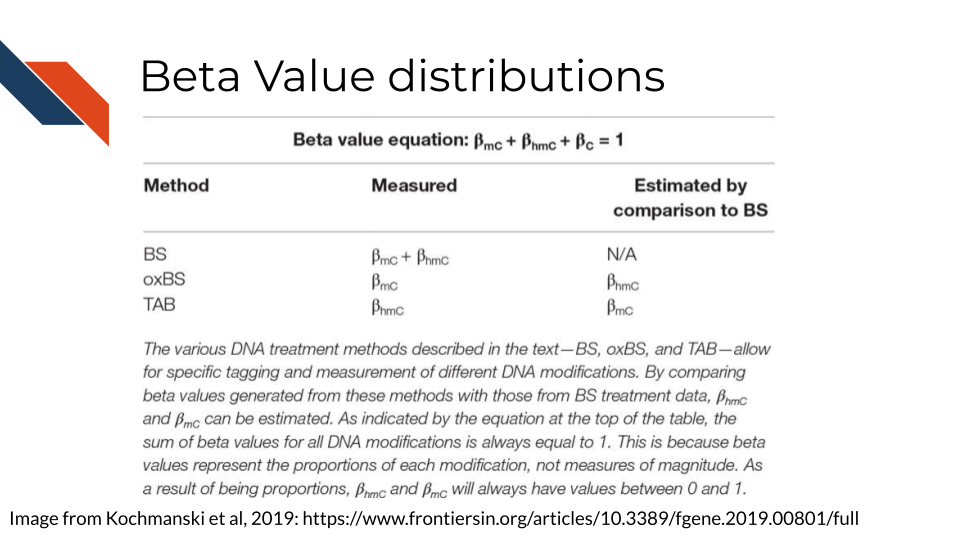
Because beta values are a ratio, by their nature, they are not normally distributed data and should be treated appropriately. This means data models (like those used by the limma package) built for RNA-seq data should not be used on methylation data. More accurately, Beta values follow a binomial distribution.



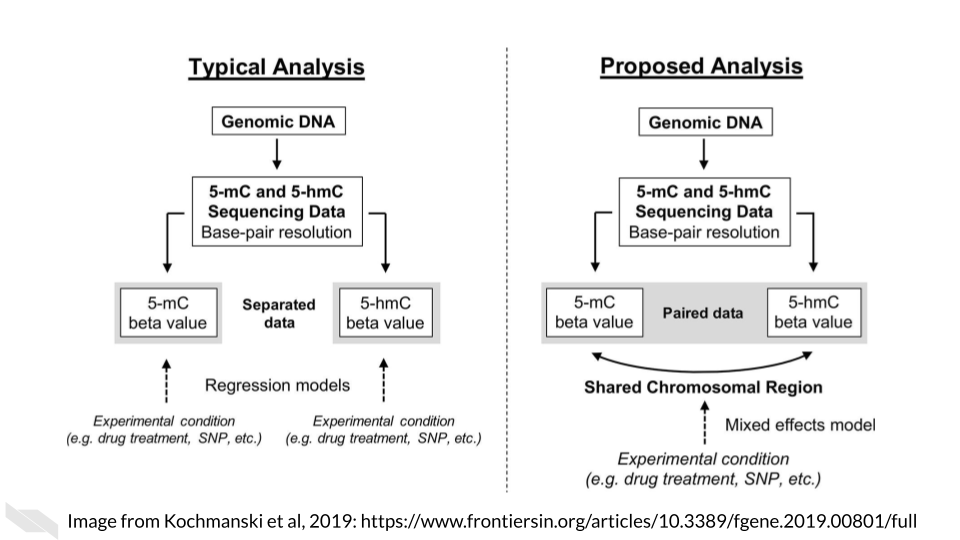
This generally involves applying a generalized linear model.

### 13.3.2 Measuring 5mC and/or 5hmC

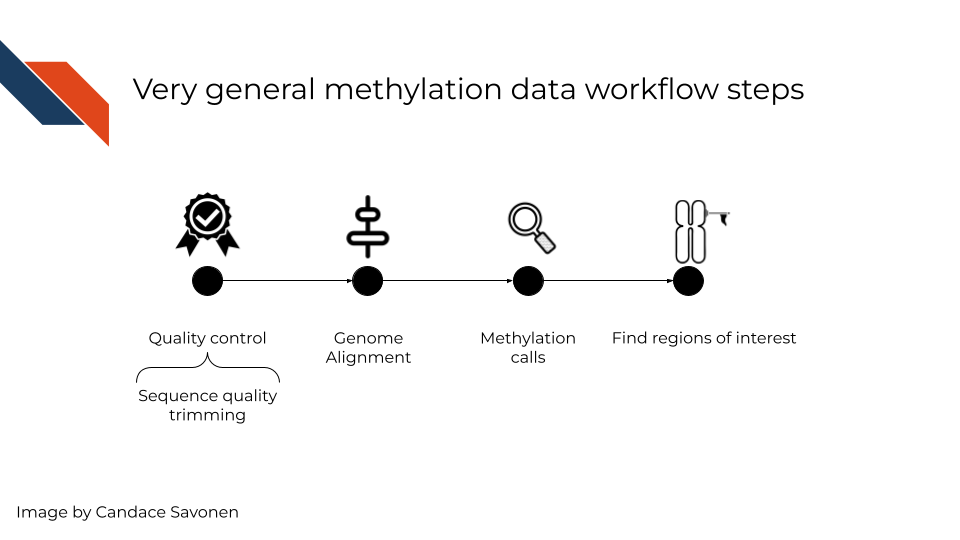
If your data and questions are interested in both 5mC and 5hmC, you will have separate sequencing datasets for each sample for both the BS and OBS processed samples. 5mC is often a step toward 5hmC conversion and therefore the 5mC and 5hmC measurements are, by nature, not independent from each other. In theory, 5mC, 5hmC and unmethylated cytosines should add up to 1.



Because of this, its been proposed that the most appropriate way to model these data is to combine them together in a model ([Kochmanski, Savonen, and Bernstein 2019](#ref-Kochmanski2019)).



## 13.4 Methylation data workflow



Like other sequencing methods, you will first need to start by quality control checks. Next, you will also need to align your sequences to the genome. Then, using the base calls, you will need to make methylation calls – which are methylated and which are not. This details of step depends on whether you are measuring 5mC and/or 5hmC methylation calls. Lastly, you will likely want to use your methylation calls as a whole to identify differentially methylated regions of interest.

## 13.5 Methylation tools!

### 13.5.1 Quality control

TODO: How should this be the same or different from general sequencing quality control checks? - [FASTQC](https://www.bioinformatics.babraham.ac.uk/projects/fastqc/)

### 13.5.2 Genome Alignment

TODO: How should this be the same or different from general sequencing alignment?

### 13.5.3 Methylation calls

TODO: What other packages/tools should be mentioned here?

* [MethylKit](https://bioconductor.org/packages/release/bioc/html/methylKit.html) - <https://compgenomr.github.io/book/data-filtering-and-exploratory-analysis.html>

### 13.5.4 Find regions of interest!

<https://compgenomr.github.io/book/extracting-interesting-regions-differential-methylation-and-segmentation.html>

### 13.5.5 Annotation of regions of interest

TODO: How does this differ from annotating genomic regions in general?

## 13.6 More resources

* [DNA methylation analysis with Galaxy tutorial](https://training.galaxyproject.org/training-material/topics/epigenetics/tutorials/methylation-seq/tutorial.html)
* [The mint pipeline](https://github.com/sartorlab/mint/blob/master/README.md) for analyzing methylation and hydroxymethylation data.
* [Book chapter about finding methylation regions of interest](https://compgenomr.github.io/book/extracting-interesting-regions-differential-methylation-and-segmentation.html)

# 14 General Data Analysis Tools

## 14.1 Learning Objectives

## 14.2 Command Line vs GUI

### 14.2.1 Bash

### 14.2.2 R

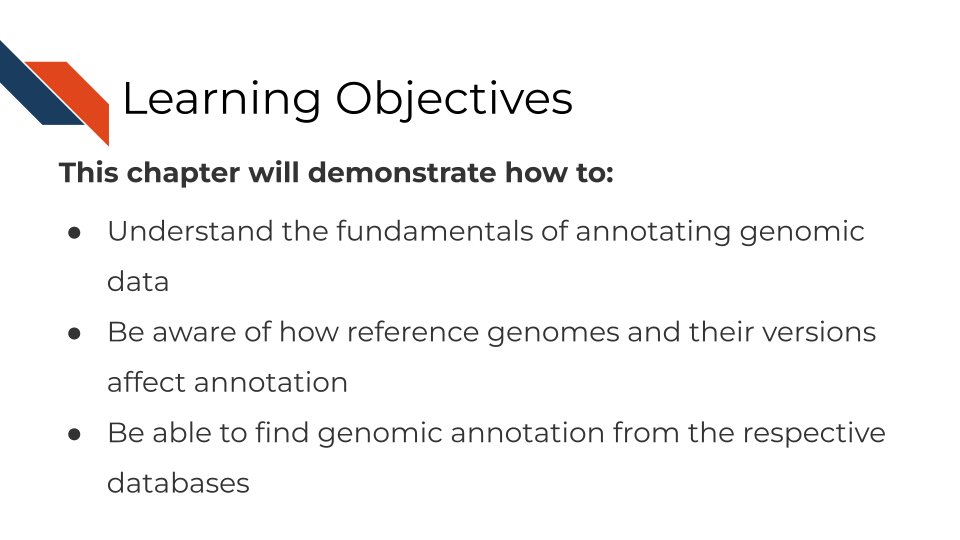
### 14.2.3 Python

## 14.3 Data Visualization Tools

## 14.4 More resources

# 15 Annotating Genomes

## 15.1 Learning Objectives



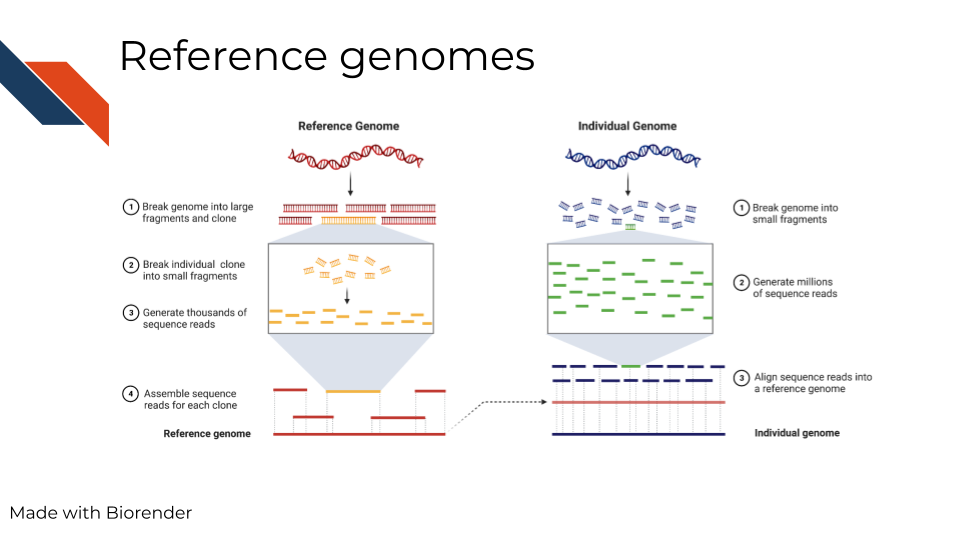
In this chapter, we are going to discuss methods that affect every genomic method and may take up the majority of your time as a genomic data analyst: Annotation.

We know that the sequencing or array data is not useful on its own – for our human minds to comprehend it and apply it to something we need a tangible piece of information to be attached to it. This is where annotation comes in. At best annotation helps you and others interpret genomic data. At its worst, its a time consuming activity that, done incorrectly, can lead to erroneous conclusions and labeling.

Proper annotation requires an understanding of how the annotation data you are using was derived as well as the realization that all annotation data is constantly changing and the confidence for these data are never 100%. Some organism’s genomes are better annotated than others but nearly all are at least somewhat incomplete.

## 15.2 What are reference genomes?

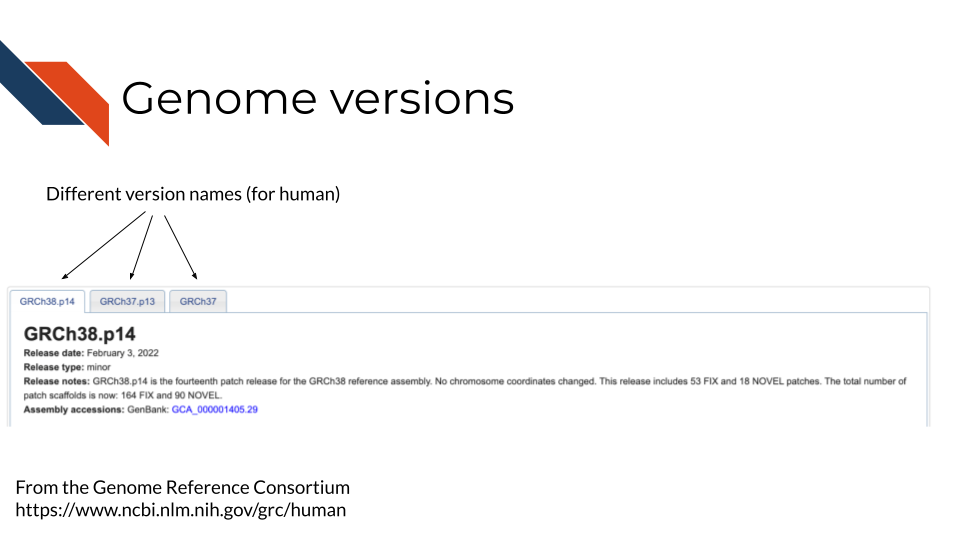
Every individual organism has its own DNA sequence that is unique to it. So how can we compare organisms to each other? In some studies, sequencing data is obtained and the genome is built de novo (aka from scratch) but this takes a lot of time and computing power. So instead, most genomic studies use the imperfect method of comparing to a reference genome. Reference genomes are built from prior data and available online. They inherently have biases in them. For example, human genomes are generally not made from diverse populations but instead from mostly males of european descent. It is inherently bad for both ethical and scientific reasons to to have [genome references that are too white](https://www.sciencenews.org/article/genetics-race-dna-databases-reference-genome-too-white). For more on the problems with reference genomes, [read this](https://genomebiology.biomedcentral.com/articles/10.1186/s13059-019-1774-4).



In summary, reference genomes are used for comparison and as a ‘source of truth’ of sorts, but its important to note that this method is biased and better alternatives need to be realized.

## 15.3 What are genome versions?

If you are familiar with software development, or have used any app before, you’re familiar with software updates and releases. Similarly, the genome has updates and releases as continued cloning and assemblies of organisms teaches us more. In the image below we are showing an example of what a genome version may be noted as (note that different databases may have different terminology – here we are showing the Genome Reference Consortium). You may also notice on their website it shows the date the genome version was released and what was fixed.

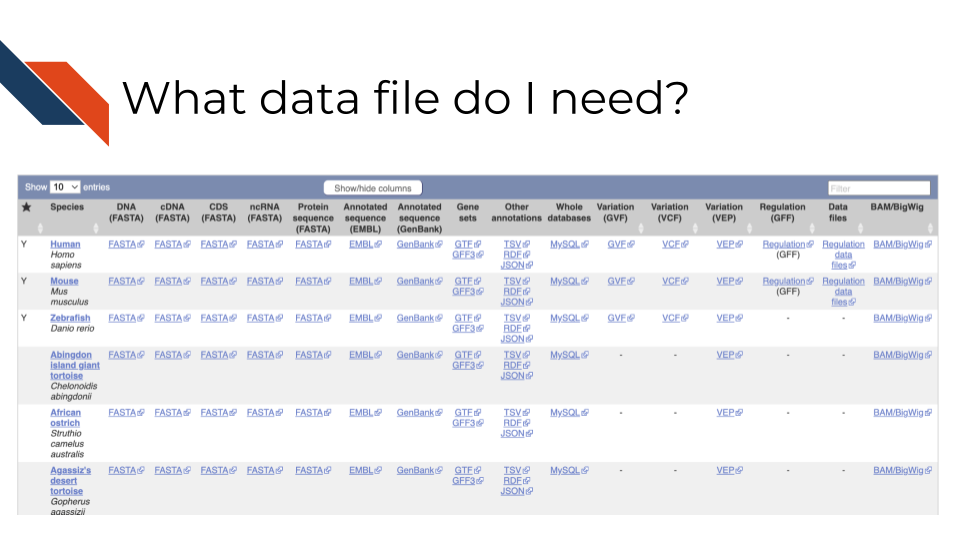


The details of how genome versions are fixed and released are not really of concern for your data analysis. This is merely to explain that genomes change and what is most important in your analysis is that:

1. You choose one genome version and consistently use it in all your analyses.
2. Choose a genome version that the rest of your field has generally had a consensus on and is also using. Generally this means sticking with major releases of a genome instead of always going with the latest version. Most databases will try to point you to their major release, so just stick with that. We will point you where you can find genome annotation for a lot of the major organisms.

## 15.4 What are the different files?

Although we can’t walk you through every organism and database set up, we will walkthrough the files and structure of one example here.



In the above screenshot, [from Ensembl](https://useast.ensembl.org/info/data/ftp/index.html), it shows different organisms in the rows, but also a variety of different files across the columns. In this example, DNA reference to the DNA sequence of the organism’s genome, but cDNA refers to complementary DNA – aka DNA that has been reversed transcribed from RNA. If you are working with RNA data you may want to use the cDNA file. Whereas CDS files are referring to only coding sequences and ncRNA files are showing only non coding sequences. Gene sets are also annotated and are in their own files. Most of these files are FASTA files. For a reminder on what these different file types are [see the previous chapter](http://hutchdatascience.org/Choosing_Genomics_Tools/a-very-general-genomics-overview.html#basic-file-formats).

Depending on the tool you are using, the data file and type you need will vary. Some tools have these data built in or are compatible with other packages that have annotation. If a tool automatically includes annotation within it, you will need to ensure that any additional tools you are using are also pulling from the same genome and version. Look into a tool’s documentation to find out what genome versions it is based on. If it doesn’t tell you at all, you don’t want to be using that tool. You cannot assume that cross genome analyses will translate.

### 15.4.1 How to download annotation files

For another database example we’ll look at the human data on ENA’s servers. Note that if you see FTP that just means “Fast Transfer Protocol” and it just means its where you can get the files themselves. For more on computing lingo, you can take our [Computing in Cancer Informatics course](https://www.itcrtraining.org/courses#h.civy2cnri95t).

[There’s many ways you can download these files and they are described here](https://ena-docs.readthedocs.io/en/latest/retrieval/file-download.html). In summary: - If you don’t feel comfortable using command line, [you can use the browser downloader for ENA here](https://www.ebi.ac.uk/ena/browser/home) - If you are using command line to write a script, then you can write use the [wget or curl instructions described here](https://ena-docs.readthedocs.io/en/latest/retrieval/file-download.html#using-wget). Be sure to read the README files to understand what it is you are downloading.

Also note that if you are working from a high power computing cluster or other online server, these annotation files may already be available to you. You don’t want to take up more computing resources by downloading extra files, so check with an administrator or informatics expert who also uses the cluster or cloud to check if the annotation files already exist in your workspace.

## 15.5 Considerations for annotating genomic data

### 15.5.1 Make sure you have the right file to start!

1. Is the annotation from the right organism? You may think this is a dumb question, but its very critical that you make sure you have the genome annotation for the organism that matches your data. Indeed the author of this has made this mistake in the past, so double check that you are using the correct organism.
2. Are all analyses utilizing coordinates from the same genome/transcriptome version? Genome versions are constantly being updated. Files from older genome versions cannot be used with newer ones (without some sort of liftover conversion). This also goes for transcriptome and genome data. All analysis need to be done using the same genomic versions so that is ensured that any chromosomal coordinates can translate between files. For example, it could be in one genome version a particular gene was said to be at chromosome base pairs 300 - 400, but in the next version its now been changed to 305 - 405. This can throw off an analysis if you are not careful. This type of annotation mapping becomes even more complicated when considering different splice variants or non-coding genes or regulatory regions that have even less confidence and annotation about them.

### 15.5.2 Be consistent in your annotations

1. If at all possible avoid making cross species analyses - unless you are an evolutionary genomics expert and understand what you are doing. But for most applications cross species analyses are hopeful wishing at best, so stick to one organism.
2. Avoid mixing genome/transcriptome versions. Yes there is liftover annotation data to help you identify what loci are parallel between releases, but its really much simpler to stick with the same version throughout your analyses’ annotations.

### 15.5.3 Be clear in your write ups!

Above all else, not matter what you end up doing, make sure that your steps, what files you use, and what tool versions you use are clear and reproducible! Be sure to clearly link to and state the database files you used and include your code and steps so others can track what you did and reproduce it. For more information on how to create reproducible analyses, you can take our reproducibility in cancer informatics courses: [Introduction to Reproducibility](https://www.itcrtraining.org/courses#h.n5yoq68qj0rz) and [Advanced Reproducibility in Cancer Informatics](https://www.itcrtraining.org/courses#h.i5zyiyjyttr4).

## 15.6 Resources you will need for annotation!

### 15.6.1 Annotation databases

* [Ensembl](https://useast.ensembl.org/downloads.html)
* [EMBL-EBI](https://www.ebi.ac.uk/services)
* [UCSCGenomeBrowser](https://genome.ucsc.edu/cgi-bin/hgGateway?hgsid=1571980135_Ym6A5aa3nDyOfZKtGishprdrhLDm)
* [NCBI Genomes download page](https://ftp.ncbi.nlm.nih.gov/genomes/)

### 15.6.2 GUI based annotation tools

* [UCSCGenomeBrowser](https://genome.ucsc.edu/cgi-bin/hgGateway?hgsid=1571980135_Ym6A5aa3nDyOfZKtGishprdrhLDm)
* [BROAD’s IGV](https://software.broadinstitute.org/software/igv/)
* [Ensembl’s biomart](https://useast.ensembl.org/info/data/biomart/index.html)

### 15.6.3 Command line based tools

#### 15.6.3.1 R-based packages:

* [annotatr](https://bioconductor.org/packages/release/bioc/html/annotatr.html)
* [ensembldb](https://bioconductor.org/packages/release/bioc/html/ensembldb.html)
* [GenomicRanges](https://www.bioconductor.org/packages/release/bioc/html/GenomicRanges.html) - useful for manipulating and identifying sequences.
* [GO.db](https://bioconductor.org/packages/3.16/data/annotation/html/GO.db.html) - Gene ontology annotation
* [org.Hs.eg.db](https://bioconductor.org/packages/release/data/annotation/html/org.Hs.eg.db.html)
* [RSamtools](https://www.bioconductor.org/packages/release/bioc/html/Rsamtools.html)
* [A full list of Bioconductors annotation packages](https://bioconductor.org/packages/3.16/data/annotation/) - contains annotation for all kinds of species and versions of genomes and transcriptomes.

#### 15.6.3.2 Python-based packages:

* [BioPython](https://biopython.org/)
* [genetrack](https://github.com/ialbert/chipexo)

### 15.6.4 More resources about genome annotation

# 16 Genomic Tool Glossary

Here’s all the tools that have been mentioned in this course or are otherwise recommended for your use. The list is in alphabetical order.

## 16.1 Bioconductor

The mission of the Bioconductor project is to develop, support, and disseminate free open source software that facilitates rigorous and reproducible analysis of data from current and emerging biological assays. We are dedicated to building a diverse, collaborative, and welcoming community of developers and data scientists.

Bioconductor uses the R statistical programming language, and is open source and open development. It has two releases each year, and an active user community. Bioconductor is also available as Docker images.

### 16.1.1 Notable Bioconductor genomics tools:

* [annotatr](https://bioconductor.org/packages/release/bioc/html/annotatr.html)
* [ensembldb](https://bioconductor.org/packages/release/bioc/html/ensembldb.html)
* [GenomicRanges](https://www.bioconductor.org/packages/release/bioc/html/GenomicRanges.html) - useful for manipulating and identifying sequences.
* [GO.db](https://bioconductor.org/packages/3.16/data/annotation/html/GO.db.html) - Gene ontology annotation
* [org.Hs.eg.db](https://bioconductor.org/packages/release/data/annotation/html/org.Hs.eg.db.html)
* [RSamtools](https://www.bioconductor.org/packages/release/bioc/html/Rsamtools.html)
* [A full list of Bioconductors annotation packages](https://bioconductor.org/packages/3.16/data/annotation/) - contains annotation for all kinds of species and versions of genomes and transcriptomes.
* [ComplexHeatmap](https://bioconductor.org/packages/release/bioc/html/ComplexHeatmap.html#:~:text=Complex%20heatmaps%20are%20efficient%20to,and%20supports%20various%20annotation%20graphics.)
* [MultiAssayExperiment](https://bioconductor.org/packages/MultiAssayExperiment/)
* [limma](https://www.bioconductor.org/packages/release/bioc/html/limma.html)
* [DESEq2](https://www.bioconductor.org/packages/release/bioc/html/DESeq2.html)
* [edgeR](https://www.bioconductor.org/packages/release/bioc/html/edgeR.html)
* [curatedTCGAData](https://bioconductor.org/packages/curatedTCGAData/)
* [cBioPortalData](https://bioconductor.org/packages/cBioPortalData/)
* [SingleCellMultiModal](https://bioconductor.org/packages/SingleCellMultiModal/)

## 16.2 Cancer Models

Patient Derived Cancer Models Finder (www.cancermodels.org) is a cancer research platform that aggregates clinical, genomic and functional data from patient-derived xenografts, organoids and cell lines. The PDCM Finder standardises, harmonises and integrates the complex and diverse data associated with PDCMs for cancer community.

Data types used are model meta data, related clinical metadata from the sample for which the model was derived, e.g. molecular and treatment-based. Data are preprocessed, consistently semantically annotated, harmonised and FAIR.

PDCM Finder contains >6200 models across 13 cancer types, including rare pediatric models (17%) and models from minority ethnic backgrounds (33%), making it the largest free to consumer and open access resource of this kind. Get started at www.cancermodels.org to browse and query models by cancer type

## 16.3 CIViC

[CIViC](www.civicdb.org) is a knowledgebase and curation interface for the clinical interpretation of variants in cancer. Evidence is curated from published literature describing the diagnostic, prognostic, predictive, predisposing, oncogenic, or functional role of variants in specific cancer types. Evidence submitted by community curators is revised and moderated by expert editors. Individual evidence is synthesized into gene summaries, variant summaries and variant-disease assertions of specific clinical relevance. Anyone can make use of CIViC knowledge through the open web interface or API. Information on how to use or contribute to CIViC is available in our help docs (docs.civicdb.org). The main distinguishing feature of CIViC compared to similar resources it is total commitment to open data sharing. All data are available in the Public Domain (CC0). The code is available for any use under an MIT license.

## 16.4 CTAT

The Trinity Cancer Transcriptome Analysis Toolkit (CTAT, <https://github.com/NCIP/Trinity_CTAT/wiki> ) provides a diverse collection of tools to gain insights into the biology of cancer through the lens of the transcriptome. Using RNA-seq as input, CTAT modules enable detection of mutations, fusion transcripts, copy number aberrations, cancer-specific splicing aberrations, and oncogenic viruses including insertions into the human genome. CTAT uses both read mapping and de novo assembly methods to analyze RNA-seq, leveraging tumor bulk and single cell transcriptomes. CTAT modules provide interactive visualizations as outputs, are easily installed for local execution or run via cloud computing (eg. Terra), have detailed user guides and tutorials, and are well-supported through user forums.

## 16.5 DeepPhe

[DeepPhe: Natural Language Processing Tools for Cancer Research](https://deepphe.github.io/)

Under development since 2014, the DeepPhe suite of software tools aims to extract deep phenotype information from the Electronic Medical Records from patients with cancer. DeepPhe combines:

1. multiple natural language processing (NLP) techniques based on cTAKES,1
2. a structured cancer information model including concepts from the NCIT and the HemOnc ontology
3. a graph data model supporting persistence of extracted details including links between patient data enabling semantically informed interpretation, aggregation, and disaggregation of key attributes,
4. visual analytics tools supporting patient- and cohort-level displays of extracted data5 including identification of patients matching key research criteria and the examination of individual patient records such as exploration of links between summary items and supporting text mentions, and
5. multiple strategies for use, including containerized REST services and GUIs for installation and pipeline execution.

DeepPhe tools are available for download and installation from the [DeepPhe website](https://deepphe.github.io/) under an open-source license for non-commercial use.

## 16.6 Genetic Cancer Risk Detector (GARDE)

[Genetic Cancer Risk Detector (GARDE)](https://reimagineehr.utah.edu/innovations/garde/) screens and identifies patients who meet National Comprehensive Cancer Network (NCCN) criteria for genetic evaluation of familial cancer risk based on their family history in the EHR using both structured data and natural language processing of free-text data. Patients identified by GARDE are imported into an EHR’s population health management dashboard (e.g., Epic’s Healthy Planet module) where genetic counseling staff review individual cases, select, and send bulk outreach messages to patients via chatbot and/or through the patient portal.

GARDE is a population clinical decision support (CDS) platform based on Fast Healthcare Interoperability Resources (FHIR) and CDS Hooks standards to support interoperability and logic sharing beyond single vendor solutions.

## 16.7 Gene Set Enrichment Analysis (GSEA)

[Gene Set Enrichment Analysis (GSEA)](https://www.gsea-msigdb.org/gsea/index.jsp) is a method to identify the coordinate activation or repression of groups of genes that share common biological functions, pathways, chromosomal locations, or regulation, thereby distinguishing even subtle differences between phenotypes or cellular states. Gene set-based enrichment analysis is now standard practice for interpreting global transcription profiling experiments and elucidating the biological mechanisms associated with disease and other biological phenotypes of interest. The method is more powerful than typical single-gene approaches to comparing phenotypes, as it can identify sets of genes (e.g., perturbation signatures or molecular pathways) that are coordinately up- or downregulated when each gene in the set may not be significantly differentially expressed. The GSEA software provides useful visualizations and reports for the exploration and interpretation of results. GSEA bundles direct access to the Molecular Signatures Database (MSigDB) – a comprehensive curated repository of annotated gene sets representing signatures derived from publications, pathway databases, and other sources of public data; MSigDB can also be used independently.

The website for the GSEA-MSigDB resource can be found at gsea-msigdb.org. To get started with GSEA you can view the GSEA User Guide, and access the GSEA software through the downloads page or through the GSEA modules available on GenePattern. See the MSigDB section of the website for more information about MSigDB and to interactively explore the gene sets and their annotations. User support for GSEA and MSigDB is available through our help forum.

## 16.8 Integrative Genomics Viewer (IGV)

The [Integrative Genomics Viewer (IGV)](https://software.broadinstitute.org/software/igv/) is a track-based browser for interactively exploring genomic data mapped to a reference genome. IGV supports all the standard genomic data types (aligned reads, variants, signal peaks, genome annotations, copy number variation, etc.) as well as sample information, such as clinical, phenotypic, or other attributes. IGV provides great flexibility in loading data, whether investigator generated or publicly available, directly from multiple disparate sources without the need for any pre-processing. Supported data sources include local file systems; web servers on the user’s intranet or the Internet; commercial cloud providers (Google, Amazon, Azure, Dropbox); web links to data in public repositories. Authentication to access private data on the web is supported with the industry standard OAuth protocol.

IGV is available in multiple forms, including both end-user applications and versions for use by developers. The IGV website at <https://igv.org> provides access to all modalities of IGV. Download and install the IGV Desktop application from the downloads page. To learn about using the application see the tutorial videos on the IGV YouTube channel and the online User Guide. The IGV-Web app is available at <https://igv.org/app>. To learn about using the app, the Help link in the menu bar provides access to the documentation, and see also the tutorial videos on the YouTube channel. The igv.js JavaScript component is for web developers who wish to embed IGV in their web apps or portals. More information can be found in the Readme file and the Wiki in the igv.js GitHub repository. IGV user support is available through the igv-help online forum and the GitHub repositories.

## 16.9 NDEx

The [Network Data Exchange (NDEx)](https://www.ndexbio.org/#/) project provides an open-source framework where scientists and organizations can store, share and publish biological network knowledge. A distinctive feature of NDEx is that it serves as a home for models that are currently available only as figures, tables, or supplementary information, such as networks produced via systematic mining and integration of large-scale molecular data.

NDEx includes features to support data distribution and access according to FAIR principles. Its full integration with Cytoscape, the popular desktop application for network analysis and visualization, provides the cloud back-end component for data I/O; so, if a network file format can be opened in Cytoscape, it can also be stored in (and retrieved from) NDEx.

NDEx can be accessed via its web user interface or programmatically, via REST API and client libraries in Python, R, Java. Web applications can interface with NDEx via JavaScript: MSigDB, CRAVAT, cBioPortal and IQuery, are all examples of web applications integrated with NDEx.

For more information, please review the About NDEx page. To get started, visit the NDEx public server: there, you can review the NDEx FAQ, access documentation, contact us, and search or browse thousands of biological network models.

## 16.10 MultiAssayExperiment

[MultiAssayExperiment](https://bioconductor.org/packages/MultiAssayExperiment/) is an R/Bioconductor package that harmonizes data management, manipulation, and subsetting of multiple experimental assays performed on an overlapping set of specimens. It supports on-disk and remote data storage, and provides reshaping tools for adaptability to arbitrary downstream analysis.

MultiAssayExperiment is distinct from alternative approaches in its focus on multi’omic data management and manipulation and in its integration with the Bioconductor ecosystem: it is used by more than 50 other Bioconductor packages, it provides a familiar Bioconductor user experience by extending concepts from SummarizedExperiment while supporting an open-ended mix of data classes for individual assays, and it allows subsetting by genomic ranges, row names, phenotypic data, and assays.

You can get started with the MultiAssayExperiment Bioconductor package documentation, or start with prebuilt MultiAssayExperiments objects from [curatedTCGAData](https://bioconductor.org/packages/curatedTCGAData/), [cBioPortalData](https://bioconductor.org/packages/cBioPortalData/), or [SingleCellMultiModal](https://bioconductor.org/packages/SingleCellMultiModal/).

## 16.11 OpenCRAVAT

[OpenCRAVAT](https://run.opencravat.org) uses variation data in many popular variant file formats and its outputs are variant annotations and visualizations. To get started go to opencravat.org. Download and run on your local machine, multi-user servers, at <https://run.opencravat.org> or in the cloud. We offer a broader selection of annotation tools than comparable software and results can be explored with an interactive GUI that provides customized filtering options, interactive tables and widgets. Use it for a single sample or a large cohort, or pull single variant reports with a structured url (Example: <https://run.opencravat.org/webapps/variantreport/index.html?chrom=chr11&pos=48123823&ref_base=A&alt_base=C> )

## 16.12 WebMeV

[WebMeV](https://webmev.tm4.org) is an online tool that facilitates analysis of large-scale RNA-seq and other multi-omic datasets by providing intuitive access to advanced analytical methods and high-performance computing for a wide range of basic, clinical, and translational researchers. Although WebMeV provides support for “bulk” RNA-seq data, single-cell RNA-seq, and other types of -omic data and provides easy access to public data resources such as The Cancer Genome Atlas (TCGA) and the Genotype-Tissue Expression project (GTEx)—as well as user-provided data. WebMeV uniquely provides a user-friendly, intuitive, interactive interface to processed analytical data uses cloud-computing elasticity for computationally intensive analyses that are increasingly required for genomic data analysis. WebMeV’s design places an emphasis on user-driven data analysis by providing users the ability to visualize, interact with, and dissect genomic data at each step in the analysis with a “point-and-click” interactive data environment. Although the primary input is normalized “count matrices,” WebMeV does include tools for data normalization and quality control and uses Dropbox and Google Drive as means of easily uploading data. Analytical methods include statistical tests for comparing cohorts, for identifying gene seats, for doing functional enrichment analysis on gene sets (GSEA), and for inferring gene regulatory network models and comparing these networks between phenotypes to understand the drivers of disease. WebMeV also provides a platform to support reproducible research and makes code for the entire system and its component methods available as open-source software code.

## 16.13 Xena

[UCSC Xena](http://xena.ucsc.edu/) is a web-based visualization tool for multi-omic data and associated clinical and phenotypic annotations.

Xena showcases seminal cancer genomics datasets from TCGA, the Pan-Cancer Atlas, GDC, PCAWG, ICGC, and more; a total of more than 1500 datasets across 50 cancer types. We support virtually any type of functional genomics data (sometimes known as level 3 or 4 data). This includes SNPs, INDELs, copy number variation, gene expression, ATAC-seq, DNA methylation, exon-, transcript-, miRNA-, lncRNA-expression and structural variants. We also support clinical data such as phenotype information, subtype classifications and biomarkers. All of our data is available for download via python or R APIs, or through our URL links.

### 16.13.1 Questions Xena can help you answer include:

* Is overexpression of this gene associated with better survival?
* What genes are differentially expressed between these two groups of samples?
* What is the relationship between mutation, copy number, expression, etc for this gene?

Our tool differentiates itself by its ability to visualize more uncommon data types, such as DNA methylation, its visual integration of multiple types of genomic data side-by-side, and its ability to easily privately visualize your own data.

Get started with our tutorials: <https://ucsc-xena.gitbook.io/project/tutorials>. If you use us please cite us: <https://www.nature.com/articles/s41587-020-0546-8>

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## ─ Session info ───────────────────────────────────────────────────────────────  
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## os Ubuntu 20.04.3 LTS   
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## ui X11   
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## tz Etc/UTC   
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##   
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## rlang 0.4.10 2022-02-15 [1] Github (r-lib/rlang@f0c9be5)   
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## stringi 1.5.3 2020-09-09 [1] RSPM (R 4.0.3)   
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## usethis 2.1.5.9000 2022-02-15 [1] Github (r-lib/usethis@57b109a)   
## withr 2.3.0 2020-09-22 [1] RSPM (R 4.0.2)   
## xfun 0.26 2022-02-15 [1] Github (yihui/xfun@74c2a66)   
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