

Developing a parsimonious predictor for binary traits in sugar beet (*Beta vulgaris*)

Filson Nazzarini · Simone Marini ·
Piergiorgio Stevanato · Nelppo Biscicari

Received: 05 August 2014 / Accepted:

Abstract Insert your abstract here. Include keywords, PACS and mathematical subject classification numbers as needed.

Keywords binary traits · genomic predictions · parsimonious predictor · sugar beet

1 Introduction

The primary goal of breeding schemes in farm animals and crops is generally to increase the agricultural output. Production traits are typically quantitative continuous variables (e.g. milk yield in dairy cattle, or per hectare yield in maize and rice). Many traits of importance in plant and animal breeding follow nonetheless a discrete categorical distribution, both ordered (e.g. calving ease in cattle, grain texture in rice) and unordered (e.g. grain pigmentation in rice, coat colour in cattle). A special case is that of binomial traits, which can take up only two different values, like disease resistance/susceptibility or

Filippo Biscarini and Nelson Nazzicari contributed equally to the work.

F. Biscarini
Fondazione Parco Tecnologico Padano
E-mail: filippo.biscarini@tecnoparco.org

S. Marini
Dipartimento di Ingegneria Industriale e dell'Informazione
Università di Pavia

N. Nazzicari
Fondazione Parco Tecnologico Padano
E-mail: nelson.nazzicari@tecnoparco.org

P. Stevanato
DAFNE, Università di Padova
24105 Padova, Italy

presence/absence of a morphological characteristic. Annual bolting (flowering) behaviour and root vigor are examples of binomial traits of agronomic importance in sugar beet (*Beta vulgaris*). [move this?]

Advances in biotechnology and genomics, and the advent of high-density molecular markers (especially single-nucleotide polymorphisms, SNP) genotyping have led to the emergence of molecular breeding [12]. One exciting application of molecular breeding is genomic selection: the possibility of predicting the genetic value and future performance of selection candidates solely from their genotypes ([11]). The predictive equations are trained on reference individuals with both genotypic and phenotypic data and then applied to selection candidates with genotypes only. Genomic selection may bring about multiple benefits in breeding programmes such as shorter breeding cycles or more efficient (e.g. traits difficult or expensive to phenotype) and more accurate (e.g. traits with low heritability) estimation of breeding values/selection ([6,8]). Key to the application of genomic selection to breeding programmes are reliable genomic predictions. The recent publication of the reference genome for *Beta vulgaris* genome [4] is facilitating the application of molecular breeding also in this crop species. Pioneering studies on genomic predictions for both continuous ([9,18]) and binary ([2]) traits in sugar beet have already been published.

Genomic predictions are being based on increasing number of molecular markers (e.g. 777K SNP-chip in cattle, 56K SNP-chip in maize, whole-genome sequence data). When a huge number of potential predictors is available, it may be useful to select a subset to limit laboratory and bioinformatics costs, and the time of analysis, while at the same time improving interpretability of results. There is therefore interest in finding the minimum necessary set of information for a specific problem. The principle of parsimony states that a model needs to be simpler than the data it explains (think for instance of K-nearest neighbors -KNN- classifier with $k=1$), and according to Occam's razor, given two models that explain the data equally well, the simplest has to be chosen ([3]).

The objective of this paper is to present the development of a parsimonious predictor for the binary trait root vigor in a population of sugar beet accessions. SNPs in the panel were ranked based on their relevance and used to classify observations: one SNP at a time was removed, progressively reducing the number of SNPs in the predictive model. We found that it was possible to strongly reduce the dimension of the predictor and still achieve high accuracy.

2 Material and methods

2.1 Plant material and SNP genotypes

The available population comprised 123 individual sugar beet (*B. vulgaris*) plants, 100 with high- and 24 with low-root vigor. Root vigor is related to nutrient uptake from the soil and plant productivity ([17]) and is recorded

as a binary trait (either high or low) based on the root elongation rate of eleven-days-old seedlings. No predetermined root elongation rate threshold was used to classify sugar beets into high or low root vigour. The classification was subjectively made upon phenotypic inspection and has nevertheless been shown to be robust over time ([17]). The plant material was provided by Lion Seeds Ltd. (UK).

All plants were genotyped for 192 SNP markers with the high-throughput marker array QuantStudio 12K Flex system coupled with Taqman OpenArray technology. Additional details on the genotyping procedure are described in Stevanato et al., 2013 ([16]). After imputation and editing (call-rate $\geq 95\%$, MAF $\leq 2.5\%$) 175 SNPs were left for the analysis. A more detailed description of SNP genotypes and editing procedure can be found in Biscarini et al. ([2]).

2.2 Predictor development procedure

A two-step approach was adopted for the construction of a parsimonious predictor for root vigor in sugar beet. First, the 175 SNP available for the analysis after data editing were ranked based on their relevance for predicting the trait under study. In the second step the set of predictors was progressively reduced by removing the least useful predictors one at a time. At each iteration logistic regression was used to classify observations with the given set of SNPs. As many classification results as the number of SNPs (i.e. 175) were therefore obtained.

2.2.1 Rank of predictors

When many predictors are available -especially if $p > n$ - it may be of interest to reduce the dimensionality of the problem by choosing the optimal subset of predictors that best describe the relationship between dependent and independent variables [or that best model the response variable, or are most informative with respect to the outcome to be predicted, etc ...]. A Bayesian model selection method, the binary outcome stochastic search (BOSS) algorithm [14], was applied to identify the best set of predictive SNPs by repeated sampling in a Markov Chain Monte Carlo (MCMC) approach. SNPs were ranked based on their probability of inclusion in the best predictive model.

In BOSS the relationship between binary observations and predictors is described by a Gaussian latent variable model with a probit link function:

$$P(Y = [0/1]|X) = \phi(X\beta) \quad (1)$$

where $P(Y = [0/1]|X)$ is the probability of having low or high root vigor given the SNP genotypes X , β is a vector of regression coefficients, and ϕ is the normal cumulative distribution function.

Priors!!

Prior distributions assigned to regression coefficients and model size, to specify prior belief on model complexity. Metropolis-Hastings sampling algorithm (check the Theory that would not die). BOSS extensively explores the model space to identify relevant predictors (sort of best model selection).

β is assigned a multivariate Gaussian prior.

[Describe latent variables? See if this is needed for variable selection and BOSS algorithm!]

The selection of predictors was performed by introducing a vector of indicator variables $\gamma = (\gamma_1, \dots, \gamma_p)$ such that $\gamma = 0$ if $\beta = 0$ and $\gamma = 1$ if $\beta \neq 0$. A predictor was included in the model if its regression coefficient was not null and the associated indicator variable $\gamma = 1$. The model space to be searched was therefore given by the 2^p possible combinations of SNPs (included/excluded).

[How are the γ chosen/determined?]

Equation 2 can be re-written as $\mathbf{Z} = \alpha \mathbf{1} + \mathbf{X}_\gamma \beta_\gamma + \epsilon$ and includes only predictors for which $\gamma = 1$ (α is the intercept).

γ obtained through Binary Outcome Stochastic Search (BOSS): a sampling scheme from $f(\gamma|Z)$.

Z : latent variables ($Z \sim N(X^T \beta)$). The threshold (latent variable) model is summarized in equation 2

$$Y_i = \begin{cases} 0 & Z_i \leq 0 \\ 1 & Z_i > 0 \end{cases} \quad \mathbf{Z} = \mathbf{X}\beta + \epsilon \quad (2)$$

Posteriors for β through Gibbs sampling (again, see the Theory that would not die).

1. initialize Z
2. sample prior for γ
3. sample $f(\beta_\gamma|\gamma, \sigma^2)$
4. sample $f(Z|Y, \beta_\gamma)$
5. restart from 2 until m iterations are completed

The first step in our approach is to rank the SNPs by their informativeness on the studied phenotype trait. To do so we used one of the outputs of the BOSS algorithm [14]. This algorithm is designed to target binary traits, and performs a model search by sampling the predictors on the basis of a general and efficient Markov Chain Monte Carlo (MCMC) technique that exploits the conjugacy structure of data and parameters.

The relationship between the observed responses and the available predictors is described by a latent variable model with a probit link. Prior distributions are assigned both to the regression coefficients and the model size, therefore allowing the user to specify a prior belief on the model complexity.

The algorithm produces a XXX

2.2.2 Selection of predictors and classification method

We take one predictor out at each iteration You put the model formula for ridge logistic regression

2.2.3 Predictive ability

Cross validation: how many times, what fractions. Explanation of error rate and other parameters (ROC?)

2.3 Comparison with another method to rank predictors

Another ranker: why use one, and its description. P value and SNP effect (as it is done in GWAS)

SNP variance [5]

2.4 Software

R [13], weka [7], perl.

3 Results

Supplementary table with BOSS rank of SNPs. A few comments on the most important SNPs (which ones, on which chromosomes, how many per chromosome ...).

Figure 1: Accuracy as a function of the number of predictors, BOSS vs logistic [improve plot: no need to go down to 0.0 in the y-axis; legend names and position; color of the lines? The “bump” at around 20-30 SNPs is not visible]

Table 1: TER, FPR, FNR for the first 30/35 SNPs + average for the rest of the SNP (error close to 0). BOSS + GWAS (6 columns)

Probability of assignment as a function of predictors: Figure 2. Better a table? Maybe in discussion?

From ROC curves only the AUC. No plot, use AUC as result in the text (e.g. comparison between ranker: overall average AUC, average AUC per # SNPs + std). Table?

4 Discussion

General overview why error rates are not evenly distributed? Reminder: it works very well because of LD and H2

Unstable below 30/40 SNPs; little “bump” around 20 SNPs: more marked with BOSS, but also visible with GWAS. Why there? SNPs with large effect on the trait, but low significance? SNPs with large effect but low LD (with the QTL)? In the latter case, the marker might sometimes be in the opposite phase. Look also at marker frequency.

Based on results, a panel of 30-35-40 SNPs is recommended for accurate prediction of root vigor (move to breeding applications? Together with development of a custom-chip?)

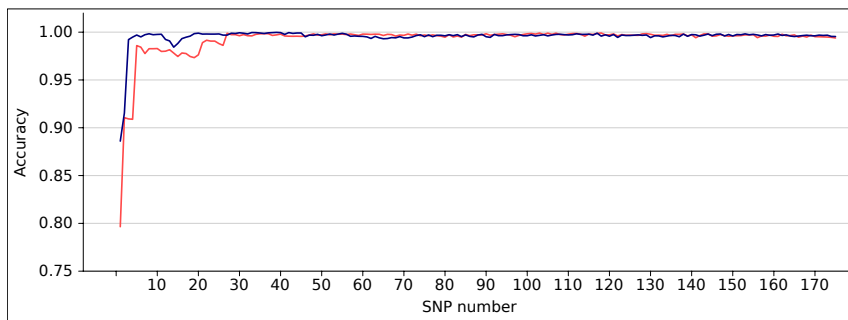


Fig. 1 Accuracy (1 - error rate) of prediction as a function of the number of SNPs included in the classifier: BOSS (blue line) vs logistic regression (red line)

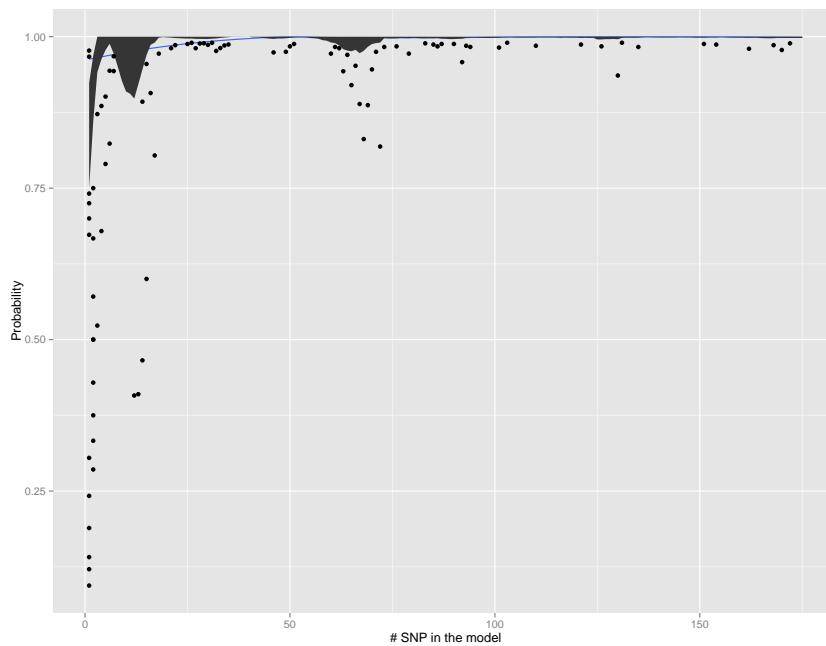


Fig. 2 Distribution of $P(Y = 1|x)$ as a function of the number of SNPs in the classifier

4.1 Relative performance of rankers

why using Pvalues and not other standard rankers (e.g. backward stepwise selection)? Because of the specific nature of the problem

Comparison of rankers: spearman correlation + plot (ranker1 vs ranker2).

Figure 4

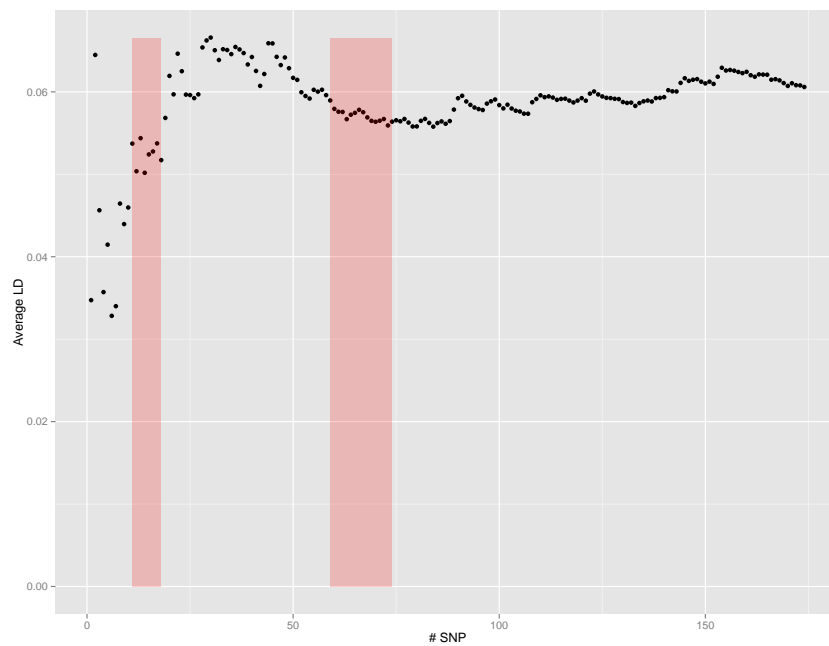


Fig. 3 Average linkage disequilibrium (LD) for increasing number of SNPs in the predictive model

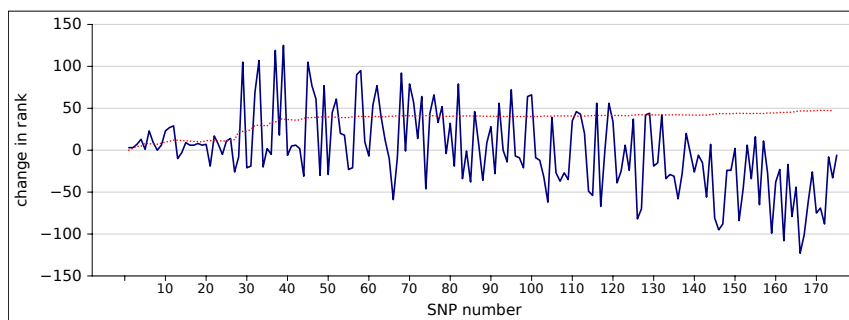


Fig. 4 Comparison of BOSS and logistic regression in terms of relative rank position of relevant SNPs

4.2 SNP effects

Manhattan plot with BOSS weights and weights from the other articles, somehow compared (same chart? two charts? only ten best?).

Do the peaks make sense from the biological perspective?

Variance of SNPs vs genetic variance: → missing heritability? (cite Brachi 2011, Manolio 2009?).

BOSS probability: 1 big peak + smaller peaks. Compare against SNP density? Maybe the big peak corresponds to a physically isolated SNP, whereas

smaller peaks correspond to a cluster of SNPs in LD which individually account for a smaller part of the variability, but together play an important predictive role.

4.3 Genotyping strategies and applications to breeding

Several technology choices are commonly available when genotyping strategies must be decided. Assuming knowledge of SNP flanking sequences, we examine four options: SNP chips, genotyping by sequencing (GBS), targeted sequencing (TS), and a commercial solution, Illumina BeadXpress.

Genotyping by sequencing is

genotyping strategies: Costs, possible technologies (GBS, SNP chip, microarrays), implications

applications to breeding: why is it important root vigor early detection. Other binomial traits (e.g. disease resistance) May be applied to bolting (another trait which exhibits binomial distribution), which has been shown to be controlled by multiple genes and influenced by environmental factors ([1]).

sugar beet: 30% of world's sugar production (cite Dohm? FAO?). Root vigor linked to yield.

Sugar beet: sugar + energy (citation?)

Other binomial traits: resistance to viral and fungal diseases, bolting (cite Dohm? Someone else?)

Breeding has shaped the genome of sugar beet (comparison with *Beta maritima*, [4]).

Extensions to multinomial traits? Examples?

potential and challenges of genomic selection in plant breeding ([10])

5 Conclusions

Concluding remarks.

Acknowledgements This research was financially supported by the Marie Curie European Reintegration Grant "NEUTRADAPT".

References

1. Abou-Elwafa, S., Büttner, B., Kopisch-Obuch, F., Jung, C., Müller, A.: Genetic identification of a novel bolting locus in *Beta vulgaris* which promotes annuality independently of the bolting gene *B*. *Molecular Breeding* **29**, 989–998 (2012)
2. Biscarini, F., Stevanato, P., Broccanello, C., Stella, A., Saccomani, M.: Genome-enabled predictions for binomial traits in sugar beet populations. *BMC Genetics* **18**(5), 1–9 (2014)
3. Chaitin, G.: The limits of reason. *Scientific American* **294**(3), 74–81 (2006)

Table 1 Total error rate (TER), false positive (FPR) and false negative (FNR) rates as a function of the number of SNPs ranked according to BOSS or logistic regression

| # SNPs | TER | FPR | FNR |
|---------|-------|-------|-------|
| 1 | 0.114 | 0.065 | 0.049 |
| 2 | 0.085 | 0.037 | 0.047 |
| 3 | 0.008 | | |
| 4 | 0.005 | | |
| 5 | 0.003 | | |
| 6 | 0.005 | | |
| 7 | 0.003 | | |
| 8 | 0.002 | | |
| 9 | 0.003 | | |
| 10 | 0.002 | | |
| 11 | 0.002 | | |
| 12 | 0.008 | | |
| 13 | 0.009 | | |
| 14 | 0.016 | | |
| 15 | 0.012 | | |
| 16 | 0.007 | | |
| 17 | 0.005 | | |
| 18 | 0.004 | | |
| 19 | 0.002 | | |
| 20 | 0.001 | | |
| 21–30 | 0.002 | | |
| 31–40 | 0.001 | | |
| 41–50 | 0.004 | | |
| 51–60 | 0.002 | | |
| 61–70 | 0.003 | | |
| 71–80 | 0.004 | | |
| 81–90 | 0.002 | | |
| 91–100 | 0.001 | | |
| 101–175 | 0.001 | | |

4. Dohm, J.C., Minoche, A.E., Holtgräwe, D., Capella-Gutiérrez, S., Zakrzewski, F., Tafer, H., Rupp, O., Sørensen, T.R., Stracke, R., Reinhardt, R., et al.: The genome of the recently domesticated crop plant sugar beet (*beta vulgaris*). *Nature* (2013)
5. Gianola, D., de Los Campos, G., Hill, W.G., Manfredi, E., Fernando, R.: Additive genetic variability and the bayesian alphabet. *Genetics* **183**(1), 347–363 (2009)
6. Goddard, M., Hayes, B.: Genomic selection. *Journal of Animal Breeding and Genetics* **124**(6), 323–330 (2007)
7. Hall, M., Frank, E., Holmes, G., Pfahringer, B., Reutemann, P., Witten, I.H.: The weka data mining software: an update. *ACM SIGKDD explorations newsletter* **11**(1), 10–18 (2009)
8. Heffner, E.L., Lorenz, A.J., Jannink, J.L., Sorrells, M.E.: Plant breeding with genomic selection: gain per unit time and cost. *Crop science* **50**(5), 1681–1690 (2010)
9. Hofheinz, N., Borchardt, D., Weissleder, K., Frisch, M.: Genome-based prediction of test cross performance in two subsequent breeding cycles. *Theoretical and Applied Genetics* **125**(8), 1639–1645 (2012)
10. Jonas, E., de Koning, D.J.: Does genomic selection have a future in plant breeding? *Trends in biotechnology* **31**(9), 497–504 (2013)
11. Meuwissen, T., Hayes, B., Goddard: Prediction of total genetic value using genome-wide dense marker maps. *Genetics* **157**(4), 1819–1829 (2001)
12. Moose, S.P., Mumm, R.H.: Molecular plant breeding as the foundation for 21st century crop improvement. *Plant physiology* **147**(3), 969–977 (2008)

13. R Development Core Team: R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria (2008). URL <http://www.R-project.org>. ISBN 3-900051-07-0
14. Russu, A., Malovini, A., Puca, A.A., Bellazzi, R.: Stochastic model search with binary outcomes for genome-wide association studies. *Journal of the American Medical Informatics Association* **19**(e1), e13–e20 (2012)
15. Saccomani, M., Stevanato, P., Trebbi, D., McGrath, J.M., Biancardi, E.: Molecular and morpho-physiological characterization of sea, ruderal and cultivated beets. *Euphytica* **169**(1), 19–29 (2009)
16. Stevanato, P., Broccanello, C., Biscarini, F., Del Corvo, M., Sablok, G., Panella, L., Stella, A., Concheri, G.: High-throughput rad-snp genotyping for characterization of sugar beet genotypes. *Plant Molecular Biology Reporter* pp. 1–6 (2013)
17. Stevanato, P., Trebbi, D., Saccomani, M.: Root traits and yield in sugar beet: identification of aflp markers associated with root elongation rate. *Euphytica* **173**(3), 289–298 (2010)
18. Würschum, T., Reif, J.C., Kraft, T., Janssen, G., Zhao, Y.: Genomic selection in sugar beet breeding populations. *BMC genetics* **14**(1), 85 (2013)