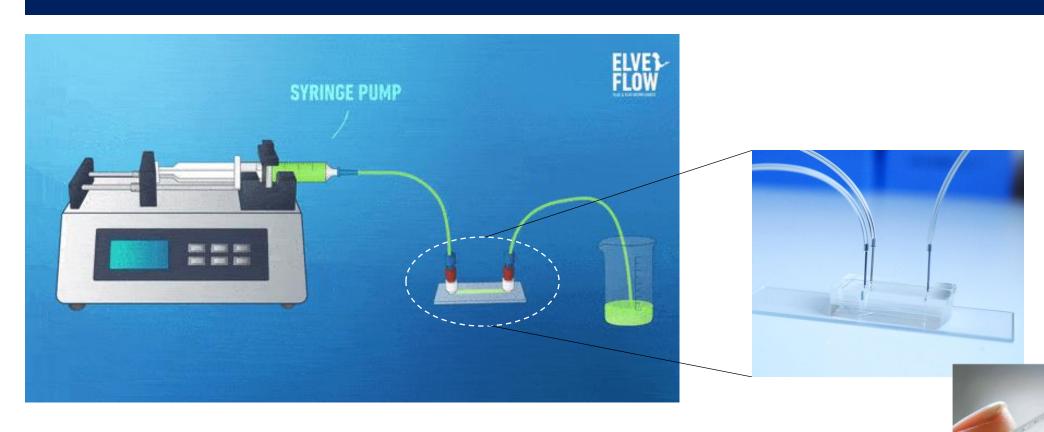
BBL 737- MICROFABRICATION

Contents

- Microfluidics
- Microfabrication technique to fabricate microfluidic devices
- **Scope of microfluidic experiments in biological system**
- Governing equations for microfluidic flows

Microfluidic Device



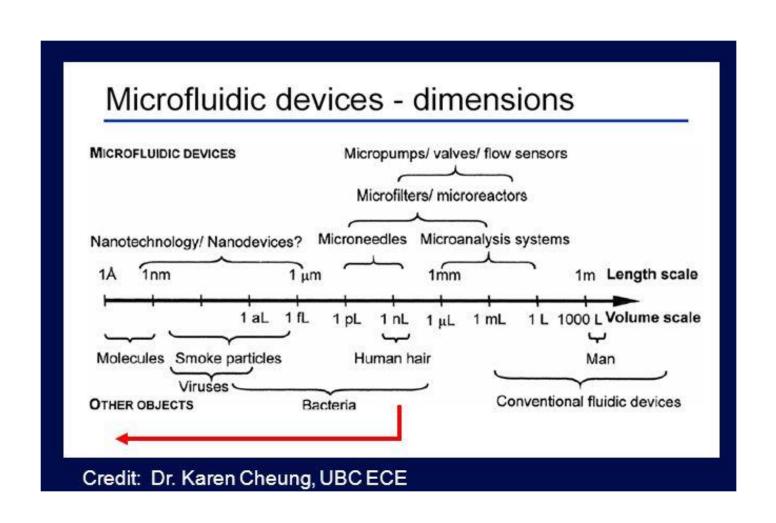
- o Macromolecular analysis (DNA, Proteins, etc.)
- Disease detection and diagnostic purposes
- Drug testing/adminitration
- Cell manipulation

Microfluidics

 Study of fluid flow with <u>characteristic length scale</u> of the order of Microns

o Processes and devices that deal with volume of fluid in the ranges between pico-liters to nano-liters

Microfluidics

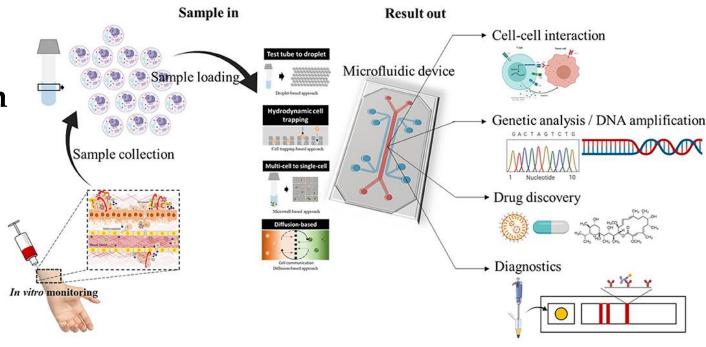


Why miniaturization?

- o Minimize material consumption and sample
- Reduction of power budget
- Faster analysis (scaling)
- Exploitation of new effects
- Required when application demands handling of very small volumes (drug administration)
- Cost/performance advantages
- o Improved reproducibility, improved accuracy and reliability

Implications in biological systems

- o Fundamental understanding of biophysical processes
- Manipulation and analysis of biological macromolecules (DNA/RNA), proteins, cell, etc.
- Biomedical diagnostics
- Drug delivery/Blood extraction

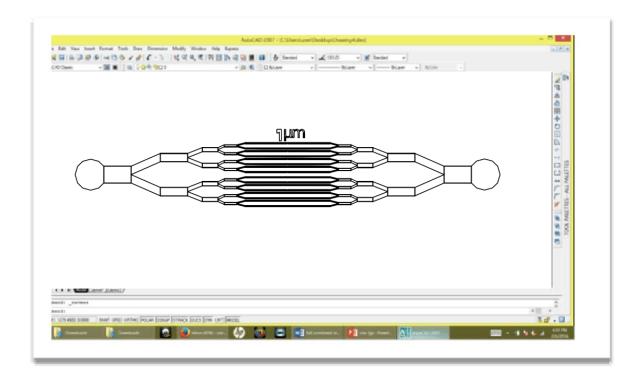


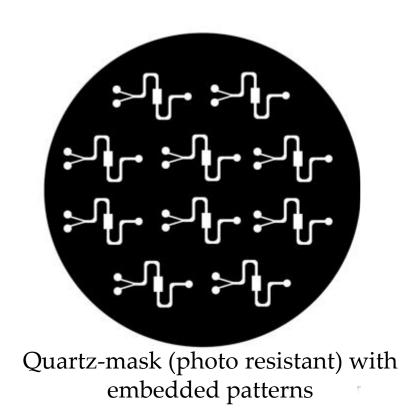
Microfabrication or Photolithography Technique

- 1. Circuit design and mask printing
- 2. Precleaning and dehydration of wafer
- 3. Wafer coating with photoresist (film deposition)
- 4. Pre-exposure soft bake
- 5. Mask-alignment
- 6. Post-exposure soft bake
- 7. Development (Wet-etching)
- 8. Hard bake
- 9. Soft-lithography
- 10. Inspection of wafer
- 11. Plasma bonding

1. Circuit design and Printing on Mask

Softwares to design the circuit – AutoCAD/FreeCAD, Clevin, DraftSight





2. Precleaning and dehydration of silicon-wafer (2", 3", 5"...)

Thin slice of wafer serve as a substrate





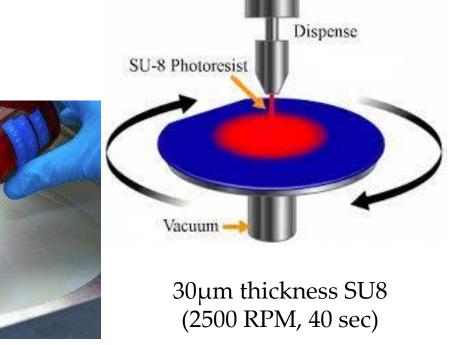
- Rinsing with acetone and IPA, blow dry with N₂ for clean wafers to remove resides on the surface
- Dehydration to remove moisture content from wafer



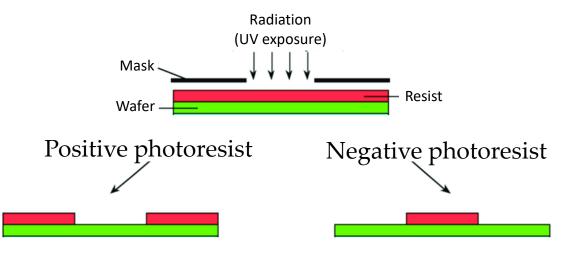
Drying at 200°C for 30 minutes

3. Spin-coating of photo-resist





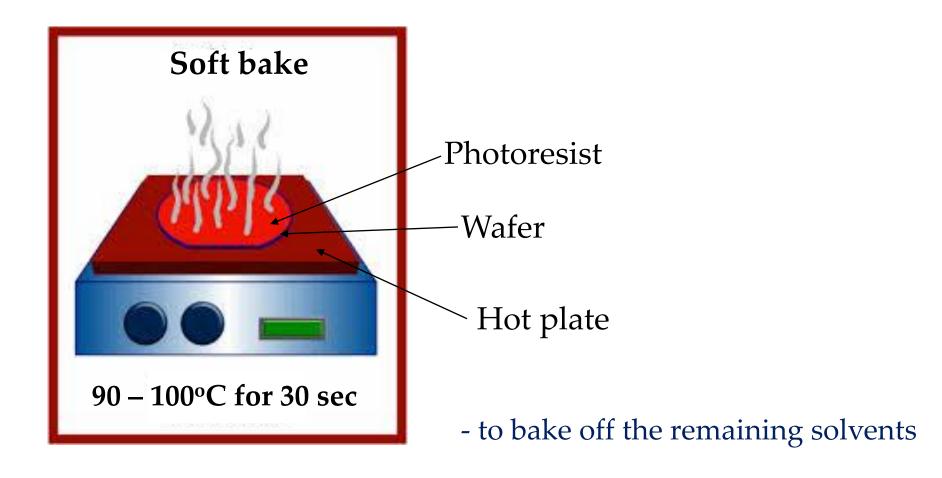
Spin-coating of photoresist...contd.



Two types of photoresists

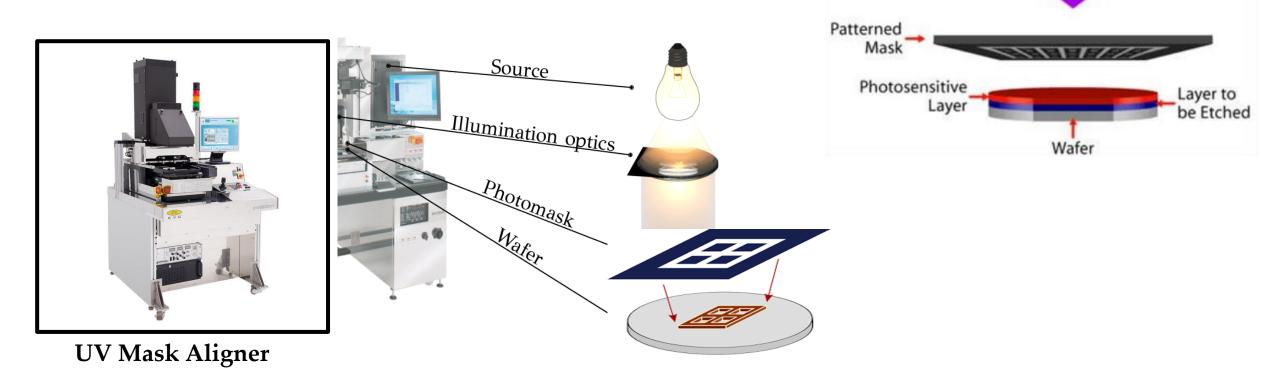
- 1. Positive photoresist portion of the photoresist that is exposed to light becomes soluble to the photoresist developer. The unexposed portion of the photoresist remains insoluble to the photoresist developer.
- 2. Negative photoresist portion of the photoresist that is exposed to light becomes insoluble to the photoresist developer (cross-linked). The unexposed portion of the photoresist is dissolved by the photoresist developer SU8

4. Pre-exposure soft bake on hot plate



UV Light

5. UV Mask-alignment



Transfer of geometric design from an optical mask to a light-sensitive photoresist coated on the wafer

6. Post-exposure soft bake on hot plate



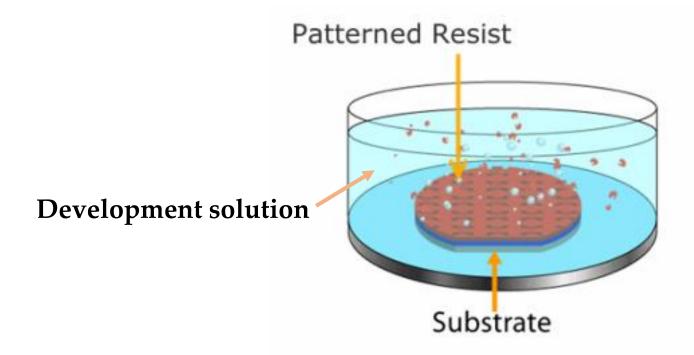
Silicon wafer coated with exposed photoresist



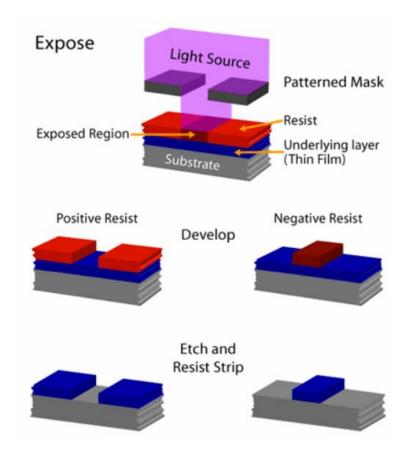
Hot plate (T = 95 °C)

- Performed immediately after exposure
- Completes the photo reaction initiated during exposure
- Same procedure as the pre-exposure soft bake:
 - T = 95 °C on hot plate
 - -t=2 min
 - Cool to room temperature

7. Development (Wet chemical etching)

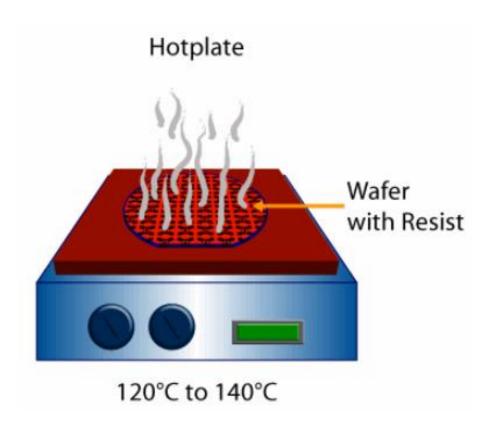


Dissolution of un-crosslinked photoresist in development solution (treatment time: 2-3 minutes)



Difference between negative and positive photoresist (Post-development)

8. Hard-bake at 135°C

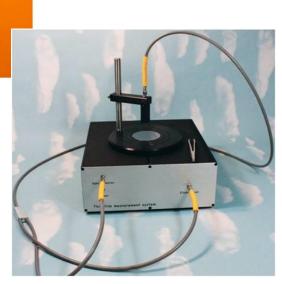


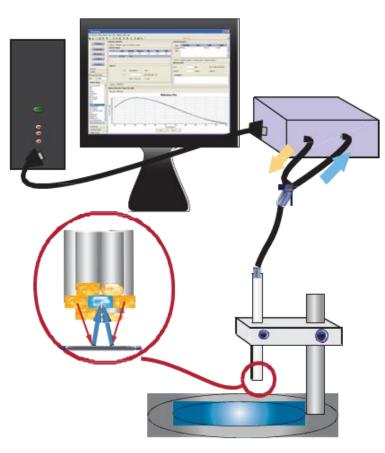
To densify the resist, improve the adhesion to the surface, and make it more resistant to wet chemical etching

9. Inspection of wafer (Thickness measurement)



Inspecting a wafer





Spectrophotometer/Profilometer/Reflectometer

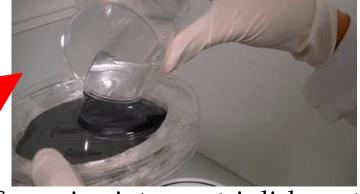
10. Soft-lithography



Degracaine a degracator

Degassing in a desiccator

Poly-Dimethyl Siloxane (PDMS) and Curing agent (10:1)

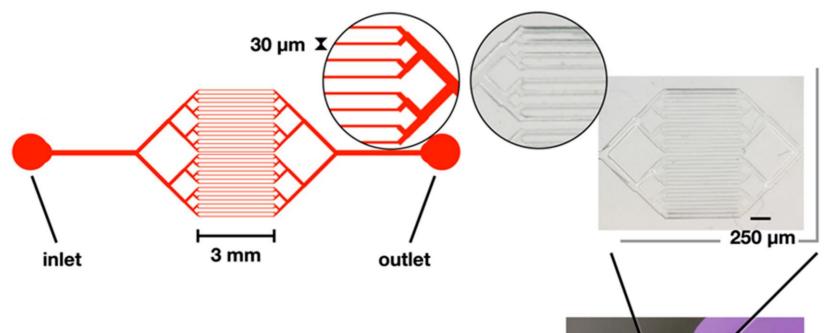


PDMS pouring into a petri-dish containing wafer

Baking for 2 hours at 65°C

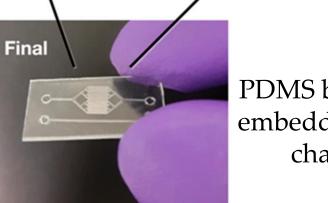


Peeling of solidified PDMS



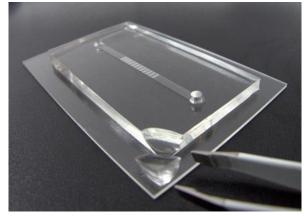
PDMS

- Inert, non-toxic, non-flammable and transparent
- Widely used material in microfluidic research
- Rapid prototyping with minimal cost
- Requires surface treatment to produce strong bond between surface



PDMS block with embedded microchannels

11. Plasma bonding (Air/Oxygen Plasma)

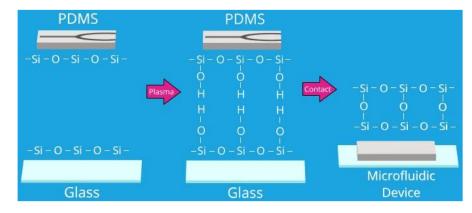




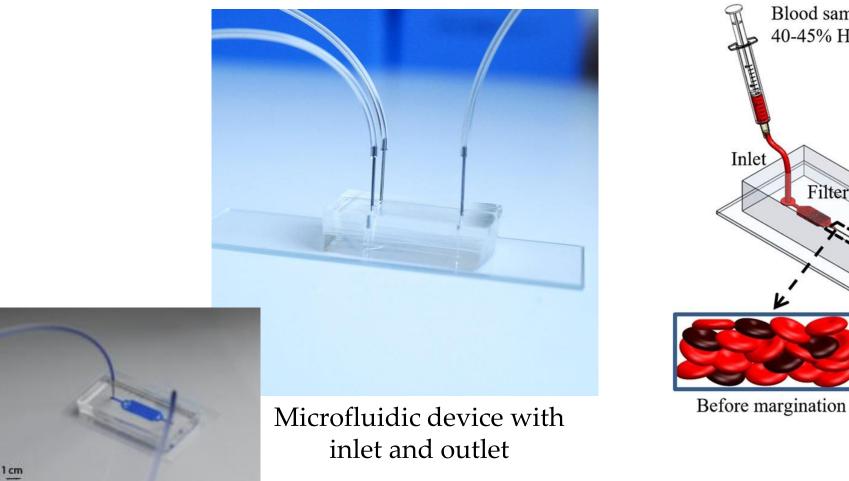


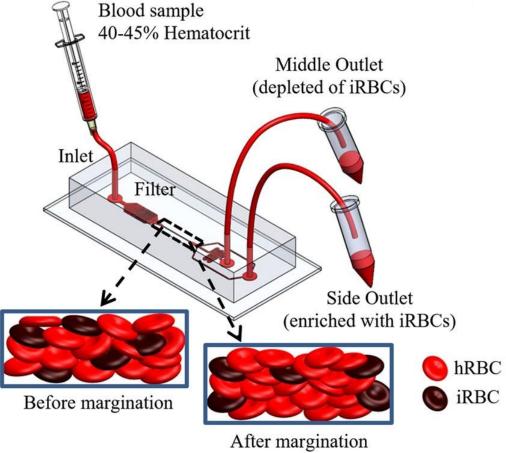
Plasma Cleaner for bonding of hydrophobic surfaces

- PDMS surface oxidises in air/oxygen
- Polar Silanol group (-Si-O-H) formation makes surface hydrophilic
- Si-O-Si bond provides strong bonding



Activation of PDMS and Glass surface

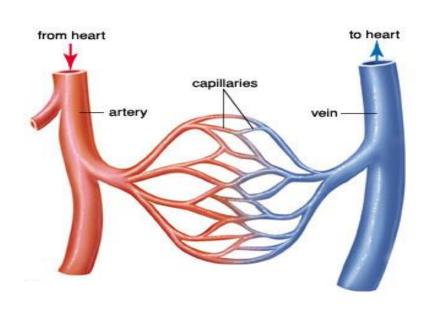


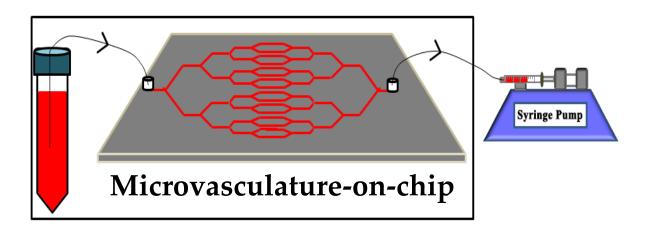


Relevance in Biological Systems

- Cell manipulation (cell response to different biophysical processes)
- Cell separation and sorting
- Drug administration
- Disease detection
- Macromolecular analysis

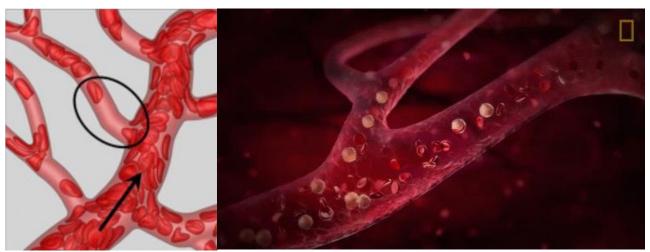
Biomimetics in Lab-On-Chip devices

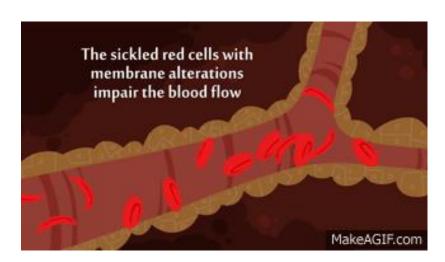


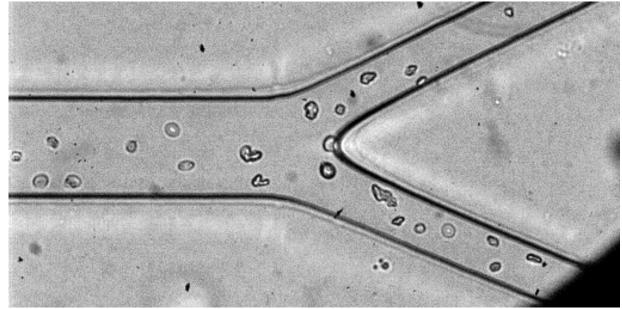


Microdevice that mimics human microvasculature

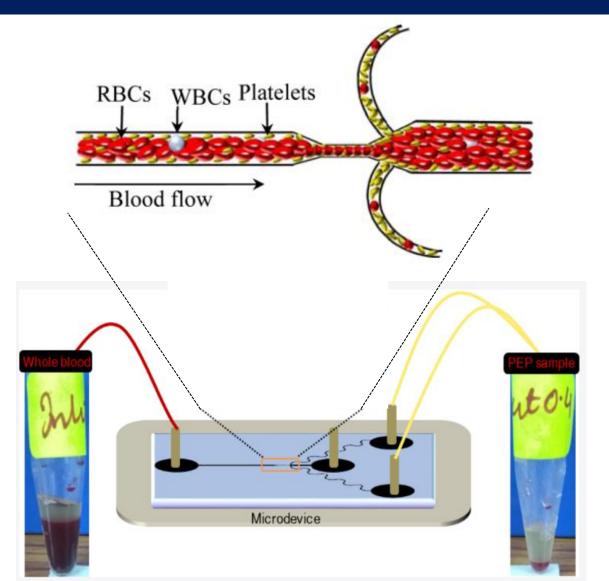
Cell manipulation in Microdevice

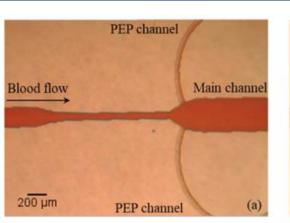


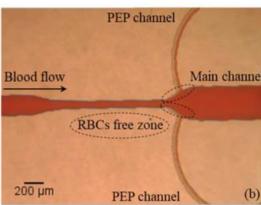


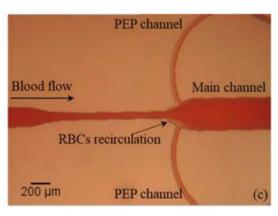


Separation of Platelets from Whole Blood using Microdevice









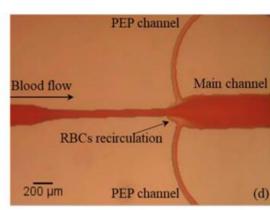
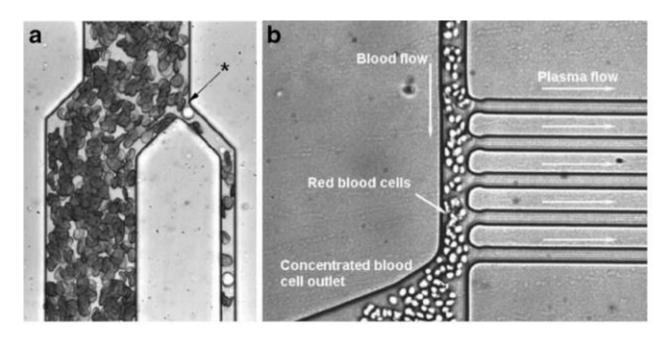


Figure 5. Experimental image of microdevice #2 with the whole blood sample at (a) 0.2 mL/min, (b) 0.4 mL/min flow rate, (c) 0.5 mL/min, and (d) 0.6 mL/min.

(Laxmi V, et. Al., I&EC Research, 2020, 59, 4792-4801)

Separation of WBCs and Plasma from Whole Blood using Microdevice

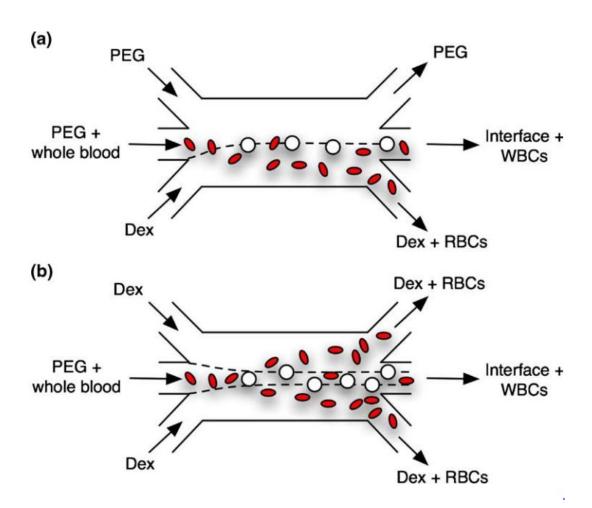


Leukocyte margination is used to separate leukocytes

(Gossett, D. R. et. al., Anal Bioannal Chem, 2010, 397, 3249-3267)

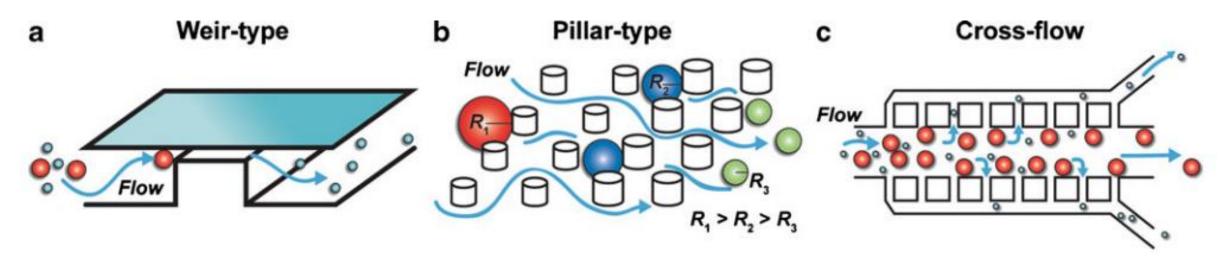
Separation of WBCs and RBCs from Whole Blood using Microdevice

- (a) The leukocytes (WBC) prefer the interface while erythrocytes (RBC) migrate to the Dex.
- (b) Increases in the surface area that blood is exposed to due to two stream interface, resulting in more effective leukocyte concentration



(SooHoo et al. Biomed microdevices 2009, 11, 323-329)

Cell-Separation from Whole Blood using Microdevice of different pattern



Filters size-exclude cellular components while allowing flow of smaller cells and molecules to pass through a planar slit.

Filters are arrays of pillars which exclude cells larger than the spacing of the pillars

Filters are arranged perpendicular to primary channel flow to avoid problems associated with obstructed flow

(Gossett, D. R. et. al., Anal Bioannal Chem, 2010, 397, 3249-3267)

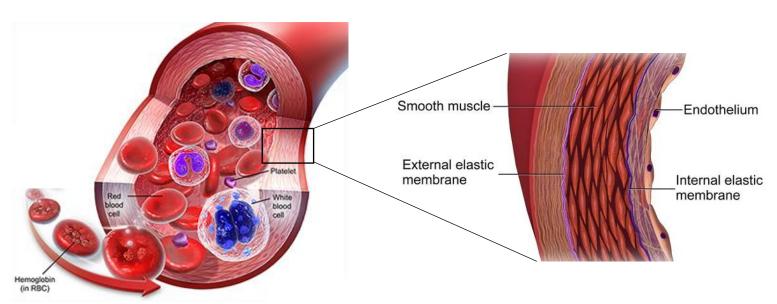
Replacement for Static Cell Assessment

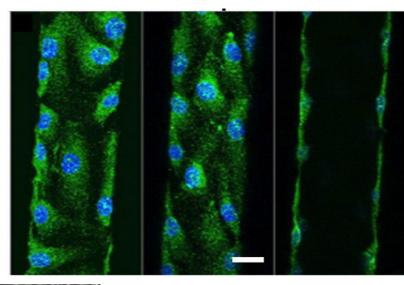


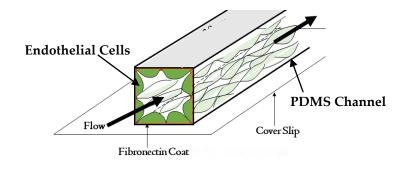


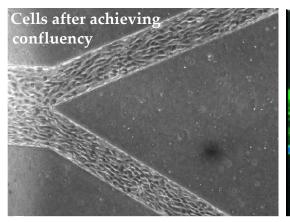
Transwell assay for biological manipulation

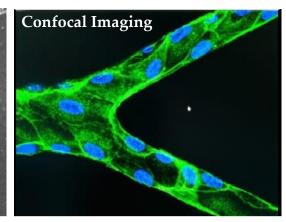
Epithelial cell culture in Microfluidic devices



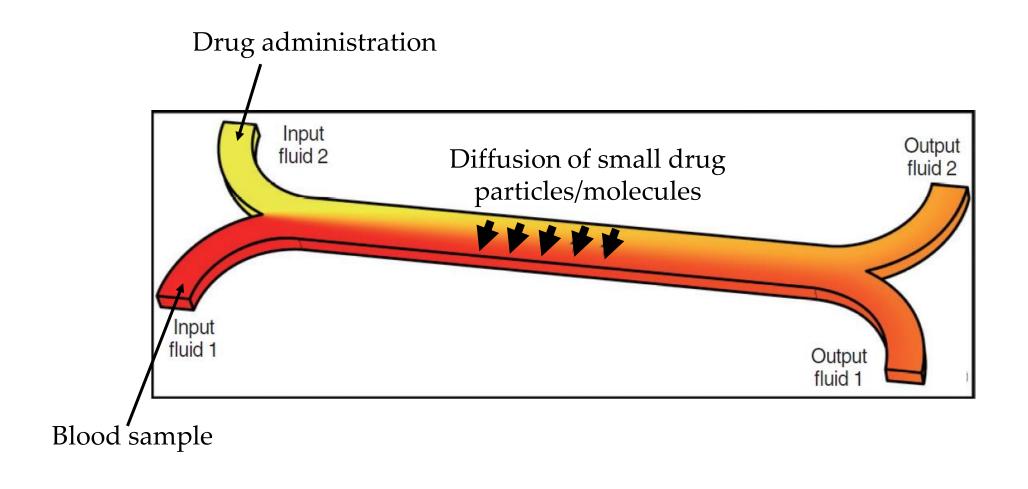




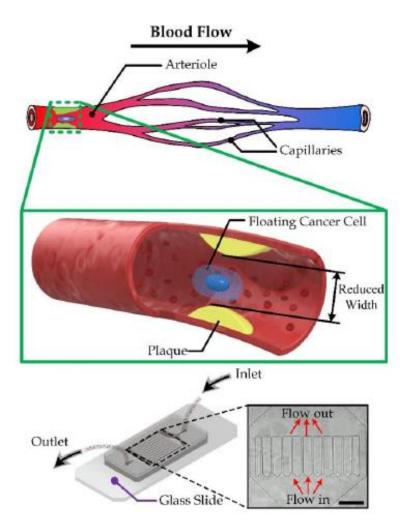




Drug Testing in Microdevices



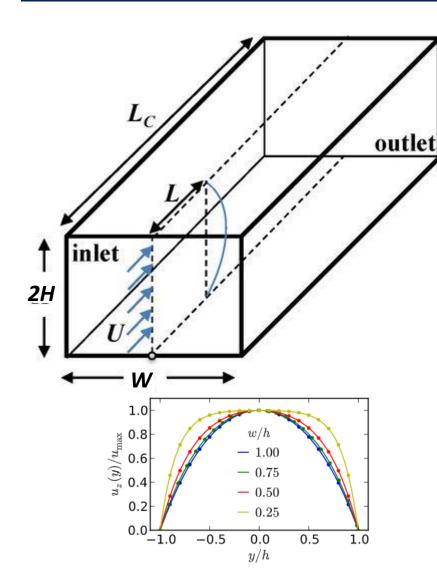
Analysis of transition speed of floating cancer cells during metastasis



Estimation of transiting speed of a deformed cell passing along a constricted narrow channel.

(Ren J, et.al, Micromachines, 2021, 13, 183)

Governing Equation for Microfluidic Flows



Velocity profile for fluid flow through microchannel due to pressure difference:

$$U(y) = \frac{\Delta P}{2\mu L_c} (H^2 - y^2)$$

Where,

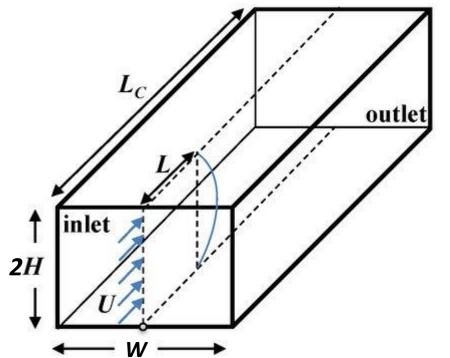
 ΔP – Pressure difference across the two ends of the channel

H – Height of the channel

μ - Fluid viscosity

L_c – Channel length

Governing Equation for Microfluidic Flows



Volumetric flow rate through the micro-channel:

$$Q = \frac{2}{3} \left(\frac{\Delta P}{L_c} \right) \frac{H^3 W}{\mu}$$

Average velocity:

$$\overline{v} = \frac{Q}{2HW}$$

Thank you