**Mite Survey Protocol**

**Objective: To collect mites on roses in Florida**

*Equipment:*

* Datasheet
* GPS with photo capability or phone with GPS and Solocator installed
* Bypass pruners
* 70% ethanol in spray bottle
* 95% ethanol for sample storage
* 500 mL polypropylene bottles
* China marker
* UFL Business Card and Project Sheet

***Warning: make sure you ask permission before sampling on private property!***

*Before Sampling:*

Before taking any cuttings, sanitize your bypass prunes with 70% ethanol, take a picture of the roses you will be sampling, fill out the datasheet and label the sampling bottle with a sampling ID, sampling number and date. ***Make sure you label your bottles on both the lid and on the side!*** Use the supplied china marker, ethanol will dissolve other markers.

*Sampling Method:*

Any species from the genus *Rosa* is suitable for our sampling. *Phyllocoptes fructiphilus* are found under the sepals of roses, so blooming flowers are preferred when sampling. If no blooms are present, try to select flowers which have recently bloomed, or which will bloom soon. Fruits and buds are better than nothing. Then cut a bloom with ~5 cm of cane underneath. Try to take enough tissue to fill the 500 mL bottle so that the lid still goes on easily. After the bottle is full, add a little (~10 mL) of the 95% ethanol, put the lid on tightly and shake the bottle for about 30 seconds so the ethanol covers the entire plant and kills any arthropods present. Before and after cutting anything, sanitize the bypass pruners by spraying them with 70% ethanol, especially when taking tissues from a different plant.

*Sampling Tips:*

Each bottle should contain a sample which represents an entire rose planting. This means, take cuttings from all sides of the roses and take flowers from different levels of the plant. If the rose patch is large, subdivide the planting into a new sampling bottle for every 12 feet of the planting, with a new GPS coordinate as well.