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### First report of the *Brevipalpus*-transmitted (Trombidiformes: Tenuipalpidae) virus, *Orchid fleck dichorhavirus* (Mononegavirales: Rhabdoviridae) infecting ornamental Nolinoideae in the Florida panhandle

Austin **Fife**1, Daniel **Carrillo**2, Gary **Knox**3, Fanny **Iriarte**4, Kishore **Dey**5, Avijit **Roy**6, Ronald **Ochoa**6, Gary **Bauchan**7 , Mathews **Paret**4, Xavier **Martini**1\*

1 University of Florida, Department of Entomology and Nematology, North Florida Research and Education Center, Quincy FL 32351

2 University of Florida, Department of Entomology and Nematology, Tropical Research and Education Center, Homestead FL 33031

3 University of Florida, Department of Environmental Horticulture, North Florida Research and Education Center, Quincy FL 32351

4 University of Florida, Department of Plant Pathology, North Florida Research and Education Center, Quincy FL 32351

5 The Florida Department of Agriculture and Consumer Services, Division of Plant Industry, Section of Plant Pathology, Doyle Conner Building, 1911 SW 34th street, Gainesville, FL 32608

6 United States Department of Agriculture - Animal and Plant Health Inspection Service, Systematic Entomology Laboratory 10300 Baltimore Ave, Bldg. 5 BARC-West, Beltsville, MD 20705

7 United States Department of Agriculture - Animal and Plant Health Inspection Service, Electron and Confocal Microscopy Unit, Bldg. 12 BARC-West, 10300 Baltimore Ave, Beltsville, MD 20705

\*Corresponding author; E-mail: xmartini@ufl.edu Phone: 850-875-7160 Fax: 352-846-6617

### Abstract

Flat mites from the genus *Brevipalpus* Donnadieu (Trombidiformes: Tenuipalpidae) are the only known vectors of dichorhaviruses and transmit *Orchid fleck dichorhavirus* (OFV) in a persistent propagative manner. OFV is the type species for the genus *Dichorhavirus* and infects 50+ spp. of Orchidaceae, Nolinoideae, and *Citrus*. During June 2020, we observed chlorotic ringspot symptoms on Giant Lilyturf (*Liriope* spp., cv. ‘Gigantea’) in a landscape of Leon County, Florida. We tested plant samples for a wide range of pathogens and ultimately identified the pathogen as *Orchid fleck dichorhavirus*. We took further samples from other *Liriope* spp., *Ophiopogon* spp., and *Aspidistra elatior* Blume (Asparagales: Nolinoideae) in Leon and Alachua county. All symptomatic plants tested positive for OFV. We recovered mites from OFV-infected plants, which belonged to *Brevipalpus californicus* sensu lato. Florida has various mite species of *Brevipalpus* and a diverse array of susceptible plant species in the landscape, both native and introduced. We suggest that OFV already has a wide distribution in Florida which will continue to spread if unchecked, representing a potential threat for many plants of economic importance for Florida.

### Keywords:

false spider mite, flat mite, *Brevipalpus*-transmitted viruses, *Liriope*, Ruscaceae, Asparagaceae, orchid, Orchidaceae, pests, ornamental plants

*Orchid fleck dichorhavirus* (OFV) is the type species for the genus *Dichorhavirus*, a group of plant-infecting, bacilliform, nuclear rhabdoviruses composed of two segments of single-stranded, negative-sense RNA (Dietzgen et al. 2014, Walker et al. 2018, Amarasinghe et al. 2019). Other members of this genus are: *Citrus chlorotic spot dichorhavirus*, *Citrus leprosis N dichorhavirus*, *Clerodendrum chlorotic spot dichorhavirus* and *Coffee ringspot dichorhavirus* (Dietzgen, Freitas-Astúa, et al. 2018). Flat mites from the genus *Brevipalpus* Donnadieu (Trombidiformes: Tenuipalpidae) are the only known vector for dichorhaviruses (Maeda 1998). These mites transmit OFV in a persistent propagative manner (Kondo et al. 2003).

OFV-infected plants exhibit various symptoms dependent on the variety of plant infected as well as the strain of the virus (Kubo et al. 2009), but symptoms typically appear as chlorotic flecks, which ultimately coalesce into larger spots or ringspot patterns (Fig. 1).

OFV was first described infecting *Cymbidium* orchids in Japan (Doi et al. 1977). Many countries have reported OFV and OFV-like rhabdoviruses infecting orchids worldwide (Kondo et al. 2003), including Australia (Lesemann and Begtrup 1971, Lesemann and Doraiswamy 1975, Gibbs 2000), Brazil (Kitajima et al. 1974, Kitajima et al. 2001), China (Peng et al. 2017), Colombia (Kubo et al. 2009), Costa Rica (Freitas-Astúa et al. 2002), Denmark (Begtrup 1972), Fiji (Pearson et al. 1993), France (Sauvêtre et al. 2018), Germany (Petzold 1971, Lesemann and Doraiswamy 1975), Korea Zheng et al. (2013), Paraguay (Ramos-González et al. 2015), South Africa (Blanchfield et al. 2001), the United States (Blanchfield et al. 2001, Bratsch et al. 2015) and Vanuatu (Pearson et al. 1993). The prevalence of OFV and its mite vector is thought to be associated with the movement of infected orchids (Dietzgen, Freitas-Astúa, et al. 2018).

OFV naturally infects 50+ spp of Orchidaceae (Kitajima et al. 2010, Peng et al. 2013), some Nolinoideae Dietzgen, Tassi, et al. (2018), and *Citrus* where it causes citrus leprosis-like symptoms Roy et al. (2020). Mechanical transmission of OFV is possible under lab conditions to various *Chenopodiaceae*, *Aizoaceae*, *Fabaceae*, and *Solanaceae* (Chang et al. 1976, Kondo et al. 2003, Peng et al. 2013).

During June 2020, chlorotic ringspot symptoms were observed on the liriopogon Giant Lilyturf *Liriope* spp., cv. ‘Gigantea’ in a landscape of Leon County, Florida (Fig. 1). Liropogons belong to the plant family Asparagaceae, subfamily Nolinoideae, which includes various monocotyledonous lilliod plants (Chase et al. 2009). Liriopogons are native to southeastern Asia, encompassing the closely-related genera *Liriope* and *Ophiopogon*, which are characterized by evergreen, long grass-like leaves, and erect flowers which produce black, purple or blue, berry-like fruits, depending on the cultivar (Lattier et al. 2014, Fantz et al. 2015). These characteristics have given rise to various common names, including: monkey grass, Aztec grass, lilyturf, mondo grass, etc. (*Table 1*). The genus *Liriope* contains eight species, four of which are commonly cultivated, while the genus *Ophiopogon* consists of about 65 species, with only a handful are available for horticulture (Nesom 2010, Lattier et al. 2014). Liripogons are considered the most important ground cover sold by the nursery industry in southeastern US (Mcharo et al. 2003).

Plant leaf samples were initially tested at the Plant Disease Diagnostic Clinic at the North Florida Research and Education Center (NFREC) in Quincy, FL. The plant samples were tested for a wide range of common plant pathogens, but the results were inconclusive. The infected materials were subsequently sent to the Florida Department of Agriculture and Consumer Services (FDACS). The presence of *Orchid fleck dichorhavirus* was confirmed via generic one step conventional RT-PCR with R2-Dicho-GF and R2-Dicho-GR primers, amplifying ~800 nt of L-gene (RNA2) amplicon from a leaf sample of infected *Liriope* (Kubo et al. 2009). RT-qPCR confirmed OFV and the RT-PCR product was sequenced using Sanger sequencing and High Throughput Sequencing (Kondo et al. 2006). Sanger sequencing of RT-PCR amplicons shared 98 % nucleotide identity with OFV strains which infect orchids (GenBank Accession numbers: AB244418 and MK522807) (Kondo et al. 2006, 2014, Cook et al. 2019). Further samples were taken from various symptomatic cultivars of *Liriope* spp., *Ophiopogon* spp., as well as *Aspidistra elatior* Blume (Asparagaceae: Nolinoideae) during subsequent visits to the initial site of collection as well as other locations in Leon county. Samples were tested via RT-PCR at the NFREC (*Table 1*) to confirm the presence of OFV.

Further surveys of Nolinoideae in Florida have revealed more sites with symptomatic plants in both Leon and Alachua counties. In addition to the discovery of the OFV-infected plants, we collected mites from symptomatic plants in Leon county which were observed with phase contrast microscopy. We encountered both eriophyoid mites and flat mites which were identified as *Brevipalpus californicus* (Banks) sensu lato, an identity which was confirmed by the FDACS via Differential Interference Contrast (DIC) microscopy. *Brevipalpus* mites have been previously associated with OFV (Dietzgen, Tassi, et al. 2018, García-Escamilla et al. 2018, Beltran-Beltran et al. 2020) and similar diseases (Kitajima et al. 2010) and are known to feed on a large variety of economically-important plants (Childers et al. 2003, Akyazi et al. 2017). Unfortunately, the *Brevipalpus* mite species complex is known to contain cryptic species (Childers and Rodrigues 2011) whose identification can be improved with molecular methods as well as more advanced microscopy techniques, such as low-temperature scanning electron microscopy (LT-SEM) (León and Nadler 2010, Skoracka et al. 2015). With that in mind, we sent additional samples of the mites to the USDA-ARS in Beltsville to observe the mites with LT-SEM techniques, which agreed with both prior identifications of *B. californicus* s.l. (Fig. 2).

The earliest mention of a virus which may have been OFV is Ko et al. (1985), who detected nuclear inclusions caused by an undescribed bacilliform rhabdovirus. Ko et al. (1985) described the spoke-wheel configurations typically associated with OFV (Chang et al. 1976), in *Brassia* orchids, but unfortunately made no mention of mites or further investigations of this virus. The first certain report of OFV in the US was made by Bratsch et al. (2015), who confirmed the presence of OFV in *Phalaenopsis* hybrids in the US, using TEM of ultrathin sections of plant tissue as well as molecular sequence analysis. In addition, Bratsch et al. (2015) detected *Brevipalpus* mites and their exoskeletons associated with OFV-infected plants. It appears that Bratsch et al. (2015) did not make a conclusive species identification, but did cite Kondo et al. (2003), suggesting that they suspected *B. californicus* as their vector.

OFV has been reported in other Nolinoideae in Australia (Mei et al. 2016, Dietzgen, Tassi, et al. 2018), including *Liriope spicata* (Thunb.) Lour. (Mei et al. 2016). Our plants are thought to belong to different species of *Liriope* which are not *L. spicata* and we are not aware of any record other than this manuscript which reports OFV infection in *Ophiopogon* plants. Unfortunately, liriopogons species are very similar in appearance and growth habit, with few useful characters used for their classification (Fantz 2008a). Furthermore, the horticultural industry has created a diverse array of cultivars of these plants, which are often mislabeled (Fantz 2008a). Aside from the taxonomic confusion created by humans, natural hybrids between *Ophiopogon* and *Liriope* have created a natural source of error for reconstructing phylogenies (Zhou et al. 2009). Together, these factors make it difficult to differentiate and identify plants in the landscape by visual inspection alone. These obfuscations of species identity may be accounted for in the future via sequence comparisons of the OFV-infected plants, but these comparisons are beyond the scope of our current report. Nonetheless, we are confident that ours is the first report of OFV infecting *Aspidistra elatior*; although (Zheng et al. 2013) mentions the association of *B. californicus* with *A. elatior* they make no mention of OFV symptoms in this plant.

### Control methods for *Brevipalpus* mites

Reducing the populations of the *Brevipalpus* mite vector may be able to reduce the spread of OFV by infective mites, but infected plants are not able to recover from the virus and must be rouged or removed. Roguing plants is a direct and efficient option to remove mite and virus populations quickly but increases in difficultly with the number of plants involved. Once infected plants have been removed, the objective will be to prevent re-infestation of the *Brevipalpus* vector via conventional pest management methods.

Most research on the integrated management of mites has been catered towards controlling various species of spider mites (Acari: Tetranychidae) (Leeuwen et al. 2015), but many of those methods can be applied to *Brevipalpus* spp. as well:

Chemical control via synthetic acaricides is commonly used to control *Brevipalpus* spp. (Leeuwen et al. 2015), but acaricides have some drawbacks. Many products control a broad class of arthropods, but often they do so indiscriminately, killing beneficial insects as well as pests (Suckling et al. 2013). Control strategies which rely heavily on chemical control have the potential to encourage pesticide resistance in mite populations and pesticide resistance has been reported in various *Brevipalpus* populations (Alves et al. 2000, Omoto et al. 2000, Campos and Omoto 2002). In addition, it is important to consider the interactions which may occur between different chemical applications or tank mixes (Vechia et al. 2018). Lastly, application costs of chemical controls can be large and increase the cost of production (Rodrigues and Machado 2000), nevertheless chemical applications are valuable tools used to consider against *Brevipalpus*.

Various biological control methods have also been studied, but their use has not been widely adopted (Messing and Brodeur 2017). Even so, some studies have found biological control methods which could be used specifically for *Brevipalpus* mites: The predatory mite *Galendromus helveolus* (Chant) (Acari: Phytoseiidae) has been reported to feed on eggs, larvae and nymphs of *B. californicus* (Chen et al. 2006) and studies of *Amblyseius largoensis* (Muma) (Acari: Phytoseiidae) demonstrated its ability to suppress populations of *B. yothersi* on citrus plants (Argolo et al. 2020). In addition, the entomopathogenic fungus *Metarhizium anisopliae* var. *acridum* was found to be pathogenic to *B. phoenicis* (Magalhães et al. 2005). In our own surveys, we have found mites which succumbed to fungal infections (Fig. 3); however the fungal species was not determined.

### Conclusion

Detecting OFV in Florida represents a concern for horticulturists who grow susceptible plants of economic importance, including orchids, *Liriope*, *Ophiopogon*, or other susceptible Asparagales spp. which are commonly used in landscaping. Florida is also home to a plethora of native and naturalized orchid species, many of which are threatened, including the famous Ghost Orchid, *Dendrophylax lindenii* (Lindl.) Benth. ex Rolfe. Furthermore, OFV represents an obstacle to overcome for the burgeoning interest in cultivating *Vanilla* in southern Florida (Chambers et al. 2019). Lastly, some OFV isolates are known to be involved with nuclear types of citrus leprosis (Roy et al. 2015), which may be a cause for concern for the citrus industry. In fact, Kitajima et al. (2011) found that the Citrus Leprosis virus (CiLV) which previously affected Florida citrus was a nuclear type of citrus leprosis, which are closely related to OFV strains (Roy et al. 2013). *B. californicus* and *B. yothersi* are both known vectors of Dichorhaviruses (OFV) and Cileviruses (Citrus Leprosis) (Knorr 1968, Kondo et al. 2003, Beltran-Beltran et al. 2020) and *B. obovatus* is a suspected vector as well (Childers et al. 2003). All three mite species/complexes are present in Florida (Childers et al. 2003, Akyazi et al. 2017) (Fig. 4); therefore, it is critical to monitor the spread of Orchid Fleck Virus and its mite vector(s) to determine what risk this virus represents for plants in Florida and the surrounding regions.

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### References

**Akyazi, R., E. A. Ueckermann, and O. E. Liburd**. **2017**. New report of *Brevipalpus yothersi* (prostigmata: Tenuipalpidae) on blueberry in Florida. Florida Entomologist. 100: 731–739.

**Alves, E. B., C. Omoto, and C. R. Franco**. **2000**. Resistência cruzada entre o dicofol e outros acaricidas em *Brevipalpus phoenicis* (Geijskes) (acari: tenuipalpidae). Anais da Sociedade Entomológica do Brasil. 29: 765–771.

**Amarasinghe, G. K., M. A. Ayllón, Y. Bào, C. F. Basler, S. Bavari, K. R. Blasdell, T. Briese, P. A. Brown, A. Bukreyev, A. Balkema-Buschmann, U. J. Buchholz, C. Chabi-Jesus, K. Chandran, C. Chiapponi, I. Crozier, R. L. de Swart, R. G. Dietzgen, O. Dolnik, J. F. Drexler, R. Dürrwald, W. G. Dundon, W. P. Duprex, J. M. Dye, A. J. Easton, A. R. Fooks, P. B. H. Formenty, R. A. M. Fouchier, J. Freitas-Astúa, A. Griffiths, R. Hewson, M. Horie, T. H. Hyndman, D. Jiāng, E. W. Kitajima, G. P. Kobinger, H. Kondō, G. Kurath, I. V. Kuzmin, R. A. Lamb, A. Lavazza, B. Lee, D. Lelli, E. M. Leroy, J. Lǐ, P. Maes, S.-Y. L. Marzano, A. Moreno, E. Mühlberger, S. V. Netesov, N. Nowotny, A. Nylund, A. L. Økland, G. Palacios, B. Pályi, J. T. Pawęska, S. L. Payne, A. Prosperi, P. L. Ramos-González, B. K. Rima, P. Rota, D. Rubbenstroth, M. Shı̄, P. Simmonds, S. J. Smither, E. Sozzi, K. Spann, M. D. Stenglein, D. M. Stone, A. Takada, R. B. Tesh, K. Tomonaga, N. Tordo, J. S. Towner, B. van den Hoogen, N. Vasilakis, V. Wahl, P. J. Walker, L.-F. Wang, A. E. Whitfield, J. V. Williams, F. M. Zerbini, T. Zhāng, Y.-Z. Zhang, and J. H. Kuhn**. **2019**. Taxonomy of the order Mononegavirales: Update 2019. Archives of Virology. 164: 1967–1980.

**Argolo, P. S., A. M. Revynthi, M. A. Canon, M. M. Berto, D. J. Andrade, İ. Döker, A. Roda, and D. Carrillo**. **2020**. Potential of predatory mites for biological control of *brevipalpus yothersi* (acari: tenuipalpidae). Biological Control. 149: 104330.

**Baker, E. W., and D. M. Tuttle**. **1987**. The false spider mites of Mexico (tenuipalpidae: acari). (technical report No. 1706). The United States Department of Agriculture - Agricultural Research Service.

**Beard, J. J., R. Ochoa, R. Bauchan G., D. Trice M., J. Redford A., W. Walters T., and C. Mitter**. **2012**. Flat mites of the world edition 2. (<http://idtools.org/id/mites/flatmites/>).

**Begtrup, J.** **1972**. Structure of a bacilliform virus in Dendrobium as revealed by negative staining. Journal of Phytopathology. 75: 268–273.

**Beltran-Beltran, A. K., M. T. Santillán-Galicia, A. W. Guzmán-Franco, D. Teliz-Ortiz, M. A. Gutiérrez-Espinoza, F. Romero-Rosales, and P. L. Robles-Garcı́a**. **2020**. Incidence of citrus leprosis virus c and orchid fleck dichorhavirus citrus strain in mites of the genus *Brevipalpus* in Mexico. Journal of Economic Entomology. 113: 1576–1581.

**Blanchfield, A. L., A. M. Mackenzie, A. Gibbs, H. Kondo, T. Tamada, and C. R. Wilson**. **2001**. Identification of orchid fleck virus by reverse transcriptase-polymerase chain reaction and analysis of isolate relationships. Journal of Phytopathology. 149: 713–718.

**Bratsch, S. A., B. E. Lockhart, and C. Ishimaru**. **2015**. Confirmation of first report of orchid fleck virus in *phalaenopsis* hybrid orchids in the USA. Plant Health Progress. 16: 146–148.

**Broussard, M. C.** **2007**. A horticultural study of *Liriope* and *Ophiopogon*: Nomenclature, morphology, and culture (PhD thesis).

**Campos, F. J., and C. Omoto**. **2002**. Experimental and Applied Acarology. 26: 243–251.

**Chambers, A. H., P. Moon, V. Edmond, and E. Bassil**. **2019**. Vanilla cultivation in southern Florida. EDIS. 2019: 7.

**Chang, M. U.** **1991**. Studies on the viruses in orchids in Korea. 2. *Dendrobium mosaic virus, Odontoglossum ringspot virus, Orchid fleck virus* and unidentified potyvirus. Korean J Plant Pathol. 7: 118–129.

**Chang, M. U., Arai. Kei, Doi. Yoji, and Yora. Kiyoshi**. **1976**. Morphology and intracellular appearance of *Orchid fleck virus*. Japanese Journal of Phytopathology. 42: 156–157.

**Chase, Mark. W., James. L. Reveal, and M. F. Fay**. **2009**. A subfamilial classification for the expanded asparagalean families amaryllidaceae, asparagaceae and xanthorrhoeaceae. Botanical Journal of the Linnean Society. 161: 132–136.

**Chen, T.-Y., J. V. French, T.-X. Liu, and J. V. da Graça**. **2006**. Predation of *Galendromus helveolus* (acari: Phytoseiidae) on *Brevipalpus californicus* (acari: tenuipalpidae). Biocontrol Science and Technology. 16: 753–759.

**Childers, C. C., and J. C. V. Rodrigues**. **2011**. An overview of *Brevipalpus* (acari: Tenuipalpidae) and the plant viruses they transmit. Zoosymposia. 6: 180–192.

**Childers, C. C., J. C. V. Rodrigues, and W. C. Welbourn**. **2003**. Host plants of *brevipalpus californicus*, *b. Obovatus*, and *b. Phoenicis* (acari: Tenuipalpidae) and their potential involvement in the spread of viral diseases vectored by these mites. Experimental and Applied Acarology. 30: 29–105.

**Cook, G., W. Kirkman, R. Clase, C. Steyn, E. Basson, P. H. Fourie, S. D. Moore, T. G. Grout, E. Carstens, and V. Hattingh**. **2019**. Orchid fleck virus associated with the first case of citrus leprosis-N in South Africa. European Journal of Plant Pathology. 155: 1373–1379.

**Dietzgen, R. G., J. Freitas-Astúa, C. Chabi-Jesus, P. L. Ramos-González, M. M. Goodin, H. Kondo, A. D. Tassi, and E. W. Kitajima**. **2018**. Dichorhaviruses in their host plants and mite vectors, pp. 119–148. *In* Advances in Virus Research. Elsevier.

**Dietzgen, R. G., J. H. Kuhn, A. N. Clawson, J. Freitas-Astúa, M. M. Goodin, E. W. Kitajima, H. Kondo, T. Wetzel, and A. E. Whitfield**. **2014**. Dichorhavirus: A proposed new genus for *Brevipalpus* mite-transmitted, nuclear, bacilliform, bipartite, negative-strand RNA plant viruses. Archives of Virology. 159: 607–619.

**Dietzgen, R. G., A. D. Tassi, J. Freitas-Astúa, and E. W. Kitajima**. **2018**. First report of orchid fleck virus and its mite vector on green cordyline. Australasian Plant Disease Notes. 13.

**Doi, Y., M. U. Chang, and K. Yora**. **1977**. Orchid fleck virus. CMI/AAB descriptions of plant viruses.

**Fantz, P. R.** **2008a**. Macrophytography of cultivated liriopogons and genera delineation. HortTechnology. 18: 334–342.

**Fantz, P. R.** **2008b**. Species of *Liriope* cultivated in the southeastern United States. HortTechnology. 18: 343–348.

**Fantz, P. R.** **2009**. Names and species of *Ophiopogon* cultivated in the southeastern United States. HortTechnology. 19: 385–394.

**Fantz, P. R., D. Carey, T. Avent, and J. Lattier**. **2015**. Inventory, descriptions, and keys to segregation and identification of liriopogons cultivated in the southeastern United States. HortScience. 50: 957–993.

**Freitas-Astúa, J., L. Moreira, C. Rivera, C. M. Rodrı́guez, and E. W. Kitajima**. **2002**. First report of orchid fleck virus in Costa Rica. Plant Disease. 86: 1402–1402.

**García-Escamilla, P., Y. Duran-Trujillo, G. Otero-Colina, G. Valdovinos-Ponce, Ma. T. Santillán-Galicia, C. F. Ortiz-Garcı́a, J. J. Velázquez-Monreal, and S. Sánchez-Soto**. **2018**. Transmission of viruses associated with cytoplasmic and nuclear leprosis symptoms by *Brevipalpus yothersi* and *B. californicus*. Tropical Plant Pathology. 43: 69–77.

**Gibbs, A.** **2000**. Viruses of orchids in Australia; their identification, biology and control. The Australian Orchid Rev. 65: 10–21.

**Kitajima, E. W., A. Blumenschein, and A. S. Costa**. **1974**. Rodlike particles associated with ringspot symptoms in several orchid species in Brazil. Journal of Phytopathology. 81: 280–286.

**Kitajima, E. W., C. M. Chagas, R. Harakava, R. F. Calegario, J. Freitas-Astúa, J. C. V. Rodrigues, and C. C. Childers**. **2011**. Citrus leprosis in Florida, USA, appears to have been caused by the nuclear type of citrus leprosis virus (CilLV-N). Virus Reviews & Research. 16.

**Kitajima, E. W., H. Kondo, A. Mackenzie, J. A. M. Rezende, R. Gioria, A. Gibbs, and T. Tamada**. **2001**. Comparative cytopathology and immunocytochemistry of Japanese, Australian and Brazilian isolates of orchid fleck virus. Journal of General Plant Pathology. 67: 231–237.

**Kitajima, E. W., J. C. V. Rodrigues, and J. Freitas-Astua**. **2010**. An annotated list of ornamentals naturally found infected by *Brevipalpus* mite-transmitted viruses. Scientia Agricola. 67: 348–371.

**Knorr, L. C.** **1968**. Studies on the etiology of leprosis in citrus. *In* International Organization of Citrus Virologists Conference Proceedings.

**Ko, N.-J., F. W. Zettler, J. R. Edwardson, and R. G. Christie**. **1985**. Light microscopic techniques for detecting orchid viruses. Acta Horticulturae. 241–254.

**Kondo, H., T. Maeda, Y. Shirako, and T. Tamada**. **2006**. Orchid fleck virus is a rhabdovirus with an unusual bipartite genome. Journal of General Virology. 87: 2413–2421.

**Kondo, H., T. Maeda, and T. Tamada**. **2003**. Orchid fleck virus: *Brevipalpus californicus* mite transmission, biological properties and genome structure. Experimental and Applied Acarology. 30: 215–223.

**Kondo, H., K. Maruyama, S. Chiba, I. B. Andika, and N. Suzuki**. **2014**. Transcriptional mapping of the messenger and leader RNAs of orchid fleck virus, a bisegmented negative-strand RNA virus. Virology. 452-453: 166–174.

**Kubo, K. S., J. Freitas-Astúa, M. A. Machado, and E. W. Kitajima**. **2009**. Orchid fleck symptoms may be caused naturally by two different viruses transmitted by *Brevipalpus*. Journal of General Plant Pathology. 75: 250–255.

**Lattier, J. D., T. G. Ranney, P. R. Fantz, and T. Avent**. **2014**. Identification, nomenclature, genome sizes, and ploidy levels of *Liriope* and *Ophiopogon* taxa. HortScience. 49: 145–151.

**Leeuwen, T. V., L. Tirry, A. Yamamoto, R. Nauen, and W. Dermauw**. **2015**. The economic importance of acaricides in the control of phytophagous mites and an update on recent acaricide mode of action research. Pesticide Biochemistry and Physiology. 121: 12–21.

**León, G. P.-P. de, and S. A. Nadler**. **2010**. What we don’t recognize can hurt us: A plea for awareness about cryptic species. Journal of Parasitology. 96: 453–464.

**Lesemann, D., and J. Begtrup**. **1971**. Elektronenmikroskopischer nachweis eines bazilliformen virus in *Phalaenopsis*. Journal of Phytopathology. 71: 257–269.

**Lesemann, D., and S. Doraiswamy**. **1975**. Bullet-shaped virus-like particles in chlorotic and necrotic leaf lesions of orchids. Journal of Phytopathology. 83: 27–39.

**Maeda, T.** **1998**. Evidence that orchid fleck virus is efficiently transmitted in a persistent manner by the mite *Brevipalpus californicus*. Abstr., 7th Int. Cong. Plant Pathol. 3.

**Magalhães, B. P., J. C. V. Rodrigues, D. G. Boucias, and C. C. Childers**. **2005**. Pathogenicity of *metarhizium anisopliae* var. Acridum to the false spider mite *brevipalpus phoenicis* (acari: tenuipalpidae). Florida Entomologist. 88: 195–198.

**Masiero, E., D. Banik, J. Abson, P. Greene, A. Slater, and T. Sgamma**. **2020**. Molecular verification of the UK national collection of cultivated *Liriope* and *Ophiopogon* plants. Plants. 9: 558.

**Mcharo, M., E. Bush, D. L. Bonte, C. Broussard, and L. Urbatsch**. **2003**. Molecular and morphological investigation of ornamental liriopogons. Journal of the American Society for Horticultural Science. 128: 575–577.

**Mei, Y., N. Bejerman, K. S. Crew, N. McCaffrey, and R. G. Dietzgen**. **2016**. First report of orchid fleck virus in lilyturf (*Liriope spicata*) in Australia. Plant Disease. 100: 1028–1028.

**Messing, R., and J. Brodeur**. **2017**. Current challenges to the implementation of classical biological control. BioControl. 63: 1–9.

**Nesom, G. L.** **2010**. Overview of *liriope* and *ophiopogon* (ruscaceae) naturalized and commonly cultivated in the USA. Phytoneuron. 56: 1–31.

**Omoto, C., E. B. Alves, and P. C. Ribeiro**. **2000**. Detecção e monitoramento da resistência de *brevipalpus phoenicis* (Geijskes) (acari: Tenuipalpidae) do dicofol. Anais da Sociedade Entomológica do Brasil. 29: 757–764.

**Pearson, M. N., G. V. H. Jackson, S. P. Pone, and R. L. J. Howitt**. **1993**. Vanilla viruses in the South Pacific. Plant Pathology. 42: 127–131.

**Peng, D. W., G. H. Zheng, Q. X. Tong, Z. Z. Zheng, and Y. L. Ming**. **2017**. First report of orchid fleck dichorhavirus from *cymbidium* sp. In China. Plant Disease. 101: 514–514.

**Peng, D. W., G. H. Zheng, Z. Z. Zheng, Q. X. Tong, and Y. L. Ming**. **2013**. Orchid fleck virus: An unclassified bipartite, negative-sense RNA plant virus. Archives of Virology. 158: 313–323.

**Petzold, H.** **1971**. Der elektronenmikroskopische nachweis eines bacilliformen virus an blattfleckenkranken *Dendrobien*. Journal of Phytopathology. 70: 43–52.

**Ramos-González, P. L., H. Sarubbi-Orue, L. Gonzales-Segnana, C. Chabi-Jesus, J. Freitas-Astúa, and E. W. Kitajima**. **2015**. Orchid fleck virus infecting orchids in Paraguay: First report and use of degenerate primers for its detection. Journal of Phytopathology. 164: 342–347.

**Rodrigues, J. C. V., and M. A. Machado**. **2000**. Virus-*Brevipalpus*-plant relationships on citrus leprosis pathosystems. Proc. Int. Soc. Citriculture Congr. 3–7.

**Roy, A., J. S. Hartung, W. L. Schneider, J. Shao, G. Leon, M. J. Melzer, J. J. Beard, G. Otero-Colina, G. R. Bauchan, R. Ochoa, and R. H. Brlansky**. **2015**. Role bending: Complex relationships between viruses, hosts, and vectors related to citrus leprosis, an emerging disease. Phytopathology. 105: 1013–1025.

**Roy, A., A. L. Stone, G. Otero-Colina, G. Wei, R. H. Brlansky, R. Ochoa, G. Bauchan, W. L. Schneider, M. K. Nakhla, and J. S. Hartung**. **2020**. Reassortment of genome segments creates stable lineages among strains of orchid fleck virus infecting citrus in Mexico. Phytopathology. 110: 106–120.

**Roy, A., A. Stone, G. Otero-Colina, G. Wei, N. Choudhary, D. Achor, J. Shao, L. Levy, M. K. Nakhla, C. R. Hollingsworth, J. S. Hartung, W. L. Schneider, and R. H. Brlansky**. **2013**. Genome assembly of citrus leprosis virus nuclear type reveals a close association with orchid fleck virus. Genome Announcements. 1.

**Sauvêtre, P., E. Veniant, G. Croq, A. D. Tassi, E. W. Kitajima, C. Chabi-Jesus, P. L. Ramos-González, J. Freitas-Astúa, and D. Navia**. **2018**. First report of orchid fleck virus in the orchid collection of jardin du luxembourg, Paris, France. Plant Disease. 102: 2670–2670.

**Skoracka, A., S. Magalhães, B. G. Rector, and L. Kuczyński**. **2015**. Cryptic speciation in the acari: A function of species lifestyles or our ability to separate species? Experimental and Applied Acarology. 67: 165–182.

**Suckling, D. M., L. D. Stringer, A. E. A. Stephens, B. Woods, D. G. Williams, G. Baker, and A. M. El-Sayed**. **2013**. From integrated pest management to integrated pest eradication: Technologies and future needs. Pest Management Science. 70: 179–189.

**Vechia, J. F. D., M. C. Ferreira, and D. J. Andrade**. **2018**. Interaction of spirodiclofen with insecticides for the control of *Brevipalpus yothersii* in citrus. Pest Management Science. 74: 2438–2443.

**Walker, P. J., K. R. Blasdell, C. H. Calisher, R. G. Dietzgen, H. Kondo, G. Kurath, B. Longdon, D. M. Stone, R. B. Tesh, N. Tordo, N. Vasilakis, and A. E. Whitfield**. **2018**. ICTV virus taxonomy profile: rhabdoviridae. Journal of General Virology. 99: 447–448.

**Wang, G.-Y., Y. Meng, J.-L. Huang, and Y.-P. Yang**. **2014**. Molecular phylogeny of *Ophiopogon* (asparagaceae) inferred from nuclear and plastid DNA sequences. Systematic Botany. 39: 776–784.

**Zheng, G. H., Z. Z. Zheng, Q. X. Tong, Y. L. Ming, and others**. **2013**. Orchid fleck virus: An unclassified bipartite, negative-sense rna plant virus. Archives of virology. 158: 313–323.

**Zhou, Q., J. Zhou, J. Chen, and X. Wang**. **2009**. Karyotype analysis of medicinal plant *Liriope spicata* var. *prolifera* (liliaceae). Biologia. 64.

### Table

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| --- | --- | --- |
| Scientific Name | Common Names | Symptoms Observed |
| *Liriope muscari* Bailey | Lilyturf, Orchardgrass, Monkeygrass | Ringspots, Chlorotic Flecking, Necrotic Lesions |
| *Liriope gigantea*\* Hume | Giant Lilyturf | Ringspots, Chlorotic Flecking, Necrotic Lesions |
| *Ophiopogon japonicus* Ker Gawl. | Dwarf Lilyturf, Mondo Grass, Snake’s beard | Ringspots, Chlorotic Flecking, Necrotic Lesions |
| *Ophiopogon intermedius*\*\* Don | Aztec Grass, ‘Argenteomarginatus’ | Ringspots, Chlorotic Flecking, Necrotic Lesions |
| *Aspidistra elatior* Blume | Cast Iron Plant, Bar-room Plant | Chlorosis, Necrotic Lesions |

Table 1: List of plants with symptoms of Orchid fleck dichorhavirus found in northern Florida. \* *L. gigantea* have been traditionally classified as seperate from *L. muscari* by Broussard (2007) and Fantz et al. (2015), although this distinction has been challenged by Wang et al. (2014) and Masiero et al. (2020). \* \* *O. intermedius* is sometimes misclassified as *Liriope muscari* ‘Variegated Evergreen Giant’ Fantz (2009) or ‘Grandiflora White’ (Fantz 2009).

### Figure captions

Fig. 1: Variety of symptoms expressed by plants infected with Orchid fleck dichorhavirus: (a) ringspot symptoms on *Liriope gigantea* (b) chlorotic flecking on *Aspidistra elatior*.

Fig. 2: LT-SEM images of *Brevipalpus californicus* sensu lato displaying various characters used for identification (Baker and Tuttle 1987, Beard et al. 2012) (a) Dorsum (b) Lateral view (c) Venter (d) Close up of distal end of leg 2, with arrows indicating paired solenidia, characteristic of the genus *Brevipalpus* (e) Enlargement of the microplates of the mite cerotegument (f) Dorsal view of the distal portion of mite abdomen (g) Dorsal view of the mite rostrum (h) Ventral view of mite rostrum, observe 3 distal setae. LT-SEM images provided by Dr. Gary Bauchan, USDA-ARS 2021

Fig. 3: LT-SEM images of unidentified fungus infecting *Brevipalpus californicus* sensu lato: (a) Infested *B. californicus* adult, dorsal (b) Detail of fungal sporangia. LT-SEM images provided by Dr. Gary Bauchan, USDA-ARS 2021

Fig. 4: Florida is home to other common pest species of *Brevipalpus* which are potential vectors of *Orchid fleck dichorhavirus*: (a) *B. yothersi*, dorsal (b) *B. yothersi*, lateral (c) *B. obovatus*, dorsal. SEM images provided by Dr. Ron Ochoa, USDA-ARS 2021

### Figures

   