20190701 orthogroup stat analysis

July 8, 2020

```
[1]: import numpy, pandas, datetime, sys, os
     import matplotlib, seaborn
     import re, math, time, scipy
     import sqlite3, IPython.display, sqlalchemy, statsmodels, ete3, svgutils.
      \hookrightarrowtransform
     import lxml.etree, gseapy, mygene
     import itertools
     import igraph
     import upsetplot
     import random
     import subprocess
     import importlib
     from svgpathtools import svg2paths
     from sqlalchemy.types import *
     import statsmodels
     from statsmodels.sandbox.regression.predstd import wls prediction std
     from networkx.drawing.nx_agraph import write_dot
     from decimal import Decimal
     sys.path.append('/Users/kef74yk/Dropbox_w/repos/kftools')
     sys.path.append('/Users/kef74yk/Dropbox_w/repos/kftools/kftools')
     import kftools
     kftools = importlib.reload(kftools)
     from kftools import kfplot
     from kftools import kfutil
     from kftools import kfstat
     font size = 8
     %matplotlib inline
     pandas.options.display.max_rows=10
     pandas.options.display.max_columns=1000
     seaborn.set_style("white")
     seaborn.set_style("ticks")
     matplotlib.rcParams['font.size'] = font_size
     #matplotlib.rcParams['font.family'] = 'Helvetica'
```

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#matplotlib.rc('font',**{'family':'sans-serif','sans-serif':['Helvetica']})
matplotlib.rcParams['svg.fonttype'] = 'none' # none, path, or sugfont

matplotlib.rc('xtick', labelsize=font_size)
matplotlib.rc('ytick', labelsize=font_size)
matplotlib.rc('font', size=font_size)
matplotlib.rc('axes', titlesize=font_size)
matplotlib.rc('axes', labelsize=font_size)
matplotlib.rc('ytick', labelsize=font_size)
matplotlib.rc('ytick', labelsize=font_size)
matplotlib.rc('legend', fontsize=font_size)
matplotlib.rc('figure', titlesize=font_size)
```

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[2]: omega_method = 'mapdnds' # 'hyphy' or 'mapdnds'
     #l1ou criterion = 'AICc'
     #expression unit = 'TPM' # 'FPKM', 'TPM'
     #intersect fpkm tpm = False
     sptree_root = 'sp_root'
     dir_data = '/Users/kef74yk/Dropbox_p/data/'
     wd = os.path.join(dir_data, "04_transcriptome_amalgamation/
     →20190701_orthogroup_stat_analysis/out/")
     dir_ensembl = os.path.join(dir_data, '/Users/kef74yk/Dropbox_p/db/Ensembl/
     →release-91/')
     #db file = os.path.join(dir ensembl, 'orthogroup/Ensembl.91.orthogroup.
     → '+l1ou criterion+'.db')
     db_file = os.path.join(dir_ensembl, 'orthogroup/Ensembl.91.orthogroup.db')
     sptree_file = os.path.join(dir_ensembl, 'timetree/species_timetree.nwk')
     phylopic_dir = os.path.join(dir_ensembl, 'phylopic/svg_files/')
     inkscape='/Applications/Inkscape.app/Contents/Resources/bin/inkscape'
     if not (os.path.exists(wd)):
         os.mkdir(wd)
     os.chdir(wd)
     font size = 8
     max_delta_intron_present=-0.5
     pcm_prefixes = ['llou_fpkm_','llou_tpm_',]
     shift_columns =
     →['l1ou_fpkm_is_shift','l1ou_tpm_is_shift','l1ou_intersect_is_shift']
     branch_categories = ['S','D','R']
     category colors = {'S':'#0033cc', 'D':'#cc0000', 'R':'#e59400'}
     organs = ['brain','heart','kidney','liver','ovary','testis']
     organ_colors = ['#1B9E77','#D95F02','#7570B3','#E7298A','#66A61E','#E6AB02']
     organ_colors_dict = dict()
     for o,c in zip(organs,organ_colors):
```

```
organ_colors_dict[o] = c
```

```
[3]: def attach neighbor stats(df, neighbor, columns=[]):
         columns = ['orthogroup', 'numerical_label',] + columns
         df_tmp = df.loc[:,columns]
         df_tmp.columns = neighbor+'_'+pandas.Series(df_tmp.columns).astype(str)
         right_on = [neighbor+'_orthogroup',neighbor+'_numerical_label']
         df = pandas.merge(df, df_tmp, left_on=['orthogroup',neighbor],__
      →right_on=right_on, how='left')
         df = df.drop(right_on, axis=1)
         return df
     def my_distplot(x, df, ax, kde, xlab, norm_hist, x_range):
         seaborn.distplot(df[x], ax=ax, kde=kde, norm_hist=norm_hist, color='black')
         ax.tick_params(axis='both', which='major', direction='in', length=6,_
      \rightarrowwidth=1)
         ax.set_xlabel(xlab)
         if norm_hist:
             ax.set_ylabel('Frequency')
         else:
             ax.set_ylabel('Count')
         if (len(x_range)>1)&(type(x_range)!=str):
             ax.set_xlim(x_range[0], x_range[1])
     def my barplot(x, y, ax, xlab, ylab):
         seaborn.barplot(x=x, y=y, ax=ax, color='gray')
         ax.set xlabel(xlab)
         ax.set_ylabel(ylab)
         ax.tick_params(axis='both', which='major', direction='in', length=6,__
      \rightarrowwidth=1)
     def add_missing_data(df, index=[], columns=[]):
             del df.index.name
         except:
             0
         try:
             del df.columns.name
         except:
         for ind in index:
             if not ind in df.index:
                 df = pandas.concat([df, pandas.DataFrame(0, index=[ind,],__
      ⇒columns=df.columns)], axis=0)
         for col in columns:
             if not col in df.columns:
```

```
df = pandas.concat([df, pandas.DataFrame(0, index=df.index,__
 df = df.sort_index(axis=0)
   df = df.sort index(axis=1)
   return df
def add_igraph_legends(svg_file, height, width, texts=[]):
   font size = 8
   font_family = 'Helvetica'
   fig = svgutils.transform.SVGFigure(str(width)+"pt", str(height)+"pt")
   figs = list()
   figs.append(svgutils.transform.fromfile(svg_file))
   plots = list()
   for i in range(len(figs)):
       plots.append(figs[i].getroot())
   plots[0].moveto(x=0, y=0, scale=1)
   txts = list()
   for i in range(len(texts)):
       txts.append(svgutils.transform.TextElement(x=0, y=font_size*(i+2),_
 -text=texts[i], size=font_size, weight="normal", font=font_family))
   fig.append(plots)
   fig.append(txts)
   fig.save(svg_file)
def calc_symmetry(pivot_table):
    if not type(pivot_table) == type(pandas.DataFrame()):
       pivoto_table = pivot_table * (numpy.eye(pivot_table.shape[0],__
→pivot_table.shape[1])==0)
       pivot_table = pandas.DataFrame(pivot_table)
   pivot_table = pivot_table.fillna(0)
   organs = list(set(pivot_table.index.tolist()).intersection(set(pivot_table.
dif = 0
   for i in itertools.combinations(organs, 2):
        if i[0]!=i[1]:
           dif = dif + numpy.abs(pivot_table.loc[i[0],i[1]] - pivot_table.
\rightarrowloc[i[1],i[0]])
    symmetry = 1 - (dif/pivot_table.sum().sum())
   return(symmetry)
def integrate_pcm_stats(df, pcm_prefix, drop=False):
   num og before = 0
   df['pcm_method'] = ''
   for pp in pcm_prefix:
       pp_columns = df.columns[df.columns.str.startswith(pp)]
       for ppc in pp_columns:
           new_column = ppc.replace(pp, '')
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if not (new_column in df.columns):
                df[new_column] = df[ppc]
            else:
                df.loc[df[new_column].isnull(), new_column] = df.
→loc[df[new_column].isnull(), ppc]
        if drop:
            df = df.drop(pp_columns, axis=1)
        col = 'regime' if 'regime' in df.columns else 'num_regime'
        df.loc[(~df[col].isnull())&(df['pcm_method']==''), 'pcm_method'] = re.
 →sub('_$','',pp)
       num_og_after = df.loc[(~df[col].isnull()), 'orthogroup'].
 →drop_duplicates().shape[0]
        print(pp, ': number of added orthogroups =', num_og_after -_
→num_og_before)
       num_og_before = num_og_after
   return df
def add_max_mu(df=pandas.DataFrame(), cols=[], new_col=''):
   df[new_col] = df.loc[:,cols].idxmax(axis=1)
   num null before = df[new col].isnull().sum()
   tmp = df.loc[:,cols].values
   row max = tmp.max(axis=1)
   for i in numpy.arange(tmp.shape[1]):
        tmp[:,i] = tmp[:,i] - row_max
   is_multiple_max = ((tmp==0).sum(axis=1)!=1)
   df.loc[is_multiple_max,new_col] = numpy.nan
   num_null_after = is_multiple_max.sum()
   print(new_col, ':', num_null_after-num_null_before, 'tie max values were_

detected¹)
   return df
def shift_freq_bootstrap(df1, var1='parent_max_organ', var2='max_organ', u
→down reg=True, up reg=True,
                         exclude_self=True, nboot=1000, nsubsample=numpy.inf):
   if exclude_self:
        df1 = df1.loc[(df1[var1]!=df1[var2]),:]
    if df1.shape[0]>nsubsample:
       df1 = df1.loc[numpy.random.choice(df1.index, nsubsample,__
 →replace=False),:]
   df1 = df1.sort_values(axis=0, by=[var1, var2])
   df1 = df1.reset_index()
   N = df1.shape[0]
   print('N after filtering =', N)
   all_organs = pandas.concat([df1[var1],df1[var2]]).dropna().unique()
   all_organs.sort()
    observed = pandas.DataFrame(df1.groupby([var1,var2])['orthogroup'].count())
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```
observed = observed.reset_index().pivot(var1, var2)
   colnames = observed.columns.get_level_values(1)
   observed = observed.T.reset_index(drop=True).T
   observed.columns = colnames
   observed = observed.fillna(0)
   observed = add_missing_data(df=observed, index=all_organs,__

→columns=all_organs)

   if exclude_self:
       observed_total = observed.sum().sum()
       index_values = observed.index
       column_values = observed.columns
       self_true = numpy.array([ i==c for i in index_values for c in_
→column_values]).reshape([index_values.shape[0],column_values.shape[0]])
   nrep = nboot
   num_var1 = df1[var1].unique().shape[0]
   num_var2 = df1[var2].unique().shape[0]
   axis = [nrep, observed.shape[0], observed.shape[1]]
   bs = numpy.zeros(axis, dtype=numpy.int64)
   replace=False
   for i in numpy.arange(nrep):
       df3 = df1.loc[:,:]
       if replace:
           df3.loc[:,var1] = df3.loc[numpy.random.choice(df3.index, size=df3.
→shape[0], replace=replace), var1].values
       df3.loc[:,var2] = df3.loc[numpy.random.choice(df3.index, size=df3.
⇒shape[0], replace=replace), var2].values
       pivot = pandas.DataFrame(df3.groupby([var1,var2])['orthogroup'].count())
       pivot = pivot.reset_index().pivot(var1, var2)
       pivot.columns = pivot.columns.get_level_values(1)
       if replace:
           if not observed.index.shape[0] == pivot.index.shape[0]:
               organs = list(set(observed.columns.tolist() + observed.index.
→tolist()))
               missings = [ pcol for pcol in organs if not pcol in pivot.index_
\hookrightarrow
               for missing in missings:
                   pivot.loc[missing,:] = 0
           if not observed.columns.shape[0] == pivot.columns.shape[0]:
               organs = list(set(observed.columns.tolist() + observed.index.
→tolist()))
               missings = [ pcol for pcol in organs if not pcol in pivot.
→columns.get_level_values(1) ]
               for missing in missings:
                   pivot['orthogroup',missing] = 0
       pivot = add_missing_data(df=pivot, index=all_organs, columns=all_organs)
       pivot = pivot.fillna(0).values
```

```
if exclude_self:
           pivot[self_true] = 0
           pivot = pivot / pivot.sum().sum() * observed_total
       bs[i,:,:] = pivot
       del df3
   del df1
   col sum = observed.sum(axis=0)
   row_sum = observed.sum(axis=1)
    col sum = numpy.expand dims(col sum, axis=0)
   row_sum = numpy.expand_dims(row_sum, axis=1)
    col_freq = col_sum / col_sum.sum().sum()
   row_freq = row_sum / row_sum.sum().sum()
   expected = numpy.array(col_freq * row_freq)
   if exclude_self:
        expected[self_true] = 0
    expected = expected / expected.sum().sum() * observed.sum().sum()
    expected = pandas.DataFrame(expected)
    expected.index = observed.index
    expected.columns = observed.columns
    corrected = observed/expected.values
    corrected = add_missing_data(df=corrected, index=all_organs,_
corrected = corrected.fillna(0)
   return(corrected,observed,expected,bs,N)
def draw_network(corrected, observed, expected, bs, self_arrow=False,_
⇒self_vertex_size=False, show_vertex_stat=False,
                edge_width=1, hide_nonsig=False, coordinates=None,_
 →test='random'):
    bs_mean = pandas.DataFrame(bs.mean(axis=0))
   bs_conf95lower = pandas.DataFrame(numpy.percentile(bs, axis=0, q=2.5))
   bs_conf95upper = pandas.DataFrame(numpy.percentile(bs, axis=0, q=97.5))
   bs_conf99lower = pandas.DataFrame(numpy.percentile(bs, axis=0, q=0.5))
   bs_conf99upper = pandas.DataFrame(numpy.percentile(bs, axis=0, q=99.5))
   bs_IQRlower = pandas.DataFrame(numpy.percentile(bs, axis=0, q=25))
   bs_IQRupper = pandas.DataFrame(numpy.percentile(bs, axis=0, q=75))
   for bs in ...
→ [bs_mean,bs_conf95lower,bs_conf95upper,bs_conf99lower,bs_conf99upper,bs_IQRlower,bs_IQRuppe
       bs.index = observed.index
       bs.columns = observed.columns
   g = igraph.Graph(directed=True)
   vertices = pandas.Series(list(set(corrected.index.tolist()+corrected.
vertex_weights = list()
   vertex_labels = list()
```

```
vertex colors = list()
  tissue_colors={'brain':'#1B9E77','heart':'#D95F02','kidney':
→ '#7570B3', 'liver': '#E7298A', 'ovary': '#66A61E', 'testis': '#E6AB02'}
  my coordinates = list()
  for organ in vertices:
       if (organ in corrected.index)&(organ in corrected.columns):
           vertex_weights.append(corrected.loc[organ,organ])
           n_parent = str(int(observed.loc[organ,:].sum().sum()))
           n_child = str(int(observed.loc[:,organ].sum().sum()))
           organ = organ.replace('-', '\n')
           if show_vertex_stat:
               vertex_labels.append(organ+'\n'+n_parent+':'+n_child)
           else:
               vertex_labels.append(organ)
       else:
           vertex_labels.append(organ)
           vertex_weights.append(1)
           if show_vertex_stat:
               organ = organ.replace('-', '\n')
               vertex_labels.append(organ)
       if organ in tissue colors.keys():
           vertex_colors.append(tissue_colors[organ])
       else:
           vertex_colors.append('darkgray')
       if coordinates is not None:
           my_coordinates.append(coordinates[organ])
   g.add_vertices(vertices)
   edge_weights = list()
   edge_colors = list()
   edge_labels = list()
   edge_curves = list()
   if test=='rank':
       df_rank = pandas.DataFrame(observed.values.flatten())
       df rank = df rank.rank()
       df_rank = numpy.reshape(df_rank.values, newshape=observed.shape)
       df_rank = df_rank - min(observed.shape) # setting baseline by excluding_
\rightarrow diagonal elements
       numpy.fill_diagonal(a=df_rank, val=0)
       df_rank = pandas.DataFrame(df_rank).astype(int)
       df_rank.index = observed.index
       df_rank.columns = observed.columns
       rank_colors = kfutil.get_rgb_gradient(ncol=int(df_rank.max().max()),u
\hookrightarrow col1=[0,0,1], col2=[1,0,0], colm=[0.5,0.5,0.5])
  for organ1 in corrected.index:
       for organ2 in corrected.columns:
           flag = True if (self_arrow) | (self_vertex_size) else organ1!=organ2
           if flag:
```

```
organ1id = vertices.loc[vertices==organ1].index.values[0]
              organ2id = vertices.loc[vertices==organ2].index.values[0]
              corrected_value = corrected.loc[organ1,organ2]
              expected_value = expected.loc[organ1,organ2]
              observed_value = observed.loc[organ1,organ2]
              if test=='random':
                  if (observed_value==int(observed_value)):
                      edge_label = str(int(observed_value))+'/'+str(int(numpy.
→round(expected_value, decimals=0)))
                  else:
                      edge_label = str(numpy.round(observed_value,_
→decimals=2))+'/'+str(numpy.round(expected_value, decimals=2))
                  color = igraph.color_name_to_rgba('#808080')
                  if observed.loc[organ1,organ2] < bs_IQRlower.</pre>
→loc[organ1,organ2]:
                      color = igraph.color_name_to_rgba('#808080')
                  if observed.loc[organ1,organ2] < bs_conf95lower.</pre>
→loc[organ1,organ2]:
                      color = igraph.color_name_to_rgba('#40C040')
                  if observed.loc[organ1,organ2] < bs conf99lower.</pre>
→loc[organ1,organ2]:
                      color = igraph.color_name_to_rgba('#00FF00')
                  if observed.loc[organ1,organ2] > bs_IQRupper.
→loc[organ1,organ2]:
                      color = igraph.color_name_to_rgba('#808080')
                  if observed.loc[organ1,organ2] > bs_conf95upper.
→loc[organ1,organ2]:
                      color = igraph.color_name_to_rgba('#C040C0')
                  if observed.loc[organ1,organ2] > bs_conf99upper.
→loc[organ1,organ2]:
                      color = igraph.color_name_to_rgba('#FF00FF')
              elif test=='chisq':
                  parentY_childY = observed.loc[(observed.index==organ1),__
parentY_childN = observed.loc[(observed.index==organ1),__

→ (observed.columns!=organ2)].sum().sum()
                  parentN_childY = observed.loc[(observed.index!=organ1),__
parentN_childN = observed.loc[(observed.index!=organ1),__
cont_table = pandas.DataFrame([[parentY_childY,__
→parentY_childN],[parentN_childY, parentN_childN]])
                  chi2_out = scipy.stats.
→chi2_contingency(observed=cont_table, correction=True)
                  chi2 = chi2 out[0]
                  pvalue = chi2_out[1]
```

```
print(organ1, organ2, chi2, pvalue)
                   print('test = chisq. This option is not yet implemented.')
               elif test=='rank':
                   rank = int(df_rank.loc[organ1,organ2])
                   rgb = rank_colors[rank-1]
                   hex_color = kfutil.rgb_to_hex(rgb[0],rgb[1],rgb[2])
                   color = igraph.color_name_to_rgba(hex_color)
                   edge label = ''
               g.add edges([(organ1id,organ2id),])
               if edge_width=='observed':
                   edge weight = 0.5
                   if observed_value >= 10:
                       edge weight = 1
                   if observed_value >= 100:
                       edge_weight = 2
               else:
                   edge_weight = edge_width
               edge_weights.append(edge_weight)
               edge_colors.append(color)
               edge_labels.append(edge_label)
               edge_curve = -0.05
               if (organ1 not in corrected.columns) | (organ2 not in corrected.
⇒index):
                   edge_curve = 0
               edge_curves.append(edge_curve)
   if hide_nonsig:
       for i in range(len(edge_labels)):
           if edge_colors[i] == igraph.color_name_to_rgba('#808080'):
               edge_labels[i] = ''
   visual style = {}
   visual_style["vertex_size"] = [ v*20 for v in vertex_weights ] ifu
⇒self vertex size else 26#27
   visual_style["vertex_label"] = vertex_labels
   visual style["vertex color"] = vertex colors
   visual_style["vertex_label_color"] = 'black'
   visual_style["vertex_label_size"] = 8 if show_vertex_stat else 8
   visual_style["edge_width"] = edge_weights
   visual_style["edge_arrow_size"] = 0.5
   visual_style["edge_label"] = edge_labels
   visual_style["edge_label_size"] = 6
   visual_style["edge_label_color"] = edge_colors
   visual_style["edge_color"] = edge_colors
   visual_style["edge_curved"] = edge_curves
   #random.seed(22) #15 #6 #9
   if coordinates is None:
       layout = g.layout("circular")#"kk"
   else:
```

```
layout = my_coordinates
    return(g,layout,visual_style)
def ols_annotations(x, y, data=None, ax=None, color='black', font_size=8,__
\rightarrowtextxy=[0.05,0.95], textva='top',
                    method='quantreg', stats=['N','slope','slope p']):
    import statsmodels.api as sm
    import statsmodels.formula.api as smf
    if data is None:
        data = pandas.DataFrame({'X':x,'Y':y})
        x = 'X'
        y = Y'
    data = data.sort_values(x)
    if method=='ols':
        X = sm.add_constant(data.loc[:,x])
        Y = data.loc[:,y]
        mod = sm.OLS(Y, X)
        res = mod.fit()
    elif method=='quantreg':
        mod = smf.quantreg(y+' ~ '+x, data)
        res = mod.fit(q=0.5)
    N = data.shape[0]
    slope = res.params[x]
    slope_p = res.pvalues[x]
    rsquared = res.rsquared_adj
    rsquared_p = res.f_pvalue
    text = ''
    for stat in stats:
        if stat=='N':
            text += 'N = \{:,\}\n'.format(N)
        if stat=='slope':
            text += 'slope = {}\n'.format('%.2f'%Decimal(slope))
        if stat=='slope_p':
            text += 'P = {}\n'.format('%.2E'%Decimal(slope_p))
        if stat=='rsquared':
            text += 'R2 = {}\n'.format('%.2f'%Decimal(rsquared))
        if stat=='rsquared p':
            text += 'P = {}\n'.format('%.2E'%Decimal(rsquared_p))
    ax.text(textxy[0], textxy[1], text, transform=ax.transAxes, va=textva,_u
→color=color, fontsize=font_size)
    xmin = data.loc[:,x].min()
    xmax = data.loc[:,x].max()
    ax.plot(data[x].values[[0,N-1]], res.predict()[[0,N-1]], color=color)
def get_quantile(observed, bs):
    q = observed.copy()
    q.loc[:,:] = numpy.nan
```

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for i in range(q.shape[0]):
        for c in range(q.shape[1]):
            gt_count = (observed.iloc[i,c]>bs[:,i,c]).sum()
            ge_count = (observed.iloc[i,c]>=bs[:,i,c]).sum()
            corrected_rank = (gt_count + ge_count)/2
            q.iloc[i,c] = corrected_rank/bs.shape[0]
    return q
def get quantile fdr(quantile, fdr threshold=0.001, tail='both'):
    q2 = quantile.copy()
    if tail=='both':
        q2[q2<0.5] = 1-q2[q2<0.5]
    f = q2.values.flat
    decimals = 0
    thresholds = numpy.array([])
    for d in numpy.arange(1,10):
        start = 1-(10**(-float(d-1)))
        end = 1-(10**(-float(d)))
        step = (10**(-float(d-1)))/1000
        tmp = numpy.arange(start, end, step)
        thresholds = numpy.concatenate((thresholds, tmp))
        if all(f==numpy.round(f, decimals=d)):
            decimals = d
            break
    if decimals!=0:
        print('Inferred number of permutations:', 10**decimals)
    else:
        print('Number of permutations could not be inferred.')
    num_data = q2.shape[0] * q2.shape[1]
    for i in numpy.arange(len(thresholds)):
        fdr = ((1-thresholds[i])*num_data)/((f > thresholds[i]).sum())
        if fdr<fdr_threshold:</pre>
            quantile_threshold = thresholds[i]
            print('FDR={}, Quantile threshold={}'.format(fdr,__
 →quantile_threshold))
            break
        assert i<(len(thresholds)-1), 'No quantile threshold satisfied FDR
    fdrs = q2 > quantile_threshold
    return fdrs
def draw_network2(observed, quantile, fdrs, self_arrow=False,_
self_vertex_size=False, show_vertex_stat=False, no_obs=True,
                 edge_width=False, edge_width_weight=None, hide_nonsig=False,_
→vertex_dict=None, coordinates=None,
                show_edge_color=True, scaled_edge_curve=False,__
 →relative_freq_edge_width=False, no_fp=False, sig_color='bluered'):
```

```
g = igraph.Graph(directed=True)
   if sig_color=='bluered':
      high_color = '#FF0000'
       low_color = '#0000FF'
   elif sig_color=='greenpink':
      high_color = '#FF00FF'
       low_color = '#00FF00'
   if vertex_dict is None:
       vertices = pandas.Series(list(set(observed.index.tolist()+observed.
vertex_colors = 'darkgray'
  else:
       vertices = pandas.Series(vertex_dict['vertex'])
       vertex_colors = pandas.Series(vertex_dict['color'])
  vertex_weights = list()
  vertex_labels = list()
  my coordinates = list()
  for v in vertices:
       if (v in observed.index)&(v in observed.columns):
           vertex_weights.append(observed.loc[v,v])
          n parent = str(int(observed.loc[v,:].sum().sum()))
           n_child = str(int(observed.loc[:,v].sum().sum()))
           v = v.replace('-', '\n')
           if show_vertex_stat:
               vertex_labels.append(v+'\n'+n_parent+':'+n_child)
           else:
               vertex_labels.append(v)
       else:
           vertex_labels.append(v)
           vertex_weights.append(1)
           if show_vertex_stat:
               v = v.replace('-', '\n')
               vertex_labels.append(v)
       if coordinates is not None:
           my_coordinates.append(coordinates[v])
  g.add_vertices(vertices)
   edge_weights = list()
   edge_colors = list()
   edge_labels = list()
  edge_curves = list()
  for v1 in observed.index:
       for v2 in observed.columns:
           flag = True if (self_arrow) | (self_vertex_size) else v1!=v2
           if (no_fp)&(v1 in ['F', 'P'])&(v2 in ['F', 'P']):
               flag = False
           if flag:
               v1id = vertices.loc[vertices==v1].index.values[0]
```

```
v2id = vertices.loc[vertices==v2].index.values[0]
               observed_value = observed.loc[v1,v2]
               edge_label = str(int(observed_value))
               color = igraph.color_name_to_rgba('#808080')
               if (fdrs.loc[v1,v2])&(quantile.loc[v1,v2]<0.5):
                   color = igraph.color_name_to_rgba(low_color)
               else:
                   if (not no_obs)&(observed.loc[v1,v2]==0):
                       continue
               if (fdrs.loc[v1,v2])&(quantile.loc[v1,v2]>0.5):
                   color = igraph.color name to rgba(high color)
               g.add_edges([(v1id,v2id),])
               edge_weight = 0.5
               if edge_width=='discrete':
                   if observed value >= 10:
                       edge_weight = 1
                   if observed_value >= 100:
                       edge_weight = 2
               elif edge_width=='identity':
                   edge_weight = observed_value+1
               elif edge_width=='log':
                   edge_weight = numpy.log2(observed_value+1)
               edge_weights.append(edge_weight)
               edge colors.append(color)
               edge_labels.append(edge_label)
               edge curve = -0.2 if (len(vertices) < 5) else -0.01
               if (v1 not in observed.columns) | (v2 not in observed.index):
                   edge_curve = 0
               if scaled_edge_curve:
                   v1coordinate = my_coordinates[v1id]
                   v2coordinate = my_coordinates[v2id]
                   euclidean_dist = numpy.linalg.norm(numpy.
→array(v1coordinate)-numpy.array(v2coordinate))
                   edge_curve = edge_curve * ((1+euclidean_dist)*2)
               edge_curves.append(edge_curve)
   if hide_nonsig:
       for i in range(len(edge_labels)):
           if edge_colors[i] == igraph.color_name_to_rgba('#808080'):
               edge labels[i] = ''
   if relative_freq_edge_width:
       edge_weights = [ ew/sum(edge_weights)*20 for ew in edge_weights ]
  visual_style = {}
  visual_style["vertex_size"] = [ v*20 for v in vertex_weights ] if_
⇒self_vertex_size else 26
  visual_style["vertex_label"] = vertex_labels
  visual_style["vertex_color"] = vertex_colors
  visual_style["vertex_label_color"] = 'black'
```

```
[4]: print('start:', "{0:%Y-%m-%d %H:%M:%S}".format(datetime.datetime.today()))
     conn = sqlalchemy.create_engine("sqlite:///"+db_file)
     #b = pandas.read_sql_query(sql="SELECT * from branch", con=conn, __

    index_col=None, coerce_float=True)

     #t = pandas.read_sql_query(sql="SELECT * from tree", con=conn, index_col=None,_
     →coerce float=True)
     b = pandas.read_sql_query(sql="SELECT * from branch", con=conn, index_col=None,_
     t = pandas.read_sql_query(sql="SELECT * from tree", con=conn, index_col=None,_
     →coerce float=True)
     conn.dispose()
     tissues = ['brain', 'heart', 'kidney', 'liver', 'ovary', 'testis']
     #b = integrate_pcm_stats(b, pcm_prefix)
     #t = integrate_pcm_stats(t, pcm_prefix)
     b.loc[:,'l1ou_intersect_is_shift'] = (b['l1ou_fpkm_is_shift'].fillna(0).
     →astype(bool))&(b['llou_tpm_is_shift'].fillna(0).astype(bool))
     b.loc[b.parent==0,'so_event_parent'] = 'No'
     b.loc[(b.spnode_coverage=='root')&(b.so_event_parent=='S'), 'so_event_parent'] =__

→ 'No'
     b['chromosome'] = b.loc[:,['A','X','Y']].idxmax(axis=1)
     for pp in pcm_prefixes:
        mus = [pp+'mu_'+o for o in_{\sqcup}]
     →['brain','heart','kidney','liver','ovary','testis']]
         b[pp+'max_mu'] = b.loc[:,mus].max(axis=1)
```

```
b[pp+'second_max_mu'] = numpy.sort(b.loc[:,mus].values)[:,-2]
   b[pp+'max second mu ratio'] = b[pp+'max mu'] - b[pp+'second max mu']
   pc = mus + [pp+'tau', __
b = attach_neighbor_stats(df=b, neighbor='parent', columns=pc)
   b = attach neighbor stats(df=b, neighbor='sister', columns=[pp+'is shift',])
for ext in [' omega',' dn',' ds']:
   cols = b.columns[b.columns.str.endswith(ext)]
   for col in cols:
       b['log_'+col] = numpy.log(b[col])
b = attach_neighbor_stats(df=b, neighbor='parent',__
columns=['l1ou_intersect_is_shift','chromosome','age','support_iqtree'])
sc =
→ ['delta_intron_present', 'hyphy_omega', 'mapdnds_omega', 'log_hyphy_omega', 'log_mapdnds_omega'
b = attach_neighbor_stats(df=b, neighbor='sister', columns=sc)
b.loc[(b.spnode_coverage.isnull()), 'spnode_coverage'] = sptree_root
for pcm_prefix in pcm_prefixes:
   for tis in tissues:
       b[pcm_prefix+'delta_mu_'+tis] = b[pcm_prefix+'mu_'+tis] -__
→b['parent_'+pcm_prefix+'mu_'+tis]
       b.loc[b.parent==0,pcm_prefix+'delta_mu_'+tis] = 0
   for prefix in ['mu_','delta_mu_']:
       targets = [ pcm_prefix+prefix+o for o in tissues ]
       b[pcm_prefix+prefix+'max'] = b.loc[:,targets].idxmax(axis=1)
       b[pcm_prefix+prefix+'max'] = b[pcm_prefix+prefix+'max'].str.
 →replace(pcm_prefix+prefix,'')
       b.loc[b.loc[:,targets].sum(axis=1)==0,pcm_prefix+prefix+'max'] = 'no'
   prefixes =
 → [pcm_prefix+'delta_mu_',pcm_prefix+'mu_','parent_'+pcm_prefix+'mu_',]
   newcols =
 → [pcm_prefix+'max_upregulation',pcm_prefix+'max_organ','parent_'+pcm_prefix+'max_organ']
   for prefix,newcol in zip(prefixes, newcols):
       cols = [ prefix+o for o in tissues ]
       b = add_max_mu(df=b, cols=cols, new_col=newcol)
       b[newcol] = b[newcol].str.replace(prefix,'')
is_parent_dup = b.so_event_parent=='D'
is_retrotransposition = (b.delta_intron_present<=max_delta_intron_present)</pre>
is_sister_retrotransposition = (b.
→sister_delta_intron_present<=max_delta_intron_present)</pre>
is_lower_delta_intron_present = (b.delta_intron_present<=b.</pre>
⇔sister_delta_intron_present)
```

```
is higher_dnds = (b[omega_method+'_omega']>=b['sister_'+omega_method+'_omega'])
b['branch_category'] = numpy.nan
b.loc[(~is_parent_dup), 'branch_category'] = 'S'
 →loc[(~is_retrotransposition)&(~is_sister_retrotransposition)&(is_parent_dup), 'pranch_catego
→loc[(is_retrotransposition)&(is_lower_delta_intron_present)&(is_parent_dup), 'branch_categor
b.loc[(b.so_event_parent.isnull()), 'branch_category'] = numpy.nan
b = attach_neighbor_stats(df=b, neighbor='parent', columns=['branch_category',])
b = attach_neighbor_stats(df=b, neighbor='sister',__

→columns=['branch_category','spnode_coverage'])
for sc in shift columns:
    conditions = True
    conditions = (conditions)&(b['spnode coverage']!=sptree root).fillna(False)
    conditions = (conditions)&(b['sister_spnode_coverage']!=sptree_root).
→fillna(False)
    conditions = (conditions)\&((b[sc]==1).fillna(False)|(b['sister_'+sc]==1).
 →fillna(False))
    conditions = (conditions) \& ((b[sc]==1).fillna(False) \& (b['sister_'+sc]==1).
 →fillna(False))
    conditions = (conditions) \& ((b['sister_branch_category'] == 'R') \& (b[sc] == 1).
→fillna(False))
    conditions =
\hookrightarrow (conditions) &~ ((b['branch_category']=='R') & (b['sister_'+sc]==1).
 →fillna(False))
    conditions = (conditions)&(b.branch_category!='No')
    b.loc[:,sc+'_pair'] = conditions
t = t.loc[(~t['l1ou_tpm_alpha_brain'].isnull())&(~t['l1ou_fpkm_alpha_brain'].
\rightarrowisnull()),:]
b = b.loc[(~b['llou tpm mu brain'].isnull())&(~b['llou fpkm mu brain'].
→isnull()),:]
b = b.loc[:,~b.columns.str.contains('phylogeneticem')]
t = t.loc[:,~t.columns.str.contains('phylogeneticem')]
print('Number of orthogroups in the tree table:', t['orthogroup'].unique().
\rightarrowshape [0])
print('Number of orthogroups in the branch table:', b['orthogroup'].unique().
\rightarrowshape [0])
criterion = 'l1ou_fpkm_alpha_brain'
is_leaf = (b['so_event']=='L')
```

```
analyzed_orthogroups = t.loc[(~t[criterion].isnull()), 'orthogroup']
num_analyzed_orthogroups = analyzed_orthogroups.shape[0]
is_analyzed = (b['orthogroup'].isin(analyzed_orthogroups))
spp = b.loc[(is_leaf)&(is_analyzed), 'node_name'].str.replace('_',' ', 1).str.
→replace('_.*','').unique()
num spp = spp.shape[0]
num_genes = b.loc[(is_leaf)&(is_analyzed),:].shape[0]
mean_num_gene_per_sp = num_genes / num_spp
num human genes = b.loc[(is leaf)&(is analyzed)&(b['node name'].str.
⇔startswith('Homo')),:].shape[0]
print('The number of species analyzed:', num_spp)
print('The number of orthogroups analyzed:', num_analyzed_orthogroups)
print('The average number of genes per species:', mean_num_gene_per_sp)
print('The number of analyzed genes:', num genes)
print('The number of analyzed human genes:', num_human_genes)
sci_names = b['spnode_coverage'].drop_duplicates()
for sn in sci names:
   if len(sn)>4:
        is_gene = (b['node_name'].str.startswith(sn)) &__
is introned gene = (is gene) & (b['num intron']>=1)
        is_intronless_gene = (is_gene) & (b['num_intron']==0)
       print(sn, '# gene:', is_gene.sum(), '# intron-containing:',
→is_introned_gene.sum(), '# intronless:', is_intronless_gene.sum())
is_root = (b['spnode_coverage'] == 'root')
is_shift = (b['llou_intersect_is_shift']==1)
num_all_shift = (~is_root&is_shift).sum()
print('Number of all shifts:', num all shift)
for event in b['branch_category'].unique():
    is event = (b['branch category'] == event)
   num_branch = ((~is_root)&(is_event)).sum()
   num_shift = ((~is_root)&is_event&is_shift).sum()
   print('Number of {} branches: {}'.format(event, num_branch))
   print('Number of {} shifts: {} or {}%'.format(event, num_shift, num_shift/
 →num_all_shift*100))
TEC cutoff = 0.5
num_higher_tec = (b.loc[is_shift,'llou_fpkm_mu_complementarity']>=TEC_cutoff).
print('Number of highly complementary shifts: {} or {}%. TEC threshold = {}'.
 →format(num_higher_tec, num_higher_tec/num_all_shift, TEC_cutoff))
```

```
print('Number of trees dated with non-RDS constraint: {}'.

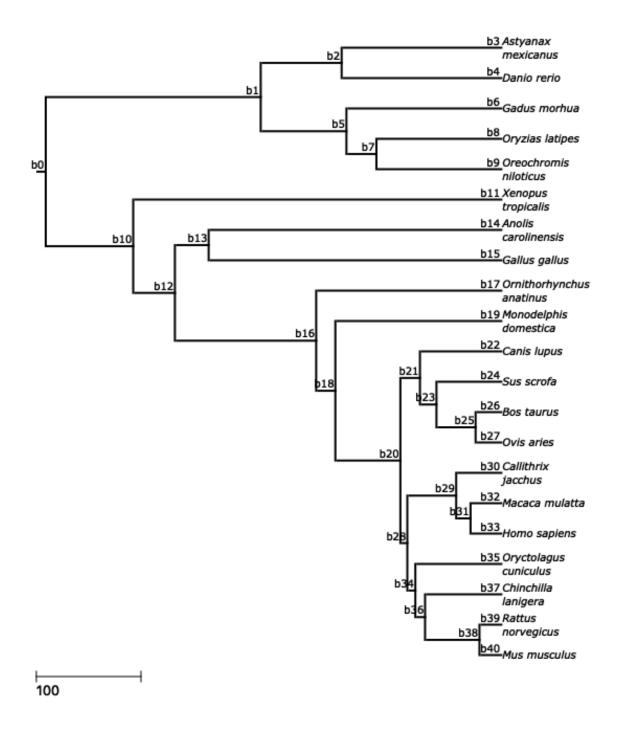
→format((t['dating_method']!='RDS').sum()))
tmp = b.loc[:,['orthogroup','mapdnds_omega','hyphy_omega']].replace([numpy.inf,_
 →-numpy.inf], numpy.nan).dropna()
sout = scipy.stats.spearmanr(tmp['mapdnds_omega'], tmp['hyphy_omega'])
pout = scipy.stats.pearsonr(tmp['mapdnds_omega'], tmp['hyphy_omega'])
print('All: Omega correlation between mapdNdS and Hyphy: {}'.format(sout))
print('All: Omega correlation between mapdNdS and Hyphy: {}'.format(pout))
print('end:', "{0:%Y-%m-%d %H:%M:%S}".format(datetime.datetime.today()))
c = 'orthogroup'
is_nonc = [ col!=c for col in t.columns ]
nonc = t.columns[is_nonc].tolist()
t = t.loc[:,[c,] + nonc]
t = t.sort_values(by=c, axis=0)
t.to_csv('orthogroup_statistics.tsv', sep='\t', index=False)
start: 2020-07-08 10:54:12
/Users/kef74yk/anaconda3/lib/python3.6/site-packages/pandas/core/series.py:679:
RuntimeWarning: divide by zero encountered in log
 result = getattr(ufunc, method)(*inputs, **kwargs)
l1ou_fpkm_max_upregulation : 584780 tie max values were detected
llou_fpkm_max_organ : O tie max values were detected
parent_l1ou_fpkm_max_organ : 0 tie max values were detected
l1ou_tpm_max_upregulation : 583600 tie max values were detected
llou_tpm_max_organ : O tie max values were detected
parent l1ou tpm max organ : O tie max values were detected
Number of orthogroups in the tree table: 15280
Number of orthogroups in the branch table: 15280
The number of species analyzed: 21
The number of orthogroups analyzed: 15280
The average number of genes per species: 15474.66666666666
The number of analyzed genes: 324968
The number of analyzed human genes: 20873
Bos_taurus # gene: 17022 # intron-containing: 15673 # intronless: 1349
Callithrix jacchus # gene: 16432 # intron-containing: 15526 # intronless: 906
Canis_lupus # gene: 16219 # intron-containing: 14775 # intronless: 1444
Chinchilla lanigera # gene: 15053 # intron-containing: 14604 # intronless: 449
Homo_sapiens # gene: 20873 # intron-containing: 19990 # intronless: 883
Macaca_mulatta # gene: 15771 # intron-containing: 14935 # intronless: 836
Mus musculus # gene: 19946 # intron-containing: 18970 # intronless: 976
Oryctolagus cuniculus # gene: 15108 # intron-containing: 13539 # intronless:
1569
```

```
Ovis_aries # gene: 15765 # intron-containing: 14504 # intronless: 1261
    Rattus_norvegicus # gene: 18573 # intron-containing: 16778 # intronless: 1795
    Sus scrofa # gene: 16871 # intron-containing: 15745 # intronless: 1126
    Anolis_carolinensis # gene: 12971 # intron-containing: 12029 # intronless: 942
    Astyanax mexicanus # gene: 15235 # intron-containing: 14319 # intronless: 916
    Danio_rerio # gene: 19628 # intron-containing: 18794 # intronless: 834
    Gallus gallus # gene: 13023 # intron-containing: 12229 # intronless: 794
    Oreochromis_niloticus # gene: 17734 # intron-containing: 16955 # intronless: 779
    Ornithorhynchus_anatinus # gene: 7467 # intron-containing: 7025 # intronless:
    442
    sp_root # gene: 0 # intron-containing: 0 # intronless: 0
    Oryzias_latipes # gene: 13633 # intron-containing: 13181 # intronless: 452
    Xenopus_tropicalis # gene: 13566 # intron-containing: 12774 # intronless: 792
    Monodelphis domestica # gene: 15424 # intron-containing: 13854 # intronless:
    1570
    Gadus_morhua # gene: 8654 # intron-containing: 8313 # intronless: 341
    Number of all shifts: 19636
    Number of S branches: 556240
    Number of S shifts: 11762 or 59.90018333672846%
    Number of D branches: 72476
    Number of D shifts: 6480 or 33.00061112242819%
    Number of R branches: 2985
    Number of R shifts: 1107 or 5.637604400081483%
    Number of nan branches: 0
    Number of nan shifts: 0 or 0.0%
    Number of highly complementary shifts: 18583 or 0.9463740069260542%. TEC
    threshold = 0.5
    Number of trees dated with non-RDS constraint: 5642
    All: Omega correlation between mapdNdS and Hyphy:
    SpearmanrResult(correlation=0.7278447626412824, pvalue=0.0)
    All: Omega correlation between mapdNdS and Hyphy: (-3.159100507681842e-06,
    0.9980921341552754)
    end: 2020-07-08 10:55:25
[5]: # prepare sptree
     sptree = ete3.PhyloNode(sptree_file, format=3)
     sptree.ladderize()
     node_id = 0
     for node in sptree.traverse(strategy='preorder'):
         node.name = node.name.replace("\'", "")
         node.id = 'b'+str(node_id)
         node_id += 1
         if node.is_root():
             node.name = sptree_root
     b['branch_id'] = 0
     for node in sptree.traverse(strategy='preorder'):
         b.loc[(b['spnode_coverage'] == node.name), 'branch_id'] = node.id
```

```
def my_layout(node):
   size_s = 6
   size_m = 10
   size_phylopic = 25
   nodeStyle = ete3.NodeStyle()
   nodeStyle["hz_line_width"] = nodeStyle["vt_line_width"] = 2
   nodeStyle["size"] = 0
   if node.is root():
       node.dist=10
   if node.is_leaf():
       if len(node.name)>15:
           leaf name = node.name.replace(' ', '\n')
       else:
           leaf_name = node.name.replace('_', ' ')
       leafnameFace = ete3.TextFace(leaf_name, ftype="Verdana", fsize=size_m,__
 ete3.add_face_to_node(face=leafnameFace, node=node, column=1,_
 →aligned=True, position="aligned")
    # branch id
   text branch = str(node.id)
   branchFace = ete3.TextFace(text_branch, fsize=size_m, fgcolor="black")
   branchFace.margin_bottom = branchFace.margin_right = branchFace.margin_top_
 →= branchFace.margin_left = 2
   ete3.add_face_to_node(face=branchFace, node=node, column=1, aligned=False,_
 →position="float")
   ete3.add_face_to_node(face=ete3.TextFace('', fsize=size_m), node=node,__
# phylopic images
   if False:
   #if node.is leaf():
       phylopic_file = [ file for file in os.listdir(phylopic_dir) if file.
 →startswith(node.name) ][0]
       paths, attributes = svg2paths(phylopic_dir+phylopic_file)
       for i in range(len(paths)):
           if i==0:
               xmin, xmax, ymin, ymax = paths[i].bbox()
           else:
               my_xmin, my_xmax, my_ymin, my_ymax = paths[i].bbox()
               xmin = my_xmin if my_xmin < xmin else xmin</pre>
               xmax = my_xmax if my_xmax > xmax else xmax
               ymin = my_ymin if my_ymin < ymin else ymin</pre>
               ymax = my_ymax if my_ymax > ymax else ymax
       width=None
       height=None
       if (ymax-ymin)>(xmax-xmin):
```

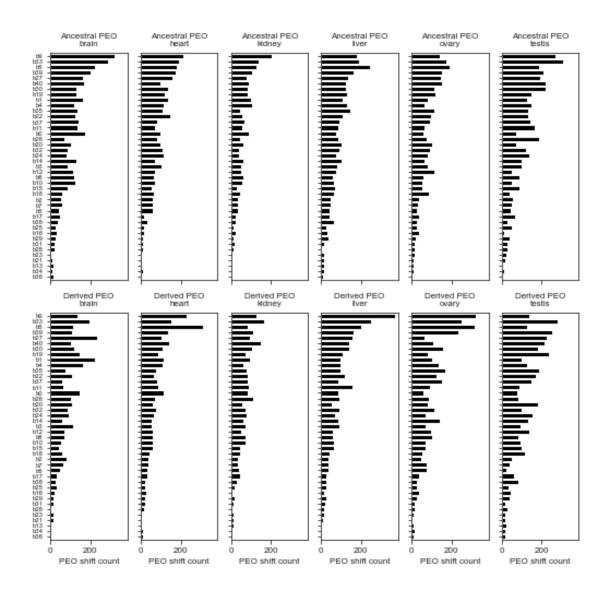
```
height=size_phylopic
                        else:
                                   width=size_phylopic
                       phylopicFace = ete3.SVGFace(phylopic_dir+phylopic_file, width=width,__
   →height=height)
                        ete3.add_face_to_node(face=phylopicFace, node=node, column=2,__
  →aligned=True, position="aligned")
           node.set_style(nodeStyle)
treeStyle = ete3.TreeStyle()
treeStyle.layout_fn = my_layout
treeStyle.scale = 1
treeStyle.min_leaf_separation = 29
treeStyle.show_leaf_name = False
treeStyle.scale_length = 100
#treeStyle.optimal_scale_level = "mid"
treeStyle.complete_branch_lines_when_necessary = False
treeStyle.allow_face_overlap = False
\textit{\#sptree.render(file\_name="annotated\_sptree.pdf", tree\_style=treeStyle, h=None, \_location for the property of the property 
 \rightarrow w=4.2, units="in", dpi=1200)
#sptree.render(file name="annotated sptree.svq", tree_style=treeStyle, h=None,_
  \rightarrow w=4.2, units="in", dpi=1200)
sptree.render(file_name="annotated_sptree.pdf", tree_style=treeStyle, h=None,_
 \hookrightarroww=3.0, units="in")
sptree.render(file_name="annotated_sptree.svg", tree_style=treeStyle, h=None,_
  \rightarroww=3.0, units="in")
sptree.render("%%inline", tree_style=treeStyle)
```

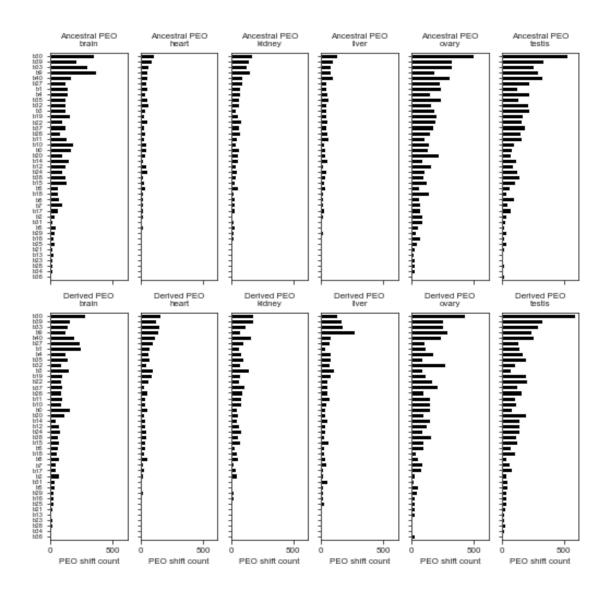
[5]:



```
[6]: for pp in pcm_prefixes:
    fig,axes = matplotlib.pyplot.subplots(nrows=2, ncols=len(organs),
    →figsize=(7.2, 7), sharex=True)
```

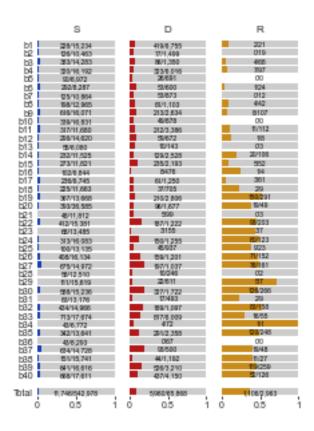
```
is_shift = (b[pp+'is_shift']==1)
  is_peo_shift = (b['parent_'+pp+'max_organ']!=b[pp+'max_organ'])
  is_target = (is_shift)&(is_peo_shift)
   #is_specific = (b[pp+'delta_tau']>0.5)
   #is_target = (is_target)&(is_specific)
  tmp = b.loc[is target,:]
   #tmp['o2o'] = tmp['parent_'+pp+'max_organ'] + '_' + tmp[pp+'max_organ']
  index_order = tmp['branch_id'].value_counts()
  for j,col in enumerate(['parent_'+pp+'max_organ',pp+'max_organ']):
       for k,organ in enumerate(organs):
           ax = axes[j,k]
           dat = tmp.loc[(tmp[col]==organ), 'branch_id'].value_counts()
           dat = dat.loc[index_order.index]
           seaborn.barplot(x=dat, y=dat.index, color='black', ax=ax)
           if k!=0:
                ax.set_yticklabels([''] * len(ax.get_yticklabels()),__
→fontsize=font_size-2)
           else:
                ax.set_yticklabels(ax.get_yticklabels(), fontsize=font_size-2)
           if j==1:
               ax.set_title('Derived PEO\n'+organ, fontsize=font_size)
               ax.set_xlabel('PEO shift count', fontsize=font_size)
           else:
               ax.set_title('Ancestral PEO\n'+organ, fontsize=font_size)
               ax.set_xlabel('', fontsize=font_size)
  fig.tight_layout()
  file_base = 'branch_wise_peo_shift_'+pp
  for ext in ['png','pdf','svg']:
       fig.savefig(file_base+'.'+ext, format=ext)
```





```
tmp = tmp.iloc[numpy.argsort([ int(s[1:]) for s in tmp.index ]),:]
placeholders = pandas.DataFrame([[numpy.nan,]*tmp.shape[1]], columns=tmp.
→columns, index=['',])
totals = pandas.DataFrame([tmp.sum(axis=0).tolist()], columns=tmp.columns,
→index=['Total',])
tmp = pandas.concat([tmp, placeholders, totals], axis=0)
for i in range(len(branch_categories)):
   tmp_my = tmp0.
→loc[(tmp0['branch_category']==branch_categories[i]),'bl_dated']
   nbranch = (tmp0['branch category'] == branch categories[i]).sum()
   nshift =
→((tmp0[shift_col]==1)&(tmp0['branch_category']==branch_categories[i])).sum()
   total_my = tmp_my.sum()
   txt = '{}; Shift frequency = {} shift/My; # shifts = {:,}; # branches = {:
 \hookrightarrow,}; Total branch MY = {:,}'
   print(txt.format(branch_categories[i], nshift/total_my, nshift, nbranch,_u
→total_my))
   tmp2 = pandas.DataFrame(tmp.loc[:,tmp.columns.
→get_level_values(0)==branch_categories[i]])
    tmp2.columns = ['no shift', 'shift']
   tmp3 = tmp2.copy()
   sum values = tmp2.sum(axis=1)
   tmp2.loc[(tmp2['shift']<min_nshift).tolist(),:] = numpy.nan</pre>
   for c in tmp2.columns:
       tmp2.loc[:,c] = tmp2.loc[:,c] / sum_values
   tmp2 = tmp2.reset_index(drop=False)
    \#tmp2 = tmp2.iloc[numpy.argsort([int(s[1:]) for s in tmp2['branch_id']]),:
   ax = kfplot.stacked_barplot(x=['shift', 'no_shift'], y='index', data=tmp2,__
 \rightarrowax=axes[i],
ax.set xlim(0, 1)
   ax.set xlabel('')
   ax.set ylabel('')
   ax.set_title(branch_categories[i], fontsize=font_size)
   ax.tick_params(axis='both', which='major', direction='out', length=2,_
→width=1, pad=1, top=False, left=False, right=False)
   xticks = [0, 0.5, 1]
   ax.set_xticks(xticks, minor=False)
   ax.set_xticklabels([ str(tick) for tick in xticks ], minor=False,_
⇔ha='center')
   ax.set_xticklabels(ax.get_xticklabels(), rotation=0, fontsize=font_size-1)
   ax.set_yticklabels(ax.get_yticklabels(), rotation=0, fontsize=font_size-1)
   for j in range(tmp3.shape[0]):
```

```
nshift = tmp3['shift'].iloc[j]
        nbranch = tmp3.iloc[j,:].sum()
        if (nshift==nshift)&(nbranch==nbranch):
            percent = numpy.round(nshift/nbranch*100, decimals=1)
             \#txt = str(percent) + '\% ('+str(int(nshift)) + '/' + str(int(nbranch)) + ')'
            txt = "{:,}/{:,}".format(int(nshift), int(nbranch))
            ax.text(x=0.5, y=j, s=txt, color='black', ha='center', va='center', u
 →fontsize=5)
    if i%num_category!=0:
        ax.get_yaxis().set_visible(False)
    for part in ['top','bottom','left','right']:
        ax.spines[part].set_visible(False)
file_base = 'proportion_shift'
for ext in ['png','pdf','svg']:
    fig.savefig(file_base+'.'+ext, format=ext)
# Chisq test
contingency_table = numpy.array([
     [tmp.loc['Total',('S',True)].astype(int), tmp.loc['Total',('S',False)].
 →astype(int)],
     [tmp.loc['Total',('D',True)].astype(int), tmp.loc['Total',('D',False)].
 →astype(int)],
     [tmp.loc['Total',('R',True)].astype(int), tmp.loc['Total',('R',False)].
 →astype(int)],
])
out = scipy.stats.chi2 contingency(observed=contingency_table, correction=True,__
 →lambda_=None)
chi,p,dof,expected = out
print('Chi-square test among 3 branch categories: chisq={:,}, P={}, dof={}'.
 →format(chi, p, dof))
S; Shift frequency = 0.00024804574942087113 shift/My; # shifts = 11,746; #
branches = 542,978; Total branch MY = 47,354,167.63812388
D; Shift frequency = 0.0016886637493336888 shift/My; # shifts = 5,960; #
branches = 65,868; Total branch MY = 3,529,417.8621123894
R; Shift frequency = 0.007029358246006006 shift/My; # shifts = 1,106; # branches
= 2,963; Total branch MY = 157,340.11004893875
/Users/kef74yk/anaconda3/lib/python3.6/site-packages/ipykernel_launcher.py:49:
RuntimeWarning: invalid value encountered in double_scalars
Chi-square test among 3 branch categories: chisq=21,064.636194292412, P=0.0,
dof=2
```



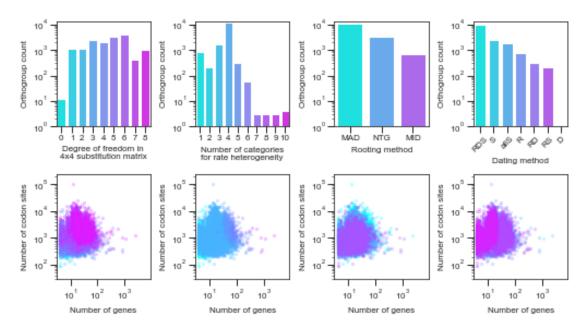
```
[8]: criterion = 'iqtree_best_BIC'
     alpha = 0.2
     cols = ['model_df','model_rate','rooting_method','dating_method']
     cmaps = ['cool','cool','cool','cool']
     hist_xlabels = ['Degree of freedom in\n4x4 substitution matrix','Number of_
      →categories\nfor rate heterogeneity', 'Rooting method', 'Dating method']
     fig,axes = matplotlib.pyplot.subplots(nrows=2, ncols=len(cols), figsize=(7.
      \rightarrow 2,4), sharex=False)
     model_df = {'JC':0,'JC69':0,'F81':3,'K80':1,'K2P':1,'HKY':4,'HKY85':4,'TN':
      →5, 'TN93':5, 'TNe':2, 'K81':2, 'K3P':2,
            'K3Pu':2,'K81u':5,'TPM2':2,'TPM2u':5,'TPM3':2,'TPM3u':5,'TIM':6,'TIMe':
      →3, 'TIM2':6, 'TIM2e':3, 'TIM3':6,
            'TIM3e':3,'TVM':7,'TVMe':4,'SYM':5,'GTR':8,}
     t['model_df'] = t[criterion]
     t['model_df'] = t['model_df'].str.replace('\+.*','')
     for key in model_df.keys():
         t.loc[t['model_df'] == key, 'model_df'] = model_df[key]
```

```
t['model_rate'] = t[criterion]
t.loc[t['model_rate'].str.contains('\+[GR]'),'model_rate'] = t.
→loc[t['model_rate'].str.contains('\+[GR]'), 'model_rate'].str.replace('.
→*\+[GR]','').str.replace('\+.*','')
t.loc[~t['model_rate'].str.contains('^[0-9]'), 'model_rate'] = 1
t.loc[:,'model rate'] = t['model rate'].astype(int)
print('The number of orthogroups analyzed for phylogeny:', (~t[criterion].
→isnull()).sum())
#print('The number of orthogroups with nonzero parsimony-informative site:', __
\hookrightarrow (t['prop_parsimony_informative_cleaned']!=0).sum())
print('The number of trees with +G models:', t[criterion].str.contains('\+G').
\rightarrowsum())
print('The number of trees with +R models:', t[criterion].str.contains('\+R').
print('The number of trees with *H models:', t[criterion].str.contains('\*H').
\rightarrowsum())
print('The number of trees with no rate heterogeneity:', (~t[criterion].str.
-replace('\+I','').str.replace('\+F','').str.contains('\+')).sum())
for j in range(len(cols)):
    col = cols[j]
    print(col)
    if col=='rooting_method':
        t[col] = t[col].str.replace('NOTUNG','NTG').str.
 →replace('midpoint','MID')
    elif col=='dating method':
        # some RDS trees were not annotated correctly.
        conditions = (t[col].isnull())&(~t['llou fpkm alpha brain'].isnull())
        t.loc[conditions,col] = 'RDS'
    print(t.loc[:,[col,'orthogroup']].groupby(col).count().
 →to_dict()['orthogroup'])
    df_unique = t[col].dropna().drop_duplicates()
    df_unique = sorted(df_unique.tolist())
    num_df = len(df_unique)
    cmap = matplotlib.cm.get_cmap(cmaps[j])
    df_rgba = cmap(numpy.arange(0,1.001, 1/num_df))
    for i in range(len(df rgba)):
        df_rgba[i][3] = alpha
    ax = axes[0,j]
    val = t[col]
    val = pandas.DataFrame(val.value_counts())
    ax = seaborn.barplot(x=val.index, y=val[col], ax=ax, palette=df rgba)
    ax.set_yscale('log', basey=10)
    ax.set_ylim(1,13000)
```

```
ax.set_xlabel(hist_xlabels[j])
    ax.set_ylabel('Orthogroup count')
    if col=='dating_method':
        ax.set_xticklabels(ax.get_xticklabels(), rotation=45)
    ax = axes[1,j]
    for i in numpy.arange(num_df):
        tmp = t.loc[t[col] == df_unique[i],:]
        if tmp.shape[0] == 1:
            tmp = tmp.append(pandas.Series(), ignore_index=True)
        ax = seaborn.regplot(x='cleaned_num_seq', y='cleaned_num_site',_

data=tmp, fit_reg=False,
                              color=df_rgba[i], scatter_kws={'alpha':
 →alpha, 'rasterized':True, 's':5}, ax=ax)
    ax.set_xscale('log', basex=10)
    ax.set_yscale('log', basey=10)
    ax.set_xlabel('Number of genes')
    ax.set_ylabel('Number of codon sites')
    ax.set_xlim(3, 2**13*1.1)
    ax.set_ylim(10**1*3, 10**6/4)
    ax.set_xticks([10,100,1000])
for ax in axes.flat:
    ax.tick_params(axis='both', which='major', direction='out', length=6, __
 →width=1, pad=2, top=False, right=False, labelsize=font_size)
    ax.tick_params(axis='both', which='minor', top=False, right=False,_u
 →labelsize=font_size)
fig.tight_layout()
outbase = 'tree_inference'
fig.savefig(outbase+".pdf", format='pdf', transparent=True)
fig.savefig(outbase+".svg", format='svg', transparent=True)
The number of orthogroups analyzed for phylogeny: 15280
The number of trees with +G models: 10880
The number of trees with +R models: 3531
The number of trees with *H models: 0
The number of trees with no rate heterogeneity: 869
model df
{0: 12, 1: 1107, 2: 1122, 3: 2371, 4: 1990, 5: 3304, 6: 3963, 7: 408, 8: 1003}
model rate
{1: 869, 2: 201, 3: 1693, 4: 12135, 5: 312, 6: 57, 7: 3, 8: 3, 9: 3, 10: 4}
rooting_method
{'MAD': 11237, 'MID': 662, 'NTG': 3381}
dating_method
{'D': 1, 'R': 758, 'RD': 299, 'RDS': 9638, 'RS': 216, 'S': 2558, 'allS': 1810}
/Users/kef74yk/anaconda3/lib/python3.6/site-packages/ipykernel_launcher.py:63:
```

DeprecationWarning: The default dtype for empty Series will be 'object' instead of 'float64' in a future version. Specify a dtype explicitly to silence this warning.



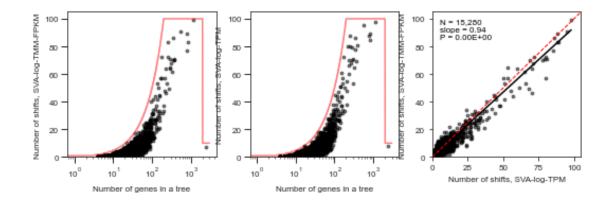
```
[9]: fig,axes = matplotlib.pyplot.subplots(nrows=1, ncols=3, figsize=(7.2,2.4),
     →sharex=False)
     axes = axes.flat
     num_leaves = numpy.arange(0,3000, 1)
     halves = numpy.floor(num leaves/2).astype(int)
     halves[halves>100] = 100
     sqroots = numpy.floor(num_leaves**0.5).astype(int)
     max nshifts = halves
     max_nshifts[max_nshifts<sqroots] = sqroots[max_nshifts<sqroots]</pre>
     df_max_nshift = pandas.DataFrame({'num_leaf':num_leaves,'max_nshift':
     →max_nshifts})
     df_max_nshift.loc[(df_max_nshift['num_leaf']>=2000),'max_nshift'] = 10
     color = 'black'
     ymax = 105
     for pp,ax in zip(pcm_prefixes,axes):
         tmp = t.loc[(~t[pp+'alpha_brain'].isnull()),:]
         tmp = pandas.merge(tmp, df_max_nshift, left_on='cleaned_num_seq',_
      →right on='num leaf')
         num_tree = tmp.shape[0]
         num_max_shift = (tmp[pp+'num_shift'] == tmp['max_nshift']).sum()
```

```
label = '(N = '+str(num_tree)+', # of trees reaching the limit = __

    '+str(num_max_shift)+')'

    print(pp, label)
    ax = seaborn.regplot('cleaned_num_seq', pp+'num_shift', data=tmp,__
 →fit_reg=False, ax=ax, color=color,
                         scatter_kws={'alpha':0.5, 'rasterized':True, 's':8},__
→label=label)
    ax.plot('num_leaf', 'max_nshift', data=df_max_nshift, color='red', alpha=0.
→5, label='Upper limit in regime shift inference')
    ax.set_ylim(0,ymax)
    ax.set_xscale('log', basex=10)
    #ax.set_yscale('log', basex=2)
    ax.tick_params(axis='both', which='major', direction='out', length=6,__
 →width=1, pad=2, top=False, right=False, labelsize=font_size)
    ax.tick_params(axis='both', which='minor', top=False, right=False,_
→labelsize=font_size)
    ax.set_xlabel('Number of genes in a tree')
    ax.set_ylabel('Number of shifts, SVA-log-TMM-FPKM' if pp=='l1ou fpkm_' else_
→'Number of shifts, SVA-log-TPM')
ax = axes[2]
seaborn.regplot('l1ou_tpm_num_shift', 'l1ou_fpkm_num_shift', t, fit_reg=False,_
⇒ax=ax, color=color,
                scatter kws={'alpha':0.5,'rasterized':True,'s':8})
ols_annotations('l1ou_tpm_num_shift', 'l1ou_fpkm_num_shift', t, ax=ax)
ax.set_xlabel('Number of shifts, SVA-log-TPM')
ax.set_ylabel('Number of shifts, SVA-log-TMM-FPKM')
ax.plot([0,ymax],[0,ymax], color='red', lw=1, linestyle='--')
ax.set_xlim(0,ymax)
ax.set_ylim(0,ymax)
fig.tight_layout(pad=0)
outbase = 'ou nshift'
fig.savefig(outbase+".pdf", format='pdf', transparent=True)
fig.savefig(outbase+".svg", format='svg', transparent=True)
```

```
l1ou\_fpkm\_ (N = 15280, # of trees reaching the limit = 0) l1ou\_tpm\_ (N = 15280, # of trees reaching the limit = 3)
```



```
[10]: import matplotlib_venn
     fig,axes = matplotlib.pyplot.subplots(nrows=1, ncols=3, figsize=(7.2,2.4),__
      →sharex=False)
     axes = axes.flat
     label_fpkm = 'FPKM'#'log TMM-SVA-FPKM'
     label_tpm = 'TPM'#'log SVA-TPM'
     colors = ['magenta','cyan']
     tmp = b.copy()
     shift_cols = ['l1ou_tpm_is_shift','l1ou_fpkm_is_shift']
     window_search_col = shift_cols[0]
     labels = ['TPM','FPKM']
     window sizes = [0,1,]
     neighbors = ['parent','sister','child1','child2']
     id_cols =_
      →['orthogroup', 'numerical_label', 'parent', 'sister', 'child1', 'child2', 'branch_category']
     tmp = b.loc[:,id_cols+shift_cols]
     for sc in shift_cols:
         tmp.loc[:,sc] = tmp.loc[:,sc].astype(bool)
     for neighbor in neighbors:
         tmp = attach_neighbor_stats(df=tmp, neighbor=neighbor,__
      tmp.loc[:,'ogb_id'] = tmp['orthogroup'] + tmp['numerical_label'].astype(str)
     def my_formatter(s):
         return "{:,}".format(s)
```



```
file_og_cor = 'orthogroup_correlation.tsv'
if (os.path.exists(file_og_cor))&(~force):
    print('Reading', file_og_cor)
    og_cor = pandas.read_csv(file_og_cor, sep='\t', header=0)
else:
    og_cor = pandas.DataFrame({'orthogroup':orthogroups,'pearson':numpy.

¬nan, 'spearman':numpy.nan})
    for i in numpy.arange(len(orthogroups)):
        tmp_og = tmp.loc[(tmp['orthogroup']==orthogroups[i]),:]
        og cor.loc[i, 'pearson'] = scipy.stats.pearsonr(x=tmp_og['hyphy_omega'],_

    y=tmp_og['mapdnds_omega'])[0]

        og_cor.loc[i,'spearman'] = scipy.stats.
 ⇒spearmanr(a=tmp_og['hyphy_omega'], b=tmp_og['mapdnds_omega']).correlation
        if i%1000==0:
            print("{0:%Y-%m-%d %H:%M:%S}".format(datetime.datetime.today()), ',u
 →processing', i, 'th orthogroups')
    og_cor.to_csv(file_og_cor, sep='\t', index=False)
\#kfplot.density\_scatter(x='hyphy\_omega', y='mapdnds\_omega', df=tmp, ax=ax, 
→cor=True, diag=False, reg family=None, hue log=True)
#seaborn.regplot('hyphy_omega', 'mapdnds_omega', data=tmp, fit_reg=False)
```

Number of branches with omega annotations: 607873 Number of orthogroups with omega annotations: 15280 Reading orthogroup_correlation.tsv

```
[12]: | params = [pp+'alpha_',pp+'sigma2_',pp+'gamma_']
     ymaxs = [0.08, 0.15, 5]
     ylabels = ['$$ (adaptation rate)', '$^2$ (expression drift variance)', '$$_{\sqcup}
      for pp in pcm_prefixes:
         fig,axes = matplotlib.pyplot.subplots(nrows=1, ncols=3, figsize=(9,3),__
      →sharex=False)
         for i in range(len(params)):
             ax = axes[i]
             df2 = t
             df3 = df2.loc[:,df2.columns.str.startswith(params[i])].melt()
             df3['variable'] = df3['variable'].str.replace(params[i],'')
             ymin = -0.001
             lw = 2
             alpha = 0.95
             colors = ['#1B9E77','#D95F02','#7570B3','#E7298A','#66A61E','#E6AB02']
             texty = (ymaxs[i] - ymin) * 0.925
```

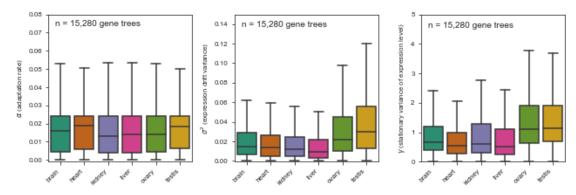
```
seaborn.boxplot(x='variable', y='value', data=df3, ax=ax,u

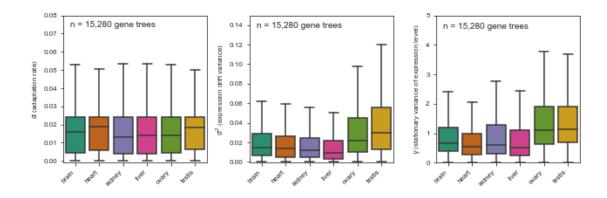
palette=colors, fliersize=0)

ax.set_ylabel(ylabels[i])
ax.set_xlabel('')
ax.set_ylim(ymin, ymaxs[i])
my_text='n = '+"{:,d}".format(df3.loc[df3.variable=='brain','variable'].

shape[0])+' gene trees'
ax.text(x=-0.2, y=texty, s=my_text, fontsize=10, ha='left', va='center')
ax.set_xticklabels(labels=ax.get_xticklabels(), rotation=45, ha='right')

matplotlib.pyplot.tight_layout()
outbase = 'sigma_boxplot_'+pp
fig.savefig(outbase+".pdf", format='pdf')
fig.savefig(outbase+".svg", format='yvg')
```





```
[13]: ys = ['log_mapdnds_omega','log_hyphy_omega']
#ylabs = ['Log $d_N/d_S$','Log $d_N/d_S$',]
ylabs = ['Log ', 'Log ']
#ys = ['log_mapdnds_omega', 'log_mapdnds_dn', 'log_mapdnds_ds']
```

```
#ylabs = ['Log $d_N/d_S$', 'Log $d_N$', 'Log $d_S$']
num_col=len(ys)
num_row=1
matplotlib.rcParams['font.size'] = 10
alpha_small=0.1
alpha large=0.95
kernel='gau'
spnodes = b.spnode coverage.unique()
is parent dup = b.so event parent=='D'
is_retrotransposition = (b.delta_intron_present<=max_delta_intron_present)</pre>
is sister retrotransposition = (b.
→sister_delta_intron_present<=max_delta_intron_present)</pre>
is lower_delta_intron_present = (b.delta_intron_present<=b.</pre>
⇒sister_delta_intron_present)
if 'flat' in dir(axes):
    axes = axes.flat
else:
    axes = [axes,]
for sc in ['l1ou_intersect_is_shift']:
    print(sc)
    fig,axes = matplotlib.pyplot.subplots(nrows=num_row, ncols=num_col,_
 →figsize=(3.4*num_col,3.6*num_row), sharex=False)
    for y,ylab,ax in zip(ys,ylabs,axes):
        print('\n', y)
        tmp = b.loc[:,['orthogroup',y]]
        \#tmp[y] = tmp[y].replace([-numpy.inf,numpy.inf], numpy.nan)
        log_var_value = tmp.groupby('orthogroup')[y].median().var()
        var_value = numpy.exp(log_var_value)
        print('Variance of ortholog median (unlog):', var_value)
        df2 = b.loc[(b[sc+'_pair']),:]
        df2 = df2.loc[~(df2[y].isnull()|df2['sister_'+y].isnull()),:]
        df2.loc[(df2['sister_branch_category']=='R'), 'branch_category'] = 'R'
        df2.loc[:,y] = df2.loc[:,y].clip(-10, 10)
        df2['dup shift'] = ''
        for bc in branch_categories:
            df2.loc[(df2.branch category==bc)&(df2[sc]==0), 'dup shift'] = bc+'-'
            df2.loc[(df2.branch_category==bc)&(df2[sc]==1), 'dup_shift'] = bc+'+'
        df2 = df2.dropna(subset=[y,], axis=0)
        orders = []
        colors = []
```

```
for bc in branch_categories:
           orders = orders + [bc+'-', bc+'+']
           colors = colors + [category_colors[bc], category_colors[bc]]
       seaborn.boxplot(x='dup_shift', y=y, data=df2, ax=ax, showfliers=False, u
→palette=colors, order=orders)
       \#ax.set xlim(-0.6, 3.6)
       \#ax.set\_xticks([0,1,2,3], minor=False)
       \#ax.set\_xticks([-0.6, -0.6, 0.5, 2.5], minor=True)
       \#ax.set\_xticklabels(['-','+','-','+',], minor=False, ha='center')
       #ax.set_xticklabels(['Exp. shift','\nPrec._
\rightarrow event', '\nSpeciation', '\nDuplication',], minor=True, ha='center')
       ax.tick params(axis='x', which='major', direction='out', length=6,,,
⇒width=1)
       ax.tick_params(axis='x', which='minor', direction='out', length=6, u
\rightarrowwidth=0)
       ax.set_xlabel('')
       pos=0
       lowest = numpy.inf
       highest = -numpy.inf
       for ev in branch categories:
           for sh in [0,1]:
                dat = df2.loc[(df2.branch_category==ev)&(df2[sc]==sh),y].values
                Q1, median, Q3 = numpy.percentile(numpy.asarray(dat), [25, 50, __
→75])
                IQR = Q3 - Q1
               loval = Q1 - 1.5 * IQR
               hival = Q3 + 1.5 * IQR
               wiskhi = numpy.compress(dat <= hival, dat)</pre>
               wisklo = numpy.compress(dat >= loval, dat)
               actual_hival = numpy.max(wiskhi)
                actual loval = numpy.min(wisklo)
               if actual_hival > highest:
                    highest = actual_hival
                if actual_loval < lowest:</pre>
                    lowest = actual_loval
                \#label\_text = "\{:,d\}".format(len(dat))
                label_text = "{}".format(numpy.round(numpy.exp(numpy.
→median(dat)), decimals=3))
               ax.text(x=pos, y=actual_hival+0.8, s=label_text,__

→fontsize=font_size, ha='center', va='center')
               pos+=1
                del dat
       #ax.set_ylim(lowest-0.1, highest+4)
       ax.set_ylim(-10.1, 10.1)
```

```
ax.set_ylabel(ylab)
        ax.tick_params(axis='both', which='major', direction='out', length=6, __
 →width=1, pad=1, top=False, right=False)
        for event in ['S','D','R']:
            xval = df2.loc[df2['dup shift'] == event+'-',y].values
             yval = df2.loc[df2['dup_shift'] == event+'+',y].values
             #out = kfstat.brunner munzel test(xval, yval,
 \rightarrow alternative="two-sided")
             (W, dof, p, Pest, Cl, Ch) = kfstat.bm_test(xval, yval, ttype=1,__
 \rightarrowalpha=0.05) #ttype, 1 = greater,-1 = lesser, 0 = two-sided
             print(event, 'Number of branch pairs =', len(xval))
            print(event, 'Brunner-Munzel stat =', W)
             print(event, 'P value =', p)
             print(event, 'Effect size and 95% CI =', Pest, Cl, Ch)
             print(event, 'Unlog median value of '+event+'- =', numpy.exp(numpy.
 →median(xval)))
             print(event, 'Unlog median value of '+event+'+ =', numpy.exp(numpy.
 →median(yval)))
    print(numpy.finfo(p))
    #del df2
    fig.tight_layout()
    outbase = 'dNdS_dN_dS_boxplot_'+sc
    fig.savefig(outbase+".pdf", format='pdf')
    fig.savefig(outbase+".svg", format='svg')
l1ou_intersect_is_shift
log_mapdnds_omega
Variance of ortholog median (unlog): 4.052727336760715
S Number of branch pairs = 11498
S Brunner-Munzel stat = 3.710725473681451
S P value = 0.0001035766799742932
S Effect size and 95% CI = 0.5141220684965032 0.5066625558204125
0.5215815811725939
S Unlog median value of S- = 0.09653370712268128
S Unlog median value of S+ = 0.10195232477385784
D Number of branch pairs = 5529
D Brunner-Munzel stat = 7.07301168594131
D P value = 8.040235144335384e-13
D Effect size and 95% CI = 0.5387574799620318 0.5280164239456809
0.5494985359783827
D Unlog median value of D- = 0.18236911096662003
D Unlog median value of D+ = 0.24378035689145436
R Number of branch pairs = 1075
```

```
R Brunner-Munzel stat = 25.299145968747506
```

- R P value = 0.0
- R Effect size and 95% CI = 0.7654909680908599 0.7449107814342645
- 0.7860711547474554
- R Unlog median value of R- = 0.035579136470610964
- R Unlog median value of R+ = 0.3942508012820513

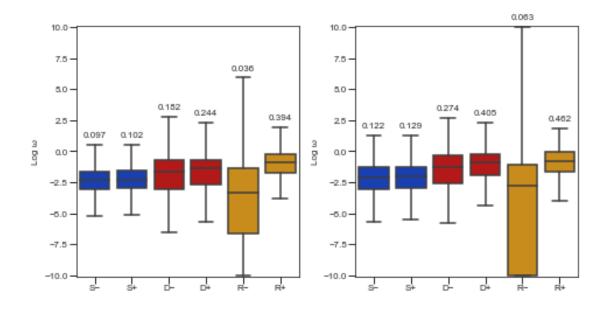
log_hyphy_omega

Variance of ortholog median (unlog): nan

- S Number of branch pairs = 11290
- S Brunner-Munzel stat = 3.342935721371553
- S P value = 0.00041516114605388843
- S Effect size and 95% CI = 0.5128365673158168 0.5053100801497592
- 0.5203630544818745
- S Unlog median value of S- = 0.12181926686467885
- S Unlog median value of S+ = 0.12919372346526056
- D Number of branch pairs = 4119
- D Brunner-Munzel stat = 9.9103346216216
- D P value = 0.0
- D Effect size and 95% CI = 0.5626106872379674 0.5502263445942862
- 0.5749950298816485
- D Unlog median value of D- = 0.2737190043195873
- D Unlog median value of D+ = 0.4051894631593254
- R Number of branch pairs = 833
- R Brunner-Munzel stat = 18.51086007403166
- R P value = 0.0
- R Effect size and 95% CI = 0.7296966805930055 0.705357239650684
- 0.7540361215353271
- R Unlog median value of R- = 0.06323508882314291
- R Unlog median value of R+ = 0.4619892444123987

Machine parameters for float64

```
precision = 15 resolution = 1.000000000000001e-15
machep = -52 eps = 2.2204460492503131e-16
negep = -53 epsneg = 1.1102230246251565e-16
minexp = -1022 tiny = 2.2250738585072014e-308
maxexp = 1024 max = 1.7976931348623157e+308
nexp = 11 min = -max
```



```
[14]: y = 'mapdnds_delta_omega'
      ylabel = 'Δ Log $d_N/d_S$\n(shift br. - sister br.)'
      ymin = -10
      ymax = 10
      alpha = 0.1
      ps = 3
      mk = 'o'
      ols_method = 'ols' # ols or quantreg
      show_xlab = False
      no\_colors = [(0.5, 0.5, 0.5), (0, 0, 0)]
      negative_colors = matplotlib.cm.get_cmap('Paired').colors[0:2]
      positive_colors = matplotlib.cm.get_cmap('Paired').colors[6:8]
      stats = ['N','slope','slope_p']
      textys = [0.3, 0.95]
      events = ['S+','D+','R+']
      titles = ['S branches', 'D branches', 'R branches']
      for sc in ['llou_intersect_is_shift',]:
          print(sc)
          for pp in pcm_prefixes:
              print(pp)
              fig,axes = matplotlib.pyplot.subplots(nrows=3, ncols=3, figsize=(4.8, 4.
       ⇔8))
              df2 = b.loc[b[sc+'_pair'],:]
              df2.loc[(df2['sister_branch_category']=='R'), 'branch_category'] = 'R'
              df2.loc[:,'dup_shift'] = ''
```

```
for bc in branch_categories:
           df2.loc[(df2.branch_category==bc)&(df2[sc]==0), 'dup_shift'] = bc+'-'
           df2.loc[(df2.branch_category==bc)&(df2[sc]==1), 'dup_shift'] = bc+'+'
       for i,event in enumerate(events):
           dat = df2.loc[(df2[sc]==1)&(df2.dup_shift==event),:]
           dat.loc[:,'mapdnds_delta_omega'] =__
→dat['log_mapdnds_omega']-dat['sister_log_mapdnds_omega']
           j = 0
           ax = axes[j,i]
           x = pp+'delta_tau'
           ax.axhline(y=0, linestyle='--', color='black', lw=0.5)
           for colors, conditions, texty in_
→zip([negative_colors,positive_colors],[(dat[x]<0),(dat[x]>0)],textys):
               seaborn.regplot(x, y, data=dat.loc[conditions,:],_
→fit_reg=False, truncate=True, ax=ax, color=colors[0], marker=mk,
                               scatter_kws={'alpha':alpha,'rasterized':
→True, 's':ps}, line_kws={'color':colors[1]})
               ols annotations(x, y, dat.loc[conditions,:], ax, colors[1],
→font_size, textxy=[0.05,texty], method=ols_method,
                               stats=stats)
           ax.set_xlim(-1,1)
           ax.set_ylim(ymin,ymax)
           if show_xlab:
               ax.set_xlabel('Shift in organ\nexp. specificity ()' if i%3==1,,
→else '')
           else:
               ax.set_xlabel('')
           ax.xaxis.set ticks([-1,0,1])
           ax.set_ylabel(ylabel if (i==0)&(j==1) else '')
           if i!=0:
               ax.yaxis.set_ticklabels(['']*len(ax.get_yticklabels()))
           ax.set_title(titles[i], fontsize=font_size)
           j = 1
           ax = axes[j,i]
           x = pp+'delta_maxmu'
           ax.axhline(y=0, linestyle='--', color='black', lw=0.5)
           for colors, conditions, texty in_
-zip([negative_colors,positive_colors],[(dat[x]<0),(dat[x]>0)],textys):
               seaborn.regplot(x, y, data=dat.loc[conditions,:],__
→fit_reg=False, truncate=True, ax=ax, color=colors[0], marker=mk,
                               scatter_kws={'alpha':alpha,'rasterized':
→True,'s':ps}, line_kws={'color':colors[1]})
```

```
ols_annotations(x, y, dat.loc[conditions,:], ax, colors[1],__

→font size, textxy=[0.05,texty], method=ols_method,
                               stats=stats)
           ax.set xlim(-15,15)
           ax.set_ylim(ymin,ymax)
           expression unit = 'FPKM' if pp=='llou fpkm' else 'TPM'
           if show xlab:
               ax.set_xlabel('Shift in max expression\nlevel (Δ Log_L)
→'+expression_unit+')' if i%3==1 else '')
           else:
               ax.set_xlabel('')
           ax.xaxis.set ticks([-10,0,10])
           ax.set_ylabel(ylabel if (i==0)&(j==1) else '')
           if i!=0:
               ax.yaxis.set_ticklabels(['']*len(ax.get_yticklabels()))
           j = 2
           ax = axes[j,i]
           x = pp+'mu_complementarity'
           ax.axhline(y=0, linestyle='--', color='black', lw=0.5)
           seaborn.regplot(x, y, data=dat, fit_reg=False, truncate=False,
→ax=ax, color=no_colors[0], marker=mk,
                           scatter_kws={'alpha':alpha,'rasterized':True,'s':
→ps}, line_kws={'color':no_colors[1]})
           ols_annotations(x, y, dat, ax, no_colors[1], font_size, textxy=[0.
→05,0.95], method=ols_method, stats=stats)
           ax.set xlim(0,1)
           ax.set ylim(ymin,ymax)
           if show_xlab:
               ax.set_xlabel('Expression complementarity\nbetween sister_
→lineages' if i%3==1 else '')
           else:
               ax.set_xlabel('')
           ax.xaxis.set_ticks([0,0.5,1])
           ax.xaxis.set_ticklabels(['0','0.5','1'])
           ax.set_ylabel(ylabel if (i==0)&(j==1) else '')
           if i!=0:
               ax.yaxis.set_ticklabels(['']*len(ax.get_yticklabels()))
       fig.tight_layout()
       #fig.subplots_adjust(left=0, right=1, bottom=0, top=1)
       outbase = 'dNdS_vs_exp_properties_'+pp+sc
       fig.savefig(outbase+".pdf", format='pdf')
       fig.savefig(outbase+".svg", format='svg')
```

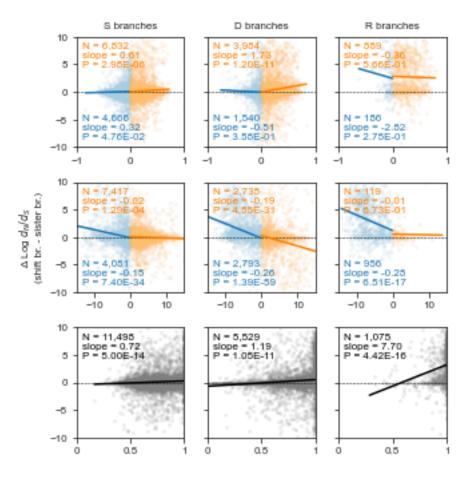
11ou_intersect_is_shift
11ou_fpkm_

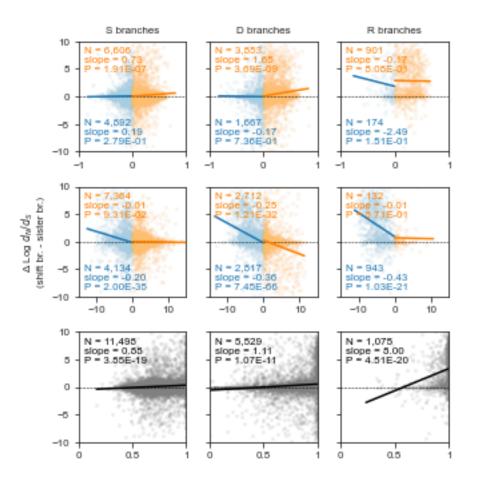
```
/Users/kef74yk/anaconda3/lib/python3.6/site-
packages/pandas/core/indexing.py:966: SettingWithCopyWarning:
A value is trying to be set on a copy of a slice from a DataFrame.
Try using .loc[row_indexer,col_indexer] = value instead
See the caveats in the documentation: https://pandas.pydata.org/pandas-
docs/stable/user guide/indexing.html#returning-a-view-versus-a-copy
  self.obj[item] = s
/Users/kef74yk/anaconda3/lib/python3.6/site-
packages/pandas/core/indexing.py:845: SettingWithCopyWarning:
A value is trying to be set on a copy of a slice from a DataFrame.
Try using .loc[row_indexer,col_indexer] = value instead
See the caveats in the documentation: https://pandas.pydata.org/pandas-
docs/stable/user_guide/indexing.html#returning-a-view-versus-a-copy
  self.obj[key] = _infer_fill_value(value)
/Users/kef74yk/anaconda3/lib/python3.6/site-
packages/pandas/core/indexing.py:845: SettingWithCopyWarning:
A value is trying to be set on a copy of a slice from a DataFrame.
Try using .loc[row_indexer,col_indexer] = value instead
See the caveats in the documentation: https://pandas.pydata.org/pandas-
docs/stable/user_guide/indexing.html#returning-a-view-versus-a-copy
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docs/stable/user_guide/indexing.html#returning-a-view-versus-a-copy
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/Users/kef74yk/anaconda3/lib/python3.6/site-
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A value is trying to be set on a copy of a slice from a DataFrame.
Try using .loc[row_indexer,col_indexer] = value instead
See the caveats in the documentation: https://pandas.pydata.org/pandas-
docs/stable/user_guide/indexing.html#returning-a-view-versus-a-copy
 self.obj[item] = s
```

```
l1ou_tpm_
/Users/kef74yk/anaconda3/lib/python3.6/site-
packages/pandas/core/indexing.py:966: SettingWithCopyWarning:
A value is trying to be set on a copy of a slice from a DataFrame.
Try using .loc[row_indexer,col_indexer] = value instead
See the caveats in the documentation: https://pandas.pydata.org/pandas-
docs/stable/user_guide/indexing.html#returning-a-view-versus-a-copy
  self.obj[item] = s
/Users/kef74yk/anaconda3/lib/python3.6/site-
packages/pandas/core/indexing.py:845: SettingWithCopyWarning:
A value is trying to be set on a copy of a slice from a DataFrame.
Try using .loc[row_indexer,col_indexer] = value instead
See the caveats in the documentation: https://pandas.pydata.org/pandas-
docs/stable/user_guide/indexing.html#returning-a-view-versus-a-copy
  self.obj[key] = _infer_fill_value(value)
/Users/kef74yk/anaconda3/lib/python3.6/site-
packages/pandas/core/indexing.py:845: SettingWithCopyWarning:
A value is trying to be set on a copy of a slice from a DataFrame.
Try using .loc[row_indexer,col_indexer] = value instead
See the caveats in the documentation: https://pandas.pydata.org/pandas-
docs/stable/user guide/indexing.html#returning-a-view-versus-a-copy
  self.obj[key] = _infer_fill_value(value)
/Users/kef74yk/anaconda3/lib/python3.6/site-
packages/pandas/core/indexing.py:966: SettingWithCopyWarning:
A value is trying to be set on a copy of a slice from a DataFrame.
Try using .loc[row_indexer,col_indexer] = value instead
See the caveats in the documentation: https://pandas.pydata.org/pandas-
docs/stable/user_guide/indexing.html#returning-a-view-versus-a-copy
  self.obj[item] = s
/Users/kef74yk/anaconda3/lib/python3.6/site-
packages/pandas/core/indexing.py:845: SettingWithCopyWarning:
A value is trying to be set on a copy of a slice from a DataFrame.
Try using .loc[row_indexer,col_indexer] = value instead
See the caveats in the documentation: https://pandas.pydata.org/pandas-
docs/stable/user_guide/indexing.html#returning-a-view-versus-a-copy
  self.obj[key] = _infer_fill_value(value)
/Users/kef74yk/anaconda3/lib/python3.6/site-
packages/pandas/core/indexing.py:966: SettingWithCopyWarning:
A value is trying to be set on a copy of a slice from a DataFrame.
Try using .loc[row_indexer,col_indexer] = value instead
```

See the caveats in the documentation: https://pandas.pydata.org/pandas-

docs/stable/user_guide/indexing.html#returning-a-view-versus-a-copy
 self.obj[item] = s





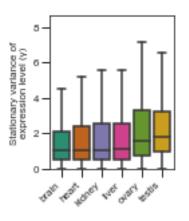
```
[15]: ylims=[8.7,4.5]
      for pp,ymax in zip(pcm_prefixes,ylims):
          print(pp)
          fig,axes = matplotlib.pyplot.subplots(nrows=1, ncols=1, figsize=(2,2.3),__
       →sharex=False)
          ax = axes
          x=pp+'gamma_'
          df3 = t.loc[:,t.columns.str.startswith(x)].melt()
          df3['variable'] = df3['variable'].str.replace(x,'')
          xmin=-0.6
          xmax=5.6
          ymin = -0.05
          lw=2
          alpha=0.95
          colors=['#1B9E77','#D95F02','#7570B3','#E7298A','#66A61E','#E6AB02']
          order = ['brain', 'heart', 'kidney', 'liver', 'ovary', 'testis']
          seaborn.boxplot(x='variable', y='value', data=df3, ax=ax, palette=colors,
       ⇒showfliers=False, order=order)
```

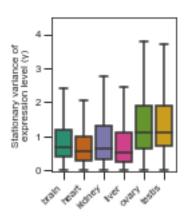
```
ax.set_ylabel('Stationary variance of\nexpression level ()')
   ax.set_xlabel('')
   ax.set_xlim(xmin, xmax)
   ax.set_ylim(ymin, ymax)
   ax.set_xticklabels(order, rotation=45, ha='right')
   xloc = xmin+((xmax-xmin)/100*4)
   yloc = ymax-((ymax-ymin)/100*2)
   my_{text}='N = '+"\{:,d\}".format(df3.loc[(df3.variable=='brain')\&(~df3.value.)]
→isnull()), 'variable'].shape[0])+' gene trees'
   fout = scipy.stats.friedmanchisquare(t[x+'brain'].values, t[x+'heart'].
→values, t[x+'kidney'].values,
                                t[x+'liver'].values, t[x+'ovary'].values,
→t[x+'testis'].values, )
   chistat = fout[0]
   chitext = '^2 = '+"{:.2e}".format(Decimal(str(chistat)))
   pval = fout[1]
   sign = '='
   if pval==0:
       pval = numpy.finfo(type(pval)).eps
       sign = '<'
   pval = "{:.2e}".format(Decimal(str(pval)))
   ptext = 'P '+sign+' '+pval
   my text = my text+'\n'+chitext
   my_text = my_text+'\n'+ptext
   #ax.text(x=xloc, y=yloc, s=my_text, fontsize=font_size, ha='left', va='top')
   ax.tick_params(axis='both', which='major', direction='out', length=6,__
→width=1, pad=2, top=False, right=False)
   matplotlib.pyplot.tight_layout()
   outbase = x+'organ_'+pp
   fig.savefig(outbase+".pdf", format='pdf', transparent=True)
   fig.savefig(outbase+".svg", format='svg', transparent=True)
   control = list()
   control = control + t[pp+'gamma brain'].dropna().tolist()
   control = control + t[pp+'gamma_heart'].dropna().tolist()
   control = control + t[pp+'gamma_kidney'].dropna().tolist()
   control = control + t[pp+'gamma_liver'].dropna().tolist()
   target = list()
   target = target + t[pp+'gamma_ovary'].dropna().tolist()
   target = target + t[pp+'gamma_testis'].dropna().tolist()
   (W, dof, p, Pest, Cl, Ch) = kfstat.bm_test(control, target)
   print(pp, 'Brunner-Munzel stat =', W)
   print(pp, 'P value =', p)
   print(pp, 'Effect size and 95% CI =', Pest, Cl, Ch)
```

```
print(pp, 'Pest - Cl =', Pest - Cl)
print(pp, 'Ch - Pest =', Ch - Pest)
print()
```

```
l1ou_fpkm_
l1ou_fpkm_ Brunner-Munzel stat = 56.767067346862625
l1ou_fpkm_ P value = 0.0
l1ou_fpkm_ Effect size and 95% CI = 0.6081602634333626 0.60442581428692
0.6118947125798052
l1ou_fpkm_ Pest - Cl = 0.003734449146442609
l1ou_fpkm_ Ch - Pest = 0.003734449146442609

l1ou_tpm_
l1ou_tpm_ Brunner-Munzel stat = 94.8233631302835
l1ou_tpm_ P value = 0.0
l1ou_tpm_ Effect size and 95% CI = 0.6736075821146624 0.6700191196059939
0.6771960446233309
l1ou_tpm_ Pest - Cl = 0.0035884625086685107
l1ou_tpm_ Ch - Pest = 0.0035884625086685107
```





```
[16]: #req_family = statsmodels.qenmod.families.family.NegativeBinomial()
     reg_family = statsmodels.genmod.families.family.Poisson()
      #req_family = statsmodels.genmod.families.family.Gaussian()
     num row=2
     num_col=2
     alpha small=0.1
     alpha_large=0.9
     kernel='gau'
      # full figure size = 9.7 length x 7.2 width in inches
     #fig,axes = matplotlib.pyplot.subplots(nrows=num_row, ncols=num_col, figsize=(2.
      \rightarrow2*num_col,2.2*num_row), sharex=False)
     xs = ['delta_tau','delta_maxmu','mu_complementarity']
     xmins = {'delta_tau':-0.45,'delta_maxmu':-11,'mu_complementarity':0}
     xmaxs = {'delta_tau':0.75,'delta_maxmu':11,'mu_complementarity':1}
     #axis= {'delta_tau':axes.flat[1],'delta_maxmu':axes.
      \rightarrow flat[2], 'mu_complementarity':axes.flat[3]}
     xlabels = {'delta_tau':'Shift in expression specificity',
                 'delta_maxmu': 'Shift in max expression level',
                 'mu_complementarity':'Expression complementarity\nbetween sister_
      →lineages'}
     zero vlines = {'delta tau':False,'delta maxmu':False,'mu complementarity':False}
     conditions = True
     conditions = conditions&(b['spnode_coverage']!='root')
     box_step = 0.15
     for sc in shift_columns:
         \#ax = axes[0,0]
         col1 = 'branch category'
         col2 = 'spnode_coverage'
         df_branch = b.loc[(b[col1]!='No'),:].pivot_table(index=col2, columns=col1,__
      →values='orthogroup', aggfunc='count').fillna(0)
         df shift = b.loc[(b[col1]!='No')&(b[sc]==1),:].pivot table(index=col2,...)
      df_bl = b.loc[(b[col1]!='No'),:].pivot_table(index=col2, columns=col1,__
      →values='bl_dated', aggfunc='sum').fillna(0)
         df prop = df shift / df bl
         df_prop = df_prop.drop(sptree_root)
         df_prop = pandas.DataFrame(df_prop.stack())
         conf95_lowers = dict()
```

```
conf95_uppers = dict()
    slopes = dict()
    for bc in branch_categories:
        dat = pandas.DataFrame(df_shift[bc])
        dat['bl'] = df_bl[bc]
        #formula = bc+'\sim1'
        formula = bc+' \sim bl-1'
        #mod = statsmodels.formula.api.glm(formula=formula, data=dat,__
 → family=reg_family, freq_weights=dat['bl'])
        mod = statsmodels.formula.api.glm(formula=formula, data=dat,__
 →family=reg_family)
        res = mod.fit()
        conf95_lowers[bc] = res.conf_int().loc['bl',0]
        conf95_uppers[bc] = res.conf_int().loc['bl',1]
        slopes[bc] = res.params['bl']
        print(sc, bc, 'num_shift =', dat[bc].sum(), 'total_bl =', dat['bl'].

¬sum(), 'shift/MY =', dat[bc].sum()/dat['bl'].sum())
    print(sc,'conf95_lower =', conf95_lowers)
    print(sc,'conf95_upper =', conf95_uppers)
    print(sc,'slope =', slopes)
llou_fpkm_is_shift S num_shift = 23985.0 total_bl = 47435809.20286555 shift/MY =
0.0005056306702268946
l1ou_fpkm_is_shift D num_shift = 9018.0 total_bl = 4739356.061232859 shift/MY =
0.001902790143531467
l1ou fpkm is shift R num shift = 1238.0 total bl = 163542.86835132877 shift/MY =
0.007569880683152035
l1ou_fpkm_is_shift conf95_lower = {'S': 2.183119445576086e-06, 'D':
6.106426293159801e-06, 'R': 0.00028209263803218287}
l1ou_fpkm_is_shift conf95_upper = {'S': 2.1947428232433673e-06, 'D':
6.184910517345311e-06, 'R': 0.0002916342426424933}
l1ou_fpkm_is_shift slope = {'S': 2.1889311344097268e-06, 'D':
6.145668405252556e-06, 'R': 0.0002868634403373381}
l1ou_tpm_is_shift S num_shift = 25518.0 total_bl = 47435809.20286555 shift/MY =
0.0005379480276360182
11ou_{tpm} is shift D num shift = 8694.0 total bl = 4739356.061232859 shift/MY =
0.0018344264257997974
l1ou_tpm_is_shift R num_shift = 1196.0 total_bl = 163542.86835132877 shift/MY =
0.007313067283562063
l1ou_tpm_is_shift conf95_lower = {'S': 2.1774986916571217e-06, 'D':
6.029995119022642e-06, 'R': 0.0002810877949159398}
liou tpm is shift conf95 upper = {'S': 2.189240661272377e-06, 'D':
6.112092914266233e-06, 'R': 0.00029071850272207754}
l1ou tpm is shift slope = {'S': 2.1833696764647494e-06, 'D':
6.0710440166444375e-06, 'R': 0.00028590314881900866}
liou intersect is shift S num shift = 11762.0 total bl = 47435809.20286555
shift/MY = 0.0002479561368859176
```

```
shift/MY = 0.0013672743546333894
     l1ou_intersect_is_shift R num_shift = 1107.0 total_bl = 163542.86835132877
     shift/MY = 0.006768867460621408
     liou intersect is shift conf95 lower = {'S': 1.9712411712291976e-06, 'D':
     5.7516899231161895e-06, 'R': 0.0002766506739228304}
     liou intersect is shift conf95 upper = {'S': 1.98823158605984e-06, 'D':
     5.848362005042187e-06, 'R': 0.00028668393881475}
     l1ou_intersect_is_shift slope = {'S': 1.9797363786445187e-06, 'D':
     5.800025964079188e-06, 'R': 0.0002816673063687902}
[17]: num_row=1
     num_col=4
     alpha_small=0.1
     alpha_large=0.9
     kernel='gau'
     box_step = 0.15
     # full figure size = 9.7 length x 7.2 width in inches
     for sc in ['llou_intersect_is_shift',]:
         for pp in pcm_prefixes:
             print(sc, pp)
             fig,axes = matplotlib.pyplot.subplots(nrows=num_row, ncols=num_col,__
      →figsize=(7.2+2.4,2.6*num_row), sharex=False)
             if pp=='l1ou_fpkm_':
                 expression_unit = 'SVA-log-TMM-FPKM'
              elif pp=='llou tpm ':
                 expression_unit = 'SVA-log-TPM'
             xs = [pp+'delta_tau',pp+'delta_maxmu',pp+'mu_complementarity']
             xmins = {pp+'delta_tau':-0.3,pp+'delta_maxmu':
      xmaxs = {pp+'delta_tau':0.6,pp+'delta_maxmu':10,pp+'mu_complementarity':
       →1}
              axis= {pp+'delta_tau':axes.flat[1],pp+'delta_maxmu':axes.

→flat[2],pp+'mu_complementarity':axes.flat[3]}
             xlabels = {pp+'delta_tau':'Change in expression\nspecificity ($Δ$)',
                        pp+'delta_maxmu':'Change in max_

→expressionlevel\n($Δ _{max}$, w/ '+expression_unit+')',
                        pp+'mu\_complementarity':'Expression \ complementarity \backslash nbetween_{\sqcup}
      zero_vlines = {pp+'delta_tau':False,pp+'delta_maxmu':
       →False,pp+'mu_complementarity':False}
             conditions = True
             conditions = conditions&(b['spnode_coverage']!='root')
```

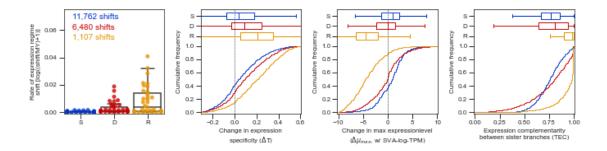
l1ou_intersect_is_shift D num_shift = 6480.0 total_bl = 4739356.061232859

```
for x in xs:
          box_position = 1 + (box_step*len(branch_categories))
          yticks = [0.0, 0.2, 0.4, 0.6, 0.8, 1.0]
          bins=numpy.arange(xmins[x]-((xmaxs[x]-xmins[x])/50),_{\square}
\rightarrowxmaxs[x]+((xmaxs[x]-xmins[x])/50), (xmaxs[x]-xmins[x])/100)
          ax = axis[x]
          if 'delta ' in x:
               ax.axvline(x=0, lw=0.5, linestyle='--', color='black')
          df_tmp=b.loc[(b[sc]==1)&(conditions),:]
          ax = kfplot.hist_boxplot(x=x, category='branch_category',__

→df=df_tmp, colors=category_colors, xlim=[xmins[x],xmaxs[x]], bins=bins,

\rightarrowalpha=0.9, box_step=0.15, ax=ax)
          ax.set xlabel(xlabels[x])
          for bc1,bc2 in itertools.combinations(branch_categories, 2):
              v1 = df_tmp.loc[(df_tmp['branch_category']==bc1),x].values
              v2 = df_tmp.loc[(df_tmp['branch_category']==bc2),x].values
              statistic,pvalue = scipy.stats.ks_2samp(v1, v2,__
→alternative='two-sided', mode='auto')
              print('{}; {}-{}: D = {:.2}, P = {:.2}'.format(x, bc1, bc2, __
⇒statistic, pvalue))
      ax = axes.flat[0]
      col1 = 'branch_category'
      col2 = 'spnode_coverage'
      df_branch = b.loc[(b[col1]!='No'),:].pivot_table(index=col2,__
df shift = b.loc[(b[col1]!='No')&(b[sc]==1),:].pivot table(index=col2,...)
df bl = b.loc[(b[col1]!='No'),:].pivot_table(index=col2, columns=col1,__
→values='bl_dated', aggfunc='sum').fillna(0)
      df prop = df shift / df bl
      df_prop = df_prop.drop(sptree_root)
      df prop = pandas.DataFrame(df prop.stack())
      df_prop = numpy.log(df_prop+1)
      colors = [ category colors[bc] for bc in branch categories ]
      seaborn.boxplot(x=df_prop.index.get_level_values(1), y=0, data=df_prop,_u
→order=branch_categories, palette=colors, ax=ax, boxprops={'facecolor':
→'None'}, showfliers=False)
      seaborn.swarmplot(x=df_prop.index.get_level_values(1), y=0,__
→data=df_prop, order=branch_categories, palette=colors, ax=ax, alpha=0.8)
      ax.set ylim(-0.001, 0.075)
      ax.set_ylabel('Rate of expression regime\nshift [log((shift/MY)+1)]')
      ax.set xlabel('')
      ymin,ymax = ax.get_ylim()
      label_y = ymax - ((ymax-ymin)*0.035)
```

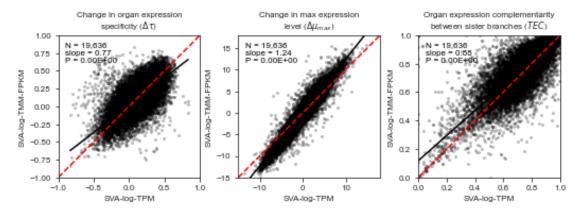
```
for ev in branch_categories:
             num shift = b.loc[(b.branch_category==ev)&(b[sc]==1)&(conditions),:
 \rightarrow] . shape [0]
             num text = "{:,d}".format(num shift)+' shifts'
             color = category_colors[ev]
             ax.text(-0.25, label y, num text, va='top', ha='left', color=color)
             label_y = label_y - ((ymax-ymin)*0.1)
         for ax in axes.flat:
             ax.tick_params(axis='both', which='major', direction='out', ___
 →length=6, width=1, pad=2, top=False, right=False)
         fig.tight layout()
         outbase = 'shiftStats2_'+pp+sc
         fig.savefig(outbase+".pdf", format='pdf', transparent=True)
         fig.savefig(outbase+".svg", format='svg', transparent=True)
llou intersect is shift llou fpkm
l1ou_fpkm_delta_tau; S-D: D = 0.12, P = 7.1e-49
llou_fpkm_delta_tau; S-R: D = 0.26, P = 6.4e-62
l1ou_fpkm_delta_tau; D-R: D = 0.16, P = 2.7e-21
l1ou_fpkm_delta_maxmu; S-D: D = 0.17, P = 3.9e-101
11ou fpkm delta maxmu; S-R: D = 0.64, P = 0.0
llou_fpkm_delta_maxmu; D-R: D = 0.48, P = 8.6e-186
llou_fpkm_mu_complementarity; S-D: D = 0.17, P = 1.8e-107
l1ou_fpkm_mu_complementarity; S-R: D = 0.59, P = 8.4e-305
llou_fpkm_mu_complementarity; D-R: D = 0.42, P = 1.5e-146
l1ou_intersect_is_shift l1ou_tpm_
l1ou_tpm_delta_tau; S-D: D = 0.1, P = 1.4e-39
llou_tpm_delta_tau; S-R: D = 0.31, P = 7.3e-84
l1ou tpm delta tau; D-R: D = 0.21, P = 2.1e-37
llou_tpm_delta_maxmu; S-D: D = 0.19, P = 3.2e-131
l1ou tpm delta maxmu; S-R: D = 0.61, P = 4.9e-323
l1ou_tpm_delta_maxmu; D-R: D = 0.48, P = 1.5e-191
l1ou_tpm_mu_complementarity; S-D: D = 0.17, P = 9e-104
l1ou_tpm_mu_complementarity; S-R: D = 0.59, P = 1.2e-305
l1ou_tpm_mu_complementarity; D-R: D = 0.43, P = 1.1e-151
           11,762 shifts
           6,480 shifts
                           R.
                                                                  1.0 -
                           1.0 -
                                              1.0
       0.04
                           0.5 -
                                              0.5
                           0.6
                                              0.6
                                                                  0.6
       0.02
                           0.4
                                              0.4
                                Change in expre
                                 specificity (\Delta T)
                                                 (Δμ<sub>max</sub>, w/ SVA-log-TMM-FPKM)
```



```
[18]: fig,axes = matplotlib.pyplot.subplots(nrows=1, ncols=3, figsize=(7.2,2.5),
       ⇒sharex=False)
      axes = axes.flat
      alpha = 0.2
      size = 4
      conditions = True
      conditions = conditions&(b['spnode_coverage']!='root')
      conditions = conditions&(b['llou_intersect_is_shift']==1)
      tmp = b.loc[conditions,:]
      xs = ['delta_tau','delta_maxmu','mu_complementarity']
      labels = ['Change in organ expression\nspecificity ($Δ$)',
                'Change in max expression\nlevel ($\Delta_{\max}$)',
                'Organ expression complementarity\nbetween sister branches ($TEC$)'
      xyranges = [[-1,1],[-15,18],[0,1]]
      for i,x in enumerate(xs):
          ax = axes[i]
          seaborn.regplot('l1ou_tpm_'+x, 'l1ou_fpkm_'+x, data=tmp, ax=ax,_

color='black', fit_reg=False,
                          scatter_kws={'alpha':alpha,'s':size,'rasterized':True})
          \#pearson_r = numpy.round(scipy.stats.pearsonr(x=tmp['l1ou_tpm_'+x], 
       \rightarrow y = tmp['llow_fpkm_' + x])[0], decimals = 2)
          \#cor\ text = "Pearson's r = {}".format(pearson r)
          #ax.set title(labels[i]+'\n'+cor text)
          ols_annotations('l1ou_tpm_'+x, 'l1ou_fpkm_'+x, data=tmp, ax=ax)
          ax.set title(labels[i])
          ax.set_xlabel('SVA-log-TPM')
          ax.set_ylabel('SVA-log-TMM-FPKM')
          ax.set_xlim(xyranges[i])
          ax.set_ylim(xyranges[i])
          ax.plot(xyranges[i], xyranges[i], linestyle='--', color='red')
```

```
fig.tight_layout(pad=0)
outbase = 'shiftStats_correlation'
fig.savefig(outbase+".pdf", format='pdf', transparent=True)
fig.savefig(outbase+".svg", format='svg', transparent=True)
```

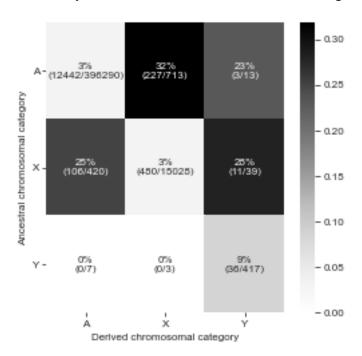


```
[19]: fig,axes = matplotlib.pyplot.subplots(nrows=1, ncols=1, figsize=(4,4),__
       ⇒sharex=False)
      shift_col = 'l1ou_intersect_is_shift'
      var1 = 'parent chromosome'
      var2 = 'chromosome'
      var_ph = 'orthogroup'
      excluded_spp =
      →'Astyanax_mexicanus|Danio_rerio|Gadus_morhua|Oryzias_latipes|Oreochromis_niloticus|Xenopus_
      excluded_spp = [ sp.replace('_','_') for sp in excluded_spp.split('|') ]
      coordinates = {
          'A':[0, -1],
          'X':[-0.8660254037844386467637, 0.5],
          'Y': [0.8660254037844386467637, 0.5],
      }
      excluded_spnodes = list()
      for n in sptree.traverse():
          contain_excluded_spp = all([ node_sp in excluded_spp for node_sp in n.
       →get_leaf_names() ])
          if contain_excluded_spp:
              excluded_spnodes.append(n.name)
      print('excluded spnodes in chromosome analysis:', excluded_spnodes)
      is_excluded_node = b['spnode_coverage'].isin(excluded_spnodes)
```

```
b2 = b.loc[~is_excluded_node,:]
is chr shift = (b2['parent chromosome']!=b2['chromosome'])
is_exp_shift = b2[shift_col].fillna(0).astype(bool)
is_all = True
# show pivot table
pivot_all = pandas.DataFrame(b2.groupby([var1,var2])[var_ph].count()).
→reset_index().pivot(var1, var2).fillna(0)
pivot_chr_shift = pandas.DataFrame(b2.loc[is_chr_shift,:].
 →groupby([var1,var2])[var_ph].count()).reset_index().pivot(var1, var2).
\rightarrowfillna(0)
pivot_exp_shift = pandas.DataFrame(b2.loc[is_exp_shift,:].
 →groupby([var1,var2])[var_ph].count()).reset_index().pivot(var1, var2).
\rightarrowfillna(0)
pivot_exp_all_ratio = pivot_exp_shift/pivot_all
pivot_exp_all_ratio.columns = pivot_exp_all_ratio.columns.get_level_values(1)
#IPython.display.display(pivot_exp_all_ratio)
pivot_out = pandas.DataFrame(numpy.zeros(pivot_exp_all_ratio.shape))
for i in numpy.arange(pivot_out.shape[0]):
   for j in numpy.arange(pivot out.shape[1]):
       percent = str(int(numpy.round(pivot_exp_all_ratio.values[i,j]*100,__
→decimals=0)))
       numerator = str(int(pivot_exp_shift.values[i,j]))
       denominator = str(int(pivot_all.values[i,j]))
       pivot_out.loc[i,j] = percent+'%\n('+numerator+'/'+denominator+')'
pivot_out.index = pivot_all.index
pivot_out.columns = pivot_all.columns
rows = pivot_all.index
cols = pivot_all.columns.get_level_values(1)
#ax.axis('tight')
#ax.axis('off')
#the_table = ax.table(cellText=pivot_out.values, colLabels=cols,__
→rowLabels=rows, loc='center')
ax = axes
seaborn.heatmap(pivot_exp_all_ratio, annot=pivot_out.values, fmt='',_
ax.set_xlabel('Derived chromosomal category')
ax.set ylabel('Ancestral chromosomal category')
ax.tick_params(pad=2, length=3, rotation=0, labelsize=font_size)
del b2
```

```
outbase = 'chromosome_heatmap'
fig.savefig(outbase+".pdf", format='pdf', transparent=True)
fig.savefig(outbase+".svg", format='svg', transparent=True)
```

```
excluded spnodes in chromosome analysis: ['10', '11', '13', 'Xenopus_tropicalis', 'Astyanax_mexicanus', 'Danio_rerio', 'Gadus_morhua', '14', '19', 'Oryzias_latipes', 'Oreochromis_niloticus', 'Anolis_carolinensis', 'Gallus_gallus', 'Ornithorhynchus_anatinus', 'Chinchilla_lanigera']
```



```
[20]: # tissue network
min_taus = [0,0.5]
min_max_second_mu_ratios = [0,] # test 0 and 1
min_bss = [0,99]
min_delta_tau = -numpy.inf
nboot = 10000
nsubsample=numpy.inf#174#sum_R
shift_col = 'llou_intersect_is_shift'
pcm_prefixes = ['llou_fpkm_', 'llou_tpm_']
edge_width_weight=0.4
hide_nonsig=True
ancestral_max_unif_tau = None # 0.3
derived_max_unif_tau = None #0.3
bbox_length = 150
margin_size = 15
```

```
fdr threshold = 0.05
calc_fdr = False
coordinates = {
    'brain':[0, -1],
    'heart': [0.8660254037844386467637, -0.5],
    'kidney': [0.8660254037844386467637, 0.5],
    'liver':[0,1],
    'ovary': [-0.8660254037844386467637, 0.5],
    'testis':[-0.8660254037844386467637, -0.5],
}
for pp,min_tau,min_max_second_mu_ratio,min_bs in itertools.

-product(pcm_prefixes[::-1], min_taus, min_max_second_mu_ratios, min_bss):
    corrected = dict()
    observed = dict()
    expected = dict()
    bs = dict()
    Ns = dict()
    fdrs = dict()
    fdr flag = True
    var1 = 'parent '+pp+'max organ'
    var2 = pp+'max_organ'
    specificity_term =__
→pp+'muRatio'+str(min_max_second_mu_ratio)+'_minTau'+str(min_tau)+'_minBS'+str(min_bs)
    print('\n', specificity_term)
    for event in branch categories+['All',]:
        print('event =', event)
        conditions = True
        conditions = conditions&(b[shift col]==1)
        if event is not 'All':
            conditions = conditions&(b.branch_category==event)
        conditions = conditions&(b.spnode_coverage!=sptree_root)
        conditions = (conditions)&(b['parent '+pp+'tau']>=min tau)
        conditions = (conditions)&(b[pp+'tau']>=min_tau)
        conditions = (conditions)&(b[pp+'delta_tau']>=min_delta_tau)
        conditions =
 →(conditions)&(b[pp+'max second mu ratio']>=min max second mu ratio)
 →(conditions)&(b['parent '+pp+'max second mu ratio']>=min max second mu ratio)
        if min_bs!=0:
            conditions = (conditions)&(b['parent_support_iqtree']>=min_bs)
        #conditions = conditions&(b.parent_reconcil_support>=0.9)
        \#conditions = conditions \& (b.max mu>=0.1)
        #conditions = conditions&(b.parent_max_mu>=0.1)
        #conditions = conditions&(b.max_mu/b.second_max_mu>=1.5)
        \#conditions = conditions \& (b.parent_max_mu/b.parent_second_max_mu>=1.5)
```

```
df2 = b.loc[conditions,:]
      print('n before filterings =', df2.shape[0])
      out = shift_freq_bootstrap(df1=df2, var1=var1, var2=var2, u
→down_reg=False,
                               exclude_self=True, nboot=nboot,_
→nsubsample=nsubsample)
      corrected[event], observed[event], expected[event], bs[event], Ns[event] = ___
∽out
      →expected[event])**2).sum(axis=0).sum(axis=0) / ((corrected[event].
→shape[0]*corrected[event].shape[1])-1-corrected[event].shape[1])))
      IPython.display.display(observed[event])
      observed[event].
→to_csv('shift_count_observed_'+specificity_term+'_'+event+'.tsv', sep='\t', 
→index=True)
      expected[event].
→to_csv('shift_count_expected_'+specificity_term+'_'+event+'.tsv', sep='\t',__
→index=True)
      corrected[event].
→to_csv('shift_count_corrected_'+specificity_term+'_'+event+'.tsv', sep='\t',
→index=True)
      observed[event].stack().
→to_csv('stack_shift_count_observed_'+specificity_term+'_'+event+'.tsv', |
→sep='\t', index=True)
      expected[event].stack().
-to_csv('stack_shift_count_expected_'+specificity_term+'_'+event+'.tsv',__
→sep='\t', index=True)
      corrected[event].stack().
→sep='\t', index=True)
      test method = 'random'
      out = draw network(corrected[event], observed[event], expected[event],
→bs[event], self_arrow=False, self_vertex_size=False,
                        show_vertex_stat=False, edge_width=1,__
→hide_nonsig=hide_nonsig,
                       coordinates=coordinates, test=test_method)
      g,layout,visual_style = out
      outbase = wd+'shift_network_'+specificity_term+'_'+test_method+'_'+event
      igraph.plot(g, target=outbase+'.svg', layout=layout, bbox=(bbox_length,_
→bbox_length), background=None, margin=margin_size, **visual_style)
      texts = []
      texts.append(event)
      texts.append('N = '+"{:,d}".format(Ns[event]))
```

```
add_igraph_legends(svg_file=outbase+'.svg', height=bbox_length,__
→width=bbox_length, texts=texts)
       command = inkscape+' --export-pdf='+wd+outbase+'.pdf'+' '+wd+outbase+'.
⇔svg'
       os.system(command)
       #IPython.display.SVG(outbase+'.svg')
       if calc_fdr:
           vertex_dict = {'vertex':organs, 'color':organ_colors}
           quantile = get_quantile(observed[event], bs[event])
           try:
               fdrs[event] = get_quantile_fdr(quantile,__
→fdr_threshold=fdr_threshold)
           except AssertionError:
               print('FDR cannot be calculated. Skipped.')
               fdr flag = False
           except ValueError:
               print('FDR cannot be calculated. Skipped.')
               fdr_flag = False
           else:
               out = draw_network2(observed[event], quantile, fdrs[event],__
→no_fp=True,
                                  self_arrow=False, self_vertex_size=False,__
⇒show_vertex_stat=False, no_obs=False,
                                  edge_width=2, edge_width_weight=5,_u
→hide nonsig=False, vertex_dict=vertex_dict, coordinates=coordinates,
                                  show_edge_color=True,_
→scaled_edge_curve=True, relative_freq_edge_width=False, sig_color='greenpink'
               g,layout,visual_style = out
               outbase = wd+'fdr_shift_network_'+specificity_term+'_'+event
               igraph.plot(g, target=outbase+'.svg', layout=layout,_
→bbox=(bbox_length, bbox_length), background=None, margin=margin_size, __
→**visual style)
               texts = []
               texts.append(event)
               texts.append('N = '+"{:,d}".format(Ns[event]))
               add_igraph_legends(svg_file=outbase+'.svg', height=bbox_length,_
→width=bbox_length, texts=texts)
               command = inkscape+' --export-pdf='+wd+outbase+'.pdf'+'_
→ '+wd+outbase+'.svg'
               os.system(command)
   if fdr_flag&calc_fdr:
       # Unidirectional table
       tmp2 = pandas.DataFrame()
```

```
for event in branch_categories:
            df_pvalue = observed[event].copy()
            df_pvalue.loc[:,:] = (observed[event].values > bs[event]).
⇒sum(axis=0) / bs[event].shape[0]
            for ind,col in itertools.product(df_pvalue.index, df_pvalue.
 →columns):
                if ind==col:
                    df_pvalue.at[ind,col] = numpy.nan
            obs = pandas.DataFrame(observed[event].stack())
            exp = pandas.DataFrame(expected[event].stack())
            pval = pandas.DataFrame(df_pvalue.stack()) * 100
            fdr = pandas.DataFrame(fdrs[event].stack()).iloc[:,0]
            tmp = pandas.concat([obs, exp, pval, fdr], axis=1)
            tmp.columns = ['Observed', 'Expected', 'Permutation-based percentile_
→rank','FDR<0.05']</pre>
            tmp = tmp.reset_index()
            tmp.columns = tmp.columns.str.replace('parent_'+pp+'max_organ',__
 →'Ancestral PEO')
            tmp.columns = tmp.columns.str.replace(pp+'max_organ', 'Derived PEO')
            tmp = tmp.dropna()
            tmp.loc[:,'Branch'] = event
            col order = ['Branch', 'Ancestral PEO', 'Derived___
→PEO', 'Observed', 'Expected', 'Permutation-based percentile rank', 'FDR<0.05']
            tmp = tmp.loc[:,col_order]
            is_sig = tmp.loc[:,'FDR<0.05'].values</pre>
            tmp.loc[is_sig,'FDR<0.05'] = 'yes'</pre>
            tmp.loc[~is_sig,'FDR<0.05'] = 'no'</pre>
            tmp2 = pandas.concat([tmp2,tmp], ignore_index=True, axis=0)
        outfile = 'shift_summary_'+specificity_term+'.tsv'
        tmp2.to_csv(outfile, sep='\t', index=True)
del corrected, expected, observed, bs, Ns
# Chi-square test
pp = 'llou_fpkm_'
mu_ratio = 0
min_tau = 0
min_bs = 0
contingency_list = list()
for bc in branch_categories:
    tsv_file = 'shift_count_observed {}muRatio{} minTau{} minBS{}_{}.tsv'
    tsv_file = tsv_file.format(pp, mu_ratio, min_tau, min_bs, bc)
    tmp = pandas.read_table(tsv_file, index_col=0)
    contingency_list.append(tmp.astype(int).values.flatten())
contingency_table = numpy.array(contingency_list)
is_observed = (contingency_table.sum(axis=0)>0)
contingency_table = contingency_table[:,is_observed]
```

```
out = scipy.stats.chi2_contingency(observed=contingency_table, correction=True,__
 →lambda_=None)
chi,p,dof,expected = out
print('Chi-square test among 3 branch categories: chisq={:,}, P={}, dof={}'.
 →format(chi, p, dof))
l1ou_tpm_muRatio0_minTau0_minBS0
event = S
n before filterings = 11746
N after filtering = 6617
sample standard deviation = 267.3594426707461
llou_tpm_max_organ
                          brain heart kidney liver ovary testis
parent_l1ou_tpm_max_organ
                            0.0 193.0
                                         214.0 139.0 492.0
brain
                                                              513.0
heart
                           91.0
                                 0.0
                                          68.0
                                                39.0 107.0
                                                              121.0
kidney
                          103.0
                                 70.0
                                          0.0 118.0 186.0
                                                              182.0
                                  34.0
                           60.0
                                         110.0
                                                  0.0 101.0
                                                              102.0
liver
                          517.0 166.0
                                         232.0 182.0
                                                        0.0
                                                              706.0
ovary
                          489.0 177.0
                                         249.0 200.0 656.0
                                                                0.0
testis
event = D
n before filterings = 5960
N after filtering = 3495
sample standard deviation = 135.37208846222526
l1ou_tpm_max_organ
                          brain heart kidney liver ovary testis
parent_l1ou_tpm_max_organ
                                          61.0
                            0.0
                                  54.0
                                                 54.0 149.0
                                                              198.0
brain
heart
                           63.0
                                 0.0
                                          23.0
                                                 25.0
                                                       60.0
                                                               89.0
kidnev
                           56.0
                                  52.0
                                          0.0
                                                82.0 130.0
                                                              119.0
liver
                           52.0
                                  32.0 75.0
                                                  0.0 117.0
                                                               96.0
                          215.0
                                  84.0
                                         129.0 121.0
                                                        0.0
                                                              427.0
ovary
                          264.0 103.0
                                         137.0 124.0 304.0
                                                                0.0
testis
event = R
n before filterings = 1106
N after filtering = 713
sample standard deviation = 34.049246167874735
                          brain heart kidney liver ovary testis
l1ou_tpm_max_organ
parent_l1ou_tpm_max_organ
brain
                            0.0
                                  16.0
                                          7.0
                                                 5.0
                                                       28.0
                                                               48.0
                                 0.0
                           15.0
                                          10.0
                                                  6.0
                                                       21.0
                                                               46.0
heart
kidnev
                           14.0
                                  24.0
                                          0.0
                                                 18.0
                                                       13.0
                                                               51.0
                            2.0
                                  1.0
                                           5.0
                                                 0.0
                                                        4.0
                                                               20.0
liver
                           33.0
                                  17.0
                                          30.0
                                                 20.0
                                                        0.0
                                                              146.0
```

ovary

testis 26.0 15.0 15.0 8.0 49.0 0.0

event = All
n before filterings = 19096
N after filtering = 11007

sample standard deviation = 436.0393924880891

l1ou_tpm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_tpm_max_organ						
brain	0.0	267.0	282.0	198.0	680.0	765.0
heart	170.0	0.0	104.0	73.0	192.0	257.0
kidney	173.0	153.0	0.0	219.0	348.0	357.0
liver	120.0	67.0	194.0	0.0	227.0	221.0
ovary	777.0	271.0	398.0	327.0	0.0	1291.0
testis	789.0	305.0	411.0	338.0	1033.0	0.0

l1ou_tpm_muRatio0_minTau0_minBS99

event = S

n before filterings = 5717

N after filtering = 3122

sample standard deviation = 125.71332017109673

l1ou_tpm_max_organ	brain	${\tt heart}$	kidney	liver	ovary	testis
parent_l1ou_tpm_max_organ						
brain	0.0	100.0	87.0	73.0	233.0	234.0
heart	38.0	0.0	23.0	21.0	46.0	51.0
kidney	55.0	37.0	0.0	51.0	76.0	75.0
liver	27.0	15.0	70.0	0.0	47.0	49.0
ovary	246.0	77.0	101.0	79.0	0.0	298.0
testis	273.0	99.0	136.0	95.0	310.0	0.0

event = D

n before filterings = 3061

N after filtering = 1799

sample standard deviation = 70.89962646450866

l1ou_tpm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_tpm_max_organ						
brain	0.0	32.0	34.0	35.0	86.0	101.0
heart	35.0	0.0	11.0	9.0	28.0	42.0
kidney	28.0	27.0	0.0	37.0	52.0	76.0
liver	32.0	20.0	35.0	0.0	58.0	48.0
ovary	119.0	45.0	63.0	62.0	0.0	250.0
testis	134.0	50.0	63.0	47.0	140.0	0.0

event = R

n before filterings = 628
N after filtering = 395

sample standard deviation = 17.742557163867247

l1ou_tpm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_tpm_max_organ						
brain	0.0	12.0	3.0	3.0	17.0	18.0
heart	7.0	0.0	4.0	4.0	12.0	22.0
kidney	6.0	17.0	0.0	10.0	7.0	28.0
liver	1.0	0.0	4.0	0.0	3.0	12.0
ovary	16.0	8.0	19.0	10.0	0.0	82.0
testis	17.0	10.0	11.0	5.0	27.0	0.0

event = All

n before filterings = 9529
N after filtering = 5386

sample standard deviation = 213.2151605464185

l1ou_tpm_max_organ	brain	heart	kidney	liver	ovary	testis
<pre>parent_l1ou_tpm_max_organ</pre>						
brain	0.0	145.0	124.0	111.0	339.0	355.0
heart	80.0	0.0	38.0	34.0	87.0	116.0
kidney	89.0	82.0	0.0	99.0	143.0	181.0
liver	63.0	35.0	111.0	0.0	111.0	111.0
ovary	386.0	133.0	186.0	151.0	0.0	636.0
testis	428.0	162.0	215.0	149.0	486.0	0.0

11ou_tpm_muRatio0_minTau0.5_minBS0

event = S

n before filterings = 5407

N after filtering = 2440

sample standard deviation = 104.02846536341698

l1ou_tpm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_tpm_max_organ						
brain	0.0	105.0	117.0	88.0	228.0	247.0
heart	30.0	0.0	19.0	14.0	36.0	47.0
kidney	40.0	20.0	0.0	51.0	66.0	59.0
liver	30.0	13.0	57.0	0.0	44.0	55.0
ovary	74.0	25.0	31.0	37.0	0.0	162.0
testis	167.0	75.0	125.0	96.0	282.0	0.0

event = D

n before filterings = 2508

N after filtering = 1142

sample standard deviation = 45.90762558051996

l1ou_tpm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_tpm_max_organ						
brain	0.0	22.0	23.0	23.0	43.0	84.0
heart	22.0	0.0	3.0	10.0	19.0	23.0
kidney	10.0	5.0	0.0	18.0	24.0	27.0
liver	20.0	12.0	36.0	0.0	48.0	45.0
ovary	35.0	9.0	27.0	24.0	0.0	109.0
testis	117.0	41.0	68.0	61.0	134.0	0.0

event = R

n before filterings = 290

N after filtering = 147

sample standard deviation = 6.262471039887569

l1ou_tpm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_tpm_max_organ						
brain	0.0	2.0	1.0	3.0	4.0	17.0
heart	5.0	0.0	2.0	2.0	5.0	20.0
kidney	1.0	1.0	0.0	0.0	0.0	4.0
liver	1.0	0.0	4.0	0.0	2.0	5.0
ovary	4.0	3.0	1.0	2.0	0.0	21.0
testis	9.0	6.0	3.0	3.0	16.0	0.0

event = All

n before filterings = 8284

N after filtering = 3762

sample standard deviation = 154.65131013359417

l1ou_tpm_max_organ	brain	heart	kidney	liver	ovary	testis
<pre>parent_l1ou_tpm_max_organ</pre>						
brain	0.0	130.0	141.0	114.0	279.0	349.0
heart	57.0	0.0	24.0	26.0	60.0	90.0
kidney	51.0	26.0	0.0	69.0	91.0	90.0
liver	51.0	25.0	98.0	0.0	95.0	106.0
ovary	113.0	37.0	60.0	63.0	0.0	295.0
testis	297.0	126.0	198.0	163.0	438.0	0.0

11ou_tpm_muRatio0_minTau0.5_minBS99

event = S

n before filterings = 2797

N after filtering = 1217

sample standard deviation = 51.15382501689103

l1ou_tpm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_tpm_max_organ						
brain	0.0	60.0	52.0	44.0	111.0	119.0
heart	12.0	0.0	8.0	8.0	16.0	14.0

kidney	24.0	11.0	0.0	27.0	26.0	27.0
liver	11.0	6.0	37.0	0.0	23.0	26.0
ovary	36.0	12.0	12.0	21.0	0.0	79.0
testis	107.0	43.0	68.0	45.0	132.0	0.0

event = D

n before filterings = 1282

N after filtering = 567

sample standard deviation = 21.755287970514175

l1ou_tpm_max_organ	brain	heart	kidney	liver	ovary	testis
<pre>parent_l1ou_tpm_max_organ</pre>						
brain	0.0	11.0	11.0	16.0	24.0	43.0
heart	15.0	0.0	3.0	6.0	9.0	11.0
kidney	6.0	4.0	0.0	5.0	9.0	17.0
liver	12.0	9.0	16.0	0.0	24.0	24.0
ovary	19.0	4.0	18.0	11.0	0.0	50.0
testis	55.0	18.0	33.0	20.0	64.0	0.0

event = R

n before filterings = 174

N after filtering = 80

sample standard deviation = 3.1484517998403234

llou_tpm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_tpm_max_organ						
brain	0.0	1.0	0.0	2.0	1.0	5.0
heart	1.0	0.0	0.0	1.0	4.0	9.0
kidney	1.0	1.0	0.0	0.0	0.0	1.0
liver	1.0	0.0	3.0	0.0	2.0	3.0
ovary	2.0	2.0	1.0	0.0	0.0	16.0
testis	6.0	4.0	1.0	2.0	10.0	0.0

event = All

n before filterings = 4294

N after filtering = 1880

sample standard deviation = 76.13306751530278

l1ou_tpm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_tpm_max_organ						
brain	0.0	72.0	63.0	62.0	137.0	167.0
heart	28.0	0.0	11.0	15.0	29.0	34.0
kidney	31.0	16.0	0.0	32.0	36.0	45.0
liver	24.0	15.0	57.0	0.0	50.0	54.0
ovary	57.0	18.0	32.0	32.0	0.0	147.0
testis	171.0	65.0	104.0	68.0	208.0	0.0

l1ou_fpkm_muRatio0_minTau0_minBS0

event = S

n before filterings = 11746

N after filtering = 6886

sample standard deviation = 245.19105957603753

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
<pre>parent_l1ou_fpkm_max_organ</pre>						
brain	0.0	377.0	238.0	295.0	308.0	459.0
heart	230.0	0.0	137.0	225.0	185.0	268.0
kidney	133.0	93.0	0.0	186.0	129.0	162.0
liver	190.0	188.0	184.0	0.0	180.0	236.0
ovary	169.0	143.0	106.0	175.0	0.0	305.0
testis	361.0	246.0	239.0	313.0	426.0	0.0

event = D

n before filterings = 5960

N after filtering = 3586

sample standard deviation = 125.45799930283056

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_fpkm_max_organ						
brain	0.0	76.0	71.0	102.0	101.0	154.0
heart	125.0	0.0	72.0	94.0	94.0	165.0
kidney	73.0	62.0	0.0	78.0	89.0	106.0
liver	128.0	133.0	128.0	0.0	155.0	194.0
ovary	104.0	65.0	72.0	126.0	0.0	191.0
testis	196.0	117.0	135.0	173.0	207.0	0.0

event = R

n before filterings = 1106

N after filtering = 746

sample standard deviation = 31.595094351223743

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_fpkm_max_organ						
brain	0.0	14.0	8.0	11.0	12.0	40.0
heart	40.0	0.0	25.0	35.0	37.0	87.0
kidney	9.0	5.0	0.0	9.0	2.0	26.0
liver	36.0	29.0	42.0	0.0	27.0	70.0
ovary	16.0	14.0	11.0	18.0	0.0	35.0
testis	17.0	21.0	10.0	10.0	30.0	0.0

event = All

n before filterings = 19096

N after filtering = 11395

sample standard deviation = 399.2952002397656

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_fpkm_max_organ						
brain	0.0	470.0	320.0	411.0	431.0	656.0
heart	398.0	0.0	235.0	361.0	319.0	523.0
kidney	216.0	167.0	0.0	276.0	229.0	297.0
liver	357.0	359.0	361.0	0.0	370.0	503.0
ovary	295.0	228.0	192.0	326.0	0.0	535.0
testis	588.0	397.0	390.0	504.0	681.0	0.0

l1ou_fpkm_muRatio0_minTau0_minBS99

event = S

n before filterings = 5717

N after filtering = 3307

sample standard deviation = 118.45128065945984

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_fpkm_max_organ						
brain	0.0	195.0	112.0	141.0	152.0	225.0
heart	98.0	0.0	62.0	85.0	77.0	90.0
kidney	65.0	51.0	0.0	102.0	61.0	87.0
liver	96.0	75.0	85.0	0.0	70.0	96.0
ovary	80.0	76.0	53.0	90.0	0.0	147.0
testis	209.0	134.0	128.0	162.0	203.0	0.0

event = D

n before filterings = 3061

N after filtering = 1826

sample standard deviation = 63.53810496821362

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_fpkm_max_organ						
brain	0.0	44.0	36.0	55.0	56.0	86.0
heart	70.0	0.0	36.0	44.0	53.0	87.0
kidney	40.0	32.0	0.0	37.0	44.0	65.0
liver	72.0	57.0	72.0	0.0	80.0	100.0
ovary	52.0	28.0	37.0	56.0	0.0	91.0
testis	101.0	58.0	67.0	74.0	96.0	0.0

event = R

n before filterings = 628

N after filtering = 408

sample standard deviation = 15.80211741497752

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_fpkm_max_organ						
brain	0.0	7.0	2.0	4.0	10.0	19.0
heart	16.0	0.0	12.0	23.0	17.0	39.0

kidney	5.0	4.0	0.0	6.0	1.0	13.0
liver	23.0	16.0	22.0	0.0	14.0	37.0
ovary	8.0	8.0	9.0	10.0	0.0	21.0
testis	14.0	16.0	7.0	7.0	18.0	0.0

event = All

n before filterings = 9529

N after filtering = 5618

sample standard deviation = 196.28866925653753

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_fpkm_max_organ						
brain	0.0	247.0	151.0	200.0	223.0	332.0
heart	185.0	0.0	110.0	153.0	148.0	218.0
kidney	111.0	91.0	0.0	147.0	110.0	166.0
liver	192.0	153.0	183.0	0.0	170.0	236.0
ovary	141.0	115.0	100.0	158.0	0.0	262.0
testis	331.0	210.0	205.0	245.0	325.0	0.0

11ou_fpkm_muRatio0_minTau0.5_minBS0

event = S

n before filterings = 4959

N after filtering = 2299

sample standard deviation = 92.23749573568381

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
<pre>parent_l1ou_fpkm_max_organ</pre>						
brain	0.0	187.0	113.0	126.0	151.0	208.0
heart	48.0	0.0	32.0	54.0	39.0	60.0
kidney	28.0	26.0	0.0	68.0	48.0	47.0
liver	45.0	46.0	82.0	0.0	51.0	79.0
ovary	34.0	19.0	17.0	34.0	0.0	55.0
testis	104.0	87.0	107.0	114.0	190.0	0.0

event = D

n before filterings = 2262

N after filtering = 1058

sample standard deviation = 38.8931588183576

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_fpkm_max_organ						
brain	0.0	34.0	17.0	34.0	38.0	61.0
heart	33.0	0.0	18.0	22.0	21.0	36.0
kidney	17.0	14.0	0.0	25.0	22.0	24.0
liver	36.0	34.0	40.0	0.0	57.0	68.0
ovary	15.0	10.0	16.0	19.0	0.0	35.0
testis	67.0	33.0	57.0	62.0	93.0	0.0

event = R
n before filterings = 264
N after filtering = 149
sample standard deviation = 6.845836834841906

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
<pre>parent_l1ou_fpkm_max_organ</pre>						
brain	0.0	4.0	1.0	0.0	1.0	9.0
heart	10.0	0.0	4.0	9.0	9.0	24.0
kidney	1.0	0.0	0.0	1.0	0.0	2.0
liver	5.0	3.0	6.0	0.0	1.0	23.0
ovary	1.0	5.0	2.0	2.0	0.0	0.0
testis	6.0	5.0	1.0	2.0	12.0	0.0

event = All
n before filterings = 7568
N after filtering = 3544
sample standard deviation = 135.0823187369566

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_fpkm_max_organ						
brain	0.0	225.0	132.0	160.0	192.0	279.0
heart	92.0	0.0	54.0	87.0	69.0	120.0
kidney	46.0	41.0	0.0	94.0	72.0	73.0
liver	86.0	83.0	129.0	0.0	110.0	172.0
ovary	51.0	34.0	35.0	55.0	0.0	90.0
testis	181.0	130.0	167.0	184.0	301.0	0.0

11ou_fpkm_muRatio0_minTau0.5_minBS99
event = S
n before filterings = 2568
N after filtering = 1168

sample standard deviation = 47.04147442583773

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_fpkm_max_organ						
brain	0.0	99.0	53.0	65.0	71.0	112.0
heart	22.0	0.0	13.0	26.0	17.0	20.0
kidney	10.0	18.0	0.0	36.0	22.0	22.0
liver	24.0	22.0	42.0	0.0	22.0	33.0
ovary	13.0	12.0	10.0	17.0	0.0	33.0
testis	66.0	54.0	61.0	61.0	92.0	0.0

event = D
n before filterings = 1179

N after filtering = 530 sample standard deviation = 19.094916912802102

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
<pre>parent_l1ou_fpkm_max_organ</pre>						
brain	0.0	20.0	5.0	17.0	18.0	33.0
heart	19.0	0.0	9.0	9.0	10.0	22.0
kidney	10.0	7.0	0.0	9.0	7.0	13.0
liver	23.0	15.0	21.0	0.0	33.0	35.0
ovary	9.0	5.0	13.0	6.0	0.0	13.0
testis	32.0	19.0	28.0	27.0	43.0	0.0

event = R

n before filterings = 149

N after filtering = 75

sample standard deviation = 3.2516885764346233

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_fpkm_max_organ						
brain	0.0	1.0	0.0	0.0	1.0	4.0
heart	2.0	0.0	2.0	6.0	4.0	13.0
kidney	1.0	0.0	0.0	0.0	0.0	1.0
liver	3.0	2.0	3.0	0.0	1.0	11.0
ovary	1.0	2.0	2.0	0.0	0.0	0.0
testis	4.0	3.0	0.0	2.0	6.0	0.0

event = All

n before filterings = 3939

N after filtering = 1791

sample standard deviation = 67.98354220222437

l1ou_fpkm_max_organ	brain	heart	kidney	liver	ovary	testis
parent_l1ou_fpkm_max_organ						
brain	0.0	120.0	58.0	82.0	90.0	150.0
heart	44.0	0.0	24.0	42.0	31.0	55.0
kidney	21.0	25.0	0.0	45.0	31.0	36.0
liver	50.0	39.0	67.0	0.0	56.0	81.0
ovary	23.0	19.0	25.0	23.0	0.0	46.0
testis	104.0	77.0	91.0	92.0	144.0	0.0

Chi-square test among 3 branch categories: chisq=530.740393856863, P=1.5244714357425737e-77, dof=58

```
[21]: pp = 'l1ou_fpkm_'
mu_ratio = 0
min_tau = 0
min_bs = 0
```

```
# Unidirectional
tmp_list = list()
for bc in branch_categories:
   for stat in ['observed', 'expected']:
       tsv_file = 'shift_count_{}_{minTau{}_minBS{}_{}.tsv'
       tsv_file = tsv_file.format(stat, pp, mu_ratio, min_tau, min_bs, bc)
       tmp = pandas.read_table(tsv_file, index_col=0)
       tmp.index.name = None
       tmp = pandas.DataFrame(tmp.stack())
       tmp.columns = [bc+' '+stat,]
       tmp_list.append(tmp)
tmp2 = pandas.concat(tmp list, axis=1)
for col in tmp2.columns[tmp2.columns.str.endswith('_observed')]:
   tmp2.loc[:,col] = tmp2.loc[:,col].astype(int)
tmp2 = tmp2.reset_index()
tmp2.columns = tmp2.columns.str.replace('level_0', 'ancestral_PEO')
tmp2.columns = tmp2.columns.str.replace('level_1', 'derived_PEO')
for bc in branch_categories:
   tmp2.loc[:,bc+'_pvalue'] = numpy.nan
   tmp2.loc[:,bc+'_chi2'] = numpy.nan
   total_obs = tmp2.loc[:,bc+'_observed'].sum()
   total_exp = tmp2.loc[:,bc+'_expected'].sum()
   assert total_obs==total_exp, 'Total counts should match.'
outfile = 'shift chisq test unidirectional.tsv'
tmp2.to_csv(outfile, sep='\t', index=True)
# Bidirectional
organ_combinations = itertools.combinations(organs, 2)
df_bi = pandas.DataFrame(organ_combinations)
df_bi.columns = ['PE01','PE02']
cols = [ bc+'_'+stat for bc,stat in itertools.product(branch_categories,_
for col in cols:
   df_bi.loc[:,col] = 0
for i in df_bi.index:
   o1 = df_bi.loc[i, 'PEO1']
   o2 = df bi.loc[i,'PEO2']
   is_target1 = ((tmp2.loc[:,'ancestral_PEO']==o1)&(tmp2.loc[:
 →, 'derived_PEO']==o2))
    is_target2 = ((tmp2.loc[:, 'ancestral_PEO']==o2)&(tmp2.loc[:
→, 'derived_PEO']==o1))
   is target = is target1 | is target2
   tmp = tmp2.loc[is_target,cols].sum(axis=0)
   df_bi.loc[i,cols] = tmp.values
```

```
for bc in branch_categories:
          total_obs = df_bi.loc[:,bc+'_observed'].sum()
         total_exp = df_bi.loc[:,bc+'_expected'].sum()
         for i in df_bi.index:
             obs = df_bi.loc[i,bc+'_observed']
              exp = df_bi.loc[i,bc+'_expected']
     outfile = 'shift_chisq_test_bidirectional.tsv'
     df_bi.to_csv(outfile, sep='\t', index=True)
     df bi.head()
[21]:
         PE01
                 PEO2 S_observed S_expected D_observed D_expected R_observed \
     0 brain
                heart
                            607.0 505.507195
                                                    201.0 192.884207
                                                                             54.0
     1 brain kidney
                            371.0 398.683574
                                                    144.0 167.185266
                                                                             17.0
     2 brain liver
                            485.0 535.961005
                                                    230.0 252.900053
                                                                             47.0
     3 brain ovary
                            477.0 530.775255
                                                    205.0 227.337199
                                                                             28.0
     4 brain testis
                            820.0 720.330343
                                                    350.0 312.114196
                                                                             57.0
        R_expected
         52.791154
     0
     1 22.351151
     2 49.070692
     3 31.958141
         50.941958
[22]: for pp,min_tau,min_max_second_mu_ratio,min_bs_in_itertools.

product(pcm_prefixes, min_taus, min_max_second_mu_ratios, min_bss):
          specificity term =
       →pp+'muRatio'+str(min_max_second_mu_ratio)+'_minTau'+str(min_tau)+'_minBS'+str(min_bs)
         print(specificity_term)
          edat = dict()
         erank = dict()
         for event in ['S','D','R']:
              infile = 'stack_shift_count_observed_'+specificity_term+'_'+event+'.tsv'
              tmp = pandas.read csv(infile, sep='\t', header=None, index col=[0,1])
              edat[event] = tmp.astype(int).values.flatten()
         is nonzero = (edat['S']+edat['D']+edat['R']!=0)
         for event in ['S','D','R']:
              edat[event] = edat[event][is_nonzero]
              #erank[event] = edat[event].argsort()
         for event1, event2 in itertools.combinations(['S','D','R'], 2):
              edat_ct = numpy.array([edat[event1],edat[event2]])
              #erank_ct = numpy.array([erank[event1], erank[event2]])
             try:
                  chisq_dat = scipy.stats.chi2 contingency(edat_ct, correction=True)
```

```
#chisq_rank = scipy.stats.chi2_contingency(erank_ct,_
 \rightarrow correction=True)
            print(event1, event2, 'raw count. P-value =', chisq_dat[1], 'chisq_
 →stat =', chisq dat[0])
             #print(event1, event2, 'rank. P-value =', chisq_rank[1], 'chisq_
 \rightarrow stat =', chisq_rank[0])
        except:
            print(event1, event2, 'raw count. Error in chisq test')
    #a.1.1.
    edat_ct = numpy.array([edat['S'],edat['D'],edat['R']])
    #erank_ct = numpy.array([erank['S'],erank['D'],erank['R']])
    chisq_dat = scipy.stats.chi2_contingency(edat_ct, correction=True)
    #chisq_rank = scipy.stats.chi2 contingency(erank_ct, correction=True)
    print('ALL raw count. P-value =', chisq_dat[1], 'chisq stat =',_
 \rightarrowchisq_dat[0])
    #print('ALL rank. P-value =', chisq_rank[1], 'chisq stat =', chisq_rank[0])
    print()
11ou_fpkm_muRatio0_minTau0_minBS0
S D raw count. P-value = 1.1645425725499324e-33 chisq stat = 232.6015555526961
S R raw count. P-value = 3.15741388058073e-54 chisq stat = 337.26871370739775
D R raw count. P-value = 7.322841049325359e-27 chisq stat = 196.83199619773367
ALL raw count. P-value = 1.5244714357425737e-77 chisq stat = 530.740393856863
l1ou fpkm muRatio0 minTau0 minBS99
S D raw count. P-value = 1.1656647990706282e-19 chisq stat = 157.77142932826834
S R raw count. P-value = 4.92143783390324e-28 chisq stat = 203.0640970783855
D R raw count. P-value = 3.0619835981501303e-06 chisq stat = 77.09101926176949
ALL raw count. P-value = 4.6986592404860774e-36 chisq stat = 309.68191950099987
llou_fpkm_muRatio0_minTau0.5_minBS0
S D raw count. P-value = 8.887362232292662e-18 chisq stat = 147.27294260834327
S R raw count. P-value = 2.941578886654832e-31 chisq stat = 220.05562051022451
D R raw count. P-value = 3.816019765323653e-14 chisq stat = 126.51240092175345
ALL raw count. P-value = 5.811232248715891e-43 chisq stat = 347.9745963183017
11ou_fpkm_muRatio0_minTau0.5_minBS99
S D raw count. P-value = 5.566817868652583e-12 chisq stat = 113.72968199442163
S R raw count. P-value = 2.377658640736338e-17 chisq stat = 144.86730756686904
D R raw count. P-value = 0.0004896507293822123 chisq stat = 60.807030670771894
ALL raw count. P-value = 4.223659765548238e-22 chisq stat = 228.57604722134585
llou_tpm_muRatioO_minTauO_minBSO
S D raw count. P-value = 1.2803596852689708e-29 chisq stat = 211.44194549979858
SR raw count. P-value = 3.2765565191475903e-44 chisq stat = 286.7964343857895
D R raw count. P-value = 1.189506679469606e-24 chisq stat = 184.99699829145175
```

```
11ou_tpm_muRatio0_minTau0_minBS99
     S D raw count. P-value = 1.0523789030835427e-14 chisq stat = 129.760188956094
     S R raw count. P-value = 2.2029154641494666e-24 chisq stat = 183.55598383597146
     D R raw count. P-value = 5.481109524503753e-08 chisq stat = 88.8422765150004
     ALL raw count. P-value = 9.238148961661861e-31 chisq stat = 279.6357445931875
     11ou_tpm_muRatio0_minTau0.5_minBS0
     S D raw count. P-value = 1.3961237112388258e-18 chisq stat = 151.7750466106425
     S R raw count. P-value = 3.0760106664249445e-14 chisq stat = 127.05739423406402
     D R raw count. P-value = 1.6272885044004317e-08 chisq stat = 92.26093633030023
     ALL raw count. P-value = 8.693352683566968e-31 chisq stat = 279.7874088371217
     11ou_tpm_muRatioO_minTau0.5_minBS99
     S D raw count. P-value = 1.3431121459082411e-07 chisq stat = 86.28509322451443
     S R raw count. P-value = 8.085657493226364e-09 chisq stat = 94.20781351567337
     D R raw count. P-value = 0.006416527451683615 chisq stat = 51.36161390751743
     ALL raw count. P-value = 2.3979096921829816e-13 chisq stat = 172.81407742787528
[23]: reverse=True
     for pp,min_tau,min_max_second_mu_ratio,min_bs in itertools.
       →product(pcm_prefixes, min_taus, min_max_second_mu_ratios, min_bss):
          specificity_term =__
      →pp+'muRatio'+str(min_max_second_mu_ratio)+'_minTau'+str(min_tau)+'_minBS'+str(min_bs)
         print(specificity term)
         observed = dict()
         for event in ['S','D','R']:
              infile = 'shift_count_observed_'+specificity_term+'_'+event+'.tsv'
              observed[event] = pandas.read_csv(infile, sep='\t', header=0,__
       \rightarrowindex_col=0)
         num\_data = len(observed['S'].index) * len(observed['S'].columns) *_{\sqcup}
       →len(observed.keys())
         df1 = pandas.DataFrame(index=numpy.arange(num_data),__
       for k in observed.keys():
             for state in ['ancestral', 'derived']:
                 if state=='ancestral':
                     axis=1
                 elif state=='derived':
                     axis=0
                 values = observed[k].sum(axis=axis)
                 values = values / values.sum()
                 next_i = i+len(values)
```

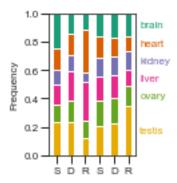
ALL raw count. P-value = 1.1858636662178189e-66 chisq stat = 474.31996529441284

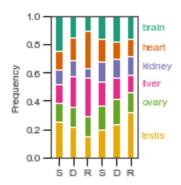
```
#tmp = pandas.DataFrame({'organ':values.index.tolist(),__
→ 'shift_value':values.tolist()})
           \#tmp['event'] = k
           #tmp['stete'] = state
           df1.loc[i:(next_i-1),'event'] = k
           df1.loc[i:(next i-1), 'state'] = state
           df1.loc[i:(next_i-1), 'organ'] = values.index.tolist()
           df1.loc[i:(next_i-1), 'shift_value'] = values.tolist()
           df1 = pandas.concat([df1,tmp], ignore_index=True, sort=False)
           i = next_i
   \#df1.loc[:, 'organ'] = df1['organ'].str.replace('unif. \n < 0.3', 'unif.')
   #colors = {'brain': '#1B9E77', 'heart': '#D95F02', 'kidney': '#7570B3', 'liver':
→ '#E7298A', 'ovary': '#66A61E', 'testis': '#E6AB02'}
   #colors =
→ ['#1B9E77', '#D95F02', '#7570B3', '#E7298A', '#66A61E', '#E6AB02', 'darkgray']
   colors = ['#1B9E77','#D95F02','#7570B3','#E7298A','#66A61E','#E6AB02',]
   fig,axes = matplotlib.pyplot.subplots(nrows=1, ncols=1, figsize=(2,2),__
→sharex=False)
   ax=axes
   df1 = df1.pivot_table(index=['state','event'], columns='organ',_
→values='shift_value', aggfunc='first').fillna(0)
   df1 = df1.loc[[ (11,12) for 11 in ['ancestral', 'derived'] for 12 in_
→branch_categories ],:]
   if reverse:
       df1 = df1.iloc[:.::-1]
       colors.reverse()
   df1.plot.bar(stacked=True, ax=ax, color=colors, legend=False,
→fontsize=font_size)
   ax.set_xlabel('', fontsize=font_size)
   ax.set_ylabel('Frequency', fontsize=font_size)
   ax.set xticks(numpy.arange(len(branch categories)*2), minor=False)
   ax.set_xticks([(len(branch_categories)-1)/2,__
→len(branch_categories)+((len(branch_categories)-1)/2)], minor=True)
   ax.set_xticklabels(branch_categories*2, minor=False, ha='center',_
→rotation=0, fontsize=font_size)
   ax.set_xticklabels(['\nAncestral','\nDerived'], minor=True, ha='center',_
→rotation=0, fontsize=font_size)
   ax.tick_params(axis='x', which='major', direction='out', length=6, width=1)
   ax.tick_params(axis='x', which='minor', direction='out', length=6, width=0)
   ax.tick_params(axis='both', which='major', direction='out', length=6,__
→width=1, pad=2, top=False, right=False)
   ax.set ylim(0, 1)
   handles, labels = ax.get_legend_handles_labels()
```

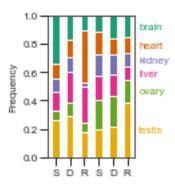
```
\#ax.legend(handles[::-1], labels[::-1], bbox to_anchor=(1,1), loc=2,
→ fontsize=font size)
  xmax = 0
  for p in ax.patches:
      xmax = max(xmax, p.get x())
  rightmost_y_coordinates = list()
  for p in ax.patches:
       if p.get_x()==xmax:
           rightmost_y_coordinates.append(p.get_y())
  rightmost_y_coordinates.append(1)
  rightmost_y_coordinates = numpy.array(rightmost_y_coordinates)[::-1]
  rightmost_y_coordinates = (rightmost_y_coordinates[1:] +__
→rightmost_y_coordinates[:-1]) / 2
   colors.reverse()
  for tis,c,y in zip(labels[::-1], colors, rightmost_y_coordinates):
       ax.text(x=(len(branch_categories)*2)-0.2, y=y, s=tis,__

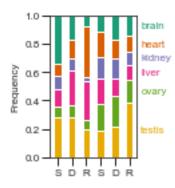
→fontsize=font_size, color=c, va='center', ha='left')
  fig.tight_layout()
  outbase = 'shift_freq_ancestral_derived_'+specificity_term
  fig.savefig(outbase+".pdf", format='pdf', transparent=True)
  fig.savefig(outbase+".svg", format='svg', transparent=True)
```

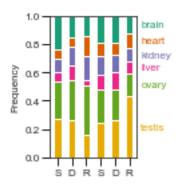
```
11ou_fpkm_muRatioO_minTauO_minBSO
11ou_fpkm_muRatioO_minTauO_minBS99
11ou_fpkm_muRatioO_minTauO.5_minBS0
11ou_fpkm_muRatioO_minTauO.5_minBS99
11ou_tpm_muRatioO_minTauO_minBS0
11ou_tpm_muRatioO_minTauO_minBS99
11ou_tpm_muRatioO_minTauO.5_minBS0
11ou_tpm_muRatioO_minTauO.5_minBS99
```

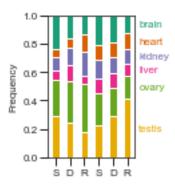


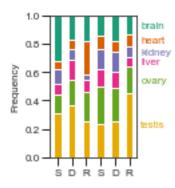


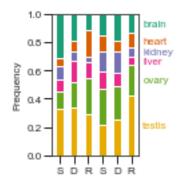










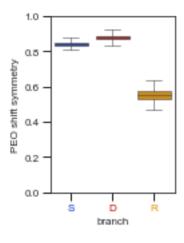


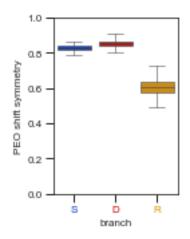
```
[24]: |ylab = 'PEO shift symmetry'
      nrep = 1000
      orders = branch_categories
      def get_bootstrap(df):
          num_category = numpy.prod(df.shape)
          fill_series = pandas.Series([0,]*num_category)
          shape = df.shape
          x = df.values.flatten()
          size = x.sum().astype(int)
          ind = numpy.arange(0,x.shape[0])
          p = x/size
          rand = numpy.random.choice(a=ind, size=size, p=p)
          count = pandas.Series(rand).value_counts().add(fill_series).fillna(0).values
          table = numpy.reshape(a=count, newshape=shape)
          return table
      for pp,min_tau,min_max_second_mu_ratio,min_bs in itertools.
       →product(pcm_prefixes, min_taus, min_max_second_mu_ratios, min_bss):
          specificity_term =_
       →pp+'muRatio'+str(min_max_second_mu_ratio)+'_minTau'+str(min_tau)+'_minBS'+str(min_bs)
          print(specificity_term)
          observed = dict()
          for event in ['S','D','R']:
              infile = 'shift_count_observed_'+specificity_term+'_'+event+'.tsv'
              observed[event] = pandas.read_csv(infile, sep='\t', header=0,_
       →index col=0)
          fig,axes = matplotlib.pyplot.subplots(nrows=1, ncols=1, figsize=(1.7,2.2),__
       →sharex=False)
          ax = axes
          #axes = axes.flat
          dat = observed
```

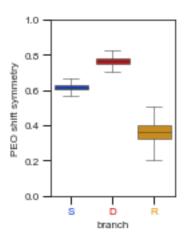
```
resampled_symmetry = dict()
  for event in branch_categories:
      resampled symmetry[event] = pandas.DataFrame(index=numpy.arange(nrep),_
resampled symmetry[event]['event'] = event
      for i in numpy.arange(nrep):
          bs_table = get_bootstrap(df=observed[event])
          resampled_symmetry[event].loc[i,'value'] = __
df_bp = pandas.concat(resampled_symmetry)
  df bp = df bp.reset index()
  df bp['x'] = 0
  x = 0
  for ev in branch_categories:
      df_bp.loc[(df_bp.event==ev),'x'] = x
      x += 1
  df_bp['value'] = df_bp['value'].astype(float)
   seaborn.boxplot(x='event', y='value', data=df_bp, fliersize=0,__
→palette=category_colors.values(),
                  order=category_colors.keys(), linewidth=0.5)
  ax.set_xlim(-0.5, len(branch_categories)-0.5)
  ax.set ylim(0, 1)
  ax.set_xlabel('branch')
  ax.set_ylabel(ylab, fontsize=font_size)
  ax.tick_params(axis='both', which='major', direction='out', length=6,__
→width=1, pad=2, top=False, right=False, labelsize=font_size)
   ax.set_xticks(numpy.arange(len(branch_categories)))
  ax.set_xticklabels(labels=branch_categories, fontsize=font_size)
   [t.set_color(category_colors[t.get_text()]) for t in ax.
→get_xticklabels(minor=False) ]
  for bc1,bc2 in itertools.combinations(branch categories, 2):
      v1 = df_bp.loc[(df_bp['event']==bc1), 'value'].values
      v2 = df_bp.loc[(df_bp['event']==bc2), 'value'].values
      statistic,pvalue = scipy.stats.ks_2samp(v1, v2,__
→alternative='two-sided', mode='auto')
      print('{}; {}-{}: D = {:.2}, P = {:.2}'.format(x, bc1, bc2, statistic,_u
→pvalue))
  fig.tight_layout(pad=0)
   outbase = 'shift_symmetry2_'+specificity_term
  fig.savefig(outbase+".pdf", format='pdf', transparent=True)
  fig.savefig(outbase+".svg", format='svg', transparent=True)
```

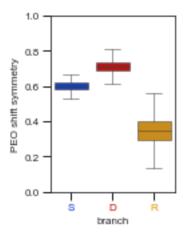
```
11ou_fpkm_muRatio0_minTau0_minBS0
3; S-D: D = 0.82, P = 0.0
```

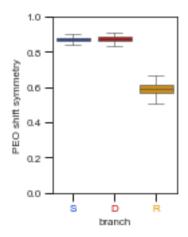
- 3; S-R: D = 1.0, P = 0.0
- 3; D-R: D = 1.0, P = 0.0
- llou_fpkm_muRatio0_minTau0_minBS99
- 3; S-D: D = 0.58, P = 2.7e-158
- 3; S-R: D = 1.0, P = 0.0
- 3; D-R: D = 1.0, P = 0.0
- 11ou_fpkm_muRatio0_minTau0.5_minBS0
- 3; S-D: D = 1.0, P = 0.0
- 3; S-R: D = 1.0, P = 0.0
- 3; D-R: D = 1.0, P = 0.0
- llou_fpkm_muRatio0_minTau0.5_minBS99
- 3; S-D: D = 0.94, P = 0.0
- 3; S-R: D = 0.99, P = 0.0
- 3; D-R: D = 1.0, P = 0.0
- 11ou_tpm_muRatio0_minTau0_minBS0
- 3; S-D: D = 0.11, P = 2.6e-05
- 3; S-R: D = 1.0, P = 0.0
- 3; D-R: D = 1.0, P = 0.0
- llou_tpm_muRatio0_minTau0_minBS99
- 3; S-D: D = 0.24, P = 3e-25
- 3; S-R: D = 1.0, P = 0.0
- 3; D-R: D = 1.0, P = 0.0
- l1ou_tpm_muRatio0_minTau0.5_minBS0
- 3; S-D: D = 0.99, P = 0.0
- 3; S-R: D = 0.65, P = 6.6e-199
- 3; D-R: D = 0.95, P = 0.0
- 11ou_tpm_muRatio0_minTau0.5_minBS99
- 3; S-D: D = 0.9, P = 0.0
- 3; S-R: D = 0.74, P = 2.3e-268
- 3; D-R: D = 0.93, P = 0.0

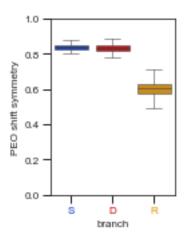


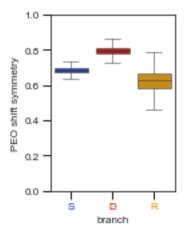


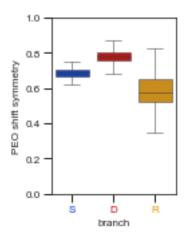












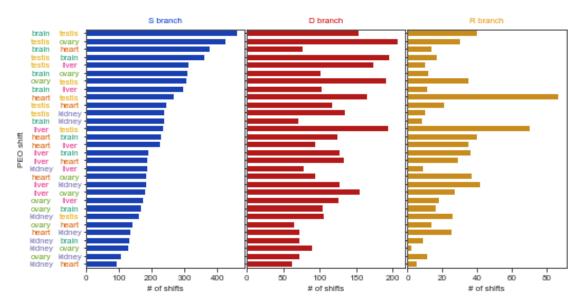
```
[25]: dfs = dict()
      for min_tau,pp,min_bs in itertools.product(min_taus,pcm_prefixes,min_bss):
          specificity_term =
       →pp+'muRatio'+str(min_max_second_mu_ratio)+'_minTau'+str(min_tau)+'_minBS'+str(min_bs)
          print(specificity term)
          for event in ['All','S','D','R']:
              tmp = pandas.
       →read_csv('stack_shift_count_observed_'+specificity_term+'_'+event+'.tsv',_
       \Rightarrowsep='\t', header=0, index_col=[0,1])
              dfs[specificity_term+'_'+event] = tmp['0']
          fig,axes = matplotlib.pyplot.subplots(nrows=1, ncols=3, figsize=(7.2,3.6),__
       →sharex=False)
          axes = axes.flat
          df plot = dict()
          for i,event in enumerate(['S','D','R']):
              tmp = dfs[specificity_term+'_'+event]
              tmp = tmp.reset_index()
              tmp.columns = ['from','to','count']
              tmp['y'] = tmp['from'] + '\rightarrow' + tmp['to']
              tmp = tmp.loc[(tmp['from']!=tmp['to']),:]
              if i==0:
                  tmp = tmp.sort_values(by='count', ascending=False)
                  sorted_y = tmp.loc[:,'y']
                  sorted_y = pandas.DataFrame({'y':sorted_y})
              else:
                  tmp = pandas.merge(sorted_y, tmp, sort=False)
              df_plot[event] = tmp
          for i, event in enumerate(['S','D','R']):
              ax = axes[i]
              color = category_colors[event]
```

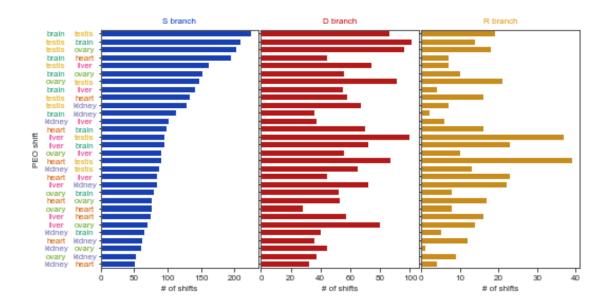
```
seaborn.barplot(x='count', y='y', data=df_plot[event], orient='h',__
 ax.set_xlabel('# of shifts')
        if event=='S':
            ax.set_ylabel('PEO shift')
            yticks = numpy.arange(0, df plot[event].shape[0])
            ax.set yticks(yticks, minor=False)
            ax.set_yticks(yticks+1e-3, minor=True)
            ax.set_yticklabels(df_plot[event]['from'], minor=False, ha='center')
            ax.set_yticklabels(df_plot[event]['to'], minor=True, ha='center')
            ax.tick_params(axis='y', which='major', direction='out', length=2,__
 \rightarrowwidth=1, pad=40)
            ax.tick_params(axis='y', which='minor', direction='out', length=0,__
 \rightarrowwidth=1, pad=15)
             [t.set_color(organ_colors_dict[t.get_text()]) for t in ax.
 →get_yticklabels(minor=False) ]
             [t.set_color(organ_colors_dict[t.get_text()]) for t in ax.
 →get_yticklabels(minor=True) ]
        else:
            ax.set_ylabel('')
            ax.tick_params(axis='y', which='major', direction='out', length=2,_
 \rightarrowwidth=1, pad=0)
             ax.set_yticklabels([''] * len(ax.get_yticklabels()))
        ax.set_title(event+' branch', fontsize=font_size, color=color)
    fig.tight_layout(pad=0)
    outbase = 'PEO_shift_hist_'+specificity_term
    fig.savefig(outbase+".pdf", format='pdf', transparent=True)
    fig.savefig(outbase+".svg", format='svg', transparent=True)
    for pair in itertools.combinations(['S','D','R'], 2):
        pout = scipy.stats.pearsonr(df_plot[pair[0]]['count'],__

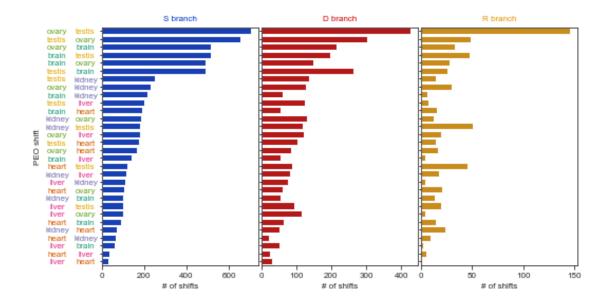
→df_plot[pair[1]]['count'])
        sout = scipy.stats.spearmanr(df_plot[pair[0]]['count'],__

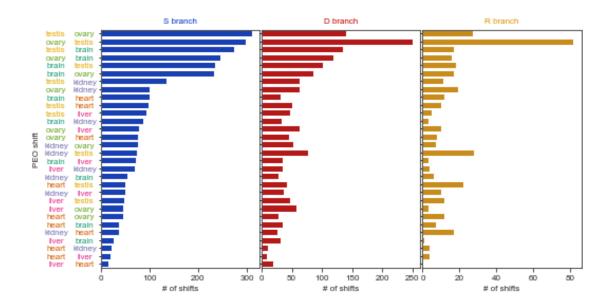
df_plot[pair[1]]['count'])
        print('{}-{}: pearsonr = {}, spearmanr = {}'.format(pair[0], pair[1],__
 →pout[0], sout.correlation))
11ou_fpkm_muRatio0_minTau0_minBS0
S-D: pearsonr = 0.601615621885016, spearmanr = 0.5981974295684188
S-R: pearsonr = 0.20616978664149052, spearmanr = 0.2769368382524662
D-R: pearsonr = 0.5270126462152529, spearmanr = 0.5649705247837534
llou_fpkm_muRatio0_minTau0_minBS99
S-D: pearsonr = 0.5309322972593387, spearmanr = 0.5482794034168859
S-R: pearsonr = 0.05154543361868519, spearmanr = 0.16124369271160363
D-R: pearsonr = 0.6556653003532937, spearmanr = 0.6545881271370931
11ou_tpm_muRatio0_minTau0_minBS0
```

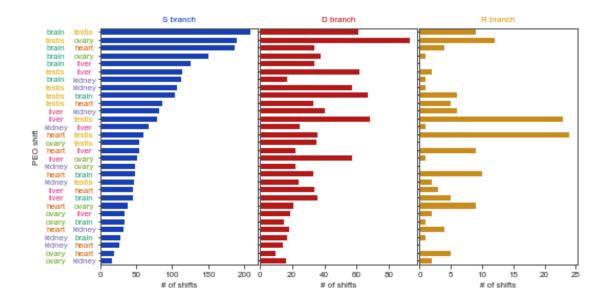
```
S-D: pearsonr = 0.9219567897634421, spearmanr = 0.8541226268521085
S-R: pearsonr = 0.7038241612457165, spearmanr = 0.6221753162382043
D-R: pearsonr = 0.8076823113985413, spearmanr = 0.6353819378663778
llou_tpm_muRatio0_minTau0_minBS99
S-D: pearsonr = 0.8809955210373743, spearmanr = 0.8283234981036648
S-R: pearsonr = 0.6230748608544636, spearmanr = 0.5867543583327695
D-R: pearsonr = 0.8597693023670354, spearmanr = 0.6422681347070208
llou_fpkm_muRatio0_minTau0.5_minBS0
S-D: pearsonr = 0.651728380289062, spearmanr = 0.7372760899569756
S-R: pearsonr = 0.12666101012379385, spearmanr = 0.14087349265134982
D-R: pearsonr = 0.3890561843953191, spearmanr = 0.34661220012149385
11ou_fpkm_muRatio0_minTau0.5_minBS99
S-D: pearsonr = 0.5951344734684358, spearmanr = 0.5794769914989731
S-R: pearsonr = 0.026809299816154538, spearmanr = 0.07749191500501597
D-R: pearsonr = 0.46103806104175044, spearmanr = 0.5117623117247345
11ou_tpm_muRatio0_minTau0.5_minBS0
S-D: pearsonr = 0.8155750539105285, spearmanr = 0.7975728956423863
S-R: pearsonr = 0.5843572432840924, spearmanr = 0.49480153486299155
D-R: pearsonr = 0.6719878172691942, spearmanr = 0.5931253949854308
llou tpm muRatio0 minTau0.5 minBS99
S-D: pearsonr = 0.8420239814752041, spearmanr = 0.7209830229639094
S-R: pearsonr = 0.48568618847029565, spearmanr = 0.4311789084897865
D-R: pearsonr = 0.695267938002528, spearmanr = 0.5991733757920211
```

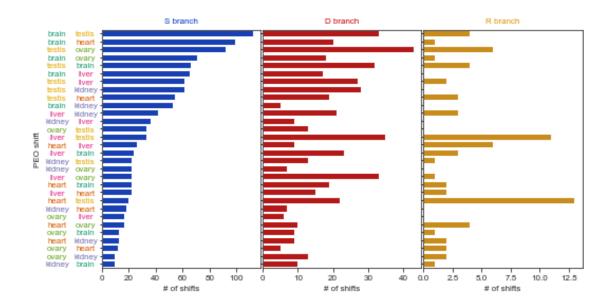


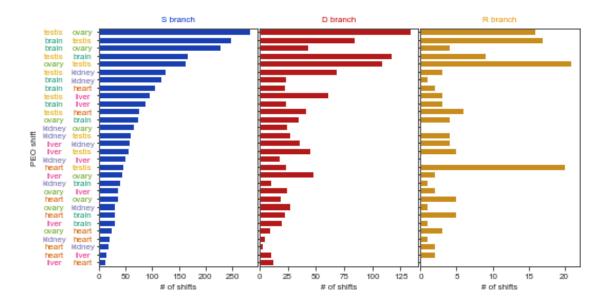


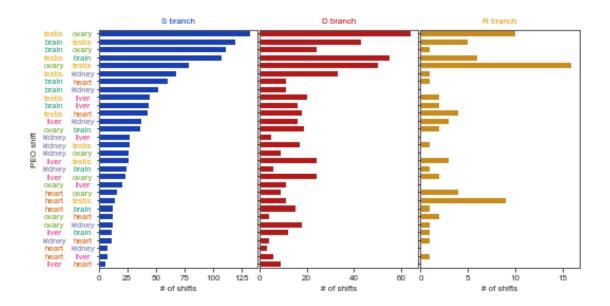












```
[26]: thresholds = [0.1,1,3,5]
      # read transcriptomes
      dir_tissue_mean = '/Users/kef74yk/Dropbox_p/db/Ensembl/release-91/
      ⇔curated_transcriptome/2018_5_1/tpm/tissue_mean'
      tcs = dict()
      for sp in spp:
          file = os.path.join(dir_tissue_mean, sp.replace(' ','_')+'.tissue.mean.tsv')
          tcs[sp] = pandas.read_csv(file, sep='\t', header=0, index_col=0)
      # calc expression ratio
      expressed_ratio = pandas.DataFrame()
      for sp,threshold in itertools.product(spp, thresholds):
          num_gene = tcs[sp].shape[0]
          tmp = (tcs[sp]>threshold).sum()/num_gene
          tmp['species'] = sp
          tmp['threshold'] = threshold
          tmp['num_gene'] = num_gene
          tmp = tmp.to_frame().T
          expressed_ratio = pandas.concat([expressed_ratio,tmp], ignore_index=True)
      # plot
      dfs = dict()
      for min_tau,pp,min_bs,threshold in itertools.
       →product(min_taus,['llou_fpkm_'],min_bss,thresholds):
          specificity_term =
       →pp+'muRatio'+str(min_max_second_mu_ratio)+'_minTau'+str(min_tau)+'_minBS'+str(min_bs)
          print(specificity term)
```

```
for event in ['All','S','D','R']:
       tmp = pandas.
→read_csv('stack_shift_count_observed_'+specificity_term+'_'+event+'.tsv', □
\Rightarrowsep='\t', header=0, index_col=[0,1])
       dfs[specificity_term+'_'+event] = tmp['0']
   fig,axes = matplotlib.pyplot.subplots(nrows=1, ncols=3, figsize=(7.2,3.6),
⇒sharex=False)
   axes = axes.flat
   df_plot = dict()
   for i,event in enumerate(['S','D','R']):
       tmp = dfs[specificity_term+'_'+event]
       tmp = tmp.reset index()
       tmp.columns = ['from', 'to', 'count']
       tmp['y'] = tmp['from'] + '\rightarrow' + tmp['to']
       tmp = tmp.loc[(tmp['from']!=tmp['to']),:]
       # Normalization by expressed gene ratio
       er = expressed ratio.
→loc[(expressed_ratio['threshold']==threshold), organs].median()
       er = er/er.mean()
       tmp.loc[:,'count_normalized'] = tmp.loc[:,'count']
       tmp.loc[:,'count_normalized'] /= er[tmp['from']].values
       tmp.loc[:,'count_normalized'] /= er[tmp['to']].values
       if i==0:
           tmp = tmp.sort_values(by='count_normalized', ascending=False)
           sorted y = tmp.loc[:,'y']
           sorted_y = pandas.DataFrame({'y':sorted_y})
       else:
           tmp = pandas.merge(sorted_y, tmp, sort=False)
       df plot[event] = tmp
   for i, event in enumerate(['S','D','R']):
       ax = axes[i]
       color = category_colors[event]
       seaborn.barplot(x='count_normalized', y='y', data=df_plot[event],_

→orient='h', color=color, ax=ax)
       #ax.set xlim(0,xymax)
       #ax.set_ylim(0,xymax)
       ax.set_xlabel('# of normalized shifts')
       if event=='S':
           ax.set_ylabel('PEO shift')
           yticks = numpy.arange(0, df_plot[event].shape[0])
           ax.set_yticks(yticks, minor=False)
           ax.set_yticks(yticks+1e-3, minor=True)
           ax.set_yticklabels(df_plot[event]['from'], minor=False, ha='center')
           ax.set_yticklabels(df_plot[event]['to'], minor=True, ha='center')
           ax.tick_params(axis='y', which='major', direction='out', length=2,_
\rightarrowwidth=1, pad=40)
```

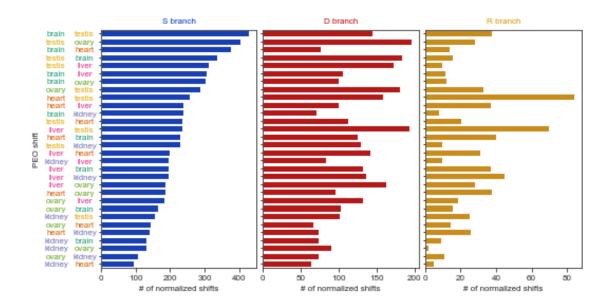
```
ax.tick_params(axis='y', which='minor', direction='out', length=0,_
 \rightarrowwidth=1, pad=15)
             [t.set_color(organ_colors_dict[t.get_text()]) for t in ax.
 →get yticklabels(minor=False) ]
             [t.set_color(organ_colors_dict[t.get_text()]) for t in ax.
 →get_yticklabels(minor=True) ]
        else:
            ax.set ylabel('')
            ax.tick_params(axis='y', which='major', direction='out', length=2, __
 \rightarrowwidth=1, pad=0)
            ax.set_yticklabels([''] * len(ax.get_yticklabels()))
        ax.set_title(event+' branch', fontsize=font_size, color=color)
    fig.tight_layout(pad=0)
    outbase =
 → 'PEO_shift_numExpressedGeneNormalized_'+specificity_term+'_expThreshold'+str(threshold)
    fig.savefig(outbase+".pdf", format='pdf', transparent=True)
    fig.savefig(outbase+".svg", format='svg', transparent=True)
    for pair in itertools.combinations(['S','D','R'], 2):
        pout = scipy.stats.pearsonr(df_plot[pair[0]]['count_normalized'],__

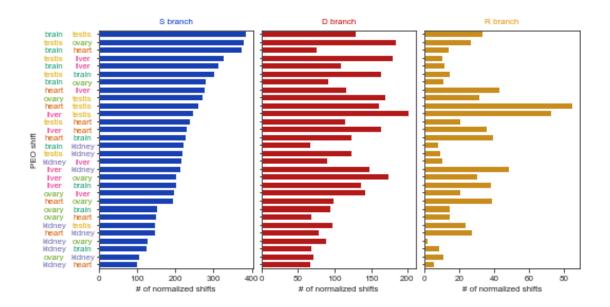
→df_plot[pair[1]]['count_normalized'])
        sout = scipy.stats.spearmanr(df_plot[pair[0]]['count_normalized'],_

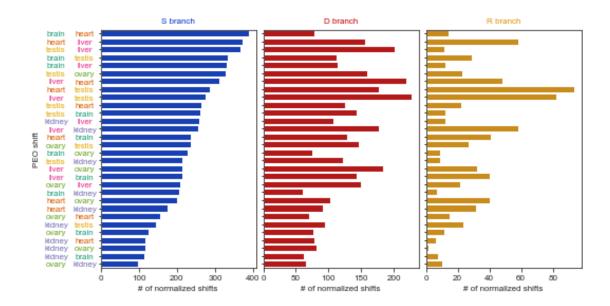
→df_plot[pair[1]]['count_normalized'])
        print('{}-{}: pearsonr = {}, spearmanr = {}'.format(pair[0], pair[1], __
 →pout[0], sout.correlation))
11ou_fpkm_muRatio0_minTau0_minBS0
S-D: pearsonr = 0.5410168526762561, spearmanr = 0.5715239154616241
S-R: pearsonr = 0.1753212146871715, spearmanr = 0.27252502780867627
D-R: pearsonr = 0.5286843487244848, spearmanr = 0.5813125695216907
l1ou fpkm muRatio0 minTau0 minBS0
S-D: pearsonr = 0.5022875522939251, spearmanr = 0.5483870967741935
S-R: pearsonr = 0.20135883538135968, spearmanr = 0.24404894327030033
D-R: pearsonr = 0.5729216714132666, spearmanr = 0.585761957730812
11ou_fpkm_muRatio0_minTau0_minBS0
S-D: pearsonr = 0.5656209274037655, spearmanr = 0.6080088987764182
S-R: pearsonr = 0.35329095449106834, spearmanr = 0.42869855394883205
D-R: pearsonr = 0.670835879217093, spearmanr = 0.6809788654060066
11ou_fpkm_muRatio0_minTau0_minBS0
S-D: pearsonr = 0.6614350376280237, spearmanr = 0.7139043381535038
S-R: pearsonr = 0.5010578221385856, spearmanr = 0.5546162402669632
D-R: pearsonr = 0.743101018784715, spearmanr = 0.7228031145717464
11ou_fpkm_muRatio0_minTau0_minBS99
S-D: pearsonr = 0.4532188119619963, spearmanr = 0.48209121245828696
S-R: pearsonr = -0.0016112755556108027, spearmanr = 0.14438264738598441
```

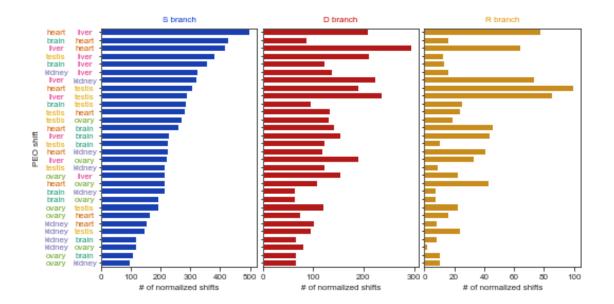
```
D-R: pearsonr = 0.6537649009676451, spearmanr = 0.6542825361512792
11ou_fpkm_muRatio0_minTau0_minBS99
S-D: pearsonr = 0.3612728360219229, spearmanr = 0.3730812013348164
S-R: pearsonr = -0.015099113643584485, spearmanr = 0.10166852057842046
D-R: pearsonr = 0.6905369778044107, spearmanr = 0.6694104560622914
l1ou fpkm muRatio0 minTau0 minBS99
S-D: pearsonr = 0.3559158040735669, spearmanr = 0.47986651835372635
S-R: pearsonr = 0.10456058871705029, spearmanr = 0.19199110122358176
D-R: pearsonr = 0.7720463772788436, spearmanr = 0.7530589543937708
llou_fpkm_muRatio0_minTau0_minBS99
S-D: pearsonr = 0.45670334552227737, spearmanr = 0.5652947719688542
S-R: pearsonr = 0.28640037372446764, spearmanr = 0.3348164627363737
D-R: pearsonr = 0.824547007618165, spearmanr = 0.7486095661846496
llou fpkm muRatio0 minTau0.5 minBS0
S-D: pearsonr = 0.62020788290504, spearmanr = 0.742018026618497
S-R: pearsonr = 0.09579146077980835, spearmanr = 0.12183993996558765
D-R: pearsonr = 0.3698530358910138, spearmanr = 0.3198932703876434
llou_fpkm_muRatio0_minTau0.5_minBS0
S-D: pearsonr = 0.5850270137481305, spearmanr = 0.7174638487208009
S-R: pearsonr = 0.07901001398799448, spearmanr = 0.12138092158580513
D-R: pearsonr = 0.36790460398377367, spearmanr = 0.3534523349663721
llou fpkm muRatio0 minTau0.5 minBS0
S-D: pearsonr = 0.5600344607496623, spearmanr = 0.6886194280012738
S-R: pearsonr = 0.10086776393164132, spearmanr = 0.16594287984344205
D-R: pearsonr = 0.40351876644490386, spearmanr = 0.4283807513617259
11ou_fpkm_muRatio0_minTau0.5_minBS0
S-D: pearsonr = 0.5640964201494536, spearmanr = 0.7205784204671858
S-R: pearsonr = 0.13964779249554532, spearmanr = 0.2652858656289121
D-R: pearsonr = 0.42688354514991617, spearmanr = 0.47978652776211306
11ou_fpkm_muRatio0_minTau0.5_minBS99
S-D: pearsonr = 0.5536278956679052, spearmanr = 0.5719688542825361
S-R: pearsonr = -0.005889481541357603, spearmanr = 0.05009218381837329
D-R: pearsonr = 0.4410980794502596, spearmanr = 0.5096823546362735
11ou_fpkm_muRatio0_minTau0.5_minBS99
S-D: pearsonr = 0.5014184826152503, spearmanr = 0.5701890989988877
S-R: pearsonr = -0.027285643398619695, spearmanr = 0.0386273638092479
D-R: pearsonr = 0.43633577919571626, spearmanr = 0.5216939891214121
11ou_fpkm_muRatio0_minTau0.5_minBS99
S-D: pearsonr = 0.45828864334425834, spearmanr = 0.6022246941045606
S-R: pearsonr = -0.011431965307467616, spearmanr = 0.09209773706517062
D-R: pearsonr = 0.4630890763654339, spearmanr = 0.5701074552961051
11ou_fpkm_muRatio0_minTau0.5_minBS99
S-D: pearsonr = 0.4517691426632375, spearmanr = 0.5701890989988877
S-R: pearsonr = 0.030580834048735288, spearmanr = 0.11950242955773359
```

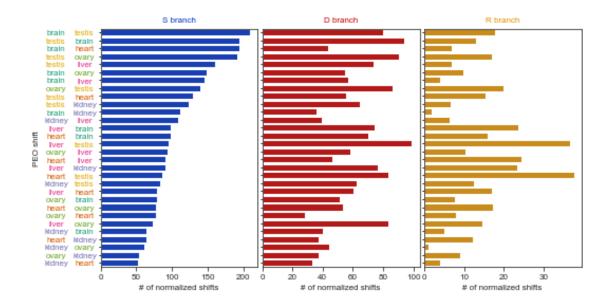
D-R: pearsonr = 0.4730841762014321, spearmanr = 0.5849329446773276

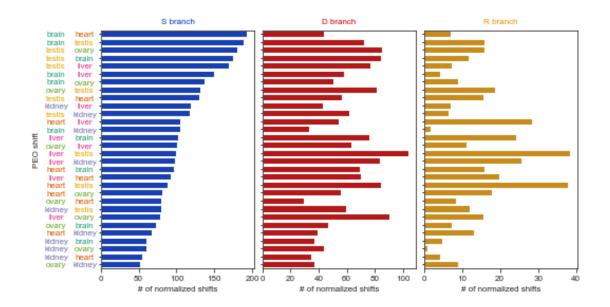


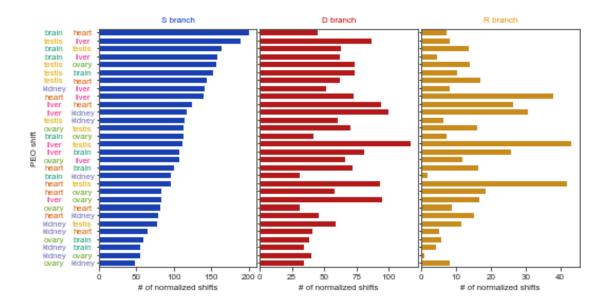


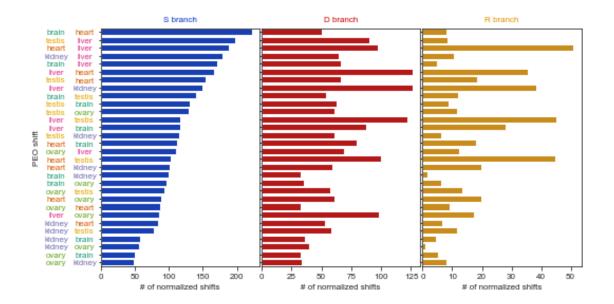


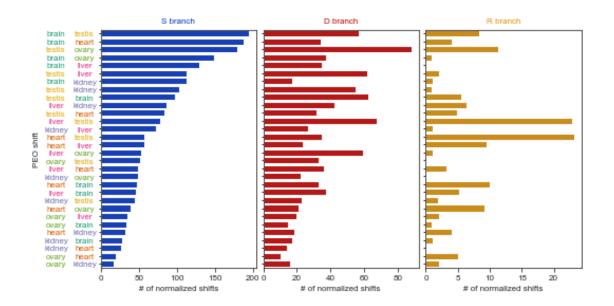


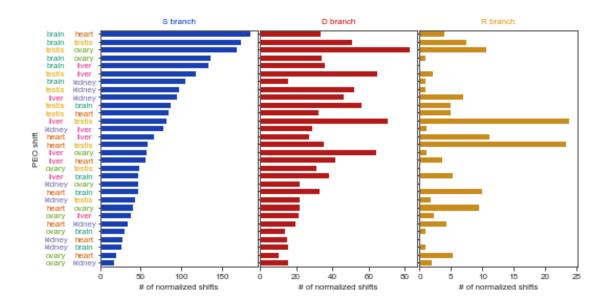


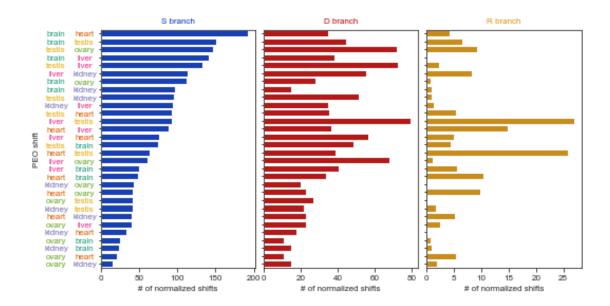


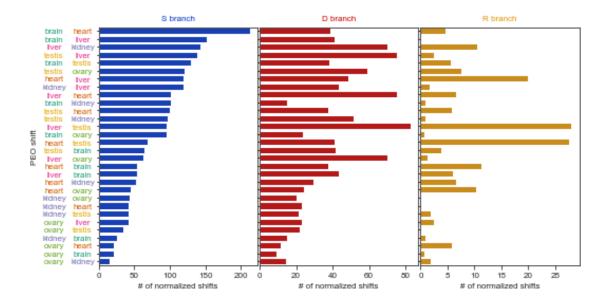


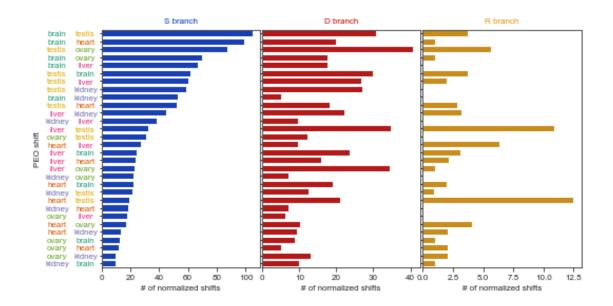


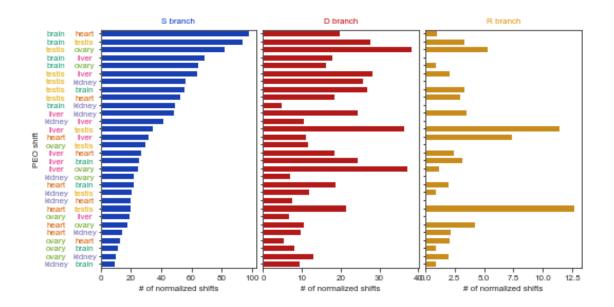


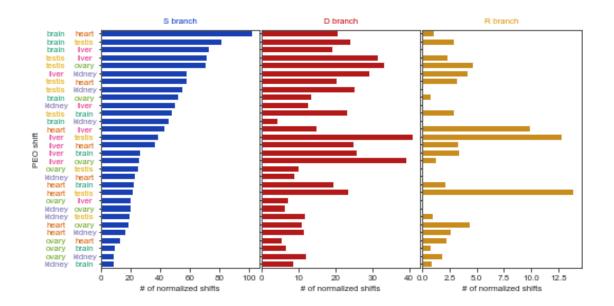


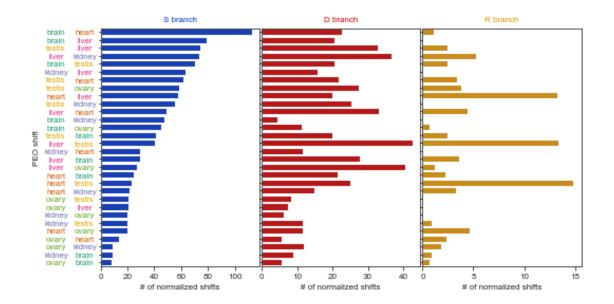












```
for threshold in thresholds:
    fig,axes = matplotlib.pyplot.subplots(nrows=1, ncols=1, figsize=(7.2,2.4),
    sharex=False)

#axes = axes.flat

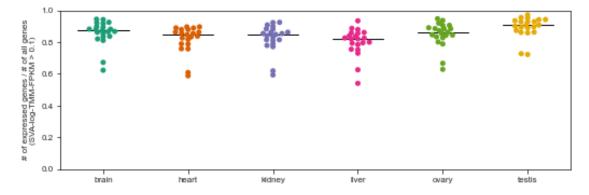
tmp1 = expressed_ratio.loc[(expressed_ratio['threshold']==threshold),organs]

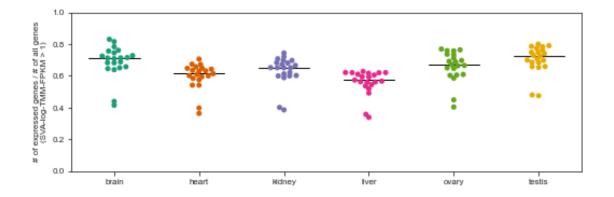
tmp = tmp1.stack().reset_index().drop('level_0', axis=1)

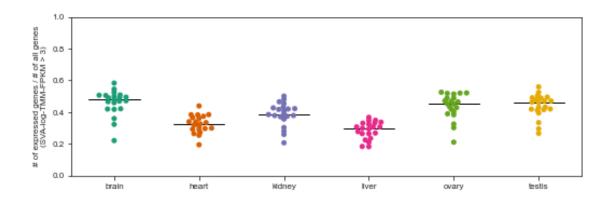
tmp.columns = ['organ','expressed_gene_ratio']

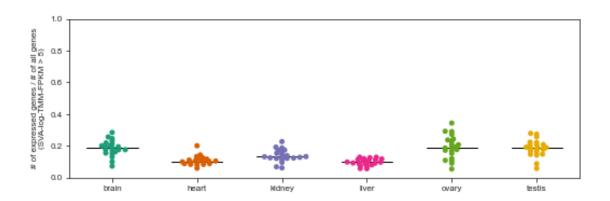
median_y = tmp1.median().values

median_x_from = numpy.arange(len(median_y))-0.3
```









```
event_combinations = list(itertools.combinations(branch_categories, 2))
print('event_combinations:', event_combinations)
for pp in pcm_prefixes:
   for min_tau in min_taus:
      for min_max_second_mu_ratio in min_max_second_mu_ratios:
          for min_bs in min_bss:
             specificity_term =
→pp+'muRatio'+str(min_max_second_mu_ratio)+'_minTau'+str(min_tau)+'_minBS'+str(min_bs)
             print(specificity_term)
             observed = dict()
             corrected = dict()
             for event in ['S','D','R']:
                infile = 
observed[event] = pandas.read_csv(infile, sep='\t',__
→header=0, index_col=0)
                infile2 =
```

```
corrected[event] = pandas.read_csv(infile2, sep='\t',__
→header=0, index_col=0)
               for event combination in event combinations:
                   tested_values = observed
                   e1 = event combination[0]
                   e2 = event combination[1]
                   tmp = tested_values[e1]; tmp1 = tmp.values.reshape(tmp.
\rightarrow shape [0] *tmp.shape [1])
                   tmp = tested_values[e2]; tmp2 = tmp.values.reshape(tmp.
\rightarrow shape [0] *tmp.shape [1])
                   ctable = numpy.array([tmp1,tmp2])
                   ctable = ctable[:,(ctable.sum(axis=0)>0)]
                   out = scipy.stats.chi2_contingency(ctable, correction=True,_
→lambda_=None)
                   print('ancestral-derived', e1, e2, 'pvalue =', out[1], __
for event_combination in event_combinations:
                   tested_values = observed
                   e1 = event_combination[0]
                   e2 = event_combination[1]
                   tmp = tested_values[e1]; tmp1 = tmp.values.sum(axis=0)
                   tmp = tested_values[e2]; tmp2 = tmp.values.sum(axis=0)
                   ctable = numpy.array([tmp1,tmp2])
                   ctable = ctable[:,(ctable.sum(axis=0)>0)]
                   out = scipy.stats.chi2_contingency(ctable, correction=True,_
→lambda =None)
                   print('only derived', e1, e2, 'pvalue =', out[1], 'chi2_
\rightarrowstat =', out[0])
               for event in branch categories:
                   tmp = observed[event]
                   tmp = tmp.values.reshape(tmp.shape[0]*tmp.shape[1])
                   tmp = tmp/tmp.sum()
                   out = scipy.stats.chisquare(tmp)
                   print('ancestral-derived', event, 'pvalue =', out[1], 'chi2_
\hookrightarrowstat =', out[0])
               for event in branch_categories:
                   tmp = observed[event]
                   tmp = tmp.values.sum(axis=0)
                   tmp = tmp/tmp.sum()
                   out = scipy.stats.chisquare(tmp)
                   print('only derived', event, 'pvalue =', out[1], 'chi2 stat⊔
\rightarrow=', out[0])
```

```
for event in branch_categories:
                     tmp = observed[event]
                     dif = 0
                     organs = list(set(tmp.index.tolist()).intersection(set(tmp.

→columns.tolist())))
                     for i in itertools.combinations(organs, 2):
                         if i[0]!=i[1]:
                             dif = dif + numpy.abs(tmp.loc[i[0],i[1]] - tmp.
 \rightarrowloc[i[1],i[0]])
                     symmetry = 1 - (dif/tmp.sum().sum())
                     print(event, 'symmetry of the number of shift =', symmetry)
                 for event in branch_categories:
                     tmp = corrected[event]
                     dif = 0
                     organs = list(set(tmp.index.tolist()).intersection(set(tmp.

→columns.tolist())))
                     for i in itertools.combinations(organs, 2):
                         if i[0]!=i[1]:
                             dif = dif + numpy.abs(tmp.loc[i[0],i[1]] - tmp.
 \rightarrowloc[i[1],i[0]])
                     symmetry = 1 - (dif/tmp.sum().sum())
                     print(event, 'symmetry of the acceleration of shift =', __
 →symmetry)
                print()
event_combinations: [('S', 'D'), ('S', 'R'), ('D', 'R')]
llou_fpkm_muRatio0_minTau0_minBS0
ancestral-derived S D pvalue = 1.1645425725499324e-33 chi2 stat =
232.6015555526961
ancestral-derived S R pvalue = 3.15741388058073e-54 chi2 stat =
337.26871370739775
ancestral-derived D R pvalue = 7.322841049325359e-27 chi2 stat =
196.83199619773367
only derived S D pvalue = 0.0006312751579895948 chi2 stat = 21.57219097992501
only derived S R pvalue = 5.727871322112838e-17 chi2 stat = 85.56596341989552
only derived D R pvalue = 4.704916901656827e-10 chi2 stat = 52.28968702326899
ancestral-derived S pvalue = 1.0 chi2 stat = 0.3880086372405372
ancestral-derived D pvalue = 1.0 chi2 stat = 0.35234780855959336
ancestral-derived R pvalue = 1.0 chi2 stat = 0.8670298787456246
only derived S pvalue = 0.9999968230540931 chi2 stat = 0.020499147605217337
only derived D pvalue = 0.9999840101891352 chi2 stat = 0.03923108052664371
only derived R pvalue = 0.998601936427763 chi2 stat = 0.24143061475321465
S symmetry of the number of shift = 0.848388033691548
D symmetry of the number of shift = 0.8856664807585053
```

```
R symmetry of the number of shift = 0.5630026809651474
S symmetry of the acceleration of shift = 0.954120276015039
D symmetry of the acceleration of shift = 0.9380191481355096
R symmetry of the acceleration of shift = 0.8420599103110884
llou_fpkm_muRatio0_minTau0_minBS99
ancestral-derived S D pvalue = 1.1656647990706282e-19 chi2 stat =
157.77142932826834
ancestral-derived S R pvalue = 4.92143783390324e-28 chi2 stat =
203.0640970783855
ancestral-derived D R pvalue = 3.0619835981501303e-06 chi2 stat =
77.09101926176949
only derived S D pvalue = 7.682996259087532e-06 chi2 stat = 31.4357002043132
only derived S R pvalue = 1.0529379252262355e-06 chi2 stat = 35.776145833250084
only derived D R pvalue = 0.02079599317957807 chi2 stat = 13.291378457948749
ancestral-derived S pvalue = 1.0 chi2 stat = 0.42782374468613504
ancestral-derived D pvalue = 1.0 chi2 stat = 0.3441118851588771
ancestral-derived R pvalue = 1.0 chi2 stat = 0.7244809688581315
only derived S pvalue = 0.9999990983378625 chi2 stat = 0.012372157949220062
only derived D pvalue = 0.9999674762800292 chi2 stat = 0.05221283421048528
only derived R pvalue = 0.9994212998676993 chi2 stat = 0.16789215686274508
S symmetry of the number of shift = 0.8333837314786816
D symmetry of the number of shift = 0.8707557502738226
R symmetry of the number of shift = 0.6274509803921569
S symmetry of the acceleration of shift = 0.9492625829490327
D symmetry of the acceleration of shift = 0.9407586790309318
R symmetry of the acceleration of shift = 0.793684663033914
llou_fpkm_muRatio0_minTau0.5_minBS0
ancestral-derived S D pvalue = 8.887362232292662e-18 chi2 stat =
147.27294260834327
ancestral-derived S R pvalue = 2.941578886654832e-31 chi2 stat =
220.05562051022451
ancestral-derived D R pvalue = 3.816019765323653e-14 chi2 stat =
126.51240092175345
only derived S D pvalue = 0.00021515460123787145 chi2 stat = 24.020476732194734
only derived S R pvalue = 1.7443494961517143e-07 chi2 stat = 39.66533050014218
only derived D R pvalue = 0.00011752793086720099 chi2 stat = 25.382482350291024
ancestral-derived S pvalue = 1.0 chi2 stat = 0.7454758872600206
ancestral-derived D pvalue = 1.0 chi2 stat = 0.5753159830046348
ancestral-derived R pvalue = 0.999999999999999999999 chi2 stat = 1.9236520877437948
only derived S pvalue = 0.9999883139452478 chi2 stat = 0.03458375249105983
only derived D pvalue = 0.99997347053871 chi2 stat = 0.04809874178551392
only derived R pvalue = 0.9958850594901416 chi2 stat = 0.3791270663483627
S symmetry of the number of shift = 0.6176598521096128
D symmetry of the number of shift = 0.7939508506616257
R symmetry of the number of shift = 0.40268456375838924
S symmetry of the acceleration of shift = 0.878119701406599
```

```
D symmetry of the acceleration of shift = 0.9102296154643834
R symmetry of the acceleration of shift = 0.510984529731863
llou_fpkm_muRatio0_minTau0.5_minBS99
ancestral-derived S D pvalue = 5.566817868652583e-12 chi2 stat =
113.72968199442163
ancestral-derived S R pvalue = 2.377658640736338e-17 chi2 stat =
144.86730756686904
ancestral-derived D R pvalue = 0.0004896507293822123 chi2 stat =
60.807030670771894
only derived S D pvalue = 0.0003013618753243799 chi2 stat = 23.257553274337308
only derived S R pvalue = 0.0010603021798619365 chi2 stat = 20.379911822941423
only derived D R pvalue = 0.06363467877368677 chi2 stat = 10.442109989645738
ancestral-derived S pvalue = 1.0 chi2 stat = 0.8074099268155377
ancestral-derived D pvalue = 1.0 chi2 stat = 0.5914845140619438
ancestral-derived R pvalue = 0.9999999999999 chi2 stat = 1.9888
only derived S pvalue = 0.999995154287944 chi2 stat = 0.02428340213923813
only derived D pvalue = 0.999970025749984 chi2 stat = 0.050523317906728385
only derived R pvalue = 0.996152161078495 chi2 stat = 0.3685333333333327
S symmetry of the number of shift = 0.6044520547945205
D symmetry of the number of shift = 0.7433962264150944
R symmetry of the number of shift = 0.4
S symmetry of the acceleration of shift = 0.8820005458503392
D symmetry of the acceleration of shift = 0.8583233271629571
R symmetry of the acceleration of shift = 0.41545364137615903
llou_tpm_muRatio0_minTau0_minBS0
ancestral-derived S D pvalue = 1.2803596852689708e-29 chi2 stat =
211.44194549979858
ancestral-derived S R pvalue = 3.2765565191475903e-44 chi2 stat =
286.7964343857895
ancestral-derived D R pvalue = 1.189506679469606e-24 chi2 stat =
184.99699829145175
only derived S D pvalue = 0.026558245513776127 chi2 stat = 12.68104914518126
only derived S R pvalue = 5.0033486447679886e-26 chi2 stat = 128.47921998711317
only derived D R pvalue = 4.510200345340426e-18 chi2 stat = 90.82403547603904
ancestral-derived S pvalue = 1.0 chi2 stat = 1.027348418530505
ancestral-derived D pvalue = 1.0 chi2 stat = 0.8780762217023703
ancestral-derived R pvalue = 1.0 chi2 stat = 1.7074585586453934
only derived S pvalue = 0.9997000426799068 chi2 stat = 0.1283606011224609
only derived D pvalue = 0.9996496843152228 chi2 stat = 0.13674446020372452
only derived R pvalue = 0.9902825003612684 chi2 stat = 0.5474547031782032
S symmetry of the number of shift = 0.8771346531660873
D symmetry of the number of shift = 0.884692417739628
R symmetry of the number of shift = 0.6030855539971949
S symmetry of the acceleration of shift = 0.946570717472631
D symmetry of the acceleration of shift = 0.9167071309305176
R symmetry of the acceleration of shift = 0.7873259282130881
```

```
11ou_tpm_muRatio0_minTau0_minBS99
ancestral-derived S D pvalue = 1.0523789030835427e-14 chi2 stat =
129.760188956094
ancestral-derived S R pvalue = 2.2029154641494666e-24 chi2 stat =
183.55598383597146
ancestral-derived D R pvalue = 5.481109524503753e-08 chi2 stat =
88.8422765150004
only derived S D pvalue = 0.00011722731137690391 chi2 stat = 25.388232956822737
only derived S R pvalue = 4.846318953685127e-14 chi2 stat = 71.56207476214485
only derived D R pvalue = 6.954310074482721e-06 chi2 stat = 31.654533039134602
ancestral-derived S pvalue = 1.0 chi2 stat = 1.0224654361332297
ancestral-derived D pvalue = 1.0 chi2 stat = 0.9731807028857056
ancestral-derived R pvalue = 1.0 chi2 stat = 1.695183464188431
only derived S pvalue = 0.9998081177192559 chi2 stat = 0.10702948757777357
only derived D pvalue = 0.999425349627942 chi2 stat = 0.1674097245675057
only derived R pvalue = 0.993818985787964 chi2 stat = 0.4506521390802757
S symmetry of the number of shift = 0.8443305573350417
D symmetry of the number of shift = 0.849360755975542
R symmetry of the number of shift = 0.6227848101265823
S symmetry of the acceleration of shift = 0.9119215046331075
D symmetry of the acceleration of shift = 0.8973572879495197
R symmetry of the acceleration of shift = 0.7334591303407625
llou_tpm_muRatio0_minTau0.5_minBS0
ancestral-derived S D pvalue = 1.3961237112388258e-18 chi2 stat =
151.7750466106425
ancestral-derived S R pvalue = 3.0760106664249445e-14 chi2 stat =
127.05739423406402
ancestral-derived D R pvalue = 1.6272885044004317e-08 chi2 stat =
92.26093633030023
only derived S D pvalue = 0.007310476845760265 chi2 stat = 15.841827457785897
only derived S R pvalue = 1.8422147302824774e-07 chi2 stat = 39.547683650746634
only derived D R pvalue = 1.9503892685336116e-05 chi2 stat = 29.382745603784713
ancestral-derived S pvalue = 1.0 chi2 stat = 1.0842219833378124
ancestral-derived D pvalue = 1.0 chi2 stat = 1.0983373256737647
ancestral-derived R pvalue = 0.99999999999999999999999 chi2 stat = 1.8371511870054145
only derived S pvalue = 0.9996251217498067 chi2 stat = 0.14057847352862132
only derived D pvalue = 0.9996390027501225 chi2 stat = 0.1384304427970715
only derived R pvalue = 0.9850273747764663 chi2 stat = 0.6612522560044426
S symmetry of the number of shift = 0.6885245901639344
D symmetry of the number of shift = 0.8108581436077058
R symmetry of the number of shift = 0.6938775510204082
S symmetry of the acceleration of shift = 0.9038234071657318
D symmetry of the acceleration of shift = 0.8945460653736539
R symmetry of the acceleration of shift = 0.6542436553654958
```

llou_tpm_muRatioO_minTau0.5_minBS99

```
ancestral-derived S D pvalue = 1.3431121459082411e-07 chi2 stat =
     86.28509322451443
     ancestral-derived S R pvalue = 8.085657493226364e-09 chi2 stat =
     94.20781351567337
     ancestral-derived D R pvalue = 0.006416527451683615 chi2 stat =
     51.36161390751743
     only derived S D pvalue = 0.08519227067820091 chi2 stat = 9.668488156358054
     only derived S R pvalue = 0.001024127642288065 chi2 stat = 20.460015000800386
     only derived D R pvalue = 0.021569047499517435 chi2 stat = 13.200677857356986
     ancestral-derived S pvalue = 1.0 chi2 stat = 1.1060314403793423
     ancestral-derived D pvalue = 1.0 chi2 stat = 0.9854987262394669
     ancestral-derived R pvalue = 0.99999999999992 chi2 stat = 2.251249999999998
     only derived S pvalue = 0.9998467030705749 chi2 stat = 0.0977071600693814
     only derived D pvalue = 0.9995873536266715 chi2 stat = 0.14619784813788342
     only derived R pvalue = 0.9891192452486938 chi2 stat = 0.574999999999998
     S symmetry of the number of shift = 0.6935086277732128
     D symmetry of the number of shift = 0.8007054673721341
     R symmetry of the number of shift = 0.675
     S symmetry of the acceleration of shift = 0.8607710100595406
     D symmetry of the acceleration of shift = 0.8794435751326903
     R symmetry of the acceleration of shift = 0.47099265100117405
[29]: fig = matplotlib.pyplot.figure(figsize=(1, 2.4))
      gs = matplotlib.gridspec.GridSpec(nrows=2, ncols=5)
      ax0 = matplotlib.pyplot.subplot(gs[0,:])
      ax1 = matplotlib.pyplot.subplot(gs[1,2])
      ax = ax0
      ax.axis('off')
      leg_handles = list()
      leg_handles.append(matplotlib.lines.Line2D([], [], color='black', label='0-10', []
      \rightarrowlinewidth=0.5))
      leg_handles.append(matplotlib.lines.Line2D([], [], color='black', __
      →label='11-100', linewidth=1))
      leg handles.append(matplotlib.lines.Line2D([], [], color='black', label='>100', []
      →linewidth=2))
      leg = ax.legend(handles=leg_handles, loc='upper center', frameon=False, u
      →fontsize=font_size)
      leg.set_title('# expression shift', prop={'weight': 'normal'})
      leg.get_title().set_fontsize(font_size)
      #leq._leqend_box.aliqn = "left"
      ax = ax1
      cmap = matplotlib.colors.ListedColormap(['#00FF00', '#40C040', '#808080', __
      bounds = [1, 2, 3, 4, 5, 6]
```

```
ticks = bounds#[1.5,2.5,3.5,4.5,5.5]

ticklabels = ['0%', '0.5%', '2.5%', '97.5%', '99.5%', '100%']

norm = matplotlib.colors.BoundaryNorm(bounds, cmap.N)

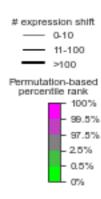
title = 'Permutation-based\npercentile rank'

cbar = matplotlib.colorbar.ColorbarBase(ax, cmap=cmap, norm=norm, orientation='vertical', ticks=ticks)

cbar.ax.set_yticklabels(ticklabels, ha='left', fontsize=font_size)

ax.set_title(title, ha='center', fontsize=font_size)

fig.savefig('colorbar_lime_magenta.svg', format='svg', transparent=True)
```



bioproject

tissue	brain	heart	kidney	liver	ovary	testis
scientific_name						
Ailuropoda melanoleuca	NaN	NaN	NaN	1.0	1.0	1.0
Anas platyrhynchos	1.0	NaN	NaN	5.0	2.0	NaN
Anolis carolinensis	4.0	3.0	2.0	3.0	2.0	2.0
Aotus nancymaae	NaN	1.0	1.0	1.0	1.0	NaN
Astyanax mexicanus	4.0	3.0	3.0	2.0	2.0	2.0

Bos taurus	3.0	5.0	7.0	27.0	7.0	11.0
Callithrix jacchus	3.0	4.0	4.0	7.0	1.0	4.0
Canis lupus	4.0	4.0	8.0	7.0	1.0	8.0
Cavia porcellus	1.0	NaN	2.0	3.0	1.0	1.0
Cercocebus atys	NaN	1.0	1.0	1.0	NaN	NaN
Chinchilla lanigera	1.0	1.0	1.0	1.0	1.0	1.0
Chlorocebus sabaeus	1.0	NaN	1.0	1.0	NaN	NaN
Danio rerio	7.0	5.0	4.0	5.0	4.0	3.0
Dasypus novemcinctus	NaN	1.0	1.0	1.0	NaN	NaN
Drosophila melanogaster	23.0	1.0	NaN	NaN	64.0	28.0
Equus caballus	1.0	NaN	1.0	1.0	NaN	6.0
Erinaceus europaeus	1.0	NaN	1.0	1.0	NaN	NaN
Felis catus	3.0	1.0	2.0	3.0	NaN	2.0
Fukomys damarensis	1.0	NaN	1.0	1.0	1.0	1.0
Gadus morhua	1.0	1.0	1.0	2.0	1.0	1.0
Gallus gallus	8.0	14.0	13.0	26.0	13.0	14.0
Gasterosteus aculeatus	6.0	1.0	5.0	3.0	NaN	1.0
Gorilla gorilla	NaN	1.0	1.0	1.0	NaN	3.0
Homo sapiens	4.0	4.0	5.0		2.0	4.0
Lepisosteus oculatus	2.0	2.0	2.0		1.0	2.0
Macaca fascicularis	2.0	1.0	3.0		NaN	1.0
Macaca mulatta	12.0	10.0	12.0		2.0	17.0
Macaca nemestrina	1.0	1.0	1.0	1.0	NaN	NaN
Meleagris gallopavo	1.0	1.0	NaN	4.0	NaN	NaN
Mesocricetus auratus	2.0	NaN	2.0	3.0	NaN	NaN
Microcebus murinus	NaN	NaN	1.0	3.0	NaN	NaN
Monodelphis domestica	7.0	5.0	5.0	7.0	4.0	8.0
Mus caroli	1.0	1.0	1.0	2.0	NaN	1.0
Mus musculus	5.0	3.0			2.0	3.0
Mus pahari	1.0	1.0	1.0			NaN
Oreochromis niloticus	3.0	2.0	3.0		2.0	2.0
Ornithorhynchus anatinus	4.0	3.0	3.0	2.0	3.0	4.0
Oryctolagus cuniculus	2.0	2.0	3.0			2.0
Oryzias latipes	3.0	2.0	1.0	6.0	3.0	4.0
Ovis aries	4.0	5.0	3.0	8.0	9.0	4.0
Pan paniscus	NaN	1.0	1.0	1.0	NaN	1.0
Pan troglodytes	2.0	4.0	4.0	6.0	NaN	3.0
Papio anubis	1.0	3.0	2.0	3.0	NaN	NaN
Petromyzon marinus	2.0	NaN	1.0	2.0	NaN	1.0
Rattus norvegicus	4.0	4.0	5.0	5.0	3.0	4.0
3	1.0	1.0				NaN
Rhinopithecus bieti			1.0	1.0	NaN 1 0	
Saimiri boliviensis	1.0	1.0	1.0	1.0	1.0	NaN
Sus scrofa	4.0	8.0	9.0	32.0	10.0	9.0
Taeniopygia guttata	2.0	NaN	NaN	NaN	NaN 1 0	1.0
Takifugu rubripes	NaN	NaN	2.0	3.0	1.0	2.0
Tupaia belangeri	1.0	NaN	NaN	2.0	NaN	1.0
Xenopus tropicalis	4.0	3.0	4.0	7.0	3.0	3.0

```
[31]: | spp = b.loc[b.so_event=='L', 'taxon'].sort_values().unique()
      exp_methods = ['tmm_rpkm','tpm']
      for exp_method in exp_methods:
          mydir = os.path.join(dir_tc, exp_method+'/', 'sra/')
          files = os.listdir(mydir)
          files = [ f for f in files if f.endswith('.tsv') ]
          dfpiv = pandas.DataFrame()
          for file in files:
              s = pandas.read_csv(mydir+file, sep='\t', header=0)
              s['count'] = 1
              piv = s.pivot_table(index='exclusion', columns='scientific_name',__
       →values='count', aggfunc=sum, fill_value=0)
              dfpiv = pandas.concat([dfpiv, piv], axis=1, sort=True)
          dfpiv = dfpiv.fillna(0).astype(int)
          dfpiv.index = dfpiv.index.str.replace('^no$', 'non_excluded')
          dfpiv = dfpiv.loc[dfpiv.index.sort_values(),dfpiv.columns.sort_values()]
          num ex = dfpiv.loc[~dfpiv.index.str.contains('non excluded'),:].sum(axis=0)
          num_nonex = dfpiv.loc['non_excluded',:]
          dfpiv.loc['exclusion_rate (%)',:] = num_ex / (num_ex+num_nonex) * 100
          last_ind = ['non_excluded','exclusion_rate (%)']
          ind = [ i for i in dfpiv.index if i not in last_ind ] + last_ind
          dfpiv = dfpiv.loc[ind,spp]
          dfpiv = dfpiv.loc[dfpiv.sum(axis=1)!=0,:].astype(int)
          outfile = 'exclusion '+exp method+'.tsv'
          dfpiv.to_csv(outfile, sep='\t', index=True)
[32]: sra_date = '2018_5_1'
      #dir_ks = os.path.join(dir_ensembl, 'kallisto_summary/', sra_date)
      #dir_fpkm = os.path.join(dir_ks, 'fpkm.tmm.kallisto.gene.log.tsv/')
      \#dir\_tpm = os.path.join(dir\_ks, 'tpm.masked.kallisto.gene.log.tsv/')
      dir tc = os.path.join(dir ensembl, 'curated transcriptome/', sra date+'/')
      dir_fpkm = os.path.join(dir_tc, 'tmm_rpkm/', 'tc/')
      dir_tpm = os.path.join(dir_tc, 'tpm/', 'tc/')
      dir_sra = os.path.join(dir_ensembl, 'sra/', sra_date)
      file_sra = os.path.join(dir_sra, 'sra_table_amalgamated_'+sra_date+'.tsv')
      df_sra = pandas.read_csv(file_sra, sep='\t', header=0)
      nsample = 1000
      # GAPDH https://www.ensembl.org/Homo_sapiens/Gene/Compara_Ortholog?db=core;
      \rightarrow q = ENSG00000111640; r = 12:6534512 - 6538374
      species_genes = [
          ['Anolis carolinensis', 'ENSACAG00000006502'],
          ['Astyanax mexicanus', 'ENSAMXG00000039361'],
          ['Bos taurus', 'ENSBTAG00000014731'],
          ['Callithrix jacchus', 'ENSCJAG00000008234'], # spermatogenic
          ['Canis lupus', 'ENSCAFG00000007743'],
```

```
['Chinchilla lanigera', 'ENSCLAG00000013783'],
    ['Danio rerio', 'ENSDARG00000043457'],
    ['Gadus morhua', 'ENSGMOG00000010369'],
    ['Gallus gallus', 'ENSGALG00000014442'],
    ['Homo sapiens', 'ENSG00000111640'],
    ['Macaca mulatta', 'ENSMMUG00000018679'],
    ['Monodelphis domestica', 'ENSMODG00000011838'],
    ['Mus musculus', 'ENSMUSG00000057666'],
    ['Oreochromis niloticus', 'ENSONIGO0000012916'],
    ['Ornithorhynchus anatinus', 'ENSOANGO000004492'],
    ['Oryctolagus cuniculus', 'ENSOCUG00000025023'],
    ['Oryzias latipes', 'ENSORLG00000012224'], # or ENSORLG00020000877, or_
→ ENSORLG00015011254
    ['Ovis aries', 'ENSOARG00000007894'],
    ['Rattus norvegicus', 'ENSRNOG00000018630'],
    ['Sus scrofa', 'ENSSSCG00000000694'],
    ['Xenopus tropicalis', 'ENSXETG00000033975'],
]
def get_subsampled_stat(gene, df_exp, df_sra, nsample, func='median',_
→verbose=False):
    conditions = True
    conditions = conditions & (df_sra['scientific_name'] == sci_name)
    conditions = conditions & (df_sra['tissue']==tissue)
    bioprojects = df_sra.loc[conditions, 'bioproject'].unique()
    num_bp = len(bioprojects)
    df sat = pandas.DataFrame({'num bp':[],'ave exp':[]})
    for i in numpy.arange(num_bp)+1:
        flag_permutation = False
        counter = 0
        for bps in itertools.combinations(bioprojects, i):
            counter +=1
            if counter>=nsample:
                flag permutation = True
                break
        if flag_permutation:
            if verbose:
                print('Combinations of {}. Permutation.'.format(i))
            for j in numpy.arange(0, nsample):
                bps = numpy.random.choice(a=bioprojects, size=i, replace=False)
                runs = df_sra.loc[(df_sra['bioproject'].isin(bps)), 'run'].
→unique()
                if func=='mean':
                    ave = df exp.loc[gene,runs].mean()
                elif func=='median':
                    ave = df_exp.loc[gene,runs].median()
```

```
df_sat = df_sat.append({'num_bp':i, 'ave_exp':ave},__
 →ignore_index=True)
        else:
            if verbose:
                print('Combinations of {}. Exhaustive. Count of combinations =_
 →{}'.format(i, counter))
            for bps in itertools.combinations(bioprojects, i):
                runs = df_sra.loc[(df_sra['bioproject'].isin(bps)), 'run'].
 →unique()
                if func=='mean':
                    ave = df_exp.loc[gene,runs].mean()
                elif func=='median':
                    ave = df_exp.loc[gene,runs].median()
                df_sat = df_sat.append({'num_bp':i,'ave_exp':ave},__
 →ignore_index=True)
    df_sat.loc[:,'num_bp'] = df_sat.loc[:,'num_bp'].astype(int)
    return df sat
dfs = dict()
for sg in species genes:
    sci name = sg[0]
    gene = sg[1]
    print(sci name)
    dfs[sci_name] = pandas.DataFrame()
    for exp_key in ['fpkm','tpm']:
        if exp_key=='fpkm':
            file_fpkm = os.path.join(dir_fpkm, sci_name.replace(' ','_')+'.tc.
→tsv')
            df_exp = pandas.read_csv(file_fpkm, sep='\t', header=0)
        elif exp_key=='tpm':
            file_tpm = os.path.join(dir_tpm, sci_name.replace(' ','_')+'.tc.

→tsv')
            df_exp = pandas.read_csv(file_tpm, sep='\t', header=0)
        df_sra2 = df_sra.loc[(df_sra['run'].isin(df_exp.columns)),:]
        if gene in df_exp.index:
            for tissue in tissues:
                #file fpkm = os.path.join(dir fpkm, sci name.replace(' ',' ')+'.
 \rightarrow gene. log. tsv')
                #file_tpm = os.path.join(dir_tpm, sci_name.replace(' ','_')+'.
\rightarrow gene. log. tsv')
                tmp = get_subsampled_stat(gene, df_exp, df_sra2, nsample)
                tmp['unit'] = exp_key
                tmp['tissue'] = tissue
                dfs[sci_name] = pandas.concat([dfs[sci_name], tmp],__
 →ignore_index=True)
        else:
```

```
print('Gene ID ({}) not found in {}'.format(gene, sci_name))
Anolis carolinensis
```

```
Astyanax mexicanus
Gene ID (ENSAMXG00000039361) not found in Astyanax mexicanus
Gene ID (ENSAMXG00000039361) not found in Astyanax mexicanus
Bos taurus
Callithrix jacchus
Canis lupus
Chinchilla lanigera
Danio rerio
Gadus morhua
Gallus gallus
Homo sapiens
Macaca mulatta
Monodelphis domestica
Mus musculus
Oreochromis niloticus
Ornithorhynchus anatinus
Oryctolagus cuniculus
Oryzias latipes
Ovis aries
Rattus norvegicus
Sus scrofa
Xenopus tropicalis
```

```
[33]: alpha=1
      for sg in species_genes:
          sci_name = sg[0]
          gene = sg[1]
          if dfs[sci_name].shape[0]==0:
              print("{} not found in input.".format(sci_name))
          else:
              print("{} found in input.".format(sci_name))
              fig,axes = matplotlib.pyplot.subplots(nrows=2, ncols=6, figsize=(7.2,2.
       →6), sharex=False, sharey=False)
              axes = axes.flat
              i = 0
              for exp_key, ylabel in_
       \rightarrowzip(['fpkm','tpm'],['Mean\nSVA-log-TMM-FPKM','Mean\nSVA-log-TPM']):
                  ymin = numpy.inf
                  ymax = -numpy.inf
                  for tissue in tissues:
                      tmp = dfs[sci_name].
       →loc[(dfs[sci_name]['unit']==exp_key)&(dfs[sci_name]['tissue']==tissue),:]
                      ymin = min(ymin, tmp['ave_exp'].min())
```

```
ymax = max(ymax, tmp['ave_exp'].max())
               yunit = (ymax-ymin)*0.025
               ymin = ymin - yunit
               ymax = ymax + yunit
           for j,tissue in enumerate(tissues):
               ax = axes[i]
               tmp = dfs[sci_name].
→loc[(dfs[sci_name]['unit']==exp_key)&(dfs[sci_name]['tissue']==tissue),:]
               count = tmp.groupby('num_bp').count()
               lower_count = count.index[count['ave_exp'] < 10].tolist()</pre>
               is_lower = (tmp['num_bp'].isin(lower_count))
               tmp2 = tmp.copy()
               tmp2.loc[~is_lower, 'ave_exp'] = numpy.nan
               seaborn.swarmplot('num_bp', 'ave_exp', data=tmp2, ax=ax, size=2,
                                 color=organ_colors[j])
               for num_bp in tmp2['num_bp'].unique():
                   y = tmp2.loc[tmp2['num_bp']==num_bp, 'ave_exp'].mean()
                   ax.plot([num_bp-1.4,num_bp-0.6], [y,y], lw=0.5,_
if any(~is_lower):
                   tmp2 = tmp.copy()
                   tmp2.loc[is_lower, 'ave_exp'] = numpy.nan
                   seaborn.boxplot('num_bp', 'ave_exp', data=tmp2, ax=ax,
                                   color=organ_colors[j], linewidth=0.5, __
→fliersize=0)
               #ax = change_seaborn_boxplot_linecolors(ax,_
\rightarrow col=organ colors[j])
               ax.set_ylim(ymin, ymax)
               if exp_key=='fpkm':
                   ax.set_title(tissue)
                   ax.set xlabel('')
                   ax.set_xticklabels(['',] * len(ax.get_xticklabels()))
               else:
                   ax.set_xlabel('# of BioProject')
               if j==0:
                   ax.set_ylabel(ylabel)
               else:
                   ax.set_ylabel('')
                   ax.set_yticklabels(['',] * len(ax.get_yticklabels()))
               while len(ax.get_xticklabels())>8:
                   xticks = ax.get_xticks()[0::2]
                   xticklabels = [ tl._text for tl in ax.get_xticklabels()[0::
→2] ]
                   ax.set_xticks(ticks=xticks)
                   ax.set_xticklabels(labels=xticklabels)
               i += 1
       fig.tight_layout()
```

```
outbase = 'mean_reversion_'+sci_name.replace(' ','_')+"_"+gene
fig.savefig(outbase+".pdf", format='pdf', transparent=True)
fig.savefig(outbase+".svg", format='svg', transparent=True)
```

Anolis carolinensis found in input.
Astyanax mexicanus not found in input.
Bos taurus found in input.
Callithrix jacchus found in input.
Canis lupus found in input.
Chinchilla lanigera found in input.

/Users/kef74yk/anaconda3/lib/python3.6/site-packages/ipykernel_launcher.py:42: UserWarning: Attempting to set identical bottom == top == 14.64062997997635 results in singular transformations; automatically expanding.
/Users/kef74yk/anaconda3/lib/python3.6/site-packages/ipykernel_launcher.py:42: UserWarning: Attempting to set identical bottom == top == 10.759435623 results in singular transformations; automatically expanding.

Danio rerio found in input.

Gadus morhua found in input.

Gallus gallus found in input.

Homo sapiens found in input.

Macaca mulatta found in input.

Monodelphis domestica found in input.

Mus musculus found in input.

Oreochromis niloticus found in input.

Ornithorhynchus anatinus found in input.

Oryctolagus cuniculus found in input.

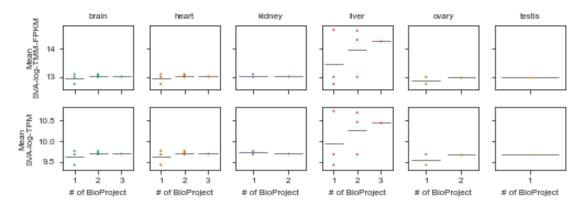
Oryzias latipes found in input.

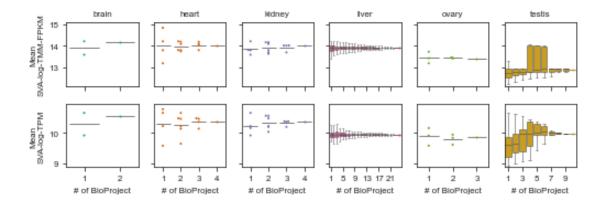
Ovis aries found in input.

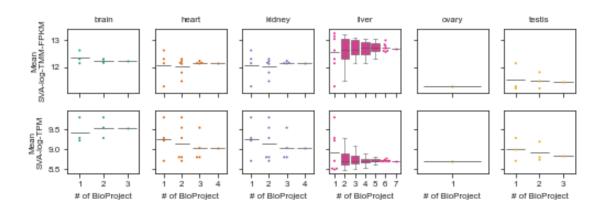
Rattus norvegicus found in input.

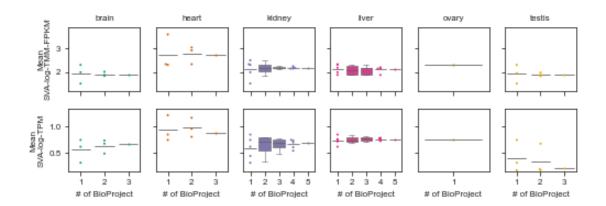
Sus scrofa found in input.

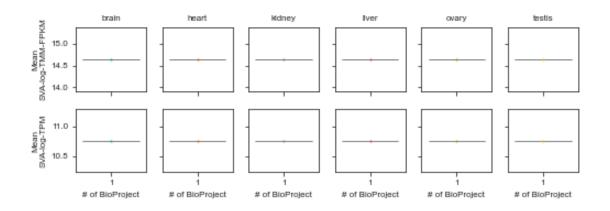
Xenopus tropicalis found in input.

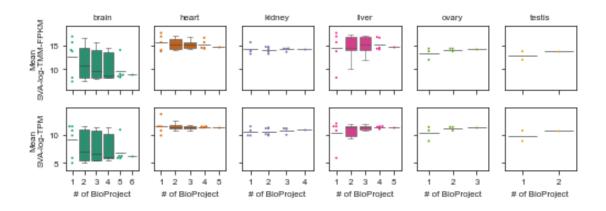


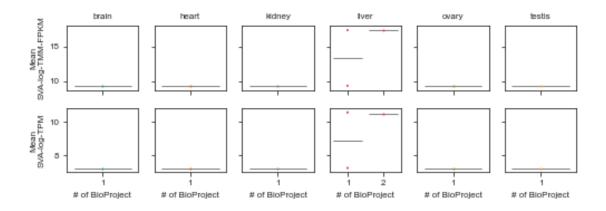


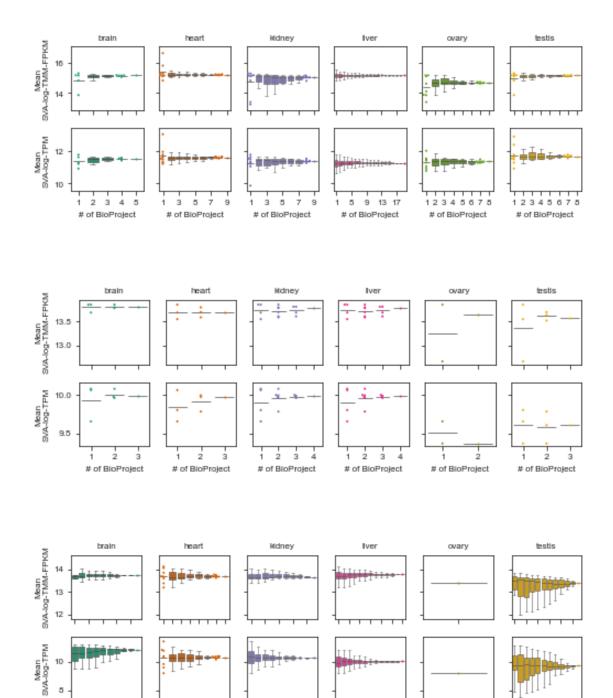












of BioProject

of BioProject

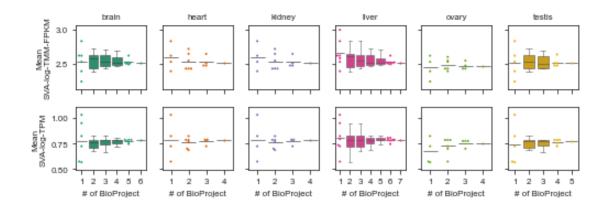
of BioProject

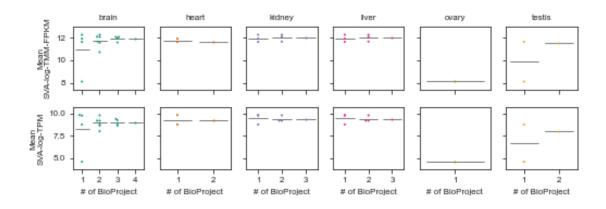
of BioProject

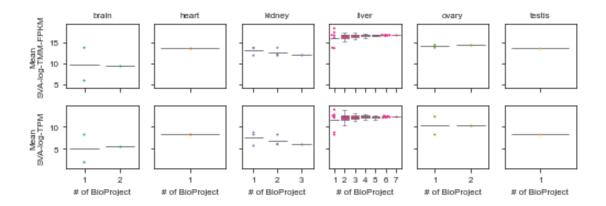
10

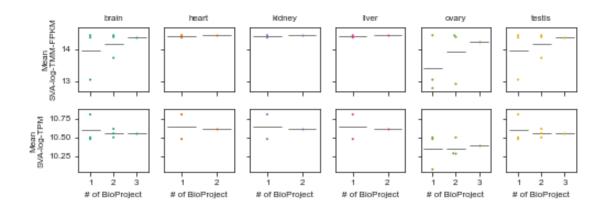
of BioProject

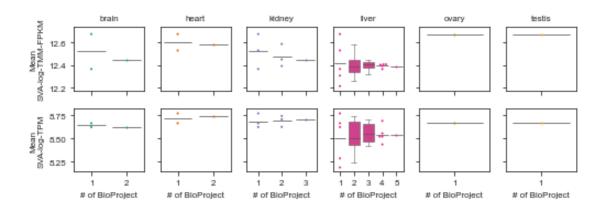
of BioProject

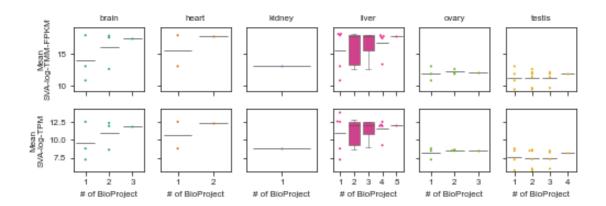


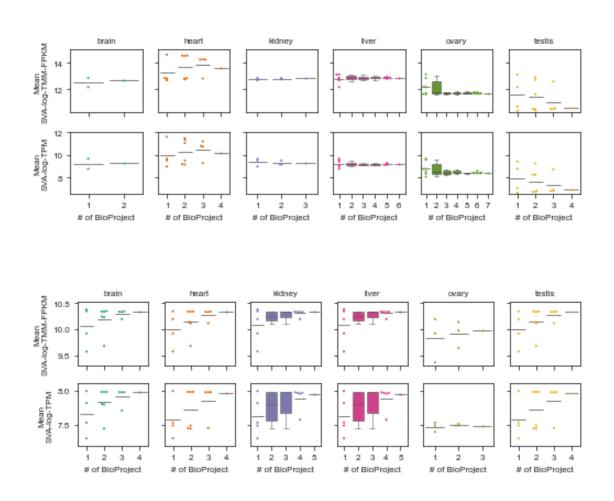


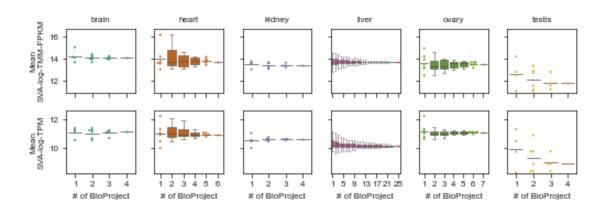


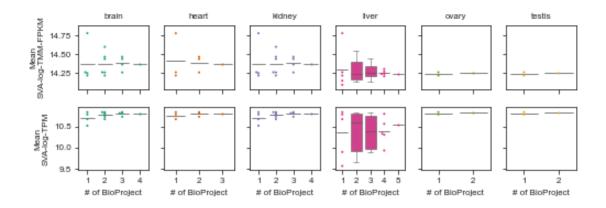












```
[34]: import gseapy
      import mygene
      #library_names = gseapy.get_library_name()
      library_names = [
          'KEGG_2019_Human',
          'KEGG_2019_Mouse',
      ]
      bidirectionals = [True,False]
      pcm_prefixes = ['llou_fpkm_','llou_tpm_']
      min_taus = [0,0.5]
      mg = mygene.MyGeneInfo()
      taxids = ','.join(b.taxid.unique().astype(str))
      new_wd = os.path.join(wd+'GO_specificity_shift')
      if not os.path.exists(new_wd):
          os.mkdir(new wd)
      os.chdir(new_wd)
      b_leaf = b.loc[(b['so_event']=='L'),:]
      for pp,bidirectional,min_tau in itertools.product(pcm_prefixes, bidirectionals,u
       →min_taus):
          organ1s = b['parent_'+pp+'max_organ'].dropna().unique()
          organ2s = b[pp+'max_organ'].dropna().unique()
          events = ['All','S','D','R']
          conditions = True
```

```
conditions = conditions&(b['llou_intersect_is_shift']==1)
   conditions = conditions&(b['spnode_coverage']!='root')
   conditions = conditions&(b[pp+'tau']>=min_tau)
   conditions = conditions&(b['parent_'+pp+'tau']>=min_tau)
  b2 = b.loc[conditions,:].reset_index()
   if bidirectional:
       organ_combinat = list(itertools.combinations(set(organ1s).
→union(set(organ2s)), 2))
       iters = [[ev,]+list(o) for ev in events for o in organ_combinat ]
  else:
       iters = itertools.product(events, organ1s, organ2s)
  for ev,organ1,organ2 in iters:
       if organ1==organ2:
           continue
       elif organ1>organ2:
           organ1prev = organ1
           organ2prev = organ2
           organ1 = organ2prev
           organ2 = organ1prev
       conditions = True
       if ev!='All':
           conditions = conditions&(b2['branch_category']==ev)
       if bidirectional:
           is_organ_combinat1 =__

→ (b2['parent_'+pp+'max_organ']==organ1)&(b2[pp+'max_organ']==organ2)
           is_organ_combinat2 =__
→(b2['parent_'+pp+'max_organ']==organ2)&(b2[pp+'max_organ']==organ1)
           conditions = conditions&(is_organ_combinat1|is_organ_combinat2)
       else:
           conditions = conditions&(b2['parent_'+pp+'max_organ']==organ1)
           conditions = conditions&(b2[pp+'max_organ']==organ2)
       b3 = b2.loc[conditions,['orthogroup',pp+'regime']].
→reset_index(drop=True)
       if b3.shape[0] == 0:
           continue
       gene ids = pandas.DataFrame()
       for i in b3.index:
           if i%1000==0:
              print(i)
           regime = b3.loc[i,pp+'regime']
           og = b3.loc[i,'orthogroup']
           tmp_gene_ids = b_leaf.
→loc[(b_leaf[pp+'regime']==regime)&(b_leaf['orthogroup']==og),['orthogroup','node_name']]
           gene_ids = pandas.concat([gene_ids,tmp_gene_ids], sort=False,__
→ignore_index=True)
```

```
gene_ids = gene_ids.drop_duplicates().

→sort_values(by=['orthogroup', 'node_name'])
       for sp in ['Homo_sapiens','Mus_musculus']:
           outdir=pp+sp+'_minTau'+str(min_tau)+'_'+organ1+'_'+organ2+'_'+ev
           if bidirectional:
               outdir = 'bidirectional '+outdir
           df_sp = gene_ids.loc[(gene_ids['node_name'].str.startswith(sp)),:]
           if df_sp.shape[0] == 0:
               continue
           df_sp.loc[:,'node_name'] = df_sp.loc[:,'node_name'].replace('.
→*_','', regex=True)
           df sp.columns = ['orthogroup', 'gene id']
           if not os.path.exists(outdir):
               os.mkdir(outdir)
           df_sp.to_csv(os.path.join(outdir, outdir+'.geneid.tsv'), sep='\t',u
→index=False)
           sp_gene_ids = df_sp.loc[:,'gene_id'].tolist()
           b_mygene = mg.querymany(sp_gene_ids, scopes='ensembl.gene',_
→fields='symbol', species=taxids, as_dataframe=True)
           if b mygene.columns[0] == 'notfound':
               b_mygene = pandas.DataFrame()
           if b mygene.shape[0]==0:
               continue
           for ln in library_names:
               if (sp=='Homo_sapiens')&('Mouse' in ln):
                   continue
               if (sp=='Mus_musculus')&('Human' in ln):
                   continue
               txt = '{} min_tau={} {} {}-{} {}: #shift={}, #leaf={},_
→#queryGene={}, {}'
               if bidirectional:
                   txt = txt+' bidirectional'
               else:
                   txt = txt+' unidirectional'
               txt = txt.format(pp, min_tau, ev,organ1,organ2,sp,b3.
→shape[0],len(gene_ids),len(sp_gene_ids),ln)
               if os.path.isfile(outdir+'/'+ln+'.'+outdir+'.enrichr.reports.
→txt'):
                   print('Skipped.', txt)
                   continue
               print(txt)
               try:
                   out = gseapy.enrichr(gene_list=b_mygene['symbol'].
→astype(str).tolist(), description=outdir, gene_sets=ln, outdir=outdir,
\rightarrowcutoff=0.05)
               except:
```

```
print('Error and retry.')
                     try:
                         out = gseapy.enrichr(gene_list=b_mygene['symbol'].
 →astype(str).tolist(), description=outdir, gene_sets=ln, outdir=outdir,
 \rightarrowcutoff=0.05)
                     except:
                         time.sleep(3)
                         print('The retry failed. Skipped.')
        print('')
print('Done!')
del b_leaf
os.chdir(wd)
0
/Users/kef74yk/anaconda3/lib/python3.6/site-
packages/pandas/core/indexing.py:1048: SettingWithCopyWarning:
A value is trying to be set on a copy of a slice from a DataFrame.
Try using .loc[row_indexer,col_indexer] = value instead
See the caveats in the documentation: https://pandas.pydata.org/pandas-
docs/stable/user_guide/indexing.html#returning-a-view-versus-a-copy
  self.obj[item_labels[indexer[info_axis]]] = value
querying 1-140...done.
Finished.
/Users/kef74yk/anaconda3/lib/python3.6/site-
packages/biothings_client/base.py:143: FutureWarning:
pandas.io.json.json_normalize is deprecated, use pandas.json_normalize instead
  df = json_normalize(obj)
140 input query terms found no hit:
        ['ENSG00000204577', 'ENSG00000244482', 'ENSG00000273991',
'ENSG00000274587', 'ENSG00000275290', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-107...done.
Finished.
107 input query terms found no hit:
        ['ENSMUSG00000089942', 'ENSMUSG0000095088', 'ENSMUSG00000073968',
'ENSMUSG0000048076', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-86...done.
```

```
Finished.
86 input query terms found no hit:
        ['ENSG00000211947', 'ENSG00000211966', 'ENSG00000255374',
'ENSG00000255837', 'ENSG00000273092', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-83...done.
Finished.
83 input query terms found no hit:
        ['ENSMUSG00000076823', 'ENSMUSG00000076839', 'ENSMUSG00000076846',
'ENSMUSG0000076858', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-229...done.
Finished.
229 input query terms found no hit:
        ['ENSG00000276033', 'ENSG00000276848', 'ENSG00000240764',
'ENSG00000172466', 'ENSG00000275528', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-181...done.
Finished.
181 input query terms found no hit:
        ['ENSMUSG00000057439', 'ENSMUSG00000051469', 'ENSMUSG00000020676',
'ENSMUSG00000045534', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-110...done.
Finished.
110 input query terms found no hit:
        ['ENSG00000104970', 'ENSG00000180573', 'ENSG00000211788',
'ENSG00000075624', 'ENSG00000163017', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-95...done.
Finished.
95 input query terms found no hit:
        ['ENSMUSG00000094420', 'ENSMUSG00000058818', 'ENSMUSG00000093969',
'ENSMUSG00000096106', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
```

querying 1-392...done.

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Finished.
392 input query terms found no hit:
        ['ENSG00000125498', 'ENSG00000167633', 'ENSG00000240403',
'ENSG00000242019', 'ENSG00000243772', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-152...done.
Finished.
152 input query terms found no hit:
        ['ENSMUSG00000073913', 'ENSMUSG0000078808', 'ENSMUSG00000115644',
'ENSMUSG0000047246', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-138...done.
Finished.
138 input query terms found no hit:
        ['ENSG00000211959', 'ENSG00000259261', 'ENSG00000204642',
'ENSG00000206452', 'ENSG00000225691', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-117...done.
Finished.
117 input query terms found no hit:
        ['ENSMUSG00000053338', 'ENSMUSG00000025479', 'ENSMUSG00000040583',
'ENSMUSG00000040650', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-174...done.
Finished.
174 input query terms found no hit:
        ['ENSG00000277177', 'ENSG00000206435', 'ENSG00000228299',
'ENSG00000203747', 'ENSG00000198211', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-112...done.
Finished.
112 input query terms found no hit:
        ['ENSMUSG00000073406', 'ENSMUSG00000020440', 'ENSMUSG00000021877',
'ENSMUSG00000051853', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-136...done.
```

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Finished.
136 input query terms found no hit:
        ['ENSG00000233999', 'ENSG00000239819', 'ENSG00000242580',
'ENSG00000243063', 'ENSG00000282310', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-125...done.
Finished.
125 input query terms found no hit:
        ['ENSMUSG00000074521', 'ENSMUSG00000078889', 'ENSMUSG00000078901',
'ENSMUSG0000043091', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
1000
querying 1-186...done.
Finished.
186 input query terms found no hit:
        ['ENSG00000273884', 'ENSG00000274311', 'ENSG00000274513',
'ENSG00000277317', 'ENSG00000277398', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-196...done.
Finished.
196 input query terms found no hit:
        ['ENSMUSG00000056782', 'ENSMUSG00000066263', 'ENSMUSG00000066269',
'ENSMUSG00000094520', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-146...done.
Finished.
146 input query terms found no hit:
        ['ENSG00000276798', 'ENSG00000127362', 'ENSG00000256436',
'ENSG00000263097', 'ENSG00000282612', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-129...done.
Finished.
129 input query terms found no hit:
        ['ENSMUSG00000076778', 'ENSMUSG00000074955', 'ENSMUSG00000007097',
'ENSMUSG00000033161', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
```

```
querying 1-187...done.
Finished.
187 input query terms found no hit:
        ['ENSG00000211957', 'ENSG00000282045', 'ENSG00000282211',
'ENSG00000282305', 'ENSG00000211599', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
querying 1-164...done.
Finished.
164 input query terms found no hit:
        ['ENSMUSG00000073028', 'ENSMUSG00000076500', 'ENSMUSG00000076501',
'ENSMUSG0000076505', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-117...done.
Finished.
117 input query terms found no hit:
        ['ENSG00000184698', 'ENSG00000273539', 'ENSG00000278042',
'ENSG00000282425', 'ENSG00000282651', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-135...done.
Finished.
135 input query terms found no hit:
        ['ENSMUSG00000050085', 'ENSMUSG00000058200', 'ENSMUSG00000094531',
'ENSMUSG00000096773', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-158...done.
Finished.
158 input query terms found no hit:
        ['ENSG00000005001', 'ENSG00000282937', 'ENSG00000127366',
'ENSG00000276541', 'ENSG00000165527', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-120...done.
Finished.
120 input query terms found no hit:
        ['ENSMUSG00000044147', 'ENSMUSG00000049758', 'ENSMUSG00000074946',
'ENSMUSG0000109528', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
```

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1000
querying 1-235...done.
Finished.
235 input query terms found no hit:
        ['ENSG00000276114', 'ENSG00000227739', 'ENSG00000151079',
'ENSG00000169432', 'ENSG00000188613', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-259...done.
Finished.
259 input query terms found no hit:
        ['ENSMUSG00000094076', 'ENSMUSG00000043366', 'ENSMUSG00000073973',
'ENSMUSG00000094822', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
0
1000
querying 1-262...done.
Finished.
262 input query terms found no hit:
        ['ENSG00000276590', 'ENSG00000124635', 'ENSG00000197903',
'ENSG00000184260', 'ENSG00000170465', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-199...done.
Finished.
199 input query terms found no hit:
        ['ENSMUSG00000043948', 'ENSMUSG00000073962', 'ENSMUSG00000046932',
'ENSMUSG00000095632', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-83...done.
Finished.
83 input query terms found no hit:
        ['ENSG00000179144', 'ENSG00000090659', 'ENSG00000143226',
'ENSG00000088881', 'ENSG00000094963', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-66...done.
Finished.
66 input query terms found no hit:
        ['ENSMUSG00000062421', 'ENSMUSG00000078606', 'ENSMUSG00000017756',
'ENSMUSG00000063694', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
```

terms.

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0
querying 1-53...done.
Finished.
53 input query terms found no hit:
        ['ENSG00000108691', 'ENSG00000213719', 'ENSG00000223639',
'ENSG00000226248', 'ENSG00000226417', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-55...done.
Finished.
55 input query terms found no hit:
        ['ENSMUSG00000076823', 'ENSMUSG00000076839', 'ENSMUSG00000076846',
'ENSMUSG00000094562', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-120...done.
Finished.
120 input query terms found no hit:
        ['ENSG00000240764', 'ENSG00000130037', 'ENSG00000114279',
'ENSG00000120049', 'ENSG00000101144', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-105...done.
Finished.
105 input query terms found no hit:
        ['ENSMUSG00000020676', 'ENSMUSG00000045534', 'ENSMUSG00000090841',
'ENSMUSG00000025221', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-73...done.
Finished.
73 input query terms found no hit:
        ['ENSG00000180573', 'ENSG00000213139', 'ENSG00000257529',
'ENSG00000188778', 'ENSG00000160447', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-66...done.
Finished.
66 input query terms found no hit:
        ['ENSMUSG00000037820', 'ENSMUSG00000042842', 'ENSMUSG00000043613',
'ENSMUSG00000021702', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
```

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0
querying 1-328...done.
Finished.
328 input query terms found no hit:
        ['ENSG00000125498', 'ENSG00000167633', 'ENSG00000240403',
'ENSG00000242019', 'ENSG00000243772', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-72...done.
Finished.
72 input query terms found no hit:
        ['ENSMUSG00000073913', 'ENSMUSG00000031085', 'ENSMUSG00000040752',
'ENSMUSG00000053093', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-83...done.
Finished.
83 input query terms found no hit:
        ['ENSG00000224305', 'ENSG00000225824', 'ENSG00000226165',
'ENSG00000228254', 'ENSG00000228813', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-72...done.
Finished.
72 input query terms found no hit:
        ['ENSMUSG00000053338', 'ENSMUSG00000025479', 'ENSMUSG00000023122',
'ENSMUSG0000071866', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-94...done.
Finished.
94 input query terms found no hit:
        ['ENSG00000198211', 'ENSG00000215048', 'ENSG00000229295',
'ENSG00000230708', 'ENSG00000010438', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-65...done.
Finished.
65 input query terms found no hit:
        ['ENSMUSG00000108827', 'ENSMUSG00000049680', 'ENSMUSG00000041453',
'ENSMUSG00000037419', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
```

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0
querying 1-87...done.
Finished.
87 input query terms found no hit:
        ['ENSG00000246705', 'ENSG00000105388', 'ENSG00000090382',
'ENSG00000215472', 'ENSG00000211792', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-75...done.
Finished.
75 input query terms found no hit:
        ['ENSMUSG00000058260', 'ENSMUSG00000069516', 'ENSMUSG00000062328',
'ENSMUSG00000090451', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-76...done.
Finished.
76 input query terms found no hit:
        ['ENSG00000230413', 'ENSG00000233095', 'ENSG00000235346',
'ENSG00000235680', 'ENSG00000237216', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-79...done.
Finished.
79 input query terms found no hit:
        ['ENSMUSG00000056782', 'ENSMUSG0000066263', 'ENSMUSG0000066269',
'ENSMUSG0000066850', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-75...done.
Finished.
75 input query terms found no hit:
        ['ENSG00000127362', 'ENSG00000070831', 'ENSG00000136531',
'ENSG00000163399', 'ENSG00000157388', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-80...done.
Finished.
80 input query terms found no hit:
        ['ENSMUSG00000074955', 'ENSMUSG00000033161', 'ENSMUSG00000019302',
'ENSMUSG00000033295', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
```

terms.

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0
querying 1-131...done.
Finished.
131 input query terms found no hit:
        ['ENSG00000211599', 'ENSG00000211611', 'ENSG00000211623',
'ENSG00000211625', 'ENSG00000211626', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-130...done.
Finished.
130 input query terms found no hit:
        ['ENSMUSG00000073028', 'ENSMUSG00000076500', 'ENSMUSG00000076501',
'ENSMUSG00000076505', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-59...done.
Finished.
59 input query terms found no hit:
        ['ENSG00000184698', 'ENSG00000169777', 'ENSG00000115386',
'ENSG00000117335', 'ENSG00000212657', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-72...done.
Finished.
72 input query terms found no hit:
        ['ENSMUSG00000050085', 'ENSMUSG00000058200', 'ENSMUSG00000094531',
'ENSMUSG00000096773', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-94...done.
Finished.
94 input query terms found no hit:
        ['ENSG00000127366', 'ENSG00000186767', 'ENSG00000139675',
'ENSG00000156508', 'ENSG00000211643', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-78...done.
Finished.
78 input query terms found no hit:
        ['ENSMUSG00000074946', 'ENSMUSG00000109528', 'ENSMUSG00000037742',
'ENSMUSG00000027669', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
```

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0
querying 1-138...done.
Finished.
138 input query terms found no hit:
        ['ENSG00000151079', 'ENSG00000169432', 'ENSG00000188613',
'ENSG00000132681', 'ENSG00000155511', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-149...done.
Finished.
149 input query terms found no hit:
        ['ENSMUSG00000043366', 'ENSMUSG00000073973', 'ENSMUSG00000094822',
'ENSMUSG00000064259', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-171...done.
Finished.
171 input query terms found no hit:
        ['ENSG00000184260', 'ENSG00000170465', 'ENSG00000170477',
'ENSG00000185479', 'ENSG00000205420', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-135...done.
Finished.
135 input query terms found no hit:
        ['ENSMUSG00000043948', 'ENSMUSG00000073962', 'ENSMUSG00000037737',
'ENSMUSG00000059430', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-57...done.
Finished.
57 input query terms found no hit:
        ['ENSG00000204577', 'ENSG00000244482', 'ENSG00000273991',
'ENSG00000274587', 'ENSG00000275290', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-37...done.
Finished.
37 input query terms found no hit:
        ['ENSMUSG00000089942', 'ENSMUSG00000095088', 'ENSMUSG00000073968',
'ENSMUSG00000048076', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
```

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0
querying 1-33...done.
Finished.
33 input query terms found no hit:
        ['ENSG00000211947', 'ENSG00000211966', 'ENSG00000255374',
'ENSG00000255837', 'ENSG00000273092', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-28...done.
Finished.
28 input query terms found no hit:
        ['ENSMUSG00000076858', 'ENSMUSG00000096908', 'ENSMUSG00000095074',
'ENSMUSG0000001847', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-108...done.
Finished.
108 input query terms found no hit:
        ['ENSG00000276033', 'ENSG00000276848', 'ENSG00000172466',
'ENSG00000275528', 'ENSG00000223865', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-70...done.
Finished.
70 input query terms found no hit:
        ['ENSMUSG00000057439', 'ENSMUSG00000051469', 'ENSMUSG00000022949',
'ENSMUSG0000109033', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-35...done.
Finished.
35 input query terms found no hit:
        ['ENSG00000104970', 'ENSG00000211788', 'ENSG00000075624',
'ENSG00000163017', 'ENSG00000184009', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-27...done.
Finished.
27 input query terms found no hit:
        ['ENSMUSG00000094420', 'ENSMUSG00000058818', 'ENSMUSG00000093969',
'ENSMUSG00000096106', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
```

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0
querying 1-59...done.
Finished.
59 input query terms found no hit:
        ['ENSG00000158373', 'ENSG00000248496', 'ENSG00000026950',
'ENSG00000111801', 'ENSG00000154370', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-60...done.
Finished.
60 input query terms found no hit:
        ['ENSMUSG00000078808', 'ENSMUSG00000115644', 'ENSMUSG00000076760',
'ENSMUSG00000060981', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-54...done.
Finished.
54 input query terms found no hit:
        ['ENSG00000211959', 'ENSG00000259261', 'ENSG00000204642',
'ENSG00000206452', 'ENSG00000225691', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-43...done.
Finished.
43 input query terms found no hit:
        ['ENSMUSG00000040583', 'ENSMUSG00000040650', 'ENSMUSG00000040660',
'ENSMUSG0000066704', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-77...done.
Finished.
77 input query terms found no hit:
        ['ENSG00000277177', 'ENSG00000206435', 'ENSG00000228299',
'ENSG00000203747', 'ENSG00000004059', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-44...done.
Finished.
44 input query terms found no hit:
        ['ENSMUSG00000073406', 'ENSMUSG00000020440', 'ENSMUSG00000021877',
'ENSMUSG00000051853', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
```

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querying 1-49...done.
Finished.
49 input query terms found no hit:
        ['ENSG00000233999', 'ENSG00000239819', 'ENSG00000242580',
'ENSG00000243063', 'ENSG00000282310', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-45...done.
Finished.
45 input query terms found no hit:
        ['ENSMUSG00000074521', 'ENSMUSG00000078889', 'ENSMUSG00000078901',
'ENSMUSG0000043091', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-101...done.
Finished.
101 input query terms found no hit:
        ['ENSG00000273884', 'ENSG00000274311', 'ENSG00000274513',
'ENSG00000277317', 'ENSG00000277398', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-104...done.
Finished.
104 input query terms found no hit:
        ['ENSMUSG00000094520', 'ENSMUSG0000060024', 'ENSMUSG00000061829',
'ENSMUSG00000094898', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-70...done.
Finished.
70 input query terms found no hit:
        ['ENSG00000276798', 'ENSG00000256436', 'ENSG00000263097',
'ENSG00000282612', 'ENSG00000206302', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-48...done.
Finished.
48 input query terms found no hit:
        ['ENSMUSG00000076778', 'ENSMUSG00000007097', 'ENSMUSG00000028664',
'ENSMUSG00000032537', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
```

```
querying 1-54...done.
Finished.
54 input query terms found no hit:
        ['ENSG00000211957', 'ENSG00000282045', 'ENSG00000282211',
'ENSG00000282305', 'ENSG00000168148', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-28...done.
Finished.
28 input query terms found no hit:
        ['ENSMUSG00000078851', 'ENSMUSG00000069265', 'ENSMUSG00000069267',
'ENSMUSG00000069273', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-54...done.
Finished.
54 input query terms found no hit:
        ['ENSG00000273539', 'ENSG00000278042', 'ENSG00000282425',
'ENSG00000282651', 'ENSG00000211786', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-56...done.
Finished.
56 input query terms found no hit:
        ['ENSMUSG00000061296', 'ENSMUSG00000115170', 'ENSMUSG00000115253',
'ENSMUSG00000030196', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-62...done.
Finished.
62 input query terms found no hit:
        ['ENSG0000005001', 'ENSG00000282937', 'ENSG00000276541',
'ENSG00000165527', 'ENSG00000163221', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-40...done.
Finished.
40 input query terms found no hit:
        ['ENSMUSG00000044147', 'ENSMUSG00000049758', 'ENSMUSG0000001020',
'ENSMUSG00000033208', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
```

```
querying 1-94...done.
Finished.
94 input query terms found no hit:
        ['ENSG00000276114', 'ENSG00000227739', 'ENSG00000206290',
'ENSG00000138758', 'ENSG00000164402', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-105...done.
Finished.
105 input query terms found no hit:
        ['ENSMUSG00000094076', 'ENSMUSG00000096516', 'ENSMUSG00000091652',
'ENSMUSG00000094553', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-89...done.
Finished.
89 input query terms found no hit:
        ['ENSG00000276590', 'ENSG00000124635', 'ENSG00000197903',
'ENSG00000183785', 'ENSG00000198033', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-60...done.
Finished.
60 input query terms found no hit:
        ['ENSMUSG00000046932', 'ENSMUSG00000115404', 'ENSMUSG00000090581',
'ENSMUSG00000095730', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-4...done.
Finished.
4 input query terms found no hit:
        ['ENSMUSG00000110469', 'ENSMUSG00000078153', 'ENSMUSG00000060019',
'ENSMUSG00000073640']
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
0
querying 1-5...done.
Finished.
5 input query terms found no hit:
```

```
['ENSMUSG00000092086', 'ENSMUSG0000096156', 'ENSMUSG00000113255',
'ENSMUSG00000096486', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-1...done.
Finished.
1 input query terms found no hit:
        ['ENSG0000125817']
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-2...done.
Finished.
2 input query terms found no hit:
        ['ENSMUSG00000068267', 'ENSMUSG00000069622']
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-5...done.
Finished.
5 input query terms found no hit:
        ['ENSG00000180138', 'ENSG00000170950', 'ENSG00000163114',
'ENSG00000189401', 'ENSG00000212643']
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-17...done.
Finished.
17 input query terms found no hit:
        ['ENSMUSG00000059343', 'ENSMUSG00000063129', 'ENSMUSG00000073730',
'ENSMUSG00000078154', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-2...done.
Finished.
2 input query terms found no hit:
        ['ENSMUSG00000044424', 'ENSMUSG00000069324']
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-3...done.
Finished.
3 input query terms found no hit:
        ['ENSG00000226784', 'ENSG00000174599', 'ENSG00000188021']
```

```
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-3...done.
Finished.
3 input query terms found no hit:
        ['ENSMUSG00000044528', 'ENSMUSG00000050148', 'ENSMUSG00000061619']
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-1...done.
Finished.
1 input query terms found no hit:
        ['ENSMUSG00000048040']
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-9...done.
Finished.
9 input query terms found no hit:
        ['ENSG00000177688', 'ENSG00000165496', 'ENSG00000276380',
'ENSG00000112273', 'ENSG00000253626', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-13...done.
Finished.
13 input query terms found no hit:
        ['ENSMUSG00000046173', 'ENSMUSG00000051732', 'ENSMUSG00000060499',
'ENSMUSG00000098559', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-1...done.
Finished.
1 input query terms found no hit:
        ['ENSG0000178597']
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-1...done.
Finished.
1 input query terms found no hit:
        ['ENSMUSG00000043430']
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
```

```
querying 1-1...done.
Finished.
1 input query terms found no hit:
        ['ENSG00000211675']
Pass "returnall=True" to return complete lists of duplicate or missing query
querying 1-1...done.
Finished.
1 input query terms found no hit:
        ['ENSMUSG0000067608']
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-3...done.
Finished.
3 input query terms found no hit:
        ['ENSG00000229937', 'ENSG00000120329', 'ENSG00000189134']
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-5...done.
Finished.
5 input query terms found no hit:
        ['ENSMUSG00000100296', 'ENSMUSG00000036463', 'ENSMUSG00000092305',
\verb|'ENSMUSG0000046717', | \verb|'ENSMUSG000||
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-1...done.
Finished.
1 input query terms found no hit:
        ['ENSG0000188042']
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-1...done.
Finished.
1 input query terms found no hit:
        ['ENSMUSG00000049866']
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-2...done.
Finished.
2 input query terms found no hit:
        ['ENSG00000177144', 'ENSG00000137040']
Pass "returnall=True" to return complete lists of duplicate or missing query
```

```
terms.
querying 1-5...done.
Finished.
5 input query terms found no hit:
        ['ENSMUSG00000099787', 'ENSMUSG0000064063', 'ENSMUSG00000074909',
'ENSMUSG00000051255', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
0
querying 1-2...done.
Finished.
2 input query terms found no hit:
        ['ENSG00000228075', 'ENSG00000250254']
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-2...done.
Finished.
2 input query terms found no hit:
        ['ENSMUSG00000054117', 'ENSMUSG00000112039']
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-25...done.
Finished.
25 input query terms found no hit:
        ['ENSG00000090659', 'ENSG00000143226', 'ENSG00000231555',
'ENSG00000094963', 'ENSG00000169908', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-25...done.
Finished.
25 input query terms found no hit:
        ['ENSMUSG00000089942', 'ENSMUSG00000110469', 'ENSMUSG00000090877',
'ENSMUSG00000074179', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-13...done.
Finished.
13 input query terms found no hit:
        ['ENSG00000100170', 'ENSG00000223609', 'ENSG00000102048',
'ENSG00000148180', 'ENSG00000109062', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
```

querying 1-11...done.

```
Finished.
11 input query terms found no hit:
        ['ENSMUSG00000031384', 'ENSMUSG0000002900', 'ENSMUSG00000052911',
'ENSMUSG00000027219', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-79...done.
Finished.
79 input query terms found no hit:
        ['ENSG00000223865', 'ENSG00000226826', 'ENSG00000237710',
'ENSG00000130037', 'ENSG00000159212', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-68...done.
Finished.
68 input query terms found no hit:
        ['ENSMUSG00000045534', 'ENSMUSG00000022949', 'ENSMUSG00000028773',
'ENSMUSG00000025221', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-16...done.
Finished.
16 input query terms found no hit:
        ['ENSG00000183801', 'ENSG00000106483', 'ENSG00000274194',
'ENSG00000277025', 'ENSG00000277733', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-13...done.
Finished.
13 input query terms found no hit:
        ['ENSMUSG00000037820', 'ENSMUSG00000011257', 'ENSMUSG00000043613',
'ENSMUSG0000010830', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-28...done.
Finished.
28 input query terms found no hit:
        ['ENSG00000248496', 'ENSG00000105679', 'ENSG00000206383',
'ENSG00000236251', 'ENSG00000006047', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-39...done.
```

```
Finished.
39 input query terms found no hit:
        ['ENSMUSG00000031085', 'ENSMUSG00000079271', 'ENSMUSG00000096793',
'ENSMUSG00000101653', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-32...done.
Finished.
32 input query terms found no hit:
        ['ENSG00000197838', 'ENSG00000231939', 'ENSG00000167755',
'ENSG00000005471', 'ENSG00000073734', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-54...done.
Finished.
54 input query terms found no hit:
        ['ENSMUSG00000053338', 'ENSMUSG00000025479', 'ENSMUSG00000040583',
'ENSMUSG00000040650', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-61...done.
Finished.
61 input query terms found no hit:
        ['ENSG00000203747', 'ENSG00000134287', 'ENSG00000230763',
'ENSG00000236693', 'ENSG00000010438', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-35...done.
Finished.
35 input query terms found no hit:
        ['ENSMUSG00000073406', 'ENSMUSG00000051853', 'ENSMUSG00000071517',
'ENSMUSG00000028645', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-24...done.
Finished.
24 input query terms found no hit:
        ['ENSG00000276192', 'ENSG00000174156', 'ENSG00000136689',
'ENSG00000121552', 'ENSG00000197142', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-34...done.
```

```
Finished.
34 input query terms found no hit:
        ['ENSMUSG00000074521', 'ENSMUSG0000078889', 'ENSMUSG00000078901',
'ENSMUSG00000058260', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-65...done.
Finished.
65 input query terms found no hit:
        ['ENSG00000277317', 'ENSG00000139648', 'ENSG00000167768',
'ENSG00000170454', 'ENSG00000170484', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-73...done.
Finished.
73 input query terms found no hit:
        ['ENSMUSG00000022986', 'ENSMUSG00000023041', 'ENSMUSG00000046834',
'ENSMUSG00000048699', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-41...done.
Finished.
41 input query terms found no hit:
        ['ENSG00000276798', 'ENSG00000206302', 'ENSG00000136531',
'ENSG00000018625', 'ENSG00000105409', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-37...done.
Finished.
37 input query terms found no hit:
        ['ENSMUSG00000076778', 'ENSMUSG0000007097', 'ENSMUSG00000033161',
\verb|'ENSMUSG00000024597', | \verb|'ENSMUSG000||
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-17...done.
Finished.
17 input query terms found no hit:
        ['ENSG00000052344', 'ENSG00000175793', 'ENSG00000211675',
'ENSG00000229077', 'ENSG00000232180', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-12...done.
```

```
Finished.
12 input query terms found no hit:
        ['ENSMUSG00000024727', 'ENSMUSG00000012187', 'ENSMUSG00000021263',
'ENSMUSG00000030498', 'ENSMUSG000
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-37...done.
Finished.
37 input query terms found no hit:
        ['ENSG00000211786', 'ENSG00000115386', 'ENSG00000279804',
'ENSG00000282212', 'ENSG00000179344', 'ENS
Pass "returnall=True" to return complete lists of duplicate or missing query
terms.
querying 1-49...
       HTTPError
                                                 Traceback (most recent call
 →last)
       <ipython-input-34-0fd6712b8370> in <module>
                       df_sp.to_csv(os.path.join(outdir, outdir+'.geneid.tsv'),__
 →sep='\t', index=False)
                       sp_gene_ids = df_sp.loc[:,'gene_id'].tolist()
        85
   ---> 86
                       b_mygene = mg.querymany(sp_gene_ids, scopes='ensembl.
 →gene', fields='symbol', species=taxids, as_dataframe=True)
        87
                       if b_mygene.columns[0] == 'notfound':
        88
                           b_mygene = pandas.DataFrame()
        ~/anaconda3/lib/python3.6/site-packages/biothings_client/base.py in_
 535
                   li_query = []
       536
                   def query_fn(qterms): return self._querymany_inner(qterms,__
 →verbose=verbose, **kwargs)
   --> 537
                   for hits in self._repeated_query(query_fn, qterms,_
 →verbose=verbose):
       538
                       if return_raw:
       539
                           out.append(hits) # hits is the raw response text
        ~/anaconda3/lib/python3.6/site-packages/biothings_client/base.py in_
 →_repeated_query(self, query_fn, query_li, verbose, **fn_kwargs)
```

```
print("querying {0}-{1}...".format(i + 1, cnt), end="")
      219
      220
                      i = cnt
  --> 221
                      from_cache, query_result = query_fn(batch, **fn_kwargs)
      222
                      yield query_result
                      if verbose:
      223
      ~/anaconda3/lib/python3.6/site-packages/biothings_client/base.py in_u

¬query_fn(qterms)
      534
                  li_dup = []
      535
                  li query = []
  --> 536
                  def query_fn(qterms): return self._querymany_inner(qterms,_
→verbose=verbose, **kwargs)
      537
                  for hits in self._repeated_query(query_fn, qterms,_
→verbose=verbose):
      538
                      if return_raw:
      ~/anaconda3/lib/python3.6/site-packages/biothings_client/base.py in_u
481
                  _kwargs.update(kwargs)
      482
                  _url = self.url + self._query_endpoint
  --> 483
                  return self._post(_url, params=_kwargs, verbose=verbose)
      484
              def _querymany(self, qterms, scopes=None, **kwargs):
      485
      ~/anaconda3/lib/python3.6/site-packages/biothings_client/base.py in_u
→_post(self, url, params, verbose)
      175
                  if self.raise for status:
      176
                      # raise requests.exceptions.HTTPError if not 200
  --> 177
                      res.raise_for_status()
                  if return raw:
      178
      179
                      return from_cache, res
      ~/anaconda3/lib/python3.6/site-packages/requests/models.py in ⊔
→raise_for_status(self)
      939
      940
                  if http_error_msg:
  --> 941
                      raise HTTPError(http_error_msg, response=self)
      942
      943
              def close(self):
      HTTPError: 502 Server Error: Bad Gateway for url: http://mygene.info/v3/
→query/
```

```
[35]: | #method = ['pvalue', 0.05]
     method = ['top', 10]
     new_wd = os.path.join(wd+'GO_specificity_shift')
     if ('b' in vars()):
         organ1s = b['parent_'+pp+'max_organ'].dropna().unique()
         organ2s = b[pp+'max_organ'].dropna().unique()
     else:
         organs = ['brain','heart','kidney','liver','ovary','testis']
         organ1s = organs
         organ2s = organs
     events = ['All','S','D','R']
     for ln in library_names:
         for sp in ['Homo_sapiens','Mus_musculus']:
             for pp,bidirectional,min_tau in itertools.product(pcm_prefixes,_
      →bidirectionals, min_taus):
                 if bidirectional:
                     summary_wd = os.path.join(new_wd,__
      else:
                     summary wd = os.path.join(new wd,___

¬'summary_unidirectional_'+method[0]+str(method[1]))
                 if not os.path.exists(summary_wd):
                     os.mkdir(summary_wd)
                 outfile =
      → 'summary_'+ln+'_'+pp+sp+'_minTau'+str(min_tau)+'_'+method[0]+str(method[1])+'.
      ⇔tsv'
                 print(outfile)
                 dfln = pandas.DataFrame()
                 if pp=='l1ou_fpkm_':
                     expression_metrics = 'SVA-log-TMM-FPKM'
                 elif pp=='l1ou_tpm_':
                     expression_metrics = 'SVA-log-TPM'
                 if bidirectional:
                     organ_combinat = list(itertools.combinations(set(organ1s).
      →union(set(organ2s)), 2))
                     iters = [ [ev,]+list(o) for ev in events for o in_
      →organ_combinat ]
                 else:
                     iters = itertools.product(events, organ1s, organ2s)
                 for ev,organ1,organ2 in iters:
                     if organ1==organ2:
                         continue
```

```
elif organ1>organ2:
                   organ1prev = organ1
                   organ2prev = organ2
                   organ1 = organ2prev
                   organ2 = organ1prev
               outdir=pp+sp+'_minTau'+str(min_tau)+'_'+organ1+'_'+organ2+'_'+ev
               if bidirectional:
                   outdir = 'bidirectional '+outdir
               ln file = os.path.join(new wd, outdir+'/'+ln+'.'+outdir+'.
⇔enrichr.reports.txt')
               if os.path.isfile(ln_file):
                   tmp = pandas.read_csv(ln_file, sep='\t', header=0)
                   if method[0] == 'pvalue':
                       tmp_sig = tmp.loc[(tmp['Adjusted P-value'] < method[1]),:]</pre>
                   elif method[0] == 'top':
                       tmp_sig = tmp.sort_values(by='Combined Score',__
→ascending=False).iloc[0:method[1],:]
                   if tmp_sig.shape[0]!=0:
                       tmp_sig.loc[:,'branch_category'] = ev
                       tmp_sig.loc[:,'organ1'] = organ1
                       tmp_sig.loc[:,'organ2'] = organ2
                       dfln = pandas.concat([dfln, tmp_sig],__
→ignore_index=True, sort=False)
           dfln.to_csv(os.path.join(summary_wd, outfile), sep='\t',_
→index=False)
```

summary KEGG 2019 Human l1ou fpkm Homo sapiens minTau0 top10.tsv summary_KEGG_2019_Human_l1ou_fpkm_Homo_sapiens_minTau0.5_top10.tsv summary_KEGG_2019_Human_l1ou_fpkm_Homo_sapiens_minTau0_top10.tsv summary_KEGG_2019_Human_l1ou_fpkm_Homo_sapiens_minTau0.5_top10.tsv summary_KEGG_2019_Human_l1ou_tpm_Homo_sapiens_minTau0_top10.tsv summary_KEGG_2019_Human_l1ou_tpm_Homo_sapiens_minTau0.5_top10.tsv summary_KEGG_2019_Human_l1ou_tpm_Homo_sapiens_minTau0_top10.tsv summary KEGG 2019 Human llou tpm Homo sapiens minTau0.5 top10.tsv summary KEGG 2019 Human llou fpkm Mus musculus minTau0 top10.tsv summary KEGG 2019 Human l1ou fpkm Mus musculus minTau0.5 top10.tsv summary KEGG 2019 Human l1ou fpkm Mus musculus minTauO top10.tsv summary KEGG 2019 Human l1ou fpkm Mus musculus minTau0.5 top10.tsv summary KEGG 2019 Human l1ou tpm Mus musculus minTau0 top10.tsv summary KEGG 2019 Human llou tpm Mus musculus minTau0.5 top10.tsv summary_KEGG_2019_Human_l1ou_tpm_Mus_musculus_minTau0_top10.tsv summary KEGG 2019 Human l1ou tpm Mus musculus minTau0.5 top10.tsv summary_KEGG_2019_Mouse_l1ou_fpkm_Homo_sapiens_minTau0_top10.tsv summary_KEGG_2019_Mouse_l1ou_fpkm_Homo_sapiens_minTau0.5_top10.tsv summary_KEGG_2019_Mouse_l1ou_fpkm_Homo_sapiens_minTau0_top10.tsv summary_KEGG_2019_Mouse_l1ou_fpkm_Homo_sapiens_minTau0.5_top10.tsv summary_KEGG_2019_Mouse_l1ou_tpm_Homo_sapiens_minTau0_top10.tsv

summary_KEGG_2019_Mouse_l1ou_tpm_Homo_sapiens_minTau0.5_top10.tsv summary_KEGG_2019_Mouse_l1ou_tpm_Homo_sapiens_minTau0_top10.tsv summary_KEGG_2019_Mouse_l1ou_tpm_Homo_sapiens_minTau0.5_top10.tsv summary_KEGG_2019_Mouse_l1ou_fpkm_Mus_musculus_minTau0_top10.tsv summary_KEGG_2019_Mouse_l1ou_fpkm_Mus_musculus_minTau0.5_top10.tsv summary_KEGG_2019_Mouse_l1ou_fpkm_Mus_musculus_minTau0.top10.tsv summary_KEGG_2019_Mouse_l1ou_fpkm_Mus_musculus_minTau0.5_top10.tsv summary_KEGG_2019_Mouse_l1ou_tpm_Mus_musculus_minTau0.top10.tsv summary_KEGG_2019_Mouse_l1ou_tpm_Mus_musculus_minTau0.5_top10.tsv summary_KEGG_2019_Mouse_l1ou_tpm_Mus_musculus_minTau0.5_top10.tsv summary_KEGG_2019_Mouse_l1ou_tpm_Mus_musculus_minTau0.5_top10.tsv summary_KEGG_2019_Mouse_l1ou_tpm_Mus_musculus_minTau0.5_top10.tsv

[]:	
[]:	