



ALMA MATER STUDIORUM
UNIVERSITÀ DI BOLOGNA

PHD PROGRAMME IN
Innovative technologies and sustainable use of Mediterranean Sea fishery and
biological resources (FishMed-PhD)

Cycle XXXVII

Settore Consorsuale: 05/B1 – ZOOLOGIA E ANTROPOLOGIA

Settore Scientifico Disciplinare: BIO/05 – ZOOLOGIA

**A comparative and evolutionary approach to
study bivalve sex determination from a
broad-phylogenetic perspective**

Candidate: Filippo Nicolini

PhD Coordinator:
prof. Stefano Goffredo

Supervisor:
prof. Andrea Luchetti

Final exam 2025

Table of contents

Table of contents	ii
List of abbreviations	1
Chapter 1: Introduction	4
1.1 The diversity of sexual processes	4
1.2 Genetic sex determination and the evolution of sex-determining genes	5
1.3 Sex determination in bivalves: a long-standing enigma	6
Chapter 2: Bivalves as emerging model systems to study the mechanisms and evolution of sex determination: a genomic point of view.	9
2.1 Open yet inspiring topics in bivalve sex determination	11
2.1.1 Transitions between environmental and genetic sex determination	11
2.1.2 Evolution of sex chromosomes	12
2.1.3 Mito-nuclear interactions	13
2.1.4 Evolution of sex-determination related genes	15
2.2 The case of the Dmrt gene family in bivalves	16
2.3 Conclusions: bivalves as new models in the study of sex determination	19
2.4 Acknowledgments	26
2.5 Data Availability	26
Chapter 3: Identification of putative sex-determination related genes in bivalves through comparative molecular evolutionary analyses	27
3.1 Introduction	28
3.2 Materials and Methods	31
3.2.1 Dataset of bivalve annotated genomes and transcriptomes	31
3.2.2 Identification and classification of Dmrt, Sox and Fox genes in bivalves .	31
3.2.3 Sequence diversity of bivalve single-copy orthogroups	33
3.2.4 Mammals and <i>Drosophila</i> spp. as test datasets	34
3.2.5 GO-term enrichment	35
3.3 Results	36
3.3.1 Genomic and transcriptomic datasets	36
3.3.2 The Dmrt, Sox, and Fox complements in bivalves	36

3.3.3 Amino acid sequence divergence of Dmrt, Sox, and Fox genes in bivalves	38
3.3.4 Dmrt, Sox, and Fox genes, and amino acid sequence divergence in the test datasets	46
3.4 Discussion	47
3.4.1 A new manually-curated and phylogenetic-based reference dataset of Dmrt, Sox, and Fox genes in bivalves	47
3.4.2 High amino acid sequence divergence identifies putative sex-determining genes	51
3.5 Conclusions	56
Chapter 4: Localisation of three sex-related genes and the germline marker <i>Vasa/Vasa</i> in the early developmental stages of <i>Mytilus galloprovincialis</i> . . .	58
4.1 Introduction	59
4.2 Materials and Methods	61
4.2.1 Time-series gene expression	61
4.2.2 Sample collection, MitoTracker staining and fixation	62
4.2.3 mRNA <i>in-situ</i> hybridization chain reaction (HCR)	63
4.2.4 Immunolocalization of Vasa	65
4.3 Results	66
4.3.1 Differential gene expression analysis of <i>Vasa</i> and SRGs in embryo time-series	66
4.3.2 mRNA <i>in-situ</i> HCR of <i>Vasa</i> and SRGs	69
4.4 Discussion	70
References	92
Appendix	93
Supplementary materials	99
Supplementary figures	99
Supplementary tables	114
Data availability	192

List of abbreviations

AASD	amino acid sequence divergence
CMS	cytoplasmatic male sterility
CUE	coupling of ubiquitin conjugation to endoplasmic reticulum degradation [domain]
DEAD/DEAH-box	Asp-Glu-Ala-Asp/Asp-Glu-Ala-His box
DGE	differential gene expression
dpf	days post fertilization
DM	<i>dsx</i> and <i>mab-3</i> [domain]
DMA	DM-associated [domain]
Dmrt	<i>doublesex</i> (<i>dsx</i>) and <i>mab-3</i> related transcription factor
Dmrt-1L	<i>Dmrt 1-like</i>
Dm-W	<i>dsx</i> and <i>mab-3</i> related gene <i>W</i>
Dmy	<i>dsx</i> and <i>mab-3</i> related gene <i>Y</i>
DSFG	Dmrt, Sox, and Fox gene
dsx	<i>doublesex</i>
DUI	doubly uniparental inheritance
ESD	environmental sex determination
FASW	filtered artificial sea water
FHA	forkhead-associated [domain]
Fox	forkhead box
GC	germ cell
GO	gene ontology
GRN	gene regulatory network
GSD	genetic sex determination

HCR	hybridization chain reaction
HeSC	heteromorphic sex chromosome
HMG	high mobility group [box domain]
HMM	hidden Markov model
HoSC	homomorphic sex chromosome
hpf	hours post fertilization

<i>mab-3</i>	<i>male abnormal-3</i>
MCL	Markov clustering algorithm
ML	maximum likelihood
mRNA-ISH	mRNA <i>in-situ</i> hybridization
Mya	million years ago

ORF	open reading frame
------------	--------------------

PBS	1× PBS with 0.1% Tween 20
PBS-Tw	1× PBS with 0.1% Tween 20
PCA	principal component analyses
PFA	paraformaldehyde
PGC	primordial germ cell

qRT-PCR	quantitative real-time polymerase chain reaction
----------------	--

RNAi	RNA interference
RT	room temperature

SC	sex chromosome
SCO	single-copy orthogroup
SD	sex determination
SDf	sex differentiation

SDG	sex-determining gene
Sox	<i>Sry</i> -related HMG-box
SRG	sex-determination related gene
<i>Sry</i>	<i>Sex-determining region of chromosome Y</i>
SSC-Tw	5× saline-sodium citrate with 0.1% Tween 20
<i>Sxl</i>	<i>Sex-lethal</i>
 TBS	1× Tris-buffered saline
TBS-Tx	1× TBS with Triton X-100
<i>tra</i>	<i>transformer</i>

Chapter 1

Introduction

1.1 The diversity of sexual processes

The process of sex determination (SD) has been traditionally associated with the very first step of gonad differentiation, where an initial trigger activates the molecular pathway that establishes organism sex. According to this view, two alternative types of SD have been recognized at first: the environmental sex determination (ESD) and the genetic sex determination (GSD), depending on whether the very first cues are of environmental or genetic origin. Conversely, all the downstream events of gonad development (i.e., after SD) have been appointed as sex differentiation (SDf), which consists of the entire set of morphogenetic, molecular, and physiological events leading to the full maturation of testes or ovaries (**Uller and Helanterä, 2011; Beukeboom and Perrin, 2014**). Lately, however, the dichotomous views of ESD/GSD and of SD/SDf have been questioned. On the one hand, a growing number of studies on non-model organisms proved that ESD and GSD represent a continuum of mixed conditions rather than two mutually exclusive phenomena. On the other, the high evolutionary dynamics and the variable expression patterns of the genes involved in the processes of gonad commitment and development make the distinction between SD and SDf of unclear utility (**Beukeboom and Perrin, 2014**).

Considering this complex scenario, **Uller and Helanterä, 2011** proposed a unified and broad-scope definition for SD, that is, “the processes within an embryo leading to the formation of differentiated gonads as either testes or ovaries”, without any actual distinction between environmental/genetic initial triggers or the downstream effectors. However, I argue that this definition should be expanded to encompass not only the embryonic stage of the animal life cycle

but also adulthood, since cases of sex reversals and sex changes (sequential hermaphroditism) legitimately express proper SD processes during post-embryonic life stages as well.

1.2 Genetic sex determination and the evolution of sex-determining genes

In its most intimate core, animal SD is the manifestation of complex gene regulatory networks where, in accordance with the Wilkins' theory (1995), the downstream actors appear to be nearly conserved both from functional and identity point of views, while the master top regulators (the commonly recognized sex determinants, such as the *Sex-determining region of chromosome Y (Sry)* in therians or the ratio between sex and autosome chromosomes in *Drosophila*) are often the most variable part (**Beukeboom and Perrin, 2014**). As a matter of fact, this evolutionary pattern of animal sex-determining cascades has been observed in major animal clades, including vertebrates (e.g., **Marshall Graves and Peichel, 2010**), insects (e.g., **Verhulst et al., 2010**), and nematodes (e.g., **Stothard and Pilgrim, 2003**).

Sex-determination related genes (SRGs) are of particular interest not only from a regulatory point of view but also because of their patterns of molecular evolution. In fact, transcriptionally sex-biased genes (including SRGs) often tend to evolve faster than unbiased genes at the level of protein sequences. In particular, male-biased genes generally show higher rate of sequence evolution in comparison to both female-biased and unbiased counterparts (reviewed in **Parsch and Ellegren, 2013; Grath and Parsch, 2016**), as it has been repeatedly observed in well-studied organisms such as fruit flies (e.g., **Meisel and Connallon, 2013**), nematodes (e.g., **Cutter and Ward, 2005**), mice (e.g., **Kousathanas et al., 2014**) and primates (e.g., **Khaitovich et al., 2005**), and in other emerging model systems, such as *Daphnia pulex* (**Eads et al., 2007**), aphids (**Purandare et al., 2014**), and two wasp species of the genus *Nasonia* (**Wang et al., 2015**). Growing evidence is however showing cases in which instead female-biased genes have higher rates of sequence evolution than male-biased genes, such as in mosquitoes of the genus *Anopheles* (**Papa et al., 2017**), and European and Manila clams of the genus *Ruditapes* (**Ghiselli et al., 2018**).

The pattern of molecular evolution of sex-biased genes is particularly evident in organisms with sex chromosomes (both in XY/ZW and X0 systems), such as fruit flies, birds and mammals,

where the so-called fast-X (or fast-Z) effect has been extensively reported for sex-chromosome associated genes (**Vicoso and Charlesworth, 2006; Mank et al., 2007; Meisel and Connallon, 2013**). This high rate of sequence evolution in sex-biased genes and sex chromosomes (SCs) can be the result of both adaptative and non-adaptative processes, since the observed higher ratio between non-synonymous and synonymous mutations (dN/dS) can be caused by natural selection, sexual selection or sexual antagonism, as well as genetic drift (**Vicoso and Charlesworth, 2006; Meisel and Connallon, 2013; Parsch and Ellegren, 2013; Grath and Parsch, 2016**).

1.3 Sex determination in bivalves: a long-standing enigma

Bivalves are the second largest clade in molluscs, counting more than 18,000 species (Catalogue of Life) distributed at all depths and in all marine environments, as well as in some freshwater habitats. Thanks to their high diversity and biological peculiarities, they have been proposed as promising model organisms for investigating a wide array of biological, ecological and evolutionary issues (**Milani and Ghiselli, 2020; Ghiselli et al., 2021**). However, despite their socio-economic and scientific importance, the knowledge concerning the molecular basis of bivalve reproduction and SD is still quite limited (**Breton et al., 2018**). Clues from various works seem to suggest that both genetic and environmental factors (e.g., temperature, food availability, and steroids) are involved in SD, and that heteromorphic sex chromosomes (HeSCs) are absent (**Breton et al., 2018; Han et al., 2022**). However, the exact process by which sex is determined and gonad commitment is established is, currently, still unknown. Actually, bivalves represent a dazzling example of how the traditional dichotomies between ESD/GSD and SD/SDf can sometimes hamper scientific research, as many bivalve species exhibit various forms of hermaphroditism and because a master environmental or genetic sex determinant inducing SDf may just not exist.

In the attempt to identify SRGs, many differential gene expression analyses have been recently performed on a variety of species covering most of the phylogenetic diversity of bivalves (e.g., **Milani et al., 2013; Zhang et al., 2014; Chen et al., 2017; Capt et al., 2018; Ghiselli et al., 2018; Shi et al., 2018**). Some of the genes that were found to be differentially expressed between gonads of different sex were systematically retrieved across species, such as those belonging to the *dsx* and *mab-3* related transcription factor (Dmrt),

Sry-related HMG-box (Sox), and forkhead box (Fox) families, which act in concert in various animal developmental processes including the SD cascade (**Marshall Graves and Peichel, 2010; Beukeboom and Perrin, 2014**). To this regard, **Zhang et al., 2014** proposed a working model for the sex-determining pathway of the Pacific oyster *Crassostrea gigas* in which: *CgSoxH* promotes male gonad development by activating *CgDsx*, which belong to the Dmrt family, and inhibiting *CgFoxL2*; *CgFoxL2*, when not inhibited by the pair *CgSoxH/CgDsx*, promotes female gonad development. Moreover, **Han et al., 2022** recently identified homomorphic sex chromosomes (HoSCs) in eight scallop species and appointed *FoxL2* as a putative SRG in *Patinopacten yessoensis* and *Chlamys farreri*. Though, much of the recent research effort on bivalve SRGs has been limited to their molecular cloning, differential transcription, and tissue localization (**Liang et al., 2019; Sun et al., 2022**). Furthermore, few works have directly investigated the biological functions of Dmrt, Sox, and Fox genes in bivalves so far, and most used post-transcriptional silencing of target mRNAs [RNA interference (RNAi)]. **Liang et al., 2019** studied the role of *Sox2* in the spermatogenesis of the Zhikong scallop *C. farreri* and found that it likely regulates proliferation of spermatogonia and apoptosis of spermatoocytes, since its knockdown resulted in the loss of male germ cells. **Wang et al., 2020** proposed that in the female gonads of the freshwater mussel *Hyriopsis cumingii*, *FoxL2* might be related to the *Wnt/β-catenin* signaling pathway, which takes part in ovarian differentiation also in vertebrates. **Sun et al., 2022** found instead that in *C. gigas*, *FoxL2* and *Dmrt1L* mRNA knockdown results in the size reduction of female and male mature gonads, respectively.

In this sense, bivalve molluscs represent a striking example of the difficulty to reconcile the traditional view of a single sex determinant with an apparent multifactorial model in which many genes and environmental cues act in concert to establish the sexual identity of the individual (**Breton et al., 2018**). Lately, much effort has been put in the characterisation of bivalve SD and a general framework is eventually taking shape. Functional assays with RNAi and CRISPR-Cas9 techniques (e.g., **Wang et al., 2020; Sun et al., 2022; Wang et al., 2022**), as well as with mRNA *in-situ* hybridization (mRNA-ISH) and immunohistochemistry (e.g., **Perez-Garcia et al., 2011; Milani et al., 2013**), are making their way into the study of bivalve biology and have been proved essential instruments also for the investigation of sex-related traits. However, very few works have made extensive use of the comparative and integrative approach in bivalve studies so far, which hampers the possibility to infer general patterns for such a vast class of organisms (**Milani and Ghiselli, 2020**). The high evolutionary

rates and plasticity of SRGs make the situation even harder, since phylogenetic and orthology inferences can lead to erroneous reconstructions in the presence of signal saturation and high sequence divergence (reviewed in **Natsidis et al., 2021; Lozano-Fernandez, 2022**).

Chapter 2

Bivalves as emerging model systems to study the mechanisms and evolution of sex determination: a genomic point of view

Filippo Nicolini^{1,2}, Fabrizio Ghiselli¹, Andrea Luchetti¹, Liliana Milani¹

¹*Department of Biological, Geological and Environmental Science, University of Bologna, Bologna (BO), Italy.*

²*Fano Marine Center, Fano (PU), Italy.*

Published in: 2023, *Genome Biology and Evolution*, 15(10):evad181.

10.1101/gbe/evad181

Abstract. Bivalves are a diverse group of molluscs that have recently attained a central role in plenty of biological research fields, thanks to their peculiar life history traits. Here we propose that bivalves should be considered as emerging model systems also in sex-determination studies, since they would allow to investigate: (i) the transition between environmental and genetic sex determination, with respect to different reproductive backgrounds and sexual systems (from species with strict gonochorism to species with various forms of hermaphroditism); (ii) the genomic evolution of sex chromosomes, considering that no heteromorphic sex chromosomes are currently known and that homomorphic sex chromosomes have been identified just in few species of scallops; (iii) the putative role of mitochondria at some level of the sex determination signaling pathway, in a mechanism that may resemble the cytoplasmatic male sterility of plants;

(iv) the evolutionary history of sex-determination related gene families with respect to other animal groups. In particular, we think that this last topic may lay the foundations for expanding our understanding of bivalve sex determination, as our current knowledge is quite fragmented and limited to few species. As a matter of fact, tracing the phylogenetic history and diversity of sex-determination related gene families (such as the Dmrt, Sox and Fox genes) would allow to perform more targeted functional experiments and genomic analyses, but also fostering the possibility of establishing a solid comparative framework.

Significance. In this perspective, we provide an examination of the phylogenetic diversity of Dmrt genes, a sex-determination related gene family, to address the importance of bivalves in sex determination studies. By analyzing their taxonomic distribution and sequence diversity, we show how such a comparative study may set a common ground plan to settle down targeted functional experiments and essays. This kind of approach should be applied more extensively in future studies, especially when dealing with understudied organisms.

Bivalves are the second largest clade in molluscs, counting more than 18,000 species (Catalogue of Life, accessed 16/12/2022) distributed at all depths and in all marine environments, as well as in some freshwater habitats. Thanks to their high diversity and peculiar biological features, they have been proposed as promising model organisms for investigating a wide array of biological, ecological, and evolutionary issues, from mitochondrial biology and evolution to the physiological plasticity under fluctuating environmental conditions (**Milani and Ghiselli, 2020; Ghiselli et al., 2021**). In this context, bivalves may serve as a compelling model system to investigate the evolution and characteristics of sex determination (SD) as well, thanks to the diversity of their reproductive modes and genomic features. Nonetheless, this research field has been largely overlooked and many aspects of bivalve reproductive biology remain uncharacterized. In this perspective, we address the topic by first examining the relevant questions that bivalves may help to answer regarding processes and patterns of SD, and then providing a case study in the field of comparative genomics.

2.1 Open yet inspiring topics in bivalve sex determination

Despite the socio-economic and scientific importance of bivalves, the knowledge concerning the genetic and molecular bases of their SD system is quite limited and its study has been mostly neglected. Yet, bivalves may constitute a novel model system in SD studies that is as intriguing and valuable as other well-established models, such as vertebrates, insects and plants (**of Sex Consortium et al., 2014**), as they may provide complementary perspectives in many aspects of SD evolutionary studies. Topics such as (i) the transition between environmental and genetic SD, (ii) the evolution of sex chromosomes, (iii) the mito-nuclear interaction, and (iv) the evolution of SD related genes, can largely benefit from the integration with bivalve studies. But many others are likely to emerge as research in the field progresses.

2.1.1 Transitions between environmental and genetic sex determination

Clues from several works seem to suggest that both genetic and environmental factors are involved in bivalve SD, thus implying that a mixed system may exist (reviewed in **Breton et al.,**

2018). The traditional dichotomy between environmental sex determination (ESD) and genetic sex determination (GSD) seems inapplicable in most bivalve species, where ESD and GSD rather represent the two ends of a continuum of mixed and plastic conditions. A weak distinction between ESD and GSD is also found in amphibians, reptiles and teleost fish, three clades in which environment-dependent SD has been largely studied. Here, the interaction—or even the transition—between the two sexual systems have been reported in many species, suggesting that sex-determining mechanisms can be extraordinary plastic (**Bachtrög et al., 2014; Capel, 2017**). Adding a representative and diverse group of Lophotrochozoa (Protostomia) to those vertebrate taxa, can widely expand the comparative framework of the investigation, allowing to better understand the evolution of SD as a whole. In bivalves, ESD has been studied mostly in oysters, where hermaphroditic species show an effect of temperature on SD (reviewed in **Breton et al., 2018; Fig. 2.1**). Oysters may indeed constitute a prolific model to examine how the SD pathways are shaped in the presence of different initial triggers and highly dynamic reproductive backgrounds. In fact, various sexual systems can be found in oysters, such as (i) strictly gonochoric population, (ii) the coexistence of simultaneous hermaphroditic with strictly gonochoric individuals in the same population, (iii) the possibility of sex change according to environmental conditions, and (iv) the presence of both parasitic dwarf males and free-living males in the same species (**Collin, 2013**). Consequently, oysters may be extremely useful to understand how epigenetic control is involved in sex change, how gene regulatory networks can sustain the occurrence of different hermaphroditic conditions within gonochoric populations, and whether certain SD systems are more labile than others (**Abbott, 2011**).

2.1.2 Evolution of sex chromosomes

So far, heteromorphic sex chromosomes (HeSCs)—i.e., sex chromosomes showing strong morphological differentiation, have never been observed in bivalves (**Breton et al., 2018**), while the first evidence of homomorphic sex chromosomes (HoSCs)—i.e., sex chromosomes showing little or no differentiation, comes from a very recent study on several scallop species, where a non-homologous origin of the SD system has been proposed for different subfamilies (**Han et al., 2022; Fig. 2.1**). Theory predicts that, once originated, sex chromosomes (SCs) will eventually turn into HeSCs, because of the recombination arrest in the sex-determining region (**Bachtrög et al., 2014; Beukeboom and Perrin, 2014; Han et al., 2022**). Nonetheless, HoSCs are much more widespread in the animal kingdom than expected, sometimes also being

of ancient age (**Bachtrog et al., 2014; Han et al., 2022**).

Species from the order Pectinida may thus be useful to investigate what determines the long-term maintenance of HoSCs and which genomic architectures and molecular dynamics prevent HeSCs from evolving in bivalves. Additionally, they may be taken as model systems to investigate the origin of SCs in relation to the sexual systems and the route by which molecular pathways have been reprogrammed in the transition between different SD mechanisms (**Han et al., 2022**).

Researchers have been addressing this topic mainly in snakes, ratites and sturgeons (**Bachtrog et al., 2014; Han et al., 2022** and references therein). Though, scallops currently hold the oldest HoSC pairs, which are dated back to about 350 million years. The system is thus of great importance to investigate the role of sex-biased gene expression and selection forces in the long-term stability of SCs (**Han et al., 2022**), as well as the intertwining between SD systems.

2.1.3 Mito-nuclear interactions

An additional pivotal topic in bivalve biology, tentatively connected to SD, regards the doubly uniparental inheritance (DUI) of mitochondria, a process in which two highly divergent mitochondrial genomes are transmitted uniparentally through the maternal and paternal lineages, respectively through eggs and sperm. This process, which has been reported in more than a hundred bivalve species from five different orders (**Fig. 2.1; Gusman et al., 2016; Capt et al., 2020**), has been proposed to interact with the major nuclear pathways that primarily establish the sexual identity, in a way that can resemble the cytoplasmatic male sterility (CMS) of plants (**Ghiselli et al., 2013; Breton et al., 2022**). In CMS, specific mitochondrial chimeric open reading frames (ORFs) cause the pollen to be sterile, while certain nuclear loci act in counterbalance to restore male fertility when occurring in the same individual. This Red-Queen scenario, in which balancing selection shapes the evolution of both CMS and restorer-of-fertility genes and keeps the two sexes viable, has been also hypothesized to be acting on bivalve DUI species (**Ghiselli et al., 2013; Xu, Iannello, et al., 2022**), where additional and effectively-transcribed ORFs have been observed in both the male-inherited and female-inherited mitochondrial lineages (**Milani et al., 2013, 2014**).

Clearly, if a functional interplay between DUI and SD in bivalves is proven, this will provide

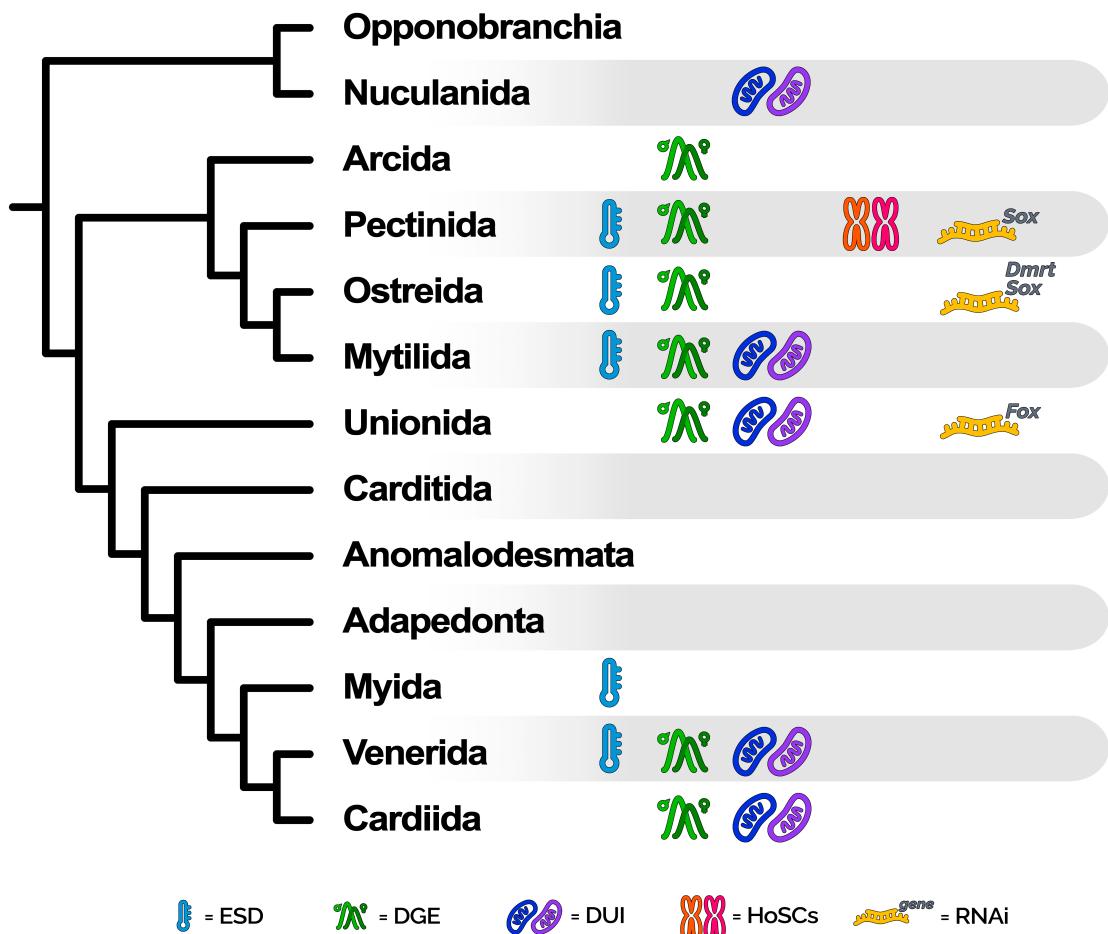


Figure 2.1 – Graphical summary of the available knowledge and experiments concerning the genetic basis of SD in bivalves, at the level of major taxonomic orders (as reported in WoRMS; accessed before or on 14/03/2023). For each bivalve clade it is reported: (i) the availability of records of ESD (ii) the availability of differential gene expression (DGE) experiments specifically intended to investigate sex-biased or sex-specific genes; (iii) whether the DUI of mitochondria has been reported in at least one species; (iv) whether HoSCs have been identified in at least one species; (v) the availability of RNA interference (RNAi) experiments for genes belonging to the Dmrt, Sox, and Fox gene families. The phylogenetic tree on the left has been drawn on the basis of the most widely accepted topology for bivalves, according to analyses based on nuclear markers and morphological data. The tips of the tree correspond to major bivalve orders, except for Opponobranchia and Anomalodesmata, which represent higher-level taxonomic ranks. References for the availability of data and experiments can be found throughout the main test.

new research questions regarding not only bivalve biology itself but also broader evolutionary topics (e.g., are there any converging trait between DUI and CMS systems? What is the degree of plasticity of such mitochondria-related SD systems? Are mitochondria-related SD systems more widespread in eukaryotes than currently thought?).

2.1.4 Evolution of sex-determination related genes

Considering this intricate scenario of SD mechanisms and the wide diversity of bivalves, in the last years many differential transcription analyses have been performed on several species with the attempt to identify the most probable sex-determination related genes (SRGs) (e.g., **Milani et al., 2013; Zhang et al., 2014; Chen et al., 2017; Capt et al., 2018; Shi et al., 2018; Fig. 2.1**). Interestingly, certain genes consistently emerged across different bivalve species as being substantially more transcribed in one sex (sex-biased) or exclusively transcribed in one sex (sex-specific), suggesting their potential involvement in the SD pathway. These genes mainly belong to the *dsx* and *mab-3* related transcription factor (Dmrt), *Sry*-related HMG-box (Sox), and forkhead box (Fox) families, which play a role in various developmental processes (including the SD cascade) in most animals (**Marshall Graves and Peichel, 2010; Bachtrog et al., 2014; Beukeboom and Perrin, 2014**). Members of these three gene families are also included in the working model for the SD regulatory network proposed for the Pacific oyster *Crassostrea gigas* by **Zhang et al. (2014)**, in which: *CgSoxH* (which belong to the Sox family) promotes male gonad development by activating *CgDsx* (which belong to the Dmrt family) and inhibiting *CgFoxL2* (which belong to the Fox family); *CgFoxL2*, when not inhibited by the pair *CgSoxH/CgDsx*, promotes female gonad development. Similarly, **Han et al. (2022)** appointed *FoxL2* as a putative SD gene in the two scallop species *Patinopacten yessoensis* and *Chlamys farreri*. If their pivotal role in SD of bivalves is confirmed, an evolutionary genomic analysis may help in better understanding why members of the above-mentioned gene families appear particularly prone to be recruited in the SD cascade also in distantly related species, as it is observed for *Dmrt1* and *Sox3* homologs in vertebrates (**Marshall Graves and Peichel, 2010; Bachtrog et al., 2014**; and the following section). Furthermore, considering the occurrence of mixed SD systems in bivalves, Dmrt, Sox, and Fox genes may provide new perspectives on the influence of different environmental cues on the molecular evolution of animal SRGs. However, to date, experiments have been limited to molecular cloning, differential transcription, and tissue localization of such genes (**Liang et al., 2019; Sun et al., 2022**), while only a few have directly investigated their biological functions in bivalves, for example through post-transcriptional silencing of target mRNAs (RNAi; **Fig. 2.1**; e.g., **Liang et al., 2019; Wang et al., 2020; Sun et al., 2022**).

Overall, Dmrt, Sox, and Fox genes are highly interesting targets to be investigated in the framework of bivalve SD and have indeed obtained much more attention than the study of SCs

or the role of environmental cues. However, much work is still to be done in order to understand their function in the SD signaling pathway and their evolutionary history.

2.2 The case of the Dmrt gene family in bivalves

Among the SRG candidates identified in bivalves, Dmrt genes (named after *doublesex* (*dsx*) from *Drosophila melanogaster* and *male abnormal-3* (*mab-3*) from *Caenorhabditis elegans*) are of particular interest. As a matter of fact, in vertebrates, besides their role in placode neurogenesis and somite patterning (reviewed in **Mawaribuchi et al., 2019**), Dmrt genes are also involved in the development of male gonads and the maintenance of the testicular function (**Sun et al., 2022**). Their role in the specification and organization of male sexual characters seems indeed to be common across Metazoa, suggesting that a similar function may have been already present in the Bilateria common ancestor (**Kopp, 2012; Beukeboom and Perrin, 2014**).

The first attempts to dig inside the phylogenetic history and diversity of bivalve Dmrt genes have been provided by **Li et al. (2018)** and **Evensen et al. (2022)**: besides retrieving all the canonical genes (i.e., *Dmrt2*, *Dmrt3* and *Dmrt4/5*), their inferences brought to light a monophyletic Dmrt group (named *Dmrt 1-like* (*Dmrt-1L*)) which appears to be private to molluscs and present in several bivalve species. The *Dmrt-1L* monophyletic group is confirmed also when expanding the analysis by mining genomes from a wider range of bivalve taxa (**Tab. 2.1; Fig. 2.2A**), suggesting that *Dmrt-1L* genes are widespread in bivalves and were likely present in their common ancestor (**Evensen et al., 2022**). In particular, *Dmrt-1L* genes can be successfully retrieved in species of the orders Mytilida, Ostreida, Pectinida, Unionida, and from *Scapharca broughtonii* (Arcida), while the opposite holds for Venerida, *Sinonovacula constricta* (Adapedonta), and *Dreissena* spp. (Myida; **Fig. 2.2B**). Clearly, the absence of *Dmrt-1L* genes demands further investigations, as it may derive from errors in genome assembly and annotations.

The present analysis also supports a higher amino acid sequence divergence of the *Dmrt-1L* orthology group with respect to the other Dmrt orthology groups (**Fig. 2.2C**), which may be explained by a higher rate of sequence evolution related to their sex-biased expression in certain species (**Zhang et al., 2014; Shi et al., 2015; Li et al., 2018; Evensen et al., 2022**). This is consistent with what has been already observed for the SRGs *Dmrt1* and *dsx*

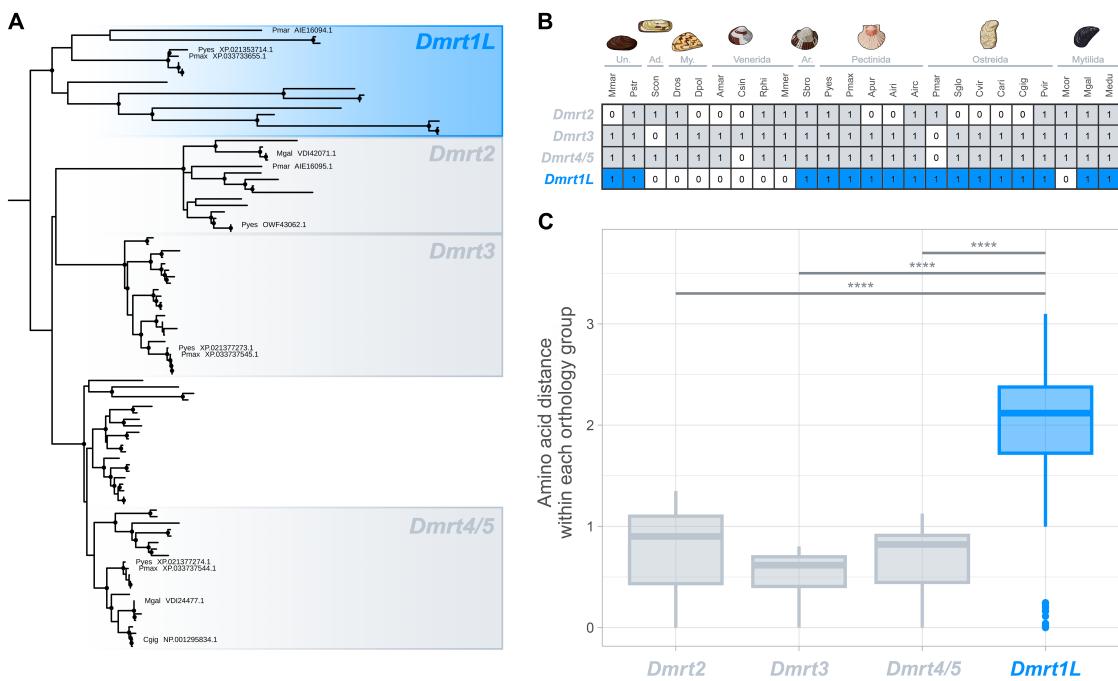


Figure 2.2 – Phylogenetic tree (A) and taxonomic distribution (B) of Dmrt genes in bivalves, and comparison of amino acid pairwise distances within Dmrt-1L and the other Dmrts (C). (A) Dmrt orthologs from bivalve genome assemblies were obtained with HMMsearch (HMMER toolkit; **Eddy, 2011**) with the Pfam HMM profile of the DM domain (PF00751). Amino acid alignment was obtained with MAFFT-DASH (**Rozewicki et al., 2019**), and manually inspected to remove poorly aligning sequences, and trimmed with trimAI (gap threshold of 60%; **Capella-Gutiérrez et al., 2009**). The phylogenetic analysis was carried out using IQ-TREE 2 (**Minh et al., 2020**) with default parameters. Nodes with bootstrap values greater than 84 are marked with filled black circles. The tree was rooted according to **Evenesen et al., 2022**. Dmrt genes analysed by **Evenesen et al., 2022** were used as reference to annotate the various orthology groups, and accession numbers are reported in the tree. The phylogenetic tree with all annotated tips and nodes can be accessed on supplementary material online. (B) Taxonomic distribution of identified Dmrt genes in bivalve genomes. Orders as reported in WoRMS (accessed before or on 14/03/2023) and in **Fig. 2.1** are specified. (C) Pairwise amino acid distances were computed for amino acid sequences within each Dmrt orthology group identified in the tree, with the R package ‘phangorn’ (**Schliep, 2011**) under the JTT substitution model. After checking for normality with the Shapiro-Wilk test ($W = 0.88544$, $p < 2.2 \times 10^{-16}$) and for group effect with the Kruskal-Wallis test ($p < 2.2 \times 10^{-16}$), the pairwise Wilcoxon rank-sum test was used to compare the distributions of pairwise amino acid distances of *Dmrt-1L* and the other Dmrts. Horizontal bars mark the significative results with $p < 2.2 \times 10^{-16}$ (****). The list of genome assemblies used for these analyses and species identifiers can be found in **Tab. 2.1**. Un.: Unionida; Ad.: Adapedonta; My.: Myida; Ar.: Arcida.

in vertebrates and *Drosophila*, respectively (e.g., **Bewick et al., 2011; Baral et al., 2019**). In fact, sex-biased genes (including SRGs) often tend to evolve faster than unbiased genes at

the level of protein sequences, either when considering male-biased (reviewed in **Parsch and Ellegren, 2013; Grath and Parsch, 2016**) or female-biased genes (e.g., **Papa et al., 2017; Ghiselli et al., 2018**). Another possible explanation for the higher amino acid divergence of *Dmrt-1L* genes may lie on their expression breadth, that is, genes with a narrow tissue-specific expression tend to evolve faster than more ubiquitous genes (**Parsch and Ellegren, 2013; Xu, Martelossi, et al., 2022**). As a matter of fact, *Dmrt-1L* genes have been found to be significantly more transcribed in the gonadic tissue (particularly in testes) in *P. yessoensis* (**Li et al., 2018**) and *C. gigas* (**Yue et al., 2021**).

Understanding the role and molecular interactions of *Dmrt-1L* genes in bivalve SD and gonad development would greatly enhance the possibility of outlining the evolutionary causes and consequences of their high amino acid divergence (**Fig. 2.2C**), for example by linking the molecular evolution to the degree of pleiotropy. However, most of our knowledge on *Dmrt-1L* biology is currently limited to the temporal and tissue localization of transcripts in a few species of bivalves (e.g., **Li et al., 2018; Yue et al., 2021**). In fact—apart from the work by **Sun et al. (2022)**, which confirmed the role of *Dmrt-1L* in the gonad development of *C. gigas* through non-invasive RNAi and found that the knocked-down phenotype results in size reduction of male gonads—no other experiments intended to elucidate the function of *Dmrt-1L* genes in bivalves have been carried out so far (**Fig. 2.1**). This clearly hinders any possible integration between molecular data with functional assays. If the role of *Dmrt-1L* as major sex determinants was confirmed, bivalves would become an intriguing clade in which investigate why, in Metazoa, certain genes (namely, the Dmrt gene family) appear particularly prone to being recruited at the top of the SD cascade. To date, this phenomenon has been widely examined in vertebrates, where *Dmrt1* genes have independently gained a primary role in male SD in fish, amphibians, and birds, and are considered candidate sex-determining genes also in monotreme mammals (**Marshall Graves and Peichel, 2010; Beukeboom and Perrin, 2014; Mawaribuchi et al., 2019**). Bivalves may provide an alternative evolutionary scenario to study the selective forces and molecular modifications that support Dmrt genes in repeatedly taking over the SD process. In fact, since *Dmrt-1L* genes seem to be restricted to molluscs (**Fig. 2.2A**), it would be intriguing to clarify if the putative involvement in the SD cascade of extant bivalve species is the result of shared ancestry or convergent evolution, which would establish a study system for the evolution of Dmrt genes parallel to that of vertebrates (see **Capel, 2017**).

Obviously, *Dmrt-1L* should not be expected to be the sole sex-determining gene. In fact,

Fox-L2 has already been appointed as the female sex-determining gene in *P. yessoensis* and *C. farreri* (**Han et al., 2022**). Consequently, we should expect that other primary genetic determinants exist, consistently with the extremely high species diversity of the clade. Thus, bivalves may additionally serve as a valuable model system to study how genes from different families take over the SD cascade and are shaped by selection.

2.3 Conclusions: bivalves as new models in the study of sex determination

SD is undoubtedly a fascinating biological and evolutionary topic as much as it is challenging to investigate. Our understanding of the causes and consequences of the SD mechanism diversity strongly relies on the study of different systems and non-model model organisms (**Bachtrog et al., 2014; Milani and Ghiselli, 2020**), which provide the foundation for depicting a comprehensive evolutionary and comparative framework in which new and coherent research perspectives can be grounded.

In recent years, bivalves have been achieving growing importance in many fields of biology, from ecology to genomics, and from environmental biomonitoring to mitochondrial studies (**Milani and Ghiselli, 2020; Ghiselli et al., 2021**), but they can be a valuable model to address also SD studies. The diversity of their life history traits provides indeed a challenging, yet extremely fascinating framework, to put the SD processes into an evolutionary context.

Bivalves can help us explain how ESD and GSD interplay with each other in response to the environmental conditions, as a mixed system of both has been proposed to act in the establishment of bivalve sexual identity (reviewed in **Breton et al., 2018**). Moreover, the occurrence of the many existing variants of hermaphroditism and gonochorism even in closely related species, or within the same population, strongly suggests that the basic SD pathway (whether genetic, environmental, or mixed) should be plastic enough to sustain the existence of individuals of both sexes, thus providing the opportunity to study how SD gene regulatory networks are shaped and selected throughout evolution and how epigenetic regulation may influence SD. The unique DUI system further poses an undeniable challenge in SD studies since it may represent an SD-linked mechanism which relies on the non-nuclear portion of the genome and may unfold many new research paths (**Milani and Ghiselli, 2020; Ghiselli**

et al., 2021). Nonetheless, much of the research effort on bivalve SD has been devolved to specific groups of socio-economic importance, such as Mytilida, Ostreida, Pectinida, and Unionida, while the other lineages of the bivalve phylogeny have been neglected (Fig. 2.1). Our understanding of the SD processes of bivalves is thus restricted and is mainly lacking a broad comparative framework in which to draw comprehensive evolutionary inferences.

Genes from the Dmrt, Sox and Fox families, which are involved in SD also in other Metazoa, may be considered excellent genomic targets to study the processes and patterns of molecular evolution in sex-biased genes, as well as of the recurrent recruitment of genes in the SD cascade. Also, identifying the major genetic regulators of SD in bivalves would burst the functional study of the interaction between ESD and GSD, by providing genetic targets that can be manipulated through RNAi and/or genome editing techniques to understand the role of environmental cues in SD. In the same way, knowing the main genetic actors of SD would allow researcher to identify SCs not only on the basis of in-silico techniques (such as k-mer based or SNP methods) but also by less-expensive wet lab protocols (such as fluorescence mRNA *in-situ* hybridization [mRNA-ISH] on metaphase chromosome plates). Furthermore, it would help to understand whether and how the mitochondrial additional ORFs of DUI species interact with the SD system, by performing thorough gene expression essays.

In conclusion, we strongly urge researchers to invest more resources in the integrative study of bivalve SD to unravel the many underlying mechanisms and expand our understanding of this biological process. Given our limited knowledge in the field, one of the first routes that should be undertaken may rely on the comparative study of SRGs of bivalves from a genomic perspective, as this kind of data is nowadays growing at a rate faster than ever. Establishing such a genomic ground plan for understudied organisms will in fact allow researchers to develop evolutionary-aware experiments with better selected genetic targets.

Table 2.1 – List of bivalve genomes from which Dmrt genes have been extracted. For each species, the accepted name and the most-common synonym (in parentheses) are reported. NCBI accession numbers are provided, when available, as well as BUSCO scores of the predicted proteomes against the ‘metazoa_odb10’ dataset (Manni et al., 2021).

Species	ID	Order	Assembly level	BUSCO score	Reference	NCBI Acc. No.
<i>Anadara (Scapharca) broughtonii</i>	Sbro	Arcida	Chromosome	C:91.2% [S:85.6%,D:5.6%] F:2.6%	Bai et al., 2019	NA
				M:6.2%		
<i>Sinonovacula constricta</i>	Scon	Adapedonta	Chromosome	C:92.5% [S:80.4%,D:12.1%] F:3.4%	Ran et al., 2019	GCA_007844125.1
				M:4.1%		
<i>Dreissena polymorpha</i>	Dpol	Myida	Chromosome	C:86.9% [S:75.1%,D:11.8%] F:6.4%	McCartney et al., 2022	GCA_020536995.1
				M:6.7%		
<i>Dreissena rostriformis</i>	Dros	Mytilida	Scaffold	C:75.2% [S:73.2%,D:2.0%] F:15.2%	Calcino et al., 2019	GCA_007657795.1
				M:9.6%		
<i>Mytilus unguiculatus (coruscus)</i>	Mcor	Mytilida	Chromosome	C:80.0% [S:79.1%,D:0.9%] F:7.7%	Yang et al., 2021	GCA_017311375.1
				M:12.3%		

Tab. 2.1 continued from previous page

Species	ID	Order	Assembly level	BUSCO score	Reference	NCBI Acc. No.
<i>Mytilus edulis</i>	Medu	Mytilida	Scaffold	C:83.7% [S:64.5%,D:19.2%] F:5.2% M:11.1%	Corrochano-Fraile et al., 2022	GCA_905397895.1
<i>Mytilus galloprovincialis</i>	Mgal	Mytilida	Scaffold	C:80.3% [S:47.5%,D:32.8%] F:8.8% M:10.9%	Gerdol et al., 2020	GCA_900618805.1
<i>Perna viridis</i>	Pvir	Mytilida	Scaffold	C:99.4% [S:99.0%,D:0.4%] F:0.2% M:0.4%	Inoue et al., 2021	GCA_018327765.1
<i>Magallana (Crassostrea) ariakensis</i>	Cari	Ostreida	Chromosome	C:94.6% [S:90.9%,D:3.7%] F:0.9% M:4.5%	Li et al., 2021	GCA_020567875.1
<i>Magallana (Crassostrea) gigas</i>	Cgig	Ostreida	Chromosome	C:98.5% [S:67.6%,D:30.9%] F:0.3% M:1.2%	Penaloza et al., 2021	GCF_902806645.1

Tab. 2.1 continued from previous page

Species	ID	Order	Assembly level	BUSCO score	Reference	NCBI Acc. No.
<i>Crassostrea virginica</i>	Cvir	Ostreida	Chromosome	C:98.1% [S:58.6%,D:39.5%] F:0.3% M:1.6%	Gómez-Chiarri et al., 2015	GCF_002022765.2
<i>Saccostrea glomerata</i>	Sglo	Ostreida	Scaffold	C:88.9% [S:85.3%,D:3.6%] F:5.1% M:6.0%	Powell et al., 2018	GCA_003671525.1
<i>Argopecten irradians concentricus</i>	Airc	Pectinida	Scaffold	C:94.8% [S:93.9%,D:0.9%] F:3.7% M:1.5%	Liu et al., 2020	GCA_004382765.1
<i>Argopecten irradians irradians</i>	Airi	Pectinida	Scaffold	C:94.8% [S:93.9%,D:0.9%] F:3.7% M:1.5%	Liu et al., 2020	GCA_004382745.1
<i>Argopecten purpuratus</i>	Apur	Pectinida	Scaffold	C:89.2% [S:88.5%,D:0.7%] F:5.0% M:5.8%	Liu et al., 2020	NA

Tab. 2.1 continued from previous page

Species	ID	Order	Assembly level	BUSCO score	Reference	NCBI Acc. No.
<i>Pecten maximus</i>	Pmax	Pectinida	Chromosome	C:98.5% [S:74.7%,D:23.8%] F:0.4% M:1.1%	Kenny et al., 2020	GCF_902652985.1
<i>Mizuhopecten (Patinopecten) yessoensis</i>	Pyes	Pectinida	Scaffold	C:98.6% [S:75.2%,D:23.4%] F:0.4% M:1.0%	Wang, Zhang, et al., 2017	GCF_002113885.1
<i>Margaritifera margaritifera</i>	Mmar	Unionida	Scaffold	C:92.6% [S:82.3%,D:10.3%] F:3.2% M:4.2%	Gomes-dos-Santos et al., 2021	GCA_015947965.1
<i>Potamius streckersoni</i>	Pstr	Unionida	Scaffold	C:74.7% [S:73.8%,D:0.9%] F:7.0% M:18.3%	Smith, 2021	GCA_016746295.1
<i>Calyptogena (Archivesica) marissinica</i>	Amar	Venerida	Chromosome	C:82.0% [S:80.0%,D:2.0%] F:6.1% M:11.9%	Ip et al., 2021	GCA_014843695.1

Tab. 2.1 continued from previous page

Species	ID	Order	Assembly level	BUSCO score	Reference	NCBI Acc. No.
<i>Cyclina sinensis</i>	Csin	Venerida	Scaffold	C:94.0% [S:83.8%,D:10.2%] F:1.9% M:4.1%	Wei et al., 2020	GCA_012932295.1
<i>Mercenaria mercenaria</i>	Mmer	Venerida	Chromosome	C:95.4% [S:70.9%,D:24.5%] F:0.5% M:4.1%	Song et al., 2021	GCF_014805675.1
<i>Ruditapes philippinarum</i>	Rphi	Venerida	Chromosome	C:83.4% [S:74.5%,D:8.9%] F:8.8% M:7.8%	Xu, Martelossi, et al., 2022	GCA_026571515.1

2.4 Acknowledgments

The authors are extremely thankful to Sofía Blanco González from the University of Vigo for her willingness to engage in discussions and for genuinely sharing her opinion on this work.

2.5 Data Availability

Analyzed data and R scripts used to generate plots can be accessed in supplementary material online deposited at the following GitHub repository: [filonico/bivalve_sex_perspective](https://github.com/filonico/bivalve_sex_perspective).

Chapter 3

Identification of putative sex-determination related genes in bivalves through comparative molecular evolutionary analyses

Filippo Nicolini^{1,2}, Mariangela Iannello¹, Giovanni Piccinini¹, Sergey Nuzhdin³,
Fabrizio Ghiselli¹, Andrea Luchetti¹, Liliana Milani¹

¹*Department of Biological, Geological and Environmental Science, University of Bologna, Bologna (BO), Italy.*

²*Fano Marine Center, Fano (PU), Italy.*

³*Department of Molecular and Computational Biology, University of Southern California, Los Angeles, CA, USA.*

In preparation.

3.1 Introduction

In sexually reproducing organisms, the modes of sex determination (SD), i.e., the process by which the male or female identity of an organism (or of the gonadic tissue) is established, is highly diverse, ranging from strictly genetic systems to environmentally-dependent processes (Haag and Doty, 2005; Uller and Helanterä, 2011; Bachtrog et al., 2014; Beukeboom and Perrin, 2014). Characterising the molecular basis of SD is crucial for understanding not only reproductive biology but also the evolutionary pressures shaping these systems (Wilkins, 1995; Ellegren and Parsch, 2007; Grath and Parsch, 2016; Nicolini, Ghiselli, et al., 2023), as sex-determination related genes (SRGs), including primary sex-determining genes (SDGs), are those responsible for the phenotypic differences of males and females, thanks to their sex-biased expression and interactions (Ellegren and Parsch, 2007; Beukeboom and Perrin, 2014; Grath and Parsch, 2016). One key aspect of SRGs is that they often exhibit accelerated rates of sequence evolution, due to their involvement in sex-related traits and reproduction. This represents the effects of sexual and/or adaptive selection, which act in sex-biased genes and produce high-divergent proteins at the interspecific level (Civetta and Singh, 1998; Ellegren and Parsch, 2007; Meisel, 2011; Grath and Parsch, 2016). Rapid sequence evolution is known for *Sex-determining region of chromosome Y (Sry)* of therians (Pamilo and O'Neill, 1997; Mawaribuchi et al., 2012), *dsx* and *mab-3* related gene *W (Dm-W)* of the African clawed frog *Xenopus laevis*, and *dsx* and *mab-3* related gene *Y (Dmy)* of the medaka fish *Oryzias latipes* (Mawaribuchi et al., 2012), all of which are master SDGs, that is, genes whose expression is primarily responsible for the establishment of the sexual fate of the organism. Evolution under episodic diversifying selection has been detected also in *Drosophila* for genes involved in the SD cascade (e.g., *Sex-lethal [Sxl]* , *transformer [tra]* , and *doublesex [dsx]*), in correspondence with its establishment in the genus common ancestor (Mullon et al., 2012; Baral et al., 2019); though, rapid sequence evolution seems to not be concerning extant amino acid sequences (Haerty et al., 2007; Baral et al., 2019), as they are globally evolving under purifying selection, especially in their catalytic domain (Mullon et al., 2012; Baral et al., 2019). Concerning the *dsx* genes, higher rates of nucleotide and amino acid sequence evolution can be however observed for male-specific regions, if compared to female-specific and oligomerization regions (Baral et al., 2019).

While SD has been extensively studied in model organisms, like mammals, insects, and nematodes, comparatively little is known about the molecular ground plans in non-model organisms. A remarkable example of this is represented by bivalve molluscs, which exhibit a wide variety of reproductive strategies and sexual systems (Breton et al., 2018). Notwithstanding the considerable importance in the human socio-economic landscape (reviewed in Haszprunar and Wanninger, 2012; Gomes-dos-Santos et al., 2020), the study of SD mechanisms in bivalves has been hampered by the striking divergence among species (Li et al., 2022), and thus largely overlooked and limited to few case studies (Breton et al., 2018; Nicolini, Ghiselli, et al., 2023). So far, no master SDG has been unambiguously identified, and the only working hypothesis on the functioning of the SD gene regulatory network is available for the Pacific oyster *Crassostrea gigas* (now *Magallana gigas*; Zhang et al., 2014). Nonetheless, the field still lacks both a robust functional investigation and an evolutionary framework in which to place the current knowledge (Nicolini, Ghiselli, et al., 2023). As a matter of fact, major efforts have been dedicated to identify sex-biased genes through differential gene expression (DGE) analyses (e.g., Milani et al., 2013; Teaniniuraitemoana et al., 2014; Zhang et al., 2014; Capt et al., 2018; Afonso et al., 2019), but very few have leveraged cutting-edge techniques to investigate their actual role in SD and/or gonad differentiation and development (e.g., Liang et al., 2019; Sun et al., 2022).

Components of the Dmrt, Sox, and Fox gene (DSFG) families are notoriously known as key actors in several developmental processes across Metazoa (Benayoun et al., 2011; Matson and Zarkower, 2012; Sarkar and Hochedlinger, 2013; Mawaribuchi et al., 2019), including SD in certain clades: the aforementioned *Dm-W*, *Dmy*, and *dsx* all belong to the *dsx* and *mab-3* related transcription factor (Dmrt) gene family, while *Sry* belongs to the *Sry*-related HMG-box (Sox) gene family; *Fox-L2*, which takes part in most of the vertebrate SD processes as a downstream effector of the female pathway, belongs to the forkhead box (Fox) gene families. Members of the DSFGs have been identified as putative SRGs also in bivalves, thanks to both DGE analyses and mRNA *in-situ* hybridization (mRNA-ISH) (e.g., Naimi et al., 2009; Li et al., 2018; Liang et al., 2019; Yue et al., 2021), suggesting that their role in morphological and sexual development is maintained also in the clade. However, the clear role of DSFGs has yet to be elucidated, probably as a consequence to the lack of (i) a systematic classification of the families and (ii) a comprehensive understanding of their evolutionary history.

In order to overcome such limitations, this study aims to perform a thorough investigation of

the DSFG families in bivalves, with the attempt to provide a high-quality resource to be used as a reference for future studies. Through the analysis of more than 40 annotated bivalve genomes and transcriptomes, we aim (i) to describe the complete set and evolutionary history of DSFGs in bivalves by means of phylogenetic inferences, manual curation, and orthology prediction; furthermore, we aim (ii) to identify DSFGs potentially involved in bivalve SD by investigating their sequence evolution in a genome-wide context. As a matter of fact, our hypothesis is that, if any of the DSFGs is directly involved in SD (i.e., is a SDG), then we should expect it to be experiencing a higher rate of sequence evolution, as already found in previous studies (**Pamilo and O'Neill, 1997; Mawaribuchi et al., 2012**) and discussed earlier; this characteristic, in turn, would be reflected in a high diversity of the extant amino acid sequences across the bivalve clade. To assess the robustness and reliability of our approach, we additionally applied our pipeline to two non-bivalve datasets, composed of mammal and *Drosophila* species, respectively (hereon referred to as the ‘mammal dataset’ and the ‘fruit fly dataset’). By choosing two clades for which SD is well characterised, we wanted to compare our results with those obtained on taxa for which a deeper and detailed knowledge is available. Particularly, mammals and *Drosophila* provide two different frameworks to study the patterns of molecular evolution in SDGs: the former is a system where SD is completely genetic (i.e., the development into a male or into a female is triggered by the up- or downregulation of *Sry* in undifferentiated gonads, respectively), while the latter is a system where SD is chromosomal, thus lacks a master SDG (the sexual fate of the individual is determined by the ratio between autosomal and X chromosomes). Hence, they represent opposing control datasets to be compared to bivalves, as it is expected that a higher rate of sequence evolution concerns only master SDGs (as *Sry* in therians; i.e., the top regulatory part of the SD cascade), but not also the downstream genes (i.e., the bottom effectors). If our method is robust, we should thus expect that, (i) in the mammalian dataset *Sry* is detected as rapidly-evolving, while (ii) in the fruit fly dataset no gene among those working within the sex-determining cascade is evolving at a higher pace. By testing the performance of the pipeline in mammals and fruit flies, we were able to assess the reliability of results in bivalves.

This work offers novel insights into the evolutionary dynamics of SRGs and contributes a valuable genomic resource for understanding SD in bivalves, one of the most ecologically and economically important groups of marine organisms. Particularly, here we provide the first extensive phylogenetic-based classification of DSFGs in bivalves, covering many species from

the major bivalve orders, along with a comprehensive investigation of their sequence evolution.

3.2 Materials and Methods

3.2.1 Dataset of bivalve annotated genomes and transcriptomes

Annotated genome assemblies of bivalves were obtained from various publicly available resources, while reference genome assemblies for gastropods and cephalopods were downloaded from NCBI (**Tab. S1**). Isoforms were removed from genome annotations using a perl script from the AGAT toolkit (v0.8.0; **Dainat et al., 2022**). Concerning *Sinonovacula constricta* (Adapedonta), the nucleotide coding sequence fasta file was not available for download. To avoid excluding the species from our analyses, the file was generated in-house by mapping the annotated protein sequences on the reference genome using miniprot (v0.13-0; **Li, 2023**). Then, the corresponding nucleotide sequences were extracted using AGAT on the resulting gff annotation file.

In order to provide an extensive identification of SRGs also for underrepresented bivalve orders (mainly belonging to the Heterodonta clade), 14 additional species represented by sequenced transcriptomes were included in the analyses. Assembled and annotated transcriptomes were obtained from **Piccinini et al., 2021** and **Iannello et al., 2023**. Briefly, raw reads were trimmed using Trimmomatic (**Bolger et al., 2014**) and assembled using Trinity (**Grabherr et al., 2011**) with default parameters. Isoforms were removed using the dedicated perl script from the Trinity utilities. Open reading frames were predicted through TransDecoder (**Haas, n.d.**), by also including diamond (**Buchfink et al., 2015**) and HMMER (v3.3.2; <http://hmmer.org/>) annotation of hits.

The resulting set of annotated genomes and transcriptomes (hereafter referred to as the “comprehensive set”) was checked for completeness using BUSCO with the Metazoa reference dataset (v5.2.2; **Manni et al., 2021**).

3.2.2 Identification and classification of Dmrt, Sox and Fox genes in bivalves

Members of DSFG families were retrieved in the comprehensive set with hmmsearch from the HMMER package (v3.3.2; <http://hmmer.org/>). The signature catalytic domains of each

family were used as queries. Specifically, hidden Markov model (HMM) profiles were built after the Pfam databases for the *dsx* and *mab-3* (DM) domain (PF00751), the high mobility group (HMG) box (PF00505) and the forkhead domain (PF00250) to retrieve members of the DSFG families, respectively. The e-value for both the per-target and the per-domain inclusion threshold was set to 1.0×10^{-5} .

Obtained hits were then annotated using (i) the PANTHER HMM standalone sequence scoring against the PANTHER library v18.0 and (ii) RPS-BLAST (v2.5.0+) against the Conserved Domain Database (CDD; pre-compiled version, downloaded from ftp.ncbi.nih.gov on 09/11/23). In both cases, hits with an e-value of 1.0×10^{-5} were retained. Genes which were correctly annotated by both systems (on the basis of the PANTHER gene family and CDD domain identifiers; **Tab. S2**) were kept for subsequent analyses.

DSFGs from *Homo sapiens*, *Drosophila melanogaster*, and *Caenorhabditis elegans* (**Tab. S3**; hereafter referred to as ‘reference species’) were retrieved from NCBI and were used as reference genes for annotation (see below). Classification and nomenclature of each family was retrieved from: **Mawaribuchi et al. (2019)** for Dmrt genes; **Phochanukul and Russell (2010)** and **Sarkar and Hochedlinger (2013)** for Sox genes; **Mazet et al. (2003)** for Fox genes.

The alignments of mollusc and reference DSFGs were guided by the aforementioned Pfam HMM profiles and performed with Clustal Omega (v1.2.3; **Sievers et al., 2011**), then trimmed with trimAl (v1.4.rev15; **Capella-Gutiérrez et al., 2009**) with a gap threshold of 40%. Resulting alignments were manually inspected to remove sequences with incomplete catalytic domains, then aligned and trimmed again as before. Phylogenetic trees were inferred using IQ-TREE (v2.1.4-beta COVID-edition; **Minh et al., 2020**) with automatic model selection (**Kalyaanamoorthy et al., 2017**), 1000 bootstrap replicates and 5 independent runs. The phylogenetic tree of Dmrt genes was midpoint rooted, as no clear homology relationship has been found with other gene families or zinc-finger proteins so far (**Wexler et al., 2014**). Phylogenetic trees of Sox and Fox gene families were rooted using two fungi mating protein A (Mat-A) sequences (XP_62685912.1, CCD57795.1) and two Amoebozoa forkhead-like domains (XP_004368148.1, XP_004333268.1), respectively (**Nakagawa et al., 2013; Heenan et al., 2016**). The rooting was performed with Gotree (v0.4.5; **Lemoine and Gascuel, 2021**). To identify and annotate bivalve homology groups within each gene family, we employed a species overlap algorithm followed by a Markov clustering algorithm (MCL) weighted by node

supports as implemented in Possvm (v1.2; **Grau-Bové and Sebé-Pedrós, 2021**). DSFGs from *H. sapiens*, *D. melanogaster*, and *C. elegans* were used as reference annotation.

In order to better establish the orthology relationships among ambiguous groups of Dmrt and Fox genes, we run a series of other phylogenetic reconstructions (see **Section 3.4**), by using the same pipeline as before. In the case of *Fox-Y* genes, we also employed Fox gene sequences from the sea urchin *Strongylocentrotus purpuratus*, as given by **Tu et al. (2006)**. All the phylogenetic trees were plotted using the R package ‘ggtree’ (**Yu et al., 2017**).

3.2.3 Sequence diversity of bivalve single-copy orthogroups

As a metrics to measure the sequence diversity of bivalve DSFGs, and test whether those putatively involved in SD show higher values than other genes, we employed the amino acid sequence divergence. As a matter of fact, this metric is fast and straightforward to obtain, as it only requires the amino acid alignment and the corresponding best-fit substitution mode.

To this purpose, we produced amino acid alignments of bivalve single-copy orthogroups (SCOs) groups and built the distribution of their median amino acid sequence divergence (AASD). Specifically, we assembled a second dataset (hereafter referred to as the ‘reduced bivalve dataset’) which includes, for each bivalve genus, only the best genomes and transcriptomes in terms of either BUSCO scores (on the ‘metazoan_odb10’ dataset; **Manni et al., 2021**) or assembly statistics (**Tab. S1**), in order to reduce computational time. *Archivesica marissinica* (now *Calyptogena marissinica*) and *Saccostrea glomerata* were also removed, as their annotated coding sequences contain many stop codons, which prevent accurate amino acid guided alignments. Genes were clustered in orthologous groups using OrthoFinder (v2.5.5; **Emms and Kelly, 2019**) with DIAMOND ultra-sensitive and default parameters. Resulting orthogroups were splitted into SCOs using DISCO (v1.3.1; **Willson et al., 2022**), and orthogroups with at least 17 species (50% of the species included in the bivalve reduced dataset) were retained. Amino acid and nucleotide sequences of SCOs were then aligned using Clustal Omega as implemented in TranslatorX (v1.1; **Abascal et al., 2010**), and jointly trimmed using trimAl with a gap threshold of 40% and the removal of spurious sequences (`-resoverlap 50 -seqoverlap 50`). Eventually, orthogroups containing (i) internal stop codons, (ii) with less than 17 species left (50% of the species included in the bivalve reduced dataset), or (iii) containing DSFGs were removed from downstream analyses. The best

amino acid substitution model was inferred for each trimmed alignment using ModelFinder as implemented in IQTREE2 (model search was restricted to matrices accepted by the ‘phangorn’ R library; i.e., Blosum62, cpREV, Dayhoff, DCMut, FLU, HIVb, HIVw, JTT, JTTDCMut, LG, mtART, mtMAM, mtREV, mtZOA, rtREV, VT, WAG) and the corresponding pairwise amino acid distances were computed with the function ‘dist.ml’ from the ‘phangorn’ R package (**Schliep, 2011**). We decided to employ the pairwise amino acid distance instead of the tip-to-tip phylogenetic distance (which accounts for a more comprehensive evolutionary signal) in order to save computational time. However, to check whether the two metrics were comparable to each other, we randomly selected 200 decomposed orthogroups (including orthogroups from the DSFGs) and computed the maximum likelihood (ML) trees using IQTREE2, with ModelSelection restricted as before. Then, the tip-to-tip pairwise distances were obtained with the R package ‘adephylo’ (**Jombart and Dray, 2010**). The same pipeline was also employed to obtain pairwise amino acid distances for each DSFG single-copy orthologous group.

The distribution of amino acid distances was then built after the median values of pairwise distances of each SCO, and genes were categorised accordingly into three groups: Group 1, consisting of genes from the 1% upper quantile of the distribution; Group 2, consisting of genes between the 1% and 5% upper quantiles; and Group 3, consisting of all the remaining genes. Group 1 and Group 2 genes will be referred to as ‘highly divergent genes’.

3.2.4 Mammals and *Drosophila* spp. as test datasets

To validate our approach for the study of bivalve SRG molecular evolution, we run the same analysis on two additional datasets, consisting of reference genomes of mammals and *Drosophila* species (**Tab. S4** and **S5**, respectively), whose sex-determining mechanisms are well studied and characterised. As a matter of fact, despite it is well known that SDGs tend to evolve faster than genes not involved in SD, the hypothesis has never been tested extensively across the entire phylogenetic diversity of a group: molecular evolution of SDGs and SRGs has mainly been tested on single species or inside the boundaries of taxonomic genera (REFERENCE). For both mammals and fruit flies, annotated genomes were downloaded from NCBI using the command-line tool ‘datasets’, then processed using the same pipeline and scripts as before (**Fig. 3.1**).

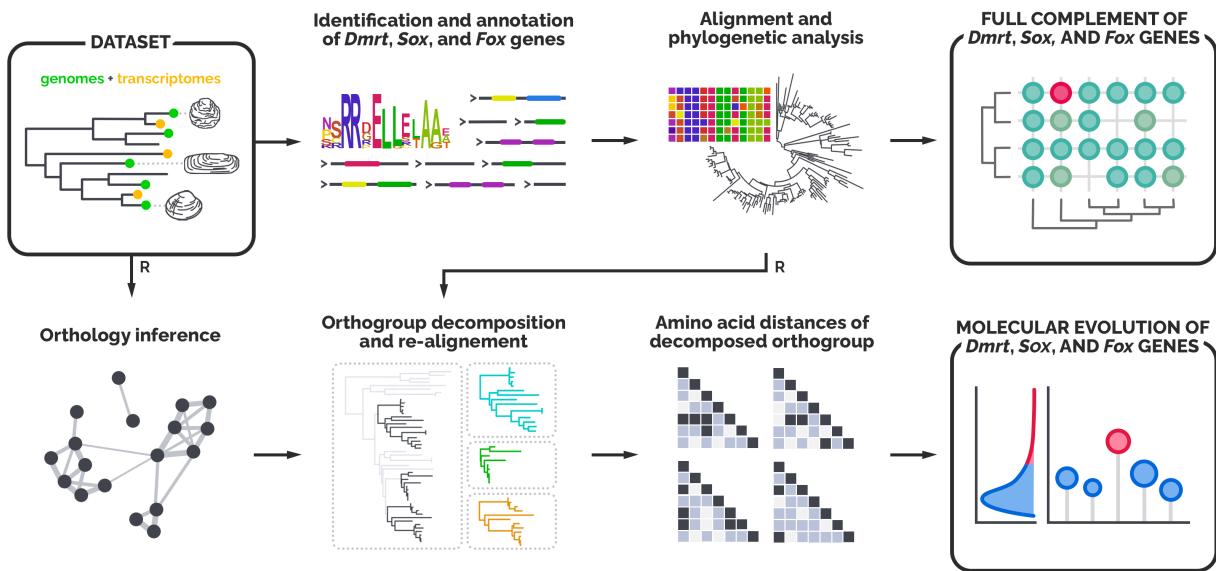


Figure 3.1 – Workflow of the analyses for the bivalve dataset. Starting from a set of both genomes and transcriptomes covering a great portion of bivalve taxonomic diversity, we first characterized the entire complement of glsdsfg genes (upper row). In particular, we used sequence annotation and phylogenetic tools to obtain reliable sequences and filter out any putative mis-assembled or mis-annotated sequence. Afterwards, we built a reduced set of transcriptomes and genomes (the reduced bivalve dataset, where we minimized the redundancy of congeneric species) from which to draw the molecular evolution patterns of orthologous genes (bottom row). In particular, after having obtained gene single-copy orthologous groups, we calculated the amino acid distances within each orthogroup and then we built the distribution of median values. The same pipeline was also employed for the mammal and the fruit fly datasets, with just two minor differences: the starting dataset was composed of only genomes, and that the reduction step (R) was not necessary.

3.2.5 GO-term enrichment

After having obtained the distributions of AASD in the three datasets (Bivalvia, Mammalia, and *Drosophila*) and having sorted SCOs genes up into 3 groups (Group 1, Group 2, and Group 3), we performed a gene ontology (GO) enrichment analysis of genes from Group 1 and genes from Group 1 + Group 2. To do so, we firstly selected one gene per SCO, giving priority to few chosen species: (i) for bivalves, we selected genes from *Pecten maximus*, or alternatively from *C. gigas*, *Hyriopsis bialata* (now *Unio delphinus*), *Tridacna squamosa*, and *Solen grandis*; (ii) for mammals, we selected genes from *H. sapiens*, or alternatively from *Bubalus bubalis*, *Panthera tigris*, *Camelus dromedarius*, and *Monodelphis domestica*; (iii) for fruit flies, we selected genes from *D. melanogaster*, or alternatively from *Drosophila hydei*, *Drosophila pseudoobscura*, and

Drosophila suzukii. By doing so, we ensured that each SCO was represented by one gene. Afterwards, we annotated the obtained datasets with the corresponding GO terms using the OMA browser (accessed 18/09/2024; **Altenhoff et al., 2024**). The GO-term enrichment of Group 1 genes and Group 1 + Group 2 genes was performed with the R package ‘topGO’ with the Fisher exact test (**Alexa and Rahnenführer, 2009**).

3.3 Results

3.3.1 Genomic and transcriptomic datasets

The complete bivalve dataset consists of 29 bivalve genomes, 14 bivalve transcriptomes, and 7 outgroup genomes (5 gastropods and 2 *Octopus* spp.; **Tab. S1**). BUSCO statistics for complete single-copy genes spanned from the 64.9% in *Modiolus modiolus* to the 99.4% of *Perna viridis*, with a median value of 94.7%. We were able to get at least one representative species for 11 different bivalve orders, covering a good proportion of the phylogenetic diversity of the clades Pteriomorpha, Palaeoheterodonta, and Imparidentia, and thus building the most extensive genomic and transcriptomic dataset for bivalve comparative analyses so far (**Tab. S1**). Unfortunately, no genomes or transcriptomes for Protobranchia, Archiheterodonta, and Anomalodesmata were available at the time of the project, thus we were not able to include any of those clades in our analysis. The reduced bivalve dataset (used for the orthology inference and the molecular evolution analysis; **Fig. 3.1**) consists instead of 36 genomes and transcriptomes (**Tab. S1**), and was built to retain just one species for each taxonomic genera.

The mammal dataset consists of 32 species and 1 outgroup (*Gallus gallus*, Aves; **Tab. S4**), and covers 12 major orders, while the fruit fly dataset consists of 17 species and 1 outgroup (*Anopheles gambiae*, Culicidae; **Tab. S5**), and covers 2 *Drosophila* subgenera (i.e., *Drosophila* and *Sophophora*). BUSCO statistics for complete single-copy genes were generally higher than those of bivalves, with a median of 98.3% for mammals and of 99.8% for fruit flies (**Tab. S4** and **S5**).

3.3.2 The Dmrt, Sox, and Fox complements in bivalves

Our annotation pipeline managed to successfully identify and annotate DSFGs in bivalves, as proved by the same analysis in mammals and fruit flies (see **Section 3.3.4**).

We retrieved four main orthology groups of Dmrt genes in bivalves (**Fig. 3.2** and **S1**; **Tab. S6**), three corresponding to the groups present in the Bilateria common ancestor (*Dmrt-2*, *Dmrt-3*, and *Dmrt-4/5*; **Mawaribuchi et al., 2019**), and one additional group with no unambiguous ortholog among reference genes, and thus putatively specific to molluscs (named *Dmrt 1-like* [*Dmrt-1L*], as per **Li et al., 2018**; **Evensen et al., 2022**). The majority of identified Dmrt genes are present in single-copy in each species, but *Dmrt-4/5*s show a group-specific expansion in Palaeoheterodonta and Heterodonta, while *Dmrt-1L* is completely absent from Heterodonta. The degree of missing data for Dmrt genes in bivalves is about 35%, with *Dmrt-2* having the highest (about 56%) and *Dmrt-4/5* the lowest (about 7%; **Tab. S7**). The coupling of ubiquitin conjugation to endoplasmic reticulum degradation (CUE)-like DM-associated (DMA) domain has been annotated in most of the *Dmrt-3* and *Dmrt-4/5* genes, while an additional DM domain has been annotated in *Dmrt-1L* genes in Mytilida and the gastropod *Pomacea canaliculata* (**Tab. S6**). Additionally, we retrieved six main orthology groups of Sox genes, none of which is restricted to molluscs or bivalves (**Fig. 3.2** and **S2**; **Tab. S6**). Five Sox groups (*Sox-B1/2*, *Sox-C*, *Sox-D*, *Sox-E*, and *Sox-F*) are those traditionally considered to be present in the Bilateria common ancestor (**Phochanukul and Russell, 2010**), while one has been identified outside mammals only recently (*Sox-H*, or *Sox-30*; **Han et al., 2010**). *Sox-B2* and *Sox-B1* have been grouped in the same clade, as in our phylogenetic reconstruction the former results in a paraphyletic group with the latter (**Fig. S2**), despite being traditionally recognised as a separate paralogy group in humans, fruit flies, and nematodes. The degree of missing data for Sox genes in bivalves is about 8%, with *Sox-H* having the highest (about 21%) and *Sox-B1/2* and *Sox-C* both having no missing genes (**Tab. S7**). The Sox N-terminal signature domain was annotated for *Sox-E* genes (**Tab. S6**). Concerning Fox genes, we retrieved 27 main orthology groups (**Fig. 3.2** and **S3**; **Tab. S6**), two of which are specific to molluscs (*Fox-OG13/NA*, *Fox-OG16/NA*). Additionally, other potential mollusc-specific Fox groups have been identified, but these have been excluded from the final orthology analysis as they are present in less than half of bivalve species (see **Section 3.2**; **Tab. S6**). The two major Fox gene subgroups, Group I (monophyletic, specific to Metazoa; includes *Fox-A*, *Fox-B*, *Fox-C*, *Fox-D*, *Fox-E*, *Fox-F*, *Fox-G*, *Fox-H*, *Fox-L1*, *Fox-L2*, *Fox-Q2*) and Group II (paraphyletic, specific to Opisthokonta; includes *Fox-O*, *Fox-P*, *Fox-J2*, *Fox-J1*, *Fox-K*, *Fox-N2/3*, *Fox-N1/4*; **Larroux et al., 2008**), have been recovered, including the four Fox genes that were present in the Bilateria common ancestor (*Fox-C*, *Fox-F*, *Fox-L1*, and *Fox-Q1*; **Shimeld et al., 2010**).

Two putative lineage-specific expansions have been recovered for *Fox-OG28/NA*, one regarding *Mytilus* spp. and one regarding the two Myida species (**Fig. 3.2**; **Fig. S3**). The degree of missing data for Fox genes in bivalves is about 22%, with *Fox-H* having the highest (about 42%) and *Fox-J1* having no missing genes (**Tab. S7**). The forkhead-associated (FHA) domain was annotated for *Fox-K* genes, the *Fox-P* coiled-coil signature domain was annotated for *Fox-P* genes, while both the forkhead N- and C-terminal signature domains were annotated for *Fox-A* genes (**Tab. S6**). Regarding bivalve species, the amount of missing data greatly differs between genomes and transcriptomes, with a mean of about 9% and about 45%, respectively. *Argopecten irradians concentricus*, *Mytilus unguiculatus* (formerly *coruscus*), and *Pecten maximus* have no missing data, while *Loripes orbiculatus* has the highest proportion (about 64%; **Tab. S7**).

3.3.3 Amino acid sequence divergence of Dmrt, Sox, and Fox genes in bivalves

In the reduced bivalve dataset, OrthoFinder collectively analysed >1.2G genes distributed in 34 species. 89.4% of these genes were placed in orthogroups, while 10.6% were not. The number of retrieved SCOs is 5, which is drastically low but can be explained considering the mixed nature of the dataset, that is, it includes both genomes and transcriptomes with highly different BUSCO scores (**Tab. S1**). In order to be able to analyse a greater number of genes, we decomposed OrthoFinder orthogroups using DISCO and eventually obtained 11k SCOs with at least 50% of the species. By running the same pipeline on DSFGs, we included in the AASD analysis 32 SCOs (**Fig. 3.2**) out of 33 initial Possvm-identified groups (*Fox-H* didn't meet the species occupancy threshold; **Fig. 3.3**).

From the distribution of median AASD, 112 genes were assigned to Group 1 (1% upper quantile), 447 to Group 2 (5% upper quantile), and 10.603 to Group 3. Most of the DSFGs (29/32) fell in Group 3 (**Fig. 3.3**), which means they have a median AASD comparable to the vast majority of other genes in bivalves (median level of the genomes). Just *Dmrt-1L*, *Sox-H*, and *Sox-F* showed higher divergences, and have been accordingly placed in Group 2. Overall, pairwise AASD proved to be a good approximation of the tip-to-tip distances ($R = 0.84, p < 2.2 \times 10^{-16}$, calculated on 200 randomly-selected trees; **Fig. 3.3C**), while it showed no influence from the alignment length ($R = 0.11$) or the number of represented species ($R = -0.23$; **Fig. 3.3D** and **3.3E**). Genes from Group 1 and Group 2 are strongly involved in

cellular regulatory processes (such as those related to the metabolism of nucleic acids, proteins, and other macromolecules), but also in development and response to external stimuli, as shown by the GO-term enrichment analysis (**Tab. 3.1** and **S10**).

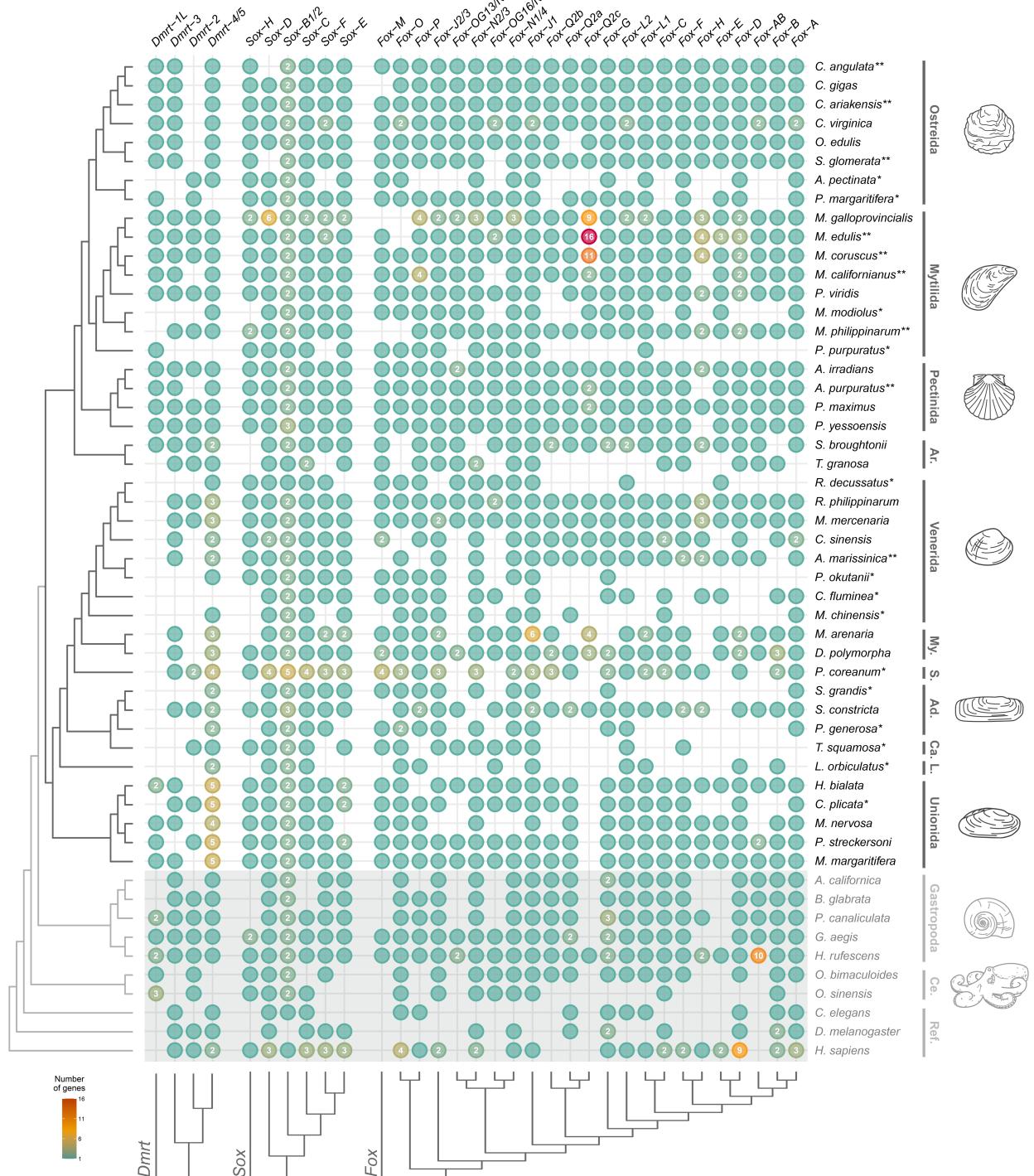


Figure 3.2 – DSFG complement in bivalves and their outgroups. Presence/absence of genes in various species are indicated by filled circles. Numbers inside each circle specify genes with 2 or more copies. The shaded area highlights non-bivalve species, belonging either to other molluscs or to the references. The phylogenetic tree of analyzed species, as inferred from literature, is shown on the left, while major taxonomic groups are reported on the right. Species represented by transcriptomic data are marked with an asterisk ('*'), and species not present in the reduced bivalve dataset are marked with two asterisks ('**'; see main text and **Fig. 3.1**); note that the two categories do not overlap. DSFG trees are shown on the bottom (full trees can be found in **Fig. S1** and **S3**). Full species names, along with all assembly and taxonomic information, can be found in **Tab. S1**. Ad.: Adapedonta; Ar.: Arcida; Ca.: Cardiida; Ce.: Cephalopoda; L.: Lucinida; My.: Myida; Ref.: reference genes; S.: Sphaeriida.

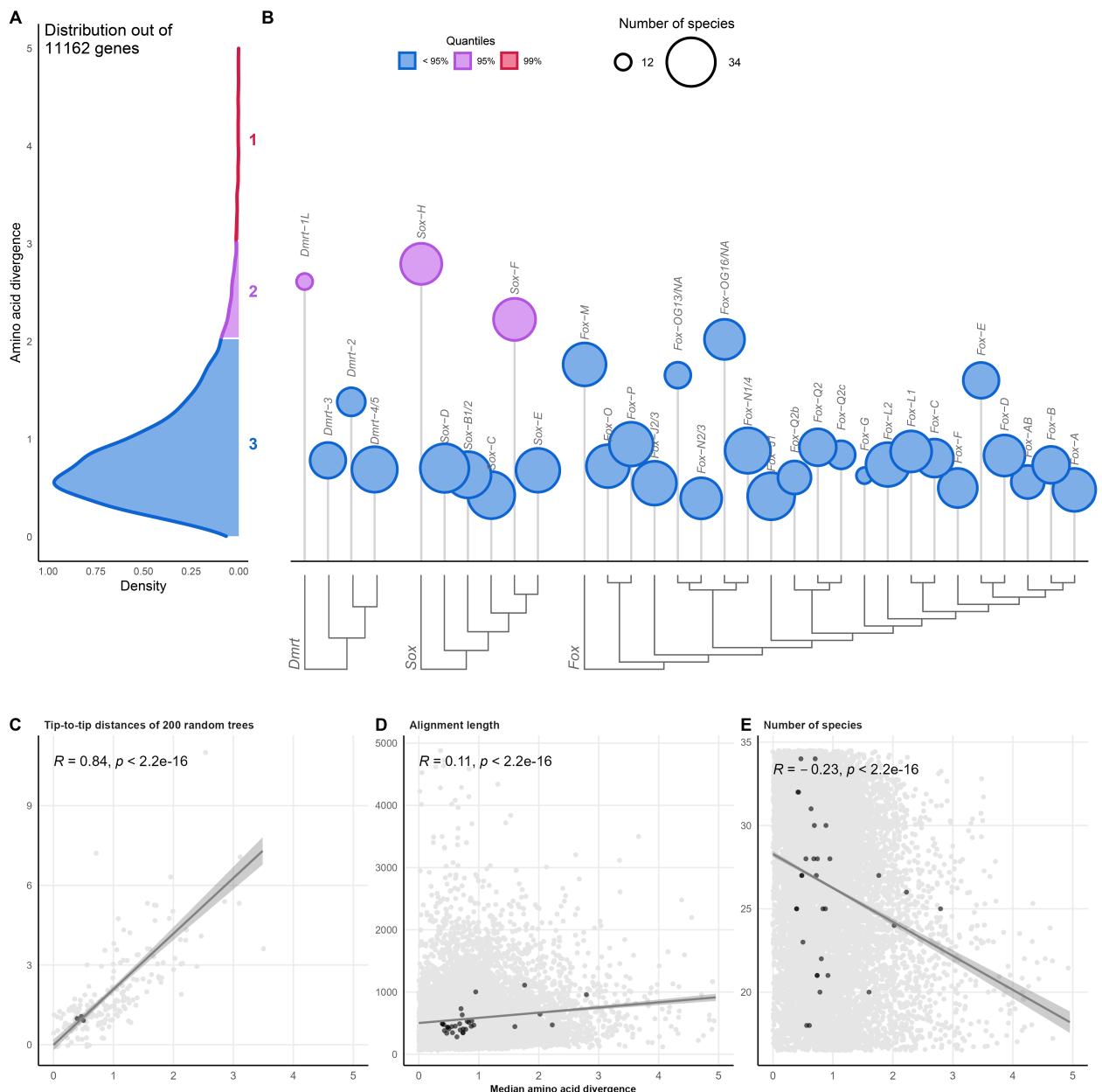


Figure 3.3 – Distribution of AASD of single-copy orthogroups in bivalves (A), including DSFGs (B), and their correlations with tip-to-tip distances (C), alignment lengths (D), and number of species (E). The distribution of AASD has been computed on the median values of pairwise distances of >11k SCOs from the reduced bivalve dataset (see main text and **Fig. 3.1**). Genes have been divided according to their median AASD value into three different groups, which are indicated by different colors and increasing numbers (Groups 1, 2, and 3). Circle heights of DSFGs show the median value of their AASD, while the size indicates the number of represented species. DSFG trees are shown on the bottom (full trees can be found in **Fig. S1** and **S3**). Darker points in C–E indicate DSFG SCOs. The correlation between the amino acid distance and the tip-to-tip distance has been computed on 200 randomly-selected orthogroups.

Table 3.1 – Top enriched GO terms for Group 1 and Group 2 genes of bivalves, mammals, and *Drosophila*. The extended version of the table, which includes also the expected number of annotated genes per GO term and all the other enriched GO terms, can be accessed in [Tab. S10](#).

Dataset	GO.ID	Term	Annotated genes	Significant genes	Corrected genes	p-value
Bivalvia	GO:0060255	regulation of macromolecule metabolic process	737	59	0.04525	
	GO:0080090	regulation of primary metabolic process	673	53	0.01818	
	GO:0019219	regulation of nucleobase-containing compound metabolic process	541	41	0.02388	
	GO:0006351	DNA-templated transcription	571	39	0.03767	
	GO:0032774	RNA biosynthetic process	579	39	0.04490	
	GO:0051252	regulation of RNA metabolic process	517	37	0.02719	
	GO:0006355	regulation of DNA-templated transcription	490	35	0.03751	
	GO:2001141	regulation of RNA biosynthetic process	491	35	0.03844	
	GO:0006950	response to stress	370	33	0.01949	
	GO:0032502	developmental process	261	27	0.04445	
	GO:0006468	protein phosphorylation	345	23	0.02483	
	GO:0031325	positive regulation of cellular metabolic process	125	17	0.00801	
	GO:0010604	positive regulation of macromolecule metabolic process	151	17	0.04047	
	GO:0051172	negative regulation of nitrogen compound metabolic process	117	16	0.00814	
	GO:0051173	positive regulation of nitrogen compound metabolic process	137	15	0.02454	
	GO:0006310	DNA recombination	66	14	0.00087	
	GO:0048513	animal organ development	83	12	0.04088	
	GO:0010629	negative regulation of gene expression	78	11	0.00048	
	GO:0023051	regulation of signaling	133	11	0.02872	
	GO:0045934	negative regulation of nucleobase-containing compound metabolic process	64	11	0.03637	
	GO:0009605	response to external stimulus	90	11	0.04544	

Tab. 3.1 continued from previous page

Dataset	GO.ID	Term	Annotated genes	Significant genes	Corrected p-value
Bivalvia	GO:0044419	biological process involved in interspecies interaction between organisms	63	11	0.04761
	GO:0006955	immune response	1297	145	0.00061
	GO:0098542	defense response to other organism	853	112	0.02066
	GO:0045087	innate immune response	647	82	8.5e-10
	GO:0001817	regulation of cytokine production	630	51	0.04660
	GO:0042742	defense response to bacterium	233	45	1.7e-07
	GO:0006954	inflammatory response	642	45	0.01735
	GO:0019221	cytokine-mediated signaling pathway	382	44	3.9e-07
	GO:0002250	adaptive immune response	342	44	1.3e-05
	GO:0001819	positive regulation of cytokine production	402	41	0.02723
	GO:0002697	regulation of immune effector process	308	37	0.04426
	GO:0042110	T cell activation	432	35	0.02564
	GO:0051607	defense response to virus	257	34	1.9e-07
	GO:0048232	male gamete generation	491	32	0.02255
	GO:0007283	spermatogenesis	478	31	0.02801
	GO:0070661	leukocyte proliferation	273	29	0.01285
	GO:0002449	lymphocyte mediated immunity	221	29	0.04833
	GO:0070663	regulation of leukocyte proliferation	212	25	0.01870
	GO:0050727	regulation of inflammatory response	300	24	0.00235
	GO:0031349	positive regulation of defense response	240	24	0.01239
	GO:0002768	immune response-regulating cell surface receptor signaling pathway	177	22	0.00336
	GO:0050829	defense response to Gram-negative bacterium	66	17	1.7e-10
	GO:0071222	cellular response to lipopolysaccharide	164	17	0.00012

Tab. 3.1 continued from previous page

Dataset	GO.ID	Term	Annotated genes		Significant genes	Corrected p-value
			Annotated genes	Significant genes		
Mammalia	GO:0010466	negative regulation of peptidase activity	163	16	0.00036	
	GO:0002429	immune response-activating cell surface receptor signalling pathway	164	16	0.00243	
	GO:1903555	regulation of tumor necrosis factor superfamily cytokine production	137	16	0.01244	
	GO:0071706	tumor necrosis factor superfamily cytokine production	137	16	0.01244	
	GO:0070665	positive regulation of leukocyte proliferation	132	16	0.02765	
	GO:0045089	positive regulation of innate immune response	113	16	0.03224	
	GO:0071356	cellular response to tumor necrosis factor	175	15	0.00219	
	GO:0002695	negative regulation of leukocyte activation	148	15	0.01151	
	GO:0002456	T cell mediated immunity	82	15	0.01605	
	GO:0002705	positive regulation of leukocyte mediated immunity	113	15	0.01837	
Drosophila	GO:0032680	regulation of tumor necrosis factor production	133	15	0.03262	
	GO:0032640	tumor necrosis factor production	133	15	0.03262	
	GO:0050866	negative regulation of cell activation	165	15	0.04048	
	GO:0000819	sister chromatid segregation	140	11	0.02927	
	GO:0070192	chromosome organization involved in meiotic cell cycle	54	9	0.00849	
	GO:0007131	reciprocal meiotic recombination	37	7	0.00066	
	GO:0007143	female meiotic nuclear division	54	6	0.02270	
	GO:0035967	cellular response to topologically incorrect protein	44	5	0.03334	
	GO:0035966	response to topologically incorrect protein	47	5	0.04266	
	GO:0007141	male meiosis I	13	4	0.00150	
	GO:0140543	positive regulation of piRNA transcription	3	3	6.9e-05	
	GO:0010526	retrotransposon silencing	8	3	0.00331	
	GO:0007130	synaptonemal complex assembly	10	3	0.00666	

Tab. 3.1 continued from previous page

Dataset	GO.ID	Term	Annotated genes		Significant genes	Corrected p-value
			Annotated genes	Significant genes		
Drosophila	GO:0030719	P granule organization	11	3	0.00888	
	GO:0071218	cellular response to misfolded protein	12	3	0.01149	
	GO:0051788	response to misfolded protein	12	3	0.01149	
	GO:0007135	meiosis II	15	3	0.02169	
	GO:0034508	centromere complex assembly	19	3	0.04094	

3.3.4 Dmrt, Sox, and Fox genes, and amino acid sequence divergence in the test datasets

The DSFG datasets retrieved in mammals and fruit flies are far more complete than those in bivalves, and most of the already-recognised orthology groups have been identified.

In mammals, we retrieved 7 Dmrt orthology groups with about 3.1% of missing data, 20 Sox orthology groups with about 8.1% of missing data, and 42 Fox orthology groups with about 4.6% of missing data (**Fig. S4A, S5, and S7; Tab. S8**). Of these, just *Sox-5* was not included in the subsequent AASD analysis, as it did not meet the 50%-species occupancy threshold. OrthoFinder analysed about 650M genes, and the number of SCOs used in the AASD analysis (thus resulting from the DISCO-based orthogroup decomposition pipeline) is >16k (**Fig. 3.4A**). From the distribution of median AASD, 163 genes were assigned to Group 1, 649 to Group 2, and 15.355 to Group 3. Most of the DSFGs (66/68) fell in Group 3 (**Fig. 3.4B**), while *Sry* and *Fox-D4* showed higher divergences, and have been accordingly placed in Group 1 and 2, respectively. Genes from Group 1 and Group 2 show a strong enrichment in immune-related functions (such as innate and adaptive immune response, defence response to bacteria and viruses, lymphocyte metabolism, etc.), but also in reproductive processes (such as spermatogenesis; **Tab. 3.1** and **S10**).

Concerning *Drosophila*, we retrieved 4 Dmrt orthology groups with about 1.7% of missing data, 7 Sox orthology groups with about 3.9% of missing data, and 17 Fox genes with about 8.3% of missing data (**Fig. S4B, S8, and S10; Tab. S9**). OrthoFinder analysed about 240M, and the distribution of median AASD was built after >12k SCOS (**Fig. 3.4C**). 126 genes were assigned to Group 1, 501 to Group 2, and 11.880 to Group 3. All of the DSFGs have been used in the AASD analysis, but none of them have been placed in Group 1 or 2, that is, all the DSFGs in *Drosophila* have an AASD comparable to the median level of the genome (**Fig. 3.4D**). Genes of Group 1 and Group 2 show a GO-term enrichment in meiotic processes, such as chromosome/chromatid organisation, and retrotransposon silencing (**Tab. 3.1** and **S10**).

3.4 Discussion

3.4.1 A new manually-curated and phylogenetic-based reference dataset of Dmrt, Sox, and Fox genes in bivalves

The annotation and characterisation process of a gene family in a certain clade of organisms may harbour many overlooked challenges (**Vizueta Moraga et al., 2020**). For example, the presence of highly-conserved catalytic domains may hamper the correct identification of the components of a gene family because of insufficient phylogenetic signal, as it is the case for Hox and ParaHox genes and their homeobox motif (**Baldwin-Brown et al., 2018; Nicolini, Martelossi, et al., 2023**). Conversely, the components of dynamic gene families characterised by abrupt and sequential duplication events may be difficult to sort into separate groups. As a matter of fact, varying levels of sequence heterogeneity and gene copy numbers makes the inference of orthologous groups hard, as for certain clans of the P450 gene family (**Dermauw et al., 2020**). Regardless of the causes, having a solid and wide phylogenetic context in which to study gene duplications and losses, and orthology relationships, is crucial to overcome these difficulties. In the same way, manual curation and visual inspection of multiple sequence alignments, phylogenetic trees, and gene structures (in terms of domain annotation, start and stop codons, and other feature representations) is helpful, despite being time-demanding and possibly low reproducible. In this study, we characterised the full complement of DSFGs in the vast class of bivalves, by leveraging sequence domain annotation, phylogenetics, and manual curation of the dataset. Our aim was to obtain the most reliable gene complements as possible, combined with a vast taxonomic dataset, a solid phylogenetic inference, an openly-available dataset of gene sequences, and a reproducible pipeline for the annotation of gene identity. By doing so, we want to provide a reliable resource for future studies of DSFGs, either focused on bivalves or generally in Metazoa.

Concerning the Dmrt gene family, we identified orthologs of the vertebrate *Dmrt-2*, *Dmrt-3*, and *Dmrt-4/5* (or *A1/A2*; **Fig. 3.2** and **S1**; **Tab. S6**), which are also expected to have been present in the Bilateria common ancestor (**Mawaribuchi et al., 2019**). **Wang et al. (2023)** found that *Dmrt-4/5* is duplicated in *Mercenaria mercenaria* and *Cyclina sinensis* (Venerida), and in *Dreissena polymorpha* (Myida), and we confirm this result by tracing back the duplication event to the split between Palaeoheterodonta (here represented by Unionida) and Heterodonta (here represented by Venerida, Myida, Sphaeriida, Adapedonta, Cardiida, and

Lucinida; **Fig. 3.2**). Furthermore, we confirm *Dmrt-1L* to be present in many bivalve species (mainly belonging to the Ostreida, Pectinida, Mytilida, and Unionida orders; **Fig. 3.2**), as well as in gastropods and *Octopus*. Though, our phylogenetic analysis did not retrieve any unambiguous orthology relationship among *Dmrt-1L* and either vertebrate *Dmrt-1* or *Drosophila dsx* genes, as instead it was proposed in previous works (Li et al., 2018; Evensen et al., 2022). As a matter of fact, the amino acid sequence of the *Dmrt-1L* DM domain does not recall that of any other Dmrt gene. Furthermore, it must be considered that various phylogenetic analyses have recovered both *Dmrt-1* and *dsx* genes to be restricted to vertebrates and arthropods, respectively (Wexler et al., 2014; Mawaribuchi et al., 2019; Panara et al., 2019), that is, they do not have any direct ortholog outside their relative clades. Thus, if *Dmrt-1L*, *dsx*, and *Dmrt-1* are true orthologs, their origin would need to be placed at least in the Bilateria common ancestor, which seems however to be not the case. All considered, we thus confirm that *Dmrt-1L* is not orthologous to *Dmrt-1* and *dsx* and is rather a mollusc-specific gene (Evensen et al., 2022). The monophyly of the group is not supported by the phylogenetic tree inferred with Dmrt genes from molluscs and the reference species (**Fig. S1**); though, it is recovered when analysing just genes from mollusc species (**Fig. S11**). To this regard, we speculate that in our analysis, the difficulty in obtaining the monophyly of *Dmrt-1L* genes may have arisen primarily because of the many *C. elegans*-restricted genes (**Tab. S3**), which are placed among the other bivalve genes (**Fig. S1**), but also because of the high AASD of *Dmrt-1L* genes (see the following section), which hampers a straight-forward phylogenetic reconstruction. Furthermore, our broad-context analysis allowed us to identify some cases of incorrect gene identification in bivalves, which have arisen because of erroneous or ambiguous annotations in previous works, as a result of limited datasets or analyses. For example, (i) the scallop-specific cluster of Dmrt genes retrieved by Wang et al. (2023) rather belongs to the *Dmrt-1L* group, and (ii) the classification of Dmrt genes in *Crassostrea* species provided by Zeng et al. (2024) needs to be revised following the one of this work: *Dmrt-1* genes are *Dmrt-4/5*; *Dmrt-2* genes are *Dmrt-3*; *Dmrt-3* genes are *Dmrt-1L*; hence, *Crassostrea* species do not have *Dmrt-2* genes.

For what concerns the Sox gene family, bivalves (or molluscs) do not show any major clade-restricted gene, as only the five Bilateria-specific Sox groups (*Sox-B1/2*, *Sox-C*, *Sox-D*, *Sox-E*, and *Sox-F*) and *Sox-H* have been identified (**Fig. 3.2** and **S2**; **Tab. S6**), in accordance with previous findings (Evensen et al., 2022; Wang and Nie, 2024; Yu et al., 2017). *Sox-*

B1/2 is clearly made up of two subgroups (i.e., *Sox-B1* and *Sox-B2*), as expected, but their respective identity could not be unambiguously established, as *Sox-B1/2* genes of reference species do not form separate clusters (**Fig. S2**). Even when inferring the phylogenetic tree only of components of the *Sox-B1/2* group from molluscs and reference species, the identity can not be properly established (**Fig. S12**).

Compared to Dmrt and Sox genes, the Fox gene family appears as the most dynamic in terms of gene presence/absence, as already shown by other works (**Wu et al., 2020; Schomburg et al., 2022; Seudre et al., 2022**). Our phylogenetic analysis successfully recovered Group I and Group II of Fox genes (**Larroux et al., 2008**), which include the four Fox genes that were present in the Bilateria common ancestor (*Fox-C*, *Fox-F*, *Fox-L1*, and *Fox-Q1*; **Fig. 3.2 and S3; Tab. S6**; Shimeld et al., 2010). To our knowledge, this is the first broad-taxonomic identification and classification of Fox genes in bivalves, as up to now they have been systematically characterised only in *C. gigas* (**Yang et al., 2014**), *Patinopacten yessoensis* (now *Mizuhopecten yessoensis*; **Wu et al., 2020**), and *Ruditapes philippinarum* (**Liu et al., 2024**). Firstly, our analysis confirms the absence in molluscs of *Fox-I*, *Fox-Q1*, *Fox-R*, *Fox-S* (**Fig. S3**), which are in fact thought to have emerged with the diversification of deuterostomes or vertebrates (**Yang et al., 2014; Wu et al., 2020; Schomburg et al., 2022; Seudre et al., 2022**). Furthermore, we have found many Fox groups that appeared as mollusc-specific and/or still-unnamed at a first analysis. However, a more in-depth investigation revealed a different scenario. *Fox-OG2/NA* appears close to the human *Fox-M* gene in the phylogenetic tree, but they do not form a monophyletic group (**Fig. S3**). However, by comparing *Fox-OG2/NA* sequences and phylogenetic tree with those analysed by **Yang et al. (2014)**, **Wu et al. (2020)**, **Schomburg et al. (2022)**, and **Seudre et al. (2022)**, it appears clear that this group of Fox genes is indeed *Fox-M*. However, our analysis has failed to retrieve a monophyletic relationship among bivalve and human *Fox-M* genes, even when inferring a tree with just *Fox-J2*, *Fox-M*, *Fox-O*, and *Fox-P* complements (**Fig. S13**), which belong to the same Fox group. Regarding the *Fox-OG39/NA* group, it does not have any homolog in reference species (**Fig. S3**) but is found to belong to the *Fox-AB* group by sequence comparison with previous works (**Yang et al., 2014; Wu et al., 2020; Seudre et al., 2022**). *Fox-AB* was formerly described only in the sea urchin *S. purpuratus* and the lancelet *Branchiostoma floridae* (**Tu et al., 2006; Yu et al., 2008**), but was later identified also in several Spiralia lineages, including molluscs (e.g., **Yang et al., 2014; Wu et al., 2020; Seudre et al., 2022**). A

similar situation concerns *Fox-OG15/NA* and *Fox-OG28/NA*, which again could not be named based on orthology relationships with the reference species genes (**Fig. S3**), but actually represent two lineage-specific expansions of the *Fox-Q2* group (named *Fox-Q2b* and *Fox-Q2c*), as already appointed in previous studies (Yang et al., 2014; Wu et al., 2020). This observation fits within the wider context of the *Fox-Q2* group expansion in Bilateria and, particularly, in Spiralia, that led to remarkable differences in their gene copy numbers across various clades (Seudre et al., 2022). Two additional Fox genes have been previously identified in bivalves, and were named *Sox-Y* and *Sox-Z* (Yang et al., 2014; Wu et al., 2020). In our analysis, these Fox groups were identified as *Fox-OG13/NA* and *Fox-OG16/NA*, after sequence comparison of Fox genes from *C. gigas* and *P. yessoensis*. On one hand, *Fox-Y* was firstly identified in *S. purpuratus* (Tu et al., 2006) and only recently in a few bivalve species (Yang et al., 2014; Wu et al., 2020). However, when analysing bivalve and *S. purpuratus* Fox genes, we failed in retrieving such a clear orthology relationship, as *S. purpuratus* *Fox-Y* does not fall within the phylogenetic range of bivalve *Fox-OG13/NA*, which contains the supposed *Fox-Y* orthologs (**Fig. S14**). Also, the forkhead domains of *Fox-OG13/NA* genes were annotated as ‘forkhead domain P’ (**Tab. S6**). On the other hand, *Fox-Z* was firstly identified in bivalves and in several other protostomes, thanks to a phylogenetic work including the brachiopod *Lingula unguis*, the annelid *Capitella teleta*, the scorpion *Centruroides sculpturatus*, and the centipede *Strigamia maritima* (Wu et al., 2020). However, later works have not recovered this Fox gene, even when analysing annelids (Seudre et al., 2022) and panarthropods (Schomburg et al., 2022) in a more focused effort. In this case, the forkhead domains were annotated as either a generic ‘forkhead domain’ or a ‘forkhead domain Q2’ (**Tab. S6**). All considered, we argue that bivalves possess two additional Fox groups (here *Fox-OG13/NA* and *Fox-OG16/NA*; **Fig. 3.2** and **S3**; **Tab. S6**) which are shared with other mollusc species, as revealed also by other authors. However, given the discordant results of the phylogenetic hypothesis and domain annotation, we think that a more thorough investigation on their orthology relationships with Fox genes from other Metazoa is needed, and thus we chose to not employ their former names *Fox-Y* and *Fox-Z*.

Besides the DSFG groups discussed so far, it must be also considered that many orphan genes have been identified (**Fig. S1** and **S3**; **Tab. S6**). For example, Wu et al. (2020) identified a duplication event of *Fox-H* genes in *C. gigas*, which has been recovered also in our analysis for the entire Ostreida clade (*Fox-OG36/NA*; **Fig. S3**). Similarly, a gene orthology

group putatively specific to Pteriomorphia has been identified among Sox genes (*Sox-OG1/NA*). Of course, these genes deserve as much attention as their widely-distributed paralogs, as they may constitute true group-specific expansions and may play fundamental roles in some biological processes. However, they have not been discussed here or included in **Fig. 3.2** for clarity purposes, but they are freely available in supplementary materials.

Overall, our analysis clearly shows the importance of adopting a wide-angle approach when characterising the members of a gene family, especially for large ones such as the Fox genes (**Schomburg et al., 2022**). As a matter of fact, the presence of duplication events and orphan genes needs to be addressed with a broad taxonomic dataset, in order to account for possible mis-annotations, gene phylogenetic mis-placements, and sequence heterogeneity. Additionally, many reference species need to be included for the gene identification process, in order to consider distantly-related genes and obtain a solid annotation. Our gene annotation pipeline also resulted to be very solid, even with non-model organisms and sub-optimal genomic and transcriptomic resources as they are those of bivalves. As a matter of fact, by running the same pipeline on two additional datasets composed of mammal and fruit fly genomes, we were able to obtain high-quality orthology groups in accordance with previous knowledge on the clades (**Fig. S5 and S10; Tab. S8 and S9**), with little or no manual curation. Furthermore, this represents also the first broad analysis of DSFGs in both mammals and fruit flies, as so far attention has been mainly dedicated to single well-studied organisms or little clades (e.g., **Jackson et al., 2010**).

3.4.2 High amino acid sequence divergence identifies putative sex-determining genes

Sex-biased genes tend to evolve more rapidly than unbiased genes at the level of their protein sequences. Accelerated rates have been observed in both male-biased genes (reviewed in **Parsch and Ellegren, 2013; Grath and Parsch, 2016**) and female-biased genes (e.g., **Papa et al., 2017; Ghiselli et al., 2018**), but also in SRGs and primary SDGs (**O’Neil and Belote, 1992; Whitfield et al., 1993; de Bono and Hodgkin, 1996**). For example, it has been shown that *Dm-W*, *Dmy*, and *Sry* (which are SDGs in the African clawed frog *X. laevis*, in the medaka fish *O. latipes*, and in eutherians, respectively) all have higher substitution rates than their paralogues (*Dmrt-1* for *Dm-W* and *Dmy*, *Sox-3* for *Sry*), particularly when

considering their DNA-binding domains (**Mawaribuchi et al., 2012**). Similarly, both a burst of positive selection and a relaxation of purifying selection has been detected in *Drosophila Sxl* in correspondence with its recruitment at the top of the sex-determining cascade. The same signs of relaxed purifying selection have been found in the downstream targets of *Sxl*, that is, *tra* and *dsx*, despite no evidence of positive selection has been detected (**Mullon et al., 2012**).

Considering these shared features of SRGs and SDGs, we decided to look for signs of accelerated sequence evolution in DSFGs of bivalves, in order to evaluate if any of them could be *a-priori* associated with SD by employing the tools of molecular evolution. However, we wanted to analyse patterns of sequence evolution not only among putative SRGs and their close paralogs, but also considering the genomic context in which these genes evolve. In fact, our aim was to check whether higher rates of sequence evolution of SRGs hold true also when compared to other genes not involved in SD and not belonging to the same gene family. To do so, we obtained the AASD median values of more than 11k SCOs from bivalve genomes (**Fig. 3.3A**), in order to build a statistical distribution to be used as a reference: if SRGs/SDGs (in this case, DSFGs) truly evolve faster than other genes, we may expect them to fall within the 5% (or even 1%) upper quantile of the distribution (**Fig. 3.3B**), i.e., within highly divergent genes (Group 1 and Group 2 genes of the distribution; see **Section 3.2**). We chose to use the AASD as a metric of sequence evolution (instead of the tip-to-tip distances of phylogenetic trees, which account for more comprehensive evolutionary models) in order to save computational time. As a matter of fact, the AASD median values proved to be a good approximation of the tip-to-tip median distances in 200 randomly-selected genes (**Fig. 3.3C**; $R = 0.84, p < 2.2 \times 10^{-6}$).

Among DSFGs, three fell within the 5% upper quantile, namely *Dmrt-1L*, *Sox-H*, and *Sox-F*. Interestingly, *Dmrt-1L* and *Sox-H* have been already proposed to be involved in the male SD pathway of *C. gigas* (*inset* in **Fig. 3.3B**; **Zhang et al., 2014**), on the basis of DGE analyses. Specifically, *Sox-H* would play a major role in *C. gigas* SD, by interacting with *Dmrt-1L* and determining the onset of the male phenotype development; at the same time, both *Sox-H* and *Dmrt-1L* would inhibit *Fox-L2*, which instead is necessary to start the female phenotype development. *Dmrt-1L* and *Sox-H* have been appointed several other times to be involved in male-gonad development and differentiation, through DGE (e.g., **Teaniniuraitemoana et al., 2014**; **Capt et al., 2018**; **Afonso et al., 2019**), mRNA-ISH (e.g., **Naimi et al., 2009**; **Li et al., 2018**; **Liang et al., 2019**; **Yue et al., 2021**) and RNA interference (RNAi) (**Liang et al., 2019**; **Sun et al., 2022**). Therefore, the high AASD of *Dmrt-1L* and *Sox-H* is coherent

with previous works, strengthening their role as putative SRGs.

The relationship between high gene AASD and the involvement in SD is particularly enforced when looking at the patterns of AASD in the test datasets, which corroborates the solidity of our analysis: (i) from one side, in the mammal dataset—which represents a strictly genetic SD system, thus with a master and rapidly-evolving SDG, one of the genes from the 5% upper quantile of the distribution is *Sry* (**Fig. 3.4A** and **3.4B**), the male sex-determining gene in eutherians (*inset* in **Fig. 3.4B**); (ii) from the other side, in the fruit fly dataset—which represents a chromosomal SD system, thus without any expected difference in the rates of sequence evolution among SRGs, none of the DSFG exhibit significantly high AASD (**Fig. 3.4C** and **3.4D**), including the downstream effector *dsx* (*inset* in **Fig. 3.4D**). Also *Sxl* and *tra*, both involved in the SD pathway of *Drosophila* (*inset* in **Fig. 3.4D**) do not belong to the group of highly-divergent genes, as they have a mean amino acid divergence of about 0.09 and 0.9, respectively (**Fig. 3.4D**). Therefore, it can be argued that both *Dmrt-1L* and *Sox-H* may not only be SRGs, but may participate in bivalve SD as primary SDGs, which is reflected in their high AASD, as it is observed for *Sry* in mammals. As a matter of fact, if they were involved in SD just as intermediate actors of the signalling cascade, then we should have not observed a high AASD, as *Drosophila Sxl*, *tra*, and *dsx* seem to suggest. Overall, these patterns of molecular evolution concerning SRGs and SDG are also supported by the way SD regulatory networks evolve. As a matter of fact, it has been proposed that the sex-determining cascades tend to arise and be established with a bottom-up mechanism (**Wilkins, 1995; Mullon et al., 2012; Beukeboom and Perrin, 2014; Capel, 2017**). This means that the regulative relationships among genes at the bottom of the cascade are settled up prior to the regulative relationships among genes at the top and, consequently, upstream regulators are progressively recruited to fine-tune diverse SD signals. These evolutionary patterns eventually produce gene-regulatory networks in which the divergence of the upstream triggers is higher than that of downstream effectors, in terms of both identity and sequence composition (**Beukeboom and Perrin, 2014**). This mechanism has been proposed for *Drosophila* species (**Mullon et al., 2012**), *C. elegans* (**Stothard and Pilgrim, 2003**), and vertebrates, despite in the latter case it has been questioned several times (reviewed in **Capel, 2017**).

At this point, two main objections can be moved against our approach: (1) the distribution of AASD is not appropriate for this kind of inference, as it does not represent the true gene evolutionary (or substitution) rates (which instead are those usually employed when dealing

with SRGs and SDGs); (2) the three datasets are not comparable one to each other, as they take into consideration very different animal groups, with different taxonomic rankings and different divergence times (thus, the patterns of AASD are the products of other confounding factors not directly related to SD). Concerning the first objection, we are aware that the AASD does not represent the evolutionary rate itself, but rather its product. However, the two features are tightly linked, as on the long term highly-divergent proteins tend to be produced by genes with high evolutionary (or substitution) rates (**Echave et al., 2016**). By performing a GO-term enrichment, it emerged that highly-divergent genes of the mammal dataset are mainly involved in the immune response and male spermatogenesis (**Tab. 3.1** and **S10**), which are two processes notoriously connected with rapid sequence evolution (i.e., higher evolutionary rates; **Swanson and Vacquier, 2002**; **Murat et al., 2023**; **Vinkler et al., 2023**). Similarly, highly-divergent genes from the fruit fly dataset show an enrichment for GO-terms associated with meiotic-related functions (such as the formation of the synaptonemal complex by the *c(2)M*, *c(3)G*, *corona*, and *corolla* proteins; **Tab. 3.1** and **S10**), which again are known to be rapidly evolving (**Hemmer and Blumenstiel, 2016**). In other words, the test datasets allow us to directly link the high AASD (as found in this work) with high rates of sequence evolution (as found in previous works), as they represent well-studied and characterised model systems. This consideration can thus be extended also to the bivalve dataset: highly-divergent genes in terms of AASD, which include some DSFGs and show an enrichment for GO-terms associated to macromolecule metabolism and morphological development (**Tab. 3.1** and **S10**), are also genes with accelerated substitution rates. Concerning the second objection, we chose two test datasets with different characteristics as we wanted to check the extent of our hypothesis (i.e., molecular evolution can be used to look for putative primary SDGs in taxonomic-wide analyses). As a matter of fact, the difference in divergence times and taxonomy ranks for bivalves and therians (Late Cambrian, about 498 million years ago [Mya], **Song et al., 2023**; and Early Mesozoic, 166–123 Mya, **Álvarez-Carretero et al., 2022**, respectively) seems to not influence the sequence diversity of SRGs, as both *Dmrt-1L/Sox-H* for bivalves and *Sry* for mammals exhibit high AASD with respect to their own distributions, regardless of their age. *Dmrt-1L* and *Sox-H* (which are mollusc- and Bilateria-specific, respectively) are undoubtedly older than *Sry* (which, instead, emerged in the Theria common ancestor; **Foster et al., 1992**), but each of them can be considered a highly-divergent gene in bivalves and mammals, respectively (i.e., genes that are included in the 5% upper quantile of bivalve and

mammal AASD distributions). Conversely, the difference in divergence times and taxonomic ranks for *Drosophila* (Paleocene/Eocene boundary, about 56 Mya; **Russo et al., 2013**) may seem to be influencing the results for the dataset, resulting in a false negative. In other words, it can be argued that: (i) the genes included in the SD cascade of *Drosophila* (such as *Sxl*, *tra*, and *dsx*; *inset* in **Fig. 3.4D**) have indeed a high AASD, which however has not been detected by our methodological approach (for example, this may be traced back to the young diversification age of *Drosophila* species if compared to bivalves); (ii) the species included in the analysis are all congeneric, thus the sequence differentiation of SRGs may exist not at the amino acid level but at the nucleotide one. To better disentangle this issue and further discuss the fruit fly dataset, we repeated the analysis of the AASD only on species of the *Crassostrea* genus (*C. gigas*, *Crassostrea angulata*, *Crassostrea ariakensis*, and *Crassostrea virginica*), which are all congeneric and much younger (Middle Cretaceous, less than 100 Mya; **Qi et al., 2023**), thus comparable to *Drosophila*. Results showed that, even when analysing a smaller bivalve dataset, encompassing only 4 species of recent origin, the high AASD of *Dmrt-1L* persists, that is, *Dmrt-1L* is still grouped together with highly-divergent genes (**Fig. S15**). The same has not been recovered for *Sox-H*, which fell in genes from Group 3 (the group corresponding to the 95% interval of the AASD distribution) but still have the second highest AASD median value among DSFGs (**Fig. S15**).

Of course we should not expect that highly-divergent genes are only those involved in SD, but may participate also in other processes (as discussed earlier and shown by GO-term enrichments; **Tab. 3.1** and **S10**). Besides the genes of interest for SD (*Dmrt-1L/Sox-H* for bivalves, and *Sry* for mammals), also other components of the DSFG families have been retrieved with a high AASD, despite they have never been linked directly to SD so far: *Sox-F* in bivalves (**Fig. 3.3B**) and *Fox-D4* in mammals (**Fig. 3.4B**). This implies that our approach can't be used to unambiguously identify SDGs alone, as high AASD is exhibited also by many other genes. Instead, the analysis is meant to be used to detect highly-divergent genes and, subsequently, by comparison with literature and a more thorough and focused functional investigation, putative SDGs among them. In this sense, the mammal dataset exemplify the importance of putting the results of our pipeline (as those of any other comparative genomics analysis) into the correct evolutionary and genomic context: among DSFGs of mammals, two genes exhibit high AASD, one of which is directly related to SD (*Sry*), while the other has a function connected with neural development (*Fox-D4*; **Klein et al., 2013**). Thus, the high

AASD may arise either because of the involvement in the upper SD pathway or because of other life-history traits connected with the gene, respectively. Regarding bivalves, *Dmrt-1L* and *Sox-H* show a sharp connection with SD as a putative primary SDG, either when considering their molecular evolutionary features or when looking at their gene expression and possible function in gonad development (Naimi et al., 2009; Teaniniuraitemoana et al., 2014; Zhang et al., 2014; Capt et al., 2018; Li et al., 2018; Afonso et al., 2019; Liang et al., 2019; Yue et al., 2021). It is difficult to further speculate on the actual involvement in SD of *Dmrt-1L* and *Sox-H* without any additional information on their biology. Nonetheless, molecular evolution proves to be a valuable tool to investigate genes putatively involved in SD, and to identify major targets onto which dedicate future research effort.

3.5 Conclusions

In preparation.

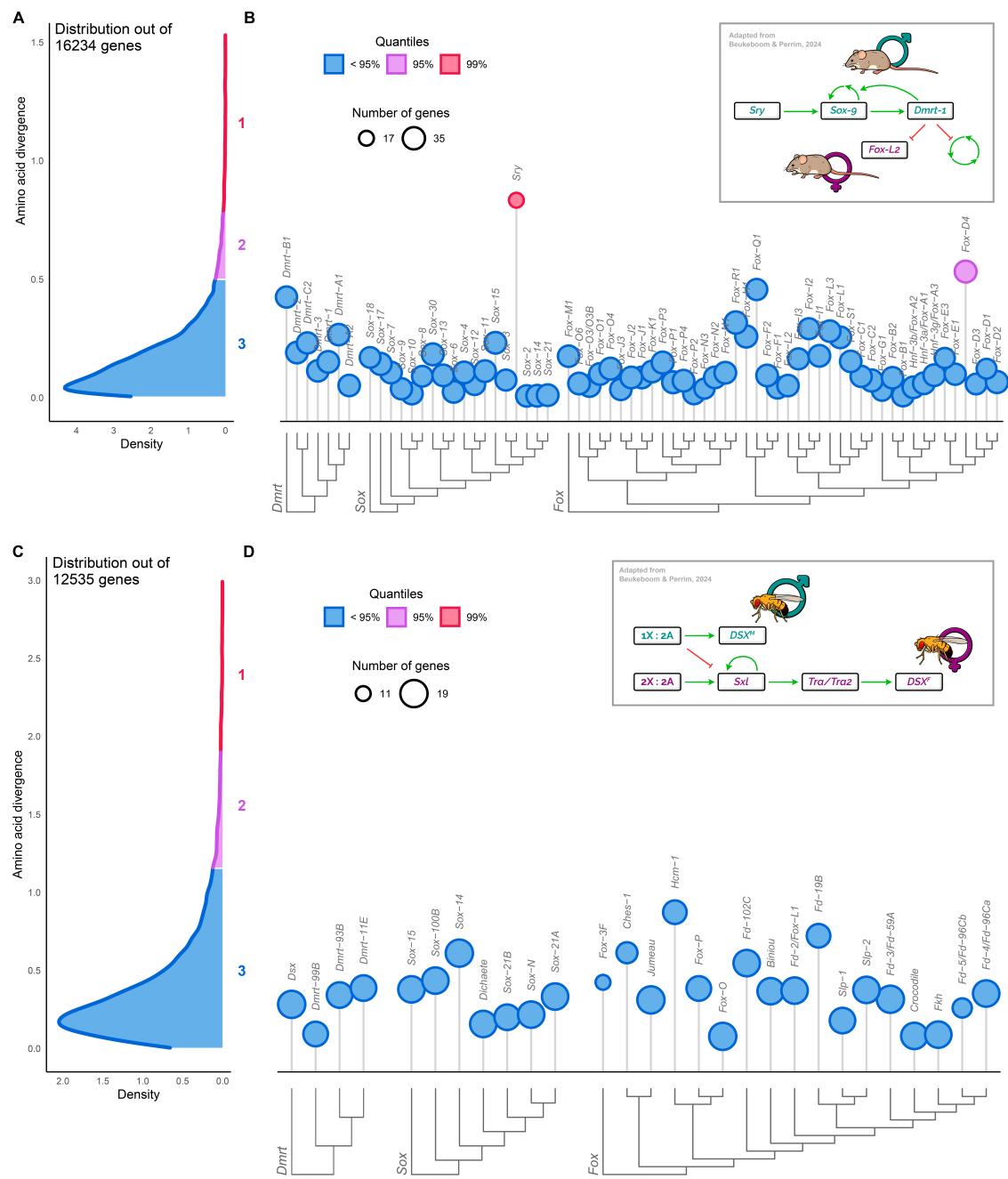


Figure 3.4 – Distribution of AASD of single-copy orthogroups in Mammalia (A) and Drosophila (C), including DSFG (B-D). The distributions of AASD in mammals and fruit flies have been computed on the median values of pairwise distances of over 16k and 12k SCOs, respectively. Genes have been divided according to their median AASD value into three different groups, which are indicated by different colors and increasing numbers (Groups 1, 2, and 3). Circle heights of DSFGs show the median value of their AASD, while the size indicates the number of represented species. DSFG trees are shown on the bottom (full trees can be found in Fig. S5–S7 for mammals and in Fig. S8–S10 for fruit flies). Insets: scheme of the sex-determination molecular pathways in *Mus musculus* and in *Drosophila melanogaster*, with shown the main genes involved (adapted from Beukeboom and Perrin, 2014). Green arrows indicate transcription activations, red arrows indicate transcription suppressions. X: sex chromosomes; A: autosomal chromosomes; *DSX^{M/F}*: *DSX* splicing variants present in males or females, respectively.

Chapter 4

Localisation of three sex-related genes and the germline marker *Vasa*/*Vasa* in the early developmental stages of *Mytilus galloprovincialis*

Filippo Nicolini^{1,2}, Sergey Nuzhdin³, Fabrizio Ghiselli¹, Andrea Luchetti¹, Liliana Milani¹

¹*Department of Biological, Geological and Environmental Science, University of Bologna, Bologna (BO), Italy.*

²*Fano Marine Center, Fano (PU), Italy.*

³*Department of Molecular and Computational Biology, University of Southern California, Los Angeles, CA, USA.*

In preparation.

4.1 Introduction

Despite the huge socio-economic and scientific importance of bivalves, the knowledge concerning the genetic and molecular bases of their sex determination (SD) system is scarce and overlooked (Breton et al., 2018; Nicolini, Ghiselli, et al., 2023). Several components of the Dmrt, Sox, and Fox gene (DSFG) families have been appointed as directly involved in SD by many works, mainly thanks to differential gene expression (DGE) analyses (e.g., Milani et al., 2013; Zhang et al., 2014; Capt et al., 2018; Shi et al., 2018), mRNA/protein visualisation (REFERENCE REFERENCE REFERENCE), RNA interference (RNAi) (REFERENCE REFERENCE REFERENCE) and quantitative real-time polymerase chain reaction (qRT-PCR) (Liang et al., 2019; Sun et al., 2022). For example, Li et al. (2018) found that *Fox-L2* and *Dmrt 1-like* (*Dmrt-1L*) are predominantly transcribed in ovaries and testes, respectively, of the Yesso scallop *Patinopacten yessoensis*, and that they contribute to establish the sexual identity of immature follicles at the molecular level prior to the morphological level. Liang et al. (2019) showed that *Sox-2* is involved in the differentiation of male gonads and spermatogenesis of the scallop *Chlamys farreri*, and that the knocked-out phenotype results in severe loss of both germ-cell mass and spermatogonia. Wang et al. (2020) speculated that *Fox-L2* is involved in the sex differentiation of female gonads in the freshwater mussel *Hyriopsis cumingii*. Overall, considerable effort has been made to characterise the transcription patterns of DSFGs of interest during the adult stage of bivalves, covering various reproductive phases, while little attention has been given to the embryo and larval stages. Nonetheless, early animal development may represent a crucial moment to the establishment of the sexual identity, as the transcription of sex-determination related genes (SRGs) and SD itself begins much earlier than the onset of gonad development and differentiation (even as early as the zygote formation; Richardson et al., 2023). In mammals, for example, the transcription of SRGs can be detected during the embryo preimplantation stage (before 4.5 days post fertilization [dpf]; reviewed in Richardson et al., 2023), while *Sex-determining region of chromosome Y (Sry)* realises its function as the male sex-determining gene (SDG) at 10.5 days post coitum (Beukeboom and Perrin, 2014). In *Drosophila melanogaster*, the early female splicing variant of *Sex-lethal (Sxl)*—which is the top regulator of the SD cascade and is activated by a mechanism of chromosome counting (*inset* in Fig. 3.4D), is transcribed during the syncytial stages of the embryo (i.e., before 2 hours post fertilization [hpf]; Salz and Erickson, 2010), when it establishes the sexual identity of the embryo through a cell-

autonomous mechanism. Therefore, the study of bivalve SD necessarily requires to consider also the early stages of embryonic and larval development, in order to obtain a comprehensive scenario of the process. Among bivalves, several species may constitute a model system particularly suitable to study the SD process during embryogenesis, because of the presence of the doubly uniparental inheritance (DUI) of mitochondria. This process—which involves the uniparental transmission of the maternal and paternal mitochondrial genomes through eggs and sperm, respectively, allows for an *a-priori* detection of the sexual identity of developing embryos, as early as the first cleavage division of the zygote: in female embryos, the sperm-inherited mitochondria assume a dispersed pattern between blastomeres; conversely, in males the sperm-inherited mitochondria stay assembled together, remain within one blastomere, and are eventually included in primordial germ cells (PGCs) (**Zouros, 2013; Ghiselli et al., 2019**).

Here we sought to expand the knowledge on the process of bivalve SD, by employing the Mediterranean mussel *Mytilus galloprovincialis* as a study system, which is a species exhibiting DUI. Particularly, we aimed to investigate the transcription patterns of three DSFG (namely, *Dmrt-1L*, *Sox-H*, and *Fox-L2*) during embryo and early larval stages. To this purpose, (i) we first performed a time-series DGE analysis by using the RNA-sequencing data published by **Miglioli et al. (2024)**; afterwards, (ii) we investigated the temporal and spatial transcription patterns of the DSFGs of interest through mRNA *in-situ* hybridization chain reaction (HCR). To obtain a more comprehensive developmental context for the transcription patterns of DSFGs, (iii) we also traced for the first time in *M. galloprovincialis* the process of the germline specification through mRNA *in-situ* HCR and immunolocalization of *Vasa*/Vasa, which is a traditionally-recognised marker of PGCs and germ cells (GCs) across Metazoa (**Extavour and Akam, 2003**). The specification and differentiation of GCs (which are part of the gonadal tissue in adults) is in fact a critical process in sexually reproducing multicellular organisms, as it provides the groundwork for the subsequent differentiation of sexually dimorphic gametes. Therefore, understanding the developmental pathway leading to the establishment of PGCs and GCs is essential to fully characterise the sex-determining process and how the sexual fate of PGCs/GCs is directed.

4.2 Materials and Methods

4.2.1 Time-series gene expression

Miglioli et al. (2024) recently produced one of the very first detailed developmental transcriptomes of the Mediterranean mussel *M. galloprovincialis*, spanning from the unfertilized oocyte to the larval stage at 72 hpf, with time points sampled every 4 hpf. A total of thirty different mRNA libraries was sequenced, consisting of fifteen developmental time points per two biological replicates each (**Tab. S11**). These data are extremely useful to thoroughly investigate the transcription patterns of genes throughout the first three days of the *M. galloprovincialis* development, to quantify the transcription level of target genes to be investigated with mRNA HCR experiments and to have an overview of the possible outcome from such analysis.

Raw reads were downloaded from the Sequence Read Archive (SRA) in NCBI (BioProject: PRJNA996031) and trimmed using Trimmomatic v0.39 (**Bolger et al., 2014**; **LEADING:5 TRAILING:5 SLIDINGWINDOW:4:15 MINLEN:65**). Read quality was checked using FastQC v0.12.1 (**Andrews et al., 2010**). Trimmed reads were mapped against the *M. galloprovincialis* annotated genome (GCA_900618805.1; **Gerdol et al., 2020**) using STAR v2.7.10b (**Dobin et al., 2013**) in ‘alignReads’ mode with default parameters. The resulting gene count matrix was extracted with StringTie v2.2.1 (**Pertea et al., 2015, 2016**) in expression estimation mode followed by the python script ‘prepDE.py’ (-1 99).

The resulting matrix was processed in R. Raw gene counts were normalised using the median of ratios method as implemented by the DESeq2 package (**Love et al., 2014**), and then transformed through the DESeq2 variance stabilising transformation (vst). Transformed gene counts were used to run a principal component analyses (PCA) and visualise sample clustering, and to plot expression values of *Vasa*, *Dmrt-1L*, *Sox-H*, and *Fox-L2* (hereafter collectively referred to as ‘target genes’). Normalised gene counts were instead used to run a time-series DGE analysis in ‘maSigPro’ (**Conesa et al., 2006**).

The entire pipeline was automated through custom python and bash scripts, which are available in a private repository on GitHub.

4.2.2 Sample collection, MitoTracker staining and fixation

Adult mussels were hand collected from various locations surrounding the AltaSea institute at the port of Los Angeles (CA, USA). Sampling took place during the spawning season of the species in California, i.e., from October 2023 to early January 2024.

Selected mussels were thoroughly cleaned from epibionts and placed in ice for approximately 30–60 minutes, then transferred in filtered artificial sea water (FASW) at 16 °C and acclimatized for 30 minutes. All the individuals were then placed in a common tank and spawning was induced by cyclical thermal shock, that is, by exposing mussels alternatively to FASW at 24–26 °C and 14–16 °C for a time of 30–40 min each. As soon as mussels started spawning, individuals were promptly removed from the common tank, carefully washed, air dried to remove contaminant gametes from the shell, and then allowed to continue spawning in isolated containers of about 250 mL with 16 °C FASW.

Both single and multiple crosses were performed: two males (M1, M2) and two females (F1, F2) were employed for single crosses; six males and six females were employed for multiple crosses, and gametes from the same sex were mixed. One hour after the spawning started, oocytes were filtered through a 75 over a 30 µm mesh, and aged in 1 L of FASW for 40–60 min, to allow them to assume a proper circular shape. Oocyte abundance was estimated under a stereomicroscope by eye counting the number of gametes in five aliquots of 1 mL, and then calculating the mean value. Sperm mitochondria were labelled with MitoTracker Red CMXRos (Thermo Fisher Scientific) at a working concentration of 500 nM for 30 min. MitoTracker is a fluorescent, vital and fixation-resistant mitochondrial dye and was used to be able to detect the sex of developing embryos (as early as the two-blastomere stage) according to the distribution pattern of sperm mitochondria (**Cao et al., 2004; Obata and Komaru, 2005**). From this step onward, samples were always kept in the dark.

Fertilisation was performed by mixing oocytes and sperm at a ratio of 1:10. Fertilisation success was checked after 20–30 min by the formation of polar bodies. The suspension was then carefully washed on a 30 µm mesh to remove excess sperm, and brought to a concentration of 250 zygotes/mL. The resulting suspension was transferred into cell-culture flasks of 40 mL and embryos/larvae were reared at 16±1 °C in the dark. Water was changed every 24 h. After 48 hpf, larvae were fed with the unicellular microalgae *Isochrysis galbana*, at a final concentration of about 100,000 cells/mL following **Helm et al. (2004)**.

Embryos/larvae were sampled at 1, 2, 3 and 4 hpf, and then every 12 h until 72 hpf. Proper development and vitality were checked under a stereomicroscope at every sampling time. After concentration with a mesh of proper size, embryos/larvae were fixed in 3.2% paraformaldehyde (PFA) in 1× PBS with 0.1% Tween 20 (PBS-Tw; 128 mM NaCl, 2 mM KCl, 8 mM Na₂HPO₄ · 2 H₂O, 2 mM KH₂PO₄) at 4 °C overnight under constant and gentle shaking. PBS was prepared with the following concentrations: 128 mM NaCl, 2 mM KCl, 8 mM Na₂HPO₄ · 2 H₂O, and 2 mM KH₂PO₄. Fixed samples were washed 3×20 min in 1× PBS with 0.1% Tween 20 (PBS-Tw) and then dehydrated 3×30 min in absolute methanol at room temperature (RT). Dehydrated samples were stored at –20 °C until usage.

4.2.3 mRNA *in-situ* hybridization chain reaction (HCR)

HCR probe design

Vasa, *Dmrt-1L*, *Sox-H*, and *Fox-L2* (the latter three are hereafter referred to as SRG) spliced-transcript nucleotide sequences of *M. galloprovincialis* were obtained from the previous analyses with OrthoFinder v2.5.5 (**Emms and Kelly, 2019**) on annotated bivalve genomes and transcriptomes (see **Section 3.2**). Accession numbers of spliced transcripts are 10B017427, 10B093608, 10B014180, and 10B094018, respectively. The ‘insitu_probe_generator’ script from the Ozpolat Lab (**Kuehn et al., 2022**) was used to generate pairs of probes specifically designed for third-generation HCR (**Choi et al., 2018**). The built-in BLASTN search against the annotated *M. galloprovincialis* transcriptome was employed to check for putative off-target bindings of probe pairs. B1-488, B2-647, B3-546, and B4-700 pairs of HCR amplifiers and fluorophores were chosen, as reported in **Tab. 4.1**. Resulting probes were synthesised by Integrated DNA Technologies (IDT™) in separate oligo pools.

mRNA *in-situ* HCR and microscope imaging

mRNA *in-situ* HCR in *M. galloprovincialis* embryos was performed following **Miglioli et al. (2024)**. All the steps were carried out in the dark to prevent MitoTracker from fading. Probe hybridization buffer, probe wash buffer and amplification buffer were manufactured by Molecular Instruments, Inc.

Dehydrated samples stored in methanol were washed 4×5 min and 1×10 min in PBS-Tw. Samples were then permeabilized for 30 min in a detergent solution (1.0% sodium dodecyl sul-

Target	Dye	HCR amplifier	HCR probe pairs	Excitation (nm)	Emission (nm)
dsDNA (nuclei)	DAPI	–	–	360	460
Sperm mitochondria	MitoTracker Red CMXRos	–	–	575	600
<i>Vasa</i> / <i>Vasa</i>	ALEXA-488/-488	B1/–	33/–	499	520
<i>Dmrt-1L</i>	ALEXA-647	B2	18	653	670
<i>Sox-H</i>	ALEXA-546	B3	22	557	575
<i>Fox-L2</i>	ALEXA-700	B4	28	685	700

Table 4.1 – Characteristics of fluorescent dyes used for each labelled target. HCR amplifiers and the number of probe sets (as in Tab. S12) are reported when applicable. Dyes for both *Vasa* and *Vasa* are reported.

fate [SDS], 0.5% Tween 20, 50 mM Tris–HCl, 1 mM ethylenediaminetetraacetic acid (EDTA), 150 mM NaCl), and washed again 2×5 min in PBS-Tw. Samples were prepared for the HCR detection stage by incubation in the probe hybridization buffer for 30 min at 37 °C. Detection stage was then performed with 4 nM of each probe set in hybridization solution overnight (>12 h) at 37 °C.

Excess probes were removed by washing 4×20 min with probe wash buffer at 37 °C and 3×5 min with 5× saline-sodium citrate with 0.1% Tween 20 (SSC-Tw; 5× SSC, 0.1% Tween 20) at RT. Samples were incubated for 30 min in the amplification buffer at RT. Hairpins were heated at 95 °C for 90 s and then snap-cooled at RT for 30 min. The amplification step of HCR was performed with 6 pmol of each hairpin in the amplification buffer overnight (>12 h) at RT.

Excess hairpins were removed by washing 2×5 min, 2×10 min, and 1×5 min with SSC-Tw. If not immediately mounted on slides, samples were stored in SSC-Tw at 4 °C. Otherwise, samples were immersed first in 50% glycerol and then in 75% glycerol, each for 30–60 min, and then mounted with VECTASHIELD®PLUS Antifade Mounting Medium with DAPI (H-2000). Slides were imaged on a Stellaris 5 Confocal Package system with the software Las X (Leica Microsystems). Each dye was imaged sequentially in a separate channel, to enhance the yield and avoid crosstalks. **Tab. 4.1** summarises the excitation and emission peaks for each dye. Images were then manipulated and post-produced using Fiji v2.14.0.

4.2.4 Immunolocalization of Vasa

M. galloprovincialis Vasa sequence was manually inspected through multiple sequence alignment with Vasa from other bivalves (data from **Chapter 3**) and several reference species (*Danio rerio* [Ddx4: NP_571132.1]; *Homo sapiens* [Ddx4: NP_077726.1]; *Mus musculus* [Ddx4: NP_001139357.1]; *D. melanogaster* [Vasa: NP_001260458.1]; *Caenorhabditis elegans* [GLH-1: NP_001262379.1, GLH-2: NP_491876.1, GLH-3: NP_491681.1, GLH-4: NP_491207.3]), to support commercial antibody specificity in *M. galloprovincialis*. The Vasa sequence from *D. rerio* was included as the polyclonal antibody was generated using the zebrafish protein variant (manufacturer indications; ab209710 by Abcam Limited). A maximum likelihood (ML) phylogenetic tree of Vasa genes and its paralog DDX3 (reference genes: *D. rerio* [Ddx3Xa: NP_001119895.1, Ddx3Xb: NP_571016.2]; *H. sapiens* [DDX3X: NP_001180346.1]; *M. musculus* [Pl10/Ddx3Xl: NP_149068.1]; *D. melanogaster* [Belle: NP_001262379.1]; *C. elegans* [LAF-1: NP_001254859.1, VBH-1: NP_001021793.1]) was built using IQTREE. The hidden Markov model (HMM) profile of the Asp-Glu-Ala-Asp/Asp-Glu-Ala-His box (DEAD/DEAH-box) signature domain for the amino acid guided alignment step, was built after the corresponding Pfam full database (PF00270). Methods are the same as in **Chapter 3**.

Vasa immunolocalization in *M. galloprovincialis* embryos was performed following **Milani et al. (2011)** with modifications. All the steps were carried out in the dark to prevent MitoTracker fluorescence from fading. Dehydrated samples stored in methanol were rinsed 3×10 min and 1×2 h in 1× Tris-buffered saline (TBS; 10 mM Tris–HCl, 155 mM NaCl), following an additional wash for 10 min with PBS. Samples were then digested for 6 min and 30 s with 0.01% pronase E (Merck) in PBS, and washed again 2×5 min in PBS. Permeabilization was performed in 1× TBS with Triton X-100 (TBS-Tx) 0.1% for 5 min at RT and in TBS-Tx 1% overnight at 4 °C.

After an additional rinse for 5 min in TBS-Tx 0.1%, non-specific binding sites were blocked with a TBS-Tx 0.1% solution containing 3% bovine serum albumin (BSA). Samples were then incubated at 4 °C for 32–48 h with primary anti-VASA/VAS antibody (polyclonal anti-VASA developed in rabbit; ab209710 by Abcam Limited), diluted 1:100.

Excess primary antibody was rinsed from samples with 4×30 min in TBS-Tx 0.1%, followed by an incubation of 1 hour in TBS-Tx 0.1% containing 3% BSA. Samples were then incubated at 4 °C for 24–32 h with secondary antibody HRP anti-rabbit in goat (Santa Cruz Biotechnology Inc.), diluted 1:400.

Excess secondary antibody was rinsed with 4×30 min in TBS-Tx 0.1% and 1×1 hour in 1%. Samples were immersed first in 50% glycerol and then in 75% glycerol, each for 30–60 min, and then mounted with VECTASHIELD®PLUS Antifade Mounting Medium with DAPI (H-2000). Slides were imaged on a Nikon A1R+ HD25 confocal microscope. Each dye was imaged sequentially in a separate channel, to enhance the yield and avoid any crosstalks. **Tab. 4.1** summarises the excitation and emission peaks for each dye. Images were then manipulated and post-produced using Fiji v2.14.0.

4.3 Results

4.3.1 Differential gene expression analysis of *Vasa* and SRGs in embryo time-series

Over 24 millions reads were mapped for each RNA-sequencing library (86.58% of the total input reads), with an average of 26.8 millions (**Tab. S11**). Of these, an average of 22 millions reads were uniquely mapped (71.86% of the total input reads), while an average of 4.5 millions were multi-mapped (14.72% of the total input reads). The average of unmapped reads was 4.1 millions (13.42% of the total input reads; **Tab. S11**). The PCA on normalised read counts returned well-clustered experimental groups for time points between 8 and 36 hpf, while for stages before 8 hpf and after 36 hpf, experimental groups are more homogeneous among each other (**Fig. 4.1A**). This situation may reflect major developmental dynamics during embryogenesis and larval development. As a matter of fact, before 8 hpf, the embryo undergoes segmentation and no big morphogenetic movements are usually detected. Between 8 and 13 hpf, instead, gastrulation begins, the embryo experiences strong morphogenetic rearrangements (such as the formation of embryonic layers) and the trochophore larva develops, all processes which are expected to be detected also at the molecular level. After 36 hpf, instead, the larva does not show any dramatic morphogenetic event, as the D-veliger is almost formed and the gross advanced larval morphology is established. The hierarchical clustering of differentially expressed genes computed by **Miglioli et al. (2024)** is concordant with this view.

Transcription levels of *Vasa*, SRGs, *Fox-B2*, and *Wnt-8a* were plotted individually (**Fig. 4.1B**) to obtain a proxy of the expected outcome of HCR. *Fox-B2* and *Wnt-8a* were employed as control genes to get support for handling of data and of the pipeline, as they were also analysed

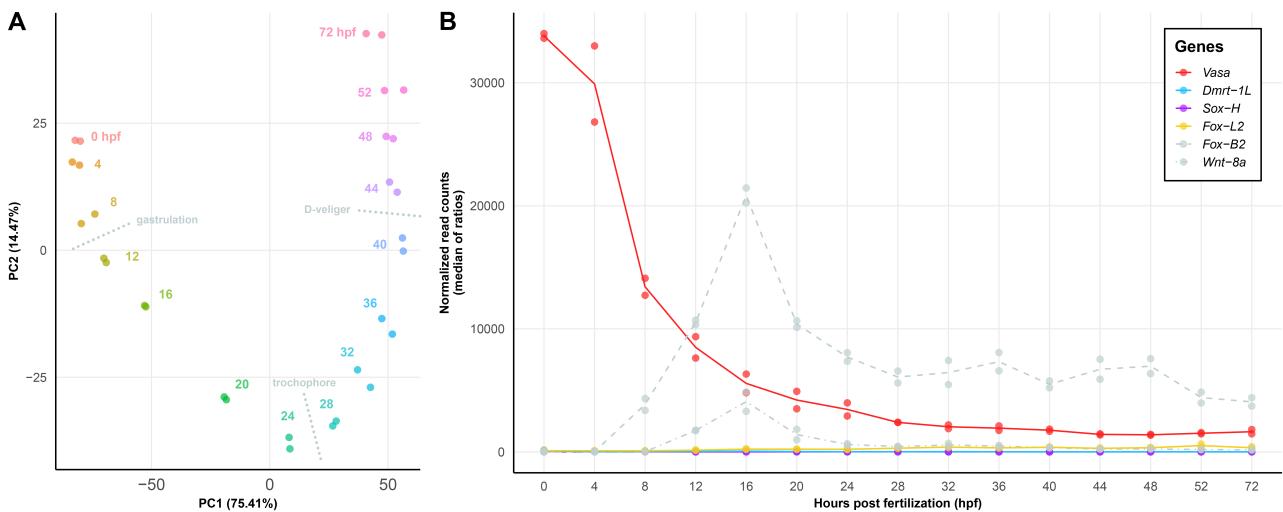


Figure 4.1 – PCA of DESeq2-normalised read counts (A) and transcription levels of target and reference genes (B). (A) Principal components (PC) 1 and 2 are plotted in the x and y axes, respectively; the proportion of variance explained by each PC is shown in parentheses. Sampled time-points are shown in different colours and are indicated by the hours hours post fertilization (hpf). Major developmental transitions are marked with dotted lines. PCA has been performed on vst-transformed, normalised read counts (DESeq2 median of ratios). (B) Transcription levels of target (*Vasa*, *Dmrt-1L*, *Sox-H*, and *Fox-L2*) and reference genes (*Wnt-8a* and *Fox-B2*) as expressed by normalised read counts (DESeq2 median of ratios).

by Miglioli et al. (2024). The transcription of both genes starts at 4 and 8 hpf, respectively, reaches a peak at 16 hpf, and then constantly decreases (fig:deseq2-B). *Vasa* transcripts are highly abundant in unfertilized oocytes and in embryos 4 hpf, then constantly decrease throughout time; conversely, *Fox-L2* transcripts increase from 12 hpf onward (fig:deseq2-B). Both *Dmrt-1L* and *Sox-H*, instead, show low or null levels of transcriptions throughout the entire time series (fig:deseq2-B).

The maSigPro DGE analysis of the *M. galloprovincialis* developmental time series found 13,067 differentially expressed genes (about 17% of the analysed genes) and clustered them into 9 different groups, according to their specific transcriptional profiles (Fig. 4.2). Among the genes of interest, only *Vasa* and *Fox-L2* showed a significantly different transcriptional profile throughout the time series, and were included in clusters 3 and 1, respectively. As already discussed, *Vasa* and *Fox-L2* transcription levels show an opposite tendency, with the former decreasing and the latter increasing throughout time. Both *Dmrt-1L* and *Sox-H* were not found to be differentially transcribed by maSigPro and, thus, were not included in any cluster. The same holds true for *Wnt-8a* and *Fox-B2*.

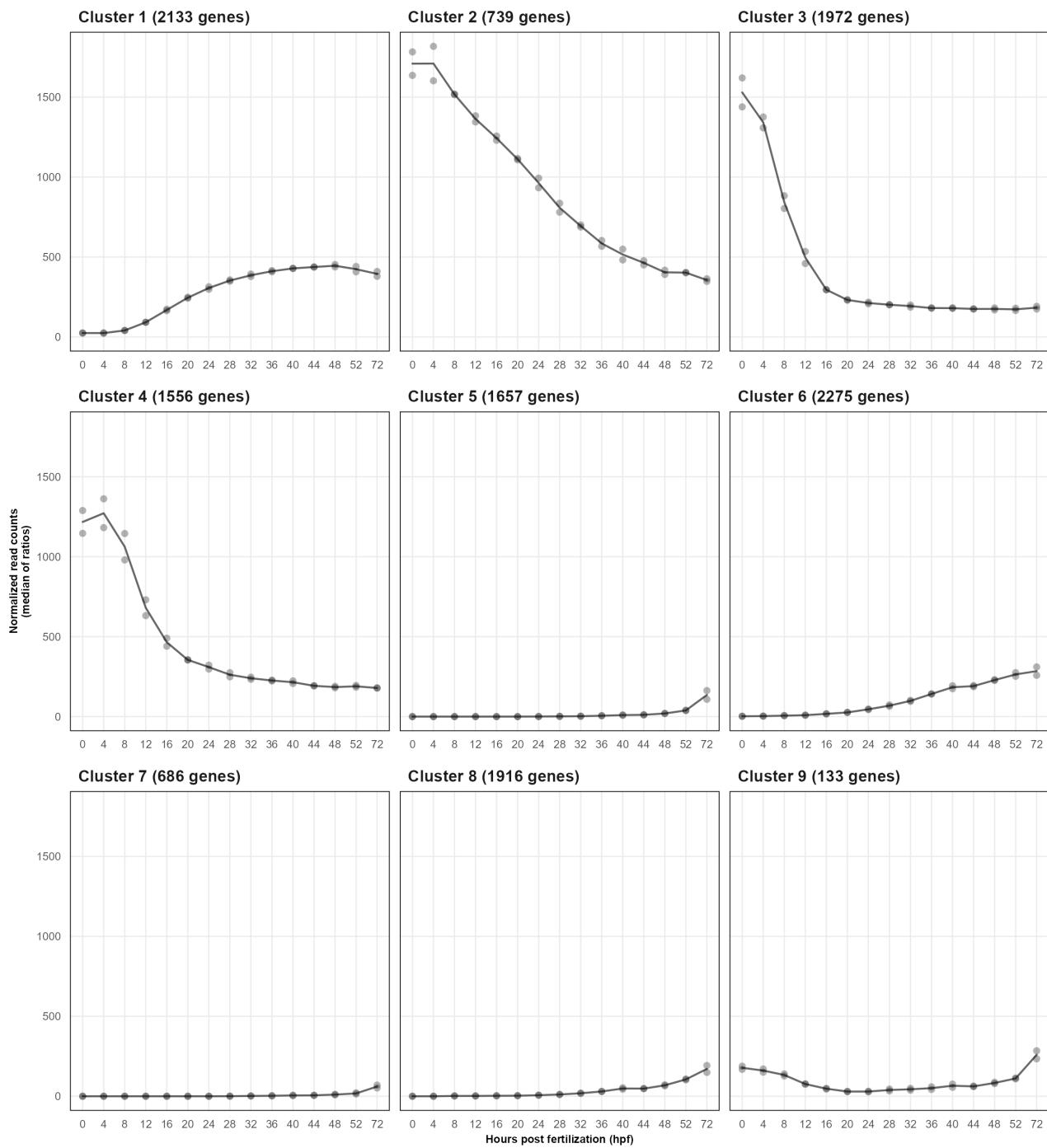


Figure 4.2 – Transcription patterns of differentially-expressed genes as inferred by maSigPro. Genes are divided into 9 different clusters according to their transcription patterns throughout 15 sampled time points. Median values of the two biological replicates are shown for each time point and represented by points. Mean values are shown for each time point and represented by solid lines.

4.3.2 mRNA *in-situ* HCR of *Vasa* and SRGs

Overall, a total of 80 adult *M. galloprovincialis* individuals were sampled and staged for thermal-shock induced spawning. Of these, 8 males and 8 females were eventually selected as parents for single (2 of each sex) and multiple (6 of each sex) crosses, on the basis of their gamete quality (i.e., presence sperm motility, and oocyte transparency and rounded shape). MitoTracker labelling was successfully retained in developing embryos of *M. galloprovincialis* until 12 hpf. After that stage, the stained sperm mitochondria were difficult to detect, and so was the dispersal pattern to establish the sexual identity.

After embryo rearing, fixation, and mRNA *in-situ* HCR of target genes, a total of 16 oocytes, 81 embryos and 33 mussel larvae were imaged (**Tab. 4.2**). Of these, on the basis of sperm mitochondria dispersal patterns, 55 were females (dispersed pattern), 28 were males (aggregated pattern) and 38 were of indeterminable sex (ambiguous pattern or unlabelled sperm mitochondria). For each stage, negative controls were also imaged (final count of 36), by staining just sperm mitochondria with MitoTracker and nuclei with DAPI, and going through the HCR protocol without adding probes in the hybridization step. Overall, a total of 137 samples were imaged.

The ‘insitu_probe_generator’ script (**Kuehn et al., 2022**) generated: (i) 33 probe pairs conjugated with hairpin B1 and ALEXA-488 for *Vasa*; (ii) 32 probe pairs conjugated with hairpin B2 and ALEXA-647 for *Dmrt-1L*; (iii) 27 probe pairs conjugated with hairpin B3 and ALEXA-546 for *Sox-H*; and (iv) 28 probe pairs conjugated with hairpin B4 and ALEXA-700 for *Fox-L2* (**Tab. 4.1** and **S12**).

HCR labelling of genes of interest proved to be concordant with results obtained from RNA-seq analysis (see **Section 4.3.1**). Concerning *Vasa*, it has been detected throughout every sampled stage (Fig. 3A): transcripts were identified quite homogeneously in the cytoplasm of unfertilized oocytes, 2-, 4-, and 8-cell embryos; in gastrulae, *Vasa* is located mainly in the ingressed cells; in trochophores, it forms a cup-like structure in the region opposite to the shell-field; in D-larvae, it is mainly retained in two central areas adjacent to the valves (right and left sides of the larvae) in a sort of a comma-shaped region. Concerning *Dmrt-1L* (Fig. 3B), final images were quite noisy and showed putative non-specific staining at the level of embryo external surface and larvae shell, which may have interfered with the true signal of HCR for this gene; in any case, no clear labelling distribution pattern was found in embryos of both

Stage	Experiment	Females	Males	Undetermined	Total
Oocytes	HCR	–	–	–	11
2-cell embryos	HCR	8	9	1	18
4-cell embryos	HCR	9	3	0	12
8-cell embryos	HCR	11	3	0	14
12-hpf embryos	HCR	7	6	0	13
Total embryos	HCR	35	21	1	57
24-hpf larvae	HCR	0	1	11	12
48-hpf larvae	HCR	0	0	11	11
72-hpf larvae	HCR	1	1	8	10
Total larvae	HCR	1	2	30	33
Oocytes	Negative control	–	–	–	5
2-cell embryos	Negative control	7	2	0	9
4-cell embryos	Negative control	7	2	0	9
8-cell embryos	Negative control	5	1	0	6
12-hpf embryos	Negative control	0	0	3	3
Total embryos	Negative control	19	5	3	27
Total larvae	Negative control	0	0	4	4
Total imaged samples	All	55	28	38	137

Table 4.2 – Number of imaged samples, divided by developmental stage, experiment, and sex.

sexes. *Sox-H* mRNAs (Fig. 3C) were not detected during the imaged developmental stages. Conversely, *Fox-L2* transcripts have been detected starting from the 8-cell stage—where they are homogeneously present, to the D-veliger larvae—where they appear to be mostly co-localized with Vasa (Fig. 3D–E). Imaging of control samples (i.e., without mRNA *in-situ* staining) can be found in Supp. Fig. S1.

4.4 Discussion

In preparation.

References

- Abascal, F., Zardoya, R., & Telford, M. J. (2010). Translatorx: Multiple alignment of nucleotide sequences guided by amino acid translations. *Nucleic acids research*, 38(suppl_2), W7–W13.
- Abbott, J. K. (2011). Intra-locus sexual conflict and sexually antagonistic genetic variation in hermaphroditic animals. *Proceedings of the Royal Society B: Biological Sciences*, 278(1703), 161–169.
- Afonso, L. F., Americo, J. A., Soares-Souza, G. B., Torres, A. L. Q., Wajsenzon, I. J. R., & Rebelo, M. d. F. (2019). Gonad transcriptome of golden mussel *Limnoperna fortunei* reveals potential sex differentiation genes. *biorxiv*, 818757.
- Albertin, C. B., Simakov, O., Mitros, T., Wang, Z. Y., Pungor, J. R., Edsinger-Gonzales, E., Brenner, S., Ragsdale, C. W., & Rokhsar, D. S. (2015). The octopus genome and the evolution of cephalopod neural and morphological novelties. *Nature*, 524(7564), 220–224.
- Alexa, A., & Rahnenführer, J. (2009). Gene set enrichment analysis with topgo. *Bioconductor Improv*, 27, 1–26.
- Altenhoff, A. M., Warwick Vesztrocy, A., Bernard, C., Train, C.-M., Nicheperovich, A., Prieto Baños, S., Julca, I., Moi, D., Nevers, Y., Majidian, S., et al. (2024). Oma orthology in 2024: Improved prokaryote coverage, ancestral and extant go enrichment, a revamped synteny viewer and more in the oma ecosystem. *Nucleic Acids Research*, 52(D1), D513–D521.
- Álvarez-Carretero, S., Tamuri, A. U., Battini, M., Nascimento, F. F., Carlisle, E., Asher, R. J., Yang, Z., Donoghue, P. C., & Dos Reis, M. (2022). A species-level timeline of mammal evolution integrating phylogenomic data. *Nature*, 602(7896), 263–267.
- Andrews, S., et al. (2010). Fastqc: A quality control tool for high throughput sequence data.

- Bachtrog, D., Mank, J. E., Peichel, C. L., Kirkpatrick, M., Otto, S. P., Ashman, T.-L., Hahn, M. W., Kitano, J., Mayrose, I., Ming, R., et al. (2014). Sex determination: Why so many ways of doing it? *PLoS biology*, 12(7), e1001899.
- Bai, C.-M., Xin, L.-S., Rosani, U., Wu, B., Wang, Q.-C., Duan, X.-K., Liu, Z.-H., & Wang, C.-M. (2019). Chromosomal-level assembly of the blood clam, *Scapharca (Anadara) broughtonii*, using long sequence reads and hi-c. *GigaScience*, 8(7), giz067.
- Baldwin-Brown, J. G., Weeks, S. C., & Long, A. D. (2018). A new standard for crustacean genomes: The highly contiguous, annotated genome assembly of the clam shrimp *Euleimnadia texana* reveals hox gene order and identifies the sex chromosome. *Genome biology and evolution*, 10(1), 143–156.
- Baral, S., Arumugam, G., Deshmukh, R., & Kunte, K. (2019). Genetic architecture and sex-specific selection govern modular, male-biased evolution of *doublesex*. *Science advances*, 5(5), eaau3753.
- Benayoun, B. A., Caburet, S., & Veitia, R. A. (2011). Forkhead transcription factors: Key players in health and disease. *Trends in Genetics*, 27(6), 224–232.
- Beukeboom, L. W., & Perrin, N. (2014). *The evolution of sex determination*. Oxford University Press.
- Bewick, A. J., Anderson, D. W., & Evans, B. J. (2011). Evolution of the closely related, sex-related genes *DM-W* and *DMRT1* in african clawed frogs (*Xenopus*). *Evolution*, 65(3), 698–712.
- Bolger, A. M., Lohse, M., & Usadel, B. (2014). Trimmomatic: A flexible trimmer for illumina sequence data. *Bioinformatics*, 30(15), 2114–2120.
- Bredemeyer, K. R., Hillier, L., Harris, A. J., Hughes, G. M., Foley, N. M., Lawless, C., Carroll, R. A., Storer, J. M., Batzer, M. A., Rice, E. S., et al. (2023). Single-haplotype comparative genomics provides insights into lineage-specific structural variation during cat evolution. *Nature genetics*, 55(11), 1953–1963.
- Breton, S., Capt, C., Guerra, D., & Stewart, D. (2018). Sex-determining mechanisms in bivalves. In J. L. Leonard (Ed.), *Transitions between sexual systems: Understanding the mechanisms of, and pathways between, dioecy, hermaphroditism and other sexual systems* (pp. 165–192). Springer International Publishing.

- Breton, S., Stewart, D. T., Brémaud, J., Havird, J. C., Smith, C. H., & Hoeh, W. R. (2022). Did doubly uniparental inheritance (dui) of mtDNA originate as a cytoplasmic male sterility (cms) system? *BioEssays*, 44(4), 2100283.
- Briones, C., Nuñez, J. J., Pérez, M., Espinoza-Rojas, D., Molina-Quiroz, C., & Guiñez, R. (2018). De novo male gonad transcriptome draft for the marine mussel *Perumytilus purpuratus* with a focus on its reproductive-related proteins. *Journal of Genomics*, 6, 127.
- Buchfink, B., Xie, C., & Huson, D. H. (2015). Fast and sensitive protein alignment using diamond. *Nature methods*, 12(1), 59–60.
- Calcino, A. D., de Oliveira, A. L., Simakov, O., Schwaha, T., Zieger, E., Wollesen, T., & Wanninger, A. (2019). The quagga mussel genome and the evolution of freshwater tolerance. *DNA Research*, 26(5), 411–422.
- Cao, L., Kenchington, E., & Zouros, E. (2004). Differential segregation patterns of sperm mitochondria in embryos of the blue mussel (*Mytilus edulis*). *Genetics*, 166(2), 883–894.
- Capel, B. (2017). Vertebrate sex determination: Evolutionary plasticity of a fundamental switch. *Nature Reviews Genetics*, 18(11), 675–689.
- Capella-Gutiérrez, S., Silla-Martínez, J. M., & Gabaldón, T. (2009). Trimal: A tool for automated alignment trimming in large-scale phylogenetic analyses. *Bioinformatics*, 25(15), 1972–1973.
- Capt, C., Bouvet, K., Guerra, D., Robicheau, B. M., Stewart, D. T., Pante, E., & Breton, S. (2020). Unorthodox features in two venerid bivalves with doubly uniparental inheritance of mitochondria. *Scientific reports*, 10(1), 1087.
- Capt, C., Renaut, S., Ghiselli, F., Milani, L., Johnson, N. A., Sietman, B. E., Stewart, D. T., & Breton, S. (2018). Deciphering the link between doubly uniparental inheritance of mtDNA and sex determination in bivalves: Clues from comparative transcriptomics. *Genome biology and evolution*, 10(2), 577–590.
- Chakraborty, M., Chang, C.-H., Khost, D. E., Vedanayagam, J., Adrián, J. R., Liao, Y., Montooth, K. L., Meiklejohn, C. D., Larracuente, A. M., & Emerson, J. (2021). Evolution of genome structure in the *Drosophila simulans* species complex. *Genome research*, 31(3), 380–396.
- Chen, H., Xiao, G., Chai, X., Lin, X., Fang, J., & Teng, S. (2017). Transcriptome analysis of sex-related genes in the blood clam *Tegillarca granosa*. *PLoS One*, 12(9), e0184584.

- Choi, H. M., Schwarzkopf, M., Fornace, M. E., Acharya, A., Artavanis, G., Stegmaier, J., Cunha, A., & Pierce, N. A. (2018). Third-generation *in situ* hybridization chain reaction: Multiplexed, quantitative, sensitive, versatile, robust. *Development*, 145(12), dev165753.
- Civetta, A., & Singh, R. S. (1998). Sex-related genes, directional sexual selection, and speciation. *Molecular biology and evolution*, 15(7), 901–909.
- Collin, R. (2013). Phylogenetic patterns and phenotypic plasticity of molluscan sexual systems. *Integrative and Comparative Biology*, 53(4), 723–735.
- Conesa, A., Nueda, M. J., Ferrer, A., & Talón, M. (2006). Masigpro: A method to identify significantly differential expression profiles in time-course microarray experiments. *Bioinformatics*, 22(9), 1096–1102.
- Corrochano-Fraile, A., Davie, A., Carboni, S., & Bekaert, M. (2022). Evidence of multiple genome duplication events in *Mytilus* evolution. *BMC genomics*, 23(1), 340.
- Cutter, A. D., & Ward, S. (2005). Sexual and temporal dynamics of molecular evolution in *C. elegans* development. *Molecular Biology and Evolution*, 22(1), 178–188.
- Dainat, J., Hereñú, D., Davis, E., Crouch, K., Sol, L., & Agostinho, N. (2022). Another gff analysis toolkit to handle annotations in any gtf/gff format (version v1. 0). *Zenodo. doi, 10.*
- de Bono, M., & Hodgkin, J. (1996). Evolution of sex determination in *Caenorhabditis*: Unusually high divergence of *tra-1* and its functional consequences. *Genetics*, 144(2), 587–595.
- Deng, T., Pang, C., Lu, X., Zhu, P., Duan, A., Tan, Z., Huang, J., Li, H., Chen, M., & Liang, X. (2016). *De novo* transcriptome assembly of the chinese swamp buffalo by rna sequencing and ssr marker discovery. *PLoS One*, 11(1), e0147132.
- Dermauw, W., Van Leeuwen, T., & Feyereisen, R. (2020). Diversity and evolution of the p450 family in arthropods. *Insect biochemistry and molecular biology*, 127, 103490.
- Dobin, A., Davis, C. A., Schlesinger, F., Drenkow, J., Zaleski, C., Jha, S., Batut, P., Chaisson, M., & Gingeras, T. R. (2013). Star: Ultrafast universal rna-seq aligner. *Bioinformatics*, 29(1), 15–21.
- Dong, C., Zhang, L., Xia, S., Sosa, D., Arsala, D., & Long, M. (2022). New gene evolution with subcellular expression patterns detected in pacbio-sequenced genomes of *Drosophila* genus. *bioRxiv*, 2022–11.

- Eads, B. D., Colbourne, J. K., Bohuski, E., & Andrews, J. (2007). Profiling sex-biased gene expression during parthenogenetic reproduction in *Daphnia pulex*. *BMC genomics*, 8, 1–14.
- Echave, J., Spielman, S. J., & Wilke, C. O. (2016). Causes of evolutionary rate variation among protein sites. *Nature Reviews Genetics*, 17(2), 109–121.
- Eddy, S. R. (2011). Accelerated profile hmm searches. *PLoS computational biology*, 7(10), e1002195.
- Elbers, J. P., Rogers, M. F., Perelman, P. L., Proskuryakova, A. A., Serdyukova, N. A., Johnson, W. E., Horin, P., Corander, J., Murphy, D., & Burger, P. A. (2019). Improving illumina assemblies with hi-c and long reads: An example with the north african dromedary. *Molecular Ecology Resources*, 19(4), 1015–1026.
- Ellegren, H., & Parsch, J. (2007). The evolution of sex-biased genes and sex-biased gene expression. *Nature Reviews Genetics*, 8(9), 689–698.
- Emms, D. M., & Kelly, S. (2019). Orthofinder: Phylogenetic orthology inference for comparative genomics. *Genome biology*, 20, 1–14.
- Evensen, K. G., Robinson, W. E., Krick, K., Murray, H. M., & Poynton, H. C. (2022). Comparative phylotranscriptomics reveals putative sex differentiating genes across eight diverse bivalve species. *Comparative Biochemistry and Physiology Part D: Genomics and Proteomics*, 41, 100952.
- Extavour, C. G., & Akam, M. (2003). Mechanisms of germ cell specification across the metazoans: Epigenesis and preformation. *Development*, 130(24), 5869–5884.
- Fan, H., Wu, Q., Wei, F., Yang, F., Ng, B. L., & Hu, Y. (2019). Chromosome-level genome assembly for giant panda provides novel insights into carnivora chromosome evolution. *Genome biology*, 20, 1–12.
- Farhat, S., Bonnivard, E., Pales Espinosa, E., Tanguy, A., Boutet, I., Guiglielmoni, N., Flot, J.-F., & Allam, B. (2022). Comparative analysis of the *Mercenaria mercenaria* genome provides insights into the diversity of transposable elements and immune molecules in bivalve mollusks. *BMC genomics*, 23(1), 192.
- Foote, A. D., Liu, Y., Thomas, G. W., Vinař, T., Alföldi, J., Deng, J., Dugan, S., van Elk, C. E., Hunter, M. E., Joshi, V., et al. (2015). Convergent evolution of the genomes of marine mammals. *Nature genetics*, 47(3), 272–275.

- Foster, J. W., Brennan, F. E., Hampikian, G. K., Goodfellow, P. N., Sinclair, A. H., Lovell-Badge, R., Selwood, L., Renfree, M. B., Cooper, D. W., & Marshall Graves, J. A. (1992). Evolution of sex determination and the y chromosome: *SRY*-related sequences in marsupials. *Nature*, 359(6395), 531–533.
- Fouret, J., Brunet, F. G., Binet, M., Aurine, N., Enchéry, F., Croze, S., Guinier, M., Goumaidi, A., Preininger, D., Volff, J.-N., et al. (2020). Sequencing the genome of indian flying fox, natural reservoir of nipah virus, using hybrid assembly and conservative secondary scaffolding. *Frontiers in Microbiology*, 11, 1807.
- Gerdol, M., Moreira, R., Cruz, F., Gómez-Garrido, J., Vlasova, A., Rosani, U., Venier, P., Naranjo-Ortiz, M. A., Murgarella, M., Greco, S., et al. (2020). Massive gene presence-absence variation shapes an open pan-genome in the mediterranean mussel. *Genome biology*, 21, 1–21.
- Ghiselli, F., Iannello, M., Piccinini, G., & Milani, L. (2021). Bivalve molluscs as model systems for studying mitochondrial biology. *Integrative and Comparative Biology*, 61(5), 1699–1714.
- Ghiselli, F., Iannello, M., Puccio, G., Chang, P. L., Plazzi, F., Nuzhdin, S. V., & Passamonti, M. (2018). Comparative transcriptomics in two bivalve species offers different perspectives on the evolution of sex-biased genes. *Genome biology and evolution*, 10(6), 1389–1402.
- Ghiselli, F., Maurizii, M. G., Reunov, A., Ariño-Bassols, H., Cifaldi, C., Pecci, A., Alexandrova, Y., Bettini, S., Passamonti, M., Franceschini, V., et al. (2019). Natural heteroplasmy and mitochondrial inheritance in bivalve molluscs. *Integrative and comparative biology*, 59(4), 1016–1032.
- Ghiselli, F., Milani, L., Guerra, D., Chang, P. L., Breton, S., Nuzhdin, S. V., & Passamonti, M. (2013). Structure, transcription, and variability of metazoan mitochondrial genome: Perspectives from an unusual mitochondrial inheritance system. *Genome biology and evolution*, 5(8), 1535–1554.
- Gomes-dos-Santos, A., Lopes-Lima, M., Castro, L. F. C., & Froufe, E. (2020). Molluscan genomics: The road so far and the way forward. *Hydrobiologia*, 847(7), 1705–1726.
- Gomes-dos-Santos, A., Lopes-Lima, M., Machado, A. M., Marcos Ramos, A., Usié, A., Bolotov, I. N., Vikhrev, I. V., Breton, S., Castro, L. F. C., da Fonseca, R. R., et al. (2021). The crown pearl: A draft genome assembly of the european freshwater pearl mussel *Margaritifera margaritifera* (linnaeus, 1758). *DNA research*, 28(2), dsab002.

- Gomes-dos-Santos, A., Lopes-Lima, M., Machado, M. A., Teixeira, A., C. Castro, L. F., & Froufe, E. (2023). Pacbio hi-fi genome assembly of the iberian dolphin freshwater mussel *Unio delphinus* spengler, 1793. *Scientific Data*, 10(1), 340.
- Gómez-Chiarri, M., Warren, W. C., Guo, X., & Proestou, D. (2015). Developing tools for the study of molluscan immunity: The sequencing of the genome of the eastern oyster, *Crassostrea virginica*. *Fish & shellfish immunology*, 46(1), 2–4.
- González, V. L., Andrade, S. C., Bieler, R., Collins, T. M., Dunn, C. W., Mikkelsen, P. M., Taylor, J. D., & Giribet, G. (2015). A phylogenetic backbone for bivalvia: An rna-seq approach. *Proceedings of the Royal Society B: Biological Sciences*, 282(1801), 20142332.
- Grabherr, M. G., Haas, B. J., Yassour, M., Levin, J. Z., Thompson, D. A., Amit, I., Adiconis, X., Fan, L., Raychowdhury, R., Zeng, Q., et al. (2011). Trinity: Reconstructing a full-length transcriptome without a genome from rna-seq data. *Nature biotechnology*, 29(7), 644.
- Grath, S., & Parsch, J. (2016). Sex-biased gene expression. *Annual review of genetics*, 50, 29–44.
- Grau-Bové, X., & Sebé-Pedrós, A. (2021). Orthology clusters from gene trees with possvm. *Molecular Biology and Evolution*, 38(11), 5204–5208.
- Gusman, A., Lecomte, S., Stewart, D. T., Passamonti, M., & Breton, S. (2016). Pursuing the quest for better understanding the taxonomic distribution of the system of doubly uniparental inheritance of mtDNA. *PeerJ*, 4, e2760.
- Haag, E. S., & Doty, A. V. (2005). Sex determination across evolution: Connecting the dots. *PLoS biology*, 3(1), e21.
- Haas, B. J. (n.d.). *Transdecoder*. <https://github.com/TransDecoder>
- Habtewold, T., Wagah, M., Tambwe, M. M., Moore, S., Windbichler, N., Christophides, G., Johnson, H., Heaton, H., Collins, J., Krasheninnikova, K., et al. (2023). A chromosomal reference genome sequence for the malaria mosquito, *Anopheles gambiae*, giles, 1902, ifakara strain. *Wellcome Open Research*, 8.
- Haerty, W., Jagadeeshan, S., Kulathinal, R. J., Wong, A., Ravi Ram, K., Sirot, L. K., Levesque, L., Artieri, C. G., Wolfner, M. F., Civetta, A., et al. (2007). Evolution in the fast lane: Rapidly evolving sex-related genes in *Drosophila*. *Genetics*, 177(3), 1321–1335.
- Han, F., Wang, Z., Wu, F., Liu, Z., Huang, B., & Wang, D. (2010). Characterization, phylogeny, alternative splicing and expression of *Sox30* gene. *BMC Molecular Biology*, 11, 1–11.

- Han, W., Liu, L., Wang, J., Wei, H., Li, Y., Zhang, L., Guo, Z., Li, Y., Liu, T., Zeng, Q., et al. (2022). Ancient homomorphy of molluscan sex chromosomes sustained by reversible sex-biased genes and sex determiner translocation. *Nature Ecology & Evolution*, 1–16.
- Hart, S. F., Yonemitsu, M. A., Giersch, R. M., Garrett, F. E., Beal, B. F., Arriagada, G., Davis, B. W., Ostrander, E. A., Goff, S. P., & Metzger, M. J. (2023). Centuries of genome instability and evolution in soft-shell clam, *Mya arenaria*, bivalve transmissible neoplasia. *Nature Cancer*, 4(11), 1561–1574.
- Haszprunar, G., & Wanninger, A. (2012). Molluscs. *Current Biology*, 22(13), R510–R514.
- Heenan, P., Zondag, L., & Wilson, M. J. (2016). Evolution of the sox gene family within the chordate phylum. *Gene*, 575(2), 385–392.
- Helm, M. M., Bourne, N., & Lovatelli, A. (2004). *Hatchery culture of bivalves: A practical manual*.
- Hemmer, L. W., & Blumenstiel, J. P. (2016). Holding it together: Rapid evolution and positive selection in the synaptonemal complex of *Drosophila*. *BMC evolutionary biology*, 16, 1–17.
- Hoskins, R. A., Carlson, J. W., Wan, K. H., Park, S., Mendez, I., Galle, S. E., Booth, B. W., Pfeiffer, B. D., George, R. A., Svirskas, R., et al. (2015). The release 6 reference sequence of the *Drosophila melanogaster* genome. *Genome research*, 25(3), 445–458.
- Iannello, M., Forni, G., Piccinini, G., Xu, R., Martelossi, J., Ghiselli, F., & Milani, L. (2023). Signatures of extreme longevity: A perspective from bivalve molecular evolution. *Genome Biology and Evolution*, 15(11), evad159.
- Inoue, K., Yoshioka, Y., Tanaka, H., Kinjo, A., Sassa, M., Ueda, I., Shizato, C., Toyoda, A., & Itoh, T. (2021). Genomics and transcriptomics of the green mussel explain the durability of its byssus. *Scientific Reports*, 11(1), 5992.
- Ip, J. C.-H., Xu, T., Sun, J., Li, R., Chen, C., Lan, Y., Han, Z., Zhang, H., Wei, J., Wang, H., et al. (2021). Host–endosymbiont genome integration in a deep-sea chemosymbiotic clam. *Molecular Biology and Evolution*, 38(2), 502–518.
- Jackson, B. C., Carpenter, C., Nebert, D. W., & Vasiliou, V. (2010). Update of human and mouse forkhead box (fox) gene families. *Human genomics*, 4, 1–8.
- Jombart, T., & Dray, S. (2010). Adephylo: Exploratory analyses for the phylogenetic comparative method. *Bioinformatics*, 26(15), 1–21.

- Kalyaanamoorthy, S., Minh, B. Q., Wong, T. K., Von Haeseler, A., & Jermiin, L. S. (2017). Modelfinder: Fast model selection for accurate phylogenetic estimates. *Nature methods*, 14(6), 587–589.
- Kenny, N. J., McCarthy, S. A., Dudchenko, O., James, K., Betteridge, E., Corton, C., Dolucan, J., Mead, D., Oliver, K., Omer, A. D., et al. (2020). The gene-rich genome of the scallop *Pecten maximus*. *Gigascience*, 9(5), giaa037.
- Khaitovich, P., Hellmann, I., Enard, W., Nowick, K., Leinweber, M., Franz, H., Weiss, G., Lachmann, M., & Paabo, S. (2005). Parallel patterns of evolution in the genomes and transcriptomes of humans and chimpanzees. *Science*, 309(5742), 1850–1854.
- Kim, B. Y., Wang, J. R., Miller, D. E., Barmina, O., Delaney, E., Thompson, A., Comeault, A. A., Peede, D., D'Agostino, E. R., Pelaez, J., et al. (2021). Highly contiguous assemblies of 101 drosophilid genomes. *Elife*, 10, e66405.
- Klein, S. L., Neilson, K. M., Orban, J., Yaklichkin, S., Hoffbauer, J., Mood, K., Daar, I. O., & Moody, S. A. (2013). Conserved structural domains in foxd4l1, a neural forkhead box transcription factor, are required to repress or activate target genes. *PLoS one*, 8(4), e61845.
- Knudsen, B., Kohn, A. B., Nahir, B., McFadden, C. S., & Moroz, L. L. (2006). Complete dna sequence of the mitochondrial genome of the sea-slug, *Aplysia californica*: Conservation of the gene order in euthyneura. *Molecular Phylogenetics and Evolution*, 38(2), 459–469.
- Kopp, A. (2012). Dmrt genes in the development and evolution of sexual dimorphism. *Trends in Genetics*, 28(4), 175–184.
- Kousathanas, A., Halligan, D. L., & Keightley, P. D. (2014). Faster-x adaptive protein evolution in house mice. *Genetics*, 196(4), 1131–1143.
- Kuehn, E., Clausen, D. S., Null, R. W., Metzger, B. M., Willis, A. D., & Özpolat, B. D. (2022). Segment number threshold determines juvenile onset of germline cluster expansion in *Platynereis dumerilii*. *Journal of Experimental Zoology Part B: Molecular and Developmental Evolution*, 338(4), 225–240.
- Lan, Y., Sun, J., Chen, C., Sun, Y., Zhou, Y., Yang, Y., Zhang, W., Li, R., Zhou, K., Wong, W. C., et al. (2021). Hologenome analysis reveals dual symbiosis in the deep-sea hydrothermal vent snail *Gigantopelta aegis*. *Nature communications*, 12(1), 1165.

- Lan, Y., Sun, J., Zhang, W., Xu, T., Zhang, Y., Chen, C., Feng, D., Wang, H., Tao, J., Qiu, J.-W., et al. (2019). Host–symbiont interactions in deep-sea chemosymbiotic vesicomyid clams: Insights from transcriptome sequencing. *Frontiers in Marine Science*, 6, 680.
- Larroux, C., Luke, G. N., Koopman, P., Rokhsar, D. S., Shimeld, S. M., & Degnan, B. M. (2008). Genesis and expansion of metazoan transcription factor gene classes. *Molecular biology and evolution*, 25(5), 980–996.
- Lemoine, F., & Gascuel, O. (2021). Gotree/goalign: Toolkit and go api to facilitate the development of phylogenetic workflows. *NAR Genomics and Bioinformatics*, 3(3), lqab075.
- Li, A., Dai, H., Guo, X., Zhang, Z., Zhang, K., Wang, C., Wang, X., Wang, W., Chen, H., Li, X., et al. (2021). Genome of the estuarine oyster provides insights into climate impact and adaptive plasticity. *Communications Biology*, 4(1), 1287.
- Li, F., Bian, L., Ge, J., Han, F., Liu, Z., Li, X., Liu, Y., Lin, Z., Shi, H., Liu, C., et al. (2020). Chromosome-level genome assembly of the east asian common octopus (*Octopus sinensis*) using pacbio sequencing and hi-c technology. *Molecular ecology resources*, 20(6), 1572–1582.
- Li, H. (2023). Protein-to-genome alignment with miniprot. *Bioinformatics*, 39(1), btad014.
- Li, J., Zhou, Y., Zhou, Z., Lin, C., Wei, J., Qin, Y., Xiang, Z., Ma, H., Zhang, Y., Zhang, Y., et al. (2020). Comparative transcriptome analysis of three gonadal development stages reveals potential genes involved in gametogenesis of the fluted giant clam (*Tridacna squamosa*). *BMC genomics*, 21, 1–16.
- Li, R., Zhang, L., Li, W., Zhang, Y., Li, Y., Zhang, M., Zhao, L., Hu, X., Wang, S., & Bao, Z. (2018). *FOXL2* and *DMRT1L* are yin and yang genes for determining timing of sex differentiation in the bivalve mollusk *Patinopecten yessoensis*. *Frontiers in Physiology*, 9, 1166.
- Li, X.-Y., Mei, J., Ge, C.-T., Liu, X.-L., & Gui, J.-F. (2022). Sex determination mechanisms and sex control approaches in aquaculture animals. *Science China Life Sciences*, 65(6), 1091–1122.
- Liang, S., Liu, D., Li, X., Wei, M., Yu, X., Li, Q., Ma, H., Zhang, Z., & Qin, Z. (2019). *SOX2* participates in spermatogenesis of zhikong scallop *Chlamys farreri*. *Scientific Reports*, 9(1), 76.

- Liao, Y., Zhang, X., Chakraborty, M., & Emerson, J. (2021). Topologically associating domains and their role in the evolution of genome structure and function in *Drosophila*. *Genome research*, 31(3), 397–410.
- Liu, C., Zhang, Y., Ren, Y., Wang, H., Li, S., Jiang, F., Yin, L., Qiao, X., Zhang, G., Qian, W., et al. (2018). The genome of the golden apple snail *Pomacea canaliculata* provides insight into stress tolerance and invasive adaptation. *Gigascience*, 7(9), giy101.
- Liu, T., Zhang, Y., Nie, H., Sun, J., & Yan, X. (2024). Characterization and expression patterns of the fox gene family under heat and cold stress in manila clam *Ruditapes philippinarum* based on genome-wide identification. *Comparative Biochemistry and Physiology Part D: Genomics and Proteomics*, 52, 101313.
- Liu, X., Li, C., Chen, M., Liu, B., Yan, X., Ning, J., Ma, B., Liu, G., Zhong, Z., Jia, Y., et al. (2020). Draft genomes of two atlantic bay scallop subspecies *Argopecten irradians irradians* and *A. i. concentricus*. *Scientific Data*, 7(1), 99.
- Love, M. I., Huber, W., & Anders, S. (2014). Moderated estimation of fold change and dispersion for rna-seq data with deseq2. *Genome biology*, 15(12), 1–21.
- Lozano-Fernandez, J. (2022). A practical guide to design and assess a phylogenomic study. *Genome Biology and Evolution*, 14(9), evac129.
- Mahajan, S., Wei, K. H.-C., Nalley, M. J., Gibilisco, L., & Bachtrog, D. (2018). De novo assembly of a young *Drosophila* y chromosome using single-molecule sequencing and chromatin conformation capture. *PLoS biology*, 16(7), e2006348.
- Mai, D., Nalley, M. J., & Bachtrog, D. (2020). Patterns of genomic differentiation in the *Drosophila nasuta* species complex. *Molecular biology and evolution*, 37(1), 208–220.
- Mank, J. E., Axelsson, E., & Ellegren, H. (2007). Fast-x on the z: Rapid evolution of sex-linked genes in birds. *Genome Research*, 17(5), 618–624.
- Manni, M., Berkeley, M. R., Seppey, M., Simão, F. A., & Zdobnov, E. M. (2021). Busco update: Novel and streamlined workflows along with broader and deeper phylogenetic coverage for scoring of eukaryotic, prokaryotic, and viral genomes. *Molecular biology and evolution*, 38(10), 4647–4654.
- Marshall Graves, J. A., & Peichel, C. L. (2010). Are homologies in vertebrate sex determination due to shared ancestry or to limited options? *Genome biology*, 11, 1–12.
- Matson, C. K., & Zarkower, D. (2012). Sex and the singular dm domain: Insights into sexual regulation, evolution and plasticity. *Nature Reviews Genetics*, 13(3), 163–174.

- Mawaribuchi, S., Ito, Y., & Ito, M. (2019). Independent evolution for sex determination and differentiation in the *DMRT* family in animals. *Biology Open*, 8(8), bio041962.
- Mawaribuchi, S., Yoshimoto, S., Ohashi, S., Takamatsu, N., & Ito, M. (2012). Molecular evolution of vertebrate sex-determining genes. *Chromosome Research*, 20, 139–151.
- Mazet, F., Yu, J.-K., Liberles, D. A., Holland, L. Z., & Shimeld, S. M. (2003). Phylogenetic relationships of the fox (forkhead) gene family in the bilateria. *Gene*, 316, 79–89.
- McCartney, M. A., Auch, B., Kono, T., Mallez, S., Zhang, Y., Obille, A., Becker, A., Abrahante, J. E., Garbe, J., Badalamenti, J. P., et al. (2022). The genome of the zebra mussel, *Dreissena polymorpha*: A resource for comparative genomics, invasion genetics, and biocontrol. *G3*, 12(2), jkab423.
- Mead, D., Fingland, K., Cripps, R., Miguez, R. P., Smith, M., Corton, C., Oliver, K., Skelton, J., Betteridge, E., Doulcan, J., et al. (2020). The genome sequence of the eastern grey squirrel, *Sciurus carolinensis* gmelin, 1788. *Wellcome Open Research*, 5.
- Meisel, R. P. (2011). Towards a more nuanced understanding of the relationship between sex-biased gene expression and rates of protein-coding sequence evolution. *Molecular Biology and Evolution*, 28(6), 1893–1900.
- Meisel, R. P., & Connallon, T. (2013). The faster-x effect: Integrating theory and data. *Trends in genetics*, 29(9), 537–544.
- Meng, J., Yang, M., Xu, F., Li, X., & Li, L. (2018). Transcriptome assembly of modiolus modiolus and comparative analysis with *Bathymodiolus platifrons*. *Acta Oceanologica Sinica*, 37, 38–45.
- Miglioli, A., Tredez, M., Boosten, M., Sant, C., Carvalho, J. E., Dru, P., Canesi, L., Schubert, M., & Dumollard, R. (2024). The mediterranean mussel *Mytilus galloprovincialis*: A novel model for developmental studies in mollusks. *Development*, 151(4), dev202256.
- Milani, L., & Ghiselli, F. (2020). Faraway, so close. the comparative method and the potential of non-model animals in mitochondrial research. *Philosophical Transactions of the Royal Society B*, 375(1790), 20190186.
- Milani, L., Ghiselli, F., Maurizii, M. G., Nuzhdin, S. V., & Passamonti, M. (2014). Paternally transmitted mitochondria express a new gene of potential viral origin. *Genome biology and evolution*, 6(2), 391–405.

- Milani, L., Ghiselli, F., Maurizii, M. G., & Passamonti, M. (2011). Doubly uniparental inheritance of mitochondria as a model system for studying germ line formation. *PLoS One*, 6(11), e28194.
- Milani, L., Ghiselli, F., Nuzhdin, S. V., & Passamonti, M. (2013). Nuclear genes with sex bias in *Ruditapes philippinarum* (bivalvia, veneridae): Mitochondrial inheritance and sex determination in dui species. *Journal of Experimental Zoology Part B: Molecular and Developmental Evolution*, 320(7), 442–454.
- Minh, B. Q., Schmidt, H. A., Chernomor, O., Schrempf, D., Woodhams, M. D., Von Haeseler, A., & Lanfear, R. (2020). Iq-tree 2: New models and efficient methods for phylogenetic inference in the genomic era. *Molecular biology and evolution*, 37(5), 1530–1534.
- Moreno, J. A., Dudchenko, O., Feigin, C. Y., Mereby, S. A., Chen, Z., Ramos, R., Almet, A. A., Sen, H., Brack, B. J., Johnson, M. R., et al. (2024). *Emx2* underlies the development and evolution of marsupial gliding membranes. *Nature*, 1–9.
- Mullon, C., Pomiankowski, A., & Reuter, M. (2012). Molecular evolution of *Drosophila Sex-lethal* and related sex determining genes. *BMC evolutionary biology*, 12, 1–11.
- Murat, F., Mbengue, N., Winge, S. B., Trefzer, T., Leushkin, E., Sepp, M., Cardoso-Moreira, M., Schmidt, J., Schneider, C., Mößinger, K., et al. (2023). The molecular evolution of spermatogenesis across mammals. *Nature*, 613(7943), 308–316.
- Murata, Y., Nikaido, M., Sasaki, T., Cao, Y., Fukumoto, Y., Hasegawa, M., & Okada, N. (2003). Afrotherian phylogeny as inferred from complete mitochondrial genomes. *Molecular phylogenetics and evolution*, 28(2), 253–260.
- Naimi, A., Martinez, A.-S., Specq, M.-L., Diss, B., Mathieu, M., & Sourdaine, P. (2009). Molecular cloning and gene expression of *Cg-Foxl2* during the development and the adult gametogenetic cycle in the oyster *Crassostrea gigas*. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 154(1), 134–142.
- Nakagawa, S., Gisselbrecht, S. S., Rogers, J. M., Hartl, D. L., & Bulyk, M. L. (2013). Dna-binding specificity changes in the evolution of forkhead transcription factors. *Proceedings of the National Academy of Sciences*, 110(30), 12349–12354.
- Natsidis, P., Kapli, P., Schiffer, P. H., & Telford, M. J. (2021). Systematic errors in orthology inference and their effects on evolutionary analyses. *Iscience*, 24(2).

- Nicolini, F., Ghiselli, F., Luchetti, A., & Milani, L. (2023). Bivalves as emerging model systems to study the mechanisms and evolution of sex determination: A genomic point of view. *Genome Biology and Evolution*, 15(10), evad181.
- Nicolini, F., Martelossi, J., Forni, G., Savojardo, C., Mantovani, B., & Luchetti, A. (2023). Comparative genomics of hox and parahox genes among major lineages of brachiopoda with emphasis on tadpole shrimps. *Frontiers in Ecology and Evolution*, 11, 1046960.
- Nie, H., Dong, S., Li, D., Zheng, M., Jiang, L., Li, X., & Yan, X. (2018). Rna-seq analysis of differentially expressed genes in the grand jackknife clam *Solen grandis* under aerial exposure. *Comparative Biochemistry and Physiology Part D: Genomics and Proteomics*, 28, 54–62.
- Obata, M., & Komaru, A. (2005). Specific location of sperm mitochondria in mussel *Mytilus galloprovincialis* zygotes stained by mitotracker. *Development, growth & differentiation*, 47(4), 255–263.
- of Sex Consortium, T., et al. (2014). Tree of sex: A database of sexual systems. *Scientific Data*, 1.
- O'Neil, M. T., & Belote, J. M. (1992). Interspecific comparison of the *transformer* gene of *Drosophila* reveals an unusually high degree of evolutionary divergence. *Genetics*, 131(1), 113–128.
- Orkin, J. D., Montague, M. J., Tejada-Martinez, D., De Manuel, M., Del Campo, J., Cheves Hernandez, S., Di Fiore, A., Fontserè, C., Hodgson, J. A., Janiak, M. C., et al. (2021). The genomics of ecological flexibility, large brains, and long lives in capuchin monkeys revealed with fecalfacs. *Proceedings of the National Academy of Sciences*, 118(7), e2010632118.
- Paggeot, L. X., DeBiase, M. B., Escalona, M., Fairbairn, C., Marimuthu, M. P., Nguyen, O., Sahasrabudhe, R., & Dawson, M. N. (2022). Reference genome for the californian ribbed mussel, *Mytilus californianus*, an ecosystem engineer. *Journal of Heredity*, 113(6), 681–688.
- Pamilo, P., & O'Neill, R. (1997). Evolution of the *Sry* genes. *Molecular biology and evolution*, 14(1), 49–55.
- Panara, V., Budd, G. E., & Janssen, R. (2019). Phylogenetic analysis and embryonic expression of panarthropod dmrt genes. *Frontiers in zoology*, 16, 1–18.

- Papa, F., Windbichler, N., Waterhouse, R. M., Cagnetti, A., D'Amato, R., Persampieri, T., Lawniczak, M. K., Nolan, T., & Papathanos, P. A. (2017). Rapid evolution of female-biased genes among four species of *Anopheles* malaria mosquitoes. *Genome research*, 27(9), 1536–1548.
- Parsch, J., & Ellegren, H. (2013). The evolutionary causes and consequences of sex-biased gene expression. *Nature Reviews Genetics*, 14(2), 83–87.
- Patnaik, B. B., Wang, T. H., Kang, S. W., Hwang, H.-J., Park, S. Y., Park, E. B., Chung, J. M., Song, D. K., Kim, C., Kim, S., et al. (2016). Sequencing, de novo assembly, and annotation of the transcriptome of the endangered freshwater pearl bivalve, *Cristaria plicata*, provides novel insights into functional genes and marker discovery. *PLoS One*, 11(2), e0148622.
- Penaloza, C., Gutierrez, A. P., Eöry, L., Wang, S., Guo, X., Archibald, A. L., Bean, T. P., & Houston, R. D. (2021). A chromosome-level genome assembly for the pacific oyster *Crassostrea gigas*. *GigaScience*, 10(3), giab020.
- Perez-Garcia, C., Moran, P., & Pasantes, J. J. (2011). Cytogenetic characterization of the invasive mussel species *Xenostrobus securis* lmk. (bivalvia: Mytilidae). *Genome*, 54(09), 771–778.
- Pertea, M., Kim, D., Pertea, G. M., Leek, J. T., & Salzberg, S. L. (2016). Transcript-level expression analysis of rna-seq experiments with hisat, stringtie and ballgown. *Nature protocols*, 11(9), 1650–1667.
- Pertea, M., Pertea, G. M., Antonescu, C. M., Chang, T.-C., Mendell, J. T., & Salzberg, S. L. (2015). Stringtie enables improved reconstruction of a transcriptome from rna-seq reads. *Nature biotechnology*, 33(3), 290–295.
- Phochanukul, N., & Russell, S. (2010). No backbone but lots of sox: Invertebrate sox genes. *The international journal of biochemistry & cell biology*, 42(3), 453–464.
- Piccinini, G., Iannello, M., Puccio, G., Plazzi, F., Havird, J. C., & Ghiselli, F. (2021). Mitonuclear coevolution, but not nuclear compensation, drives evolution of oxphos complexes in bivalves. *Molecular Biology and Evolution*, 38(6), 2597–2614.
- Powell, D., Subramanian, S., Suwansa-Ard, S., Zhao, M., O'Connor, W., Raftos, D., & Elizur, A. (2018). The genome of the oyster *Saccostrea* offers insight into the environmental resilience of bivalves. *DNA Research*, 25(6), 655–665.

- Purandare, S. R., Bickel, R. D., Jaquière, J., Rispe, C., & Brisson, J. A. (2014). Accelerated evolution of morph-biased genes in pea aphids. *Molecular biology and evolution*, 31(8), 2073–2083.
- Putnam, H. M., Trigg, S. A., White, S. J., Spencer, L. H., Vadopalas, B., Natarajan, A., Hetzel, J., Jaeger, E., Soohoo, J., Gallardo-Escárate, C., et al. (2022). Dynamic dna methylation contributes to carryover effects and beneficial acclimatization in geoduck clams. *bioRxiv*, 2022–06.
- Qi, H., Cong, R., Wang, Y., Li, L., & Zhang, G. (2023). Construction and analysis of the chromosome-level haplotype-resolved genomes of two *Crassostrea* oyster congeners: *Crassostrea angulata* and *Crassostrea gigas*. *GigaScience*, 12, giad077.
- Ran, Z., Li, Z., Yan, X., Liao, K., Kong, F., Zhang, L., Cao, J., Zhou, C., Zhu, P., He, S., et al. (2019). Chromosome-level genome assembly of the razor clam *Sinonovacula constricta* (lamarck, 1818). *Molecular ecology resources*, 19(6), 1647–1658.
- Ranz, J. M., Go, A. C., González, P. M., Clifton, B. D., Gomes, S., Jaberyzadeh, A., Woodbury, A., Chan, C., Gandasetiawan, K. A., Jayasekera, S., et al. (2023). Gene expression differentiation in the reproductive tissues of *Drosophila willistoni* subspecies and their hybrids. *Molecular Ecology*, 32(13), 3605–3623.
- Renschler, G., Richard, G., Valsecchi, C. I. K., Toscano, S., Arrigoni, L., Ramírez, F., & Akhtar, A. (2019). Hi-c guided assemblies reveal conserved regulatory topologies on x and autosomes despite extensive genome shuffling. *Genes & development*, 33(21-22), 1591–1612.
- Richardson, V., Engel, N., & Kulathinal, R. J. (2023). Comparative developmental genomics of sex-biased gene expression in early embryogenesis across mammals. *Biology of sex Differences*, 14(1), 30.
- Rogers, R. L., Grizzard, S. L., Titus-McQuillan, J. E., Bockrath, K., Patel, S., Wares, J. P., Garner, J. T., & Moore, C. C. (2021). Gene family amplification facilitates adaptation in freshwater unionid bivalve *Megalonaia nervosa*. *Molecular Ecology*, 30(5), 1155–1173.
- Rozewicki, J., Li, S., Amada, K. M., Standley, D. M., & Katoh, K. (2019). Mafft-dash: Integrated protein sequence and structural alignment. *Nucleic acids research*, 47(W1), W5–W10.
- Russo, C. A., Mello, B., Frazão, A., & Voloch, C. M. (2013). Phylogenetic analysis and a time tree for a large drosophilid data set (diptera: Drosophilidae). *Zoological Journal of the Linnean Society*, 169(4), 765–775.

- Salz, H., & Erickson, J. W. (2010). Sex determination in *Drosophila*: The view from the top. *Fly*, 4(1), 60–70.
- Sanchez-Flores, A., Peñaloza, F., Carpintero-Ponce, J., Nazario-Yepiz, N., Abreu-Goodger, C., Machado, C. A., & Markow, T. A. (2016). Genome evolution in three species of cactophilic *Drosophila*. *G3: Genes, Genomes, Genetics*, 6(10), 3097–3105.
- Sarkar, A., & Hochedlinger, K. (2013). The sox family of transcription factors: Versatile regulators of stem and progenitor cell fate. *Cell stem cell*, 12(1), 15–30.
- Schliep, K. P. (2011). Phangorn: Phylogenetic analysis in r. *Bioinformatics*, 27(4), 592–593.
- Schomburg, C., Janssen, R., & Prpic, N.-M. (2022). Phylogenetic analysis of forkhead transcription factors in the panarthropoda. *Development genes and evolution*, 232(1), 39–48.
- Seudre, O., Martín-Zamora, F. M., Rapisarda, V., Luqman, I., Carrillo-Baltodano, A. M., & Martín-Durán, J. M. (2022). The *Fox* gene repertoire in the annelid *Owenia fusiformis* reveals multiple expansions of the *foxQ2* class in spiralia. *Genome Biology and Evolution*, 14(10), evac139.
- Shi, J., Hong, Y., Sheng, J., Peng, K., & Wang, J. (2015). De novo transcriptome sequencing to identify the sex-determination genes in *Hyriopsis schlegelii*. *Bioscience, Biotechnology, and Biochemistry*, 79(8), 1257–1265.
- Shi, Y., Liu, W., & He, M. (2018). Proteome and transcriptome analysis of ovary, intersex gonads, and testis reveals potential key sex reversal/differentiation genes and mechanism in scallop *Chlamys nobilis*. *Marine Biotechnology*, 20, 220–245.
- Shimeld, S. M., Boyle, M. J., Brunet, T., Luke, G. N., & Seaver, E. C. (2010). Clustered fox genes in lophotrochozoans and the evolution of the bilaterian fox gene cluster. *Developmental biology*, 340(2), 234–248.
- Shimizu, K., Negishi, L., Ito, T., Touma, S., Matsumoto, T., Awaji, M., Kurumizaka, H., Yoshitake, K., Kinoshita, S., Asakawa, S., et al. (2022). Evolution of nacre-and prisms-related shell matrix proteins in the pen shell, *Atrina pectinata*. *Comparative Biochemistry and Physiology Part D: Genomics and Proteomics*, 44, 101025.
- Sievers, F., Wilm, A., Dineen, D., Gibson, T. J., Karplus, K., Li, W., Lopez, R., McWilliam, H., Remmert, M., Söding, J., et al. (2011). Fast, scalable generation of high-quality protein multiple sequence alignments using clustal omega. *Molecular systems biology*, 7(1), 539.

- Smith, C. H. (2021). A high-quality reference genome for a parasitic bivalve with doubly uniparental inheritance (bivalvia: Unionida). *Genome Biology and Evolution*, 13(3), evab029.
- Song, H., Guo, X., Sun, L., Wang, Q., Han, F., Wang, H., Wray, G. A., Davidson, P., Wang, Q., Hu, Z., et al. (2021). The hard clam genome reveals massive expansion and diversification of inhibitors of apoptosis in bivalvia. *BMC biology*, 19, 1–20.
- Song, H., Wang, Y., Shao, H., Li, Z., Hu, P., Yap-Chiongco, M. K., Shi, P., Zhang, T., Li, C., Wang, Y., et al. (2023). Scaphopoda is the sister taxon to bivalvia: Evidence of ancient incomplete lineage sorting. *Proceedings of the National Academy of Sciences*, 120(40), e2302361120.
- Stammnitz, M. R., Gori, K., Kwon, Y. M., Harry, E., Martin, F. J., Billis, K., Cheng, Y., Baez-Ortega, A., Chow, W., Comte, S., et al. (2023). The evolution of two transmissible cancers in tasmanian devils. *Science*, 380(6642), 283–293.
- Stothard, P., & Pilgrim, D. (2003). Sex-determination gene and pathway evolution in nematodes. *Bioessays*, 25(3), 221–231.
- Sun, D., Yu, H., & Li, Q. (2022). Examination of the roles of *Foxl2* and *Dmrt1* in sex differentiation and gonadal development of oysters by using rna interference. *Aquaculture*, 548, 737732.
- Sun, J., Zhang, Y., Xu, T., Zhang, Y., Mu, H., Zhang, Y., Lan, Y., Fields, C. J., Hui, J. H. L., Zhang, W., et al. (2017). Adaptation to deep-sea chemosynthetic environments as revealed by mussel genomes. *Nature ecology & evolution*, 1(5), 0121.
- Swanson, W. J., & Vacquier, V. D. (2002). The rapid evolution of reproductive proteins. *Nature reviews genetics*, 3(2), 137–144.
- Teaniniuraitemoana, V., Huvet, A., Levy, P., Klopp, C., Lhuillier, E., Gaertner-Mazouni, N., Gueguen, Y., & Le Moullac, G. (2014). Gonad transcriptome analysis of pearl oyster *Pinctada margaritifera*: Identification of potential sex differentiation and sex determining genes. *BMC genomics*, 15, 1–20.
- Teng, W., Fu, H., Li, Z., Zhang, Q., Xu, C., Yu, H., Kong, L., Liu, S., & Li, Q. (2023). Parallel evolution in *Crassostrea* oysters along the latitudinal gradient is associated with variation in multiple genes involved in adipogenesis. *Molecular Ecology*, 32(19), 5276–5287.
- Tu, Q., Brown, C. T., Davidson, E. H., & Oliveri, P. (2006). Sea urchin forkhead gene family: Phylogeny and embryonic expression. *Developmental biology*, 300(1), 49–62.

- Tvedte, E. S., Gasser, M., Sparklin, B. C., Michalski, J., Hjelmen, C. E., Johnston, J. S., Zhao, X., Bromley, R., Tallon, L. J., Sadzewicz, L., et al. (2021). Comparison of long-read sequencing technologies in interrogating bacteria and fly genomes. *G3*, 11(6), jkab083.
- Uller, T., & Helanterä, H. (2011). From the origin of sex-determining factors to the evolution of sex-determining systems. *The Quarterly review of biology*, 86(3), 163–180.
- Verhulst, E. C., van de Zande, L., & Beukeboom, L. W. (2010). Insect sex determination: It all evolves around *transformer*. *Current opinion in genetics & development*, 20(4), 376–383.
- Vicoso, B., & Charlesworth, B. (2006). Evolution on the x chromosome: Unusual patterns and processes. *Nature Reviews Genetics*, 7(8), 645–653.
- Vilstrup, J. T., Seguin-Orlando, A., Stiller, M., Ginolhac, A., Raghavan, M., Nielsen, S. C., Weinstock, J., Froese, D., Vasiliev, S. K., Ovodov, N. D., et al. (2013). Mitochondrial phylogenomics of modern and ancient equids. *PloS one*, 8(2), e55950.
- Vinkler, M., Fiddaman, S. R., Těšický, M., O'Connor, E. A., Savage, A. E., Lenz, T. L., Smith, A. L., Kaufman, J., Bolnick, D. I., Davies, C. S., et al. (2023). Understanding the evolution of immune genes in jawed vertebrates. *Journal of evolutionary biology*, 36(6), 847–873.
- Vizueta Moraga, J., Sánchez-Gracia, A., & Rozas Liras, J. A. (2020). Bitacora: A comprehensive tool for the identification and annotation of gene families in genome assemblies. *Molecular Ecology Resources*, 2020, vol. 20, num. 5, p. 1445-1452.
- Wang, C., Wallerman, O., Arendt, M.-L., Sundström, E., Karlsson, Å., Nordin, J., Mäkeläinen, S., Pielberg, G. R., Hanson, J., Ohlsson, Å., et al. (2021). A novel canine reference genome resolves genomic architecture and uncovers transcript complexity. *Communications biology*, 4(1), 185.
- Wang, G., Dong, S., Guo, P., Cui, X., Duan, S., & Li, J. (2020). Identification of *Foxl2* in freshwater mussel *Hyriopsis cumingii* and its involvement in sex differentiation. *Gene*, 754, 144853.
- Wang, J., & Nie, H. (2024). Genome-wide identification and expression analysis of sox gene family in the manila clam (*Ruditapes philippinarum*). *Comparative Biochemistry and Physiology Part D: Genomics and Proteomics*, 50, 101244.
- Wang, M., Xia, J., Jawad, M., Wei, W., Gui, L., Liang, X., Yang, J.-L., & Li, M. (2022). Transcriptome sequencing analysis of sex-related genes and miRNAs in the gonads of *Mytilus coruscus*. *Frontiers in Marine Science*, 9, 1013857.

- Wang, Q., Cao, T., & Wang, C. (2023). Genome-wide identification and expression analysis of dmrt genes in bivalves. *BMC genomics*, 24(1), 457.
- Wang, S., Zhang, J., Jiao, W., Li, J., Xun, X., Sun, Y., Guo, X., Huan, P., Dong, B., Zhang, L., et al. (2017). Scallop genome provides insights into evolution of bilaterian karyotype and development. *Nature ecology & evolution*, 1(5), 0120.
- Wang, X., Werren, J. H., & Clark, A. G. (2015). Genetic and epigenetic architecture of sex-biased expression in the jewel wasps *Nasonia vitripennis* and *giraulti*. *Proceedings of the National Academy of Sciences*, 112(27), E3545–E3554.
- Wang, X., Liu, Z., & Wu, W. (2017). Transcriptome analysis of the freshwater pearl mussel (*Cristaria plicata*) mantle unravels genes involved in the formation of shell and pearl. *Molecular genetics and genomics*, 292, 343–352.
- Wei, M., Ge, H., Shao, C., Yan, X., Nie, H., Duan, H., Liao, X., Zhang, M., Chen, Y., Zhang, D., et al. (2020). Chromosome-level clam genome helps elucidate the molecular basis of adaptation to a buried lifestyle. *IScience*, 23(6).
- Wexler, J. R., Plachetzki, D. C., & Kopp, A. (2014). Pan-metazoan phylogeny of the dmrt gene family: A framework for functional studies. *Development Genes and Evolution*, 224, 175–181.
- Whitfield, L. S., Lovell-Badge, R., & Goodfellow, P. N. (1993). Rapid sequence evolution of the mammalian sex-determining gene sry. *Nature*, 364 (6439), 713–715.
- Wilkins, A. S. (1995). Moving up the hierarchy: A hypothesis on the evolution of a genetic sex determination pathway. *Bioessays*, 17(1), 71–77.
- Willson, J., Roddur, M. S., Liu, B., Zaharias, P., & Warnow, T. (2022). Disco: Species tree inference using multicopy gene family tree decomposition. *Systematic biology*, 71(3), 610–629.
- Wu, S., Zhang, Y., Li, Y., Wei, H., Guo, Z., Wang, S., Zhang, L., & Bao, Z. (2020). Identification and expression profiles of fox transcription factors in the yesso scallop (*patinopecten yessoensis*). *Gene*, 733, 144387.
- Xiong, Y., Brandley, M. C., Xu, S., Zhou, K., & Yang, G. (2009). Seven new dolphin mitochondrial genomes and a time-calibrated phylogeny of whales. *BMC Evolutionary Biology*, 9, 1–13.

- Xu, R., Iannello, M., Havird, J. C., Milani, L., & Ghiselli, F. (2022). Lack of transcriptional coordination between mitochondrial and nuclear oxidative phosphorylation genes in the presence of two divergent mitochondrial genomes. *Zoological Research*, 43(1), 111.
- Xu, R., Martelossi, J., Smits, M., Iannello, M., Peruzza, L., Babbucci, M., Milan, M., Dunham, J. P., Breton, S., Milani, L., et al. (2022). Multi-tissue rna-seq analysis and long-read-based genome assembly reveal complex sex-specific gene regulation and molecular evolution in the manila clam. *Genome Biology and Evolution*, 14(12), evac171.
- Yang, J.-L., Feng, D.-D., Liu, J., Xu, J.-K., Chen, K., Li, Y.-F., Zhu, Y.-T., Liang, X., & Lu, Y. (2021). Chromosome-level genome assembly of the hard-shelled mussel *Mytilus coruscus*, a widely distributed species from the temperate areas of east asia. *Gigascience*, 10(4), giab024.
- Yang, M., Xu, F., Liu, J., Que, H., Li, L., & Zhang, G. (2014). Phylogeny of forkhead genes in three spiralians and their expression in pacific oyster *Crassostrea gigas*. *Chinese journal of oceanology and limnology*, 32(6), 1207–1223.
- Yu, J., Zhang, L., Li, Y., Li, R., Zhang, M., Li, W., Xie, X., Wang, S., Hu, X., & Bao, Z. (2017). Genome-wide identification and expression profiling of the sox gene family in a bivalve mollusc *Patinopecten yessoensis*. *Gene*, 627, 530–537.
- Yu, J.-K., Mazet, F., Chen, Y.-T., Huang, S.-W., Jung, K.-C., & Shimeld, S. M. (2008). The fox genes of *Branchiostoma floridae*. *Development genes and evolution*, 218, 629–638.
- Yue, C., Li, Q., & Yu, H. (2021). Variance in expression and localization of sex-related genes *CgDsx*, *CgBHMG1* and *CgFoxl2* during diploid and triploid pacific oyster *Crassostrea gigas* gonad differentiation. *Gene*, 790, 145692.
- Yuen, B., Polzin, J., & Petersen, J. M. (2019). Organ transcriptomes of the lucinid clam *Loripes orbiculatus* (poli, 1791) provide insights into their specialised roles in the biology of a chemosymbiotic bivalve. *BMC genomics*, 20, 1–14.
- Zeng, Y., Zheng, H., He, C., Zhang, C., Zhang, H., & Zheng, H. (2024). Genome-wide identification and expression analysis of dmrt gene family and their role in gonad development of pacific oyster (*crassostrea gigas*). *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 269, 110904.
- Zhang, N., Xu, F., & Guo, X. (2014). Genomic analysis of the pacific oyster (*Crassostrea gigas*) reveals possible conservation of vertebrate sex determination in a mollusc. *G3: Genes, Genomes, Genetics*, 4(11), 2207–2217.

- Zhou, Y., Shearwin-Whyatt, L., Li, J., Song, Z., Hayakawa, T., Stevens, D., Fenelon, J. C., Peel, E., Cheng, Y., Pajpach, F., et al. (2021). Platypus and echidna genomes reveal mammalian biology and evolution. *Nature*, 592(7856), 756–762.
- Zhu, C., Zhang, L., Ding, H., & Pan, Z. (2019). Transcriptome-wide identification and characterization of the *Sox* gene family and microsatellites for *Corbicula fluminea*. *PeerJ*, 7, e7770.
- Zouros, E. (2013). Biparental inheritance through uniparental transmission: The doubly uniparental inheritance (dui) of mitochondrial dna. *Evolutionary Biology*, 40, 1–31.

Appendix

The appendix includes the titles and abstracts of the papers published during my PhD that are not part of this thesis.

Taxonomic revision of the Australian stick insect genus *Candovia* (Phasmida: Necrosciinae): insight from molecular systematics and species-delimitation approaches.

Giobbe Forni^{1,2}, Alex Cussigh^{1,2}, Paul D. Brock³, Braxton R. Jones⁴, Filippo Nicolini¹, Jacopo Martelossi¹, Andrea Luchetti¹, Barbara Mantovani¹

¹*Department of Biological, Geological and Environmental Sciences, University of Bologna, Bologna, Italy.*

²*Department of Agricultural and Environmental Sciences, University of Milan, Milano, Italy.*

³*The Natural History Museum, Cromwell Road, London, UK.*

⁴*School of Life and Environmental Sciences, The University of Sydney, Sydney NSW 2006, Australia.*

Published in: 2023, *Zoological Journal of the Linnean Society*, 197:189–210. 10.1093/zoolinnean/zlac074

Abstract. The Phasmida genus *Candovia* comprises nine traditionally recognized species, all endemic to Australia. In this study, *Candovia* diversity is explored through molecular species-delimitation analyses using the *COI_{Fol}* gene fragment and phylogenetic inferences leveraging seven additional mitochondrial and nuclear loci. Molecular results were integrated with morphological observations, leading us to confirm the already described species and to the delimitation of several new taxa and of the new genus *Paracandovia*. New *Candovia* species from various parts of Queensland and New South Wales are described and illustrated (*C. alata* sp. nov., *C. byfieldensis* sp. nov., *C. dagleishae* sp. nov., *C. eungellensis* sp. nov., *C. karasi* sp. nov., *C. koensi* sp. nov. and *C. wollumbinensis* sp. nov.). New combinations are proposed and species removed from synonymy with the erection of the new genus *Paracandovia* (*P. cercata* stat. rev., comb. nov., *P. longipes* stat. rev., comb. nov., *P. pallida* comb. nov., *P. peridromes* comb. nov., *P. tenera* stat. rev., comb. nov.). Phylogenetic analyses suggest that the egg capitulum may have independently evolved multiple times throughout the evolutionary history of these insects. Furthermore, two newly described species represent the first taxa with fully developed wings in this previously considered apterous clade.

Comparative genomics of *Hox* and *ParaHox* genes among major lineages of Branchiopoda with emphasis on tadpole shrimps.

Filippo Nicolini^{1,2}, Jacopo Martelossi¹, Giobbe Forni³,
Castrense Savojardo⁴, Barbara Mantovani¹, Andrea Luchetti¹

¹*Department of Biological, Geological and Environmental Sciences, University of Bologna, Bologna, Italy.*

²*Fano Marine Center, Fano (PU), Italy.*

³*Department of Agricultural and Environmental Sciences, University of Milan, Milan, Italy.*

⁴*Department of Pharmacy and Biotechnology, University of Bologna, Bologna, Italy.*

Published in: 2023, *Frontiers in Ecology and Evolution*, 11:1046960.

10.3389/fevo.2023.1046960

Abstract. *Hox* and *ParaHox* genes (HPHGs) are key developmental genes that pattern regional identity along the anterior–posterior body axis of most animals. Here, we identified HPHGs in tadpole shrimps (Pancrustacea, Branchiopoda, Notostraca), an iconic example of the so-called “living fossils” and performed a comparative genomics analysis of HPHGs and the *Hox* cluster among major branchiopod lineages. Notostraca possess the entire *Hox* complement, and the *Hox* cluster seems to be split into two different subclusters, although we were not able to support this finding with chromosome-level assemblies. However, the genomic structure of *Hox* genes in Notostraca appears more derived than that of *Daphnia* spp., which instead retains the plesiomorphic condition of a single compact cluster. Spinicaudata and *Artemia franciscana* show instead a *Hox* cluster subdivided across two or more genomic scaffolds with some orthologs either duplicated or missing. Yet, branchiopod HPHGs are similar among the various clades in terms of both intron length and number, as well as in their pattern of molecular evolution. Sequence substitution rates are in fact generally similar for most of the branchiopod *Hox* genes and the few differences we found cannot be traced back to natural selection, as they are not associated with any signals of diversifying selection or substantial switches in selective modes. Altogether, these findings do not support a significant stasis in the Notostraca *Hox* cluster and further confirm how morphological evolution is not tightly associated with genome dynamics.

Multiple and diversified transposon lineages contribute to early and recent bivalve genome evolution.

Jacopo Martelossi¹, Filippo Nicolini^{1,2}, Simone Subacchi¹, Daniela Pasquale¹, Fabrizio Ghiselli¹, Andrea Luchetti¹

¹*Department of Biological, Geological and Environmental Sciences, University of Bologna, Bologna, Italy.*

²*Fano Marine Center, Fano (PU), Italy.*

Published in: 2023, *BMC Biology*, 21:145. 10.1186/s12915-023-01632-z

Abstract. **Background.** Transposable elements (TEs) can represent one of the major sources of genomic variation across eukaryotes, providing novel raw materials for species diversification and innovation. While considerable effort has been made to study their evolutionary dynamics across multiple animal clades, molluscs represent a substantially understudied phylum. Here, we take advantage of the recent increase in mollusc genomic resources and adopt an automated TE annotation pipeline combined with a phylogenetic tree-based classification, as well as extensive manual curation efforts, to characterize TE repertoires across 27 bivalve genomes with a particular emphasis on DDE/D class II elements, long interspersed nuclear elements (LINEs), and their evolutionary dynamics. **Results.** We found class I elements as highly dominant in bivalve genomes, with LINE elements, despite less represented in terms of copy number per genome, being the most common retroposon group covering up to 10% of their genome. We mined 86,488 reverse transcriptases (RVT) containing LINE coming from 12 clades distributed across all known superfamilies and 14,275 class II DDE/D-containing transposons coming from 16 distinct superfamilies. We uncovered a previously underestimated rich and diverse bivalve ancestral transposon complement that could be traced back to their most recent common ancestor that lived about 500 Mya. Moreover, we identified multiple instances of lineage-specific emergence and loss of different LINEs and DDE/D lineages with the interesting cases of CR1-Zenon, Proto2, RTE-X, and Academ elements that underwent a bivalve-specific amplification likely associated with their diversification. Finally, we found that this LINE diversity is maintained in extant species by an equally diverse set of long-living and potentially active elements, as suggested by their evolutionary history and transcription profiles

in both male and female gonads. **Conclusions.** We found that bivalves host an exceptional diversity of transposons compared to other molluscs. Their LINE complement could mainly follow a “stealth drivers” model of evolution where multiple and diversified families are able to survive and co-exist for a long period of time in the host genome, potentially shaping both recent and early phases of bivalve genome evolution and diversification. Overall, we provide not only the first comparative study of TE evolutionary dynamics in a large but understudied phylum such as Mollusca, but also a reference library for ORF-containing class II DDE/D and LINE elements, which represents an important genomic resource for their identification and characterization in novel genomes.

Towards a time-tree solution for Branchiopoda diversification: a jackknife assessment of fossil age priors.

Niccolò Righetti^{1*}, Filippo Nicolini^{2*}, Giobbe Forni², Andrea Luchetti²

¹*Laboratoire de Biologie Computationnelle et Quantitative (LCQB), Sorbonne Université, CNRS, IBPS, UMR7238, Paris, France.*

²*Department of Biological, Geological and Environmental Sciences, University of Bologna, Bologna, Italy.*

* the authors equally contributed to this work.

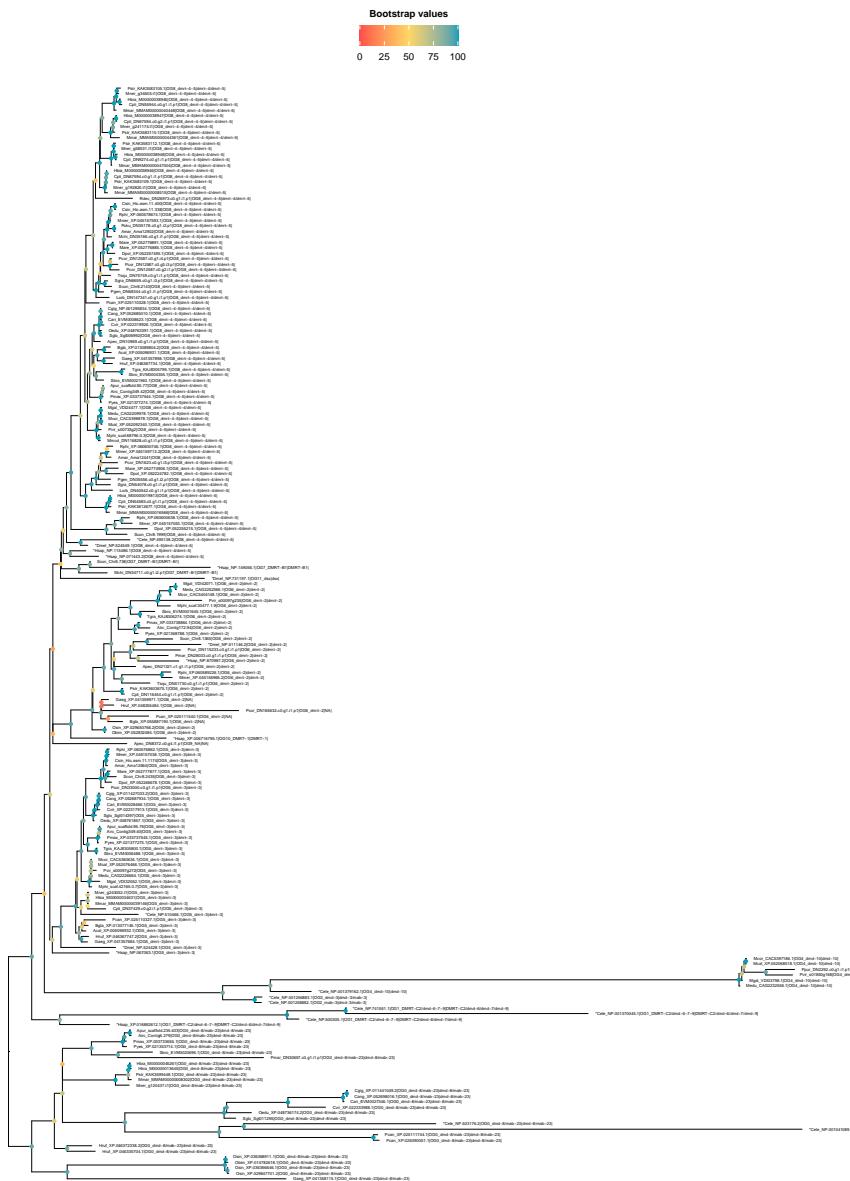
Submitted for peer-review.

Abstract. An understanding of Branchiopoda's evolutionary history is crucial for a comprehensive knowledge of the Pancrustacea tree of life, given their close evolutionary relationship with Hexapoda. Despite significant advances in molecular and morphological phylogenetics that have resolved much of the branchiopod backbone topology, a reliable temporal framework remains elusive. Key challenges include a sparse fossil record, long-term morphological stasis, and past topological inconsistencies. Leveraging a Bayesian Inference approach and the most extensive phylogenomic dataset for branchiopod to date, encompassing 46 species and over 130 genes, we inferred a time-calibrated phylogenetic tree. Furthermore, to strengthen the confidence in our divergence times estimation, we assessed the impact of age priors, topological uncertainties, and gene trees which are discordant from the species trees. Our results are largely consistent with the fossil record and with previous studies, indicating that Branchiopoda originated between 400 and 500 million years ago, and the orders of large branchiopods diversified during the Mesozoic. Concerning Cladocera, results remain problematic, with a sharper uncertainty in the diversification time with respect to the fossil record. Though, the jackknife resampling of fossils and the other sensitivity analyses proved our calibration method to be robust, suggesting that the difficulties in obtaining a paleontological-consistent time tree may be hindered by the variability in branchiopod substitution rates and topological instability within certain clades.

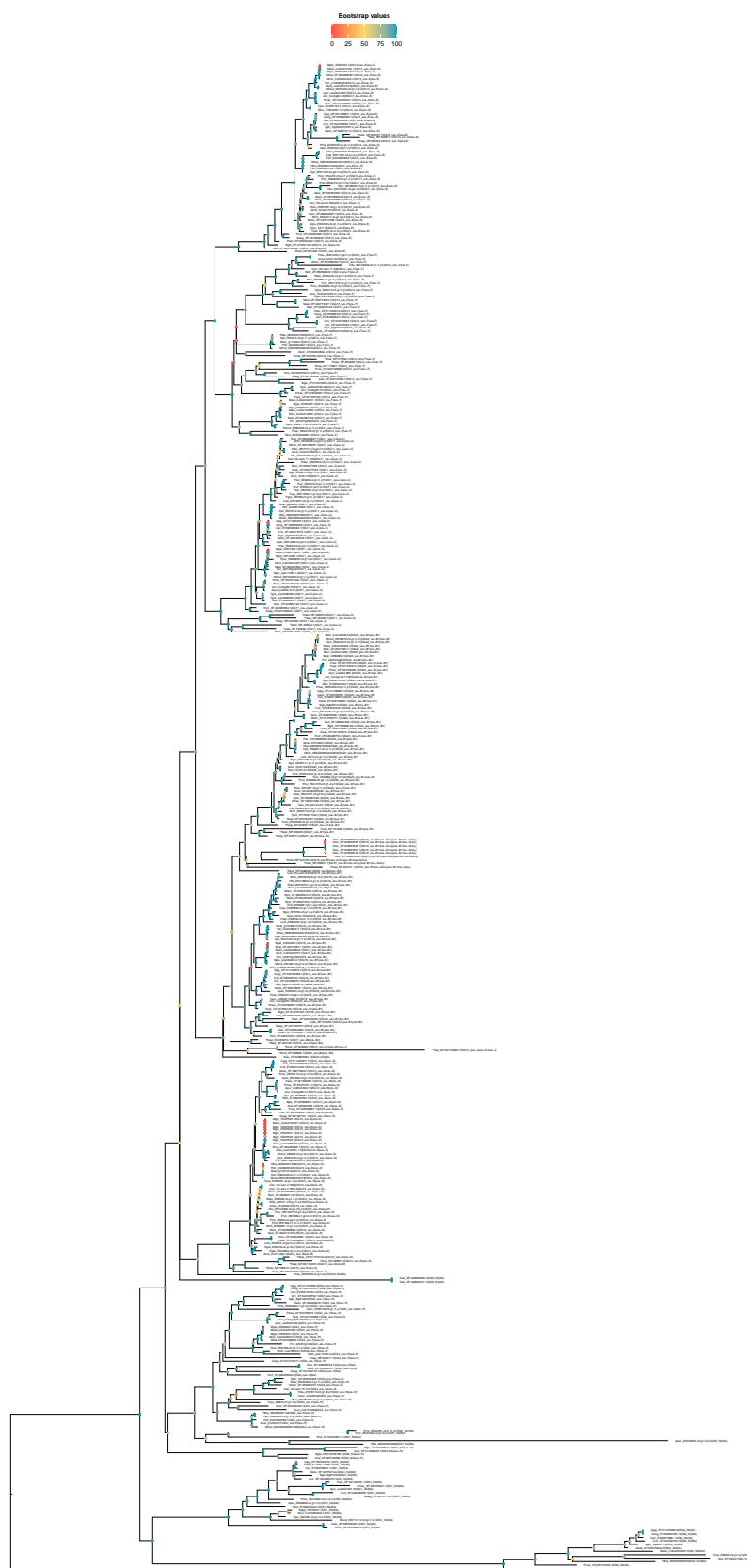
Supplementary materials

Supplementary figures

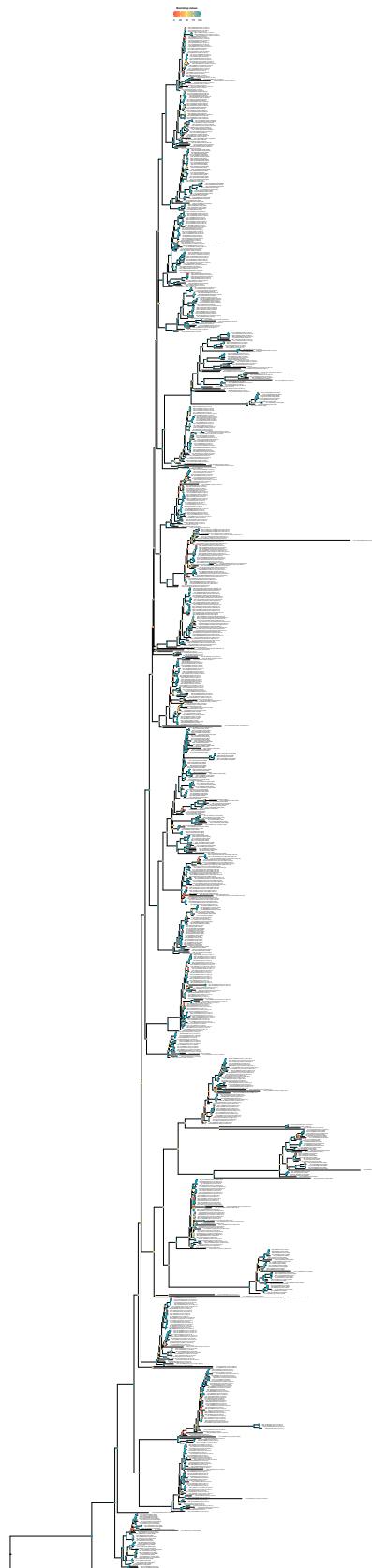
High-quality supplementary figures are available at the following GitHub repository: [LINK](#) [LINK](#) [LINK](#).



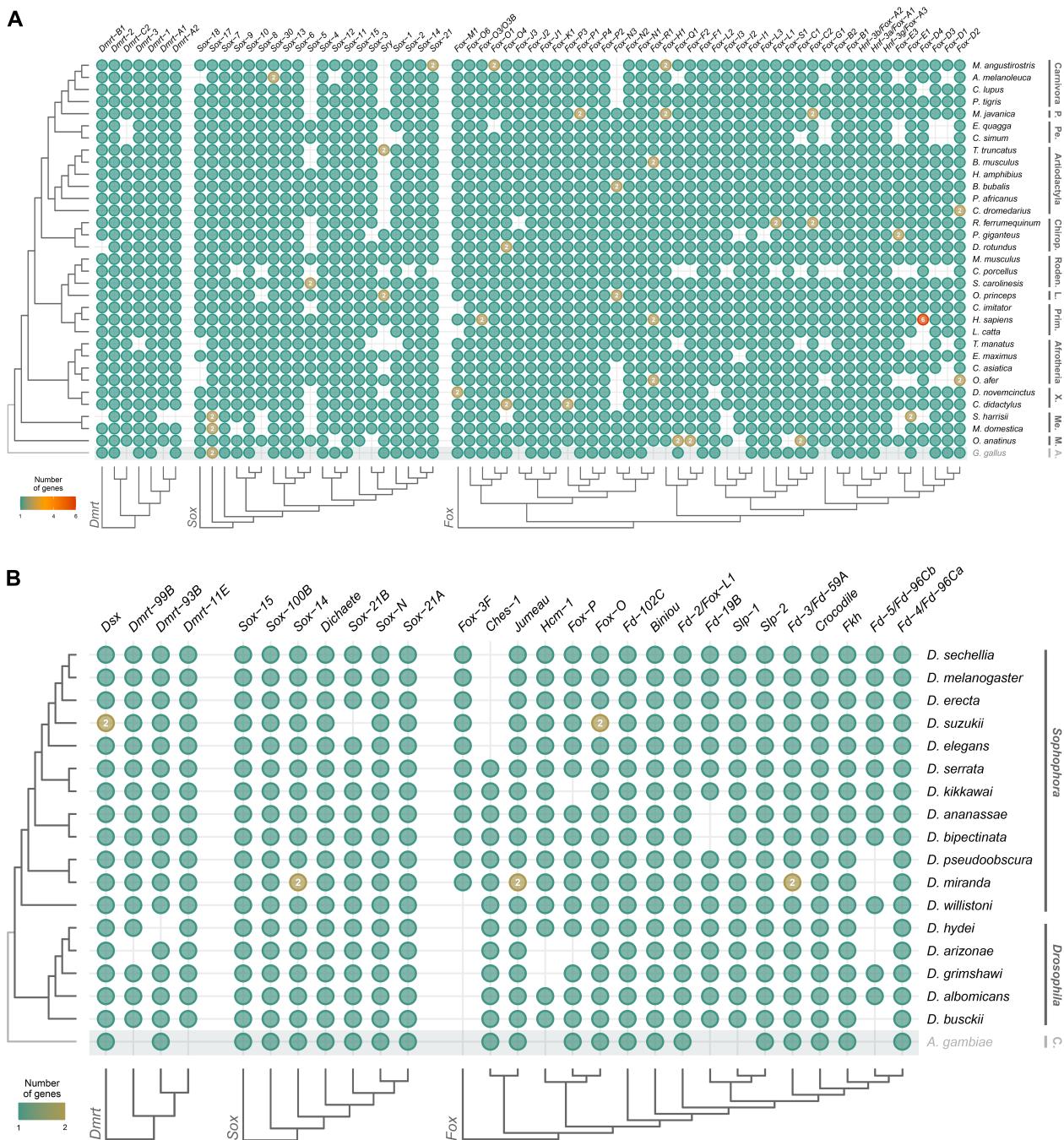
Supplementary Figure S1 – maximum likelihood (ML) phylogenetic tree of the Dmrt gene family in molluscs, including the possvm orthology inference. Reference genes from *Homo sapiens*, *Caenorhabditis elegans*, and *Drosophila melanogaster* are marked with an asterisk at the beginning of the tip names. Species ID can be found in **Tab. S1**. The tree has been midpoint rooted. Bootstrap values are shown for each node.



Supplementary Figure S2 – ML phylogenetic tree of the Sox gene family in molluscs, including the possvm orthology inference. Reference genes from *H. sapiens*, *C. elegans*, and *D. melanogaster* are marked with an asterisk at the beginning of the tip names. Species ID can be found in **Tab. S1**. Bootstrap values are shown for each node.

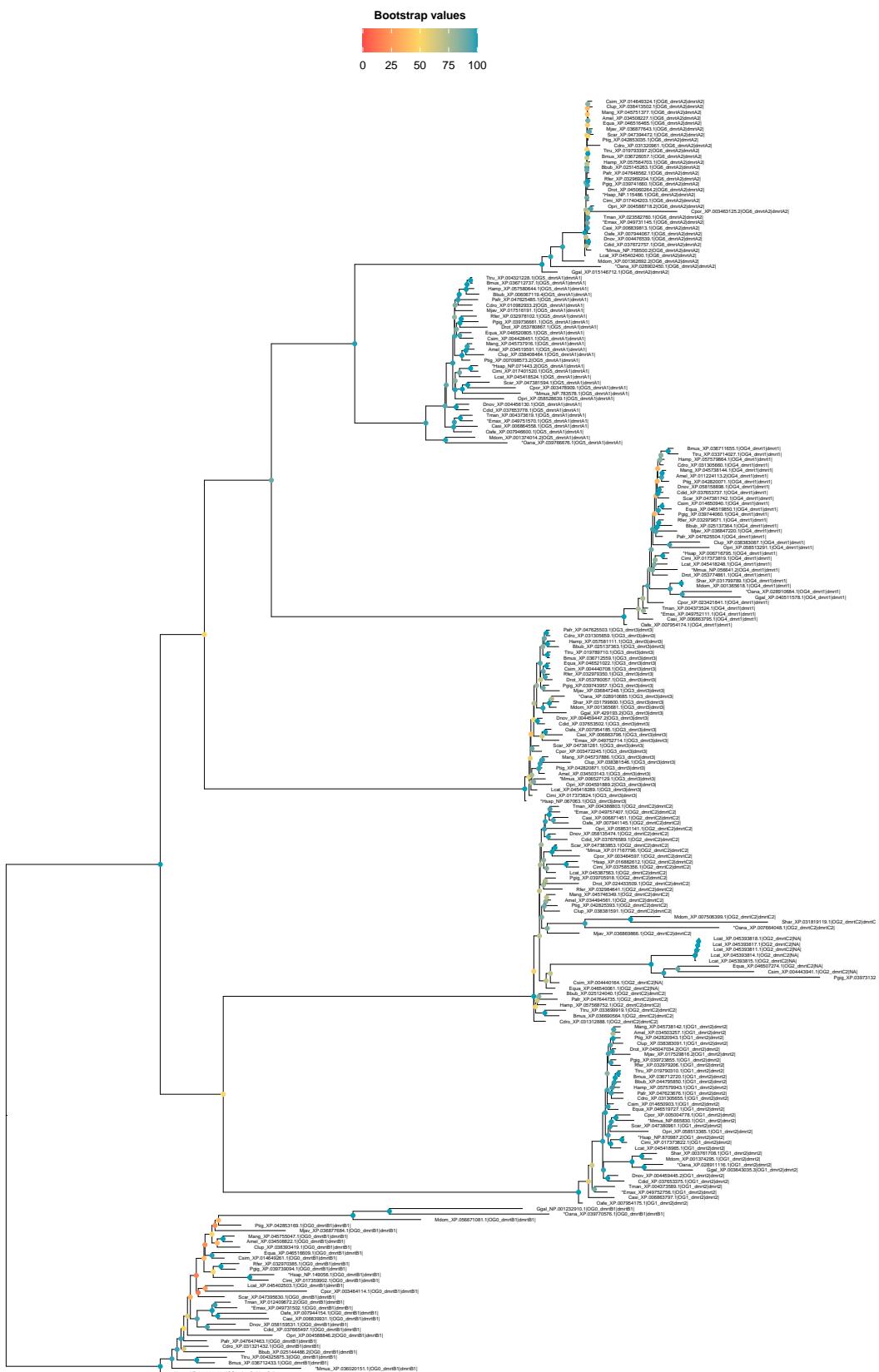


Supplementary Figure S3 – ML phylogenetic tree of the Fox gene family in molluscs, including the possvm orthology inference. Reference genes from *H. sapiens*, *C. elegans*, and *D. melanogaster* are marked with an asterisk at the beginning of the tip names. Species ID can be found in **Tab. S1**. Bootstrap values are shown for each node.

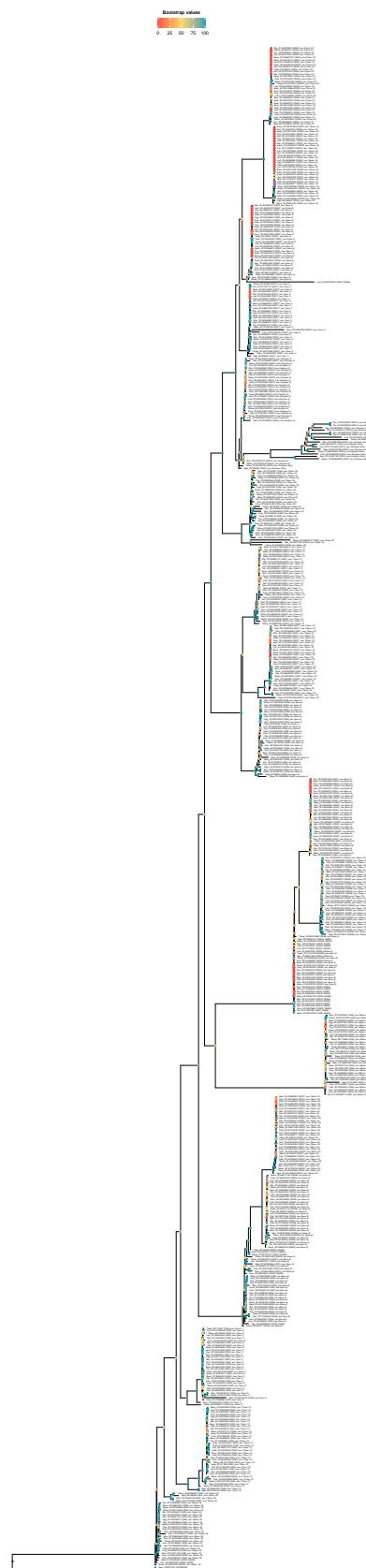


Supplementary Figure S4 – The DSFG complement in Mammalia (A) and Drosophila spp (B).

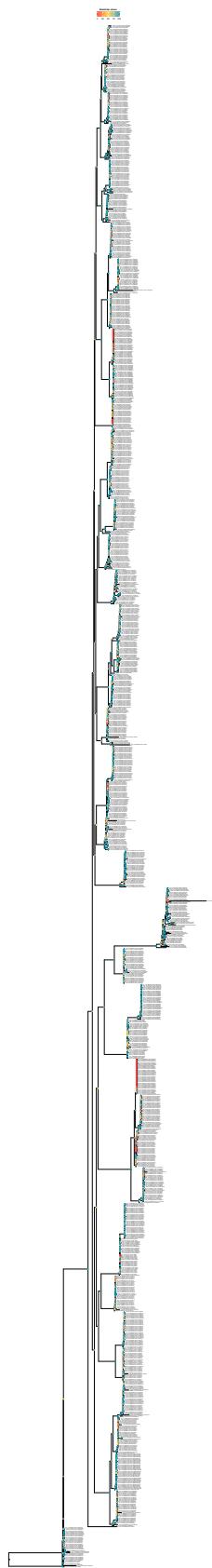
Presence/absence of genes in various species are indicated by filled circles. Numbers inside each circle specify genes with 2 or more copies. The shaded area highlights outgroup species, *Gallus gallus* (Aves) for mammals and *Anopheles gambiae* (Culicidae) for fruit flies. The phylogenetic tree of analysed species, as inferred from literature, is shown on the left, while major taxonomic groups are reported on the right. All species are represented by genomic data. Dmrt, Sox, and Fox gene (DSFG) trees are shown on the bottom (full trees can be found in Fig. S5 and S7). Full species names for both mammals and fruit flies, along with all assembly and taxonomic information, can be found in Tab. S4 and S5, respectively. A.: Aves; Chirop.: Chiroptera; L.: Lagomorpha; M.: Monotremata; Me.: Metatheria; P.: Pholidota; Pe.: Perissodactyla; Prim.: Primates; Roden.: Rodentia; X.: Xenarthra; C.: Culicidae.



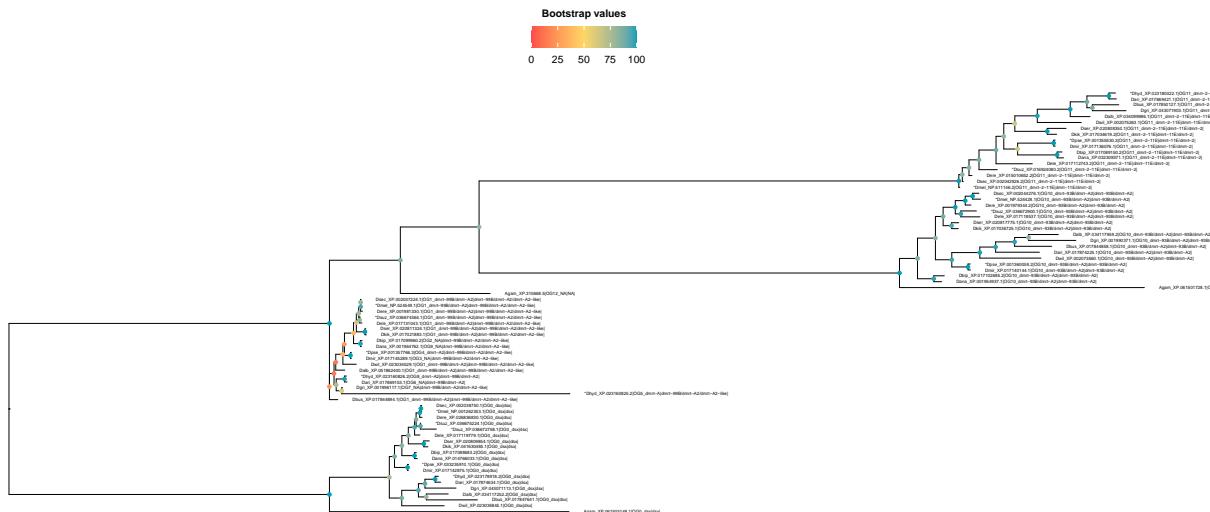
Supplementary Figure S5 – ML phylogenetic tree of the dsx and mab-3 related transcription factor (Dmrt) gene family in mammals, including the Possvm orthology inference. Reference genes from *H. sapiens*, *Mus musculus*, *Elephas maximus indicus*, and *Ornithorhynchus anatinus* are marked with an asterisk at the beginning of the tip names. Species ID can be found in **Tab. S4**. The tree has been midpoint rooted. Bootstrap values are shown for each node.



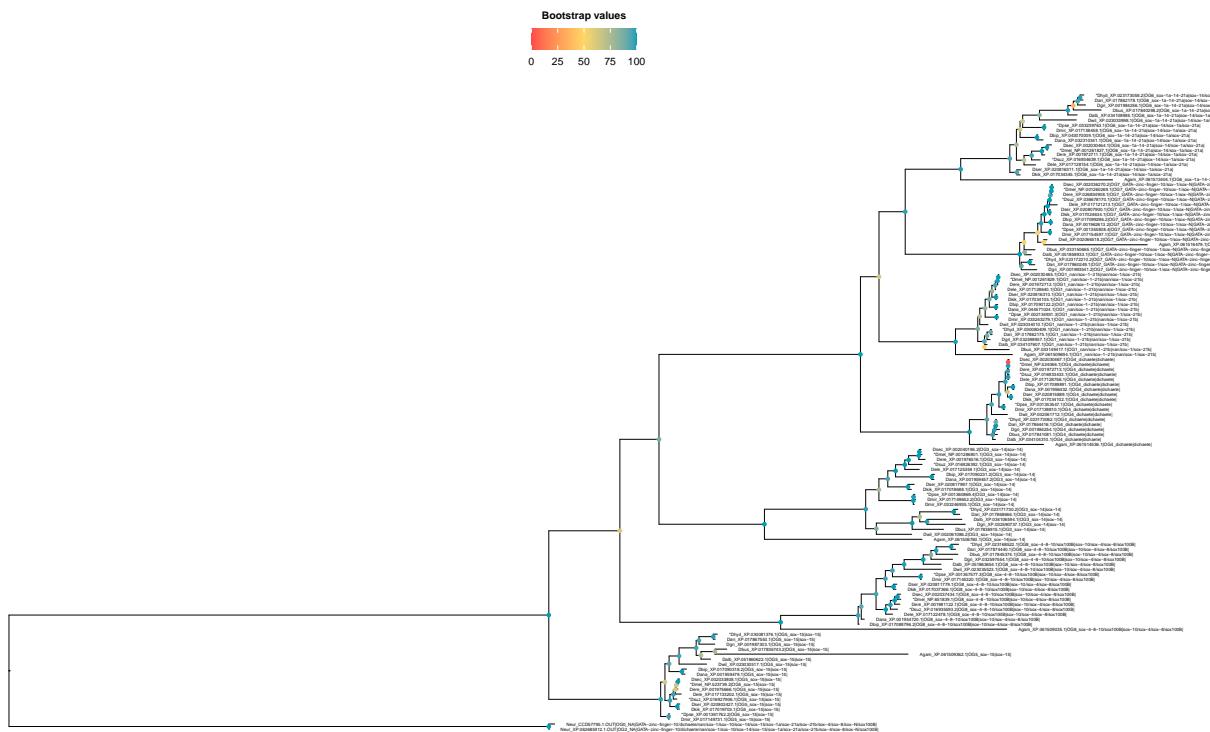
Supplementary Figure S6 – ML phylogenetic tree of the Sry-related HMG-box (Sox) gene family in mammals, including the Possvm orthology inference. Reference genes from *H. sapiens*, *M. musculus*, *E. maximus indicus*, and *O. anatinus* are marked with an asterisk at the beginning of the tip names. Species ID can be found in **Tab. S4**. Bootstrap values are shown for each node.



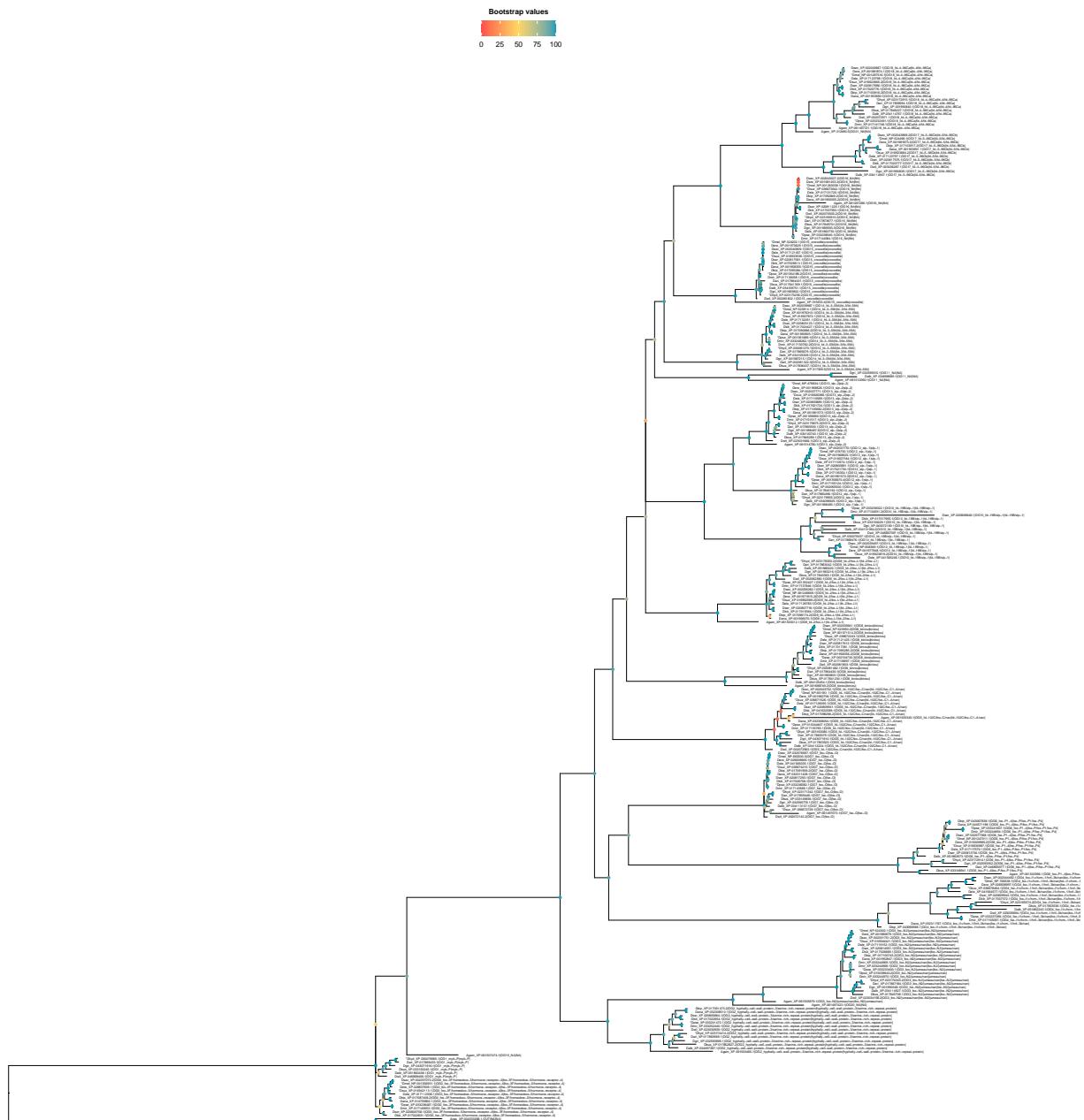
Supplementary Figure S7 – ML phylogenetic tree of the forkhead box (Fox) gene family in mammals, including the Possvm orthology inference. Reference genes from *H. sapiens*, *M. musculus*, *E. maximus*, *I. indicus*, and *O. anatinus* are marked with an asterisk at the beginning of the tip names. Species ID can be found in **Tab. S4**. Bootstrap values are shown for each node.



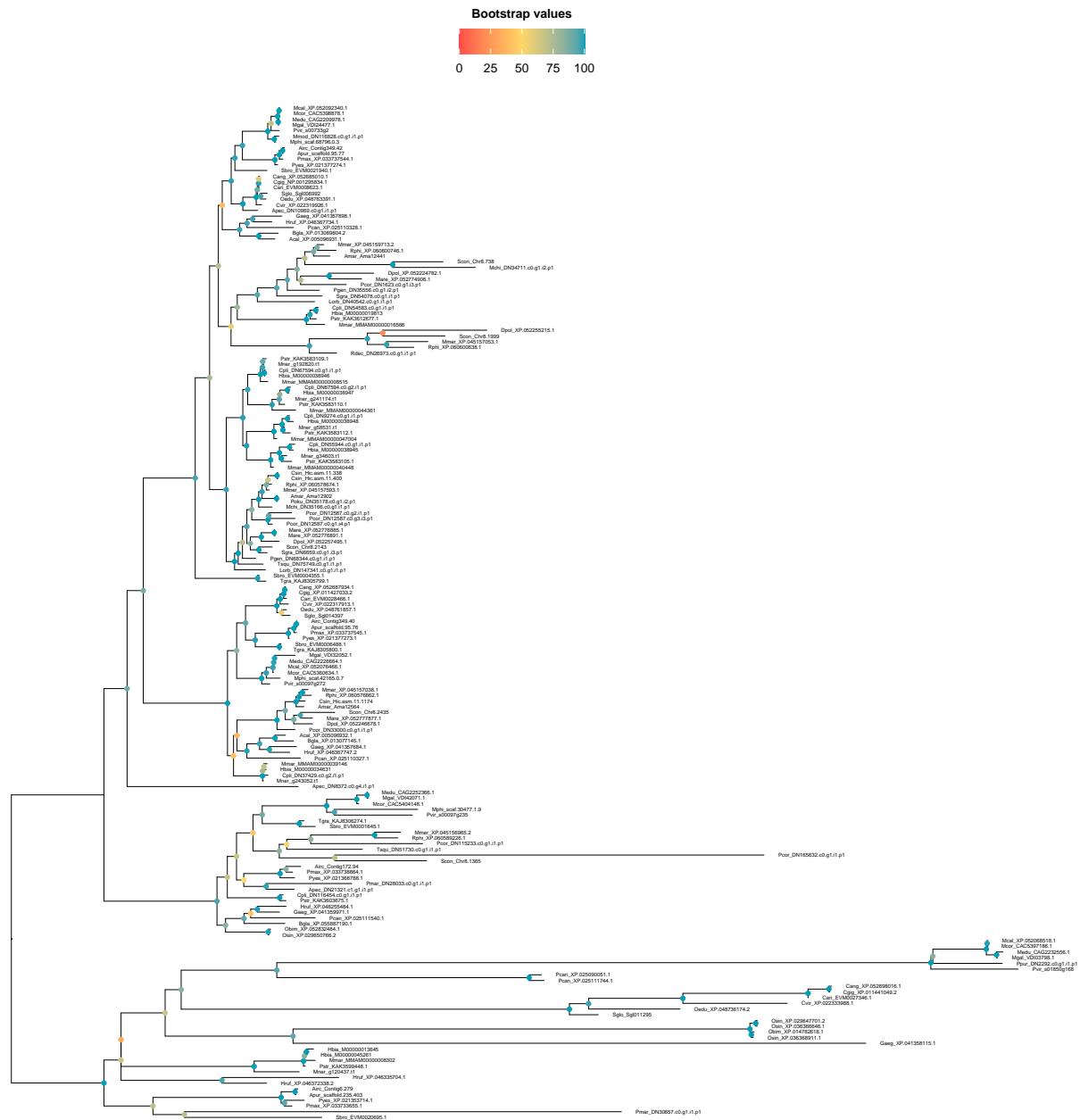
Supplementary Figure S8 – ML phylogenetic tree of the Dmrt gene family in fruit flies, including the Possvm orthology inference. Reference genes from *D. melanogaster*, *Drosophila hydei*, *Drosophila pseudoobscura*, and *Drosophila suzukii* are marked with an asterisk at the beginning of the tip names. Species ID can be found in **Tab. S5**. The tree has been midpoint rooted. Bootstrap values are shown for each node.



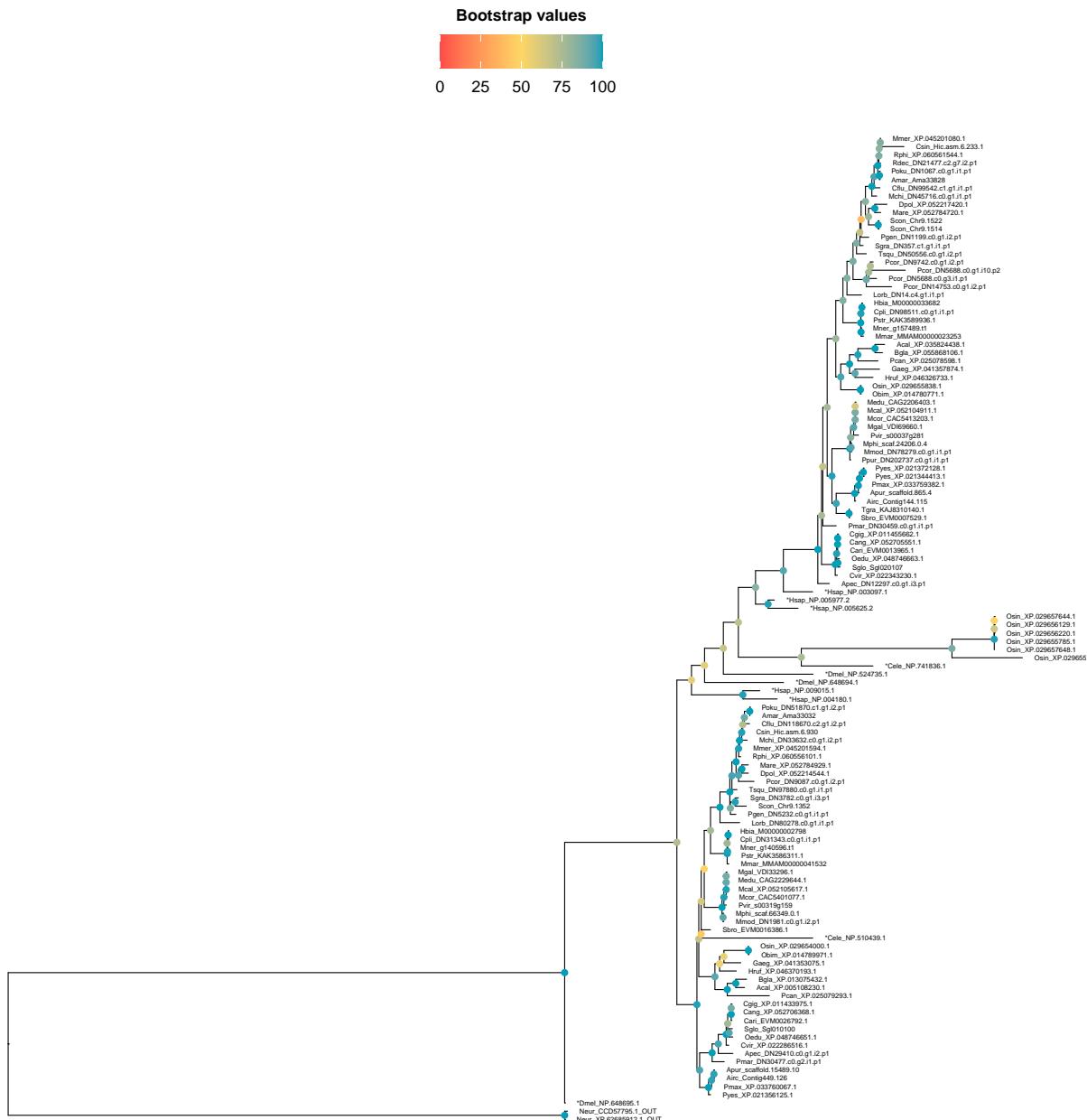
Supplementary Figure S9 – ML phylogenetic tree of the Sox gene family in fruit flies, including the Possvm orthology inference. Reference genes from *D. melanogaster*, *D. hydei*, *D. pseudoobscura*, and *D. suzukii* are marked with an asterisk at the beginning of the tip names. Species ID can be found in **Tab. S5**. Bootstrap values are shown for each node.



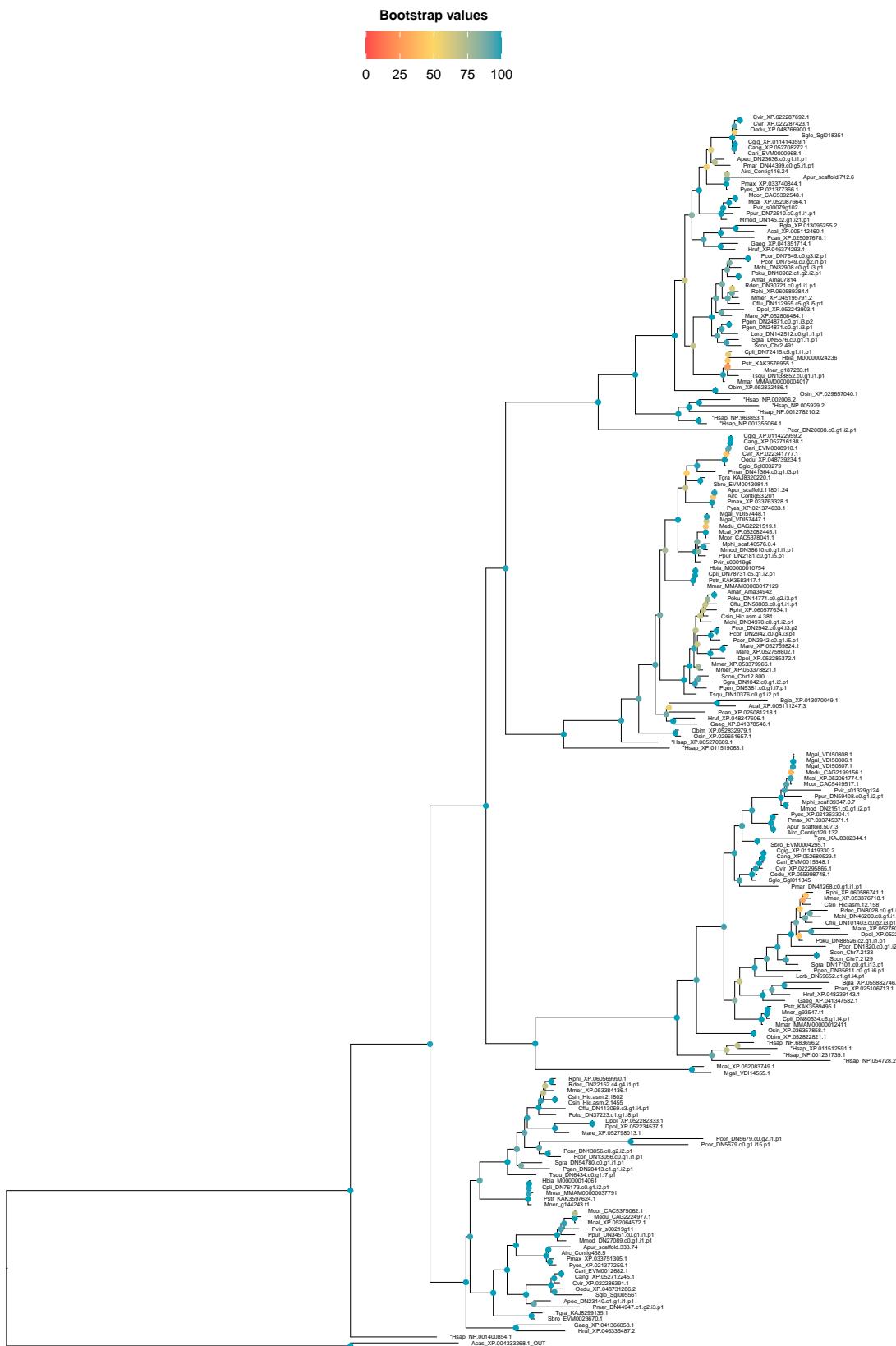
Supplementary Figure S10 – ML phylogenetic tree of the Fox gene family in fruit flies, including the Possvm orthology inference. Reference genes from *D. melanogaster*, *D. hydei*, *D. pseudoobscura*, and *D. suzukii* are marked with an asterisk at the beginning of the tip names. Species ID can be found in **Tab. S5**. Bootstrap values are shown for each node.



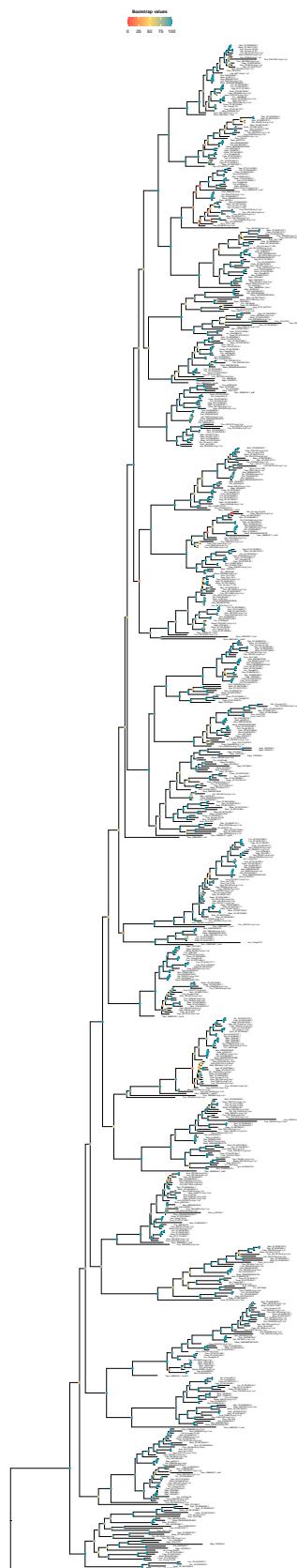
Supplementary Figure S11 – ML phylogenetic tree of the Dmrt gene family in mollusc species.
 Species ID can be found in **Tab. S1**. The tree has been midpoint rooted. Bootstrap values are shown for each node.



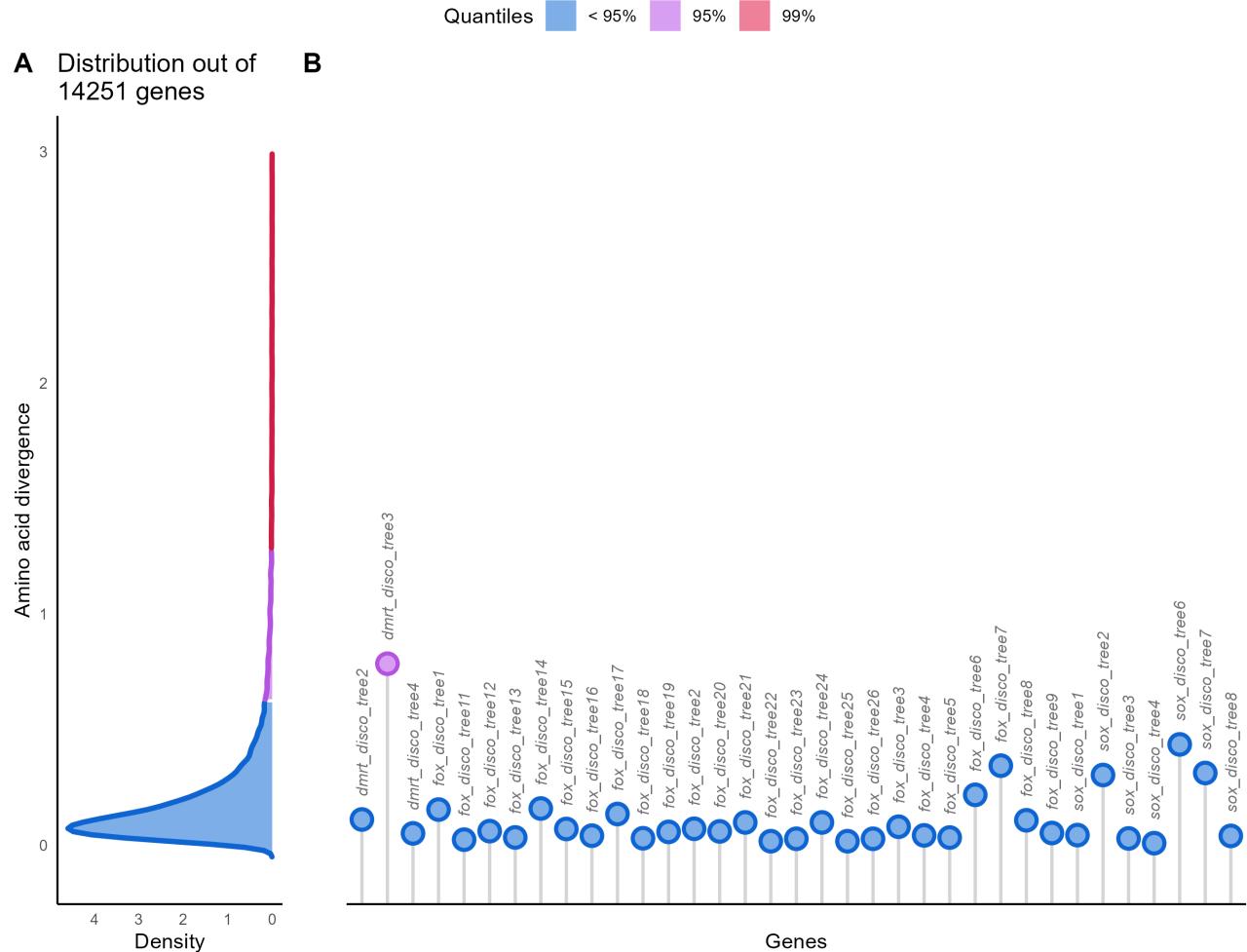
Supplementary Figure S12 – ML phylogenetic tree of Sox-B1 and Sox-B2 genes in mollusc and reference species. Reference genes from *H. sapiens*, *C. elegans*, and *D. melanogaster* are marked with an asterisk at the beginning of the tip names. Species ID can be found in **Tab. S1**. Bootstrap values are shown for each node.



Supplementary Figure S13 – ML phylogenetic tree of Fox-J2, Fox-M, Fox-O, and Fox-P genes in mollusc and reference species. Reference genes from *H. sapiens*, *C. elegans*, and *D. melanogaster* are marked with an asterisk at the beginning of the tip names. Species ID can be found in **Tab. S1**. Bootstrap values are shown for each node.



Supplementary Figure S14 – ML phylogenetic tree of the Fox gene family in bivalves and the sea urchin *Strongylocentrotus purpuratus* (Spur) ML phylogenetic tree of the Fox gene family in bivalves and the sea urchin *S. purpuratus* (Spur). Reference genes from *S. purpuratus* are marked with an asterisk at the beginning of the tip names. Species ID can be found in **Tab. S1**. *S. purpuratus* genes are those given by **Tu et al., 2006**. Bootstrap values are shown for each node.



Supplementary Figure S15 – Distribution of amino acid sequence divergence (AASD) of single-copy orthogroups in *Crassostrea gigas*, *Crassostrea angulata*, *Crassostrea ariakensis*, and *Crassostrea virginica* (A), including DSFG (B). The distribution of AASD in *Crassostrea* has been computed on the median values of pairwise distances of over 14k single-copy orthogroups (SCOs). Circle heights of DSFGs show the median value of their AASD. *Dmrt-1L* genes are indicated as ‘dmrt₋disco₋tree3’.

Supplementary tables

All the supplementary tables are available in a parsable version at the following GitHub repository: [LINK](#) [LINK](#) [LINK](#).

Supplementary Table S1 – Genomic and transcriptomic data of bivalves and other molluscs. For each species, the relative ID, taxonomic information, BUSCO statistics, NCBI accession number, and source publication are reported. Biv: Bivalvia; Ca: Cenogastropoda; Cep: Cephalopoda; Co: Coleoidea; Gas: Gastro poda; Gen: Genome; He: Heterobranchia; Im: Imparidentia; Im: Heterobranchia; Ne: Neomphaliones; Pa: Palaeoheterodontia; Pt: Pteriomorpha; Tra: Transcriptome; Ve: Veti gastropoda

Species	ID	Class	Group	Order	Type	Reduced dataset	BUSCO statistics (metazoa.ncbi10)	NCBI acc. no.	Reference	Annotation source
<i>Magallana (Crassostrea) angulata</i>	Cang	Biv	Pt	Ostreida	Gen	No	C:99.1%[S:97.1%,D:2.0%] F:0.3%,M:0.6%	GCF_025612915.1	Teng et al., 2023	NCBI
<i>Magallana (Crassostrea) gigas</i>	Cgig	Biv	Pt	Ostreida	Gen	Yes	C:98.2%[S:93.1%,D:5.1%] F:0.4%,M:1.4%	GCF_902806645.1	Penaloza et al., 2021	NCBI
<i>Magallana (Crassostrea) ariakensis</i>	Cari	Biv	Pt	Ostreida	Gen	No	C:94.8%[S:91.2%,D:3.6%] F:0.7%,M:4.5%	GCA_020567875.1	Li et al., 2021	FigShare
<i>Crassostrea virginica</i>	Cvir	Biv	Pt	Ostreida	Gen	Yes	C:98.2%[S:73.1%,D:25.1%] F:0.3%,M:1.5%	GCF_002022765.2	Gómez-Chiarri et al., 2015	NCBI
<i>Ostrea edulis</i>	Oedu	Biv	Pt	Ostreida	Gen	Yes	C:98.7%[S:97.8%,D:0.9%] F:0.5%,M:0.8%	GCF_947568905.1	Darwin Tree of Life	NCBI
<i>Saccostrea glomerata</i>	Sgio	Biv	Pt	Ostreida	Gen	No	C:89.1%[S:85.5%,D:3.6%] F:4.9%,M:6.0%	GCA_003671525.1	Powell et al., 2018	dbSRGOG
<i>Atrina pectinata</i>	Apec	Biv	Pt	Ostreida	Tra	Yes	C:95.6%[S:93.1%,D:2.5%] F:1.9%,M:2.5%	DRR348924,-25,-26	Shimizu et al., 2022	–
<i>Pinctada margaritifera</i>	Pmar	Biv	Pt	Ostreida	Tra	Yes	C:94.3%[S:93.9%,D:0.4%] F:1.7%,M:4.0%	SRR1039667 SRR1041217	Teaniniuraitemoana et al., 2014	–
<i>Mytilus galloprovincialis</i>	Mgal	Biv	Pt	Mytilida	Gen	Yes	C:80.5%[S:50.4%,D:30.1%] F:8.6%,M:10.9%	GCA_900618805.1	Gerdol et al., 2020	NCBI
<i>Mytilus edulis</i>	Medu	Biv	Pt	Mytilida	Gen	No	C:83.8%[S:70.9%,D:12.9%] F:5.1%,M:11.1%	GCA_905397895.1	Corrochano-Fraile et al., 2022	NCBI
<i>Mytilus unguiculatus (coruscus)</i>	Mcor	Biv	Pt	Mytilida	Gen	No	C:80.8%[S:78.8%,D:2.0%] F:4.3%,M:14.9%	GCA_011752425.2	Yang et al., 2021	NCBI
<i>Mytilus californianus</i>	Mcal	Biv	Pt	Mytilida	Gen	No	C:96.2%[S:95.0%,D:1.2%] F:0.4%,M:3.4%	GCF_021869535.1	Paggeot et al., 2022	NCBI
<i>Perna viridis</i>	Pvir	Biv	Pt	Mytilida	Gen	Yes	C:99.4%[S:99.0%,D:0.4%] F:0.2%,M:0.4%	GCA_018327765.1	Inoue et al., 2021	Google Drive
<i>Modiolus modiolus</i>	Mmod	Biv	Pt	Mytilida	Tra	Yes	C:95.7%[S:92.3%,D:3.4%] F:2.1%,M:2.2%	SRR5043294	Meng et al., 2018	–

Tab. S1 continued from previous page

Species	ID	Class	Group	Order	Type	Reduced dataset	BUSCO statistics ('metazoa_odb10')	NCBI acc. no.	Reference	Annotation source
<i>Modiolus philippinarum</i>	Mphi	Biv	Pt	Mytilida	Gen	No	C:64.9%[S:63.0%,D:1.9%], F:18.8%,M:16.3%	GCA_002080025.1	Sun et al., 2017	Dryad
<i>Perumytilus purpuratus</i>	Ppur	Biv	Pt	Mytilida	Tra	Yes	C:84.2%[S:83.3%,D:0.9%], F:11.8%,M:4.0%	SRR4343820	Briones et al., 2018	–
<i>Argoppecten irradians concentricus</i>	Airc	Biv	Pt	Pectinida	Gen	Yes	C:94.9%[S:94.0%,D:0.9%], F:3.6%,M:1.5%	GCA_004382765.1	Liu et al., 2020	Dryad
<i>Argoppecten purpuratus</i>	Apur	Biv	Pt	Pectinida	Gen	No	C:89.2%[S:88.6%,D:0.6%], F:5.0%,M:5.8%	–	Liu et al., 2020	GigaDB
<i>Pecten maximus</i>	Pmax	Biv	Pt	Pectinida	Gen	Yes	C:98.5%[S:94.5%,D:4.0%], F:0.4%,M:1.1%	GCF_902052985.1	Kenny et al., 2020	NCBI
<i>Mizuhopecten (Patinopecten) yessoensis</i>	Pyes	Biv	Pt	Pectinida	Gen	Yes	C:98.3%[S:96.1%,D:2.2%], F:0.5%,M:1.2%	GCF_002113885.1	Wang, Zhang, et al., 2017	NCBI
<i>Anadara (Scapharca) broughtoni</i>	Sbro	Biv	Pt	Arcida	Gen	Yes	C:91.2%[S:85.8%,D:5.0%], F:2.6%,M:6.2%	–	Bai et al., 2019	GigaDB
<i>Tegillarca granosa</i>	Tgra	Biv	Pt	Arcida	Gen	Yes	C:70.6%[S:61.3%,D:9.3%], F:11.7%,M:17.7%	GCA_029721355.1	–	NCBI
<i>Ruditapes decussatus</i>	Rdec	Biv	Im	Venerida	Tra	Yes	C:84.8%[S:84.1%,D:0.7%], F:7.3%,M:7.9%	SRR527740,-41,-43,-44,-47,-51,-52,-57	Ghiselli et al., 2018	–
<i>Ruditapes philippinarum</i>	Rphi	Biv	Im	Venerida	Gen	Yes	C:97.8%[S:85.5%,D:12.3%], F:0.7%,M:1.5%	GCF_026571515.1	Xu, Martelossi, et al., 2022	NCBI
<i>Meretrix mercenaria</i>	Mmer	Biv	Im	Venerida	Gen	Yes	C:96.0%[S:89.8%,D:6.2%], F:1.0%,M:3.0%	GCF_021730395.1	Farhat et al., 2022	NCBI
<i>Cyclina sinensis</i>	Csin	Biv	Im	Venerida	Gen	Yes	C:94.1%[S:83.9%,D:10.2%], F:1.8%,M:4.1%	GCA_012932295.1	Wei et al., 2020	Dryad
<i>Calyptogena (Archivesica) marisnica</i>	Amar	Biv	Im	Venerida	Gen	No	C:82.1%[S:80.1%,D:2.0%], F:6.0%,M:11.9%	GCA_014443695.1	Ip et al., 2021	FigShare
<i>Phreagena okutanii</i>	Poku	Biv	Im	Venerida	Tra	Yes	C:92.9%[S:85.8%,D:7.1%], F:3.0%,M:4.1%	SRR7156763,-64,-65,-66,-67,-68	Lan et al., 2019	–
<i>Corbicula fluminea</i>	Cflu	Biv	Im	Venerida	Tra	Yes	C:83.7%[S:79.9%,D:3.8%], F:10.3%,M:6.0%	SRR1559272 SRR5512046	Gonzalez et al., 2015 Zhu et al., 2019	–
<i>Mactra chinensis</i>	Mchi	Biv	Im	Venerida	Tra	Yes	C:81.5%[S:80.8%,D:0.7%], F:10.2%,M:8.3%	SRR1263980	–	–

Tab. S1 continued from previous page

Species	ID	Class	Group	Order	Type	Reduced dataset	BUSCO statistics ('metazoa_odb10')	NCBI acc. no.	Reference	Annotation source
<i>Mya arenaria</i>	Mare	Biv	Im	Myida	Gen	Yes	C:98.5%{S:80.4%,D:18.1%}, F:0.4%,M:1.1%	GCF_026914265.1	Hart et al., 2023	NCBI
<i>Dreissena polymorpha</i>	Dpol	Biv	Im	Myida	Gen	Yes	C:97.2%{S:80.1%,D:17.1%}, F:0.4%,M:2.4%	GCF_02053695.1	McCartney et al., 2022	NCBI
<i>Pisidium coreanum</i>	Pcor	Biv	Im	Sphaeriida	Tra	Yes	C:94.5%{S:81.6%,D:12.9%}, F:3.6%,M:1.9%	SRR6474597	–	–
<i>Solen grandis</i>	Sgra	Biv	Im	Adapedonta	Tra	Yes	C:92.7%{S:90.0%,D:2.7%}, F:2.5%,M:4.8%	SRR5484647, SRR5485368, SRR5499447	Nie et al., 2018	–
<i>Sinonorvacula constricta</i>	Scon	Biv	Im	Adapedonta	Gen	Yes	C:90.8%{S:79.2%,D:11.6%}, F:3.5%,M:5.7%	GCA_007844125.1	Ran et al., 2019	Dryad
<i>Panopaea generosa</i>	Pgen	Biv	Im	Adapedonta	Tra	Yes	C:84.1%{S:81.9%,D:2.2%}, F:9.7%,M:6.2%	SRR12218869, -70	Putnam et al., 2022	–
<i>Tridacna squamosa</i>	Tsqu	Biv	Im	Cardiida	Tra	Yes	C:89.7%{S:86.8%,D:2.8%}, F:3.5%,M:6.8%	SRR10824682, -65	Li, Zhou, et al., 2020	–
<i>Loripes orbicularis</i>	Lorb	Biv	Im	Lucinida	Tra	Yes	C:76.1%{S:74.9%,D:1.2%}, F:14.3%,M:9.6%	SRR10002336, -38, -39, -47	Yuen et al., 2019	–
<i>Hyriopsis bivalata</i> (<i>Unio delphinus</i>)	Hbia	Biv	Pa	Unionida	Gen	Yes	C:97.5%{S:94.9%,D:2.6%}, F:2.0%,M:0.5%	GCA_029339505.1	Gomes-dos-Santos et al., 2023	FigShare
<i>Cristaria plicata</i>	Cpli	Biv	Pa	Unionida	Tra	Yes	C:93.6%{S:92.8%,D:0.8%}, F:2.1%,M:4.3%	SRR2175468 SRR3095781	Pathak et al., 2016 Wang, Liu, and Wu, 2017	–
<i>Megalonaia nervosa</i>	Mner	Biv	Pa	Unionida	Gen	Yes	C:65.0%{S:63.3%,D:1.7%}, F:14.0%,M:21.0%	GCA_016617855.1	Rogers et al., 2021	Dryad
<i>Potamilius streckeri</i>	Pstr	Biv	Pa	Unionida	Gen	Yes	C:94.9%{S:93.4%,D:1.5%}, F:1.2%,M:3.9%	GCA_016746295.1	Smith, 2021	NCBI
<i>Margaritifera margaritifera</i>	Mmar	Biv	Pa	Unionida	Gen	Yes	C:92.6%{S:92.1%,D:0.5%}, F:3.0%,M:4.4%	GCA_015947965.1	Gomes-dos-Santos et al., 2021	FigShare
<i>Aplysia californica</i>	Acal	Gas	He	Aplysiida	Gen	No	C:97.8%{S:97.0%,D:0.8%}, F:0.7%,M:1.5%	GCF_000002075.1	Knudsen et al., 2006	NCBI
<i>Biomphalaria glabrata</i>	Bglra	Gas	—	—	Gen	No	C:98.9%{S:98.2%,D:0.7%}, F:0.1%,M:1.0%	GCF_947242115.1	–	NCBI
<i>Pomacea canaliculata</i>	Pcan	Gas	Ca	Architaenioglossa	Gen	No	C:98.2%{S:97.0%,D:1.2%}, F:0.4%,M:1.4%	GCF_003073045.1	Liu et al., 2018	NCBI

Tab. S1 continued from previous page

Species	ID	Class	Group	Order	Type	Reduced dataset	BUSCO statistics ('metazoa_odb10')	NCBI acc. no.	Reference	Annotation source
<i>Giantopelta aegis</i>	Gaeig	Gas	Ne	Neomphalida	Gen	No	C:98.4%[S:94.2%,D:4.2%] F:0.8%,M:0.8%	GCF_016097555.1	Lan et al., 2021	NCBI
<i>Haliotis rufescens</i>	Hruf	Gas	Ve	Lepetellida	Gen	No	C:99.0%[S:98.3%,D:0.7%] F:0.0%,M:1.0%	GCF_023055435.1	—	NCBI
<i>Octopus bimaculoides</i>	Obim	Cep	Co	Octopoda	Gen	No	C:94.9%[S:94.4%,D:0.5%] F:2.3%,M:2.8%	GCF_001194135.2	Albertin et al., 2015	NCBI
<i>Octopus sinensis</i>	Osin	Cep	Co	Octopoda	Gen	No	C:98.1%[S:96.9%,D:1.2%] F:0.9%,M:1.0%	GCF_006345805.1	Li, Bian, et al., 2020	NCBI

Supplementary Table S2 – DSFG family and domain identifiers (IDs) in PANTHER and CDD, respectively. After having retrieved putative DSFGs on the basis of hidden Markov model (HMM) profiles, IDs have been used to retain only reliable hits.

Gene family	PANTHER/CDD	ID	Description
Dmrt	CDD	gnl—CDD—214606	Doublesex DNA-binding motif
	CDD	gnl—CDD—422850	DM DNA binding domain
	PANTHER	PTHR12322	DOUBLESEX AND MAB-3 RELATED TRANSCRIPTION FACTOR DMRT PROTEIN CBR-MAB-23
	PANTHER	PTHR12322-SF115	DOUBLESEX- AND MAB-3-RELATED TRANSCRIPTION FACTOR 1
	PANTHER	PTHR12322-SF116	DOUBLESEX- AND MAB-3-RELATED TRANSCRIPTION FACTOR DMD-4
	PANTHER	PTHR12322-SF118	DOUBLESEX- AND MAB-3-RELATED TRANSCRIPTION FACTOR DMD-9B
	PANTHER	PTHR12322-SF123	DOUBLESEX- AND MAB-3-RELATED TRANSCRIPTION FACTOR 2
	PANTHER	PTHR12322-SF53	DOUBLESEX- AND MAB-3-RELATED TRANSCRIPTION FACTOR A1
	PANTHER	PTHR16897-SF71	STRESS RESPONSE PROTEIN NST1
	PANTHER	PTHR46888-SF11	RIBONUCLEASE H
Sox	CDD	gnl—CDD—432488	SOX transcription factor
	CDD	gnl—CDD—432558	Sox developmental protein N terminal
	CDD	gnl—CDD—438790	high mobility group (HMG)-box found in group B SRY-related high-mobility group (HMG) box (Sox) transcription factors
	CDD	gnl—CDD—438837	high mobility group (HMG)-box found in sex-determining region Y (SRY)-box (Sox) transcription factors
	CDD	gnl—CDD—438838	high mobility group (HMG)-box found in group A, group B and group G of SRY-related high-mobility group (HMG) box (Sox) transcription factors
	CDD	gnl—CDD—438839	high mobility group (HMG)-box found in group C SRY-related high-mobility group (HMG) box (Sox) transcription factors
	CDD	gnl—CDD—438840	high mobility group (HMG)-box found in group E SRY-related high-mobility group (HMG) box (Sox) transcription factors
	CDD	gnl—CDD—438841	high mobility group (HMG)-box found in group F SRY-related high-mobility group (HMG) box (Sox) transcription factors
	CDD	gnl—CDD—438842	high mobility group (HMG)-box found in sex-determining region Y (SRY)-box 30 (SOX30) and similar proteins
	CDD	gnl—CDD—438843	high mobility group (HMG)-box found in sex-determining region Y (SRY)-box 15 (SOX15) and similar proteins
	CDD	gnl—CDD—438844	high mobility group (HMG)-box found in sex-determining region Y (SRY)-box 15 (SOX15) and similar proteins
	CDD	gnl—CDD—438845	high mobility group (HMG)-box found in sex-determining region Y (SRY)-box 4 (SOX4) and similar proteins
	CDD	gnl—CDD—438846	high mobility group (HMG)-box found in sex-determining region Y (SRY)-box 11 (SOX11) and similar proteins
	CDD	gnl—CDD—438847	high mobility group (HMG)-box found in sex-determining region Y (SRY)-box 12 (SOX12) and similar proteins
	CDD	gnl—CDD—438849	high mobility group (HMG)-box found in sex-determining region Y (SRY)-box 7 (SOX7) and similar proteins
	CDD	gnl—CDD—438850	high mobility group (HMG)-box found in sex-determining region Y (SRY)-box 17 (SOX17) and similar proteins
	PANTHER	PTHR10270-SF107	TRANSCRIPTION FACTOR SOX-14
	PANTHER	PTHR10270-SF161	SOX DOMAIN-CONTAINING PROTEIN DICHAETE-RELATED
	PANTHER	PTHR10270-SF199	SEX-DETERMINING REGION Y PROTEIN
	PANTHER	PTHR10270-SF231	TRANSCRIPTION FACTOR SOX-2
	PANTHER	PTHR10270-SF227	TRANSCRIPTION FACTOR SOX-4
	PANTHER	PTHR10270-SF313	TRANSCRIPTION FACTOR SOX-21
	PANTHER	PTHR10270-SF315	TRANSCRIPTION FACTOR SOX-1A-RELATED
	PANTHER	PTHR10270-SF317	TRANSCRIPTION FACTOR SOX-15-RELATED
	PANTHER	PTHR10270-SF322	TRANSCRIPTION FACTOR SOX-3
	PANTHER	PTHR10270-SF324	TRANSCRIPTION FACTOR SOX-3
	PANTHER	PTHR10270-SF326	TRANSCRIPTION FACTOR SOX-3
	PANTHER	PTHR10270-SF326	TRANSCRIPTION FACTOR SOX-3
	PANTHER	PTHR45789-SF2	TRANSCRIPTION FACTOR SOX-9
	PANTHER	PTHR45789-SF2	TRANSCRIPTION FACTOR SOX-9
	PANTHER	PTHR45803-SF1	TRANSCRIPTION FACTOR SOX-9
	PANTHER	PTHR45803-SF2	TRANSCRIPTION FACTOR SOX-9
	PANTHER	PTHR45803-SF5	TRANSCRIPTION FACTOR SOX-9
	PANTHER	PTHR45803-SF5	TRANSCRIPTION FACTOR SOX-9
	PANTHER	PTHR45803-SF1	TRANSCRIPTION FACTOR SOX-9
	PANTHER	PTHR47279-SF1	TRANSCRIPTION FACTOR SOX-30
	PANTHER	PTHR47279-SF1	TRANSCRIPTION FACTOR SOX-30
Fox	CDD	gnl—CDD—410788	Forkhead (FH) domain found in Forkhead box (FOX) family of transcription factors and similar proteins
	CDD	gnl—CDD—410789	Forkhead (FH) domain found in Forkhead box protein A (FOXA) subfamily
	CDD	gnl—CDD—410790	Forkhead (FH) domain found in Forkhead box protein B (FOXB) subfamily
	CDD	gnl—CDD—410791	Forkhead (FH) domain found in Forkhead box protein C (FOXC) subfamily
	CDD	gnl—CDD—410792	Forkhead (FH) domain found in Forkhead box protein D (FOXD) subfamily
	CDD	gnl—CDD—410793	Forkhead (FH) domain found in the Forkhead box protein E (FOXE) subfamily
	CDD	gnl—CDD—410794	Forkhead (FH) domain found in the Forkhead box protein F (FOXF) subfamily
	CDD	gnl—CDD—410795	Forkhead (FH) domain found in the Forkhead box protein G (FOXG) subfamily
	CDD	gnl—CDD—410796	Forkhead (FH) domain found in the Forkhead box protein H (FOXH) subfamily
	CDD	gnl—CDD—410797	Forkhead (FH) domain found in Forkhead box protein J1 (FOXJ1) and similar proteins
	CDD	gnl—CDD—410798	Forkhead (FH) domain found in Forkhead box proteins FOXJ2, FOXJ3 and similar proteins
	CDD	gnl—CDD—410799	Forkhead (FH) domain found in the Forkhead box protein I (FOXI) subfamily
	CDD	gnl—CDD—410800	Forkhead (FH) domain found in the Forkhead box protein K (FOXK) subfamily

Tab. S2 continued from previous page

Gene family	PANTHER/CDD	ID	Description
	CDD	gnl—CDD—410801	Forkhead (FH) domain found in Forkhead box protein L1 (FOXL1) and similar proteins
	CDD	gnl—CDD—410802	Forkhead (FH) domain found in Forkhead box protein L2 (FOXL2) and similar proteins
	CDD	gnl—CDD—410803	Forkhead (FH) domain found in the Forkhead box protein M (FOXM1) subfamily
	CDD	gnl—CDD—410804	Forkhead (FH) domain found in Forkhead box protein N1 (FOXN1) and similar proteins
	CDD	gnl—CDD—410805	Forkhead (FH) domain found in Forkhead box protein N2 (FOXN2) and similar proteins
	CDD	gnl—CDD—410806	Forkhead (FH) domain found in the Forkhead box protein O (FOXO) subfamily
	CDD	gnl—CDD—410807	Forkhead (FH) domain found in the Forkhead box protein P (FOXP) subfamily
	CDD	gnl—CDD—410808	Forkhead (FH) domain found in Forkhead box protein Q1 (FOXQ1) and similar proteins
	CDD	gnl—CDD—410809	Forkhead (FH) domain found in Forkhead box protein Q2 (FOXQ2) and similar proteins
	CDD	gnl—CDD—410810	Forkhead (FH) domain found in the Forkhead box protein R (FOXR) subfamily
	CDD	gnl—CDD—410811	Forkhead (FH) domain found in Forkhead box protein S1 (FOXS1)
	CDD	gnl—CDD—410812	Forkhead (FH) domain found in Forkhead box protein A2 (FOXA2) and similar proteins
	CDD	gnl—CDD—410813	Forkhead (FH) domain found in Forkhead box protein A3 (FOXA3) and similar proteins
	CDD	gnl—CDD—410814	Forkhead (FH) domain found in Forkhead box protein B1 (FOXB1) and similar proteins
	CDD	gnl—CDD—410816	Forkhead (FH) domain found in Forkhead box protein B2 (FOXB2) and similar proteins
	CDD	gnl—CDD—410817	Forkhead (FH) domain found in Forkhead box protein C1 (FOXC1) and similar proteins
	CDD	gnl—CDD—410818	Forkhead (FH) domain found in Forkhead box protein C2 (FOXC2) and similar proteins
	CDD	gnl—CDD—410819	Forkhead (FH) domain found in Forkhead box protein D3 (FOXD3) and similar proteins
	CDD	gnl—CDD—410820	Forkhead (FH) domain found in Forkhead box protein D4 (FOXD4) and similar proteins
	CDD	gnl—CDD—410821	Forkhead (FH) domain found in Forkhead box protein E1 (FOXE1) and similar proteins
	CDD	gnl—CDD—410822	Forkhead (FH) domain found in Forkhead box protein F2 (FOXF2) and similar proteins
	CDD	gnl—CDD—410823	Forkhead (FH) domain found in Forkhead box protein J2 (FOXJ2) and similar proteins
	CDD	gnl—CDD—410824	Forkhead (FH) domain found in Forkhead box protein K2 (FOXK2) and similar proteins
	CDD	gnl—CDD—410825	Forkhead (FH) domain found in Forkhead box protein L1 (FOXL1) and similar proteins
	CDD	gnl—CDD—410826	Forkhead (FH) domain found in Forkhead box protein L2 (FOXL2) and similar proteins
	CDD	gnl—CDD—410827	Forkhead (FH) domain found in Forkhead box protein M (FOXM1) and similar proteins
	CDD	gnl—CDD—410828	Forkhead (FH) domain found in Forkhead box protein N1 (FOXN1) and similar proteins
	CDD	gnl—CDD—410829	Forkhead (FH) domain found in Forkhead box protein N2 (FOXN2) and similar proteins
	CDD	gnl—CDD—410830	Forkhead (FH) domain found in Forkhead box protein O1 (FOXO1)
	CDD	gnl—CDD—410831	Forkhead (FH) domain found in Forkhead box protein O4 (FOXO4) and similar proteins
	CDD	gnl—CDD—410832	Forkhead (FH) domain found in Forkhead box protein P1 (FOXP1)
	CDD	gnl—CDD—410833	Forkhead (FH) domain found in Forkhead box protein P2 (FOXP2) and similar proteins
	CDD	gnl—CDD—410834	Forkhead (FH) domain found in Forkhead box protein P3 (FOXP3) and similar proteins
	CDD	gnl—CDD—410835	Forkhead (FH) domain found in Forkhead box protein P4 (FOXP4) and similar proteins
	CDD	gnl—CDD—410836	
	CDD	gnl—CDD—410837	
	CDD	gnl—CDD—410838	
	CDD	gnl—CDD—410839	
	CDD	gnl—CDD—410840	
	CDD	gnl—CDD—410841	
PANTHER		PTHR1829	FORKHEAD BOX PROTEIN
PANTHER		PTHR1829-SF142	FORKHEAD BOX PROTEIN E3
PANTHER		PTHR1829-SF156	FORKHEAD BOX PROTEIN Q1
PANTHER		PTHR1829-SF206	FORKHEAD BOX PROTEIN B1
PANTHER		PTHR1829-SF209	FORKHEAD BOX PROTEIN D2
PANTHER		PTHR1829-SF335	FORKHEAD BOX PROTEIN L1
PANTHER		PTHR1829-SF340	FORKHEAD BOX PROTEIN L2
PANTHER		PTHR1829-SF342	FORKHEAD BOX PROTEIN D1
PANTHER		PTHR1829-SF348	FORKHEAD BOX PROTEIN D3
PANTHER		PTHR1829-SF361	FORKHEAD BOX PROTEIN PES-1
PANTHER		PTHR1829-SF398	FORKHEAD TRANSCRIPTION FACTOR FH-9
PANTHER		PTHR1829-SF412	FORKHEAD BOX CL-RELATED
PANTHER		PTHR1829-SF416	FORKHEAD BOX PROTEIN N3-LIKE PROTEIN
PANTHER		PTHR1829-SF417	FORKHEAD BOX PROTEIN N4
PANTHER		PTHR1829-SF319	FORKHEAD BOX PROTEIN N2
PANTHER		PTHR1829-SF399	FORKHEAD BOX PROTEIN N3
PANTHER		PTHR1829-SF401	FORKHEAD BOX PROTEIN N2
PANTHER		PTHR1829-SF402	FORKHEAD BOX PROTEIN O
PANTHER		PTHR1829-SF406	FORKHEAD BOX PROTEIN O
PANTHER		PTHR1829-SF407	FORKHEAD BOX P-ISOFORM C
PANTHER		PTHR1829-SF408	FORKHEAD BOX PROTEIN P1
PANTHER		PTHR1829-SF409	FORKHEAD BOX PROTEIN C
PANTHER		PTHR1829-SF410	FORKHEAD BOX PROTEIN K2
PANTHER		PTHR1829-SF411	FORKHEAD BOX PROTEIN K1
PANTHER		PTHR1829-SF412	FORKHEAD BOX PROTEIN BINOU
PANTHER		PTHR1829-SF413	FORKHEAD BOX PROTEIN BINOU

Fox

Tab. S2 continued from previous page

Gene family	PANTHER/CDD	ID	Description
Fox	PANTHER	PTH46617	FORKHEAD BOX PROTEIN G1
	PANTHER	PTH46617-SF3	FORKHEAD BOX PROTEIN G1
	PANTHER	PTH46721	FORKHEAD BOX PROTEIN N1
	PANTHER	PTH46721-SF2	FORKHEAD BOX N1
	PANTHER	PTH46805	FORKHEAD BOX PROTEIN J1
	PANTHER	PTH46878	FORKHEAD BOX PROTEIN M1
	PANTHER	PTH46878-SF1	FORKHEAD BOX PROTEIN M1
	PANTHER	PTH47316	FORKHEAD BOX PROTEIN H1
	PANTHER	PTH47316-SF1	FORKHEAD BOX PROTEIN H1

Supplementary Table S3 – List of DSFGs from reference species used to assess the identity of DSFGs in molluscs. NCBI accession numbers are reported in parenthesis. Each row represents an orthology group.

Homo sapiens	Drosophila melanogaster	Caenorhabditis elegans	Group
Dmrt gene family			
<i>DMRT1</i> (NP_068770.2)	-	-	1
<i>DMRT2</i> (NP_006548.1)	<i>dmrt11E</i> (NP_511146.2)	-	2
<i>DMRT3</i> (NP_067063.1)	<i>dmrt93B</i> (NP_524428.1)	<i>dmd-4</i> (NP_510466.1)	3
<i>DMRT4/A1</i> (NP_071443.2)	<i>dmrt99b</i> (NP_524549.1)	<i>dmd-5</i> (NP_495138.2)	A1/2
<i>DMRT5/A2</i> (NP_115486.1)			
<i>DMRT6/B1</i> (NP_149056.1)	-	-	-
<i>DMRT7/C2</i> (NP_001035373.1)	-	-	-
<i>DMRT8/C1</i> (NP_149042.2)	-	-	-
-	<i>dsx</i> (NP_731197.1)	-	-
-	-	<i>mab3</i> (NP_001256882.1)	-
-	-	<i>dmd-3</i> (NP_001256883.1)	-
-	-	<i>dmd-6</i> (NP_001370045.1)	-
-	-	<i>dmd-7</i> (NP_741551.1)	-
-	-	<i>dmd-8</i> (NP_503176.2)	-
-	-	<i>dmd-9</i> (NP_500305.1)	-
-	-	<i>dmd-11</i> (NP_001379162.1)	-
-	-	<i>mab-23</i> (NP_001041089.1)	-
Sox gene family			
<i>SRY</i> (NP_003131.1)	-	-	A
<i>SOX3</i> (NP_005625.2)			
<i>SOX2</i> (NP_003097.1)	<i>dichaete</i> (NP_524066.1)	<i>sox3</i> (NP_510439.1)	B1
<i>SOX1</i> (NP_005977.2)	<i>soxN</i> (NP_524735.1)	<i>sox2</i> (NP_741836.1)	
<i>SOX14</i> (NP_004180.1)	<i>sox21a</i> (NP_648694.1)		B2
<i>SOX21</i> (NP_009015.1)	<i>sox21b</i> (NP_648695.1)		
<i>SOX11</i> (NP_003099.1)			
<i>SOX12</i> (NP_008874.2)	<i>sox14</i> (NP_476894.1)	<i>sem-2</i> (NP_740846.1)	C
<i>SOX4</i> (NP_003098.1)			
<i>SOX13</i> (NP_005677.2)			
<i>SOX5</i> (NP_008871.3)	<i>sox10f</i> (NP_726612.1)	<i>egl-13</i> (NP_001024918.1)	D
<i>SOX6</i> (NP_001139291.2)			
<i>SOX9</i> (NP_000337.1)			
<i>SOX8</i> (NP_055402.2)	<i>sox110b</i> (NP_651839.1)	-	E
<i>SOX10</i> (NP_008872.1)			
<i>SOX18</i> (NP_060889.1)			
<i>SOX7</i> (NP_113627.1)	<i>sox15</i> (NP_523739.2)	-	F
<i>SOX17</i> (NP_071899.1)			
<i>SOX15</i> (NP_008873.1)	-	-	G
<i>SOX30</i> (NP_848511.1)	-	-	H
Fox gene family			
<i>FOXA1/HNF-3α</i> (NP_004487.2)			
<i>FOXA2/HNF-3β</i> (NP_068556.2)	<i>forkhead/fkh</i> (NP_524542.1)	<i>pha-4/Ce-fkh1</i> (NP_001041114.1)	A
<i>FOXA3/HNF-3γ</i> (NP_004488.2)			
<i>FOXB1</i> (NP_036314.2)			
<i>FOXB2</i> (NP_001013757.1)	<i>fd96Ca/fd4</i> (NP_524495.1) <i>fd96Cb/fd5</i> (NP_524496.1)	<i>lin-31</i> (NP_494704.1)	B
<i>FOXC1/MF1/FKHL7</i> (NP_001444.2)			
<i>FOXC2/MFH1</i> (NP_005242.1)	<i>crocodile/fd1</i> (NP_524202.1)	-	C
<i>FOXD1/FREAC4</i> (NP_004463.1)			
<i>FOXD2/FREAC9</i> (NP_004465.3)			
<i>FOXD3</i> (NP_036315.1)	<i>fd59A/fd3</i> (NP_523814.1)	<i>unc-130</i> (NP_496411.1)	D
<i>FOXD4</i> (NP_997188.2)			
<i>FOXE1/TITF2</i> (NP_004464.2)	-	-	E
<i>FOXE3</i> (NP_036318.1)			
<i>FOXF1</i> (NP_001442.2)			
<i>FOXF2</i> (NP_001443.1)	<i>binious/FoxF</i> (NP_523950.2)	<i>let-381/F26B1.7</i> (NP_491826.1)	F
<i>FOXG1/BF1/HBF2</i> (NP_005240.3)	<i>slp1</i> (NP_476730.1) <i>slp2</i> (NP_476834.1) <i>fd19B/cg9571</i> (NP_608369.1)	<i>fkh2/T14G12.4</i> (NP_508644.1)	G
<i>FOXH1/FAST1</i> (NP_003914.1)	-	-	H
<i>FOXI1/FREAC6/HFH3</i> (NP_036320.2)	-	-	I
<i>FOXJ1</i> (NP_001445.2)	-	-	J1
<i>FOXJ2</i> (XP_011519063.1)	-	-	J2
<i>FOXJ3</i> (XP_005270689.1)	-	-	J3
<i>FOXK1/LF1</i> (NP_001032242.1)	<i>foxK/LD16137</i> (NP_001261701.1)	-	K
<i>FOXK2</i> (NP_004505.2)			
<i>FOXL1</i> (NP_005241.1)	<i>foxL1/fd2</i> (NP_523912.1)	-	L1

Tab. S3 continued from previous page

Homo sapiens	Drosophila melanogaster	Caenorhabditis elegans	Group
Fox gene family			
<i>FOXL2</i> (NP_075555.1)	-	-	L2
<i>FOXM1</i> (NP_001400854.1)	-	-	M
<i>FOXN1/WHN</i> (NP_001356298.1)	<i>jumeau</i> (NP_524302.1)	-	N1/4
<i>FOXN4</i> (NP_998761.2)			
<i>FOXN2/HTLF</i> (NP_001362376.1)	<i>ches-1</i> (NP_511071.3)	-	N2/3
<i>FOXN3/CHES1</i> (NP_001078940.1)			
<i>FOXO1</i> (NP_002006.2)			
<i>FOXO3</i> (NP_963853.1)	-	<i>daf-16</i> (NP_001364785.1)	O
<i>FOXO3B</i> (NP_001355064.1)			
<i>FOXP1</i> (NP_001231739.1)			
<i>FOXP2</i> (NP_683696.2)			
<i>FOXP3</i> (NP_054728.2)	<i>foxP/cg16899</i> (NP_001247011.1)	<i>F26D12.1</i> (NP_001293813.1)	P
<i>FOXP4</i> (XP_011512591.1)			
<i>FOXQ/HFH11</i> (NP_150285.3)	-	-	Q1
-	<i>fd102C/cd11152</i> (NP_651951.1)	<i>fkh-10/C25A1.2</i> (NP_492676.2)	Q2
<i>FOXS1/FREAC10</i> (NP_004109.1)	-	-	S
-	-	<i>PES-1</i> (NP_001023406.1)	-
-	-	<i>B0286.5/FKH-6</i> (NP_494775.1)	-
-	-	<i>F40H3.4/FKH-8</i> (NP_001254107.1)	-
-	-	<i>C29F7.4/FKH-3</i> (NP_001294822.1)	-
-	-	<i>K03C7.2/FKH-9</i> (NP_001024760.1)	-

Supplementary Table S4 – Genomic data of mammals used to retrieve DSFGs and compute AASD of SCOs. For each species, the relative ID, taxonomic information, BUSCO statistics, NCBI accession number, and source publication are reported.

Species	ID	Class	Group	Order	Type	BUSCO statistics ('mammalia.ncbi10')	NCBI acc. no.	Reference
<i>Gallus gallus</i>	Ggal	Aves	Neognathae	Galliformes	Genome	C:99.0%[S:98.6%,D:0.4%],[F:0.2%,M:0.8%]	GCF_01669485.2	Vertebrate Genome Project
<i>Clivochilis asiatica</i>	Casi	Mammalia	Astrotheria	Afroscordida	Genome	C:98.0%[S:97.4%,D:0.6%],[F:1.1%,M:0.9%]	GCF_000296735.1	Murata et al., 2003
<i>Elephas maximus indicus</i>	Emax	Mammalia	Astrotheria	Proboscidea	Genome	C:98.8%[S:98.3%,D:0.6%],[F:0.4%,M:0.7%]	GCF_024166365.1	Vertebrate Genome Project
<i>Trichechus manatus latirostris</i>	Tman	Mammalia	Astrotheria	Sirenia	Genome	C:96.1%[S:95.7%,D:0.4%],[F:1.8%,M:2.1%]	GCF_000243295.1	Foote et al., 2015
<i>Ocypterus afer afer</i>	Oafe	Mammalia	Euarchontoglires	Tubulidentata	Genome	C:96.5%[S:96.0%,D:0.5%],[F:1.9%,M:1.6%]	GCF_000298275.1	–
<i>Ochotona princeps</i>	Opri	Mammalia	Euarchontoglires	Lagomorpha	Genome	C:98.3%[S:96.4%,D:1.9%],[F:0.5%,M:1.2%]	GCF_03043755.1	Vertebrate Genome Project
<i>Cebus imitator</i>	Cimi	Mammalia	Euarchontoglires	Primates	Genome	C:97.3%[S:95.1%,D:2.2%],[F:1.7%,M:1.0%]	GCF_001604975.1	Orkin et al., 2021
<i>Homo sapiens</i>	Hsap	Mammalia	Euarchontoglires	Primates	Genome	C:99.6%[S:97.3%,D:2.3%],[F:0.2%,M:0.2%]	GCF_000001405.40	Genome Reference Consortium
<i>Lemur catta</i>	Lcat	Mammalia	Euarchontoglires	Rodentia	Genome	C:98.3%[S:97.2%,D:1.1%],[F:0.4%,M:1.3%]	GCF_00240605.2	Vertebrate Genome Project
<i>Cavia porcellus</i>	Cpor	Mammalia	Euarchontoglires	Rodentia	Genome	C:96.4%[S:95.7%,D:0.7%],[F:1.7%,M:1.9%]	GCF_000151735.1	The Genome Sequencing Platform
<i>Mus musculus</i>	Mmus	Mammalia	Euarchontoglires	Rodentia	Genome	C:99.4%[S:98.7%,D:1.7%],[F:0.2%,M:0.4%]	GCF_000001635.27	Genome Reference Consortium
<i>Sciurus carolinensis</i>	Scar	Mammalia	Euarchontoglires	Rodentia	Genome	C:99.1%[S:96.9%,D:2.2%],[F:0.3%,M:0.6%]	GCF_002686445.1	Mead et al., 2020
<i>Bubalus bubalis</i>	Bbub	Mammalia	Eurasiatheria	Artiodactyla	Genome	C:98.7%[S:97.0%,D:1.7%],[F:0.6%,M:0.7%]	GCF_019923935.1	Deng et al., 2016
<i>Balaenoptera musculus</i>	Bmus	Mammalia	Eurasiatheria	Artiodactyla	Genome	C:98.4%[S:95.7%,D:2.7%],[F:0.6%,M:1.0%]	GCF_00987245.2	Genome 10K
<i>Camelus dromedarius</i>	Cdro	Mammalia	Eurasiatheria	Artiodactyla	Genome	C:98.7%[S:96.3%,D:0.4%],[F:0.7%,M:0.6%]	GCF_00008125.2	Elbers et al., 2019
<i>Hippopotamus amphibius kiboko</i>	Hamp	Mammalia	Eurasiatheria	Artiodactyla	Genome	C:98.7%[S:95.2%,D:3.5%],[F:0.5%,M:0.8%]	GCF_0030028045.1	Vertebrate Genome Project
<i>Phacochoerus africanus</i>	Paf	Mammalia	Eurasiatheria	Artiodactyla	Genome	C:98.8%[S:98.3%,D:0.5%],[F:0.6%,M:0.6%]	GCF_016906955.1	–
<i>Tursiops truncatus</i>	Trtu	Mammalia	Eurasiatheria	Artiodactyla	Genome	C:97.3%[S:95.2%,D:2.1%],[F:1.1%,M:1.6%]	GCF_011762505.1	Xiong et al., 2009
<i>Aluterops melanoleucus</i>	Amel	Mammalia	Eurasiatheria	Carnivora	Genome	C:97.3%[S:96.6%,D:0.7%],[F:1.3%,M:1.4%]	GCF_00207445.2	Fan et al., 2019
<i>Canis lupus familiaris</i>	Clup	Mammalia	Eurasiatheria	Carnivora	Genome	C:98.5%[S:96.7%,D:1.8%],[F:0.6%,M:0.9%]	GCF_011006635.1	Wang et al., 2021
<i>Mirounga angustirostris</i>	Mang	Mammalia	Eurasiatheria	Carnivora	Genome	C:96.7%[S:94.5%,D:2.2%],[F:1.9%,M:1.4%]	GCF_021288785.2	Morenlo et al., 2024
<i>Panthera tigris</i>	Ptig	Mammalia	Eurasiatheria	Carnivora	Genome	C:99.4%[S:98.9%,D:0.5%],[F:0.6%,M:0.3%]	GCF_018350195.1	Biedermann et al., 2023
<i>Desmodus rotundus</i>	Drot	Mammalia	Eurasiatheria	Chiroptera	Genome	C:98.2%[S:97.2%,D:1.0%],[F:0.5%,M:1.3%]	GCF_02262495.1	Bat 1K
<i>Pteropus giganteus</i>	Pgig	Mammalia	Eurasiatheria	Chiroptera	Genome	C:97.2%[S:96.9%,D:0.3%],[F:1.1%,M:1.7%]	GCF_002729225.1	Fouret et al., 2020
<i>Rhinolophus ferrumequinum</i>	Rfer	Mammalia	Eurasiatheria	Perissodactyla	Genome	C:99.2%[S:97.9%,D:1.3%],[F:0.3%,M:0.5%]	GCF_004115265.2	Vertebrate Genome Project
<i>Cetotherium simum simum</i>	Csim	Mammalia	Eurasiatheria	Perissodactyla	Genome	C:98.8%[S:96.6%,D:0.2%],[F:0.6%,M:0.3%]	GCF_000283155.1	–
<i>Equus quagga</i>	Equa	Mammalia	Eurasiatheria	Pholidota	Genome	C:98.5%[S:95.0%,D:3.5%],[F:0.5%,M:1.0%]	GCF_02161305.1	Vilstrup et al., 2013
<i>Manis javanica</i>	Mjav	Mammalia	Mammalia	Dasyuromorphia	Genome	C:95.7%[S:93.7%,D:2.0%],[F:1.9%,M:2.4%]	GCF_014570535.1	–
<i>Sarcophilus harriii</i>	Shar	Mammalia	Mammalia	Didelphimorphia	Genome	C:95.5%[S:94.5%,D:1.0%],[F:0.5%,M:1.3%]	GCF_00263505.1	Stammnitz et al., 2023
<i>Monodelphis domestica</i>	Mdom	Mammalia	Mammalia	Metatheria	Genome	C:95.1%[S:92.3%,D:2.8%],[F:0.9%,M:3.6%]	GCF_002787165.1	Vertebrate Genome Project
<i>Omittorhynchus anatinus</i>	Qana	Mammalia	Mammalia	Prototheria	Genome	C:92.3%[S:91.2%,D:1.1%],[F:1.4%,M:6.3%]	GCF_00415215.2	Zhou et al., 2021
<i>Dasyurus novemcinctus</i>	Dnov	Mammalia	Mammalia	Xenarthra	Genome	C:96.9%[S:94.3%,D:2.6%],[F:0.7%,M:2.4%]	GCF_030445035.1	Vertebrate Genome Project
<i>Choloepus didactylus</i>	Cdid	Mammalia	Mammalia	Xenarthra	Genome	C:97.8%[S:91.9%,D:5.9%],[F:0.7%,M:1.5%]	GCF_015220235.1	Vertebrate Genome Project

Species	ID	Family	Subgenus	Type	BUSCO statistics ('diptera_odb10')	NCBI acc. no.	Reference
<i>Anopheles gambiae</i>							
<i>Drosophila sechellia</i>	Agam	Culicidae	Cellia	Genome	C:99.4%[S:99.1%,D:0.3%],F:0.1%,M:0.5%	GCF_943734735.2	Habitewold et al., 2023
<i>Drosophila melanogaster</i>	Dsec	Drosophilidae	Sophophora	Genome	C:99.9%[S:99.3%,D:0.6%],F:0.0%,M:0.1%	GCF_004382195.2	Chakraborty et al., 2021
<i>Drosophila erecta</i>	Dmel	Drosophilidae	Sophophora	Genome	C:100.0%[S:99.7%,D:0.3%],F:0.0%,M:0.0%	GCF_000001215.4	Hoskins et al., 2015
<i>Drosophila bipunctinata</i>	Dere	Drosophilidae	Sophophora	Genome	C:99.9%[S:99.5%,D:0.4%],F:0.0%,M:0.1%	GCF_003286155.1	dong2022new ““textit{-Drosophila suzukii” & Dsuz
<i>Drosophila ananassae</i>	Dbip	Drosophilidae	Sophophora	Genome	C:99.9%[S:99.2%,D:0.7%],F:0.0%,M:0.1%	GCF_018153845.1	Kim et al., 2021
<i>Drosophila pseudoobscura</i>	Dana	Drosophilidae	Sophophora	Genome	C:99.6%[S:99.3%,D:0.3%],F:0.0%,M:0.4%	GCF_017639315.1	Tvedte et al., 2021
<i>Drosophila miranda</i>	Dpose	Drosophilidae	Sophophora	Genome	C:99.7%[S:98.8%,D:0.9%],F:0.1%,M:0.2%	GCF_009870125.1	Liao et al., 2021
<i>Drosophila willistoni</i>	Dmir	Drosophilidae	Sophophora	Genome	C:99.8%[S:98.6%,D:1.2%],F:0.1%,M:0.1%	GCF_003369915.1	Mahajan et al., 2018
<i>Drosophila arizoneae</i>	Dwil	Drosophilidae	Sophophora	Genome	C:99.6%[S:98.4%,D:1.2%],F:0.0%,M:0.4%	GCF_018902025.1	Ranz et al., 2023
<i>Drosophila hydei</i>	Dari	Drosophilidae	Drosophila	Genome	C:95.7%[S:95.3%,D:0.4%],F:1.2%,M:3.1%	GCF_001654025.1	Sanchez-Flores et al., 2016
<i>Drosophila grimshawi</i>	Dhyd	Drosophilidae	Drosophila	Genome	C:99.7%[S:97.5%,D:2.2%],F:0.1%,M:0.2%	GCF_0032285905.1	Dong et al., 2022
<i>Drosophila albomicans</i>	Dgri	Drosophilidae	Drosophila	Genome	C:99.9%[S:99.2%,D:0.7%],F:0.0%,M:0.1%	GCF_018153295.1	Kim et al., 2021
<i>Drosophila busckii</i>	Dalb	Drosophilidae	Drosophila	Genome	C:99.9%[S:99.1%,D:0.8%],F:0.0%,M:0.1%	GCF_009650485.2	Mai et al., 2020
	Dbus	Drosophilidae	Drosophila	Genome	C:98.1%[S:97.4%,D:0.7%],F:0.3%,M:1.6%	GCF_011750605.1	Renschler et al., 2019

Supplementary Table S6 – Complete set of DSFGs in bivalves. For each gene, the species ID (Sp. ID) as in Tab. S1, the accession number (Gene ID), the Possm-based annotation, and the CDD domains (including their Psm-ID) are indicated.

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Psm-ID)	Additional domains (Psm-ID)	Notes
Airc	Contig6_279	Dmrt	Dmrt-OGO/NA	Doublesex DNA-binding motif (214606)	-	Annotated as Dmrt-1L
	scaffid_235_403	Dmrt	Dmrt-OGO/NA	Doublesex DNA-binding motif (214606)	-	Annotated as Dmrt-1L
Apur	XP_052698046.1	Dmrt	Dmrt-OGO/NA	Doublesex DNA-binding motif (214606)	-	Annotated as Dmrt-1L
Cang	EVMO0027346.1	Dmrt	Dmrt-OGO/NA	Doublesex DNA-binding motif (214606)	-	Annotated as Dmrt-1L
Carl	XP_001441049.2	Dmrt	Dmrt-OGO/NA	Doublesex DNA-binding motif (214606)	-	Annotated as Dmrt-1L
Cgig	XP_022333988.1	Dmrt	Dmrt-OGO/NA	Doublesex DNA-binding motif (214606)	-	Annotated as Dmrt-1L
Cvir	XP_041358115.1	Dmrt	Dmrt-OGO/NA	Doublesex DNA-binding motif (214606)	-	Annotated as Dmrt-1L
Gaeq	M00000013645	Dmrt	Dmrt-OGO/NA	Doublesex DNA-binding motif (214606)	-	Annotated as Dmrt-1L
Hbia	M00000045261	Dmrt	Dmrt-OGO/NA	Doublesex DNA-binding motif (214606)	-	Annotated as Dmrt-1L
Hbifia	XP_046372338.2	Dmrt	Dmrt-OGO/NA	Doublesex DNA-binding motif (214606)	-	Annotated as Dmrt-1L
Hruf	XP_046335704.1	Dmrt	Dmrt-OGO/NA	Doublesex DNA-binding motif (214606)	-	Annotated as Dmrt-1L
Hruf	XP_052068518.1	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Mcal	CAC3397186.1	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Mcor	CAG22320556.1	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Medu	VD103798.1	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606; partial)
Migal	MMAN00000008302	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Mimar	g120437_t1	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Mner	XP_0147782618.1	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Obim	XP_048736174.2	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Oedu	XP_036368911.1	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Osin	XP_036366646.1	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Osin	XP_0296477012	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Pcan	XP_025090051.1	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Pcan	XP_025111744.1	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Pmar	DN306577_0.g1.i1.p1	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Pmar	XP_033733655.1	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Pmar	Ppur	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Pstr	KAK359948.1	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606; partial)
Pvir	s01850g168	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606; partial)
Pyes	XP_021353714.1	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Sbro	EVM0020695.1	Dmrt	Dmrt-OG4/NA	Doublesex DNA-binding motif (214606)	-	Doublesex DNA-binding motif (214606)
Sgio	Sgi011295	Dmrt	Dmrt-2	Dmrt-2	-	-
Arc	Contig172_94	Dmrt	Dmrt-2	Dmrt-2	-	-
Apcc	DN1321_0.g1.i1.p1	Dmrt	Dmrt-2	Dmrt-2	-	-
Bgia	XP_053887190.1	Dmrt	Dmrt-2	Dmrt-2	-	-
Cphi	DN116454_c0.g1.i1.p1	Dmrt	Dmrt-2	Dmrt-2	-	-
Gaeq	XP_041359971.1	Dmrt	Dmrt-2	Dmrt-2	-	-
Hruf	XP_0482532484.1	Dmrt	Dmrt-2	Dmrt-2	-	-
Mcor	CAC5404148.1	Dmrt	Dmrt-2	Dmrt-2	-	-
Medu	CAG223205366.1	Dmrt	Dmrt-2	Dmrt-2	-	-
Mgal	VD142071.1	Dmrt	Dmrt-2	Dmrt-2	-	-
Mmer	XP_045156965.2	Dmrt	Dmrt-2	Dmrt-2	-	-
Mphi	scaf_304771.9	Dmrt	Dmrt-2	Dmrt-2	-	-
Obim	XP_052832484.1	Dmrt	Dmrt-2	Dmrt-2	-	-
Osin	XP_029650766.2	Dmrt	Dmrt-2	Dmrt-2	-	-
Pcan	DN105632_c0.g1.i1.p1	Dmrt	Dmrt-2	Dmrt-2	-	-
Pcor	DN115233_c0.g1.i1.p1	Dmrt	Dmrt-2	Dmrt-2	-	-
Pcor	DN28033_c0.g1.i1.p1	Dmrt	Dmrt-2	Dmrt-2	-	-
Pmax	XP_0337338864.1	Dmrt	Dmrt-2	Dmrt-2	-	-
Pstr	KAK3603675.1	Dmrt	Dmrt-2	Dmrt-2	-	-
Pvir	s0097g235	Dmrt	Dmrt-2	Dmrt-2	-	-
Pyes	XP_021368788.1	Dmrt	Dmrt-2	Dmrt-2	-	-
Rphi	XP_060589226.1	Dmrt	Dmrt-2	Dmrt-2	-	-
Sbro	EVM001645.1	Dmrt	Dmrt-2	Dmrt-2	-	-
Scon	Ch8_1365	Dmrt	Dmrt-2	Dmrt-2	-	-
Tgra	KAJ83306274.1	Dmrt	Dmrt-2	Dmrt-2	-	-
Tsqu	DN51730_c0.g1.i1.p1	Dmrt	Dmrt-2	Dmrt-2	-	-
Acal	XP_05096932.1	Dmrt	Dmrt-3	Dmrt-3	-	-
Acal	Contig349_40	Dmrt	Dmrt-3	Dmrt-3	-	-
Amar	Ama12564	Dmrt	Dmrt-3	Dmrt-3	-	-
Apur	scaff_9576	Dmrt	Dmrt-3	Dmrt-3	-	-
BglA	XP_03077145.1	Dmrt	Dmrt-3	Dmrt-3	-	-
Cang	XP_052687934.1	Dmrt	Dmrt-3	Dmrt-3	-	-

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (P _{ssm-ID})	Additional domains (P _{ssm-ID})	Notes
Cari	EVMO028466.1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Cegig	XP_001427033.2	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Cph	DN37429_0.g2.i1.p1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Csin	Hic-asn..11.1r74	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Cvir	XP_022317913.1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Dpol	XP_052246678.1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Gaeg	XP_041357684.1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Hbia	M0000034631	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Hruf	XP_046367747.2	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mare	XP_052777877.1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mcal	XP_052076466.1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mcor	CAC5360634.1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Medu	CAG2226664.1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mgal	VD32052.1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mmar	MMA/M0000039146	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mmer	XP_045157038.1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mner	g243_052_t1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mphi	seaf2165_0.7	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Oedu	XP_0487161857.1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Pcan	XP_025110327.1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Pcor	DN33000_50.g1.i1.p1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Pmax	XP_033737545.1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Pvir	s00097g272	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Rph1	XP_060576882.1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Sbro	EVMO006488.1	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Scor	Ch8_2435	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Sgio	Sej014397	Dmrt	Dmrt-3	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Tgra	KAJ8305800.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Acal	XP_05096931.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Airc	Contig349_42	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Amar	Ama12441	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Amar	Ana12902	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Apuc	DN10969_dog1.i1.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Apur	scaffid95_77	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Bglia	XP_013088904.2	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Cang	XP_052685010.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Cari	EVMO0084823.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Cegig	NP_001298834.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Cphi	DN67594_0.g1.i1.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Cphi	DN55944_0.g1.i1.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Cphi	DN54583_0.g1.i1.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Cphi	DN9274_0.g1.i1.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Cphi	DN67594_0.g2.i1.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Cphi	Hic-asn..11.400	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Csin	Hic-asn..11.338	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Cvir	XP_022319926.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Dpol	XP_052255215.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Dpol	XP_052224782.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Dpol	XP_052257495.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Gaeig	XP_041357898.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Hbia	M0000038945	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Hbia	M0000038946	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Hbia	M0000019813	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Hbia	M0000038947	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Hruf	XP_046367734.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Lorb	DN147341_c0.g1.i1.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Lorb	DN40542_0.g1.i1.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mare	XP_052776891.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mare	XP_052776906.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mare	XP_052776885.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mcal	XP_052092340.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mchi	DN35166_d0.g1.i1.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mcor	CAC5398878.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Medu	CAG2209978.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mgal	VD124477.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mmar	MMAM/M000000040488	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	
Mmar	MMAM/M000000003515	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270600)	

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (P _{ssm-ID})	Additional domains (P _{ssm-ID})	Notes
Mmar	MMAN00000016566	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270602)	
Mmar	MMAN00000047004	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270602)	
Mmar	MMAN00000044361	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270602)	
Mmer	XP_045157593.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Mmer	XP_045157053.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Mmer	XP_045159713.2	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Mmed	DN16828_c0.g1.i1.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Mmer	g346_03.tl	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602)	
Mmer	g585_31.tl	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Mmer	g241_174.tl	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Mmer	g192_820.tl	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Mphi	scf-68796_0.3	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Oedu	XP_048763391.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602, 270465)	
Pcan	XP_025110328.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Pcor	DN12587_c0.g13.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602)	
Pcor	DN1623_c0.g1.i13.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602)	
Pcor	DN15587_c0.g1.i4.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270465)	
Pgen	DN68344_c0.g1.i1.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270465)	
Pgen	XP_033737544.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Pmax	DN35556_c0.g1.i2.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Poku	DN35178_c0.g1.i2.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Pstr	KAK3612677.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Pstr	KAK388310.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Pstr	KAK3883112.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Pstr	KAK3883109.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Pvir	s0073382	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Pyes	XP_021377274.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270465)	
Rdec	DN26973_c0.g1.i1.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Rphi	XP_060600638.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600)	
Rphi	XP_060578674.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602)	
Sbro	EVM0004355.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270602)	
Scon	Ch8-1999	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270465)	
Scon	Ch8-2143	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270465)	
Sglo	Sg106992	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270465)	
Sgra	DN54078_c0.g1.i1.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270465)	
Sgra	DN66569_c0.g1.i3.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270465)	
Tgra	KAJ3305799.1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270465)	
Tsequ	DN75749_c0.g1.i1.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270465)	
Apec	DN8372_c0.g4.i1.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270465)	
Mchi	DN34711_c0.g1.i2.p1	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270465)	
Scon	Ch8-738	Dmrt	Dmrt-4/5	Doublesex DNA-binding motif (214606)	CUE-like DMA domain (270553, 270601, 270600, 270465)	
Acal	XP_050597243.2	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Airc	Contieg_157	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Amar	Ama08751	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Apec	DN107972_c0.g1.i1.p1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Apur	scaffold124.7	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Brg1	XP_03067134.2	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Cang	XP_0527025.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Cari	EVM0004613.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Cflu	DN101169_c0.g1.i1.p1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Crig	XP_01413445.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Cpli	DN47094_c0.g4.i1.p1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Csin	Hic_asn_10.437	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Csin	XP_022333322.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Cvir	XP_022334050.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Dpol	XP_0522272379.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Gaeq	XP_04352484.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
M00000018167	Fox	Fox-A	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Hbia	XP_046371021.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Mare	XP_052769228.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Mccl	XP_052106467.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Mchi	DN35955_c0.g4.i1.p1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Mcor	CAC5374046.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Medu	CAG2201348.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Psm-ID)	Additional domains (Psm-ID)	Notes
Mgar	VD11457.1	Fox	Fox-A	Forkhead domain A1 (410812)	-	
Mmar	MMAM00000008663	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552; partial)	
Mmer	XP_045173733.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552; partial)	
Mmod	DN103780.c0.g.i1.p1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872; partial); HNF3 C-terminal domain (430552)	
Mmer	g192217.tl	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872; partial); HNF3 C-terminal domain (430552)	
Mphi	scaf4682.0	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal A1 (410812)	
Obin	XP_014788201.2	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872)	
Oedu	XP_048735259.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552; partial)	
Pcan	XP_025090786.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872; partial); HNF3 C-terminal domain (430552)	
Pcor	DN3402.c0.g.i1.p1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872; partial)	
Pgen	DN174637.c0.g.i1.p1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872; partial)	
Pmar	DN30866.c0.g.i1.p1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872; partial)	
Pmax	XP_033734080.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Pstr	KAK3597847.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552; partial)	
Pvir	XP_021361791.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Pyes	XP_060590755.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Rphi	EVMM0003194.1	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Sbro	Ch4-2670	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Scor	Sg008464	Fox	Fox-A	Forkhead domain A1 (410812)	Forkhead N-terminal (369872); HNF3 C-terminal domain (430552)	
Sgra	DN705052.c0.g.i1.p1	Fox	Fox-B	Forkhead domain A1 (410812)	Forkhead N-terminal (369872; partial); HNF3 C-terminal domain (430552)	
Acal	XP_005089018.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Airc	Contig636.38	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Aupr	scaffold313.50	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Bglj	XP_013078204.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Cang	XP_0527003533.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Cari	EVMM0003536.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Cflu	DN98613.c0.g.i1.p1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Cgig	XP_011445364.2	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Csin	Hic-asrn.16.1347	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Cvir	XP_022334612.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Dpol	XP_052233250.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Dpol	XP_052256324.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Dpol	XP_052281977.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Gaeg	XP_041361159.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Hbia	M000000209836	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Hruf	XP_045358590.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Lorb	DN5589.c3.g.i1.p1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Mare	XP_052791461.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Mcal	XP_052100219.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Mcor	CAC5382565.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Medu	CAG2229716.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Mgar	MMAM00000015629	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Mmer	scaf10920.0	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Mphi	XP_052832317.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Obim	XP_048732871.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Oedu	DN23979.c0.g.i1.p1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Pcor	XP_033749587.1	Fox	Fox-B	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Pmax	KAK3607900.1	Fox	Fox-C	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Pstr	s001399g215	Fox	Fox-C	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Pvir	XP_021357620.1	Fox	Fox-C	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Rphi	XP_060564412.1	Fox	Fox-C	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Scor	Ch5-12	Fox	Fox-C	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Sgra	Sg012012	Fox	Fox-C	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Tgra	KAJ3304921.1	Fox	Fox-C	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Acal	XP_005106277.1	Fox	Fox-C	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Airc	Contig58.63	Fox	Fox-C	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Aupr	Amar	Fox	Fox-C	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Bglj	scaf1094	Fox	Fox-C	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Cang	XP_055890240.1	Fox	Fox-C	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Cari	XP_052715579.1	Fox	Fox-C	Forkhead domain B2 (410817)	Forkhead N-terminal (369872; partial)	
Cflu	DN95576.c0.g.i1.p1	Fox	Fox-C	Forkhead domain C (410795)	Forkhead N-terminal (369872; partial)	

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Pssm-ID)	Additional domains (Pssm-ID)	Notes
Cggg	XP_011417585.2	Fox	Fox-C	Forkhead domain Li	(410801)	
Cpli	DN157725.c0.g1.i1.p1	Fox	Fox-C	Forkhead domain Li	(410801)	
Csin	Hic.asrn.17.1387	Fox	Fox-C	Forkhead domain Li	(410801)	
Csin	Hic.asrn.17.1443	Fox	Fox-C	Forkhead domain Li	(410801)	
Cvir	XP_022346235.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Dpol	XP_052247387.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Gaeg	XP_041377097.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Hbia	M0000031058	Fox	Fox-C	Forkhead domain Li	(410801)	
Hruf	XP_046372770.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Mare	XP_052819073.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Mcal	XP_052063314.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Mchi	DN13809.c0.g1.i1.p1	Fox	Fox-C	Forkhead domain Li	(410801)	
Mcor	CAC5374004.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Medu	CAG2206844.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Mgal	VD122482.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Mmar	MMAN00000355616	Fox	Fox-C	Forkhead domain Li	(410801)	
Mmer	XP_045194706_2	Fox	Fox-C	Forkhead domain Li	(410801)	
Mner	g82158.t1	Fox	Fox-C	Forkhead domain Li	(410801)	
Mphi	scf.69501.0	Fox	Fox-C	Forkhead domain Li	(410801)	
Obim	XP_04748040.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Oedu	XP_048762038.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Osin	XP_029653806.2	Fox	Fox-C	Forkhead domain Li	(410801)	
Pcan	XP_025115697.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Pcor	DN14158.c0.g2.i1.p1	Fox	Fox-C	Forkhead domain Li	(410801)	
Pcor	DN14158.c0.g5.i1.p1	Fox	Fox-C	Forkhead domain Li	(410801)	
Pmax	XP_033755061.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Pstr	KAK3590993.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Pvir	s020238g12	Fox	Fox-C	Forkhead domain Li	(410801)	
Pyes	XP_021346967.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Rphi	XP_060597004.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Sbro	EVM0022192.1	Fox	Fox-C	Forkhead domain Li	(410801)	
Scor	Chr1.448	Fox	Fox-C	Forkhead domain Li	(410801)	
Sglo	Sg1009485	Fox	Fox-C	Forkhead domain Li	(410801)	
Tgra	KAJ3303551.1	Fox	Fox-C	Forkhead domain C2	(410819)	
Acal	XP_033824261.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Airc	Contig1003.15	Fox	Fox-D	Forkhead domain D4	(410822)	
Amar	Ama11686	Fox	Fox-D	Forkhead domain D3	(410821)	
Apc	DN8782.c0.g1.i1.p1	Fox	Fox-D	Forkhead domain D4	(410822)	
Apur	scffold13962.11	Fox	Fox-D	Forkhead domain D4	(410822)	
Bglia	XP_013096936.2	Fox	Fox-D	Forkhead domain D4	(410822)	
Cang	XP_052688370.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Cari	EVM0005770.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Ceig	XP_011446328.2	Fox	Fox-D	Forkhead domain D4	(410822)	
Cphl	DN233774.c0.g1.i1.p1	Fox	Fox-D	Forkhead domain D4	(410822)	
Csin	Hic.asrn.11.1425	Fox	Fox-D	Forkhead domain D4	(410822)	
Cvir	XP_022346146.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Dpol	XP_052256038.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Dpol	XP_052256569.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Gaeg	XP_041356731.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Hbia	M0000030583	Fox	Fox-D	Forkhead domain D4	(410822)	
Hruf	XP_046329290.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Lorb	DN224803.c0.g1.i1.p1	Fox	Fox-D	Forkhead domain D4	(410822)	
Mare	XP_052777467.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Mcal	XP_052777725.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Mcal	XP_052096202.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Meal	XP_052075808.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Mgal	CAC5382691.1	Fox	Fox-D	Forkhead domain D3	(410821)	
Mgal	VDH93066.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Mcor	CAC5407497.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Medu	CAG2204666.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Medu	CAG2248150.1	Fox	Fox-D	Forkhead domain D4	(410822)	
CAG2203862.1		Fox	Fox-D	Forkhead domain D4	(410822)	
Meal	VDI07735.1	Fox	Fox-D	Forkhead domain D4	(410822)	
Mgal	MMAN0000002467	Fox	Fox-D	Forkhead domain D4	(410822)	
Mmar	XP_045157233.2	Fox	Fox-D	Forkhead domain D4	(410822)	
Mmer	g192986.t1	Fox	Fox-D	Forkhead domain D4	(410822)	
Mphi	scf.69491.12	Fox	Fox-D	Forkhead domain D4	(410822)	
Mphi	scf.42856.0.4	Fox	Fox-D	Forkhead domain D4	(410822)	
Obim	XP_052826256.1	Fox	Fox-D	Forkhead domain D4	(410822)	

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Pssm-ID)	Additional domains (Pssm-ID)	Notes
Oedu	XP_048762457.1	Fox	Fox-D	Forkhead domain D4 (410822)		
Pcan	XP_025110523.1	Fox	Fox-D	Forkhead domain D4 (410822)		
Pcor	DN15187_c0_g1.i5.p1	Fox	Fox-D	Forkhead domain D4 (410822)		
Pmar	DN31265_c0_g1.i1.p1	Fox	Fox-D	Forkhead domain D4 (410822)		
Pmax	XP_033737945.1	Fox	Fox-D	Forkhead domain D4 (410822)		
Pstr	KAK3592139.1	Fox	Fox-D	Forkhead domain D4 (410822)		
Pvir	s01422g51	Fox	Fox-D	Forkhead domain D4 (410822)		
Pves	s0097g340	Fox	Fox-D	Forkhead domain D4 (410822)		
Rphi	XP_021345225.1	Fox	Fox-D	Forkhead domain D4 (410822)		
Sbro	XP_060588828.1	Fox	Fox-D	Forkhead domain D4 (410822)		
Scon	EVM0002351.1	Fox	Fox-D	Forkhead domain D4 (410822)		
Serlo	Chr8:2069	Fox	Fox-D	Forkhead domain D4 (410822)		
Tgra	Sg1013024	Fox	Fox-D	Forkhead domain D4 (410822)		
Airc	KAJ83306624.1	Fox	Fox-E	Forkhead domain E (410793)		
Contig989.19	Contig989.19	Fox	Fox-E	Forkhead domain E (410793)		
Amar	Amar12850	Fox	Fox-E	Forkhead domain E (410793)		
Apur	scaffold_25.23	Fox	Fox-E	Forkhead domain E (410793)		
Cang	XP_052638878.1	Fox	Fox-E	Forkhead domain E (410793)		
Cari	EVM0003339.1	Fox	Fox-E	Forkhead domain E (410793)		
Chlu	DN108936_c2_g1.i1.p1	Fox	Fox-E	Forkhead domain E (410793)		
Cejig	XP_011444776.2	Fox	Fox-E	Forkhead domain E (410793)		
Cvrr	XP_022319236.1	Fox	Fox-E	Forkhead domain E (410793)		
Dpol	XP_052288560.1	Fox	Fox-E	Forkhead domain E (410793)		
Hbia	M0000038943	Fox	Fox-E	Forkhead domain E (410793)		
Hruf	XP_0463535578.2	Fox	Fox-E	Forkhead domain E (410793)		
Mare	XP_052778423.1	Fox	Fox-E	Forkhead domain E (410793)		
Mcal	XP_052075782.1	Fox	Fox-E	Forkhead domain E (410793)		
Mcor	CAC5384360.1	Fox	Fox-E	Forkhead domain E (410793)		
Medu	CAG2217852.1	Fox	Fox-E	Forkhead domain E (410793)		
Medu	CAG2194171.1	Fox	Fox-E	Forkhead domain E (410793)		
Megl	CAG2199036.1	Fox	Fox-E	Forkhead domain E (410793)		
Mnar	VDH90460.1	Fox	Fox-E	Forkhead domain E (410793)		
Mmer	MMAM0000033594.4	Fox	Fox-E	Forkhead domain E (410793)		
Mmod	XP_045157592.2	Fox	Fox-E	Forkhead domain E (410793)		
Mner	DN117568_c0_g1.i1.p1	Fox	Fox-E	Forkhead domain E (410793)		
Mphi	g2411620_t1	Fox	Fox-E	Forkhead domain E (410793)		
Oedu	s00131587.0.4	Fox	Fox-E	Forkhead domain E (410793)		
Pmar	XP_048762491.1	Fox	Fox-E	Forkhead domain E (410793)		
Pmax	DN27017_c0_g1.i1.p1	Fox	Fox-E	Forkhead domain E (410793)		
Pstr	XP_033737819.1	Fox	Fox-E	Forkhead domain E (410793)		
Pvir	KAK3583103.1	Fox	Fox-E	Forkhead domain E (410793)		
Pves	s0013145g54	Fox	Fox-E	Forkhead domain E (410793)		
Rdec	XP_021378858.1	Fox	Fox-E	Forkhead domain E (410793)		
Rphi	DN24595_c4_g1.i1.p1	Fox	Fox-E	Forkhead domain E (410793)		
Sbro	XP_060578687.1	Fox	Fox-E	Forkhead domain E (410793)		
Serlo	EVM0010028.1	Fox	Fox-E	Forkhead domain E (410793)		
Sgio	Sej009305	Fox	Fox-F	Forkhead domain F1 (410823)		
Acal	XP_005105969.2	Fox	Fox-F	Forkhead domain F1 (410794)		
Airc	Contig1133.18	Fox	Fox-F	Forkhead domain F1 (410794)		
Amar	Ama39500	Fox	Fox-F	Forkhead domain F1 (410823)		
Amar	Ana32615	Fox	Fox-F	Forkhead domain F1 (410823)		
Apc	DN75342_c0_g1.i1.p1	Fox	Fox-F	Forkhead domain F1 (410823)		
Apur	scaffold_860_37	Fox	Fox-F	Forkhead domain F1 (410794)		
Bglia	XP_055892380.1	Fox	Fox-F	Forkhead domain F1 (410823)		
Cang	XP_052712246.1	Fox	Fox-F	Forkhead domain F1 (410794)		
Cari	EVM0011190.1	Fox	Fox-F	Forkhead domain F1 (410794)		
Cejig	XP_011445317.1	Fox	Fox-F	Forkhead domain F1 (410794)		
Cpli	DN7628_c0_g1.i1.p1	Fox	Fox-F	Forkhead domain F1 (410794)		
Csin	Hic_asm_17_158	Fox	Fox-F	Forkhead domain F1 (410794)		
Cvrr	XP_022335664.1	Fox	Fox-F	Forkhead domain F1 (410794)		
Dpol	XP_052232755.1	Fox	Fox-F	Forkhead domain F1 (410794)		
Gaeq	XP_041375666.1	Fox	Fox-F	Forkhead domain F1 (410794)		
Hbia	M0000007664	Fox	Fox-F	Forkhead domain F1 (410794)		
Hruf	XP_046372649.1	Fox	Fox-F	Forkhead domain F1 (410794)		
Mare	XP_052815084.1	Fox	Fox-F	Forkhead domain F1 (410794)		
Mcal	XP_052060477.1	Fox	Fox-F	Forkhead domain F1 (410794)		
Mcor	CAC5387332.1	Fox	Fox-F	Forkhead domain F1 (410794)		
Medu	CAG2252875.1	Fox	Fox-F	Forkhead domain F1 (410794)		
Megl	VD121852.1	Fox	Fox-F	Forkhead domain F1 (410794)		

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Pssm-ID)	Additional domains (Pssm-ID)	Notes
Mmar	MMAV00000030848	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Mmer	XP_045194642.1	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Mmod	DN104261_c0.g1.i1.p1	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Mmer	g106129.t1	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Mphi	scaf.40546.0.2	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Obim	XP_047775391	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Oedu	XP_0487322022.1	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Pcan	XP_025116010.1	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Pcor	DN180603_c0.g1.i1.p1	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Rphi	DN129940_c0.g1.i1.p1	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Pmar	DN14344_c0.g1.i1.p1	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Pmax	XP_033755005.1	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Pstr	KAK3601654.1	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Pvir	sl133835x10	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Pyes	XP_021358008.1	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Rphi	XP_060601663.1	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Sbro	EVM0015186.1	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Scor	Chr11.927	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Scor	Chr11.810	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Sgio	Sgi005267	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Tgra	KAJ8302829.1	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Tsqu	DN137576_c0.g1.i1.p1	Fox	Fox-F	Fox	Fox-F	Forkhead domain F (410794)
Acal	XP_005099252.2	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Acal	XP_005099233.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Airc	Contig625_38	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Amar	Ama10381	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Apec	DN10836.c0.g1.i1.p1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Apur	scaffold_36470_28	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Bglia	XP_055879295.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Cang	XP_052699015.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Cari	EVM0001891.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Cflu	DN104980_c0.g1.i2.p1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Cgig	XP_011427689.2	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Cipl	DN158419_c0.g1.i1.p1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Csin	Hic.assn.10.1034	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Cvir	XP_022334541.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Dpol	XP_052270224.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Dpol	XP_052270147.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Gaeq	XP_041354930.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Gaeq	XP_041354700.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Hbia	M00000035850	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Hruf	XP_046371537.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Mcal	XP_052104484.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Mcor	CAC5405696.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Medu	CAG219343.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Mgal	VD124297.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Mmar	MMAV0000030730	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Mmer	XP_045162348.2	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Mmod	DN60588_c0.g1.i1.p1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Mmer	g13325_t1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Mphi	scaf.15017.0.3	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Obim	XP_052824484.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Oedu	XP_048737541.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Pcan	XP_025105677.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Pcan	XP_025106039.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Pcor	XP_025105724.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Pcor	DN81635_c0.g2.i1.p1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Pgen	DN81635_c0.g1.i1.p1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Pmar	DN12984_c0.g1.i1.p1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Pmax	XP_033734631.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Poku	DN41090_c0.g2.i1.p1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Pstr	PKA3604690.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Pvir	s00333g35	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Pyes	XP_021363790.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Rphi	XP_0605889805.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Sbro	EVM0011335.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)
Sbro	EVM0012906.1	Fox	Fox-G	Fox	Fox-G	Forkhead domain G (410795)

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Pssm-ID)	Additional domains (Pssm-ID)	Notes
Scon	Chr3:2805	Fox	Fox-G	Forkhead domain G (410795)		
Sgio	Sg1014601	Fox	Fox-G	Forkhead domain G (410795)		
Sgra	DN49488-C0.g1.i1.p1	Fox	Fox-G	Forkhead domain G (410795)		
Airc	Contig18244.2	Fox	Fox-H	Forkhead domain H (410796)		
Airc	Contig178.106	Fox	Fox-H	Forkhead domain H (410796)		
Amar	Ama16564	Fox	Fox-H	Forkhead domain H (410796)		
Amar	Ama05868	Fox	Fox-H	Forkhead domain H (410796)		
Cang	XP_052684756.1	Fox	Fox-H	Forkhead domain H (410796)		
Cari	EVM0021377.1	Fox	Fox-H	Forkhead domain H (410796)		
Cflu	DN1138466_c0.g1.i1.p1	Fox	Fox-H	Forkhead domain H (410796)		
Cgig	XP_03431325.1	Fox	Fox-H	Forkhead domain H (410796)		
Hic.asrn.14.811	Hic.asrn.14.811	Fox	Fox-H	Forkhead domain H (410796)		
Cvir	XP_022314590.1	Fox	Fox-H	Forkhead domain H (410796)		
Hbia	M00000018729	Fox	Fox-H	Forkhead domain H (410796)		
Hruf	XP_048254913.1	Fox	Fox-H	Forkhead domain H (410796)		
Hruf	XP_048255113.1	Fox	Fox-H	Forkhead domain H (410796)		
Mcor	CAC409624.1	Fox	Fox-H	Forkhead domain H (410796)		
Mcor	CAC3397897.1	Fox	Fox-H	Forkhead domain H (410796)		
Mcor	CAC5397906.1	Fox	Fox-H	Forkhead domain H (410796)		
Mcor	CAC5403969.1	Fox	Fox-H	Forkhead domain H (410796)		
Medu	CAG2228903.1	Fox	Fox-H	Forkhead domain H (410796)		
Medu	CAG2188004.1	Fox	Fox-H	Forkhead domain H (410796)		
Medu	CAG2252853.1	Fox	Fox-H	Forkhead domain H (410796)		
Medu	CAG2220596.1	Fox	Fox-H	Forkhead domain H (410796)		
Mgal	VD162725.1	Fox	Fox-H	Forkhead domain H (410796)		
Mgal	VDH3947.1	Fox	Fox-H	Forkhead domain H (410796)		
Mgal	VD120844.1	Fox	Fox-H	Forkhead domain H (410796)		
Mmar	MM-MAN000000022684.	Fox	Fox-H	Forkhead domain H (410796)		
Mmer	XP_053378226.1	Fox	Fox-H	Forkhead domain H (410796)		
Mmer	XP_045194303.2	Fox	Fox-H	Forkhead domain H (410796)		
Mmer	XP_045196985.2	Fox	Fox-H	Forkhead domain H (410796)		
Mphi	g213542.t1	Fox	Fox-H	Forkhead domain H (410796)		
Mphi	scaf.17325.0.4	Fox	Fox-H	Forkhead domain H (410796)		
Mphi	s28666.1.1	Fox	Fox-H	Forkhead domain H (410796)		
Oedu	XP_048759429.2	Fox	Fox-H	Forkhead domain H (410796)		
Pcan	XP_025075984.1	Fox	Fox-H	Forkhead domain H (410796)		
Pcor	DN116957.co.g1.i1.p1	Fox	Fox-H	Forkhead domain H (410796)		
Pmax	XP_033755807.1	Fox	Fox-H	Forkhead domain H (410796)		
Pstr	KAK3603859.1	Fox	Fox-H	Forkhead domain H (410796)		
Pvir	s75596g33	Fox	Fox-H	Forkhead domain H (410796)		
Pvir	s00234g131	Fox	Fox-H	Forkhead domain H (410796)		
Rphi	XP_060558970.1	Fox	Fox-H	Forkhead domain H (410796)		
Rphi	XP_060567331.1	Fox	Fox-H	Forkhead domain H (410796)		
Rphi	XP_060604067.1	Fox	Fox-H	Forkhead domain H (410796)		
Sbro	EVMM0016618.1	Fox	Fox-H	Forkhead domain H (410796)		
Sbro	EVMM0013817.1	Fox	Fox-H	Forkhead domain H (410796)		
Scon	Chr11.1359	Fox	Fox-H	Forkhead domain H (410796)		
Scon	Chr2.2082	Fox	Fox-H	Forkhead domain H (410796)		
Sgio	Sg1013003	Fox	Fox-H	Forkhead domain H (410796)		
Acal	XP_05108651.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Airc	Contig775.5	Fox	Fox-J1	Forkhead domain J1 (410797)		
Amar	Ama02822	Fox	Fox-J1	Forkhead domain J1 (410797)		
Apcc	DN20109_c0.g1.i6.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Apur	scaffold_797.10	Fox	Fox-J1	Forkhead domain J1 (410797)		
Brla	XP_013064514.2	Fox	Fox-J1	Forkhead domain J1 (410797)		
Cang	XP_052689275.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Cari	EVMM0033558.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Cflu	DN127407_c0.g2.i1.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Cgig	XP_011445234.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Cpli	DN65792_c0.g1.i1.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Csin	Hic.asrn.0.1540	Fox	Fox-J1	Forkhead domain J1 (410797)		
Cvir	XP_022319181.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Dpol	XP_0522319268.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Geg	XP_041362703.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Hbia	M0000003225	Fox	Fox-J1	Forkhead domain J1 (410797)		
Hruf	XP_046330175.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Lorb	DN146717_c0.g1.i1.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Mare	XP_052764636.1	Fox	Fox-J1	Forkhead domain J1 (410797)		

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Psm-ID)	Additional domains (Psm-ID)	Notes
Mare	XP_052816854.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Mare	XP_052764667.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Mare	XP_052775202.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Mare	XP_052775217.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Mare	XP_052775230.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Mcal	XP_052068038.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Mchi	DN41592.0.g1.i5.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Mcor	CAC40574.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Medu	CAG2242807.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Mgal	VD12691.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Mmar	MMAM0000019873	Fox	Fox-J1	Forkhead domain J1 (410797)		
Mmer	XP_045212565.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Mmod	DN23659.0.g1.i1.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Mner	gl198765.t1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Mphi	scfa:33310.2.9	Fox	Fox-J1	Forkhead domain J1 (410797)		
Obim	XP_052824622.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Oedu	XP_048763213.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Osin	XP_029638410.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Pcan	XP_025059423.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Pcor	DN4891.0.g1.i2.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Pcor	DN480.c0.s2.i1.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Pcor	DN18891.0.g2.i2.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Pgen	DN2399.c8.g1.i1.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Pmar	DN32837.cl.g1.i1.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Pmax	XP_03375259.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Poku	DN19777.02.g1.i1.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Ppur	DN2521.c0.g1.i4.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Pstr	KAK3579229.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Pvir	s01693g10	Fox	Fox-J1	Forkhead domain J1 (410797)		
Pyes	XP_021351058.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Rdec	DN22834.c0.g1.i1.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Robi	XP_060587750.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Sbro	EVM0018668.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Scen	Chr1_3201	Fox	Fox-J1	Forkhead domain J1 (410797)		
Scen	Chr1_3198	Fox	Fox-J1	Forkhead domain J1 (410797)		
Sgio	Sgi000050	Fox	Fox-J1	Forkhead domain J1 (410797)		
Sgra	DN11939.c0.g1.i11.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Tgra	KA.J83.18321.1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Tsqu	DN6625.c2.g1.i2.p1	Fox	Fox-J1	Forkhead domain J1 (410797)		
Acal	XP_005111247.3	Fox	Fox-J2/3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Contig53.201	Fox	Fox-J2/3				
Amar	Ama34942	Fox	Fox-J2/3			
Apur	scffold_11801.24	Fox	Fox-J2/3			
Bjia	XP_03070049.1	Fox	Fox-J2/3			
Cang	DN625.c2.g1.i2.p1	Fox	Fox-J2/3			
Cari	EVMM0008910.1	Fox	Fox-J2/3			
Arc	Contig53.201	Fox	Fox-J2/3			
Cflu	DN58808.c0.g1.i1.p1	Fox	Fox-J2/3			
Cgig	XP_011422995.2	Fox	Fox-J2/3			
Cpli	DN78731.c5.g1.i2.p1	Fox	Fox-J2/3			
Mare	XP_052759824.1	Fox	Fox-J2/3			
Csin	Hic.asn.4.381	Fox	Fox-J2/3			
Cvir	XP_022341777.1	Fox	Fox-J2/3			
Dpol	XP_052285372.1	Fox	Fox-J2/3			
Gaeg	XP_041378546.1	Fox	Fox-J2/3			
Hbia	M00000010754	Fox	Fox-J2/3			
Hruf	XP_048247606.1	Fox	Fox-J2/3			
Mare	XP_052759824.1	Fox	Fox-J2/3			
Mcal	VD157447.1	Fox	Fox-J2/3			
Mgal	VD157448.1	Fox	Fox-J2/3			
Mchi	MMAV0000001729	Fox	Fox-J2/3			
Mcor	CAC5378041.1	Fox	Fox-J2/3			
Medu	CAG2221519.1	Fox	Fox-J2/3			
Mgal	MMer	Fox	Fox-J2/3			
Mmer	XP_053379966.1	Fox	Fox-J2/3			
Mmod	DN3610.c0.g1.i1.p1	Fox	Fox-J2/3			
Mphi	scfa:40576.0.4	Fox	Fox-J2/3			
Obim	XP_052832979.1	Fox	Fox-J2/3			

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Psm-ID)	Additional domains (Psm-ID)	Notes
Oedu	XP_048739234.1	Fox	Fox-J2.3	Forkhead domain J3 (410826)		
Osin	XP_029651657.1	Fox	Fox-J2.3	Forkhead domain J3 (410826)		
Pcan	XP_025081218.1	Fox	Fox-J2.3	Forkhead domain J3 (410826)		
Pcor	DN2942_c0_g1.i5.p1	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Pcor	DN2942_c0_g4.i3.p1	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Pcor	DN2942_c0_g4.i3.p2	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Pgen	DN5381_c0_g1.i7.p1	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Pmar	DN41364_c0_g1.i3.p1	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Pmax	XP_033763328.1	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Poku	DN14771_c0_g2.i3.p1	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Ppur	DN2181_c0_g1.i5.p1	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Pstr	KAK38383417.1	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Pvir	s0019gg6	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Pyes	XP_021374633.1	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Rphi	XP_0605777634.1	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Sbro	EVM0013081.1	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Scor	Chr12_800	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Sglo	Sg1003279	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Sgra	DN1042_c0_g1.i2.p1	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Tgra	KAJ3320220.1	Fox	Fox-J2.3	Forkhead domain FOXJ2, FOXJ3 (410798)		
Tsqu	DN10376_c0_g1.i2.p1	Fox	Fox-K	Forkhead domain K (410826)		
Acal	XP_005092494.1	Fox	Fox-K	Forkhead domain K (410826)		
Apur	XP_013090285.1	Fox	Fox-K	Forkhead domain K (410826)		
Bgia	XP_052688140.1	Fox	Fox-K	Forkhead domain K (410826)		
Cang	EVMM00000012630	Fox	Fox-K	Forkhead domain K (410826)		
Cari	XP_001416098.1	Fox	Fox-K	Forkhead domain K (410826)		
Cgig	DN61350_c0_g1.i1.p1	Fox	Fox-K	Forkhead domain K (410826)		
Cpli	DN61350_c0_g1.i1.p1	Fox	Fox-K	Forkhead domain K (410826)		
Cvir	XP_022316096.1	Fox	Fox-K	Forkhead domain K (410826)		
Gaeg	XP_041362451.1	Fox	Fox-K	Forkhead domain K (410826)		
Hbif	M00000000333	Fox	Fox-K	Forkhead domain K (410826)		
Hruf	XP_048246693.1	Fox	Fox-K	Forkhead domain K (410826)		
Mmar	MMAM00000012630	Fox	Fox-K	Forkhead domain K (410826)		
Mphi	XP_14580_i.11	Fox	Fox-K	Forkhead domain K (410826; partial)		
Obim	XP_014772374.1	Fox	Fox-L	Forkhead domain L (410826)		
Oedu	XP_048761057.2	Fox	Fox-L	Forkhead domain L (410826)		
Osin	XP_029646877.1	Fox	Fox-L	Forkhead domain L (410826)		
Rdec	DN21696_c3_g1.i1.p1	Fox	Fox-L	Forkhead domain L (410826)		
Sgio	Sg100589	Fox	Fox-L	Forkhead domain L (410826)		
Acal	XP_012940028.1	Fox	Fox-L	Forkhead domain L (410826)		
Contig58.64		Fox	Fox-L	Forkhead domain L (410826)		
Amar	Amal17914	Fox	Fox-L	Forkhead domain L (410826)		
Apec	DN74037_c0_g1.i1.p1	Fox	Fox-L	Forkhead domain L (410826)		
Apulf	scaffold_122_2	Fox	Fox-L	Forkhead domain L (410826)		
Bgia	XP_055890278.1	Fox	Fox-L	Forkhead domain L (410826)		
Cang	XP_0526718636.1	Fox	Fox-L	Forkhead domain L (410826)		
Cari	EVM0019009.1	Fox	Fox-L	Forkhead domain L (410826)		
Cgig	XP_011417586.2	Fox	Fox-L	Forkhead domain L (410826)		
Cpli	DN15469_c0_g4.i1.p1	Fox	Fox-L	Forkhead domain L (410826)		
Cvir	Hic_asn_17_1225	Fox	Fox-L	Forkhead domain L (410826)		
Dpol	XP_05225043.1	Fox	Fox-L	Forkhead domain L (410826)		
Gaeg	XP_001375667.1	Fox	Fox-L	Forkhead domain L (410826)		
Hbif	M0000031057	Fox	Fox-L	Forkhead domain L (410826)		
Hruf	XP_046344397.2	Fox	Fox-L	Forkhead domain L (410826)		
Lorb	DN104456_c0_g1.i1.p1	Fox	Fox-L	Forkhead domain L (410826)		
Mare	XP_052817977.1	Fox	Fox-L	Forkhead domain L (410826)		
Mcal	XP_05225043.1	Fox	Fox-L	Forkhead domain L (410826)		
Mcov	DN51324_c0_g1.i1.p1	Fox	Fox-L	Forkhead domain L (410826)		
Medu	CAC5374005.1	Fox	Fox-L	Forkhead domain L (410826)		
Mgal	CAG2206845.1	Fox	Fox-L	Forkhead domain L (410826)		
Mgal	VD122484.1	Fox	Fox-L	Forkhead domain L (410826)		
Mgal	VDH97507.1	Fox	Fox-L	Forkhead domain L (410826)		
Mmar	MMAN00000028776	Fox	Fox-L	Forkhead domain L (410826)		
Mmer	XP_053402988.1	Fox	Fox-L	Forkhead domain L (410826)		
Mmod	DN51324_c0_g1.i1.p1	Fox	Fox-L	Forkhead domain L (410826)		
Mner	g268924.t1	Fox	Fox-L	Forkhead domain L (410826)		
Mphi	scaf69950_0.0	Fox	Fox-L	Forkhead domain L (410826)		
Obim	XP_0147785001.1	Fox	Fox-L	Forkhead domain L (410826)		

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Pssm-ID)	Additional domains (Pssm-ID)	Notes
Oedu	XP_048762056_2	Fox	Fox-L1	Forkhead domain L1 (410801)		
Pcan	XP_025076243_1	Fox	Fox-L1	Forkhead domain L1 (410801)		
Pcor	DN23326_0_c0_g1_i1_p1	Fox	Fox-L1	Forkhead domain L1 (410801)		
Pcor	DN181497_c0_g1_i1_p1	Fox	Fox-L1	Forkhead domain L1 (410801)		
Pmar	DN30135_c0_g1_i1_p1	Fox	Fox-L1	Forkhead domain L1 (410801)		
Pmax	XP_033755384_1	Fox	Fox-L1	Forkhead domain L1 (410801)		
Ppar	DN7931_c0_g1_i1_p1	Fox	Fox-L1	Forkhead domain L1 (410801)		
Pstr	KAK3390991_1	Fox	Fox-L1	Forkhead domain L1 (410801)		
Pvir	s02023g11	Fox	Fox-L1	Forkhead domain L1 (410801)		
Pyes	XP_021346965_1	Fox	Fox-L1	Forkhead domain L1 (410801)		
Rphi	XP_060608039_1	Fox	Fox-L1	Forkhead domain L1 (410801)		
Sbro	EVM0016190_1	Fox	Fox-L1	Forkhead domain L1 (410801)		
Scon	Chr1-1868	Fox	Fox-L1	Forkhead domain L1 (410801)		
Sglo	Sg1009486	Fox	Fox-L1	Forkhead domain L1 (410801)		
Acal	XP_05101910_2	Fox	Fox-L2	Forkhead domain L2 (410802)		
Airc	Contig551_34	Fox	Fox-L2	Forkhead domain L2 (410802)		
Amar	Ama34673	Fox	Fox-L2	Forkhead domain L2 (410802)		
Apur	scaffold84_159	Fox	Fox-L2	Forkhead domain L2 (410802)		
Bgia	XP_05586510_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Cang	EVMD0021728_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Cari	DN127322_6_5_2_12_p1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Cflu	NP_001293827_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Cggg	DN75086_c5_g1_i2_p1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Cpli	His_asm_4_274	Fox	Fox-L2	Forkhead domain L2 (410802)		
Csin	XP_022345405_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Cvir	XP_022345173_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Dpol	XP_052212727_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Gaeq	XP_041378252_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Hbia	M0000035173	Fox	Fox-L2	Forkhead domain L2 (410802)		
Hrf	XP_048250295_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Lorb	DN129129_c0_g1_i1_p1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Mare	XP_052760962_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Mcal	XP_052082415_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Mcor	CAC5401149_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Medu	CAG2239672_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Mgal	VD149865_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Mgal	VD149864_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Mmar	MMAN00000016212	Fox	Fox-L2	Forkhead domain L2 (410802)		
Mmer	XP_04516164_2	Fox	Fox-L2	Forkhead domain L2 (410802)		
Mmod	DN2410_c0_g1_i1_p1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Mner	g832_35_t1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Mphi	scfa50301_0_3	Fox	Fox-L2	Forkhead domain L2 (410802)		
Obim	XP_047386648_2	Fox	Fox-L2	Forkhead domain L2 (410802)		
Oedu	XP_048729555_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Pcan	XP_025083514_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Pcor	DN3937_c0_g1_i2_p1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Pgen	DN134171_c0_g1_i1_p1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Pmar	EVM0017513_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Pmax	XP_033724493_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Pstr	KAK3602726_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Pvir	s00246g193	Fox	Fox-L2	Forkhead domain L2 (410802)		
Pyes	XP_021353421_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Rdec	DN37_c29_g1_i1_p1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Rphi	XP_060586301_1	Fox	Fox-L2	Forkhead domain L2 (410802)		
Sbro	Contig281_47	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Scon	Ama23426	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Sglo	DN27027_c0_g1_i2_p1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Tsequ	scaffold17_163	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Acal	XP_05091040_1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Airc	Contig281_47	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Amar	Apuc	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Apuc	scaffold17_163	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Bgia	XP_058390100_1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Cang	XP_052711314_1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Cari	EVM002431_1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Cggg	XP_0343303195_1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Pssm-ID)	Additional domains (Pssm-ID)	Notes
Cpli	DN78331.cl.g.i1.p1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Csin	Hic.asm.2.101	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Cvir	XP_022292787.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Dpol	XP_052270473.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Gaeg	XP_041365083.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Hbia	M00000027642	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Hruf	XP_04221610.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Mare	XP_052801997.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Mcal	XP_052066630.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Mchi	DN2972.c0.g.i1.p1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Mcor	CAC5383890.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Medu	CAG2257106.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Mgal	VDH93464.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Mgal	VDH93462.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Mgal	VDH93463.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Mmar	MMAM0000018109	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Mmer	XP_045177340.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Mmod	DN2507.c0.g2.i1.p1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Mphi	scf.69935.0.10	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Obim	XP_052825413.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Oedu	XP_055996035.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Osin	XP_029638459.2	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Pcan	XP_025087495.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Pcor	DN55558.c0.g2.i1.p1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Pcor	DN55558.c0.g1.i1.p1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Pgen	DN145626.c0.i1.i1.p1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Pmar	DN30748.c0.g1.i1.p1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Pmax	XP_033751425.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Poku	DN45531.c0.g1.i1.p1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Ppur	DN199653.c0.g1.i1.p1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Pstr	KAK3387366.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Pvir	s24333g45	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Pyes	XP_021371548.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Rdec	DN20122.c0.g1.i1.p1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Rphi	XP_060606622.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Sbro	EVM0009578.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Scon	Chr14.2061	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Sglo	Sg100456	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Sgra	DN11935.c0.g2.i3.p1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Tgra	KAJ8298705.1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Tsqu	DN22139.c0.g1.i1.p1	Fox	Fox-N1/4	Forkhead domain N1 (410804)		
Acal	XP_005099217.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Airc	Contig117.153	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Amar	Amar9979	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Apuc	DN11918.c0.g1.i10.p1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Apuc	scf49.22	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Bjga	XP_013084252.2	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Cang	XP_052698143.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Carl	EVM0016469.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Cflu	DN125734.cl.g1.i18.p1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Cgig	XP_034324255.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Cpfl	DN79231.c0.g1.i7.p1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Csin	Hic.asm.10.136	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Cvir	XP_022331107.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Dpol	XP_052270965.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Gaeg	XP_041353111.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Hbia	M0000030949	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Hruf	XP_046351344.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Mare	XP_052767715.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Mcal	XP_052107190.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Mchi	DN39446.c1.g1.i1.p1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Mcor	CAC5378437.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Medu	CAG2233611.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Mgal	VDI80286.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Mgal	VDI80289.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Mgal	VDI80287.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Mmar	MMAM0000016688	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Mmer	XP_053376884.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Mmod	DN2418.c0.g1.i131.p1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Pssm-ID)	Additional domains (Pssm-ID)	Notes
Mner	g186153.t2	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Mphi	scaf_37509_0.5	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Obim	XP_052822674.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Oedu	XP_048733022.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Osin	XP_029633348.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Pcan	XP_025106088.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Pcor	DN105240_c0.i1.i1.p1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Pcor	DN5191_c0.g2.i7.p1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Pcor	DN18451_c0.g2.i3.p1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Pgen	DN14328_c0.g1.i3.p1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Pmar	DN42157_c1.g1.i3.p1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Pmax	XP_033734749.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Poku	DN17429_c4.i2.p1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Ppur	DN5075_c0.g1.i1.p1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Pstr	KAK395953.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Pvir	s00410g05	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Pyes	XP_021366994.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Rdec	DN2296_c2.g1.i1.p1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Rphi	XP_060552999.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Sgio	Sgi013452	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Sgra	DN13133_c0.g2.i8.p1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Tgra	KAJ8308641.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Tgra	KAJ8316727.1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Tsqu	DN75347_c0.g1.i2.p1	Fox	Fox-N2/3	Forkhead domain N3 (410833)		
Acal	XP_005112460.1	Fox	Fox-O	Forkhead domain O (410806)		
Airc	Contig116_24	Fox	Fox-O	Forkhead domain O (410806)		
Amar	Amat07814	Fox	Fox-O	Forkhead domain O (410806)		
Apec	DN23636_c0.g1.i1.p1	Fox	Fox-O	Forkhead domain O (410806)		
Apur	scaffold_712.6	Fox	Fox-O	Forkhead domain O (410834)		
Bgia	XP_01309255.2	Fox	Fox-O	Forkhead domain O (410833)		
Cang	EVMD00009468.1	Fox	Fox-O	Forkhead domain O (410833)		
Cari	DN112955_c5.g3.i5.p1	Fox	Fox-O	Forkhead domain O (410806)		
Cflu	XP_011414359.1	Fox	Fox-O	Forkhead domain O (410806)		
Cgg	DN72415_c5.g1.i1.p1	Fox	Fox-O	Forkhead domain O (410806)		
Cpli	XP_022287692.1	Fox	Fox-O	Forkhead domain O (410806)		
Cvir	XP_022287423.1	Fox	Fox-O	Forkhead domain O (410806)		
Dpov	XP_052243903.1	Fox	Fox-O	Forkhead domain O (410806)		
Geeg	XP_0413351714.1	Fox	Fox-O	Forkhead domain O (410806)		
Hbia	M0000024236	Fox	Fox-O	Forkhead domain O (410834)		
Hruf	XP_046374293.1	Fox	Fox-O	Forkhead domain O (410806)		
Lorb	DN142512_c0.g1.i1.p1	Fox	Fox-O	Forkhead domain O (410806)		
Mare	XP_052808484.1	Fox	Fox-O	Forkhead domain O (410806)		
Meal	XP_052087664.1	Fox	Fox-O	Forkhead domain O (410806)		
Mchi	DN32908_c0.g1.i3.p1	Fox	Fox-O	Forkhead domain O (410806)		
Oedu	CAC539258.1	Fox	Fox-O	Forkhead domain O (410806)		
Mmar	MMAM00000004017	Fox	Fox-O	Forkhead domain O (410806)		
Mmer	XP_045195791.2	Fox	Fox-O	Forkhead domain O (410806)		
Mmod	DN145_c2.g1.i2.p1	Fox	Fox-O	Forkhead domain O (410806)		
Mner	g187283.t1	Fox	Fox-O	Forkhead domain O (410835)		
Obim	XP_052832486.1	Fox	Fox-O	Forkhead domain O (410806)		
Oedu	XP_048766900.1	Fox	Fox-O	Forkhead domain O (410806)		
Osin	XP_029657040.1	Fox	Fox-O	Forkhead domain O (410834)		
Pcan	XP_025097678.1	Fox	Fox-O	Forkhead domain O (410806)		
Pcor	DN24871_c0.g1.i3.p2	Fox	Fox-O	Forkhead domain O (410806)		
Pgen	DN44399_c0.g5.i1.p1	Fox	Fox-O	Forkhead domain O (410806)		
Pmar	XP_033740844.1	Fox	Fox-O	Forkhead domain O (410806)		
Pmax	DN10962_c1.g2.i2.p1	Fox	Fox-O	Forkhead domain O (410806)		
Pcor	DN752510_c0.g1.i1.p1	Fox	Fox-O	Forkhead domain O (410806)		
Ppur	Pstr	KAK3576955.1	Fox	Forkhead domain O (410806)		
Pvir	s000798.02	Fox	Fox-O	Forkhead domain O (410806)		
Pyes	XP_021377366.1	Fox	Fox-O	Forkhead domain O (410806)		
Rdec	DN30721_c0.g1.i1.p1	Fox	Fox-O	Forkhead domain O (410806)		
Rphi	XP_0605889384.1	Fox	Fox-O	Forkhead domain O (410806)		
Scon	Chr2.491	Fox	Fox-O	Forkhead domain O (410806)		

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Pssm-ID)	Additional domains (Pssm-ID)	Notes
Sglo	Sglo18351	Fox	Fox-O	Forkhead domain O (410834)		
Sgra	DN5576_c0_g1.i1.p1	Fox	Fox-O	Forkhead domain O (410806)		
Tsqu	DN138882_c0_g1.i1.p1	Fox	Fox-O	Forkhead domain O (410806)		
Airc	Contig3461.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Airc	Contig330.72	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Apur	scaffold_576_108	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Cang	XP_052672828.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Cari	EV/M0015778.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Cgig	XP_011412482.2	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Cvir	XP_022300144.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Dpol	XP_052234997.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Dpol	XP_052237166.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Gaeg	XP_041362068.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Hbia	M00000010651	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Hruf	XP_046328651.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Hruf	XP_052088402.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Mcac	CAC338314.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Medu	CAG2250347.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Mgal	VD10563.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Mmar	MMAM00000027087	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Mmer	XP_045182963.2	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Mmod	DN9753_c0_g1.i1.p1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Mphi	scaf66119.0.21	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Oedu	XP_048731527.2	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Pmar	DN32892_c0_g1.i1.p1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Pmax	XP_033727511.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Petr	KAK3609024.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Pvir	s01296g51	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Pyes	XP_0021354438.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Rlef	DN23702_c2_g1.i2.p1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Rphi	XP_060566633.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Sbro	EV/M0009544.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Scor	Chr1-409	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Sgio	Sgi004484	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Tgra	KAJ322379.1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Tsqu	DN207442_c0_g1.i1.p1	Fox	Fox-OG13/NA	Forkhead domain P (410807)		
Acal	Contig85.21	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Airc	Ama19770	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Apur	scaffold_360_14	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Bglj	XP_013071662.2	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Cang	XP_052676257.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Cari	EV/M00106916.3	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Cgig	DN34749_c0_g1.i1.p1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Cphi	DN34749_c0_g1.i1.p1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Csin	Hic-asm_12.159	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Cvir	XP_022296913.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Dpol	XP_052253230.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Dpol	XP_052253240.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Gaeg	XP_041347345.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Hbia	M00000000012410	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Mmar	XP_0451663371.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Mmer	scaf67833.0.2	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Mphi	XP_014772941.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Obim	XP_048742700.2	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Medu	VD10563.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Mgal	MMAM00000015843	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Mmar	XP_046382017.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Mmer	scaf67833.0.2	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Pcor	DN7667_c0_g1.i1.p1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Pcor	DN7667_c0_g3.i1.p1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Pmax	XP_033744896.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Psm-ID)	Additional domains (Psm-ID)	Notes
Pstr	KAK3389497.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Pyes	XP_021371037.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Rphi	XP_060586724.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Sbro	EVM00050406.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Sbro	EVM0006433.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Ch7-1624	Chr7-1624	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Scon	Sg024307	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Sgo	Airc	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Cpfh	Contig.1425.7	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Apur	scaffold_604.173	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Cang	XP_052676026.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Cari	EVM0023364.1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Cflu	DN107758.c5.g1.i2.p1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410758)	Annotated as Fox-Q2b	
Cgig	XP_0419927657.2	Fox	Fox-OG15/NA	Forkhead domain Q2 (410758)	Annotated as Fox-Q2b	
Cpbf	DN70609.0.g1.i2.p1	Fox	Fox-OG15/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2b	
Cvfr	XP_022321288.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Cvir	XP_022295893.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Dpol	XP_052222623.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Gaeq	XP_041365712.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Hbia	M0000030826	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Hruf	DN243786.c0.g1.i1.p1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Lorb	XP_052802309.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Mare	XP_05370465.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Mcor	CAG2202185.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Medu	CAG2246856.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Mgal	VD124665.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Mmar	MMAR00000032793	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Mmer	XP_045177123.2	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Mphi	scaf.70200.0.4	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Obim	XP_04771053.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Oeud	XP_048728661.2	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Osin	DM43205.c4.g1.i2.p1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Pmar	XP_033752233.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Pmax	DN4666.c0.g1.i2.p1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Ppur	DN4666.c0.g1.i2.p1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Pstr	KAK3581527.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Pvir	s00194g51	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Pyes	XP_021353413.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Rdec	DN22502.c0.g4.i1.p1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Rphi	XP_060599562.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Rphi	XP_060601370.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Sbro	EVM0002125.1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Scon	Chr14.1628	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Tsqu	DN2384.c1.g1.i1.p1	Fox	Fox-OG15/NA	Forkhead domain (410758)	Annotated as Fox-Q2b	
Airc	Contig.438.5	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Apec	DN23140.c1.g1.i1.p1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Apur	scaffold_333.74	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Cang	XP_052712245.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Cari	EVM0012482.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Cflu	DN113069.c3.g1.i4.p1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Cgig	XP_041366058.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Cpbf	DN76173.c0.g1.i2.p1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Csin	Hic-asn.2.1802	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Csin	Hic-asn.2.1455	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Cvfr	XP_022286391.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Dpol	XP_052282333.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Dpol	XP_052234537.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Gaeq	XP_048731286.2	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Hbia	M0000014061	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Hruf	XP_045336487.2	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Mare	XP_052798013.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Mcal	XP_052064572.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Mcor	CAC5375062.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Medu	CAG2224977.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Mmar	MMAR00000037791	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Mmer	XP_0533384136.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Mmod	DN27089.c0.g1.i1.p1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Mner	g144-243.t1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Pcor	DN5679.c0.g2.i1.p1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Pssm-ID)	Additional domains (Pssm-ID)	Notes
Pcor	DN15679_c0_g1.i15.p1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Pcor	DN113056_c0_g1.i1.p1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Pcor	DN113056_c0_g2.i2.p1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Pgen	DN128413_c1_g1.i2.p1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Pmar	DN44947_c1_g2.i3.p1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Pmax	XP_033751305.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Poku	DN37223_c1_g1.i8.p1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Ppur	DN3451_c0_g1.i1.p1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Pstr	KAK3597624.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Pvir	S00219g11	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Pyes	XP_021377259.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Rdec	DN2152_c4_g1.i1.p1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Rphi	XP_06056990.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Sbro	EVM0023670.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Sgio	Sg1005361	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Sgra	DN54780_c0_g1.i1.p1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Tgra	KAJ3299135.1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Tstu	DN6134_c0_g1.i7.p1	Fox	Fox-OG2/NA	Forkhead domain M (410803)	Annotated as Fox-M	
Airc	Contig465.41	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Amar	Ama25953	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Apur	scaffold_381.16	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Cang	XP_052700156.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Cari	EVM0004465.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Cejig	XP_011435457.2	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Csin	Hic-asm.16.939	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Cvir	XP_022334263.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Dpol	XP_052278575.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Dpol	XP_052278576.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Hrfu	XP_046341176.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mare	XP_052791887.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mare	XP_052791890.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mare	XP_052791888.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mare	XP_052791891.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mcal	XP_052098761.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mcal	XP_052099555.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mcor	CAC5419385.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mcor	CAC5380823.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mcor	CAC5370920.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mcor	CAC5419389.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mcor	CAC5419386.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mcor	CAC5419381.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mcor	CAC5419382.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Medu	CAC5419383.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Medu	CAC5419387.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Medu	CAC5419380.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Medu	CAC5419388.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Medu	CAG2214460.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Medu	CAG2194706.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Medu	CAG2198066.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Medu	CAG2198058.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Medu	CAG2198055.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Medu	CAG2214461.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Medu	CAG2198060.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Medu	CAG2198063.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Medu	CAG2198061.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Medu	CAG2198064.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Medu	CAG2198062.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mgal	VD130250.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mgal	VD13959.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mgal	VD115906.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mgal	VD102348.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mgal	VD15903.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Pssm-ID)	Additional domains (Pssm-ID)	Notes
Mgal	VD102347.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mgal	VD115905.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mgal	VD102349.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mgal	VD115904.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mimer	XP_053405087.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mmod	DN6982_C0_g1.i3.p1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Mphi	scfa:15444_01	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Oedu	XP_056021213.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Sbro	DN39963_C0_g1.i1.p1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Pmax	XP_033751006.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Pmax	XP_033749723.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Pvir	s03457g48	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Pves	XP_0221360588.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Rphi	XP_060588777.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
EVM0013029.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c		
Cang	Ch5_396.1	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Scon	Se1013625	Fox	Fox-OG28/NA	Forkhead domain Q2 (410809)	Annotated as Fox-Q2c	
Acal	XP_005102249.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Airc	Contig636_41	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Amar	Amar2012	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Apur	scaff:13_52	Fox	Fox-OG39/NA	Forkhead domain E (410793)	Annotated as Fox-AB	
Bgla	XP_055873435.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Cang	XP_052699279.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Cari	EVM0018541.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Ceig	XP_011441298.1	Fox	Fox-OG39/NA	Forkhead domain E (410793)	Annotated as Fox-AB	
Cvfr	XP_022334048.1	Fox	Fox-OG39/NA	Forkhead domain E (410793)	Annotated as Fox-AB	
Dpol	XP_052278569.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Gaeig	XP_041375984.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Hbia	M00000015535	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Hruf	XP_046350707.2	Fox	Fox-OG39/NA	Forkhead domain E (410793)	Annotated as Fox-AB	
Hruf	XP_046350710.1	Fox	Fox-OG39/NA	Forkhead domain E (410793)	Annotated as Fox-AB	
Hruf	XP_046350686.2	Fox	Fox-OG39/NA	Forkhead domain E (410793)	Annotated as Fox-AB	
Hruf	XP_046350709.2	Fox	Fox-OG39/NA	Forkhead domain E (410793)	Annotated as Fox-AB	
Hruf	XP_046350688.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Hruf	XP_046350660.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Hruf	XP_046350687.2	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Hruf	XP_046350714.2	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Hruf	XP_046350712.2	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Hruf	XP_046350708.2	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Mare	XP_052795236.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Mcal	XP_052102496.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Mcor	CAC5414394.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Medu	CAG2193762.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Mgal	VD15942.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Mmar	MMAN00000023830	Fox	Fox-OG39/NA	Forkhead domain E (410793)	Annotated as Fox-AB	
Mmer	XP_045215157.2	Fox	Fox-OG39/NA	Forkhead domain E (410793)	Annotated as Fox-AB	
Mphi	scfa:1711_0.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Oedu	XP_048737442.2	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Pcan	XP_025078030.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Pmax	XP_033750901.1	Fox	Fox-OG39/NA	Forkhead domain E (410793)	Annotated as Fox-AB	
Pstr	KAK3601439.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Tgra	KAK3601419.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Sbro	s00198g393	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Pvir	XP_021357612.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Pyes	XP_060589081.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Rphi	EVM0003782.1	Fox	Fox-OG39/NA	Forkhead domain L1 (410801)	Annotated as Fox-AB	
Scon	Chr5_18	Fox	Fox-OG39/NA	Forkhead domain P2 (410839)	-	
Sglo	Sg_004401	Fox	Fox-OG39/NA	Forkhead domain P2 (410839)	-	
Tgra	KAJ3304916.1	Fox	Fox-OG39/NA	Forkhead domain P2 (410839)	-	
Airc	Contig120_132	Fox	Fox-P	Fox-P	-	
Apur	scaff:120_507.3	Fox	Fox-P	Fox-P	-	
Bgla	XP_055832746.1	Fox	Fox-P	Fox-P	-	
Cang	XP_02680529.1	Fox	Fox-P	Fox-P	-	
Cari	EVM0015348.1	Fox	Fox-P	Fox-P	-	
Cflu	DN101403_c0_2.13.p1	Fox	Fox-P	Fox-P	-	
Cgig	XP_01419350.2	Fox	Fox-P	Fox-P	-	
Cipl	DN80534_c6_g1.i4.p1	Fox	Fox-P	Fox-P	-	
Csin	Hic_asn_12.158	Fox	Fox-P	Fox-P	-	

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Pssm-ID)	Additional domains (Pssm-ID)	Notes
Cvir	XP_022295865.1	Fox	Fox-P	Forkhead domain P (4108397)	FOXP coiled-coil domain (465036)	
Dpol	XP_052252888.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Gaeq	XP_041347582.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Hruf	XP_046239143.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Lorb	DN59652.cl.g1.i4.p1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mare	XP_052780914.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mcal	XP_052101097.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mcal	XP_052083749.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mcal	XP_052100969.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mcal	XP_052061774.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mchi	DM46200.co.g1.i1.p1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mcor	CAC5419516.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Medu	CAG2199156.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mgal	VD14555.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mgal	VD150808.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mgal	VD150806.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mgal	VD150807.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mmar	MMAM00000012411.	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mmer	XP_05337678.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mmod	DN2151.co.g1.i2.p1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mner	g93547.tl	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Mphi	scfa:39347.0.7	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Obim	XP_052822821.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Oedu	XP_059998748.1	Fox	Fox-P	Forkhead domain P (4108397)	FOXP coiled-coil domain (465036)	
Osin	XP_036357888.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Pcan	XP_052106733.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Pcor	DN1820.co.g1.i26.p1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Pgen	DN3611.co.g1.i6.p1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Pmar	DN4268.co.g1.i1.p1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Phax	XP_033748371.1	Fox	Fox-P	Forkhead domain P (4108397)	FOXP coiled-coil domain (465036)	
Pour	DN88526.c2.g1.i1.p1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Pstr	KAK3589495.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Pvir	s01329g.124	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Pves	XP_021363304.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Rdec	DN8028.co.g1.i1.p1	Fox	Fox-P	Forkhead domain P (4108397)	FOXP coiled-coil domain (465036, partial)	
Rphi	XP_060586741.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Sbro	EVM0004295.1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Scon	Chr7:2129	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036, partial)	
Sglo	Sg011345	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Sgra	DN17101.co.g1.i13.p1	Fox	Fox-P	Forkhead domain P2 (410839)	FOXP coiled-coil domain (465036)	
Tgra	KAJ8302344.1	Fox	Fox-P	Forkhead domain Q2 (410839)	FOXP coiled-coil domain (465036)	
Acal	XP_005099459.2	Fox	Fox-Q2	Contig1420.28	Annotated as Fox-Q2a	
Arc	Ama2905	Fox	Fox-Q2	Hic-asym.16.4	Annotated as Fox-Q2a	
Amar	scaffold_332_35	Fox	Fox-Q2	BglA	Annotated as Fox-Q2a	
Apur	XP_055865367.1	Fox	Fox-Q2	XP_052699620.1	Annotated as Fox-Q2a	
BglA	XP_052699620.1	Fox	Fox-Q2	EVM0002665.1	Annotated as Fox-Q2a	
Cang	Cari	XP_011425762.2	Fox	Fox-Q2	Annotated as Fox-Q2a	
Cgig	Cphi	DN105612.co.g1.i1.p1	Fox	Fox-Q2	Annotated as Fox-Q2a	
Csin	Hic-asym.16.4	Fox	Fox-Q2	XP_022333968.1	Annotated as Fox-Q2a	
Cvir	XP_041363029.1	Fox	Fox-Q2	DPol	Annotated as Fox-Q2a	
Dpol	XP_052280806.1	Fox	Fox-Q2	Geag	Annotated as Fox-Q2a	
Gaeq	XP_041363041.1	Fox	Fox-Q2	XP_0000035328	Annotated as Fox-Q2a	
Hbfa	VD174621.1	Fox	Fox-Q2	M0000035328	Annotated as Fox-Q2a	
Hruf	XP_046373579.2	Fox	Fox-Q2	Mgal	Annotated as Fox-Q2a	
Mcal	XP_052101305.1	Fox	Fox-Q2	MMAR0000000686	Annotated as Fox-Q2a	
Mchi	DN22486.co.g1.i1.p1	Fox	Fox-Q2	XP_045215524.2	Annotated as Fox-Q2a	
Mcor	CAC5383792.1	Fox	Fox-Q2	g20553.tl	Annotated as Fox-Q2a	
Medu	CAG2191193.1	Fox	Fox-Q2	Scf:22910.1.1	Annotated as Fox-Q2a	
Mgal	MMAR0000000686	Fox	Fox-Q2	XP_04767584.1	Annotated as Fox-Q2a	
Mmar		Fox	Fox-Q2			
Mmer		Fox	Fox-Q2			
Mphi		Fox	Fox-Q2			
Obim		Fox	Fox-Q2			

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Pssm-ID)	Additional domains (Pssm-ID)	Notes
Pcan	XP_025078472.1	Fox	Fox-Q2	Forkhead domain Q2 (410809)	Annotated as Fox-Q2a	
Pcor	DN199235_c0_g1.i1.p1	Fox	Fox-Q2	Forkhead domain Q2 (410809)	Annotated as Fox-Q2a	
Pmar	DN49466_c0_g1.i1.p1	Fox	Fox-Q2	Forkhead domain Q2 (410809)	Annotated as Fox-Q2a	
Pmax	XP_033751003.1	Fox	Fox-Q2	Forkhead domain Q2 (410809)	Annotated as Fox-Q2a	
Pstr	KAK3395133.1	Fox	Fox-Q2	Forkhead domain Q2 (410809)	Annotated as Fox-Q2a	
Pvir	s00115g23	Fox	Fox-Q2	Forkhead domain Q2 (410809)	Annotated as Fox-Q2a	
Pyes	XP_0213424668.1	Fox	Fox-Q2	Forkhead domain Q2 (410809)	Annotated as Fox-Q2a	
Rphi	XP_060571531.1	Fox	Fox-Q2	Forkhead domain Q2 (410809)	Annotated as Fox-Q2a	
Sbro	EVM0023378.1	Fox	Fox-Q2	Forkhead domain Q2 (410809)	Annotated as Fox-Q2a	
Scn	Ch5.1974	Fox	Fox-Q2	Forkhead domain Q2 (410809)	Annotated as Fox-Q2a	
Scn	Ch5.2105	Fox	Fox-Q2	Forkhead domain Q2 (410809)	Annotated as Fox-Q2a	
Sgio	Sg009183	Fox	Fox-Q2	Forkhead domain Q2 (410809)	Annotated as Fox-Q2a	
Acal	XP_005109004.3	Fox	-	-	-	
Airc	Contig879.9	Fox	-	-	-	
Amar	Ama25952	Fox	-	-	-	
Amar	Amav23735	Fox	-	-	-	
Cang	XP_052676682.1	Fox	-	-	-	
Cang	XP_052680288.1	Fox	-	-	-	
Cang	XP_052677368.1	Fox	-	-	-	
Cari	EVM001935.1	Fox	-	-	-	
Cari	EVM0027332.1	Fox	-	-	-	
Cgig	XP_033068261.1	Fox	-	-	-	
Cgig	XP_011447567.2	Fox	-	-	-	
Cpli	DN157619.c0_g1.i1.p1	Fox	-	-	-	
Cvir	XP_022300767.1	Fox	-	-	-	
Cvir	XP_022300750.1	Fox	-	-	-	
Dpol	XP_052277921.1	Fox	-	-	-	
Dpol	XP_052227296.1	Fox	-	-	-	
Gaeq	XP_041366967.1	Fox	-	-	-	
Gaeq	XP_041378820.1	Fox	-	-	-	
Gaeq	XP_041347225.1	Fox	-	-	-	
Gaeq	XP_041375925.1	Fox	-	-	-	
Gaeq	XP_041375913.1	Fox	-	-	-	
Gaeq	XP_041379015.1	Fox	-	-	-	
Gaeq	M0000018946	Fox	-	-	-	
Hbia	XP_052791886.1	Fox	-	-	-	
Mare	XP_052771066.1	Fox	-	-	-	
Mare	XP_052098820.1	Fox	-	-	-	
Mcal	MMA00000049704.	Fox	-	-	-	
Mcor	CAC5419379.1	Fox	-	-	-	
Medu	CAG2194707.1	Fox	-	-	-	
Medu	CAG2208945.1	Fox	-	-	-	
Mgal	VD15902.1	Fox	-	-	-	
Mgal	VD152978.1	Fox	-	-	-	
Mmar	XP_045210636.2	Fox	-	-	-	
Mmer	XP_045189131.2	Fox	-	-	-	
Oedu	gl59704_t1	Fox	-	-	-	
Mner	XP_036359188.1	Fox	-	-	-	
Mphi	scfa.46189.0.0	Fox	-	-	-	
Mphi	scfa.15444.0.2	Fox	-	-	-	
Mphi	scfa.27787.1.10	Fox	-	-	-	
Obim	XP_014777604.1	Fox	-	-	-	
Pcor	DN115905_c0_g1.i1.p1	Fox	-	-	-	
Pcor	XP_033750561.1	Fox	-	-	-	
Pmax	KAK3385306.1	Fox	-	-	-	
Pstr	s00585g48	Fox	-	-	-	
Pyes	XP_021348419.1	Fox	-	-	-	
Rdec	DN23525_c0_g1.i1.p1	Fox	-	-	-	
Rphi	XP_060585776.1	Fox	-	-	-	
Scn	Ch5.397	Fox	-	-	-	
Sgio	Sg01575	Fox	-	-	-	
Tsequ	DN32960_c0_g1.i1.p1	Fox	-	-	-	
Acal	XP_033824685.1	Sox	-	-	-	

Helix loop helix domain (19764)

High mobility group box (43830)

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Psm-ID)	Additional domains (Psm-ID)	Notes
Acal	XP_012946205_1	Sox	-	High mobility group box (438820)	-	
Acal	XP_005105939_1	Sox	-	High mobility group box (438820)	-	
Apec	DN48806_0_cgl.i1.p1	Sox	-	High mobility group box (438820)	-	
Apec	DN108003_cgl.i1.p1	Sox	-	High mobility group box (438820)	-	
scaffid	391_70	Sox	-	High mobility group box (438820)	-	
Apur	XP_013078241_2	Sox	-	High mobility group box (438820)	-	
Bgfa	XP_013078156_1	Sox	-	High mobility group box (438820)	-	
Bgfa	XP_052697281_1	Sox	-	High mobility group box (438820)	-	Helix loop helix domain (197674)
Cang	XP_052713622_1	Sox	-	High mobility group box (438820)	-	Helix loop helix domain (197674, partial)
Carl	EVM0018891_1	Sox	-	High mobility group box (438820)	-	
Cari	EVM0005567_1	Sox	-	High mobility group box (438820)	-	Helix loop helix domain (197674)
Cgig	XP_014258692_2	Sox	-	High mobility group box (438820)	-	CW-type Zinc Finger (46218)
Cgig	XP_034335819_1	Sox	-	High mobility group box (438820)	-	Helix loop helix domain (197674)
Cvir	XP_022330758_1	Sox	-	High mobility group box (438820)	-	
Cvir	XP_022339079_1	Sox	-	High mobility group box (438820)	-	
Dpov	XP_052271004_1	Sox	-	High mobility group box (438820)	-	
Gaeq	XP_041377139_1	Sox	-	High mobility group box (438820)	-	
Hbia	M000000038049	Sox	-	High mobility group box (438820)	-	
Hbia	M00000004998	Sox	-	High mobility group box (438820)	-	
Hruf	XP_046329595_1	Sox	-	High mobility group box (438820)	-	
Hruf	XP_048236511_1	Sox	-	High mobility group box (438820)	-	
Mcor	CAC384832_1	Sox	-	High mobility group box (438820)	-	
Medu	CAG2253429_1	Sox	-	High mobility group box (438820)	-	
Mgal	VD178477_1	Sox	-	High mobility group box (438820)	-	
Mmod	DN13112_c0.g1.i1.p1	Sox	-	High mobility group box (438820)	-	
Obim	XP_0477859_1	Sox	-	High mobility group box (438820)	-	
Oedu	XP_048738250_1	Sox	-	High mobility group box (438820)	-	
Oedu	XP_048752144_2	Sox	-	High mobility group box (438820)	-	
Osin	XP_036358200_1	Sox	-	High mobility group box (438820)	-	
Osin	XP_029654541_1	Sox	-	High mobility group box (438820)	-	
Osin	XP_0296565568_1	Sox	-	High mobility group box (438820)	-	
Osin	XP_029657644_1	Sox	-	High mobility group box (438820)	-	
Osin	XP_029656220_1	Sox	-	High mobility group box (438820)	-	Helix loop helix domain (197674)
Osin	XP_029657648_1	Sox	-	High mobility group box (438820)	-	
Osin	XP_029655036_1	Sox	-	High mobility group box (438820)	-	
Osin	XP_029655785_1	Sox	-	High mobility group box (438820)	-	
Osin	XP_029656129_1	Sox	-	High mobility group box (438820)	-	
Osin	XP_029654991_1	Sox	-	High mobility group box (438820)	-	
Pcan	XP_025104729_1	Sox	-	High mobility group box (438820)	-	
Pcor	DN32781_c0.g1.i1.p1	Sox	-	High mobility group box (438820)	-	
Pcor	DN19364_c0.g1.i2.p1	Sox	-	High mobility group box (438820)	-	
Pmar	DN33290_c0.g1.i2.p1	Sox	-	High mobility group box (438820)	-	
Pmar	DN35008_c0.g1.i4.p1	Sox	-	High mobility group box (438820)	-	
Pmax	XP_033758821_1	Sox	-	High mobility group box (438820)	-	
Ppur	DN4784_c0.g1.i4.p1	Sox	-	High mobility group box (438820)	-	
Pyes	XP_021347051_1	Sox	-	High mobility group box (438820)	-	
Sbro	EVM0018824_1	Sox	-	High mobility group box (438820)	-	
Sglo	Sg1009175	Sox	-	High mobility group box (438820)	-	
Sglo	Sg1012029	Sox	-	High mobility group box (438820)	-	
Tseq	DN639_c0.g1.i1.p1	Sox	-	High mobility group box (438820)	-	Helix loop helix domain (197674)
Acal	XP_005108230_1	Sox	-	High mobility group box B (438790)	-	
Acal	XP_03624438_1	Sox	-	High mobility group box B (438790)	-	
Contig49_126	contig49_126	Sox	-	High mobility group box B (438790)	-	
Airc	Contig14_115	Sox	-	High mobility group box B (438790)	-	
Amar	Ama33032	Sox	-	High mobility group box B (438790)	-	
Amar	Ama33828	Sox	-	High mobility group box B (438790)	-	
Amar	DN29410_c0.g1.i2.p1	Sox	-	High mobility group box B (438790)	-	
Apec	DN12297_c0.g1.i3.p1	Sox	-	High mobility group box B (438790)	-	
Apec	scaffid_15389_10	Sox	-	High mobility group box B (438790)	-	
Apur	scaffid_865_4	Sox	-	High mobility group box B (438790)	-	
Bgfa	XP_03075432_1	Sox	-	High mobility group box B (438790)	-	
Cang	XP_053868106_1	Sox	-	High mobility group box B (438790)	-	
Cang	XP_052706368_1	Sox	-	High mobility group box B (438790)	-	
Cari	XP_02705551_1	Sox	-	High mobility group box B (438790)	-	
Cari	EVM0013965_1	Sox	-	High mobility group box B (438790)	-	
Cflu	DN18670_c2.g1.i2.p1	Sox	-	High mobility group box B (438790)	-	
Cflu	DN98542_c1.g1.i1.p1	Sox	-	High mobility group box B (438790)	-	
Cgig	XP_01433975_1	Sox	-	High mobility group box B (438790)	-	

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Psm-ID)	Additional domains (Psm-ID)	Notes
Cgg	XP_011455662.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Cpli	DN31343_c0_g1.i1.p1	Sox	Sox-B1/2	High mobility group box B (438790)		
Cpli	DN95511_c0_g1.i1.p1	Sox	Sox-B1/2	High mobility group box B (438790)		
Csin	Hic.asm.6.930	Sox	Sox-B1/2	High mobility group box B (438790)		
Csin	Hic.asm.6.233.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Cvir	XP_022286516.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Cvir	XP_022342320.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Cvir	XP_052214544.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Dpol	XP_052217420.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Gaeq	XP_041353075.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Gaeq	XP_041357874.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Hbia	M00000002798	Sox	Sox-B1/2	High mobility group box B (438790)		
Hbia	M00000033682	Sox	Sox-B1/2	High mobility group box B (438790)		
Hruf	XP_046370193.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Hruf	XP_046326733.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Lorb	DN80278_c0_g1.i1.p1	Sox	Sox-B1/2	High mobility group box B (438790)		
Lorb	DN14_c4_g1.i1.p1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mare	XP_052784929.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mare	XP_052784720.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mcal	XP_052105617.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mcchi	XP_052104911.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mchi	DN33632_c0_g1.i2.p1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mchi	DN45716_c0_g1.i1.p1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mcor	CAC5401077.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mcor	CAC5413203.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Medu	CAG229644.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Medu	CAG2206403.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mgal	VD33296.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mgal	VD16960.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mmar	MMAM00000041532	Sox	Sox-B1/2	High mobility group box B (438790)		
Mmar	MMAM00000023253	Sox	Sox-B1/2	High mobility group box B (438790)		
Mmer	XP_045201594.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mmer	XP_045201080.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mmod	DN1981_c0_g1.i2.p1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mmod	DN73279_c0_g1.i1.p1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mner	g140596_t1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mner	g157489_t1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mphi	scfa_663490.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Mphi	scfa_24206_0.4	Sox	Sox-B1/2	High mobility group box B (438790)		
Obim	XP_014738997.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Obim	XP_014780771.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Oedu	XP_048746651.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Oedu	XP_048746663.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Osin	XP_029654000.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Osin	XP_029655838.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Pcan	XP_025079293.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Pcan	XP_025078598.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Pcor	DN9587_c0_g1.i2.p1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pcor	DN14753_c0_g1.i2.p1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pcor	DN5688_c0_g1.i10.p2	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pcor	DN5688_c0_g3.i1.p1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pcor	DN9742_c0_g1.12.p1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pgen	DN5232_c0_g1.i1.p1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pgen	DN1199_c0_g1.i12.p1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pmar	DN30477_c0_g2.i1.p1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pmar	DN30459_c0_g1.i1.p1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pmax	XP_033759382.1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pmax	DN51870_c1_g1.i2.p1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Poku	DN1067_c0_g1.i1.p1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Poku	DN202737_c0_g1.i1.p1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Ppur	KAK3386311.1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pstr	KAK3389936.1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pvir	s00319g159	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pvir	s00319g281	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pyes	XP_021356125.1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pyes	XP_021344413.1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Pyes	XP_021372128.1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		
Rdec	DN21477_c2_g7.i2.p1	Sox	Sox-B1/2	High mobility group box A, B and C (438837)		

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Psm-ID)	Additional domains (Psm-ID)	Notes
Rphi	XP_060556101.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Rphi	XP_060561544.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Sbro	EVM0016386.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Sbro	EVM0007529.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Scon	Ch9-1352	Sox	Sox-B1/2	High mobility group box B (438790)		
Scon	Ch9-1522	Sox	Sox-B1/2	High mobility group box B (438790)		
Scon	Ch9-1514	Sox	Sox-B1/2	High mobility group box B (438790)		
Sgio	Sg101000	Sox	Sox-B1/2	High mobility group box B (438790)		
Sgio	Sg1020107	Sox	Sox-B1/2	High mobility group box B (438790)		
Sgra	DN3782_c0_g1.i3.p1	Sox	Sox-B1/2	High mobility group box B (438790)		
Sgra	DN357_c1_g1.i1.p1	Sox	Sox-B1/2	High mobility group box B (438790)		
Tgra	KAJ8310140.1	Sox	Sox-B1/2	High mobility group box B (438790)		
Tsqu	DN97880_c0_g1.i1.p1	Sox	Sox-B1/2	High mobility group box B (438790)		
Tsqu	DN50556_c0_g1.i2.p1	Sox	Sox-B1/2	High mobility group box B (438790)		
Airc	Contig807.70	Sox	Sox-C	High mobility group box C (438838)		
Amar	Ama12726	Sox	Sox-C	High mobility group box C (438838)		
Apec	DN12286_c0_g3.i1.p1	Sox	Sox-C	High mobility group box C (438838)		
Apur	scaffold_16_61	Sox	Sox-C	High mobility group box C (438838)		
Cang	XP_052689209.1	Sox	Sox-C	High mobility group box C (438838)		
Cari	EVMO025346.1	Sox	Sox-C	High mobility group box C (438838)		
Cflu	DN126276_c0_g1.i1.p1	Sox	Sox-C	High mobility group box C (438838)		
Cgig	XP_011445203.1	Sox	Sox-C	High mobility group box C (438838)		
Cipl	DN19112_c0_g1.i1.p1	Sox	Sox-C	High mobility group box C (438838)		
Csin	Hic_asn_11_1009	Sox	Sox-C	High mobility group box C (438838)		
Cvir	XP_022317619.1	Sox	Sox-C	High mobility group box C (438838)		
Dpol	XP_052257395.1	Sox	Sox-C	High mobility group box C (438838)		
Gaeg	XP_041358324.1	Sox	Sox-C	High mobility group box C (438838)		
Hbia	M0000037669	Sox	Sox-C	High mobility group box C (438838)		
Hruf	XP_046365064.1	Sox	Sox-C	High mobility group box C (438838)		
Lorb	DN14941_c0_g1.i1.p1	Sox	Sox-C	High mobility group box C (438838)		
Mare	XP_052777703.1	Sox	Sox-C	High mobility group box C (438838)		
Mcal	XP_052087802.1	Sox	Sox-C	High mobility group box C (438838)		
Mchi	DM4798_c0_g4.i1.p1	Sox	Sox-C	High mobility group box C (438838)		
Mcor	CAC5424030.1	Sox	Sox-C	High mobility group box C (438838)		
Medu	CAG2189937.1	Sox	Sox-C	High mobility group box C (438838)		
Mgal	VD14153.1	Sox	Sox-C	High mobility group box C (438838)		
Mgal	VD14462.1	Sox	Sox-C	High mobility group box C (438838)		
Mmar	MMWA0000036315	Sox	Sox-C	High mobility group box C (438838)		
Mmer	XP_045158937.1	Sox	Sox-C	High mobility group box C (438838)		
Mmod	DN104308_c0_g1.i1.p1	Sox	Sox-C	High mobility group box C (438838)		
Mmer	g26404.t1	Sox	Sox-C	High mobility group box C (438838)		
Mphi	scf_17954.1.5	Sox	Sox-C	High mobility group box C (438838)		
Oedu	XP_048762549.1	Sox	Sox-C	High mobility group box C (438838)		
Osin	XP_029654195.1	Sox	Sox-C	High mobility group box C (438838)		
Pean	XP_025110204.1	Sox	Sox-C	High mobility group box C (438838)		
Pcor	DN2429_c2_g1.i2.p1	Sox	Sox-C	High mobility group box C (438838)		
Pcor	DN929_c2_f1.i1.p1	Sox	Sox-C	High mobility group box C (438838)		
Pcor	DN2572_c0_g1.i1.p1	Sox	Sox-C	High mobility group box C (438838)		
Pcor	DN353_c2_g3.i1.p1	Sox	Sox-C	High mobility group box C (438838)		
Pgen	DN738_c0_g1.i1.p1	Sox	Sox-C	High mobility group box C (438838)		
Pmar	DN29124_c0_g2.i1.p1	Sox	Sox-C	High mobility group box C (438838)		
Pmax	XP_033737425.1	Sox	Sox-C	High mobility group box C (438838)		
Poku	DN71015_c0_g2.i1.p1	Sox	Sox-C	High mobility group box C (438838)		
Ppur	DN88859_c0_g1.i1.p1	Sox	Sox-C	High mobility group box C (438838)		
Pstr	KAK3610995.1	Sox	Sox-C	High mobility group box C (438838)		
Pvir	s00145g243	Sox	Sox-C	High mobility group box C (438838)		
Pys	XP_021356242.1	Sox	Sox-C	High mobility group box C (438838)		
Rdec	DN52924_c0_g1.i1.p1	Sox	Sox-C	High mobility group box C (438838)		
Rphi	XP_06055827.1	Sox	Sox-C	High mobility group box C (438838)		
Sbro	EVM0006311.1	Sox	Sox-C	High mobility group box C (438838)		
Scon	Ch8-1790	Sox	Sox-C	High mobility group box C (438838)		
Sgio	Sg1000072	Sox	Sox-C	High mobility group box C (438838)		
Sgra	KAJ8306264.1	Sox	Sox-C	High mobility group box C (438838)		
Tgra	KAJ8306264.1	Sox	Sox-C	High mobility group box C (438838)		
Tsqu	DN11669_c1_g1.i2.p1	Sox	Sox-D	High mobility group box (438839)		
Acal	XP_03824396.1	Sox	Sox-D	High mobility group box (438839)		
Airc	Contig290.5.1	Sox	Sox-D	High mobility group box (438839)		
Amar	Ama23921	Sox	Sox-D	High mobility group box (438839)		

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Psm-ID)	Additional domains (Psm-ID)	Notes
Apec	DN1990_c0_g1.i10.p1	Sox	Sox-D	High mobility group box (438339)		
Apur	scaffold_393.10	Sox	Sox-D	High mobility group box (438339)		
Bgia	XP_055899647.1	Sox	Sox-D	High mobility group box (438339)		
Cari	EVM0012405.1	Sox	Sox-D	High mobility group box (438339)		
Cflu	DN124582_c0.g1..i15.p1	Sox	Sox-D	High mobility group box (438339)		
Cgig	XP_001425377.1	Sox	Sox-D	High mobility group box (438339)		
Cph	DN64448_c0.g1..i1.p1	Sox	Sox-D	High mobility group box (438339)		
Csin	Hic-asn_2.1656	Sox	Sox-D	High mobility group box (438339)		
Cvir	Hi-casn_2.1600.2	Sox	Sox-D	High mobility group box (438339)		
Dpol	XP_022302926.1	Sox	Sox-D	High mobility group box (438339)		
Dpol	XP_052213125.1	Sox	Sox-D	High mobility group box (438339)		
Gaeq	XP_0041367101.1	Sox	Sox-D	High mobility group box (438339)		
Hbia	M00000014008	Sox	Sox-D	High mobility group box (438339)		
Hruf	XP_0463290046.1	Sox	Sox-D	High mobility group box (438339)		
Lorb	DN15537_c0.g2.i3.p1	Sox	Sox-D	High mobility group box (438339)		
Mare	XP_052800695.1	Sox	Sox-D	High mobility group box (438339)		
Mcal	XP_052065962.1	Sox	Sox-D	High mobility group box (438339)		
Mchi	DN3691_c1.g1..i4.p1	Sox	Sox-D	High mobility group box (438339)		
Mcor	CAC5366270.1	Sox	Sox-D	High mobility group box (438339)		
Medu	CAG2197887.1	Sox	Sox-D	High mobility group box (438339)		
Mgal	VD47525.1	Sox	Sox-D	High mobility group box (438339)		
Mgal	VD147529.1	Sox	Sox-D	High mobility group box (438339)		
Mgal	VD147528.1	Sox	Sox-D	High mobility group box (438339)		
Mgal	VD47527.1	Sox	Sox-D	High mobility group box (438339)		
Mgal	VD47526.1	Sox	Sox-D	High mobility group box (438339)		
Mgal	VD147530.1	Sox	Sox-D	High mobility group box (438339)		
Mmar	MMAW000000004319	Sox	Sox-D	High mobility group box (438339)		
Mmer	XP_053384999.1	Sox	Sox-D	High mobility group box (438339)		
Mmod	DN588_c0.g1..i9.p1	Sox	Sox-D	High mobility group box (438339)		
Mner	g103_147..t2	Sox	Sox-D	High mobility group box (438339)		
Mphi	scaf_42181..1..3	Sox	Sox-D	High mobility group box (438339)		
Obim	XP_052828391.1	Sox	Sox-D	High mobility group box (438339)		
Oedu	XP_048779633.1	Sox	Sox-D	High mobility group box (438339)		
Osin	XP_029644081.1	Sox	Sox-D	High mobility group box (438339)		
Pcan	XP_025088657.1	Sox	Sox-D	High mobility group box (438339)		
Pcor	DN353_c2.g2..i1.p1	Sox	Sox-D	High mobility group box (438339)		
Pcor	DN1386_c1..g1..i4.p1	Sox	Sox-D	High mobility group box (438339)		
Pcor	DN13571_c0..g1..i1.p1	Sox	Sox-D	High mobility group box (438339)		
Pcor	DN1386_c1..g2..i2.p1	Sox	Sox-D	High mobility group box (438339)		
Pgen	DN24654_c0..g1..i2.p1	Sox	Sox-D	High mobility group box (438339)		
Pmar	DN40112_c0..g1..i4.p1	Sox	Sox-D	High mobility group box (438339)		
Pmax	XP_033751614.1	Sox	Sox-D	High mobility group box (438339)		
Poku	DN371_c1..g4..i1.p1	Sox	Sox-D	High mobility group box (438339)		
Ppur	DN33319_c0..g1..i1.p1	Sox	Sox-D	High mobility group box (438339)		
Pstr	KAK3605548.1	Sox	Sox-D	High mobility group box (438339)		
Pvir	s00219g102	Sox	Sox-D	High mobility group box (438339)		
Pves	XP_021368061.1	Sox	Sox-D	High mobility group box (438339)		
Rdec	DN8993_c0..g1..i1.p1	Sox	Sox-D	High mobility group box (438339)		
Rphi	XP_05606041..i10..1	Sox	Sox-D	High mobility group box (438339)		
Sbro	EVM0000795.1	Sox	Sox-D	High mobility group box (438339)		
Scen	Ch14..562..1	Sox	Sox-D	High mobility group box (438339)		
Sgra	DN16138_c0..g1..i16..p1	Sox	Sox-D	High mobility group box (438339)		
Tgra	KAJ3298781.1	Sox	Sox-D	High mobility group box (438339)		
Tsqu	DN50531_c0..g1..i1.p1	Sox	Sox-E	High mobility group box E (438340)		
Acal	XP_005102100.1	Sox	Sox-E	High mobility group box E (438340)		
Airc	Contig52..209	Sox	Sox-E	High mobility group box E (438340)		
Amar	Amar01107	Sox	Sox-E	High mobility group box E (438340)		
Apec	DN4330_c0..g1..i1.p1	Sox	Sox-E	High mobility group box E (438340)		
Apur	scaffold_488..7	Sox	Sox-E	High mobility group box E (438340)		
Bgia	XP_013091187.2	Sox	Sox-E	High mobility group box E (438340)		
Cang	XP_052688335..1	Sox	Sox-E	High mobility group box E (438340)		
Cari	EVM00005846..1	Sox	Sox-E	High mobility group box E (438340)		
Cflu	DN110407_c5..g2..i1..p1	Sox	Sox-E	High mobility group box E (438340)		
Cgig	NP_001293801.1	Sox	Sox-E	High mobility group box E (438340)		
Cphi	DN71393_c0..g2..i1..p1	Sox	Sox-E	High mobility group box E (438340)		
Cphi	DN71393_c0..g1..i2..p1	Sox	Sox-E	High mobility group box E (438340)		
Csin	Hic-asn_0..353	Sox	Sox-E	High mobility group box E (438340)		
Cvir	XP_022312895..1	Sox	Sox-E	High mobility group box E (438340)		
Dpol	XP_0522264587..1	Sox	Sox-E	High mobility group box E (438340)		

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Psm-ID)	Additional domains (Psm-ID)	Notes
Gaegeg	XP_041362638.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Hbia	M0000012324	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586; partial)	
Hbia	M0000012325	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586; partial)	
Hruf	XP_0463593661	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586; partial)	
Mare	XP_052786944.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586; partial)	
Mare	XP_052783666.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586; partial)	
Mcal	XP_052068536.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Mchi	DN42011_c0_g1.i2.p1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Mmod	CAC5402442.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Mphi	XP_056019113.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586; partial)	
Oedu	XP_025091262.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Pcan	VD182092.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Mgal	VD182090.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586; partial)	
Mmar	MMA00000042410	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586; partial)	
Mmer	XP_045213705.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Mmod	DN78330_c0_g1.i1.p1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Mphi	scf.25.414.0.6	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Oedu	XP_056019113.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586; partial)	
Pcor	DN4274_c0_g1.i3.p1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Pcor	DN96098_c0_g1.i1.p1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Pmar	DN30335_c0_g1.i1.p1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Poku	DN81807_c0_g1.i7.p1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Pour	DN46000_c0_g1.i1.p1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Pstr	KAK3610785.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Pvir	sl.13688467.4	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Pyes	XP_021348843.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586; partial)	
Rphi	XP_060604697.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Sbro	EVM0002110.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Scor	Chr1.75	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Sglo	Sg024297	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586; partial)	
Sgra	DN24263_c0_g1.i1.p1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586)	
Tgra	KAJ83317914.1	Sox	Sox-E	High mobility group box E (438840)	Sox developmental protein N terminal (463586; partial)	
Tsqu	DN8973_c2_g1.i2.p1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Acal	XP_05107482.1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Arc	Contig80.101	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Amar	Ama11616	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Apur	scaffold-546_32	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Bglia	XP_013074628.2	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Cang	XP_0526883434.1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Cari	EVM0006823.1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Cflu	DN1399006_c0.l1.11.p1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Cggg	XP_011448074.2	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Cipli	DN4414_c0.g1.i1.p1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Csin	Hic_asm_11.549	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Cvir	XP_022319962.1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Dpol	XP_0522774104.1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Gaegeg	XP_041359436.1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Hbia	M0000015459	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Hruf	XP_046357912.1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Mare	XP_052774544.1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Mare	XP_052774361.1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Mcal	XP_052061059.1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Mgal	CAC5414609.1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Medu	CAG2242031.1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Mmod	DN80495_c0_g1.i1.p1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Mnre	g115494_t1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Mphi	scf.61.114.0.13	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Obim	XP_052825684.1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Oedu	XP_048764319.2	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	
Pcan	XP_025109598.1	Sox	Sox-F	High mobility group box F (438840)	Sox developmental protein N terminal (463586)	

Tab. S6 continued from previous page

Sp ID	Gene ID	Group	Annotation	Main catalytic domain (Psm-ID)	Additional domains (Psm-ID)	Notes
Pcor	DN11375_c0_g1.i1.p1	Sox	Sox-F	High mobility group box F (438841)		
Pcor	DN29649_c0_g1.i3.p1	Sox	Sox-F	High mobility group box F (438841)		
Pcor	DN5688_c2_g1.i3.p1	Sox	Sox-F	High mobility group box F (438841)		
Pgen	DN144332_c0_g1.i1.p1	Sox	Sox-F	High mobility group box F (438841)		
Pmar	DN24748_c0_g1.i1.p1	Sox	Sox-F	High mobility group box F (438841)		
Pmax	XP_033738287_1	Sox	Sox-F	High mobility group box F (438841)		
Poku	DN14229_c1_g2.i1.p1	Sox	Sox-F	High mobility group box F (438841)		
Pstr	KAK3383243_1	Sox	Sox-F	High mobility group box F (438841)		
Pvir	s00137g284	Sox	Sox-F	High mobility group box F (438841)		
Rdec	XP_021378109_1	Sox	Sox-F	High mobility group box F (438841)		
Rphi	DN53443_c0_g1.i1.p1	Sox	Sox-F	High mobility group box F (438841)		
Sbro	XP_060559438_1	Sox	Sox-F	High mobility group box F (438841)		
Scon	EVM0000861_1	Sox	Sox-F	High mobility group box F (438841)		
Sglo	Ch8-897	Sox	Sox-F	High mobility group box F (438841)		
Sgra	Sglo1005442	Sox	Sox-F	High mobility group box F (438841)		
Sgra	DN5013_c3_g1.i7.p1	Sox	Sox-F	High mobility group box F (438841)		
Airc	Contig1525_38	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Amar	Ama26724	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Apec	DN93182_c0_g1.i1.p1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Apur	scaffold768_3	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Cang	XP_052703370_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Cari	EVM0018164_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Cggg	XP_011415859_3	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Cphi	DN80002_c0_g1.i1.p1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Csin	Hic_asm_15_1471	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Cvir	XP_022338738_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Dpol	XP_052226448_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Gaeq	XP_041370217_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Gaeq	XP_041369137_1	Sox	Sox-H	High mobility group box Sox-30 (438820)		
Hbia	M00000001184	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Hruf	XP_046358520_2	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Mcal	XP_052099860_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Mcor	CAC540604_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Medu	CAG2257203_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Mgal	VD130324_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Mgal	VD130323_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Mmar	MMAAM0000015662	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Mmer	XP_053407277_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Mmer	g125_224_t1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Mphi	scaf12010_0_4	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Mphi	scf:59202_0_9	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Obim	XP_052832677_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Oedu	XP_056006679_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Osin	XP_035368794_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Pcor	DN186446_c0_g1.i1.p1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Pcor	DN40950_c1_g1.i2.p1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Pmax	XP_033735683_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Poku	DN16718_c0_g1.i6.p1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Ppur	DN7268_c0_g1.i1.p1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Pstr	KAK3382760_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Pvir	s00451g108	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Pyes	XP_021340986_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Rdec	DN22482_c4_g1.i1.p1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Rphi	XP_060578490_1	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Scor	Ch15_1899	Sox	Sox-H	High mobility group box Sox-30 (438820)		
Sglo	Sglo010047	Sox	Sox-H	High mobility group box Sox-30 (438842)		
Tequ	DN874_c7_g1.i1.p1	Sox	Sox-H	High mobility group box Sox-30 (438842)		

Proportions of missing data in both DSFGs and bivalve species. Bivalve species represented by transcriptomic data are high

tions of missing data in both DSFGs and bivalve species

Species		Genes	
Species	% missing data (out of 33 DSFGs)	Group	% missing data (out of 43 bivalve species)
<i>A. irradians concentricus</i>	0.000000	<i>Dmrt-1L</i>	
<i>A. marissinica</i>	21.212121	<i>Dmrt-3</i>	
<i>A. pectinata*</i>	48.484848	<i>Dmrt-2</i>	
<i>A. purpuratus</i>	6.060606	<i>Dmrt-4/5</i>	
<i>C. angulata</i>	6.060606	<i>Fox-A</i>	
<i>C. ariakensis</i>	3.030303	<i>Fox-B</i>	
<i>C. fluminea*</i>	42.424242	<i>Fox-C</i>	
<i>C. gigas</i>	6.060606	<i>Fox-D</i>	
<i>C. plicata*</i>	21.212121	<i>Fox-E</i>	
<i>C. sinensis</i>	21.212121	<i>Fox-F</i>	
<i>C. virginica</i>	3.030303	<i>Fox-G</i>	
<i>D. polymorpha</i>	9.090909	<i>Fox-H</i>	
<i>H. bialata</i>	9.090909	<i>Fox-J1</i>	
<i>L. orbiculatus*</i>	63.636364	<i>Fox-J2/3</i>	
<i>M. arenaria</i>	21.212121	<i>Fox-L1</i>	
<i>M. californianus</i>	9.090909	<i>Fox-L2</i>	
<i>M. chinensis*</i>	57.575758	<i>Fox-N1/4</i>	
<i>M. coruscus</i>	0.000000	<i>Fox-N2/3</i>	
<i>M. edulis</i>	3.030303	<i>Fox-O</i>	
<i>M. galloprovincialis</i>	6.060606	<i>Fox-P</i>	
<i>M. margaritifera</i>	6.060606	<i>Fox-Q2</i>	
<i>M. mercenaria</i>	3.030303	<i>Fox-OG13/NA</i>	
<i>M. modiolus*</i>	36.363636	<i>Fox-OG15/NA</i>	
<i>M. nervosa</i>	27.272727	<i>Fox-OG16/NA</i>	
<i>M. philippinarum</i>	9.090909	<i>Fox-OG2/NA</i>	
<i>O. edulis</i>	6.060606	<i>Fox-OG28/NA</i>	
<i>P. coreanum*</i>	18.181818	<i>Fox-OG39/NA</i>	
<i>P. generosa*</i>	54.545455	<i>Sox-B1/2</i>	
<i>P. margaritifera*</i>	21.212121	<i>Sox-C</i>	
<i>P. maximus</i>	0.000000	<i>Sox-D</i>	
<i>P. okutanii*</i>	54.545455	<i>Sox-E</i>	
<i>P. purpuratus*</i>	54.545455	<i>Sox-F</i>	
<i>P. streckersoni</i>	6.060606	<i>Sox-H</i>	
<i>P. viridis</i>	3.030303		
<i>P. yessoensis</i>	3.030303		
<i>R. decussatus*</i>	51.515152		
<i>R. philippinarum</i>	3.030303		
<i>S. broughtonii</i>	12.121212		
<i>S. constricta</i>	12.121212		
<i>S. glomerata</i>	9.090909		
<i>S. grandis*</i>	54.545455		
<i>T. granosa</i>	42.424242		
<i>T. squamosa*</i>	48.484848		

Supplementary Table S8 – Complete set of DSFGs in mammals. For each gene, the species ID (Sp. ID) as in **Tab. S4**, the accession number (Gene ID), and the Possvm-based annotation are indicated.

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Hamp	XP_057601513.1	Dmrt	Dmrt-B1	Cimi	XP_017353003.1	Fox	Fox-C1
Hsap	NP_149056.1	Dmrt	Dmrt-B1	Oana	XP_028912285.1	Fox	Fox-C1
Oafe	XP_007944154.1	Dmrt	Dmrt-B1	Dnov	XP_058141148.1	Fox	Fox-C1
Amel	XP_0345008822.1	Dmrt	Dmrt-B1	Bbub	XP_025120521.1	Fox	Fox-C1
Bbub	XP_025144486.2	Dmrt	Dmrt-B1	Drot	XP_053776986.1	Fox	Fox-C1
Casi	XP_006839931.1	Dmrt	Dmrt-B1	Rfer	XP_032971521.1	Fox	Fox-C1
Pgig	XP_039739094.1	Dmrt	Dmrt-B1	Mdom	XP_007488114.2	Fox	Fox-C1

Tab. S8 continued from previous page

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Cimi	XP_017359902.1	Dmrt	Dmrt-B1	Casi	XP_006870626.1	Fox	Fox-C1
Mmus	XP_036020151.1	Dmrt	Dmrt-B1	Pafn	XP_047652737.1	Fox	Fox-C1
Bmus	XP_036712433.1	Dmrt	Dmrt-B1	Bmus	XP_036726614.1	Fox	Fox-C1
Oana	XP_039770576.1	Dmrt	Dmrt-B1	Hamp	XP_057555406.1	Fox	Fox-C1
Scar	XP_047395630.1	Dmrt	Dmrt-B1	Lcat	XP_045407522.1	Fox	Fox-C1
Mjav	XP_036877684.1	Dmrt	Dmrt-B1	Opri	XP_058524289.1	Fox	Fox-C1
Ptig	XP_042853169.1	Dmrt	Dmrt-B1	Cdid	XP_037699408.1	Fox	Fox-C1
Lcat	XP_045402503.1	Dmrt	Dmrt-B1	Hsap	NP_001444.2	Fox	Fox-C1
Emax	XP_049731502.1	Dmrt	Dmrt-B1	Mjav	XP_036867643.1	Fox	Fox-C1
Ttru	XP_004325875.3	Dmrt	Dmrt-B1	Scar	XP_047413053.1	Fox	Fox-C1
Mang	XP_045755047.1	Dmrt	Dmrt-B1	Cdro	XP_031291159.1	Fox	Fox-C1
Pafn	XP_047647463.1	Dmrt	Dmrt-B1	Mmus	NP_032618.2	Fox	Fox-C1
Opri	XP_004588846.2	Dmrt	Dmrt-B1	Emax	XP_049738278.1	Fox	Fox-C1
Cpor	XP_003464114.1	Dmrt	Dmrt-B1	Oafe	XP_007933566.1	Fox	Fox-C1
Dnov	XP_058159531.1	Dmrt	Dmrt-B1	Mang	XP_045722432.1	Fox	Fox-C1
Cdid	XP_037665497.1	Dmrt	Dmrt-B1	Ttru	XP_019800382.2	Fox	Fox-C1
Equa	XP_046516609.1	Dmrt	Dmrt-B1	Oana	XP_028909462.1	Fox	Fox-C1
Mdom	XP_056671081.1	Dmrt	Dmrt-B1	Equa	XP_046496475.1	Fox	Fox-C1
Csim	XP_014649261.1	Dmrt	Dmrt-B1	Pgig	XP_039725399.1	Fox	Fox-C1
Cdro	XP_031321432.1	Dmrt	Dmrt-B1	Shar	XP_023352145.2	Fox	Fox-C1
Rfer	XP_032970385.1	Dmrt	Dmrt-B1	Ptig	XP_042841201.1	Fox	Fox-C1
Clup	XP_038393419.1	Dmrt	Dmrt-B1	Amel	XP_034516093.1	Fox	Fox-C1
Tman	XP_012409672.2	Dmrt	Dmrt-B1	Clup	XP_038439885.1	Fox	Fox-C1
Ggal	NP_001232910.1	Dmrt	Dmrt-B1	Scar	XP_047385678.1	Fox	Fox-C2
Oafe	XP_007954175.1	Dmrt	Dmrt-2	Mjav	XP_036849635.1	Fox	Fox-C2
Hsap	NP_870987.2	Dmrt	Dmrt-2	Cdro	XP_031292483.1	Fox	Fox-C2
Dnov	XP_004459445.2	Dmrt	Dmrt-2	Ttru	XP_033700545.1	Fox	Fox-C2
Shar	XP_003761708.1	Dmrt	Dmrt-2	Tman	XP_023588281.1	Fox	Fox-C2
Cpor	XP_005004778.1	Dmrt	Dmrt-2	Csim	XP_004437101.1	Fox	Fox-C2
Ttru	XP_019790310.1	Dmrt	Dmrt-2	Pafn	XP_047648754.1	Fox	Fox-C2
Ggal	XP_003643035.3	Dmrt	Dmrt-2	Mjav	XP_036849634.1	Fox	Fox-C2
Mdom	XP_001374295.1	Dmrt	Dmrt-2	Rfer	XP_032982997.1	Fox	Fox-C2
Oana	XP_028911116.1	Dmrt	Dmrt-2	Pgig	XP_039744659.1	Fox	Fox-C2
Tman	XP_004373589.1	Dmrt	Dmrt-2	Ggal	NP_001382975.1	Fox	Fox-C2
Equa	XP_046519727.1	Dmrt	Dmrt-2	Clup	XP_038394038.1	Fox	Fox-C2
Bmus	XP_036712720.1	Dmrt	Dmrt-2	Opri	XP_058530600.1	Fox	Fox-C2
Cdid	XP_037653375.1	Dmrt	Dmrt-2	Amel	XP_034495216.1	Fox	Fox-C2
Opri	XP_058513365.1	Dmrt	Dmrt-2	Bmus	XP_036688981.1	Fox	Fox-C2
Lcat	XP_045418965.1	Dmrt	Dmrt-2	Lcat	XP_045389072.1	Fox	Fox-C2
Cimi	XP_017373822.1	Dmrt	Dmrt-2	Cpor	XP_005008627.1	Fox	Fox-C2
Rfer	XP_032979206.1	Dmrt	Dmrt-2	Mmus	NP_038547.2	Fox	Fox-C2
Pgig	XP_039723855.1	Dmrt	Dmrt-2	Oafe	XP_007937406.1	Fox	Fox-C2
Ptig	XP_042820943.1	Dmrt	Dmrt-2	Cdid	XP_0376711788.1	Fox	Fox-C2
Hamp	XP_057579943.1	Dmrt	Dmrt-2	Bbub	XP_006080917.3	Fox	Fox-C2
Pafn	XP_047623676.1	Dmrt	Dmrt-2	Emax	XP_049720645.1	Fox	Fox-C2
Cdro	XP_031305655.1	Dmrt	Dmrt-2	Rfer	XP_032982975.1	Fox	Fox-C2
Mmus	NP_665830.1	Dmrt	Dmrt-2	Equa	XP_046539000.1	Fox	Fox-C2
Csim	XP_014650903.1	Dmrt	Dmrt-2	Ptig	XP_042825379.1	Fox	Fox-C2
Casi	XP_006863797.1	Dmrt	Dmrt-2	Mang	XP_045746247.1	Fox	Fox-C2
Emax	XP_049752756.1	Dmrt	Dmrt-2	Cimi	XP_017399160.1	Fox	Fox-C2
Mjav	XP_017529816.2	Dmrt	Dmrt-2	Drot	XP_053772216.1	Fox	Fox-C2
Amel	XP_034503257.1	Dmrt	Dmrt-2	Hsap	NP_005242.1	Fox	Fox-C2
Clup	XP_038383091.1	Dmrt	Dmrt-2	Hamp	XP_057568935.1	Fox	Fox-C2
Mang	XP_045738142.1	Dmrt	Dmrt-2	Mdom	XP_001365891.1	Fox	Fox-C2
Bbub	XP_044795850.1	Dmrt	Dmrt-2	Dnov	XP_004450287.3	Fox	Fox-C2
Drot	XP_045047034.2	Dmrt	Dmrt-2	Casi	XP_006860181.1	Fox	Fox-C2
Scar	XP_047380961.1	Dmrt	Dmrt-2	Oana	XP_039769457.1	Fox	Fox-C2
Cdro	XP_031312888.1	Dmrt	Dmrt-C2	Shar	XP_031806033.1	Fox	Fox-C2
Clup	XP_038381591.1	Dmrt	Dmrt-C2	Oana	XP_028921129.1	Fox	Hnf-3g/Fox-A3
Csim	XP_004440164.1	Dmrt	Dmrt-C2	Hsap	NP_004488.2	Fox	Hnf-3g/Fox-A3
Casi	XP_006871451.1	Dmrt	Dmrt-C2	Pafn	XP_047645608.1	Fox	Hnf-3g/Fox-A3
Mang	XP_045746349.1	Dmrt	Dmrt-C2	Ptig	XP_042825317.1	Fox	Hnf-3g/Fox-A3
Bmus	XP_036690564.1	Dmrt	Dmrt-C2	Mang	XP_045746361.1	Fox	Hnf-3g/Fox-A3
Lcat	XP_045387563.1	Dmrt	Dmrt-C2	Mmus	NP_032286.1	Fox	Hnf-3g/Fox-A3
Hamp	XP_057568752.1	Dmrt	Dmrt-C2	Casi	XP_006871384.1	Fox	Hnf-3g/Fox-A3
Cdid	XP_037676589.1	Dmrt	Dmrt-C2	Shar	XP_031817519.1	Fox	Hnf-3g/Fox-A3
Pafn	XP_047644735.1	Dmrt	Dmrt-C2	Oafe	XP_007941248.1	Fox	Hnf-3g/Fox-A3
Mmus	XP_017167796.1	Dmrt	Dmrt-C2	Ttru	XP_033701564.1	Fox	Hnf-3g/Fox-A3
Equa	XP_046507274.1	Dmrt	-	Mang	XP_045746361.1	Fox	Hnf-3g/Fox-A3
Oafe	XP_007941145.1	Dmrt	Dmrt-C2	Clup	XP_038384456.1	Fox	Hnf-3g/Fox-A3
Tman	XP_004388803.1	Dmrt	Dmrt-C2	Opri	XP_004597990.1	Fox	Hnf-3g/Fox-A3
Drot	XP_024433509.1	Dmrt	Dmrt-C2	Hamp	XP_057566925.1	Fox	Hnf-3g/Fox-A3
Oana	XP_007664048.1	Dmrt	Dmrt-C2	Rfer	XP_032982943.1	Fox	Hnf-3g/Fox-A3
Ttru	XP_033699919.1	Dmrt	Dmrt-C2	Dnov	XP_004481594.1	Fox	Hnf-3g/Fox-A3
Amel	XP_034944561.1	Dmrt	Dmrt-C2	Emax	XP_049757192.1	Fox	Hnf-3g/Fox-A3
Opri	XP_058531141.1	Dmrt	Dmrt-C2	Pgig	XP_039722184.1	Fox	Hnf-3g/Fox-A3
Pgig	XP_039731236.1	Dmrt	-	Tman	XP_004381648.1	Fox	Hnf-3g/Fox-A3
Lcat	XP_045393814.1	Dmrt	-	Mdom	XP_001364242.1	Fox	Hnf-3g/Fox-A3
Lcat	XP_045393815.1	Dmrt	-	Bmus	XP_036689576.1	Fox	Hnf-3g/Fox-A3
Lcat	XP_045393818.1	Dmrt	-	Equa	XP_046539522.1	Fox	Hnf-3g/Fox-A3
Scar	XP_047383853.1	Dmrt	Dmrt-C2	Bbub	XP_025125137.1	Fox	Hnf-3g/Fox-A3
Pgig	XP_039705918.1	Dmrt	Dmrt-C2	Csim	XP_004440228.1	Fox	Hnf-3g/Fox-A3
Hsap	XP_016882612.1	Dmrt	Dmrt-C2	Cimi	XP_017358010.1	Fox	Hnf-3g/Fox-A3
Equa	XP_046540061.1	Dmrt	-	Scar	XP_047384662.1	Fox	Hnf-3g/Fox-A3
Mjav	XP_036869866.1	Dmrt	Dmrt-C2	Cdid	XP_037675543.1	Fox	Hnf-3g/Fox-A3
Rfer	XP_032984641.1	Dmrt	Dmrt-C2	Lcat	XP_045387339.1	Fox	Hnf-3g/Fox-A3
Shar	XP_031819119.1	Dmrt	Dmrt-C2	Cdro	XP_010985920.2	Fox	Hnf-3g/Fox-A3
Emax	XP_049757407.1	Dmrt	Dmrt-C2	Cpor	XP_003464639.1	Fox	Hnf-3g/Fox-A3
Lcat	XP_045393817.1	Dmrt	-	Amel	XP_002928546.3	Fox	Hnf-3g/Fox-A3
Ptig	XP_042825393.1	Dmrt	Dmrt-C2	Drot	XP_024425331.1	Fox	Hnf-3g/Fox-A3
Bbub	XP_025124040.1	Dmrt	Dmrt-C2	Ggal	XP_025006986.1	Fox	Hnf-3a/Fox-A1
Cimi	XP_037585356.1	Dmrt	Dmrt-C2	Pafn	XP_047652638.1	Fox	Hnf-3a/Fox-A1
Cpor	XP_003464597.1	Dmrt	Dmrt-C2	Emax	XP_049755462.1	Fox	Hnf-3a/Fox-A1
Csim	XP_004443941.1	Dmrt	-	Cimi	XP_017386340.1	Fox	Hnf-3a/Fox-A1
Mdom	XP_007506399.1	Dmrt	Dmrt-C2	Csim	XP_004436444.1	Fox	Hnf-3a/Fox-A1
Dnov	XP_058135474.1	Dmrt	Dmrt-C2	Cpor	XP_013010026.1	Fox	Hnf-3a/Fox-A1
Lcat	XP_045393811.1	Dmrt	-	Ttru	XP_004326816.1	Fox	Hnf-3a/Fox-A1
Hsap	NP_067063.1	Dmrt	Dmrt-3	Lcat	XP_045424245.1	Fox	Hnf-3a/Fox-A1
Rfer	XP_032979350.1	Dmrt	Dmrt-3	Scar	XP_047398474.1	Fox	Hnf-3a/Fox-A1
Cimi	XP_017373824.1	Dmrt	Dmrt-3	Mjav	XP_017504124.1	Fox	Hnf-3a/Fox-A1
Oafe	XP_007954185.1	Dmrt	Dmrt-3	Mang	XP_045757926.1	Fox	Hnf-3a/Fox-A1
Cdid	XP_037653502.1	Dmrt	Dmrt-3	Mmus	XP_017170451.1	Fox	Hnf-3a/Fox-A1

Tab. S8 continued from previous page

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Mjav	XP_036847248.1	Dmrt	Dmrt-3	Casi	XP_006835393.1	Fox	Hnf-3a/Fox-A1
Mmus	XP_006527199.1	Dmrt	Dmrt-3	Dnov	XP_004453895.1	Fox	Hnf-3a/Fox-A1
Equa	XP_046521022.1	Dmrt	Dmrt-3	Cdro	XP_010982088.2	Fox	Hnf-3a/Fox-A1
Drot	XP_053780057.1	Dmrt	Dmrt-3	Clup	XP_038400478.1	Fox	Hnf-3a/Fox-A1
Hamp	XP_057581111.1	Dmrt	Dmrt-3	Hsap	NP_004487.2	Fox	Hnf-3a/Fox-A1
Lcat	XP_045418289.1	Dmrt	Dmrt-3	Ptig	XP_042845515.1	Fox	Hnf-3a/Fox-A1
Clup	XP_038381546.1	Dmrt	Dmrt-3	Pgig	XP_039712440.1	Fox	Hnf-3a/Fox-A1
Bmus	XP_036712559.1	Dmrt	Dmrt-3	Shar	XP_003756460.3	Fox	Hnf-3a/Fox-A1
Oana	XP_028910685.1	Dmrt	Dmrt-3	Amel	XP_019662027.2	Fox	Hnf-3a/Fox-A1
Casi	XP_006863796.1	Dmrt	Dmrt-3	Oafe	XP_007936759.1	Fox	Hnf-3a/Fox-A1
Scar	XP_047381281.1	Dmrt	Dmrt-3	Hamp	XP_057579078.1	Fox	Hnf-3a/Fox-A1
Ptig	XP_042820871.1	Dmrt	Dmrt-3	Bbub	XP_006074313.1	Fox	Hnf-3a/Fox-A1
Cpor	XP_003472245.1	Dmrt	Dmrt-3	Equa	XP_046511415.1	Fox	Hnf-3a/Fox-A1
Mdom	XP_001365681.1	Dmrt	Dmrt-3	Tman	XP_004376553.1	Fox	Hnf-3a/Fox-A1
Csim	XP_004440708.1	Dmrt	Dmrt-3	Rfer	XP_032965023.1	Fox	Hnf-3a/Fox-A1
Emax	XP_049752714.1	Dmrt	Dmrt-3	Drot	XP_053783670.1	Fox	Hnf-3a/Fox-A1
Opri	XP_004591889.2	Dmrt	Dmrt-3	Opri	XP_004584835.2	Fox	Hnf-3a/Fox-A1
Shar	XP_031799800.1	Dmrt	Dmrt-3	Mdom	XP_003339496.3	Fox	Hnf-3a/Fox-A1
Cdro	XP_031305659.1	Dmrt	Dmrt-3	Cdid	XP_037690784.1	Fox	Hnf-3a/Fox-A1
Bbub	XP_025137363.1	Dmrt	Dmrt-3	Bmus	XP_036697173.1	Fox	Hnf-3a/Fox-A1
Pafr	XP_047625503.1	Dmrt	Dmrt-3	Oana	XP_028934365.1	Fox	Hnf-3a/Fox-A1
Mang	XP_045737886.1	Dmrt	Dmrt-3	Shar	XP_031825225.1	Fox	Fox-E1
Dnov	XP_004459447.2	Dmrt	Dmrt-3	Casi	XP_0068633203.1	Fox	Fox-E1
Pgig	XP_039743957.1	Dmrt	Dmrt-3	Ggal	XP_040561338.1	Fox	Fox-E1
Ttru	XP_019789710.1	Dmrt	Dmrt-3	Oana	XP_007660203.2	Fox	Fox-E1
Ggal	XP_429193.2	Dmrt	Dmrt-3	Mdom	XP_001372714.1	Fox	Fox-E1
Amel	XP_034503143.1	Dmrt	Dmrt-3	Shar	XP_031824414.1	Fox	Fox-E1
Oafe	XP_007954174.1	Dmrt	Dmrt-1	Mang	XP_045737945.1	Fox	Fox-E1
Mdom	XP_001365618.1	Dmrt	Dmrt-1	Hamp	XP_057576142.1	Fox	Fox-E1
Tman	XP_004373524.1	Dmrt	Dmrt-1	Lcat	XP_045418451.1	Fox	Fox-E1
Cdro	XP_031305660.1	Dmrt	Dmrt-1	Equa	XP_046500267.1	Fox	Fox-E1
Bmus	XP_036711655.1	Dmrt	Dmrt-1	Hsap	NP_004464.2	Fox	Fox-E1
Shar	XP_031799789.1	Dmrt	Dmrt-1	Amel	XP_019650476.2	Fox	Fox-E1
Equa	XP_046519850.1	Dmrt	Dmrt-1	Csim	XP_004423293.1	Fox	Fox-E1
Bbub	XP_025137364.1	Dmrt	Dmrt-1	Drot	XP_053778384.1	Fox	Fox-E1
Hsap	XP_006716795.1	Dmrt	Dmrt-1	Cimi	XP_017383334.1	Fox	Fox-E1
Pafr	XP_047625504.1	Dmrt	Dmrt-1	Cdro	XP_031305931.1	Fox	Fox-E1
Drot	XP_053774861.1	Dmrt	Dmrt-1	Rfer	XP_032978155.1	Fox	Fox-E1
Cpor	XP_023421841.1	Dmrt	Dmrt-1	Bbub	XP_006080090.4	Fox	Fox-E1
Mang	XP_045738144.1	Dmrt	Dmrt-1	Cdid	XP_037654444.1	Fox	Fox-E1
Emax	XP_049752111.1	Dmrt	Dmrt-1	Ttru	XP_019804445.2	Fox	Fox-E1
Clup	XP_038383087.1	Dmrt	Dmrt-1	Dnov	XP_004454020.1	Fox	Fox-E1
Ptig	XP_042820071.1	Dmrt	Dmrt-1	Clup	XP_038408835.1	Fox	Fox-E1
Mmus	NP_056641.2	Dmrt	Dmrt-1	Pafr	XP_047623959.1	Fox	Fox-E1
Opri	XP_058513291.1	Dmrt	Dmrt-1	Opri	XP_004581237.3	Fox	Fox-E1
Csim	XP_014650940.1	Dmrt	Dmrt-1	Scar	XP_047379621.1	Fox	Fox-E1
Pgig	XP_039744060.1	Dmrt	Dmrt-1	Pgig	XP_039711385.1	Fox	Fox-E1
Ggal	XP_040511578.1	Dmrt	Dmrt-1	Mmus	NP_899121.1	Fox	Fox-E1
Oana	XP_028910684.1	Dmrt	Dmrt-1	Emax	XP_049752198.1	Fox	Fox-E1
Amel	XP_011224113.2	Dmrt	Dmrt-1	Bmus	XP_036710285.1	Fox	Fox-E1
Ttru	XP_033714027.1	Dmrt	Dmrt-1	Ptig	XP_042821250.1	Fox	Fox-E1
Cdid	XP_037653737.1	Dmrt	Dmrt-1	Mjav	XP_036860072.1	Fox	Fox-E1
Hamp	XP_057579864.1	Dmrt	Dmrt-1	Ggal	NP_990282.3	Fox	Fox-D3
Rfer	XP_032979671.1	Dmrt	Dmrt-1	Drot	XP_053776629.1	Fox	Fox-D3
Casi	XP_006863795.1	Dmrt	Dmrt-1	Cdro	XP_031321142.1	Fox	Fox-D3
Scar	XP_047381742.1	Dmrt	Dmrt-1	Ttru	XP_019774777.1	Fox	Fox-D3
Mjav	XP_036847220.1	Dmrt	Dmrt-1	Oana	XP_007663453.2	Fox	Fox-D3
Lcat	XP_045418248.1	Dmrt	Dmrt-1	Amel	XP_034505950.1	Fox	Fox-D3
Cimi	XP_017373819.1	Dmrt	Dmrt-1	Clup	XP_038391469.1	Fox	Fox-D3
Dnov	XP_058158898.1	Dmrt	Dmrt-1	Cdid	XP_037682960.1	Fox	Fox-D3
Oana	XP_039766676.1	Dmrt	Dmrt-A1	Mmus	NP_034555.3	Fox	Fox-D3
Cimi	XP_017401520.1	Dmrt	Dmrt-A1	Cimi	XP_017397049.1	Fox	Fox-D3
Mdom	XP_001374014.2	Dmrt	Dmrt-A1	Casi	XP_006862529.1	Fox	Fox-D3
Mang	XP_045737916.1	Dmrt	Dmrt-A1	Emax	XP_049731927.1	Fox	Fox-D3
Bmus	XP_036712737.1	Dmrt	Dmrt-A1	Ptig	XP_042852934.1	Fox	Fox-D3
Tman	XP_004373619.1	Dmrt	Dmrt-A1	Scar	XP_047371747.1	Fox	Fox-D3
Hsap	NP_071443.2	Dmrt	Dmrt-A1	Bmus	XP_036709961.1	Fox	Fox-D3
Emax	XP_049751570.1	Dmrt	Dmrt-A1	Opri	XP_058516851.1	Fox	Fox-D3
Drot	XP_053780867.1	Dmrt	Dmrt-A1	Dnov	XP_004448976.1	Fox	Fox-D3
Scar	XP_047381594.1	Dmrt	Dmrt-A1	Mjav	XP_036880296.1	Fox	Fox-D3
Hamp	XP_057580644.1	Dmrt	Dmrt-A1	Tman	XP_043639185.1	Fox	Fox-D3
Ttru	XP_004321228.1	Dmrt	Dmrt-A1	Hsap	NP_036315.1	Fox	Fox-D3
Opri	XP_058528639.1	Dmrt	Dmrt-A1	Lcat	XP_045402384.1	Fox	Fox-D3
Rfer	XP_032978102.1	Dmrt	Dmrt-A1	Bbub	XP_025144350.1	Fox	Fox-D3
Amel	XP_034519591.1	Dmrt	Dmrt-A1	Mang	XP_045751362.1	Fox	Fox-D3
Clup	XP_038408464.1	Dmrt	Dmrt-A1	Hamp	XP_057560293.1	Fox	Fox-D3
Oafe	XP_007946600.1	Dmrt	Dmrt-A1	Pafr	XP_047648179.1	Fox	Fox-D3
Pafr	XP_047625485.1	Dmrt	Dmrt-A1	Shar	XP_031824242.1	Fox	Fox-D3
Pgig	XP_039736661.1	Dmrt	Dmrt-A1	Pgig	XP_039739159.1	Fox	Fox-D3
Cpor	XP_0034787909.1	Dmrt	Dmrt-A1	Rfer	XP_032969240.1	Fox	Fox-D3
Mmus	NP_783578.1	Dmrt	Dmrt-A1	Mdom	XP_056670078.1	Fox	Fox-D3
Lcat	XP_045418524.1	Dmrt	Dmrt-A1	Scar	XP_047393167.1	Fox	Fox-N3
Bbub	XP_006067119.4	Dmrt	Dmrt-A1	Opri	XP_058511345.1	Fox	Fox-N3
Cdro	XP_010982933.2	Dmrt	Dmrt-A1	Cpor	XP_003472517.1	Fox	Fox-N3
Csim	XP_004428451.1	Dmrt	Dmrt-A1	Mmus	XP_006516295.1	Fox	Fox-N3
Mjav	XP_017516191.1	Dmrt	Dmrt-A1	Hsap	NP_001078940.1	Fox	Fox-N3
Ptig	XP_007098573.2	Dmrt	Dmrt-A1	Lcat	XP_045418142.1	Fox	Fox-N3
Casi	XP_006864558.1	Dmrt	Dmrt-A1	Cdro	XP_031310026.1	Fox	Fox-N3
Cdid	XP_037653778.1	Dmrt	Dmrt-A1	Rfer	XP_032965297.1	Fox	Fox-N3
Equa	XP_046520805.1	Dmrt	Dmrt-A1	Bbub	XP_045018818.1	Fox	Fox-N3
Dnov	XP_004456130.1	Dmrt	Dmrt-A1	Equa	XP_046503458.1	Fox	Fox-N3
Ggal	XP_015146712.1	Dmrt	Dmrt-A2	Pgig	XP_039739926.1	Fox	Fox-N3
Amel	XP_034508227.1	Dmrt	Dmrt-A2	Bmus	XP_036697221.1	Fox	Fox-N3
Opri	XP_004588718.2	Dmrt	Dmrt-A2	Ttru	XP_019775018.1	Fox	Fox-N3
Oana	XP_028902450.1	Dmrt	Dmrt-A2	Mjav	XP_036847649.1	Fox	Fox-N3
Bbub	XP_025145263.1	Dmrt	Dmrt-A2	Csim	XP_014645970.1	Fox	Fox-N3
Oafe	XP_007944067.1	Dmrt	Dmrt-A2	Drot	XP_024424133.1	Fox	Fox-N3
Dnov	XP_004476539.1	Dmrt	Dmrt-A2	Opri	XP_058516899.1	Fox	Fox-N3
Cdid	XP_037672757.1	Dmrt	Dmrt-A2	Cimi	XP_017380034.1	Fox	Fox-N3
Pgig	XP_039741660.1	Dmrt	Dmrt-A2	Bbub	XP_006061543.1	Fox	Fox-N3
Hsap	NP_115486.1	Dmrt	Dmrt-A2	Pafr	XP_047652286.1	Fox	Fox-N3
Mmus	NP_758500.2	Dmrt	Dmrt-A2	Hamp	XP_057589083.1	Fox	Fox-N3

Tab. S8 continued from previous page

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Ttru	XP_019793397.2	Dmrt	Dmrt-A2	Cimi	XP_017372373.1	Fox	Fox-R2
Hamp	XP_057564703.1	Dmrt	Dmrt-A2	Hsap	NP_940853.1	Fox	Fox-R2
Csim	XP_014649324.1	Dmrt	Dmrt-A2	Mdom	XP_007478014.2	Fox	Fox-I2
Bmus	XP_036726057.1	Dmrt	Dmrt-A2	Mjav	XP_017495569.2	Fox	Fox-I2
Clup	XP_038413502.1	Dmrt	Dmrt-A2	Shar	XP_031812885.1	Fox	Fox-I2
Mjav	XP_036877643.1	Dmrt	Dmrt-A2	Cdro	XP_031317856.1	Fox	Fox-I2
Paf	XP_047648562.1	Dmrt	Dmrt-A2	Cdid	XP_037661256.1	Fox	Fox-I2
Cdro	XP_031320961.1	Dmrt	Dmrt-A2	Hamp	XP_057593248.1	Fox	Fox-I2
Cimi	XP_017404203.1	Dmrt	Dmrt-A2	Ttru	XP_019795179.2	Fox	Fox-I2
Scar	XP_047394472.1	Dmrt	Dmrt-A2	Ptig	XP_042815793.1	Fox	Fox-I2
Mang	XP_045751377.1	Dmrt	Dmrt-A2	Bmus	XP_036685524.1	Fox	Fox-I2
Casi	XP_006839813.1	Dmrt	Dmrt-A2	Oafe	XP_007935078.1	Fox	Fox-I2
Tman	XP_023582760.1	Dmrt	Dmrt-A2	Bbub	XP_025129910.2	Fox	Fox-I2
Drot	XP_045060264.2	Dmrt	Dmrt-A2	Cimi	XP_017393376.1	Fox	Fox-I2
Lcat	XP_045402400.1	Dmrt	Dmrt-A2	Lcat	XP_045424382.1	Fox	Fox-I2
Rfer	XP_032969204.1	Dmrt	Dmrt-A2	Mmus	NP_899016.2	Fox	Fox-I2
Emax	XP_049731145.1	Dmrt	Dmrt-A2	Opri	XP_004579799.2	Fox	Fox-I2
Mdom	XP_001362692.2	Dmrt	Dmrt-A2	Scar	XP_047407992.1	Fox	Fox-I2
Cpor	XP_003463125.2	Dmrt	Dmrt-A2	Tman	XP_004375038.1	Fox	Fox-I2
Equa	XP_046516465.1	Dmrt	Dmrt-A2	Dnov	XP_004479387.2	Fox	Fox-I2
Ptig	XP_042853035.1	Dmrt	Dmrt-A2	Amel	XP_034517693.1	Fox	Fox-I2
Cimi	XP_017353253.1	Fox	-	Rfer	XP_032986720.1	Fox	Fox-I2
Cimi	XP_037600788.1	Fox	-	Equa	XP_046508191.1	Fox	Fox-I2
Tman	XP_023591424.1	Fox	Fox-M1	Casi	XP_006867537.1	Fox	Fox-I2
Bbub	XP_025138819.3	Fox	Fox-M1	Paf	XP_047617388.1	Fox	Fox-I2
Casi	XP_006862752.1	Fox	Fox-M1	Csim	XP_014649530.1	Fox	Fox-I2
Emax	XP_049738130.1	Fox	Fox-M1	Drot	XP_045047252.2	Fox	Fox-I2
Oafe	XP_007935251.1	Fox	Fox-M1	Hsap	NP_997309.2	Fox	Fox-I2
Drot	XP_053777572.1	Fox	Fox-M1	Mang	XP_045732277.1	Fox	Fox-I2
Ttru	XP_019781496.1	Fox	Fox-M1	Cpor	XP_003479857.1	Fox	Fox-I2
Csim	XP_014649484.1	Fox	Fox-M1	Clup	XP_038433915.1	Fox	Fox-I2
Paf	XP_047643777.1	Fox	Fox-M1	Ggal	NP_990523.3	Fox	Fox-D1
Oana	XP_028910212.1	Fox	Fox-M1	Lcat	XP_045421654.1	Fox	Fox-D1
Mang	XP_045727136.1	Fox	Fox-M1	Oana	XP_028933047.1	Fox	Fox-D1
Opri	XP_058511953.1	Fox	Fox-M1	Cimi	XP_017383122.1	Fox	Fox-D1
Pgig	XP_039725073.1	Fox	Fox-M1	Drot	XP_053781190.1	Fox	Fox-D1
Bmus	XP_036723013.1	Fox	Fox-M1	Paf	XP_047630421.1	Fox	Fox-D1
Dnov	XP_058156702.1	Fox	Fox-M1	Clup	XP_038385555.1	Fox	Fox-D1
Mjav	XP_017520172.2	Fox	Fox-M1	Shar	XP_031822455.1	Fox	Fox-D1
Amel	XP_034501532.1	Fox	Fox-M1	Bmus	XP_036703665.1	Fox	Fox-D1
Equa	XP_046512461.1	Fox	Fox-M1	Mdom	XP_056680722.1	Fox	Fox-D1
Dnov	XP_012379785.1	Fox	Fox-M1	Ttru	XP_019784789.1	Fox	Fox-D1
Clup	XP_038432360.1	Fox	Fox-M1	Cdid	XP_037657638.1	Fox	Fox-D1
Mdom	XP_056665089.1	Fox	Fox-M1	Mmus	NP_032268.2	Fox	Fox-D1
Rfer	XP_032974271.1	Fox	Fox-M1	Scar	XP_047412450.1	Fox	Fox-D1
Hamp	XP_057558870.1	Fox	Fox-M1	Dnov	XP_058161988.1	Fox	Fox-D1
Cdro	XP_031300170.1	Fox	Fox-M1	Hsap	NP_004463.1	Fox	Fox-D1
Ptig	XP_042847499.1	Fox	Fox-M1	Cdro	XP_031302569.1	Fox	Fox-D1
Shar	XP_031795171.1	Fox	Fox-M1	Rfer	XP_032965774.1	Fox	Fox-D1
Ggal	XP_046799087.1	Fox	Fox-M1	Emax	XP_049721422.1	Fox	Fox-D1
Cdid	XP_037702327.1	Fox	Fox-M1	Bbub	XP_025126650.2	Fox	Fox-D1
Mmus	NP_032047.4	Fox	Fox-M1	Pgig	XP_039714034.1	Fox	Fox-D1
Cpor	XP_013000601.1	Fox	Fox-M1	Hamp	XP_057554809.1	Fox	Fox-D1
Scar	XP_047405644.1	Fox	Fox-M1	Mjav	XP_036882073.1	Fox	Fox-D1
Ggal	XP_015153061.3	Fox	Fox-O6	Ptig	XP_042844167.1	Fox	Fox-D1
Cimi	XP_017397276.1	Fox	Fox-O6	Mang	XP_045742895.1	Fox	Fox-D1
Mdom	XP_001381514.2	Fox	Fox-O6	Opri	XP_058512313.1	Fox	Fox-D1
Paf	XP_047645485.1	Fox	Fox-O6	Ptig	XP_042823025.1	Fox	-
Mmus	NP_918949.1	Fox	Fox-O6	Cdro	XP_031325602.1	Fox	-
Emax	XP_049731901.1	Fox	Fox-O6	Dnov	XP_058140947.1	Fox	-
Scar	XP_047415675.1	Fox	Fox-O6	Cdid	XP_037666595.1	Fox	-
Ptig	XP_042852325.1	Fox	Fox-O6	Shar	XP_023357319.1	Fox	Fox-N3
Ttru	XP_033692835.1	Fox	Fox-O6	Mdom	XP_056666520.1	Fox	Fox-N3
Rfer	XP_032969239.1	Fox	Fox-O6	Oana	XP_028914735.1	Fox	Fox-N3
Tman	XP_023582811.1	Fox	Fox-O6	Ggal	XP_015143223.2	Fox	Fox-N3
Pgig	XP_039732650.1	Fox	Fox-O6	Ggal	XP_425185.2	Fox	Fox-I1
Cdro	XP_010997575.2	Fox	Fox-O6	Cpor	XP_003473379.2	Fox	Fox-I1
Bmus	XP_036684744.1	Fox	Fox-O6	Cdro	XP_031293629.1	Fox	Fox-I1
Casi	XP_008687666.1	Fox	Fox-O6	Pgig	XP_039732217.1	Fox	Fox-I1
Oana	XP_028936057.1	Fox	Fox-O6	Paf	XP_047640916.1	Fox	Fox-I1
Hsap	NP_001278210.2	Fox	Fox-O6	Lcat	XP_045408637.1	Fox	Fox-I1
Clup	XP_038413629.1	Fox	Fox-O6	Cimi	XP_017403198.1	Fox	Fox-I1
Oafe	XP_007950907.1	Fox	Fox-O6	Oafe	XP_007937941.1	Fox	Fox-I1
Bbub	XP_044801109.1	Fox	Fox-O6	Csim	XP_004428597.1	Fox	Fox-I1
Hamp	XP_057605223.1	Fox	Fox-O6	Cdid	XP_037655181.1	Fox	Fox-I1
Cpor	XP_04999818.1	Fox	Fox-O6	Hsap	NP_036320.2	Fox	Fox-I1
Dnov	XP_058159776.1	Fox	Fox-O6	Ptig	XP_007077705.2	Fox	Fox-I1
Mang	XP_045753906.1	Fox	Fox-O6	Mdom	XP_001370284.1	Fox	Fox-I1
Csim	XP_014639249.1	Fox	Fox-O6	Ttru	XP_033711041.1	Fox	Fox-I1
Amel	XP_034509638.1	Fox	Fox-O6	Shar	XP_003756860.1	Fox	Fox-I1
Shar	XP_031818190.1	Fox	Fox-O6	Rfer	XP_032952356.1	Fox	Fox-I1
Opri	XP_004591579.1	Fox	Fox-O6	Mjav	XP_017510459.2	Fox	Fox-I1
Mjav	XP_036858263.1	Fox	Fox-O6	Casi	XP_006864891.1	Fox	Fox-I1
Lcat	XP_045404045.1	Fox	Fox-O6	Mmus	NP_076396.3	Fox	Fox-I1
Drot	XP_053776876.1	Fox	Fox-O6	Scar	XP_047412385.1	Fox	Fox-I1
Equa	XP_046516690.1	Fox	Fox-O6	Drot	XP_024432893.2	Fox	Fox-I1
Cdid	XP_037673411.1	Fox	Fox-O6	Hamp	XP_057605245.1	Fox	Fox-I1
Oana	XP_028918467.1	Fox	Fox-H1	Amel	XP_02931169.2	Fox	Fox-I1
Mjav	XP_017499188.2	Fox	Fox-H1	Opri	XP_004587500.1	Fox	Fox-I1
Scar	XP_053599327.1	Fox	Fox-H1	Mang	XP_045743383.1	Fox	Fox-I1
Cpor	XP_003463743.1	Fox	Fox-H1	Equa	XP_046524041.1	Fox	Fox-I1
Bmus	XP_036686706.1	Fox	Fox-H1	Bmus	XP_036703672.1	Fox	Fox-I1
Mjav	XP_036859882.1	Fox	Fox-H1	Oana	XP_028905836.1	Fox	Fox-I1
Shar	XP_031803442.1	Fox	Fox-H1	Tman	XP_04371270.1	Fox	Fox-I1
Emax	XP_049710232.1	Fox	Fox-H1	Emax	XP_049714816.1	Fox	Fox-I1
Dnov	XP_058132182.1	Fox	Fox-H1	Clup	XP_038389675.1	Fox	Fox-I1
Rfer	XP_032982232.1	Fox	Fox-H1	Bbub	XP_006075587.1	Fox	Fox-I1
Tman	XP_004387459.1	Fox	Fox-H1	Dnov	XP_004460267.2	Fox	Fox-I1
Opri	XP_004580951.4	Fox	Fox-H1	Ggal	NP_99023.1	Fox	Fox-D2
Cdid	XP_037659123.1	Fox	Fox-H1	Emax	XP_049731922.1	Fox	Fox-D2
Mang	XP_045751338.1	Fox	Fox-H1	Cdro	XP_031320938.1	Fox	Fox-D2
Ttru	XP_033698705.1	Fox	Fox-H1	Ptig	XP_042852978.1	Fox	Fox-D2

Tab. S8 continued from previous page

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Ptig	XP_042829602.1	Fox	Fox-H1	Cdid	XP_037683211.1	Fox	Fox-D2
Bbub	XP_006064665.1	Fox	Fox-H1	Clup	XP_038413506.1	Fox	Fox-D2
Mmus	NP_032015.1	Fox	Fox-H1	Mang	XP_045751380.1	Fox	Fox-D2
Clup	XP_038411781.1	Fox	Fox-H1	Oafe	XP_007944900.1	Fox	Fox-D2
Cimi	XP_017367179.1	Fox	Fox-H1	Drot	XP_053776598.1	Fox	Fox-D2
Drot	XP_024428211.1	Fox	Fox-H1	Equa	XP_046519295.1	Fox	Fox-D2
Amel	XP_034524715.1	Fox	Fox-H1	Hamp	XP_057551251.1	Fox	Fox-D2
Mang	XP_045751622.1	Fox	Fox-H1	Cpor	XP_003461518.1	Fox	Fox-D2
Oafe	XP_007954253.1	Fox	Fox-H1	Csim	XP_004438679.1	Fox	Fox-D2
Hsap	NP_003914.1	Fox	Fox-H1	Casi	XP_006839796.1	Fox	Fox-D2
Csim	XP_004443058.1	Fox	Fox-H1	Scar	XP_053599256.1	Fox	Fox-D2
Cdro	XP_010997045.2	Fox	Fox-H1	Shar	XP_031825021.1	Fox	Fox-D2
Casi	XP_006830949.1	Fox	Fox-H1	Cimi	XP_017365579.1	Fox	Fox-D2
Pafr	XP_047641595.1	Fox	Fox-H1	Mdom	XP_003340123.1	Fox	Fox-D2
Equa	XP_046497693.1	Fox	Fox-H1	Bmus	XP_036724630.1	Fox	Fox-D2
Ppig	XP_039702105.1	Fox	Fox-H1	Pafr	XP_047645114.1	Fox	Fox-D2
Hamp	XP_057592839.1	Fox	Fox-H1	Cdro	XP_031319575.1	Fox	Fox-D2
Mdom	XP_016288146.2	Fox	Fox-H1	Mjav	XP_036877843.1	Fox	Fox-D2
Lcat	XP_045417128.1	Fox	Fox-H1	Hsap	NP_004465.3	Fox	Fox-D2
Ggal	XP_001234496.5	Fox	Fox-O3/Fox-O3B	Dnov	XP_004447059.1	Fox	Fox-D2
Mdom	XP_001368493.2	Fox	Fox-O3/Fox-O3B	Amel	XP_034511495.1	Fox	Fox-D2
Bmus	XP_036727351.1	Fox	Fox-O3/Fox-O3B	Oafe	XP_007944917.1	Fox	Fox-D2
Equa	XP_046532634.1	Fox	Fox-O3/Fox-O3B	Bbub	XP_0251144570.1	Fox	Fox-D2
Emax	XP_049752199.1	Fox	Fox-O3/Fox-O3B	Ttru	XP_019793278.1	Fox	Fox-D2
Hsap	NP_001355064.1	Fox	Fox-O3/Fox-O3B	Rfer	XP_032969197.1	Fox	Fox-D2
Shar	XP_023358910.2	Fox	Fox-O3/Fox-O3B	Tman	XP_004371825.1	Fox	Fox-D2
Ppig	XP_039725886.1	Fox	Fox-O3/Fox-O3B	Oana	XP_028902568.1	Fox	Fox-D2
Cdid	XP_037698233.1	Fox	Fox-O3/Fox-O3B	Opri	XP_004598554.3	Fox	Fox-D2
Cimi	XP_037599874.1	Fox	Fox-O3/Fox-O3B	Ppig	XP_039741502.1	Fox	Fox-D2
Bbub	XP_044780301.2	Fox	Fox-O3/Fox-O3B	Mmus	NP_032619.1	Fox	Fox-D2
Opri	XP_058515970.1	Fox	Fox-O3/Fox-O3B	Lcat	XP_045401421.1	Fox	Fox-D2
Pafr	XP_047618159.1	Fox	Fox-O3/Fox-O3B	Csim	XP_004433028.1	Fox	-
Mmus	NP_062714.1	Fox	Fox-O3/Fox-O3B	Rfer	XP_032946066.1	Fox	-
Oafe	XP_007939014.1	Fox	Fox-O3/Fox-O3B	Ppig	XP_039715338.1	Fox	-
Clup	XP_038410808.1	Fox	Fox-O3/Fox-O3B	Emax	XP_049754885.1	Fox	Fox-N3
Dnov	XP_058163213.1	Fox	Fox-O3/Fox-O3B	Tman	XP_023587457.1	Fox	Fox-N3
Scar	XP_047415677.1	Fox	Fox-O3/Fox-O3B	Cdid	XP_037678324.1	Fox	-
Ptig	XP_042842859.1	Fox	Fox-O3/Fox-O3B	Dnov	XP_004462180.1	Fox	-
Mang	XP_045721230.1	Fox	Fox-O3/Fox-O3B	Bbub	XP_006068529.1	Fox	-
Casi	XP_006840010.1	Fox	Fox-O3/Fox-O3B	Mang	XP_045748670.1	Fox	-
Mjav	XP_036868157.1	Fox	Fox-O3/Fox-O3B	Pafr	XP_047620398.1	Fox	-
Cpor	XP_023419223.1	Fox	Fox-O3/Fox-O3B	Bmus	XP_036694953.1	Fox	-
Hsap	NP_963853.1	Fox	Fox-O3/Fox-O3B	Cdro	XP_010983165.1	Fox	-
Drot	XP_024407555.2	Fox	Fox-O3/Fox-O3B	Hamp	XP_057573282.1	Fox	-
Lcat	XP_045400090.1	Fox	Fox-O3/Fox-O3B	Rfer	XP_032949909.1	Fox	-
Ttru	XP_033723495.1	Fox	Fox-O3/Fox-O3B	Equa	XP_046528957.1	Fox	-
Oana	XP_001511165.3	Fox	Fox-O3/Fox-O3B	Ppig	XP_039707370.1	Fox	-
Csim	XP_014636634.1	Fox	Fox-O3/Fox-O3B	Amel	XP_002930558.1	Fox	-
Cdro	XP_031311979.1	Fox	Fox-O3/Fox-O3B	Mjav	XP_017524585.2	Fox	-
Tman	XP_023583398.1	Fox	Fox-O3/Fox-O3B	Clup	XP_038442440.1	Fox	-
Amel	XP_034524842.1	Fox	Fox-O3/Fox-O3B	Mdom	XP_007483490.1	Fox	-
Rfer	XP_032957487.1	Fox	Fox-O3/Fox-O3B	Shar	XP_023358615.2	Fox	-
Hamp	XP_057593622.1	Fox	Fox-O3/Fox-O3B	Ggal	NP_001382146.1	Fox	-
Shar	XP_023352140.2	Fox	Fox-Q1	Mjav	XP_036853444.1	Fox	-
Mang	XP_045721917.1	Fox	Fox-Q1	Pafr	XP_047615141.1	Fox	-
Oana	XP_039766532.1	Fox	Fox-Q1	Bmus	XP_036692188.1	Fox	-
Mdom	XP_007488113.2	Fox	Fox-Q1	Ttru	XP_033703709.1	Fox	-
Oana	XP_039766897.1	Fox	Fox-Q1	Bbub	XP_006053445.2	Fox	-
Lcat	XP_045407521.1	Fox	Fox-Q1	Oana	XP_039768943.1	Fox	Fox-N2
Csim	XP_014639916.1	Fox	Fox-Q1	Shar	XP_031806310.1	Fox	Fox-N2
Rfer	XP_032969223.1	Fox	Fox-Q1	Ppig	XP_039743145.1	Fox	Fox-N2
Emax	XP_049738363.1	Fox	Fox-Q1	Emax	XP_049726307.1	Fox	Fox-N2
Scar	XP_047414319.1	Fox	Fox-Q1	Casi	XP_006839498.1	Fox	Fox-N2
Drot	XP_024408099.2	Fox	Fox-Q1	Csim	XP_004436724.1	Fox	Fox-N2
Ggal	XP_015137671.3	Fox	Fox-Q1	Cdro	XP_031322832.1	Fox	Fox-N2
Clup	XP_038440419.1	Fox	Fox-Q1	Amel	XP_011215591.1	Fox	Fox-N2
Bmus	XP_036726386.1	Fox	Fox-Q1	Bmus	XP_036729273.1	Fox	Fox-N2
Hamp	XP_057556649.1	Fox	Fox-Q1	Tman	XP_023591138.1	Fox	Fox-N2
Ttru	XP_019800554.1	Fox	Fox-Q1	Dnov	XP_023444629.2	Fox	Fox-N2
Cdid	XP_037700042.1	Fox	Fox-Q1	Lcat	XP_045405515.1	Fox	Fox-N2
Cdro	XP_031291204.1	Fox	Fox-Q1	Hsap	XP_04730063.1	Fox	Fox-N2
Equa	XP_046496965.1	Fox	Fox-Q1	Opri	XP_058523737.1	Fox	Fox-N2
Mjav	XP_036867494.1	Fox	Fox-Q1	Cimi	XP_017404014.1	Fox	Fox-N2
Ptig	XP_042841203.1	Fox	Fox-Q1	Clup	XP_038407168.1	Fox	Fox-N2
Bbub	XP_025120519.3	Fox	Fox-Q1	Drot	XP_053780764.1	Fox	Fox-N2
Mmus	NP_032265.3	Fox	Fox-Q1	Mang	XP_045740097.1	Fox	Fox-N2
Cimi	XP_017352994.1	Fox	Fox-Q1	Scar	XP_047378662.1	Fox	Fox-N2
Opri	XP_058528571.1	Fox	Fox-Q1	Equa	XP_046519098.1	Fox	Fox-N2
Dnov	XP_058141147.1	Fox	Fox-Q1	Ptig	XP_042837276.1	Fox	Fox-N2
Oafe	XP_007933584.1	Fox	Fox-Q1	Pafr	XP_047638079.1	Fox	Fox-N2
Ppig	XP_039725409.1	Fox	Fox-Q1	Cdid	XP_037662960.1	Fox	Fox-N2
Casi	XP_006870628.1	Fox	Fox-Q1	Ggal	XP_046794679.1	Fox	Fox-N2
Pafr	XP_047651460.1	Fox	Fox-Q1	Oafe	XP_007952184.1	Fox	Fox-N2
Hsap	NP_150285.3	Fox	Fox-Q1	Mmus	XP_036016210.1	Fox	Fox-N2
Cimi	XP_037597639.1	Fox	Fox-G1	Mjav	XP_017530752.1	Fox	Fox-N2
Lcat	XP_045411043.1	Fox	Fox-G1	Hamp	XP_057599130.1	Fox	Fox-N2
Cdro	XP_010984479.2	Fox	Fox-G1	Cpor	XP_003473126.1	Fox	Fox-N2
Rfer	XP_032965585.1	Fox	Fox-G1	Bbub	XP_044781825.1	Fox	Fox-N2
Drot	XP_024407046.1	Fox	Fox-G1	Mdom	XP_001375500.1	Fox	Fox-N2
Pafr	XP_047651592.1	Fox	Fox-G1	Ttru	XP_033694467.1	Fox	Fox-N2
Oana	XP_039770060.1	Fox	Fox-G1	Rfer	XP_032981124.1	Fox	Fox-N2
Ppig	XP_039726380.1	Fox	Fox-G1	Emax	XP_049728576.1	Fox	Fox-R2
Dnov	XP_058149613.1	Fox	Fox-G1	Tman	XP_004391284.1	Fox	Fox-R2
Mdom	XP_001364896.1	Fox	Fox-G1	Shar	XP_031815965.1	Fox	Fox-R1
Mang	XP_045757915.1	Fox	Fox-G1	Drot	XP_024426718.1	Fox	Fox-R1
Mjav	XP_017501197.2	Fox	Fox-G1	Mdom	XP_001380644.1	Fox	Fox-R1
Emax	XP_049754324.1	Fox	Fox-G1	Oana	XP_028931756.1	Fox	Fox-R1
Ttru	XP_019779874.1	Fox	Fox-G1	Casi	XP_006834052.1	Fox	Fox-R1
Cdid	XP_037690644.1	Fox	Fox-G1	Equa	XP_046540933.1	Fox	Fox-R1
Hsap	NP_005240.3	Fox	Fox-G1	Mmus	NP_001028641.1	Fox	Fox-R1
Shar	XP_031808132.1	Fox	Fox-G1	Cpor	XP_003472699.1	Fox	Fox-R1
Ptig	XP_042845427.1	Fox	Fox-G1	Bbub	XP_044785576.1	Fox	Fox-R1

Tab. S8 continued from previous page

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Scar	XP_047396708.1	Fox	Fox-G1	Pgig	XP_039722531.1	Fox	Fox-R1
Clup	XP_038400306.1	Fox	Fox-G1	Rfer	XP_032976774.1	Fox	Fox-R1
Hamp	XP_057580105.1	Fox	Fox-G1	Ptig	XP_042814206.1	Fox	Fox-R1
Casi	XP_006835434.1	Fox	Fox-G1	Bmus	XP_0367211689.1	Fox	Fox-R1
Opri	XP_058521348.1	Fox	Fox-G1	Bmus	XP_036715648.1	Fox	Fox-R1
Bmus	XP_036700190.1	Fox	Fox-G1	Cdro	XP_031299399.1	Fox	Fox-R1
Bbub	XP_025126917.1	Fox	Fox-G1	Pafr	XP_047609433.1	Fox	Fox-R1
Mmusp	NP_001153584.1	Fox	Fox-G1	Hsap	XP_016873064.1	Fox	Fox-R1
Ggal	NP_989659.2	Fox	Fox-O1	Cdid	XP_037695751.1	Fox	Fox-R1
Cimi	XP_017397972.1	Fox	Fox-O1	Csim	XP_004427297.1	Fox	Fox-R1
Rfer	XP_032960350.1	Fox	Fox-O1	Emax	XP_049713316.1	Fox	Fox-R1
Casi	XP_006873780.1	Fox	Fox-O1	Mjav	XP_017505694.2	Fox	Fox-R1
Pgig	XP_039695386.1	Fox	Fox-O1	Hsap	NP_859072.1	Fox	Fox-R1
Dnov	XP_058132895.1	Fox	Fox-O1	Lcat	XP_045412286.1	Fox	Fox-R1
Ttru	XP_019792245.1	Fox	Fox-O1	Dnov	XP_023443955.1	Fox	Fox-R1
Oana	XP_001512968.3	Fox	Fox-O1	Amel	XP_034521812.1	Fox	Fox-R1
Hsap	NP_002006.2	Fox	Fox-O1	Hamp	XP_057603312.1	Fox	Fox-R1
Mang	XP_045725787.1	Fox	Fox-O1	Tman	XP_004385753.1	Fox	Fox-R1
Bmus	XP_036687548.1	Fox	Fox-O1	Oafe	XP_007934779.1	Fox	Fox-R1
Amel	XP_034519677.1	Fox	Fox-O1	Clup	XP_038391436.1	Fox	Fox-R1
Scar	XP_047409558.1	Fox	Fox-O1	Scar	XP_047373976.1	Fox	Fox-R1
Mmusp	NP_062713.2	Fox	Fox-O1	Opri	XP_058519782.1	Fox	Fox-R1
Opri	XP_012782130.2	Fox	Fox-O1	Ttru	XP_033717853.1	Fox	Fox-R1
Cdid	XP_037656524.1	Fox	Fox-O1	Oafe	XP_007947037.1	Fox	Fox-R1
Mjav	XP_036867814.1	Fox	Fox-O1	Mang	XP_045748841.1	Fox	Fox-R1
Cdro	XP_031321792.1	Fox	Fox-O1	Cimi	XP_017378961.1	Fox	Fox-R1
Lcat	XP_045423168.1	Fox	Fox-O1	Lcat	XP_045383006.1	Fox	Fox-K2
Clup	XP_038429080.1	Fox	Fox-O1	Casi	XP_006869677.1	Fox	Fox-K2
Cpor	XP_023416198.1	Fox	Fox-O1	Bmus	XP_011247520.1	Fox	Fox-K2
Drot	XP_024424320.2	Fox	Fox-O1	Drot	XP_024409582.3	Fox	Fox-K2
Mdom	XP_001368312.2	Fox	Fox-O1	Cpor	XP_013002484.1	Fox	Fox-K2
Shar	XP_03764601.3	Fox	Fox-O1	Tman	XP_023584943.1	Fox	Fox-K2
Csim	XP_004443237.1	Fox	Fox-O1	Scar	XP_047400835.1	Fox	Fox-K2
Tman	XP_023597991.1	Fox	Fox-O1	Opri	XP_058532091.1	Fox	Fox-K2
Emax	XP_049709381.1	Fox	Fox-O1	Oafe	XP_007957978.1	Fox	Fox-K2
Pafr	XP_047611907.1	Fox	Fox-O1	Hsap	NP_004505.2	Fox	Fox-K2
Bbub	XP_006065837.2	Fox	Fox-O1	Emax	XP_049715130.1	Fox	Fox-K2
Ptig	XP_007097795.2	Fox	Fox-O1	Cimi	XP_017380207.1	Fox	Fox-K2
Hamp	XP_057563333.1	Fox	Fox-O1	Hamp	XP_057574401.1	Fox	-
Oafe	XP_007943433.1	Fox	Fox-O1	Hamp	XP_057571581.1	Fox	-
Mang	XP_045725786.1	Fox	Fox-O1	Lcat	XP_045397194.1	Fox	Fox-K1
Ggal	XP_015134143.3	Fox	Fox-O4	Bmus	XP_036681953.1	Fox	Fox-K1
Casi	XP_006868595.1	Fox	Fox-O4	Pgig	XP_039734750.1	Fox	Fox-K1
Mdom	XP_056665439.1	Fox	Fox-O4	Equa	XP_046521295.1	Fox	Fox-K1
Csim	XP_004439923.1	Fox	Fox-O4	Hsap	NP_001032242.1	Fox	Fox-K1
Cdid	XP_037677588.1	Fox	Fox-O4	Emax	XP_049758808.1	Fox	Fox-K1
Hsap	NP_005929.2	Fox	Fox-O4	Oana	XP_028905534.1	Fox	Fox-K1
Emax	XP_049727631.1	Fox	Fox-O4	Opri	XP_012786071.2	Fox	Fox-K1
Ptig	XP_007099158.2	Fox	Fox-O4	Cimi	XP_017389267.1	Fox	Fox-K1
Drot	XP_053773393.1	Fox	Fox-O4	Drot	XP_053782069.1	Fox	Fox-K1
Bmus	XP_036696517.1	Fox	Fox-O4	Cpor	XP_023420906.1	Fox	Fox-K1
Oafe	XP_007957073.1	Fox	Fox-O4	Dnov	XP_004454003.2	Fox	Fox-K1
Cimi	XP_017372253.1	Fox	Fox-O4	Scar	XP_047389516.1	Fox	Fox-K1
Mjav	XP_017525885.1	Fox	Fox-O4	Csim	XP_004440941.1	Fox	Fox-K1
Rfer	XP_032969557.1	Fox	Fox-O4	Mmus	NP_951031.2	Fox	Fox-K1
Pafr	XP_047621021.1	Fox	Fox-O4	Shar	XP_031796898.1	Fox	Fox-K1
Mmus	NP_061259.1	Fox	Fox-O4	Hamp	XP_057604533.1	Fox	Fox-K1
Mang	XP_045735285.1	Fox	Fox-O4	Mang	XP_045731846.1	Fox	Fox-K1
Pgig	XP_039697652.1	Fox	Fox-O4	Casi	XP_006859933.1	Fox	Fox-K1
Scar	XP_047393133.1	Fox	Fox-O4	Clup	XP_038395513.1	Fox	Fox-K1
Oana	XP_039768253.1	Fox	Fox-O4	Cdid	XP_03769880.1	Fox	Fox-K1
Dnov	XP_058146903.1	Fox	Fox-O4	Pafr	XP_047635407.1	Fox	Fox-K1
Cdid	XP_037678715.1	Fox	Fox-O4	Tman	XP_004380896.3	Fox	Fox-K1
Amel	XP_002929104.1	Fox	Fox-O4	Amel	XP_011219634.2	Fox	Fox-K1
Tman	XP_023590438.1	Fox	Fox-O4	Mjav	XP_036866572.1	Fox	Fox-K1
Clup	XP_038443581.1	Fox	Fox-O4	Ttru	XP_019802935.2	Fox	Fox-K1
Cpor	XP_013009181.1	Fox	Fox-O4	Cdro	XP_031327189.1	Fox	Fox-K1
Lcat	XP_045394627.1	Fox	Fox-O4	Mdom	XP_056662440.1	Fox	Fox-K1
Hamp	XP_057575075.1	Fox	Fox-O4	Ptig	XP_042828249.1	Fox	Fox-K1
Bbub	XP_006076433.1	Fox	Fox-O4	Rfer	XP_032951743.1	Fox	Fox-K1
Ttru	XP_033705152.1	Fox	Fox-O4	Oafe	XP_007941047.1	Fox	Fox-K1
Equa	XP_046529856.1	Fox	Fox-O4	Ggal	XP_015149844.1	Fox	Fox-K1
Opri	XP_004595290.1	Fox	Fox-O4	Bbub	XP_025131152.1	Fox	Fox-K1
Drot	XP_053773548.1	Fox	Fox-O4	Hsap	NP_001400854.1	Fox	Fox-M1
Cdro	XP_010977055.2	Fox	Fox-O4	Lcat	XP_045410542.1	Fox	-
Pgig	XP_039724773.1	Fox	Fox-J3	Ggal	XP_046760565.1	Fox	-
Mdom	XP_007492986.2	Fox	Fox-J3	Oana	XP_028912611.1	Fox	Fox-K2
Lcat	XP_045401678.1	Fox	Fox-J3	Clup	XP_038402133.1	Fox	-
Shar	XP_031818195.1	Fox	Fox-J3	Amel	XP_019662714.1	Fox	-
Equa	XP_046517148.1	Fox	Fox-J3	Mang	XP_045744166.1	Fox	-
Mjav	XP_036858181.1	Fox	Fox-J3	Equa	XP_046532308.1	Fox	-
Oafe	XP_007953745.1	Fox	Fox-J3	Ptig	XP_042846258.1	Fox	-
Mmus	NP_766287.1	Fox	Fox-J3	Oafe	XP_007942676.1	Fox	-
Drot	XP_024410422.2	Fox	Fox-J3	Casi	XP_006839645.1	Fox	-
Tman	XP_023582916.1	Fox	Fox-J3	Mmus	XP_011246143.1	Fox	Fox-R2
Dnov	XP_004477472.1	Fox	Fox-J3	Opri	XP_058514738.1	Fox	-
Hsap	XP_005270689.1	Fox	Fox-J3	Scar	XP_047391089.1	Fox	-
Csim	XP_004443185.1	Fox	Fox-J3	Ggal	XP_015137672.3	Fox	-
Scar	XP_047408133.1	Fox	Fox-J3	Ggal	XP_015141419.2	Fox	-
Cimi	XP_017397267.1	Fox	Fox-J3	Ggal	XP_015144532.3	Fox	-
Bmus	XP_036706277.1	Fox	Fox-J3	Ggal	XP_015136219.2	Fox	-
Cdid	XP_037683454.1	Fox	Fox-J3	Cpor	XP_00499985.1	Fox	-
Cpor	XP_004999975.1	Fox	Fox-J3	Cdid	XP_037667673.1	Fox	Sox-18
Oana	XP_028921226.1	Fox	Fox-J3	Opri	XP_004586020.2	Fox	Sox-18
Bbub	XP_044801100.1	Fox	Fox-J3	Clup	XP_038427314.1	Fox	Sox-18
Ttru	XP_019793563.1	Fox	Fox-J3	Scar	XP_047396502.1	Fox	Sox-18
Casi	XP_006839889.1	Fox	Fox-J3	Cpor	XP_003462830.1	Fox	Sox-18
Clup	XP_038413618.1	Fox	Fox-J3	Mjav	XP_036862488.1	Fox	Sox-18
Emax	XP_049734920.1	Fox	Fox-J3	Bmus	XP_036681681.1	Fox	Sox-18
Pafr	XP_047645481.1	Fox	Fox-J3	Cdro	XP_031289217.1	Fox	Sox-18
Mang	XP_054368911.1	Fox	Fox-J3	Lcat	XP_045384104.1	Fox	Sox-18
Opri	XP_058535693.1	Fox	Fox-J3	Cimi	XP_017371142.1	Fox	Sox-18

Tab. S8 continued from previous page

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Amel	XP_011233273.1	Fox	Fox-J3	Equa	XP_046535409.1	Sox	Sox-18
Hamp	XP_057563297.1	Fox	Fox-J3	Bbub	XP_025119695.1	Sox	Sox-18
Ptig	XP_042852753.1	Fox	Fox-J3	Csim	XP_004430565.1	Sox	Sox-18
Cdro	XP_031320784.1	Fox	Fox-J3	Rfer	XP_032951103.1	Sox	Sox-18
Oana	XP_028910333.1	Fox	Fox-J2	Drot	XP_053783310.1	Sox	Sox-18
Hamp	XP_057559090.1	Fox	Fox-J2	Pgig	XP_039720817.1	Sox	Sox-18
Pafr	XP_047643563.1	Fox	Fox-J2	Ttru	XP_019806802.1	Sox	Sox-18
Ttru	XP_033722947.1	Fox	Fox-J2	Pafr	XP_047626512.1	Sox	Sox-18
Pgig	XP_039730188.1	Fox	Fox-J2	Mang	XP_045742130.1	Sox	Sox-18
Hsap	XP_011519063.1	Fox	Fox-J2	Mmus	NP_033262.2	Sox	Sox-18
Equa	XP_046526439.1	Fox	Fox-J2	Ptig	XP_042835603.1	Sox	Sox-18
Shar	XP_031794189.1	Fox	Fox-J2	Hsap	NP_060889.1	Sox	Sox-18
Scar	XP_047406645.1	Fox	Fox-J2	Hamp	XP_057559896.1	Sox	Sox-18
Cimi	XP_017367793.1	Fox	Fox-J2	Dnov	XP_058143328.1	Sox	Sox-18
Csim	XP_004438721.1	Fox	Fox-J2	Shar	XP_023350160.2	Sox	Sox-18
Emax	XP_049738321.1	Fox	Fox-J2	Oana	XP_028925867.1	Sox	Sox-18
Tman	XP_004387308.1	Fox	Fox-J2	Mdom	XP_007475557.3	Sox	Sox-18
Bmus	XP_036722276.1	Fox	Fox-J2	Ggal	NP_089640.1	Sox	Sox-18
Dnov	XP_004455323.1	Fox	Fox-J2	Ggal	XP_040551910.1	Sox	Sox-17
Mmus	NP_068699.1	Fox	Fox-J2	Opri	XP_004580649.2	Sox	Sox-17
Bbub	XP_025138942.2	Fox	Fox-J2	Ggal	NP_001034415.2	Sox	Sox-17
Casi	XP_006875525.1	Fox	Fox-J2	Mdom	XP_001379706.1	Sox	Sox-17
Mjav	XP_017507797.1	Fox	Fox-J2	Cdid	XP_037658799.1	Sox	Sox-17
Drot	XP_053777818.1	Fox	Fox-J2	Lcat	XP_045416951.1	Sox	Sox-17
Mdom	XP_001364301.2	Fox	Fox-J2	Mmus	NP_001276393.1	Sox	Sox-17
Rfer	XP_032973158.1	Fox	Fox-J2	Tman	XP_004372647.1	Sox	Sox-17
Cdro	XP_031300018.1	Fox	Fox-J2	Rfer	XP_032981648.1	Sox	Sox-17
Opri	XP_004596477.2	Fox	Fox-J2	Cpor	XP_005002064.1	Sox	Sox-17
Cpor	XP_003470329.1	Fox	Fox-J2	Oafe	XP_007950410.1	Sox	Sox-17
Ptig	XP_042847649.1	Fox	Fox-J2	Oana	XP_028925620.1	Sox	Sox-17
Ggal	XP_046759046.1	Fox	Fox-J2	Mdom	XP_001368625.2	Sox	Sox-17
Amel	XP_034501007.1	Fox	Fox-J2	Amel	XP_002925690.2	Sox	Sox-17
Oafe	XP_007935347.1	Fox	Fox-J2	Scar	XP_047417202.1	Sox	Sox-17
Mang	XP_045729866.1	Fox	Fox-J2	Hsap	NP_071899.1	Sox	Sox-17
Lcat	XP_045410352.1	Fox	Fox-J2	Cimi	XP_017387282.1	Sox	Sox-17
Cdid	XP_037701021.1	Fox	Fox-J2	Mjav	XP_036855229.1	Sox	Sox-17
Clup	XP_038432619.1	Fox	Fox-J2	Shar	XP_003759826.2	Sox	Sox-17
Opri	XP_004592944.1	Fox	Fox-J1	Bbub	XP_025121191.1	Sox	Sox-17
Bmus	XP_036692694.1	Fox	Fox-J1	Casi	XP_006871910.1	Sox	Sox-17
Ttru	XP_033703524.1	Fox	Fox-J1	Shar	XP_031802254.1	Sox	Sox-17
Mdom	XP_016286122.1	Fox	Fox-J1	Clup	XP_038435168.1	Sox	Sox-17
Pafr	XP_047614731.1	Fox	Fox-J1	Csim	XP_004435766.1	Sox	Sox-17
Mjav	XP_017529912.2	Fox	Fox-J1	Emax	XP_049716401.1	Sox	Sox-17
Cpor	XP_003461484.1	Fox	Fox-J1	Cdro	XP_010987841.2	Sox	Sox-17
Mmus	XP_006532332.1	Fox	Fox-J1	Ptig	XP_042829097.1	Sox	Sox-17
Ptig	XP_042823405.1	Fox	Fox-J1	Bmus	XP_036685683.1	Sox	Sox-17
Equa	XP_046530684.1	Fox	Fox-J1	Pgig	XP_039724865.1	Sox	Sox-17
Clup	XP_038402010.1	Fox	Fox-J1	Mang	XP_045752504.1	Sox	Sox-17
Emax	XP_049715791.1	Fox	Fox-J1	Pafr	XP_047639872.1	Sox	Sox-17
Oana	XP_028912588.1	Fox	Fox-J1	Hamp	XP_057592218.1	Sox	Sox-17
Tman	XP_004374194.1	Fox	Fox-J1	Equa	XP_046498273.1	Sox	Sox-17
Drot	XP_045046069.2	Fox	Fox-J1	Dnov	XP_058131520.1	Sox	Sox-17
Hsap	NP_001445.2	Fox	Fox-J1	Drot	XP_024434049.3	Sox	Sox-17
Pgig	XP_039715456.1	Fox	Fox-J1	Ttru	XP_019780097.1	Sox	Sox-17
Cimi	XP_017391310.1	Fox	Fox-J1	Oana	XP_028909114.1	Sox	Sox-7
Rfer	XP_032946522.1	Fox	Fox-J1	Bbub	XP_006064379.2	Sox	Sox-7
Lcat	XP_045381684.1	Fox	Fox-J1	Csim	XP_014645382.1	Sox	Sox-7
Dnov	XP_004454510.2	Fox	Fox-J1	Mdom	XP_001373591.1	Sox	Sox-7
Ggal	NP_001308464.2	Fox	Fox-J1	Casi	XP_006864394.1	Sox	Sox-7
Hamp	XP_057572756.1	Fox	Fox-J1	Ptig	XP_042839510.1	Sox	Sox-7
Cdro	XP_031325009.1	Fox	Fox-J1	Tman	XP_004382257.1	Sox	Sox-7
Oafe	XP_007957820.1	Fox	Fox-J1	Clup	XP_038429590.1	Sox	Sox-7
Amel	XP_034497541.1	Fox	Fox-J1	Pgig	XP_039731999.1	Sox	Sox-7
Cdid	XP_037665215.1	Fox	Fox-J1	Ggal	XP_046795199.1	Sox	Sox-7
Shar	XP_031821724.1	Fox	Fox-J1	Cimi	XP_017395944.1	Sox	Sox-7
Scar	XP_047399365.1	Fox	Fox-J1	Mmus	NP_035576.1	Sox	Sox-7
Csim	XP_004432853.1	Fox	Fox-J1	Emax	XP_049722562.1	Sox	Sox-7
Casi	XP_006869617.1	Fox	Fox-J1	Amel	XP_011214939.2	Sox	Sox-7
Bbub	XP_006045356.1	Fox	Fox-J1	Rfer	XP_032989201.1	Sox	Sox-7
Mang	XP_045743961.1	Fox	Fox-J1	Equa	XP_046512534.1	Sox	Sox-7
Oana	XP_039768325.1	Fox	Fox-P3	Ttru	XP_033713462.1	Sox	Sox-7
Pgig	XP_039699607.1	Fox	Fox-P3	Cpor	XP_003479745.1	Sox	Sox-7
Csim	XP_014646458.1	Fox	Fox-P3	Shar	XP_003758123.1	Sox	Sox-7
Equa	XP_046528495.1	Fox	Fox-P3	Cdid	XP_037668898.1	Sox	Sox-7
Cpor	XP_023417629.1	Fox	Fox-P3	Hamp	XP_057576011.1	Sox	Sox-7
Rfer	XP_032973683.1	Fox	Fox-P3	Bmus	XP_036712190.1	Sox	Sox-7
Oafe	XP_007956564.1	Fox	Fox-P3	Scar	XP_047405527.1	Sox	Sox-7
Shar	XP_023363009.2	Fox	Fox-P3	Drot	XP_053767954.1	Sox	Sox-7
Bmus	XP_036695836.1	Fox	Fox-P3	Pafr	XP_047615897.1	Sox	Sox-7
Mdom	XP_056665055.1	Fox	Fox-P3	Hsap	NP_113627.1	Sox	Sox-7
Cimi	XP_037589291.1	Fox	Fox-P3	Mang	XP_045736052.1	Sox	Sox-7
Emax	XP_049728302.1	Fox	Fox-P3	Dnov	XP_058144023.1	Sox	Sox-7
Drot	XP_053773440.1	Fox	Fox-P3	Cdro	XP_031297983.1	Sox	Sox-7
Mjav	XP_036852243.1	Fox	Fox-P3	Lcat	XP_045391548.1	Sox	Sox-7
Mmus	NP_001186276.1	Fox	Fox-P3	Opri	XP_004579156.2	Sox	Sox-7
Ttru	XP_033704944.1	Fox	Fox-P3	Oafe	XP_007936906.1	Sox	Sox-7
Cdid	XP_037677418.1	Fox	Fox-P3	Mjav	XP_036854859.1	Sox	Sox-7
Bbub	XP_006073707.2	Fox	Fox-P3	Pafr	XP_047609577.1	Sox	Sox-30
Casi	XP_006876748.1	Fox	Fox-P3	Ptig	XP_007077656.3	Sox	Sox-30
Scar	XP_047391408.1	Fox	Fox-P3	Bmus	XP_036704125.1	Sox	Sox-30
Hamp	XP_057574707.1	Fox	Fox-P3	Oana	XP_039766108.1	Sox	Sox-30
Lcat	XP_045393738.1	Fox	Fox-P3	Cimi	XP_017372578.1	Sox	Sox-30
Pafr	XP_047620716.1	Fox	Fox-P3	Oafe	XP_007948121.1	Sox	Sox-30
Amel	XP_011221377.1	Fox	Fox-P3	Cdro	XP_031305277.1	Sox	Sox-30
Dnov	XP_023446473.1	Fox	Fox-P3	Emax	XP_049730174.1	Sox	Sox-30
Ptig	XP_007096061.2	Fox	Fox-P3	Hamp	XP_05758120.1	Sox	Sox-30
Opri	XP_058514303.1	Fox	Fox-P3	Lcat	XP_045408644.1	Sox	Sox-30
Clup	NP_001161933.1	Fox	Fox-P3	Shar	XP_031809461.1	Sox	Sox-30
Mang	XP_045729506.1	Fox	Fox-P3	Bbub	XP_006058618.4	Sox	Sox-30
Cdid	XP_037677000.1	Fox	Fox-P3	Drot	XP_024432555.2	Sox	Sox-30
Hsap	NP_054728.2	Fox	Fox-P3	Clup	XP_038390830.1	Sox	Sox-30
Cdro	XP_031301084.1	Fox	Fox-P3	Cpor	XP_003473352.1	Sox	Sox-30

Tab. S8 continued from previous page

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Tman	XP_004376890.1	Fox	Fox-P3	Ttru	XP_019780387.1	Sox	Sox-30
Oana	XP_028909446.1	Fox	Fox-F2	Opri	XP_004587550.2	Sox	Sox-30
Oana	XP_028912286.1	Fox	Fox-F2	Mdom	XP_001379720.1	Sox	Sox-30
Opri	XP_012786079.2	Fox	Fox-L3	Equa	XP_046521338.1	Sox	Sox-30
Mdom	XP_007498348.2	Fox	Fox-L3	Csim	XP_004428646.1	Sox	Sox-30
Drot	XP_045040885.2	Fox	Fox-L3	Cdid	XP_037655441.1	Sox	Sox-30
Rfer	XP_032952385.1	Fox	Fox-L3	Ggal	XP_414564.1	Sox	Sox-30
Scar	XP_047390130.1	Fox	Fox-L3	Dnov	XP_058162315.1	Sox	Sox-30
Oana	XP_001511921.2	Fox	Fox-L3	Scar	XP_047412799.1	Sox	Sox-30
Ttru	XP_004328913.2	Fox	Fox-L3	Mmus	NP_775560.1	Sox	Sox-30
Dnov	XP_058141662.1	Fox	Fox-L3	Amel	XP_019650665.2	Sox	Sox-30
Mmus	NP_001182057.1	Fox	Fox-L3	Pgig	XP_039732492.1	Sox	Sox-30
Hsap	NP_001361767.1	Fox	Fox-L3	Mjav	XP_036881068.1	Sox	Sox-30
Emax	XP_049761077.1	Fox	Fox-L3	Mang	XP_045743539.1	Sox	Sox-30
Mang	XP_045728168.1	Fox	Fox-L3	Rfer	XP_032952708.1	Sox	Sox-30
Mjav	XP_036864912.1	Fox	Fox-L3	Tman	XP_004371314.1	Sox	Sox-30
Shar	XP_012398242.1	Fox	Fox-L3	Hsap	NP_848511.1	Sox	Sox-30
Cimi	XP_017363768.1	Fox	Fox-L3	Casi	XP_006863994.1	Sox	Sox-30
Hamp	XP_057605555.1	Fox	Fox-L3	Amel	XP_034508949.1	Sox	Sox-30
Pafr	XP_047635063.1	Fox	Fox-L3	Scar	XP_047389638.1	Sox	Sox-8
Tman	XP_004386032.1	Fox	Fox-L3	Bmus	XP_036681892.1	Sox	Sox-8
Csim	XP_004440991.1	Fox	Fox-L3	Pafr	XP_047635134.1	Sox	Sox-8
Equa	XP_046523569.1	Fox	Fox-L3	Drot	XP_024409185.2	Sox	Sox-8
Lcat	XP_045398087.1	Fox	Fox-L3	Mjav	XP_017502547.2	Sox	Sox-8
Cpor	XP_003469985.1	Fox	Fox-L3	Clup	XP_038394815.1	Sox	Sox-8
Bmus	XP_036680082.1	Fox	Fox-L3	Rfer	XP_032957935.1	Sox	Sox-8
Bbub	XP_044792209.1	Fox	Fox-L3	Equa	XP_046523802.1	Sox	Sox-8
Clup	XP_038394791.1	Fox	Fox-L3	Amel	XP_034525488.1	Sox	Sox-8
Casi	XP_006859950.1	Fox	Fox-L3	Cimi	XP_017366444.1	Sox	Sox-8
Cdid	XP_037669698.1	Fox	Fox-L3	Csim	XP_004438275.1	Sox	Sox-8
Ptig	XP_042828254.1	Fox	Fox-L3	Bbub	XP_025130606.3	Sox	Sox-8
Ggal	XP_425229.3	Fox	Fox-L3	Lcat	XP_045397007.1	Sox	Sox-8
Cdro	XP_031327057.1	Fox	Fox-L3	Hamp	XP_057551434.1	Sox	Sox-8
Amel	XP_034525410.1	Fox	Fox-L3	Pgig	XP_039740781.1	Sox	Sox-8
Oafe	XP_007941358.1	Fox	Fox-L3	Hsap	NP_055402.2	Sox	Sox-8
Cdro	XP_031326970.1	Fox	Fox-P1	Mang	XP_045730580.1	Sox	Sox-8
Oafe	XP_007944266.1	Fox	Fox-P1	Cdro	XP_010989436.2	Sox	Sox-8
Ttru	XP_033721167.1	Fox	Fox-P1	Ptig	XP_042827523.1	Sox	Sox-8
Amel	XP_011217264.1	Fox	Fox-P1	Ttru	XP_033696379.1	Sox	Sox-8
Mdom	XP_007500148.1	Fox	Fox-P1	Bmus	XP_036722097.1	Sox	—
Opri	XP_058534843.1	Fox	Fox-P1	Ttru	XP_019791993.1	Sox	—
Shar	XP_031801298.1	Fox	Fox-P1	Oana	XP_028908350.1	Sox	Sox-4
Pafr	XP_047635064.1	Fox	Fox-P1	Opri	XP_058523457.1	Sox	Sox-4
Mjav	XP_036875087.1	Fox	Fox-P1	Ggal	NP_989815.2	Sox	Sox-4
Hsap	NP_001231739.1	Fox	Fox-P1	Casi	XP_006860848.1	Sox	Sox-4
Casi	XP_006874684.1	Fox	Fox-P1	Lcat	XP_045407834.1	Sox	Sox-4
Emax	XP_049718943.1	Fox	Fox-P1	Ttru	XP_033719780.1	Sox	Sox-4
Mjav	XP_036860335.1	Fox	Fox-P1	Cpor	XP_003468908.2	Sox	Sox-4
Pgig	XP_039707610.1	Fox	Fox-P1	Mdom	XP_007487941.1	Sox	Sox-4
Drot	XP_045048358.1	Fox	Fox-P1	Scar	XP_047414294.1	Sox	Sox-4
Mmus	XP_030110934.1	Fox	Fox-P1	Hsap	NP_003098.1	Sox	Sox-4
Tman	XP_023583661.1	Fox	Fox-P1	Bbub	XP_025121975.3	Sox	Sox-4
Cimi	XP_017398291.1	Fox	Fox-P1	Hamp	XP_057555091.1	Sox	Sox-4
Rfer	XP_032987798.1	Fox	Fox-P1	Mmus	NP_033264.2	Sox	Sox-4
Scar	XP_047390767.1	Fox	Fox-P1	Drot	XP_053776704.1	Sox	Sox-4
Ptig	XP_042835351.1	Fox	Fox-P1	Oafe	XP_007950280.1	Sox	Sox-4
Oana	XP_039766065.1	Fox	Fox-P1	Dnov	XP_012385001.3	Sox	Sox-4
Equa	XP_046523540.1	Fox	Fox-P1	Clup	XP_038440195.1	Sox	Sox-4
Clup	XP_038421798.1	Fox	Fox-P1	Equa	XP_046497050.1	Sox	Sox-4
Mang	XP_054360611.1	Fox	Fox-P1	Amel	XP_034516148.1	Sox	Sox-4
Bmus	XP_036724195.1	Fox	Fox-P1	Cdro	XP_031290990.1	Sox	Sox-4
Cpor	XP_005005313.1	Fox	Fox-P1	Emax	XP_049732340.1	Sox	Sox-4
Cdid	XP_037656169.1	Fox	Fox-P1	Csim	XP_004432138.1	Sox	Sox-4
Hamp	XP_057561364.1	Fox	Fox-P1	Mjav	XP_036850086.1	Sox	Sox-4
Lcat	XP_045385591.1	Fox	Fox-P1	Pgig	XP_039722376.1	Sox	Sox-4
Bbub	XP_044789782.1	Fox	Fox-P1	Ptig	XP_042841543.1	Sox	Sox-4
Csim	XP_004419976.1	Fox	Fox-P1	Bmus	XP_036725088.1	Sox	Sox-4
Dnov	XP_004482088.1	Fox	Fox-P1	Tman	XP_023594181.1	Sox	Sox-4
Ggal	XP_040502196.1	Fox	Fox-P1	Pafr	XP_047651758.1	Sox	Sox-4
Tman	XP_004385433.1	Fox	Fox-N1	Mang	XP_045722257.1	Sox	Sox-4
Pafr	XP_047614767.1	Fox	Fox-N1	Rfer	XP_032971451.1	Sox	Sox-4
Cdid	XP_037665986.1	Fox	Fox-N1	Cdid	XP_037700378.1	Sox	Sox-4
Casi	XP_006874270.1	Fox	Fox-N1	Cimi	XP_017399572.1	Sox	Sox-4
Emax	XP_049715494.1	Fox	Fox-N1	Shar	XP_031802646.1	Sox	Sox-4
Scar	XP_047401997.1	Fox	Fox-N1	Shar	XP_003775357.4	Sox	Sox-15
Oafe	XP_007935599.1	Fox	Fox-N1	Dnov	XP_004468798.1	Sox	Sox-15
Ggal	XP_415816.5	Fox	Fox-N1	Mdom	XP_056673573.1	Sox	Sox-15
Mmus	XP_006532328.1	Fox	Fox-N1	Oana	XP_039766818.1	Sox	Sox-15
Oana	XP_039770471.1	Fox	Fox-N1	Clup	XP_038392636.1	Sox	Sox-15
Mdom	XP_001375832.1	Fox	Fox-N1	Mjav	XP_017527261.1	Sox	Sox-15
Dnov	XP_004450005.2	Fox	Fox-N1	Cdid	XP_037664941.1	Sox	Sox-15
Shar	XP_031823539.1	Fox	Fox-N1	Pgig	XP_039703166.1	Sox	Sox-15
Cpor	XP_023420283.1	Fox	Fox-N1	Pafr	XP_047613485.1	Sox	Sox-15
Lcat	XP_045381339.1	Fox	Fox-N1	Cimi	XP_037595583.1	Sox	Sox-15
Bbub	XP_025136790.1	Fox	Fox-N1	Hamp	XP_057571542.1	Sox	Sox-15
Opri	XP_004593965.2	Fox	Fox-N1	Equa	XP_046531541.1	Sox	Sox-15
Bmus	XP_036693663.1	Fox	Fox-N1	Ptig	XP_042823564.1	Sox	Sox-15
Drot	XP_045055001.2	Fox	Fox-N1	Cpor	XP_030466299.1	Sox	Sox-15
Amel	XP_034496649.1	Fox	Fox-N1	Rfer	XP_032947056.1	Sox	Sox-15
Pgig	XP_039742030.1	Fox	Fox-N1	Cimi	XP_037595583.1	Sox	Sox-15
Rfer	XP_032947344.1	Fox	Fox-N1	Hamp	XP_057571542.1	Sox	Sox-15
Mang	XP_045744008.1	Fox	Fox-N1	Equa	XP_046531541.1	Sox	Sox-15
Equa	XP_046533715.1	Fox	Fox-N1	Ptig	XP_042823564.1	Sox	Sox-15
Cimi	XP_037599212.1	Fox	Fox-N1	Lcat	XP_045426075.1	Sox	Sox-15
Mjav	XP_017515466.1	Fox	Fox-N1	Cdro	XP_03123594.1	Sox	Sox-15
Hamp	XP_057571196.1	Fox	Fox-N1	Mang	XP_045746055.1	Sox	Sox-15
Hsap	XP_011523660.1	Fox	Fox-N1	Drot	XP_024420518.1	Sox	Sox-15
Ptig	XP_007076110.1	Fox	Fox-N1	Tman	XP_004376270.1	Sox	Sox-15
Ttru	XP_033703421.1	Fox	Fox-N1	Bbub	XP_006062923.4	Sox	Sox-15
Cdro	XP_031324225.1	Fox	Fox-N1	Hsap	NP_008873.1	Sox	Sox-15
Csim	XP_014645264.1	Fox	Fox-N1	Oafe	XP_007950453.1	Sox	Sox-15
Clup	XP_038404162.1	Fox	Fox-N1	Scar	XP_047400352.1	Sox	Sox-15

Tab. S8 continued from previous page

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Shar	XP_003760278.3	Fox	Fox-F2	Mmus	NP_033261.1	Sox	Sox-15
Drot	XP_024408147.2	Fox	Fox-F2	Csim	XP_004433180.1	Sox	Sox-15
Mdom	XP_001378770.3	Fox	Fox-F2	Amel	XP_019660673.2	Sox	Sox-15
Casi	XP_006870627.1	Fox	Fox-F2	Casi	XP_006863359.1	Sox	Sox-15
Equa	XP_046496582.1	Fox	Fox-F2	Rfer	XP_032974648.1	Sox	-
Bmus	XP_036725085.1	Fox	Fox-F2	Pgig	XP_039738421.1	Sox	-
Lcat	XP_045408674.1	Fox	Fox-F2	Dnov	XP_058138887.1	Sox	-
Opri	XP_012785618.3	Fox	Fox-F2	Drot	XP_024431130.1	Sox	-
Tman	XP_012414385.1	Fox	Fox-F2	Ggal	XP_015151955.3	Sox	Sox-12
Mjav	XP_036867662.1	Fox	Fox-F2	Equa	XP_046533915.1	Sox	Sox-12
Cdro	XP_010975130.2	Fox	Fox-F2	Emax	XP_049725681.1	Sox	Sox-12
Hsap	NP_001443.1	Fox	Fox-F2	Rfer	XP_032951289.1	Sox	Sox-12
Emax	XP_049738291.1	Fox	Fox-F2	Cpor	XP_005006612.1	Sox	Sox-12
Pgig	XP_039725405.1	Fox	Fox-F2	Bmus	XP_036683151.1	Sox	Sox-12
Csim	XP_004419240.1	Fox	Fox-F2	Tman	XP_004370660.1	Sox	Sox-12
Cdid	XP_037700041.1	Fox	Fox-F2	Bbub	XP_025119497.1	Sox	Sox-12
Dnov	XP_023447171.2	Fox	Fox-F2	Mang	XP_045741538.1	Sox	Sox-12
Cimi	XP_017352993.2	Fox	Fox-F2	Opri	XP_058535679.1	Sox	Sox-12
Oafe	XP_007933567.1	Fox	Fox-F2	Cdid	XP_037667390.1	Sox	Sox-12
Ptig	XP_042841202.1	Fox	Fox-F2	Pafr	XP_047627358.1	Sox	Sox-12
Mmus	NP_034355.2	Fox	Fox-F2	Mmus	NP_035568.1	Sox	Sox-12
Scar	XP_047414417.1	Fox	Fox-F2	Csim	XP_014645525.1	Sox	Sox-12
Bbub	XP_025120520.3	Fox	Fox-F2	Ttru	XP_004331924.3	Sox	Sox-12
Mang	XP_045721916.1	Fox	Fox-F2	Scar	XP_047398506.1	Sox	Sox-12
Amel	XP_034516095.1	Fox	Fox-F2	Ptig	XP_042836628.1	Sox	Sox-12
Rfer	XP_032969138.1	Fox	Fox-F2	Shar	XP_031810003.1	Sox	Sox-12
Clup	XP_038439937.1	Fox	Fox-F2	Pgig	XP_039719928.1	Sox	Sox-12
Ttru	XP_004311990.3	Fox	Fox-F2	Casi	XP_006860781.1	Sox	Sox-12
Pafr	XP_047651459.1	Fox	Fox-F2	Hsap	NP_008874.2	Sox	Sox-12
Hamp	XP_057555617.1	Fox	Fox-F2	Oafe	XP_007932707.1	Sox	Sox-12
Ggal	XP_046759228.1	Fox	-	Mjav	XP_017521282.2	Sox	Sox-12
Mdom	XP_007491677.2	Fox	-	Clup	XP_038427830.1	Sox	Sox-12
Shar	XP_031817418.1	Fox	-	Dnov	XP_058142829.1	Sox	Sox-12
Oana	XP_028911073.1	Fox	Fox-L2	Hamp	XP_057559844.1	Sox	Sox-12
Rfer	XP_032988626.1	Fox	Fox-L2	Lcat	XP_045385476.1	Sox	Sox-12
Ggal	NP_001012630.1	Fox	Fox-L2	Drot	XP_024415008.3	Sox	Sox-12
Mdom	XP_007494005.2	Fox	Fox-L2	Cdro	XP_031289888.1	Sox	Sox-12
Emax	XP_049726166.1	Fox	Fox-L2	Cimi	XP_017370646.1	Sox	Sox-12
Hsap	NP_075555.1	Fox	Fox-L2	Mdom	XP_007474545.2	Sox	Sox-12
Amel	XP_034517711.1	Fox	Fox-L2	Oana	XP_028907426.1	Sox	Sox-11
Drot	XP_024421665.2	Fox	Fox-L2	Cdro	XP_031322369.1	Sox	Sox-11
Mang	XP_045719675.1	Fox	Fox-L2	Pafr	XP_047635202.1	Sox	Sox-11
Tman	XP_004381516.1	Fox	Fox-L2	Cdid	XP_037669298.1	Sox	Sox-11
Casi	XP_006846679.1	Fox	Fox-L2	Mdom	XP_007476236.1	Sox	Sox-11
Csim	XP_004419996.1	Fox	Fox-L2	Dnov	XP_058143908.1	Sox	Sox-11
Mmus	NP_036150.1	Fox	Fox-L2	Mang	XP_045739658.1	Sox	Sox-11
Pafr	XP_047639332.1	Fox	Fox-L2	Casi	XP_006864036.1	Sox	Sox-11
Bbub	XP_025146742.1	Fox	Fox-L2	Tman	XP_004373138.1	Sox	Sox-11
Opri	XP_012783325.2	Fox	Fox-L2	Hamp	XP_057599420.1	Sox	Sox-11
Shar	XP_031815678.1	Fox	Fox-L2	Pgig	XP_039710941.1	Sox	Sox-11
Hamp	XP_057595697.1	Fox	Fox-L2	Cimi	XP_017397486.1	Sox	Sox-11
Cimi	XP_017379188.1	Fox	Fox-L2	Lcat	XP_045404807.1	Sox	Sox-11
Cdid	XP_037683851.1	Fox	Fox-L2	Emax	XP_049759749.1	Sox	Sox-11
Equa	XP_046508064.1	Fox	Fox-L2	Ptig	XP_042835747.1	Sox	Sox-11
Mjav	XP_036847828.1	Fox	Fox-L2	Shar	XP_031807046.1	Sox	Sox-11
Cdro	XP_031299319.1	Fox	Fox-L2	Cpor	XP_03464995.1	Sox	Sox-11
Lcat	XP_045411920.1	Fox	Fox-L2	Mjav	XP_017522349.2	Sox	Sox-11
Clup	XP_038426115.1	Fox	Fox-L2	Scar	XP_047397943.1	Sox	Sox-11
Cpor	XP_023415973.1	Fox	Fox-L2	Opri	XP_004582643.2	Sox	Sox-11
Ptig	XP_042812852.1	Fox	Fox-L2	Clup	XP_038416510.1	Sox	Sox-11
Oafe	XP_007945699.1	Fox	Fox-L2	Drot	XP_024408195.2	Sox	Sox-11
Ttru	XP_019789813.1	Fox	Fox-L2	Ggal	NP_001382976.1	Sox	Sox-11
Pgig	XP_039692762.1	Fox	Fox-L2	Equa	XP_046516078.1	Sox	Sox-11
Bmus	XP_036707130.1	Fox	Fox-L2	Amel	XP_034515849.1	Sox	Sox-11
Scar	XP_047420926.1	Fox	Fox-L2	Hsap	NP_003099.1	Sox	Sox-11
Oana	XP_028902621.2	Fox	Fox-L3	Oafe	XP_007933317.1	Sox	Sox-11
Bmus	XP_036729925.1	Fox	Fox-L3	Bmus	XP_036729557.1	Sox	Sox-11
Lcat	XP_045404844.1	Fox	Fox-L3	Bbub	XP_025117398.3	Sox	Sox-11
Casi	XP_006875916.1	Fox	Fox-L3	Rfer	XP_032980233.1	Sox	Sox-11
Dnov	XP_004472353.2	Fox	Fox-L3	Ttru	XP_019798147.1	Sox	Sox-11
Emax	XP_049713621.1	Fox	Fox-L3	Csim	XP_014647619.1	Sox	Sox-11
Csim	XP_004437309.2	Fox	Fox-L3	Mmus	NP_033260.4	Sox	Sox-11
Cimi	XP_017392139.1	Fox	Fox-L3	Mang	NP_989526.1	Sry	-
Bbub	XP_025116942.2	Fox	Fox-L3	Shar	XP_023362772.2	Sox	Sox-3
Pgig	XP_039708536.1	Fox	Fox-L3	Ptig	XP_042831086.1	Sox	Sox-3
Cdid	XP_037663497.1	Fox	Fox-L3	Dnov	XP_058147425.1	Sox	Sox-3
Tman	XP_004390712.1	Fox	Fox-L3	Rfer	XP_032973530.1	Sox	Sox-3
Clup	NP_001129118.1	Fox	Fox-L3	Oafe	XP_007949915.1	Sox	Sox-3
Opri	XP_004590794.2	Fox	Fox-L3	Csim	XP_014651689.1	Sox	Sox-3
Pafr	XP_047635149.1	Fox	Fox-L3	Drot	XP_024434131.1	Sry	-
Hamp	XP_057599300.1	Fox	Fox-L3	Hsap	NP_003131.1	Sry	-
Mang	XP_045739697.1	Fox	Fox-L3	Equa	XP_046528503.1	Sox	Sox-3
Amel	XP_002922377.3	Fox	Fox-L3	Emax	XP_049728984.1	Sox	Sox-3
Ttru	XP_033694400.1	Fox	Fox-L3	Opri	XP_058519480.1	Sox	Sox-3
Scar	XP_047393964.1	Fox	Fox-L3	Ttru	XP_033705620.1	Sox	Sox-3
Hsap	NP_001129121.1	Fox	Fox-L3	Clup	XP_038442551.1	Sox	Sox-3
Mmus	NP_001094934.1	Fox	Fox-L3	Oana	XP_001520781.2	Sry	-
Rfer	XP_032980088.1	Fox	Fox-L3	Hsap	NP_005625.2	Sox	Sox-3
Ptig	XP_042837881.1	Fox	Fox-L3	Mjav	XP_036859504.1	Sox	Sox-3
Equa	XP_046517777.1	Fox	Fox-L3	Mdom	XP_056665898.1	Sry	-
Cdro	XP_010996180.2	Fox	Fox-L3	Emax	XP_049729443.1	Sry	-
Mjav	XP_017536768.2	Fox	Fox-L3	Cdro	XP_031300930.1	Sox	Sox-3
Oafe	XP_007952295.1	Fox	Fox-L3	Pgig	XP_039735027.1	Sry	-
Drot	XP_024408151.2	Fox	Fox-L3	Shar	XP_031801149.1	Sry	-
Oana	XP_028931929.2	Fox	Fox-L1	Lcat	XP_045393696.1	Sox	Sox-3
Bbub	XP_006047558.1	Fox	Fox-L1	Cimi	XP_017374345.1	Sry	-
Mdom	XP_056674514.1	Fox	Fox-L1	Hamp	XP_0575744656.1	Sox	Sox-3
Oafe	XP_007937407.1	Fox	Fox-L1	Mmus	NP_033263.2	Sox	Sox-3
Mjav	XP_036849629.1	Fox	Fox-L1	Amel	XP_034504946.1	Sox	Sox-3
Rfer	XP_032984676.1	Fox	Fox-L1	Cdid	XP_037678811.1	Sox	Sox-3
Hamp	XP_057569166.1	Fox	Fox-L1	Bmus	XP_036696323.1	Sox	Sox-3
Pgig	XP_039744660.1	Fox	Fox-L1	Ttru	XP_033706224.1	Sry	-

Tab. S8 continued from previous page

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Dnov	XP_058135467.1	Fox	Fox-L1	Bbub	XP_025132502.3	Sox	Sox-3
Csim	XP_004437100.1	Fox	Fox-L1	Cdid	XP_037680520.1	Sox	Sry
Casi	XP_006860180.1	Fox	Fox-L1	Cpor	XP_003464754.2	Sox	Sox-3
Opri	XP_004584130.2	Fox	Fox-L1	Scar	XP_047393176.1	Sox	Sry
Clup	XP_038394040.1	Fox	Fox-L1	Opri	XP_058515261.1	Sox	Sry
Hsap	NP_005241.1	Fox	Fox-L1	Pafr	XP_047620863.1	Sox	Sox-3
Drot	XP_024411663.1	Fox	Fox-L1	Mmus	NP_035694.1	Sox	Sry
Amel	XP_002913428.1	Fox	Fox-L1	Drot	XP_053773489.1	Sox	Sox-3
Cdro	XP_031292484.1	Fox	Fox-L1	Mdom	XP_007507433.1	Sox	Sox-3
Cimi	XP_017399161.1	Fox	Fox-L1	Scar	XP_047392867.1	Sox	Sox-3
Cdid	XP_037671791.1	Fox	Fox-L1	Oafe	XP_007957280.1	Sox	Sry
Ttru	XP_004312143.1	Fox	Fox-L1	Cimi	XP_017355427.1	Sox	Sox-3
Rfer	XP_032984241.1	Fox	Fox-L1	Ttru	XP_033706225.1	Sox	Sry
Ptig	XP_042825075.1	Fox	Fox-L1	Mang	XP_045726266.1	Sox	Sox-3
Bmus	XP_036688240.1	Fox	Fox-L1	Casi	XP_006875033.1	Sox	Sox-3
Tman	XP_004377800.1	Fox	Fox-L1	Opri	XP_058515255.1	Sox	Sry
Mang	XP_045746246.1	Fox	Fox-L1	Mjav	XP_036857050.1	Sox	Sry
Shar	XP_031806032.1	Fox	Fox-L1	Dnov	XP_058142693.1	Sox	Sox-8
Mmus	NP_032050.2	Fox	Fox-L1	Casi	XP_006873911.1	Sox	Sox-8
Paf	XP_047648182.1	Fox	Fox-L1	Cdid	XP_037670405.1	Sox	Sox-8
Equa	XP_046536521.1	Fox	Fox-L1	Emax	XP_049759218.1	Sox	Sox-8
Scar	XP_047385703.1	Fox	Fox-L1	Tman	XP_023584101.1	Sox	Sox-8
Ggal	XP_001231599.5	Fox	Fox-L1	Oafe	XP_007955685.1	Sox	Sox-8
Emax	XP_049720646.1	Fox	Fox-L1	Mjav	XP_017510601.1	Sox	-
Lcat	XP_045389091.1	Fox	Fox-L1	Clup	XP_038432983.1	Sox	-
Ggal	XP_004948470.2	Fox	Fox-P4	Ptig	XP_042847878.1	Sox	-
Casi	XP_006860505.1	Fox	Fox-P4	Mang	XP_054363126.1	Sox	-
Equa	XP_046496143.1	Fox	Fox-P4	Amel	XP_019650085.1	Sox	-
Cdid	XP_037698927.1	Fox	Fox-P4	Ggal	NP_098664.1	Sox	Sox-1
Mdom	XP_056674181.1	Fox	Fox-P4	Mjav	XP_036880873.1	Sox	Sox-1
Oafe	XP_007934458.1	Fox	Fox-P4	Amel	XP_034519706.1	Sox	Sox-1
Opri	XP_058519292.1	Fox	Fox-P4	Lcat	XP_045422689.1	Sox	Sox-1
Mmus	XP_017173175.1	Fox	Fox-P4	Oana	XP_028904097.1	Sox	Sox-1
Rfer	XP_032956861.1	Fox	Fox-P4	Opri	XP_058526612.1	Sox	Sox-1
Hsap	XP_011512591.1	Fox	Fox-P4	Cdid	XP_037656244.1	Sox	Sox-1
Ptig	XP_042842255.1	Fox	Fox-P4	Scar	XP_047410448.1	Sox	Sox-1
Ttru	XP_033720169.1	Fox	Fox-P4	Mdom	XP_007501341.1	Sox	Sox-1
Mang	XP_045722826.1	Fox	Fox-P4	Equa	XP_046519750.1	Sox	Sox-1
Mjav	XP_036861021.1	Fox	Fox-P4	Ttru	XP_019776483.2	Sox	Sox-1
Cimi	XP_017402725.1	Fox	Fox-P4	Clup	XP_038425993.1	Sox	Sox-1
Clup	XP_0384009938.1	Fox	Fox-P4	Cdro	XP_031322193.1	Sox	Sox-1
Shar	XP_031820748.1	Fox	Fox-P4	Shar	XP_031814590.1	Sox	Sox-1
Lcat	XP_045397843.1	Fox	Fox-P4	Oafe	XP_007940578.1	Sox	Sox-1
Ptig	XP_039710787.1	Fox	Fox-P4	Rfer	XP_032959657.1	Sox	Sox-1
Drot	XP_053769706.1	Fox	Fox-P4	Mmus	NP_033259.2	Sox	Sox-1
Bmus	XP_036725368.1	Fox	Fox-P4	Dnov	XP_058132394.1	Sox	Sox-1
Amel	XP_002914563.3	Fox	Fox-P4	Bmus	XP_036687021.1	Sox	Sox-1
Csim	XP_004424223.1	Fox	Fox-P4	Paf	XP_047612043.1	Sox	Sox-1
Oana	XP_028925009.1	Fox	Fox-P4	Drot	XP_045042640.2	Sox	Sox-1
Hamp	XP_057555834.1	Fox	Fox-P4	Ptig	XP_042814197.1	Sox	Sox-1
Paf	XP_047653356.1	Fox	Fox-P4	Cimi	XP_017357713.1	Sox	Sox-1
Cpor	XP_003473920.1	Fox	Fox-P4	Mang	XP_045723818.1	Sox	Sox-1
Scar	XP_047414507.1	Fox	Fox-P4	Casi	XP_006851579.1	Sox	Sox-1
Emax	XP_049747581.1	Fox	Fox-P4	Cpor	XP_005007254.3	Sox	Sox-1
Tman	XP_004379494.1	Fox	Fox-P4	Hsap	NP_005977.2	Sox	Sox-1
Dnov	XP_058162686.1	Fox	Fox-P4	Hamp	XP_057563730.1	Sox	Sox-1
Bbub	XP_025125991.2	Fox	Fox-P4	Ptig	XP_039721974.1	Sox	Sox-1
Cdro	XP_031290589.1	Fox	Fox-P4	Bbub	XP_025118363.1	Sox	Sox-1
Bmus	XP_036718879.1	Fox	Fox-P2	Emax	XP_049723386.1	Sox	Sox-1
Ttru	XP_019780690.1	Fox	Fox-P2	Mdom	XP_001373727.1	Sox	-
Lcat	XP_045420440.1	Fox	Fox-P2	Shar	XP_003762017.1	Sox	-
Ptig	XP_039716577.1	Fox	Fox-P2	Hamp	XP_057557078.1	Sox	Sox-5
Csim	XP_004418850.1	Fox	Fox-P2	Equa	XP_046497249.1	Sox	Sox-5
Shar	XP_031794706.1	Fox	Fox-P2	Bbub	XP_045021534.1	Sox	Sox-5
Equa	XP_046526335.1	Fox	Fox-P2	Csim	XP_014646859.1	Sox	Sox-5
Cdro	XP_031311093.1	Fox	Fox-P2	Cdro	XP_031299822.1	Sox	Sox-5
Amel	XP_034526782.1	Fox	Fox-P2	Tman	XP_023593782.1	Sox	Sox-5
Ptig	XP_042826228.1	Fox	Fox-P2	Emax	XP_049741103.1	Sox	Sox-5
Opri	XP_058510875.1	Fox	Fox-P2	Oafe	XP_007944956.1	Sox	Sox-5
Cpor	XP_013013912.1	Fox	Fox-P2	Paf	XP_047643337.1	Sox	Sox-5
Casi	XP_006859359.1	Fox	Fox-P2	Casi	XP_006867010.1	Sox	Sox-5
Oana	XP_028928988.1	Fox	Fox-P2	Mdom	XP_001368820.2	Sox	Sox-2
Paf	XP_047618861.1	Fox	Fox-P2	Csim	XP_004424739.1	Sox	Sox-2
Oafe	XP_007942246.1	Fox	Fox-P2	Emax	XP_049730807.1	Sox	Sox-2
Hsap	NP_683696.2	Fox	Fox-P2	Casi	XP_006869879.1	Sox	Sox-2
Mdom	XP_007504226.1	Fox	Fox-P2	Ptig	XP_042855191.1	Sox	Sox-2
Ggal	XP_025007337.1	Fox	Fox-P2	Ggal	NP_990519.3	Sox	Sox-2
Dnov	XP_058153204.1	Fox	Fox-P2	Clup	XP_038439395.1	Sox	Sox-2
Bbub	XP_044802585.1	Fox	Fox-P2	Equa	XP_046515240.1	Sox	Sox-2
Cimi	XP_017390531.1	Fox	Fox-P2	Hamp	XP_057595873.1	Sox	Sox-2
Rfer	XP_032954948.1	Fox	Fox-P2	Hsap	NP_003097.1	Sox	Sox-2
Hamp	XP_057586867.1	Fox	Fox-P2	Shar	XP_031813587.1	Sox	Sox-2
Mmus	XP_036021645.1	Fox	Fox-P2	Mang	XP_045719633.1	Sox	Sox-2
Mjav	XP_036883368.1	Fox	Fox-P2	Scar	XP_047420710.1	Sox	Sox-2
Emax	XP_049750594.1	Fox	Fox-P2	Tman	XP_023592536.1	Sox	Sox-2
Cdid	XP_037692269.1	Fox	Fox-P2	Bbub	XP_006056297.2	Sox	Sox-2
Drot	XP_053782699.1	Fox	Fox-P2	Rfer	XP_032989610.1	Sox	Sox-2
Scar	XP_047417295.1	Fox	Fox-P2	Lcat	XP_045395708.1	Sox	Sox-2
Mang	XP_045720337.1	Fox	Fox-P2	Opri	XP_058518203.1	Sox	Sox-2
Tman	XP_004382624.1	Fox	Fox-P2	Oana	XP_028928807.1	Sox	Sox-2
Clup	XP_038413332.1	Fox	Fox-P2	Bmus	XP_036705414.1	Sox	Sox-2
Cdid	XP_037688024.1	Fox	-	Mmus	NP_035573.3	Sox	Sox-2
Dnov	XP_058148549.1	Fox	-	Cdid	XP_037695755.1	Sox	Sox-2
Ggal	XP_414186.5	Fox	Fox-F1	Ptig	XP_039706608.1	Sox	Sox-2
Emax	XP_049720665.1	Fox	Fox-F1	Oafe	XP_007934094.1	Sox	Sox-2
Mdom	XP_001365832.4	Fox	Fox-F1	Paf	XP_047643165.1	Sox	Sox-2
Hsap	NP_001442.2	Fox	Fox-F1	Dnov	XP_004477690.1	Sox	Sox-2
Clup	XP_038394042.1	Fox	Fox-F1	Cimi	XP_017393930.1	Sox	Sox-2
Mjav	XP_017511561.2	Fox	Fox-F1	Drot	XP_053774095.1	Sox	Sox-2
Cimi	XP_017399155.2	Fox	Fox-F1	Amel	XP_034518206.1	Sox	Sox-2
Bbub	XP_006047564.2	Fox	Fox-F1	Cdro	XP_010974669.2	Sox	Sox-2
Dnov	XP_058135594.1	Fox	Fox-F1	Mjav	XP_017496634.2	Sox	Sox-2

Tab. S8 continued from previous page

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Drot	XP_053772163.1	Fox	Fox-F1	Ttru	XP_004311832.3	Sox	Sox-2
Tman	XP_004377797.2	Fox	Fox-F1	Oana	XP_028931165.1	Sox	Sox-14
Bmus	XP_036688072.1	Fox	Fox-F1	Mdom	XP_007493983.1	Sox	Sox-14
Oana	XP_028931561.1	Fox	Fox-F1	Scar	XP_047419052.1	Sox	Sox-14
Pafr	XP_047645925.1	Fox	Fox-F1	Cdid	XP_037683894.1	Sox	Sox-14
Scar	XP_047385701.1	Fox	Fox-F1	Bmus	XP_036707689.1	Sox	Sox-14
Lcat	XP_045389195.1	Fox	Fox-F1	Mjav	XP_017531607.1	Sox	Sox-14
Shar	XP_003758519.2	Fox	Fox-F1	Casi	XP_006846671.1	Sox	Sox-14
Amel	XP_002913450.2	Fox	Fox-F1	Pafr	XP_047610665.1	Sox	Sox-14
Opri	XP_058530279.1	Fox	Fox-F1	Hamp	XP_05795112.1	Sox	Sox-14
Pgig	XP_039744668.1	Fox	Fox-F1	Amel	XP_002923375.1	Sox	Sox-14
Casi	XP_006860182.1	Fox	Fox-F1	Csim	XP_004419391.1	Sox	Sox-14
Mmus	NP_034556.2	Fox	Fox-F1	Cpor	XP_003476788.1	Sox	Sox-14
Cdid	XP_037671853.1	Fox	Fox-F1	Hsap	NP_004180.1	Sox	Sox-14
Cpor	XP_003460772.2	Fox	Fox-F1	Clup	XP_038426113.1	Sox	Sox-14
Equa	XP_046538978.1	Fox	Fox-F1	Bbub	XP_025147052.1	Sox	Sox-14
Ptig	XP_007082882.2	Fox	Fox-F1	Rfer	XP_032988974.1	Sox	Sox-14
Csim	XP_014648135.1	Fox	Fox-F1	Equa	XP_046511648.1	Sox	Sox-14
Cdro	XP_031292478.1	Fox	Fox-F1	Drot	XP_024422013.1	Sox	Sox-14
Rfer	XP_032985403.1	Fox	Fox-F1	Lcat	XP_045395088.1	Sox	Sox-14
Mang	XP_045746573.2	Fox	Fox-F1	Opri	XP_00458396.1	Sox	Sox-14
Hamp	XP_057568154.1	Fox	Fox-F1	Pgig	XP_039692761.1	Sox	Sox-14
Oafe	XP_007937340.1	Fox	Fox-F1	Ptig	XP_042813213.1	Sox	Sox-14
Ttru	XP_033700961.1	Fox	Fox-F1	Ttru	XP_019789673.1	Sox	Sox-14
Mdom	XP_056669097.1	Fox	-	Cimi	XP_017383705.1	Sox	Sox-14
Shar	XP_031805673.1	Fox	-	Emax	XP_049726472.1	Sox	Sox-14
Ggal	XP_425714.5	Fox	Fox-S1	Mang	XP_045719678.1	Sox	Sox-14
Clup	XP_038427866.1	Fox	Fox-S1	Oafe	XP_007935393.1	Sox	Sox-14
Hsap	NP_004109.1	Fox	Fox-S1	Ggal	NP_990092.1	Sox	Sox-14
Pgig	XP_039719969.1	Fox	Fox-S1	Tman	XP_004381489.1	Sox	Sox-14
Bmus	XP_036682343.1	Fox	Fox-S1	Dnov	XP_004453833.1	Sox	Sox-14
Mdom	XP_001364156.2	Fox	Fox-S1	Shar	XP_012400053.1	Sox	Sox-14
Amel	XP_002913251.1	Fox	Fox-S1	Mmus	NP_035570.1	Sox	Sox-14
Shar	XP_031809982.1	Fox	Fox-S1	Cdro	XP_010975059.1	Sox	Sox-14
Bbub	XP_025119474.1	Fox	Fox-S1	Opri	XP_058526666.1	Sox	Sox-21
Casi	XP_006860769.1	Fox	Fox-S1	Mang	XP_045726097.1	Sox	Sox-21
Cdro	XP_010992198.2	Fox	Fox-S1	Rfer	XP_032959615.1	Sox	Sox-21
Mang	XP_045741514.1	Fox	Fox-S1	Bmus	XP_036687740.1	Sox	Sox-21
Pafr	XP_047627330.1	Fox	Fox-S1	Dnov	XP_058132486.1	Sox	Sox-21
Dnov	XP_004464085.1	Fox	Fox-S1	Equa	XP_046520974.1	Sox	Sox-21
Opri	XP_004585763.2	Fox	Fox-S1	Drot	XP_053772754.1	Sox	Sox-21
Cpor	XP_003476682.1	Fox	Fox-S1	Opri	XP_058526666.1	Sox	Sox-21
Mjav	XP_017509202.2	Fox	Fox-S1	Cdid	XP_037656473.1	Sox	Sox-21
Csim	XP_004442549.2	Fox	Fox-S1	Csim	XP_014637259.1	Sox	Sox-21
Rfer	XP_032950864.1	Fox	Fox-S1	Shar	XP_031814415.1	Sox	Sox-21
Equa	XP_046536068.1	Fox	Fox-S1	Ttru	XP_033699347.1	Sox	Sox-21
Scar	XP_047396619.1	Fox	Fox-S1	Tman	XP_004371490.1	Sox	Sox-21
Tman	XP_023581200.1	Fox	Fox-S1	Mmus	NP_808421.1	Sox	Sox-21
Lcat	XP_045385439.1	Fox	Fox-S1	Oana	XP_001512271.3	Sox	Sox-21
Emax	XP_049725743.1	Fox	Fox-S1	Clup	XP_038425312.1	Sox	Sox-21
Mmus	NP_034356.1	Fox	Fox-S1	Ptig	XP_042813926.1	Sox	Sox-21
Ttru	XP_033696063.1	Fox	Fox-S1	Amel	XP_034519700.1	Sox	Sox-21
Ptig	XP_007073610.2	Fox	Fox-S1	Casi	XP_006832065.1	Sox	Sox-21
Hamp	XP_057559118.1	Fox	Fox-S1	Hamp	XP_057563720.1	Sox	Sox-21
Drot	XP_024415665.3	Fox	Fox-S1	Cdro	XP_031322066.1	Sox	Sox-21
Cimi	XP_017370299.1	Fox	Fox-S1	Mang	XP_045726110.1	Sox	Sox-21
Oafe	XP_007932722.1	Fox	Fox-S1	Pgig	XP_039693387.1	Sox	Sox-21
Cdid	XP_037667492.1	Fox	Fox-S1	Mjav	XP_036880860.1	Sox	Sox-21
Ggal	XP_015135808.1	Fox	Fox-B2	Scar	XP_047410577.1	Sox	Sox-21
Pafr	XP_047625507.1	Fox	Fox-B2	Pafr	XP_047612344.1	Sox	Sox-21
Shar	XP_012398564.2	Fox	Fox-B2	Oafe	XP_007942198.1	Sox	Sox-21
Csim	XP_004439686.1	Fox	Fox-B2	Cimi	XP_017399045.1	Sox	Sox-21
Scar	XP_047379799.1	Fox	Fox-B2	Mdom	XP_007501455.1	Sox	Sox-21
Cdro	XP_031305772.1	Fox	Fox-B2	Bbub	XP_025118466.2	Sox	Sox-21
Equa	XP_046519944.1	Fox	Fox-B2	Lcat	XP_045423694.1	Sox	Sox-21
Bbub	XP_006067725.3	Fox	Fox-B2	Hsap	NP_009015.1	Sox	Sox-21
Cimi	XP_017398753.1	Fox	Fox-B2	Emax	XP_049709053.1	Sox	Sox-21
Mmus	NP_032049.1	Fox	Fox-B2	Scar	XP_047405480.1	Sox	Sox-5
Pgig	XP_039724438.1	Fox	Fox-B2	Opri	XP_058511892.1	Sox	Sox-5
Lcat	XP_045418554.1	Fox	Fox-B2	Scar	XP_047405480.1	Sox	Sox-5
Casi	XP_006834992.1	Fox	Fox-B2	Mmus	XP_006506994.1	Sox	Sox-5
Mdom	XP_003341714.1	Fox	Fox-B2	Bmus	XP_036693736.1	Sox	Sox-9
Mjav	XP_036847034.1	Fox	Fox-B2	Hamp	XP_057573119.1	Sox	Sox-9
Oana	XP_001508095.2	Fox	Fox-B2	Ggal	NP_989612.1	Sox	Sox-9
Mang	XP_045737891.1	Fox	Fox-B2	Drot	XP_024429046.1	Sox	Sox-9
Drot	XP_053771141.1	Fox	Fox-B2	Tman	XP_004374235.1	Sox	Sox-9
Dnov	XP_023440163.2	Fox	Fox-B2	Cimi	XP_017391090.1	Sox	Sox-9
Opri	XP_058513381.1	Fox	Fox-B2	Clup	NP_001002978.1	Sox	Sox-9
Ttru	XP_019789709.1	Fox	Fox-B2	Dnov	XP_058140625.1	Sox	Sox-9
Cdid	XP_037654560.1	Fox	Fox-B2	Opri	XP_004593006.1	Sox	Sox-9
Tman	XP_023593359.1	Fox	Fox-B2	Casi	XP_006872359.1	Sox	Sox-9
Amel	XP_034502902.1	Fox	Fox-B2	Cdro	XP_031304465.1	Sox	Sox-9
Bmus	XP_036712759.1	Fox	Fox-B2	Mjav	XP_017519930.2	Sox	Sox-9
Clup	XP_038381543.1	Fox	Fox-B2	Csim	XP_004432762.1	Sox	Sox-9
Emax	XP_049751166.1	Fox	Fox-B2	Ptig	XP_042822833.1	Sox	Sox-9
Hsap	NP_001013757.1	Fox	Fox-B2	Equa	XP_046530969.1	Sox	Sox-9
Rfer	XP_032978089.1	Fox	Fox-B2	Emax	XP_049717106.1	Sox	Sox-9
Hamp	XP_057580851.1	Fox	Fox-B2	Oana	XP_001506094.2	Sox	Sox-9
Oafe	XP_007936768.1	Fox	Fox-B2	Rfer	XP_032945891.1	Sox	Sox-9
Ptig	XP_042819403.1	Fox	Fox-B2	Ttru	XP_033703168.1	Sox	Sox-9
Oana	XP_001518148.2	Fox	Fox-B1	Amel	XP_019660087.2	Sox	Sox-9
Clup	XP_038436513.1	Fox	Fox-B1	Mang	XP_045743969.1	Sox	Sox-9
Shar	XP_003755656.2	Fox	Fox-B1	Pafr	XP_047614296.1	Sox	Sox-9
Ttru	XP_033708504.1	Fox	Fox-B1	Scar	XP_047398689.1	Sox	Sox-9
Mjav	XP_017523883.1	Fox	Fox-B1	Bbub	XP_025135172.2	Sox	Sox-9
Casi	XP_006831648.1	Fox	Fox-B1	Shar	XP_003768593.1	Sox	Sox-9
Rfer	XP_032964940.1	Fox	Fox-B1	Hsap	NP_000337.1	Sox	Sox-9
Tman	XP_004374676.1	Fox	Fox-B1	Mmus	NP_035578.3	Sox	Sox-9
Hsap	NP_036314.2	Fox	Fox-B1	Oafe	XP_007957879.1	Sox	Sox-9
Amel	XP_034516791.1	Fox	Fox-B1	Cdid	XP_037664998.1	Sox	Sox-9
Pgig	XP_039720931.1	Fox	Fox-B1	Lcat	XP_045425568.1	Sox	Sox-9
Cdro	XP_031308829.1	Fox	Fox-B1	Pgig	XP_039715158.1	Sox	Sox-9

Tab. S8 continued from previous page

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Opri	XP_004578087.1	Fox	Fox-B1	Ggal	XP_015139949.1	Sox	Sox-10
Mang	XP_045755904.1	Fox	Fox-B1	Mmus	NP_035567.1	Sox	Sox-10
Cimi	XP_017387405.1	Fox	Fox-B1	Csim	XP_004437902.1	Sox	Sox-10
Dnov	XP_012376939.1	Fox	Fox-B1	Hsap	NP_008872.1	Sox	Sox-10
Equa	XP_046508174.1	Fox	Fox-B1	Mjav	XP_036862891.1	Sox	Sox-10
Mdom	XP_001365592.1	Fox	Fox-B1	Oafe	XP_007939944.1	Sox	Sox-10
Scar	XP_047396474.1	Fox	Fox-B1	Opri	XP_004589571.1	Sox	Sox-10
Emax	XP_049708649.1	Fox	Fox-B1	Scar	XP_047407584.1	Sox	Sox-10
Bmus	XP_036700853.1	Fox	Fox-B1	Mang	XP_045728505.1	Sox	Sox-10
Oafe	XP_007956505.1	Fox	Fox-B1	Cdro	XP_031319106.1	Sox	Sox-10
Bbub	XP_006043154.1	Fox	Fox-B1	Shar	XP_031797111.1	Sox	Sox-10
Ggal	XP_004943811.1	Fox	Fox-B1	Cimi	XP_017364446.1	Sox	Sox-10
Cpor	XP_013000887.1	Fox	Fox-B1	Ttru	XP_033721970.1	Sox	Sox-10
Ptig	XP_042843621.1	Fox	Fox-B1	Dnov	XP_004466267.1	Sox	Sox-10
Mmus	NP_071773.2	Fox	Fox-B1	Ptig	XP_042849176.1	Sox	Sox-10
Csim	XP_004421699.1	Fox	Fox-B1	Tman	XP_004373905.1	Sox	Sox-10
Hamp	XP_057575814.1	Fox	Fox-B1	Cpor	XP_003470533.2	Sox	Sox-10
Lcat	XP_045411390.1	Fox	Fox-B1	Hamp	XP_057599512.1	Sox	Sox-10
Cdid	XP_037691321.1	Fox	Fox-B1	Oana	XP_028934366.1	Sox	Sox-10
Drot	XP_024424310.1	Fox	Fox-B1	Amel	XP_034500270.1	Sox	Sox-10
Pafr	XP_047625395.1	Fox	Fox-B1	Casi	XP_006865257.1	Sox	Sox-10
Oana	XP_028925820.1	Fox	Hnf-3b/Fox-A2	Bbub	XP_006071428.1	Sox	Sox-10
Mdom	XP_001382097.1	Fox	Hnf-3b/Fox-A2	Cdid	XP_037702433.1	Sox	Sox-10
Clup	XP_038427348.1	Fox	Hnf-3b/Fox-A2	Lcat	XP_045408856.1	Sox	Sox-10
Amel	XP_002924170.1	Fox	Hnf-3b/Fox-A2	Equa	XP_046502255.1	Sox	Sox-10
Cimi	XP_017396970.1	Fox	Hnf-3b/Fox-A2	Pgig	XP_039733644.1	Sox	Sox-10
Mjav	XP_036878344.1	Fox	Hnf-3b/Fox-A2	Clup	XP_038406650.1	Sox	Sox-10
Ttru	XP_019783169.2	Fox	Hnf-3b/Fox-A2	Rfer	XP_032973012.1	Sox	Sox-10
Mang	XP_045739756.1	Fox	Hnf-3b/Fox-A2	Emax	XP_049740365.1	Sox	Sox-10
Shar	XP_003758186.1	Fox	Hnf-3b/Fox-A2	Drot	XP_024434928.2	Sox	Sox-10
Emax	XP_049725634.1	Fox	Hnf-3b/Fox-A2	Pafr	XP_047642247.1	Sox	Sox-10
Scar	XP_047397766.1	Fox	Hnf-3b/Fox-A2	Mdom	XP_001381534.3	Sox	Sox-10
Pgig	XP_039730271.1	Fox	Hnf-3b/Fox-A2	Bmus	XP_036720803.1	Sox	Sox-10
Equa	XP_046535478.1	Fox	Hnf-3b/Fox-A2	Hsap	XP_011519134.2	Sox	Sox-5
Hsap	NP_068556.2	Fox	Hnf-3b/Fox-A2	Lcat	XP_045410158.1	Sox	Sox-5
Mmus	NP_001277994.1	Fox	Hnf-3b/Fox-A2	Oana	XP_028925314.1	Sox	Sox-13
Lcat	XP_045385044.1	Fox	Hnf-3b/Fox-A2	Pafr	XP_047607942.1	Sox	Sox-13
Drot	XP_045050691.1	Fox	Hnf-3b/Fox-A2	Shar	XP_003767655.1	Sox	Sox-13
Cdid	XP_037668003.1	Fox	Hnf-3b/Fox-A2	Ggal	XP_015154614.2	Sox	Sox-13
Tman	XP_004376477.1	Fox	Hnf-3b/Fox-A2	Mdom	XP_056671743.1	Sox	Sox-13
Casi	XP_006860717.1	Fox	Hnf-3b/Fox-A2	Ttru	XP_033708141.1	Sox	Sox-13
Oafe	XP_007948556.1	Fox	Hnf-3b/Fox-A2	Cpor	XP_013013443.1	Sox	Sox-13
Pafr	XP_047627543.1	Fox	Hnf-3b/Fox-A2	Bmus	XP_036684089.1	Sox	Sox-13
Hamp	XP_057558835.1	Fox	Hnf-3b/Fox-A2	Mang	XP_054363913.1	Sox	Sox-13
Dnov	XP_023439993.2	Fox	Hnf-3b/Fox-A2	Tman	XP_023586088.1	Sox	Sox-13
Bmus	XP_036681859.1	Fox	Hnf-3b/Fox-A2	Hamp	XP_057585117.1	Sox	Sox-13
Rfer	XP_032950275.1	Fox	Hnf-3b/Fox-A2	Cdro	XP_031294667.1	Sox	Sox-13
Cpor	XP_003476486.1	Fox	Hnf-3b/Fox-A2	Drot	XP_024430737.2	Sox	Sox-13
Opri	XP_004585707.2	Fox	Hnf-3b/Fox-A2	Dnov	XP_058130986.1	Sox	Sox-13
Bbub	XP_044783908.1	Fox	Hnf-3b/Fox-A2	Mjav	XP_017506787.2	Sox	Sox-13
Cdro	XP_031290193.1	Fox	Hnf-3b/Fox-A2	Amel	XP_034522993.1	Sox	Sox-13
Csim	XP_004442004.1	Fox	Hnf-3b/Fox-A2	Mmus	XP_011246247.1	Sox	Sox-13
Ptig	XP_042835681.1	Fox	Hnf-3b/Fox-A2	Scar	XP_047376454.1	Sox	Sox-13
Ggal	XP_046794381.1	Fox	Hnf-3b/Fox-A2	Ptig	XP_007086583.2	Sox	Sox-13
Oana	XP_028902569.1	Fox	Fox-E3	Clup	XP_038441905.1	Sox	Sox-13
Emax	XP_049731498.1	Fox	Fox-E3	Oafe	XP_007954219.1	Sox	Sox-13
Hsap	NP_036318.1	Fox	Fox-E3	Opri	XP_05852035.1	Sox	Sox-13
Mang	XP_045751381.1	Fox	Fox-E3	Emax	XP_049714298.1	Sox	Sox-13
Hamp	XP_057562286.1	Fox	Fox-E3	Equa	XP_046537707.1	Sox	Sox-13
Drot	XP_053777352.1	Fox	Fox-E3	Casi	XP_006834281.1	Sox	Sox-13
Ttru	XP_033709174.1	Fox	Fox-E3	Hsap	NP_005677.2	Sox	Sox-13
Pafr	XP_047647470.1	Fox	Fox-E3	Cimi	XP_037593260.1	Sox	Sox-13
Ptig	XP_042852491.1	Fox	Fox-E3	Bbub	XP_025133526.2	Sox	Sox-13
Clup	XP_038413565.1	Fox	Fox-E3	Cdid	XP_037680910.1	Sox	Sox-13
Rfer	XP_032971209.1	Fox	Fox-E3	Lcat	XP_045391966.1	Sox	Sox-13
Bmus	XP_036690899.1	Fox	Fox-E3	Csim	XP_004425199.1	Sox	Sox-13
Pgig	XP_039712722.1	Fox	Fox-E3	Rfer	XP_032948937.1	Sox	Sox-13
Bbub	XP_025144571.1	Fox	Fox-E3	Pgig	XP_039737775.1	Sox	Sox-13
Cdro	XP_031321430.1	Fox	Fox-E3	Ggal	XP_025006442.1	Sox	Sox-6
Tman	XP_023582802.1	Fox	Fox-E3	Emax	XP_049746477.1	Sox	Sox-6
Pgig	XP_039738800.1	Fox	Fox-E3	Ttru	XP_019804010.1	Sox	Sox-6
Mmus	NP_056573.1	Fox	Fox-E3	Hamp	XP_057604501.1	Sox	Sox-6
Mjav	XP_036877848.1	Fox	Fox-E3	Csim	XP_032419042.1	Sox	Sox-6
Casi	XP_006839917.1	Fox	Fox-E3	Rfer	XP_032975562.1	Sox	Sox-6
Dnov	XP_058160703.1	Fox	Fox-E3	Shar	XP_031797909.1	Sox	Sox-6
Lcat	XP_045401422.1	Fox	Fox-E3	Bbub	XP_045018884.1	Sox	Sox-6
Cimi	XP_017365578.1	Fox	Fox-E3	Dnov	XP_058161675.1	Sox	Sox-6
Cdid	XP_037672792.1	Fox	Fox-E3	Oana	XP_039767308.1	Sox	Sox-6
Opri	XP_004598546.1	Fox	Fox-E3	Hsap	NP_001139291.2	Sox	Sox-6
Scar	XP_047375686.1	Fox	Fox-E3	Bmus	XP_036717651.1	Sox	Sox-6
Oana	XP_001516678.2	Fox	Fox-D4	Mjav	XP_036882886.1	Sox	Sox-6
Pgig	XP_039744744.1	Fox	Fox-D4	Amel	XP_034501998.1	Sox	Sox-6
Hamp	XP_057576439.1	Fox	Fox-D4	Equa	XP_046493708.1	Sox	Sox-6
Pafr	XP_047625506.1	Fox	Fox-D4	Mdom	XP_007497087.2	Sox	Sox-6
Hsap	NP_001119806.1	Fox	Fox-D4	Cpor	XP_023419042.1	Sox	Sox-6
Emax	XP_049752498.1	Fox	Fox-D4	Pgig	XP_039707456.1	Sox	Sox-6
Cdid	XP_037706680.1	Fox	Fox-D4	Oafe	XP_007955001.1	Sox	Sox-6
Hsap	NP_954714.2	Fox	Fox-D4	Cdid	XP_037695265.1	Sox	Sox-6
Rfer	XP_032979406.1	Fox	Fox-D4	Casi	XP_006865846.1	Sox	Sox-6
Ttru	XP_033714036.1	Fox	Fox-D4	Cimi	XP_017381104.1	Sox	Sox-6
Ptig	XP_042820135.1	Fox	Fox-D4	Mmus	XP_036008726.1	Sox	Sox-6
Cdro	XP_010998361.2	Fox	Fox-D4	Ptig	XP_042815591.1	Sox	Sox-6
Mang	XP_045739597.1	Fox	Fox-D4	Pafr	XP_047632165.1	Sox	Sox-6
Scar	XP_047380997.1	Fox	Fox-D4	Tman	XP_023593283.1	Sox	Sox-6
Drot	XP_053785855.1	Fox	Fox-D4	Drot	XP_053780580.1	Sox	Sox-6
Hsap	NP_001078945.1	Fox	Fox-D4	Mang	XP_054368007.1	Sox	Sox-6
Amel	XP_034503132.1	Fox	Fox-D4	Csim	XP_014642191.1	Sox	Sox-6
Oafe	XP_007954771.1	Fox	Fox-D4	Scar	XP_047372506.1	Sox	Sox-6
Equa	XP_046519849.1	Fox	Fox-D4	Lcat	XP_045413549.1	Sox	Sox-6
Bbub	XP_025137372.3	Fox	Fox-D4	Cdro	XP_010975526.2	Sox	Sox-6
Dnov	XP_023444807.2	Fox	Fox-D4	Clup	XP_038425054.1	Sox	Sox-6
Bmus	XP_036712053.1	Fox	Fox-D4	Emax	XP_049725376.1	Sox	Sox-18

Tab. S8 continued from previous page

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Hsap	NP_036316.1	Fox	Fox-D4	Casi	XP_006873390.1	Sox	-
Opri	XP_004591937.2	Fox	Fox-D4	Oafe	XP_007952883.1	Sox	-
Cimi	XP_017398902.1	Fox	Fox-D4	Mmus	NP_035577.1	Sox	Sox-8
Mdom	XP_001373972.1	Fox	Fox-D4	Opri	XP_004596857.2	Sox	-
Mmus	NP_032048.1	Fox	Fox-D4	Cpor	XP_003478479.1	Sox	-
Hsap	NP_997188.2	Fox	Fox-D4	Oana	XP_028913325.1	Sox	Sox-8
Hsap	NP_954586.4	Fox	Fox-D4	Ggal	NP_990062.2	Sox	-
Csim	XP_014650927.1	Fox	Fox-D4	Cpor	XP_023421073.1	Sox	-
Cpor	XP_013010584.1	Fox	Fox-D4	Cimi	XP_017378281.2	Sox	-
Mjav	XP_036847234.1	Fox	Fox-D4	Cdid	XP_037702077.1	Sox	-
Casi	XP_006863895.1	Fox	Fox-D4	Shar	XP_031794321.1	Sox	-
Mang	XP_045757341.1	Fox	-	Mdom	XP_016280870.1	Sox	-
Clup	XP_038401447.1	Fox	-	Ggal	XP_040553151.1	Sox	-
Amel	XP_019655962.1	Fox	-	Oana	XP_028913966.1	Sox	Sox-5
Clup	XP_038439326.1	Fox	-	Lcat	XP_045422920.1	Sox	-
Ggal	NP_001382914.1	Fox	Fox-C1				

Supplementary Table S9 – Complete set of DSFGs in *Drosophila*. For each gene, the species ID (Sp. ID) as in Tab. S5, the accession number (Gene ID), and the Possvm-based annotation are indicated.

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Agam	XP_061505148.1	Dmrt	Dsx	Dbus	XP_017845192.1	Fox	Slp-1
Dere	XP_026836830.1	Dmrt	Dsx	Dgri	XP_001988485.1	Fox	Slp-1
Dkik	XP_041630485.1	Dmrt	Dsx	Dhyd	XP_023179855.2	Fox	Slp-1
Dmir	XP_017142875.1	Dmrt	Dsx	Dari	XP_017860496.1	Fox	Slp-1
Dgri	XP_043071113.1	Dmrt	Dsx	Dana	XP_001961572.2	Fox	Slp-1
Dsec	XP_002038750.1	Dmrt	Dsx	Dmir	XP_017155124.1	Fox	Slp-1
Dser	XP_020809854.1	Dmrt	Dsx	Dbip	XP_017105354.1	Fox	Slp-1
Dele	XP_017119779.1	Dmrt	Dsx	Dmel	NP_476730.1	Fox	Slp-1
Dalb	XP_034117252.2	Dmrt	Dsx	Dere	XP_001968625.1	Fox	Slp-1
Dwil	XP_023035845.1	Dmrt	Dsx	Dsec	XP_002037770.1	Fox	Slp-1
Dhyd	XP_023178918.2	Dmrt	Dsx	Dkik	XP_017021730.1	Fox	Slp-1
Dmel	NP_001262353.1	Dmrt	Dsx	Dpse	XP_001356670.4	Fox	Slp-1
Dpse	XP_033235910.1	Dmrt	Dsx	Dwil	XP_002065500.1	Fox	Slp-1
Dari	XP_017874634.1	Dmrt	Dsx	Dser	XP_020800881.1	Fox	Slp-1
Dbus	XP_017847641.1	Dmrt	Dsx	Dsuz	XP_016927184.1	Fox	Slp-1
Dbib	XP_017088683.2	Dmrt	Dsx	Agam	XP_061514780.1	Fox	Slp-2
Dsuz	XP_036675224.1	Dmrt	Dsx	Dalb	XP_034100740.1	Fox	Slp-2
Dsuz	XP_036667275.1	Dmrt	Dsx	Dele	XP_017110569.1	Fox	Slp-2
Dana	XP_014766033.1	Dmrt	Dsx	Dere	XP_001968626.1	Fox	Slp-2
Dbus	XP_017844894.1	Dmrt	Dmrt-99B	Dbus	XP_017845289.1	Fox	Slp-2
Dmel	NP_524549.1	Dmrt	Dmrt-99B	Dsec	XP_002037771.1	Fox	Slp-2
Dere	XP_001981330.1	Dmrt	Dmrt-99B	Dpse	XP_001356669.3	Fox	Slp-2
Dwil	XP_023034529.1	Dmrt	Dmrt-99B	Dmir	XP_017151517.1	Fox	Slp-2
Dkik	XP_017021883.1	Dmrt	Dmrt-99B	Dkik	XP_017021724.1	Fox	Slp-2
Dser	XP_020811324.1	Dmrt	Dmrt-99B	Dana	XP_001961573.1	Fox	Slp-2
Dalb	XP_051862400.1	Dmrt	Dmrt-99B	Dari	XP_017860500.1	Fox	Slp-2
Dsuz	XP_036674364.1	Dmrt	Dmrt-99B	Dser	XP_020800889.1	Fox	Slp-2
Dsec	XP_002037224.1	Dmrt	Dmrt-99B	Dbib	XP_017105682.2	Fox	Slp-2
Dele	XP_017131043.1	Dmrt	Dmrt-99B	Dhyd	XP_023179875.2	Fox	Slp-2
Dbib	XP_017099960.2	Dmrt	Dmrt-99B	Dsuz	XP_016926388.1	Fox	Slp-2
Dmir	XP_017145289.1	Dmrt	Dmrt-99B	Dwil	XP_023031666.1	Fox	Slp-2
Dpse	XP_001357766.3	Dmrt	Dmrt-99B	Dgri	XP_001988487.3	Fox	Slp-2
Dhyd	XP_023160825.2	Dmrt	Dmrt-99B	Dmel	NP_476834.1	Fox	Slp-2
Dari	XP_017869153.1	Dmrt	Dmrt-99B	Dgri	XP_317309.5	Fox	Fd-3/Fd-59A
Dgri	XP_001996117.1	Dmrt	Dmrt-99B	Dbib	XP_017090886.2	Fox	Fd-3/Fd-59A
Dhyd	XP_023160826.2	Dmrt	Dmrt-99B	Dpse	XP_001361889.1	Fox	Fd-3/Fd-59A
Dana	XP_001964762.1	Dmrt	Dmrt-99B	Dmel	NP_523814.1	Fox	Fd-3/Fd-59A
Agam	XP_061501728.1	Dmrt	Dmrt-93B	Dsuz	XP_016927872.1	Fox	Fd-3/Fd-59A
Dalb	XP_034117959.2	Dmrt	Dmrt-93B	Dele	XP_017132051.1	Fox	Fd-3/Fd-59A
Dpse	XP_001360059.2	Dmrt	Dmrt-93B	Dser	XP_020803123.1	Fox	Fd-3/Fd-59A
Dkik	XP_017036725.1	Dmrt	Dmrt-93B	Dwil	XP_02061322.3	Fox	Fd-3/Fd-59A
Dbib	XP_017102685.2	Dmrt	Dmrt-93B	Dgri	XP_001987215.1	Fox	Fd-3/Fd-59A
Dbus	XP_017844858.1	Dmrt	Dmrt-93B	Dana	XP_001960803.1	Fox	Fd-3/Fd-59A
Dsuz	XP_036672900.1	Dmrt	Dmrt-93B	Dmir	XP_033248262.1	Fox	Fd-3/Fd-59A
Dser	XP_020817775.1	Dmrt	Dmrt-93B	Dalb	XP_034105309.1	Fox	Fd-3/Fd-59A
Dari	XP_017874225.1	Dmrt	Dmrt-93B	Dhyd	XP_030021270.1	Fox	Fd-3/Fd-59A
Dgri	XP_001990371.1	Dmrt	Dmrt-93B	Dmir	XP_017150782.2	Fox	Fd-3/Fd-59A
Dwil	XP_002073560.1	Dmrt	Dmrt-93B	Dkik	XP_017024427.1	Fox	Fd-3/Fd-59A
Dmel	NP_524428.1	Dmrt	Dmrt-93B	Dari	XP_017865679.1	Fox	Fd-3/Fd-59A
Dmir	XP_017140144.1	Dmrt	Dmrt-93B	Dsec	XP_002039987.1	Fox	Fd-3/Fd-59A
Dsec	XP_002044276.1	Dmrt	Dmrt-93B	Dere	XP_001976310.1	Fox	Fd-3/Fd-59A
Dele	XP_017118537.1	Dmrt	Dmrt-93B	Dbus	XP_017836437.1	Fox	Fd-3/Fd-59A
Dana	XP_001954937.1	Dmrt	Dmrt-93B	Agam	XP_315933.4	Fox	Crocodile
Dere	XP_001979344.2	Dmrt	Dmrt-93B	Dalb	XP_034109751.1	Fox	Crocodile
Dmel	NP_511146.2	Dmrt	Dmrt-11E	Dele	XP_017121437.1	Fox	Crocodile
Dsec	XP_002042926.2	Dmrt	Dmrt-11E	Dmel	NP_524202.1	Fox	Crocodile
Dwil	XP_002075263.1	Dmrt	Dmrt-11E	Dsec	XP_002040809.1	Fox	Crocodile
Dpse	XP_001355530.3	Dmrt	Dmrt-11E	Dsuz	XP_016933536.1	Fox	Crocodile
Dbib	XP_017089150.2	Dmrt	Dmrt-11E	Dari	XP_017864431.1	Fox	Crocodile
Dser	XP_020808350.1	Dmrt	Dmrt-11E	Dmir	XP_017139058.1	Fox	Crocodile
Dmir	XP_017136076.1	Dmrt	Dmrt-11E	Dhyd	XP_023173236.2	Fox	Crocodile
Dana	XP_002309371.1	Dmrt	Dmrt-11E	Dbib	XP_017095284.1	Fox	Crocodile
Dere	XP_015010652.2	Dmrt	Dmrt-11E	Dere	XP_001973629.1	Fox	Crocodile
Dbus	XP_017850127.1	Dmrt	Dmrt-11E	Dwil	XP_002061802.1	Fox	Crocodile
Dsuz	XP_016924080.2	Dmrt	Dmrt-11E	Dser	XP_020817581.1	Fox	Crocodile
Dalb	XP_034099986.1	Dmrt	Dmrt-11E	Dbus	XP_017841359.1	Fox	Crocodile
Dgri	XP_043071903.1	Dmrt	Dmrt-11E	Dpse	XP_001354188.2	Fox	Crocodile
Dele	XP_017112743.2	Dmrt	Dmrt-11E	Dkik	XP_017029613.1	Fox	Crocodile
Dhyd	XP_023180022.1	Dmrt	Dmrt-11E	Dana	XP_001958355.1	Fox	Crocodile
Dkik	XP_017034619.2	Dmrt	Dmrt-11E	Dgri	XP_001983802.1	Fox	Crocodile
Dari	XP_017869421.1	Dmrt	Dmrt-11E	Dere	XP_001973629.1	Fox	Crocodile
Agam	XP_310668.5	Dmrt	-	Dmir	XP_017144084.1	Fox	Fkh
				Dari	XP_017873677.1	Fox	Fkh

Tab. S9 continued from previous page

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Dkik	XP_017022631.1	Fox	Fox-3F	Dpse	XP_033238940.1	Fox	Fkh
Dsec	XP_002037072.2	Fox	Fox-3F	Dwil	XP_002070503.2	Fox	Fkh
Dser	XP_020800708.1	Fox	Fox-3F	Dsuz	XP_036673542.1	Fox	Fkh
Dbip	XP_017087408.2	Fox	Fox-3F	Dalb	XP_051862733.1	Fox	Fkh
Dere	XP_026837609.1	Fox	Fox-3F	Dbip	XP_017092849.2	Fox	Fkh
Dana	XP_014759802.1	Fox	Fox-3F	Dbus	XP_017849701.2	Fox	Fkh
Dpse	XP_032384871.1	Fox	Fox-3F	Dsec	XP_002043027.2	Fox	Fkh
Dmir	XP_017145653.1	Fox	Fox-3F	Dgri	XP_001989593.3	Fox	Fkh
Dmel	NP_001356931.1	Fox	Fox-3F	Dana	XP_001955055.2	Fox	Fkh
Dele	XP_017112308.1	Fox	Fox-3F	Dkik	XP_017037933.1	Fox	Fkh
Dsuz	XP_016942113.1	Fox	Fox-3F	Dser	XP_02081120.1	Fox	Fkh
Dwil	XP_046868406.1	Fox	-	Dere	XP_001981453.3	Fox	Fkh
Dbus	XP_033150245.1	Fox	-	Agam	XP_061497286.1	Fox	Fkh
Dhyd	XP_030079965.1	Fox	-	Dmel	NP_001263038.1	Fox	Fkh
Dari	XP_017869433.1	Fox	-	Dele	XP_017131720.1	Fox	Fkh
Dgri	XP_043071916.1	Fox	-	Dhyd	XP_023165610.2	Fox	Fkh
Dalb	XP_051860438.1	Fox	-	Dalb	XP_034114947.1	Fox	Fd-5/Fd-96Cb
Agam	XP_061503465.1	Fox	Ches-1	Dele	XP_017123797.1	Fox	Fd-5/Fd-96Cb
Dbip	XP_017091473.2	Fox	Ches-1	Dgri	XP_001990839.1	Fox	Fd-5/Fd-96Cb
Dkik	XP_017022654.1	Fox	Ches-1	Dana	XP_001953691.1	Fox	Fd-5/Fd-96Cb
Dalb	XP_034097387.1	Fox	Ches-1	Dere	XP_001981873.2	Fox	Fd-5/Fd-96Cb
Dhyd	XP_023174414.2	Fox	Ches-1	Dkik	XP_017020777.1	Fox	Fd-5/Fd-96Cb
Dana	XP_032308510.1	Fox	Ches-1	Dwil	XP_023036287.1	Fox	Fd-5/Fd-96Cb
Dwil	XP_023030939.1	Fox	Ches-1	Dbip	XP_017103917.2	Fox	Fd-5/Fd-96Cb
Dser	XP_020805864.1	Fox	Ches-1	Dser	XP_020817575.1	Fox	Fd-5/Fd-96Cb
Dgri	XP_032593899.1	Fox	Ches-1	Dsec	XP_002043869.2	Fox	Fd-5/Fd-96Cb
Dbus	XP_017852827.2	Fox	Ches-1	Dsuz	XP_016923664.2	Fox	Fd-5/Fd-96Cb
Dari	XP_017869569.1	Fox	Ches-1	Dmel	NP_524496.1	Fox	Fd-5/Fd-96Cb
Dpse	XP_033241472.1	Fox	Ches-1	Agam	XP_061497721.1	Fox	Fd-4/Fd-96Ca
Dmir	XP_033252440.1	Fox	Ches-1	Dana	XP_001953690.1	Fox	Fd-4/Fd-96Ca
Agam	XP_061505879.1	Fox	Jumeau	Dmel	NP_001287516.1	Fox	Fd-4/Fd-96Ca
Dalb	XP_034114927.1	Fox	Jumeau	Dsuz	XP_016923665.2	Fox	Fd-4/Fd-96Ca
Dgri	XP_001990048.1	Fox	Jumeau	Dpse	XP_033232491.1	Fox	Fd-4/Fd-96Ca
Dmir	XP_033244968.1	Fox	Jumeau	Dser	XP_020817866.1	Fox	Fd-4/Fd-96Ca
Dsec	XP_002031751.2	Fox	Jumeau	Dele	XP_017123798.1	Fox	Fd-4/Fd-96Ca
Dser	XP_020814051.1	Fox	Jumeau	Dkik	XP_017020776.1	Fox	Fd-4/Fd-96Ca
Dari	XP_017867184.1	Fox	Jumeau	Dwil	XP_020207297.1	Fox	Fd-4/Fd-96Ca
Dkik	XP_017026689.1	Fox	Jumeau	Dalb	XP_034114767.1	Fox	Fd-4/Fd-96Ca
Dana	XP_001952947.1	Fox	Jumeau	Dgri	XP_001990840.1	Fox	Fd-4/Fd-96Ca
Dmir	XP_033244970.1	Fox	Jumeau	Dbus	XP_017846227.1	Fox	Fd-4/Fd-96Ca
Dere	XP_001980679.1	Fox	Jumeau	Dpse	XP_017141748.1	Fox	Fd-4/Fd-96Ca
Dele	XP_017119152.1	Fox	Jumeau	Dhdy	XP_023172915.1	Fox	Fd-4/Fd-96Ca
Dsuz	XP_016944421.1	Fox	Jumeau	Dari	XP_017856594.1	Fox	Fd-4/Fd-96Ca
Dhyd	XP_023174343.2	Fox	Jumeau	Dsec	XP_002043867.1	Fox	Fd-4/Fd-96Ca
Dwil	XP_023031582.2	Fox	Jumeau	Dere	XP_001981874.1	Fox	Fd-4/Fd-96Ca
Dpse	XP_033233400.1	Fox	Jumeau	Dmir	XP_017141748.1	Fox	Fd-4/Fd-96Ca
Dpse	XP_015038643.2	Fox	Jumeau	Agam	XP_061507474.1	Fox	-
Dmir	XP_033244969.1	Fox	Jumeau	Agam	XP_061497423.1	Fox	-
Dmel	NP_524302.1	Fox	Jumeau	Agam	XP_312480.5	Fox	-
Dbip	XP_017100743.2	Fox	Jumeau	Dana	XP_044571024.1	Sox	Sox-21B
Dbus	XP_017845570.8	Fox	Jumeau	Dari	XP_017862175.1	Sox	Sox-21B
Dbip	XP_043069566.1	Fox	Hcm-1	Dhyd	XP_030080409.1	Sox	Sox-21B
Dsec	XP_002044492.1	Fox	Hcm-1	Dbus	XP_033149417.1	Sox	Sox-21B
Dere	XP_026838997.1	Fox	Hcm-1	Dere	XP_001972712.1	Sox	Sox-21B
Dhyd	XP_023165574.2	Fox	Hcm-1	Dmir	XP_033243279.1	Sox	Sox-21B
Dele	XP_041564577.1	Fox	Hcm-1	Dmel	NP_001261829.1	Sox	Sox-21B
Dsuz	XP_036678464.1	Fox	Hcm-1	Dele	XP_017128640.1	Sox	Sox-21B
Dalb	XP_051862243.1	Fox	Hcm-1	Dpse	XP_002134931.3	Sox	Sox-21B
Dana	XP_02311767.1	Fox	Hcm-1	Dwip	XP_023034010.1	Sox	Sox-21B
Dpse	XP_033237266.1	Fox	Hcm-1	Dsec	XP_002030465.1	Sox	Sox-21B
Dmel	NP_726538.1	Fox	Hcm-1	Dalb	XP_034107607.1	Sox	Sox-21B
Dmir	XP_017155587.1	Fox	Hcm-1	Dpse	XP_017090122.2	Sox	Sox-21B
Dser	XP_020809944.1	Fox	Hcm-1	Agam	XP_061509694.1	Sox	Sox-21B
Dbus	XP_017853536.1	Fox	Hcm-1	Dkik	XP_017034105.1	Sox	Sox-21B
Dwip	XP_023035694.1	Fox	Hcm-1	Dgri	XP_032598957.1	Sox	Sox-21B
Dkik	XP_017037372.1	Fox	Hcm-1	Dser	XP_020816310.1	Sox	Sox-21B
Dwil	XP_002072563.1	Fox	Fd-102C	Dgri	XP_032590737.1	Sox	Sox-14
Dhyd	XP_023163080.1	Fox	Fd-102C	Dhdy	XP_023171730.2	Sox	Sox-14
Dbip	XP_017098296.2	Fox	Fd-102C	Dbus	XP_017836915.1	Sox	Sox-14
Dmel	NP_651951.1	Fox	Fd-102C	Dpse	XP_001360869.4	Sox	Sox-14
Dsuz	XP_036671526.1	Fox	Fd-102C	Dkik	XP_017018688.1	Sox	Sox-14
Dele	XP_017126055.1	Fox	Fd-102C	Dpse	XP_017090231.2	Sox	Sox-14
Dbus	XP_017853233.1	Fox	Fd-102C	Dser	XP_020817997.1	Sox	Sox-14
Dsec	XP_002043702.1	Fox	Fd-102C	Dmir	XP_033246935.1	Sox	Sox-14
Agam	XP_061505343.1	Fox	Fd-102C	Dsec	XP_002040195.2	Sox	Sox-14
Dere	XP_001982706.1	Fox	Fd-102C	Dmel	NP_001286801.1	Sox	Sox-14
Dmir	XP_017155765.1	Fox	Fd-102C	Dmir	XP_017149652.2	Sox	Sox-14
Dana	XP_02308054.1	Fox	Fd-102C	Dsuz	XP_016926392.1	Sox	Sox-14
Dalb	XP_034112224.1	Fox	Fd-102C	Dari	XP_017868666.1	Sox	Sox-14
Dpse	XP_015044407.1	Fox	Fd-102C	Dalb	XP_034106594.1	Sox	Sox-14
Dkik	XP_041632589.1	Fox	Fd-102C	Dere	XP_001976516.1	Sox	Sox-14
Dgri	XP_043071810.1	Fox	Fd-102C	Dwip	XP_002061086.2	Sox	Sox-14
Dser	XP_020809941.1	Fox	Fd-102C	Agam	XP_061506760.1	Sox	Sox-14
Dari	XP_017869379.1	Fox	Fd-102C	Dele	XP_017125359.1	Sox	Sox-14
Agam	XP_061502996.1	Fox	Fox-P	Dana	XP_001959457.2	Sox	Sox-14
Dhyd	XP_023172914.1	Fox	Fox-P	Dpse	XP_001353547.1	Sox	Dichaete
Dbus	XP_033148941.1	Fox	Fox-P	Dele	XP_017128756.1	Sox	Dichaete
Dbip	XP_043067838.1	Fox	Fox-P	Dmir	XP_017138810.1	Sox	Dichaete
Dgri	XP_032593352.2	Fox	Fox-P	Dalb	XP_034104310.1	Sox	Dichaete
Dana	XP_044571198.1	Fox	Fox-P	Dsec	XP_002030467.1	Sox	Dichaete
Dalb	XP_051862673.1	Fox	Fox-P	Dgri	XP_001984254.1	Sox	Dichaete
Dmir	XP_033244694.1	Fox	Fox-P	Dana	XP_001956432.1	Sox	Dichaete
Dmel	NP_001247011.1	Fox	Fox-P	Dmel	NP_524066.1	Sox	Dichaete
Dere	XP_015009985.2	Fox	Fox-P	Dere	XP_001972713.1	Sox	Dichaete
Dele	XP_017117570.1	Fox	Fox-P	Dpse	XP_017089891.1	Sox	Dichaete
Dsuz	XP_016930987.1	Fox	Fox-P	Agam	XP_061514536.1	Sox	Dichaete
Dpse	XP_032341607.1	Fox	Fox-P	Dbus	XP_017841081.1	Sox	Dichaete
Dwil	XP_046865977.1	Fox	Fox-P	Dkik	XP_017034102.1	Sox	Dichaete
Dsec	XP_032577368.1	Fox	Fox-P	Dwil	XP_02061712.1	Sox	Dichaete
Dser	XP_020813734.1	Fox	Fox-P	Dari	XP_017864416.1	Sox	Dichaete
Dwil	XP_002072142.2	Fox	fox-O	Dhyd	XP_023173062.1	Sox	Dichaete
Agam	XP_061497073.1	Fox	fox-O	Dser	XP_020815889.1	Sox	Dichaete

Tab. S9 continued from previous page

Species ID	Gene ID	Group	Annotation	Species ID	Gene ID	Group	Annotation
Dari	XP_017859446.1	Fox	fox-O	Dsuz	XP_016933433.1	Sox	Dichaete
Dsuz	XP_036674215.1	Fox	fox-O	Dere	XP_001975666.1	Sox	Sox-15
Dsuz	XP_036672729.1	Fox	fox-O	Dele	XP_017133202.1	Sox	Sox-15
Dgri	XP_032595778.1	Fox	fox-O	Dbus	XP_017835743.2	Sox	Sox-15
Dbib	XP_017091959.2	Fox	fox-O	Dwil	XP_023030517.1	Sox	Sox-15
Dmir	XP_017143568.1	Fox	fox-O	Dalb	XP_051860622.1	Sox	Sox-15
Dalb	XP_034113157.1	Fox	fox-O	Dana	XP_001959479.1	Sox	Sox-15
Dsec	XP_032576567.1	Fox	fox-O	Dbib	XP_017090318.2	Sox	Sox-15
Dhyd	XP_023171342.1	Fox	fox-O	Dari	XP_017867550.1	Sox	Sox-15
Dbus	XP_033149836.1	Fox	fox-O	Dmir	XP_017149731.1	Sox	Sox-15
Dere	XP_026839669.1	Fox	fox-O	Dsec	XP_002033808.1	Sox	Sox-15
Dkik	XP_017026758.1	Fox	fox-O	Agam	XP_061509362.1	Sox	Sox-15
Dser	XP_020817293.1	Fox	fox-O	Dmel	NP_523739.2	Sox	Sox-15
Dpse	XP_03236382.1	Fox	fox-O	Dser	XP_020802427.1	Sox	Sox-15
Dele	XP_041565035.1	Fox	fox-O	Dhyd	XP_030081376.1	Sox	Sox-15
Dana	XP_032311428.1	Fox	fox-O	Dsuz	XP_016927906.1	Sox	Sox-15
Dmel	NP_650330.3	Fox	fox-O	Dpse	XP_001361762.2	Sox	Sox-15
Agam	XP_001688749.2	Fox	Binioi	Dkik	XP_017019703.1	Sox	Sox-15
Dmel	NP_523950.2	Fox	Binioi	Dgri	XP_001987303.1	Sox	Sox-15
Dalb	XP_034105454.1	Fox	Binioi	Dsec	XP_002030464.1	Sox	Sox-21A
Dser	XP_020817612.1	Fox	Binioi	Dgri	XP_001984256.1	Sox	Sox-21A
Dele	XP_017121429.1	Fox	Binioi	Dsuz	XP_016934639.1	Sox	Sox-21A
Dkik	XP_017017381.1	Fox	Binioi	Dmir	XP_017138458.1	Sox	Sox-21A
Dana	XP_001958356.2	Fox	Binioi	Dkik	XP_017034345.1	Sox	Sox-21A
Dari	XP_017864430.1	Fox	Binioi	Dbus	XP_017840298.2	Sox	Sox-21A
Dbus	XP_017841230.1	Fox	Binioi	Dari	XP_017862178.1	Sox	Sox-21A
Dwil	XP_02061803.1	Fox	Binioi	Dalb	XP_034108985.1	Sox	Sox-21A
Dgri	XP_001983803.1	Fox	Binioi	Dser	XP_020816311.1	Sox	Sox-21A
Dere	XP_001971514.2	Fox	Binioi	Agam	XP_061513404.1	Sox	Sox-21A
Dsuz	XP_036672243.1	Fox	Binioi	Dele	XP_017128154.1	Sox	Sox-21A
Dpse	XP_002134730.3	Fox	Binioi	Dana	XP_032310341.1	Sox	Sox-21A
Dhyd	XP_030081482.1	Fox	Binioi	Dhyd	XP_023173058.2	Sox	Sox-21A
Dbib	XP_017095285.2	Fox	Binioi	Dere	XP_001972711.1	Sox	Sox-21A
Dmir	XP_017139097.1	Fox	Binioi	Dbip	XP_043070009.1	Sox	Sox-21A
Dsec	XP_002035641.1	Fox	Binioi	Dwil	XP_023033998.1	Sox	Sox-21A
Agam	XP_061503012.1	Fox	Fd-2/Fox-L1	Dmel	NP_001246609.1	Sox	Sox-21A
Dmel	NP_001246609.1	Fox	Fd-2/Fox-L1	Dpse	XP_033239763.1	Sox	Sox-21A
Dhyd	XP_023178350.2	Fox	Fd-2/Fox-L1	Dwil	XP_002066518.2	Sox	Sox-N
Dbus	XP_017840363.1	Fox	Fd-2/Fox-L1	Dser	XP_020807900.1	Sox	Sox-N
Dkik	XP_017019394.1	Fox	Fd-2/Fox-L1	Dsuz	XP_036678170.1	Sox	Sox-N
Dser	XP_020807718.1	Fox	Fd-2/Fox-L1	Dbus	XP_033150665.1	Sox	Sox-N
Dari	XP_017863042.1	Fox	Fd-2/Fox-L1	Dsec	XP_002036270.2	Sox	Sox-N
Dsec	XP_002035260.1	Fox	Fd-2/Fox-L1	Dgri	XP_001993541.2	Sox	Sox-N
Dere	XP_001971815.2	Fox	Fd-2/Fox-L1	Dele	XP_017121213.1	Sox	Sox-N
Dalb	XP_051860220.1	Fox	Fd-2/Fox-L1	Dari	XP_017860249.1	Sox	Sox-N
Dana	XP_001956070.1	Fox	Fd-2/Fox-L1	Dalb	XP_051858933.1	Sox	Sox-N
Dsuz	XP_016932589.2	Fox	Fd-2/Fox-L1	Dkik	XP_017024634.1	Sox	Sox-N
Dele	XP_017126783.1	Fox	Fd-2/Fox-L1	Agam	XP_061516479.1	Sox	Sox-N
Dgri	XP_001983216.1	Fox	Fd-2/Fox-L1	Dmir	XP_017154597.1	Sox	Sox-N
Dpse	XP_001352427.1	Fox	Fd-2/Fox-L1	Dbip	XP_017099296.2	Sox	Sox-N
Dwil	XP_02062380.1	Fox	Fd-2/Fox-L1	Dere	XP_026834908.1	Sox	Sox-N
Dbip	XP_017098174.2	Fox	Fd-2/Fox-L1	Dmel	NP_001260269.1	Sox	Sox-N
Dmir	XP_017137846.1	Fox	Fd-2/Fox-L1	Dhyd	XP_023172210.2	Sox	Sox-N
Dere	XP_001977848.1	Fox	Fd-19B	Dpse	XP_001355808.4	Sox	Sox-N
Dhyd	XP_030079337.1	Fox	Fd-19B	Dana	XP_001962613.2	Sox	Sox-N
Dmel	NP_608369.1	Fox	Fd-19B	Dbus	XP_017845374.1	Sox	Sox-100B
Dsec	XP_002039497.1	Fox	Fd-19B	Dari	XP_017874440.1	Sox	Sox-100B
Dsuz	XP_016923819.2	Fox	Fd-19B	Agam	XP_061509035.1	Sox	Sox-100B
Dele	XP_041565246.1	Fox	Fd-19B	Dhyd	XP_023168522.1	Sox	Sox-100B
Dpse	XP_03239022.1	Fox	Fd-19B	Dkik	XP_017037366.1	Sox	Sox-100B
Dmir	XP_017134931.2	Fox	Fd-19B	Dana	XP_001954720.1	Sox	Sox-100B
Dari	XP_017869476.1	Fox	Fd-19B	Dele	XP_017122478.1	Sox	Sox-100B
Dgri	XP_043072169.1	Fox	Fd-19B	Dser	XP_020811779.1	Sox	Sox-100B
Dalb	XP_034101064.2	Fox	Fd-19B	Dbip	XP_017088796.2	Sox	Sox-100B
Dwil	XP_046867097.1	Fox	Fd-19B	Dmir	XP_017145320.1	Sox	Sox-100B
Dser	XP_020808848.1	Fox	Fd-19B	Dalb	XP_051863654.1	Sox	Sox-100B
Dkik	XP_017017955.1	Fox	Fd-19B	Dmel	NP_651839.1	Sox	Sox-100B
Dbus	XP_033150229.1	Fox	Fd-19B	Dsec	XP_002037434.1	Sox	Sox-100B
Agam	XP_061512060.1	Fox	-	Dere	XP_001981122.1	Sox	Sox-100B
Dalb	XP_034098689.1	Fox	-	Dsuz	XP_016935593.2	Sox	Sox-100B
Dgri	XP_032595015.1	Fox	-	Dwil	XP_023035523.1	Sox	Sox-100B
Dalb	XP_034098929.1	Fox	Slp-1	Dgri	XP_032597554.1	Sox	Sox-100B
Dele	XP_017110574.1	Fox	Slp-1	Dpse	XP_001357577.3	Sox	Sox-100B

Supplementary Table S10 – All the enriched GO terms for Group 1 and Group 2 genes of bivalves, mammals, and Drosophila.

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Bivalvia	Group 1 + Group 2	GO:0060255	regulation of macromolecule metabolic process	737	59	31.91	0.04525
Bivalvia	Group 1 + Group 2	GO:0080909	regulation of primary metabolic process	673	53	29.14	0.01818
Bivalvia	Group 1 + Group 2	GO:0019219	regulation of nucleobase-containing compound metabolic process	541	41	23.42	0.02388
Bivalvia	Group 1 + Group 2	GO:0006351	DNA-templated transcription	571	39	24.72	0.03767
Bivalvia	Group 1 + Group 2	GO:0032774	RNA biosynthetic process	579	39	25.07	0.04490
Bivalvia	Group 1 + Group 2	GO:0051252	regulation of RNA metabolic process	517	37	22.38	0.02719
Bivalvia	Group 1 + Group 2	GO:0006355	regulation of DNA-templated transcription	490	35	21.22	0.03751
Bivalvia	Group 1 + Group 2	GO:2001141	regulation of RNA biosynthetic process	491	35	21.26	0.03844
Bivalvia	Group 1 + Group 2	GO:0006950	response to stress	370	33	16.02	0.01949
Bivalvia	Group 1 + Group 2	GO:0032502	developmental process	261	27	11.3	0.04445
Bivalvia	Group 1 + Group 2	GO:0006468	protein phosphorylation	345	23	14.94	0.02483
Bivalvia	Group 1 + Group 2	GO:0031325	positive regulation of cellular metabolic process	125	17	5.41	0.00801
Bivalvia	Group 1 + Group 2	GO:0010604	positive regulation of macromolecule metabolic process	151	17	6.54	0.04047
Bivalvia	Group 1 + Group 2	GO:0051172	negative regulation of nitrogen compound metabolic process	117	16	5.07	0.00814
Bivalvia	Group 1 + Group 2	GO:0051173	positive regulation of nitrogen compound metabolic process	137	15	5.93	0.02454
Bivalvia	Group 1 + Group 2	GO:0006310	RNA recombination	66	14	2.86	0.00087
Bivalvia	Group 1 + Group 2	GO:0048513	animal organ development	83	12	3.59	0.04088
Bivalvia	Group 1 + Group 2	GO:0010629	negative regulation of gene expression	78	11	3.38	0.00048
Bivalvia	Group 1 + Group 2	GO:0023051	regulation of signaling	133	11	5.76	0.02872
Bivalvia	Group 1 + Group 2	GO:0045934	negative regulation of nucleobase-containing compound metabolic process	64	11	2.77	0.03637
Bivalvia	Group 1 + Group 2	GO:0059605	response to external stimulus	90	11	3.9	0.04544
Bivalvia	Group 1 + Group 2	GO:0044419	biological process involved in interspecies interaction between organisms	63	11	2.73	0.04761
Bivalvia	Group 1 + Group 2	GO:0006915	apoptotic process	95	10	4.11	0.00768
Bivalvia	Group 1 + Group 2	GO:0009966	regulation of signal transduction	120	10	5.2	0.03451
Bivalvia	Group 1 + Group 2	GO:0006417	regulation of translation	52	9	2.25	0.00033
Bivalvia	Group 1 + Group 2	GO:0045892	negative regulation of DNA-templated transcription	59	9	2.55	0.02968
Bivalvia	Group 1 + Group 2	GO:1902679	negative regulation of RNA biosynthetic process	59	9	2.55	0.02968
Bivalvia	Group 1 + Group 2	GO:0009607	response to biotic stimulus	55	9	2.38	0.03211
Bivalvia	Group 1 + Group 2	GO:0051253	negative regulation of RNA metabolic process	61	9	2.64	0.03719
Bivalvia	Group 1 + Group 2	GO:0006952	defense response	58	9	2.51	0.04163
Bivalvia	Group 1 + Group 2	GO:0006302	double-strand break repair	52	9	2.25	0.04860
Bivalvia	Group 1 + Group 2	GO:0080134	regulation of response to stress	52	9	2.25	0.04860
Bivalvia	Group 1 + Group 2	GO:0010564	regulation of cell cycle process	43	8	1.86	0.00669
Bivalvia	Group 1 + Group 2	GO:0042981	regulation of apoptotic process	70	8	3.03	0.01024
Bivalvia	Group 1 + Group 2	GO:0043067	regulation of programmed cell death	72	8	3.12	0.01205
Bivalvia	Group 1 + Group 2	GO:0048384	positive regulation of response to stimulus	61	8	2.64	0.03998
Bivalvia	Group 1 + Group 2	GO:0071310	cellular response to organic substance	58	9	2.51	0.04163
Bivalvia	Group 1 + Group 2	GO:0006302	regulation of gene expression	52	9	2.25	0.04860
Bivalvia	Group 1 + Group 2	GO:0045944	positive regulation of transcription by RNA polymerase II	34	7	1.47	0.02662
Bivalvia	Group 1 + Group 2	GO:1901987	regulation of cell cycle phase transition	38	6	1.65	0.00535
Bivalvia	Group 1 + Group 2	GO:2000779	regulation of double-strand break repair	29	6	1.26	0.02368
Bivalvia	Group 1 + Group 2	GO:0051247	positive regulation of protein metabolic process	11	6	0.48	0.02430
Bivalvia	Group 1 + Group 2	GO:0091248	negative regulation of protein metabolic process	54	6	2.34	0.02818
Bivalvia	Group 1 + Group 2	GO:0098557	import into cell	55	6	2.38	0.03053
Bivalvia	Group 1 + Group 2	GO:0010628	positive regulation of gene expression	56	6	2.42	0.03300
Bivalvia	Group 1 + Group 2	GO:0045944	positive regulation of transcription by RNA polymerase II	59	6	2.55	0.04118
Bivalvia	Group 1 + Group 2	GO:1902531	regulation of cell cycle phase transition	35	6	1.52	0.04666
Bivalvia	Group 1 + Group 2	GO:0044770	negative regulation of transcription by RNA polymerase II	31	5	1.34	0.00988
Bivalvia	Group 1 + Group 2	GO:0000122	mRNA catabolic process	35	5	1.52	0.01640
Bivalvia	Group 1 + Group 2	GO:0051607	defense response to virus	54	6	2.34	0.02818
Bivalvia	Group 1 + Group 2	GO:1901990	regulation of mitotic cell cycle phase transition	18	5	0.78	0.02436
Bivalvia	Group 1 + Group 2	GO:0098557	response to nitrogen compound	5	4	0.22	0.00537
Bivalvia	Group 1 + Group 2	GO:0006401	RNA catabolic process	41	5	1.78	0.03072
Bivalvia	Group 1 + Group 2	GO:0030155	regulation of cell division	44	5	1.91	0.04013
Bivalvia	Group 1 + Group 2	GO:0043066	embryonic organ development	28	4	1.21	0.03102
Bivalvia	Group 1 + Group 2	GO:0048568	negative regulation of programmed cell death	29	4	1.26	0.03480
Bivalvia	Group 1 + Group 2	GO:0007517	muscle organ development	12	4	0.52	0.00129
Bivalvia	Group 1 + Group 2	GO:0051607	cell migration	29	4	1.26	0.03480
Bivalvia	Group 1 + Group 2	GO:0032101	regulation of response to external stimulus	30	4	1.3	0.03883
Bivalvia	Group 1 + Group 2	GO:0010569	regulation of double-strand break repair via homologous recombination	21	4	0.22	0.00075
Bivalvia	Group 1 + Group 2	GO:0042274	ribosomal small subunit biogenesis	44	5	0.91	0.01148
Bivalvia	Group 1 + Group 2	GO:0043066	negative regulation of apoptotic process	28	4	1.21	0.03102
Bivalvia	Group 1 + Group 2	GO:0043069	negative regulation of programmed cell death	29	4	1.26	0.03480
Bivalvia	Group 1 + Group 2	GO:0016477	cell migration	12	4	0.52	0.00129
Bivalvia	Group 1 + Group 2	GO:0007192	regulation of response to external stimulus	30	4	1.3	0.03883
Bivalvia	Group 1 + Group 2	GO:0007669	positive regulation of neurogenesis	5	4	0.22	0.00075
Bivalvia	Group 1 + Group 2	GO:0007368	determination of left/right symmetry	7	3	0.3	0.00247
Bivalvia	Group 1 + Group 2	GO:0001819	positive regulation of cytokine production	7	3	0.3	0.00247
Bivalvia	Group 1 + Group 2	GO:0070192	chromosome organization involved in meiotic cell cycle	8	3	0.35	0.00247
Bivalvia	Group 1 + Group 2	GO:0045132	meiotic chromosome segregation	8	3	0.35	0.00382

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Bivalvia	Group 1 + Group 2	GO:0042326	negative regulation of phosphorylation	10	3	0.43	0.00768
Bivalvia	Group 1 + Group 2	GO:0008285	negative regulation of cell population proliferation	10	3	0.43	0.00768
Bivalvia	Group 1 + Group 2	GO:0022604	regulation of cell morphogenesis	10	3	0.43	0.00768
Bivalvia	Group 1 + Group 2	GO:0001894	tissue homeostasis	10	3	0.43	0.00768
Bivalvia	Group 1 + Group 2	GO:0003007	heart morphogenesis	10	3	0.43	0.00768
Bivalvia	Group 1 + Group 2	GO:0051093	negative regulation of developmental process	11	3	0.48	0.01022
Bivalvia	Group 1 + Group 2	GO:0001501	skeletal system development	11	3	0.48	0.01022
Bivalvia	Group 1 + Group 2	GO:0042327	positive regulation of phosphorylation	12	3	0.52	0.01320
Bivalvia	Group 1 + Group 2	GO:0010562	positive regulation of phosphorus metabolic process	13	3	0.56	0.01662
Bivalvia	Group 1 + Group 2	GO:0045010	actin nucleation	13	3	0.56	0.01662
Bivalvia	Group 1 + Group 2	GO:0045937	positive regulation of phosphate metabolic process	13	3	0.56	0.01662
Bivalvia	Group 1 + Group 2	GO:0007127	meiosis I	14	3	0.61	0.02049
Bivalvia	Group 1 + Group 2	GO:0031400	negative regulation of protein modification process	14	3	0.61	0.02049
Bivalvia	Group 1 + Group 2	GO:0061982	meiosis I cell cycle process	14	3	0.61	0.02049
Bivalvia	Group 1 + Group 2	GO:0097190	apoptotic signaling pathway	16	3	0.69	0.02958
Bivalvia	Group 1 + Group 2	GO:0040008	regulation of growth	16	3	0.69	0.02958
Bivalvia	Group 1 + Group 2	GO:0051345	positive regulation of hydrolase activity	17	3	0.74	0.03479
Bivalvia	Group 1 + Group 2	GO:0010257	NADH dehydrogenase complex assembly	17	3	0.74	0.03479
Bivalvia	Group 1 + Group 2	GO:0032981	mitochondrial respiratory chain complex I assembly	17	3	0.74	0.03479
Bivalvia	Group 1 + Group 2	GO:0032980	regulation of protein localization	18	3	0.78	0.04045
Bivalvia	Group 1 + Group 2	GO:0005976	polysaccharide metabolic process	18	3	0.78	0.04045
Bivalvia	Group 1 + Group 2	GO:0048729	tissue morphogenesis	19	3	0.82	0.04655
Bivalvia	Group 1 + Group 2	GO:0018022	peptidyl-lysine methylation	19	3	0.82	0.04655
Bivalvia	Group 1 + Group 2	GO:0000041	transition metal ion transport	19	3	0.82	0.04655
Bivalvia	Group 1 + Group 2	GO:0032488	Cdc42 protein signal transduction	2	2	0.09	0.00187
Bivalvia	Group 1 + Group 2	GO:0022600	digestive system signal transduction	2	2	0.09	0.00187
Bivalvia	Group 1 + Group 2	GO:0007097	nuclear migration	2	2	0.09	0.00187
Bivalvia	Group 1 + Group 2	GO:0032232	negative regulation of actin filament bundle assembly	2	2	0.09	0.00187
Bivalvia	Group 1 + Group 2	GO:10905168	positive regulation of double-strand break repair via homologous recombination	2	2	0.09	0.00187
Bivalvia	Group 1 + Group 2	GO:0002064	epithelial cell development	3	2	0.13	0.00544
Bivalvia	Group 1 + Group 2	GO:0061383	trabecula morphogenesis	3	2	0.13	0.00544
Bivalvia	Group 1 + Group 2	GO:0010830	regulation of myocyte differentiation	3	2	0.13	0.00544
Bivalvia	Group 1 + Group 2	GO:0010833	telomere maintenance via telomere lengthening	3	2	0.13	0.00544
Bivalvia	Group 1 + Group 2	GO:0000959	mitochondrial RNA metabolic process	3	2	0.13	0.00544
Bivalvia	Group 1 + Group 2	GO:0033617	mitochondrial cytochrome c oxidase assembly	3	2	0.13	0.00544
Bivalvia	Group 1 + Group 2	GO:2000179	positive regulation of neural precursor cell proliferation	3	2	0.13	0.00544
Bivalvia	Group 1 + Group 2	GO:0050777	negative regulation of immune response	3	2	0.13	0.00544
Bivalvia	Group 1 + Group 2	GO:0007095	mitotic G2 DNA damage checkpoint signaling	3	2	0.13	0.00544
Bivalvia	Group 1 + Group 2	GO:2000736	regulation of stem cell differentiation	3	2	0.13	0.00544
Bivalvia	Group 1 + Group 2	GO:0016233	telomere capping	3	2	0.13	0.00544
Bivalvia	Group 1 + Group 2	GO:0045910	negative regulation of DNA recombination	3	2	0.13	0.00544
Bivalvia	Group 1 + Group 2	GO:0051701	biological process involved in interaction with host	3	2	0.13	0.00544
Bivalvia	Group 1 + Group 2	GO:0046677	response to antibiotic	4	2	0.17	0.01058
Bivalvia	Group 1 + Group 2	GO:0046620	regulation of organ growth	4	2	0.17	0.01058
Bivalvia	Group 1 + Group 2	GO:0035023	negative regulation of BMP signaling pathway	4	2	0.17	0.01058
Bivalvia	Group 1 + Group 2	GO:10901678	iron coordination entity transport	4	2	0.17	0.01058
Bivalvia	Group 1 + Group 2	GO:0007519	skeletal muscle tissue development	4	2	0.17	0.01058
Bivalvia	Group 1 + Group 2	GO:0032507	maintenance of protein location in cell	4	2	0.17	0.01058
Bivalvia	Group 1 + Group 2	GO:0007416	synapse assembly	4	2	0.17	0.01058
Bivalvia	Group 1 + Group 2	GO:0098781	ncRNA transcription	4	2	0.17	0.01058
Bivalvia	Group 1 + Group 2	GO:0035023	regulation of Rho protein signal transduction	4	2	0.17	0.01058
Bivalvia	Group 1 + Group 2	GO:0003054	atrioventricular valve formation	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:10901678	atrioventricular valve morphogenesis	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0090101	negative regulation of transmembrane receptor protein serine/threonine kinase signaling pathway	4	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0010001	glial cell differentiation	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0090288	negative regulation of cellular response to growth factor stimulus	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0030510	regulation of BMP signaling pathway	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0061371	determination of heart left/right asymmetry	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0042063	gliogenesis	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0007162	negative regulation of cell adhesion	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0042026	protein refolding	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0007129	homologous chromosome pairing at meiosis	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0001947	heart looping	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0035050	embryonic heart tube development	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0035265	organ growth	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0030968	endoplasmic reticulum unfolded protein response	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0003188	heart valve formation	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0003171	atrioventricular valve development	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0003179	heart valve morphogenesis	5	2	0.22	0.01712

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Bivalvia	Group 1 + Group 2	GO:0003143	embryonic heart tube morphogenesis	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0090287	regulation of cellular response to growth factor stimulus	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0060538	skeletal muscle organ development	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0034620	cellular response to unfolded protein	5	2	0.22	0.01712
Bivalvia	Group 1 + Group 2	GO:0030509	BMP signaling pathway	6	2	0.26	0.02496
Bivalvia	Group 1 + Group 2	GO:0006360	transcription by RNA polymerase I	6	2	0.26	0.02496
Bivalvia	Group 1 + Group 2	GO:0045186	maintenance of protein location	6	2	0.26	0.02496
Bivalvia	Group 1 + Group 2	GO:0045143	homologous chromosome segregation	6	2	0.26	0.02496
Bivalvia	Group 1 + Group 2	GO:0001889	liver development	6	2	0.26	0.02496
Bivalvia	Group 1 + Group 2	GO:0071772	response to BMP	6	2	0.26	0.02496
Bivalvia	Group 1 + Group 2	GO:0071773	cellular response to BMP stimulus	6	2	0.26	0.02496
Bivalvia	Group 1 + Group 2	GO:0090092	regulation of transmembrane receptor protein serine/threonine kinase signaling pathway	6	2	0.26	0.02496
Bivalvia	Group 1 + Group 2	GO:0003170	heart valve development	6	2	0.26	0.02496
Bivalvia	Group 1 + Group 2	GO:0003012	muscle system process	6	2	0.26	0.02496
Bivalvia	Group 1 + Group 2	GO:0030490	maturaton of SSU-rRNA	7	2	0.3	0.03395
Bivalvia	Group 1 + Group 2	GO:0018105	peptidyl-serine phosphorylation	7	2	0.3	0.03395
Bivalvia	Group 1 + Group 2	GO:0032465	regulation of cytoskeleton	7	2	0.3	0.03395
Bivalvia	Group 1 + Group 2	GO:0061448	connective tissue development	7	2	0.3	0.03395
Bivalvia	Group 1 + Group 2	GO:0018209	peptidyl-serine modification	7	2	0.3	0.03395
Bivalvia	Group 1 + Group 2	GO:0032102	negative regulation of response to external stimulus	7	2	0.3	0.03395
Bivalvia	Group 1 + Group 2	GO:0061008	hematopoietic system development	7	2	0.3	0.03395
Bivalvia	Group 1 + Group 2	GO:0048038	regulation of developmental growth	7	2	0.3	0.03395
Bivalvia	Group 1 + Group 2	GO:0010212	response to ionizing radiation	7	2	0.3	0.03395
Bivalvia	Group 1 + Group 2	GO:0034329	cell junction assembly	7	2	0.3	0.03395
Bivalvia	Group 1 + Group 2	GO:1001652	response to peptide	8	2	0.35	0.04400
Bivalvia	Group 1 + Group 2	GO:0048732	gland development	8	2	0.35	0.04400
Bivalvia	Group 1 + Group 2	GO:0033157	regulation of intracellular protein transport	8	2	0.35	0.04400
Bivalvia	Group 1 + Group 2	GO:0051302	regulation of cell division	8	2	0.35	0.04400
Bivalvia	Group 1 + Group 2	GO:0001822	kidney development	8	2	0.35	0.04400
Bivalvia	Group 1 + Group 2	GO:0050808	synapse organization	8	2	0.35	0.04400
Bivalvia	Group 1 + Group 2	GO:0035967	cellular response to topologically incorrect protein	8	2	0.35	0.04400
Bivalvia	Group 1 + Group 2	GO:0031032	actomyosin structure organization	8	2	0.35	0.04400
Bivalvia	Group 1 + Group 2	GO:0001503	osification	8	2	0.35	0.04400
Bivalvia	Group 1 + Group 2	GO:0002711	polysaccharide biosynthetic process	8	2	0.35	0.04400
Bivalvia	Group 1 + Group 2	GO:0008893	regulation of Notch signaling pathway	8	2	0.35	0.04400
Bivalvia	Group 1 + Group 2	GO:0001822	negative regulation of cell differentiation	8	2	0.35	0.04400
Bivalvia	Group 1 + Group 2	GO:0050808	regulation of defense response to virus by host	8	2	0.35	0.04400
Bivalvia	Group 1 + Group 2	GO:0035967	semaphorin-phekin signaling pathway involved in bone trabecula morphogenesis	8	2	0.35	0.04400
Bivalvia	Group 1 + Group 2	GO:0002720	negative regulation of organ growth	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0046622	animal organ maturation	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0048799	O antigen metabolic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0046402	malate transport	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0015743	regulatory ncRNA 3'-end processing	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0043628	inhibitory synapse assembly	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:1904862	mitochondrial RNA processing	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0000963	positive regulation of defense response to virus by host	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0002230	regulation of skeletal muscle fiber development	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0048742	positive regulation of skeletal muscle fiber development	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0048743	regulation of extrinsic apoptotic signaling pathway via death domain receptors	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:1902041	response to host immune response	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0075136	response to host	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:1904862	semaphorin-phekin signaling pathway involved in neuron projection guidance	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0003687	osteoblast proliferation	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:1902287	lipopolysaccharide metabolic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0048742	regulation of osteoblast proliferation	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0033688	regulation of extrinsic apoptotic signaling pathway via death domain receptors	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0030655	protein poly-ADP-ribosylation	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0070212	beta-lactam antibiotic catabolic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0001100	negative regulation of exit from mitosis	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0043144	extrinsic apoptotic signaling pathway via death domain receptors	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0031848	sn(s)RNA processing	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0010526	protection from non-homologous end joining at telomere	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0061668	retrotransposon silencing	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0035518	mitochondrial ribosome assembly	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0035518	protein localization to synapse	1	1	0.04	0.04330

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Bivalvia	Group 1 + Group 2	GO:0061430	bone trabecula morphogenesis	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0006356	regulation of transcription by RNA polymerase I	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:101269	lipopolysaccharide metabolic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0035622	intramitotic bile duct development	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0016444	somatic cell DNA recombination	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:101271	lipopolysaccharide biosynthetic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0952173	response to defenses of other organism	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0003382	epithelial cell morphogenesis	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0070977	bone maturation	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0031571	mitotic G1 DNA damage checkpoint signaling	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0043247	telomere maintenance in response to DNA damage	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0010669	epitethelial structure maintenance	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0009301	snRNA transcription	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0042149	cellular response to glucose starvation	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:1002914	regulation of protein polyubiquitination	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:1002915	negative regulation of protein polyubiquitination	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0032196	transposition	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0032197	retrotransposition	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0034964	box H/ACA sno(s)RNA processing	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0007168	receptor guanylyl cyclase signaling pathway	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0017001	antibiotic catabolic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0030277	maintenance of gastrointestinal epithelium	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:00040495	box H/ACA sno(s)RNA 3'-end processing	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:10903513	endoplasmic reticulum to cytosol transport	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:11900481	mitochondrial tRNA synthesis	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0044919	mitotic G1/S transition checkpoint signaling	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0051155	positive regulation of striated muscle cell differentiation	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0007140	male meiotic nuclear division	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0050772	positive regulation of axogenesis	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0090646	mitochondrial tRNA processing	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0099172	presynapse organization	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0032065	maintenance of protein location in cell cortex	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:1004152	regulation of retrograde protein transport, ER to cytosol	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0097222	mitochondrial mRNA polyadenylation	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0051149	positive regulation of muscle cell differentiation	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0020033	antigenic variation	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0052200	responsible for host defenses	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0016074	sno(s)RNA metabolic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0043931	osmification involved in bone maturation	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0035279	miRNA-mediated gene silencing by mRNA destabilization	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0061009	common bile duct development	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0099054	presynapse assembly	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0048641	regulation of skeletal muscle tissue development	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0048643	positive regulation of skeletal muscle tissue development	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:1001861	regulation of muscle tissue development	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0010863	positive regulation of muscle tissue development	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0048634	regulation of muscle organ development	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0031120	snRNA pseudouridine synthase	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0031126	sn(s)RNA 3'-end processing	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0033979	box H/ACA sno(s)RNA metabolic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0071966	fungiform-type cell wall polysaccharide metabolic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0042783	evasion of host immune response	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0030870	retrograde protein transport, ER to cytosol	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0051274	beta-D-glucan biosynthetic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0051278	fungiform-type cell wall polysaccharide biosynthetic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0009103	lipopolysaccharide biosynthetic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0009292	nuclear matrix anchoring at nuclear membrane	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0071947	protein deubiquitination involved in ubiquitin-dependent protein catabolic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0045643	protein ubiquitination involved in transcription by RNA polymerase I	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0048144	fibroblast proliferation	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0048145	regulation of fibroblast proliferation	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0048147	negative regulation of fibroblast proliferation	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0072340	lactam catabolic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0006074	D->D-glucan metabolic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0006075	(1->3)-beta-D-glucan biosynthetic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0016699	antibiotic metabolic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0072338	lactam metabolic process	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:0044650	adhesion of symbiont to host cell	1	1	0.04	0.04330
Bivalvia	Group 1 + Group 2	GO:1002414	protein localization to cell junction	1	1	0.04	0.04330

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Bivalvia	Group 1 + Group 2	GO:0009272	fungal-type cell wall biogenesis	1	1	0.04	0.0430
Bivalvia	Group 1 + Group 2	GO:0009243	O antigen biosynthetic process	1	1	0.04	0.0430
Bivalvia	Group 1 + Group 2	GO:0043878	nuclear matrix organization	1	1	0.04	0.0430
Bivalvia	Group 1 + Group 2	GO:0009245	lipid A biosynthetic process	1	1	0.04	0.0430
Bivalvia	Group 1 + Group 2	GO:0044406	adhesion of symbiont to host	1	1	0.04	0.0430
Bivalvia	Group 1 + Group 2	GO:0032978	protein insertion into membrane from inner side	1	1	0.04	0.0430
Bivalvia	Group 1 + Group 2	GO:0032979	protein insertion into mitochondrial inner membrane from matrix	1	1	0.04	0.0430
Bivalvia	Group 1 + Group 2	GO:0046493	lipid A metabolic process	1	1	0.04	0.0430
Bivalvia	Group 1 + Group 2	GO:0072695	regulation of DNA recombination at telomere	1	1	0.04	0.0430
Bivalvia	Group 1 + Group 2	GO:1000044	regulation of protein K63-linked ubiquitination	1	1	0.04	0.0430
Bivalvia	Group 1 + Group 2	GO:0935871	protein K11-linked ubiquitination	1	1	0.04	0.0430
Bivalvia	Group 1 + Group 2	GO:1900455	negative regulation of DNA recombination at telomere	1	1	0.04	0.0430
Bivalvia	Group 1 + Group 2	GO:0048239	negative regulation of DNA recombination at telomere	1	1	0.04	0.0430
Drosophila	Group 1 + Group 2	GO:0045132	meiotic chromosome segregation	64	11	2.63	0.00145
Drosophila	Group 1 + Group 2	GO:0008119	sister chromatid segregation	140	11	5.75	0.02927
Drosophila	Group 1 + Group 2	GO:0070192	chromosome organization involved in meiotic cell cycle	54	9	2.22	0.00849
Drosophila	Group 1 + Group 2	GO:10007131	reciprocal meiotic recombination	37	7	1.52	0.00066
Drosophila	Group 1 + Group 2	GO:0007143	female meiotic nucle division	54	6	2.22	0.02270
Drosophila	Group 1 + Group 2	GO:0035967	cellular response to topologically incorrect protein	44	5	1.81	0.03334
Drosophila	Group 1 + Group 2	GO:0045161	meiotic chromosome segregation	47	5	0.49	0.04266
Drosophila	Group 1 + Group 2	GO:0007141	male meiosis I	13	4	0.53	0.00150
Drosophila	Group 1 + Group 2	GO:0140543	positive regulation of piRNA transcription	3	3	0.12	6.9e-05
Drosophila	Group 1 + Group 2	GO:0010852	retrotransposon silencing	8	3	0.33	0.00331
Drosophila	Group 1 + Group 2	GO:0007130	synaptonemal complex assembly	10	3	0.41	0.00666
Drosophila	Group 1 + Group 2	GO:0030719	P granule organization	11	3	0.45	0.00888
Drosophila	Group 1 + Group 2	GO:0035916	kinetochore assembly	12	3	0.49	0.01149
Drosophila	Group 1 + Group 2	GO:0051718	cellular response to misfolded protein	12	3	0.49	0.01149
Drosophila	Group 1 + Group 2	GO:0007138	response to misfolded protein	12	3	0.49	0.01149
Drosophila	Group 1 + Group 2	GO:0034808	meiosis II	15	3	0.62	0.02169
Drosophila	Group 1 + Group 2	GO:0010850	centromere complex assembly	19	3	0.78	0.04094
Drosophila	Group 1 + Group 2	GO:0007130	male germ-line cyst formation	2	2	0.08	0.00168
Drosophila	Group 1 + Group 2	GO:0061964	negative regulation of entry into reproductive diapause	5	2	0.21	0.01551
Drosophila	Group 1 + Group 2	GO:0051382	kinetochore assembly	5	2	0.21	0.01551
Drosophila	Group 1 + Group 2	GO:0051712	entry into reproductive diapause	6	2	0.25	0.02264
Drosophila	Group 1 + Group 2	GO:0051712	ER-associated misfolded protein catabolic process	6	2	0.25	0.02264
Drosophila	Group 1 + Group 2	GO:0007130	regulation of entry into reproductive diapause	6	2	0.25	0.02264
Drosophila	Group 1 + Group 2	GO:0048136	histone H4-K16 acetylation	6	2	0.25	0.02264
Drosophila	Group 1 + Group 2	GO:0007130	entry into diapause	7	2	0.29	0.03084
Drosophila	Group 1 + Group 2	GO:0990834	response to odorant	7	2	0.29	0.03084
Drosophila	Group 1 + Group 2	GO:0051177	meiotic sister chromatid cohesion	8	2	0.33	0.04002
Drosophila	Group 1 + Group 2	GO:0042795	snRNA transcription by RNA polymerase II	8	2	0.33	0.04002
Drosophila	Group 1 + Group 2	GO:0022611	dormancy process	8	2	0.33	0.04002
Drosophila	Group 1 + Group 2	GO:0009301	snRNA transcription	8	2	0.33	0.04002
Drosophila	Group 1 + Group 2	GO:0045144	meiotic sister chromatid segregation	8	2	0.33	0.04002
Drosophila	Group 1 + Group 2	GO:0001015	snRNA transcription by RNA polymerase II	1	1	0.04	0.04109
Drosophila	Group 1 + Group 2	GO:0072765	meiotic DNA repair synthesis involved in reciprocal meiotic recombination	1	1	0.04	0.04109
Drosophila	Group 1 + Group 2	GO:0009302	centromere localization	1	1	0.04	0.04109
Drosophila	Group 1 + Group 2	GO:0051308	(s)s RNA transcription	1	1	0.04	0.04109
Drosophila	Group 1 + Group 2	GO:0098653	male meiosis chromosome separation	1	1	0.04	0.04109
Drosophila	Group 1 + Group 2	GO:0051415	centromere clustering	1	1	0.04	0.04109
Mammalia	Group 1 + Group 2	GO:0006955	microtubule nucleation by interphase microtubule organizing center	1	1	0.04	0.04109
Mammalia	Group 1 + Group 2	GO:0098542	immune response	145	44	48.02	0.00061
Mammalia	Group 1 + Group 2	GO:0042795	defense response to other organism	853	112	31.98	0.02066
Mammalia	Group 1 + Group 2	GO:0001817	innate immune response	647	82	8.5e-10	0.04660
Mammalia	Group 1 + Group 2	GO:0001819	regulation of cytokine production	630	51	23.33	0.04426
Mammalia	Group 1 + Group 2	GO:0027424	defense response to bacterium	233	45	8.63	1.7e-07
Mammalia	Group 1 + Group 2	GO:0006954	inflammatory response	642	45	23.77	0.01735
Mammalia	Group 1 + Group 2	GO:0019221	cytokine-mediated signaling pathway	382	44	14.14	3.9e-07
Mammalia	Group 1 + Group 2	GO:0002250	adaptive immune response	1297	44	48.02	0.00061
Mammalia	Group 1 + Group 2	GO:0098542	defense response to virus	342	44	12.66	1.3e-07
Mammalia	Group 1 + Group 2	GO:0070661	male gamete generation	491	32	18.18	0.02255
Mammalia	Group 1 + Group 2	GO:0001819	spematogenesis	478	31	17.7	0.02801
Mammalia	Group 1 + Group 2	GO:0002449	leukocyte proliferation	273	29	10.11	0.01285
Mammalia	Group 1 + Group 2	GO:0070663	lymphocyte mediated immunity	221	29	8.18	0.04833
Mammalia	Group 1 + Group 2	GO:0050727	regulation of leukocyte proliferation	212	25	7.85	0.01870
Mammalia	Group 1 + Group 2	GO:0031349	regulation of inflammatory response	300	24	11.11	0.00235
Mammalia	Group 1 + Group 2	GO:0048232	positive regulation of defense response	240	24	8.89	0.01239

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Mammalia	Group 1 + Group 2	GO:0002768	immune response-regulating cell surface receptor signaling pathway	177	22	6.55	0.00336
Mammalia	Group 1 + Group 2	GO:0050829	defense response to Gram-negative bacterium	66	17	2.44	1.7e-10
Mammalia	Group 1 + Group 2	GO:0071222	cellular response to lipopolysaccharide	164	17	6.07	0.00012
Mammalia	Group 1 + Group 2	GO:0010466	negative regulation of peptidase activity	163	16	6.04	0.00036
Mammalia	Group 1 + Group 2	GO:0002429	immune response-activating cell surface receptor signaling pathway	164	16	6.07	0.00243
Mammalia	Group 1 + Group 2	GO:1903555	regulation of tumor necrosis factor superfamily cytokine production	137	16	5.07	0.01244
Mammalia	Group 1 + Group 2	GO:0971706	tumor necrosis factor superfamily cytokine production	137	16	5.07	0.01244
Mammalia	Group 1 + Group 2	GO:0070665	positive regulation of leukocyte proliferation	132	16	4.89	0.02765
Mammalia	Group 1 + Group 2	GO:0045089	positive regulation of innate immune response	113	16	4.18	0.03224
Mammalia	Group 1 + Group 2	GO:0071356	cellular response to tumor necrosis factor	175	15	6.48	0.00219
Mammalia	Group 1 + Group 2	GO:0002955	negative regulation of leukocyte activation	148	15	5.48	0.01151
Mammalia	Group 1 + Group 2	GO:0002456	T cell mediated immunity	82	15	3.04	0.01605
Mammalia	Group 1 + Group 2	GO:0002705	positive regulation of leukocyte mediated immunity	113	15	4.18	0.01837
Mammalia	Group 1 + Group 2	GO:0032680	regulation of tumor necrosis factor production	133	15	4.92	0.03262
Mammalia	Group 1 + Group 2	GO:0052860	tumor necrosis factor production	133	15	4.92	0.03262
Mammalia	Group 1 + Group 2	GO:0050866	negative regulation of cell activation	165	15	6.11	0.04048
Mammalia	Group 1 + Group 2	GO:0013141	regulation of cell killing	71	14	2.63	0.00628
Mammalia	Group 1 + Group 2	GO:0001818	negative regulation of cytokine production	225	14	8.33	0.04065
Mammalia	Group 1 + Group 2	GO:0050830	defensive response to Gram-positive bacterium	87	13	3.22	1.8e-05
Mammalia	Group 1 + Group 2	GO:0002286	regulation of tumor necrosis factor production	94	13	3.48	0.02934
Mammalia	Group 1 + Group 2	GO:0050809	cellular response to tumor necrosis factor	52	13	1.93	0.04957
Mammalia	Group 1 + Group 2	GO:0007259	negative regulation of cell activation	134	12	4.96	0.00411
Mammalia	Group 1 + Group 2	GO:00019731	regulation of cytokine production	130	12	4.81	0.01823
Mammalia	Group 1 + Group 2	GO:00019731	antibacterial humoral response	40	11	1.48	1.4e-07
Mammalia	Group 1 + Group 2	GO:00019731	natural killer cell activation	66	11	2.44	2.7e-05
Mammalia	Group 1 + Group 2	GO:0002100	T cell activation involved in immune response	68	11	3.15	0.00029
Mammalia	Group 1 + Group 2	GO:0002102	positive regulation of T cell proliferation	48	11	1.78	0.00080
Mammalia	Group 1 + Group 2	GO:0050832	defense response to fungus	98	11	3.63	0.00099
Mammalia	Group 1 + Group 2	GO:0071342	cellular response to type II interferon	120	11	4.44	0.00494
Mammalia	Group 1 + Group 2	GO:0008584	male gonad development	42	11	1.56	0.00768
Mammalia	Group 1 + Group 2	GO:0002820	negative regulation of adaptive immune response	162	11	6	0.03860
Mammalia	Group 1 + Group 2	GO:0043123	positive regulation of canonical NF-kappaB signal transduction	76	11	2.81	0.04485
Mammalia	Group 1 + Group 2	GO:0042100	B cell proliferation	29	10	1.07	4.7e-08
Mammalia	Group 1 + Group 2	GO:0002102	detection of chemical stimulus involved in sensory perception of bitter taste	80	10	2.96	0.00071
Mammalia	Group 1 + Group 2	GO:0030893	neutrophil chemotaxis	47	10	1.74	0.00078
Mammalia	Group 1 + Group 2	GO:0050832	complement activation	84	10	3.11	0.00105
Mammalia	Group 1 + Group 2	GO:0006956	positive regulation of tumor necrosis factor production	84	10	3.11	0.00105
Mammalia	Group 1 + Group 2	GO:0032760	cellular response to type II interferon	122	10	4.52	0.01515
Mammalia	Group 1 + Group 2	GO:0071347	positive regulation of inflammatory response [...]	39	10	1.44	0.03055
Mammalia	Group 1 + Group 2	GO:0002823	cellular response to interleukin-1	62	10	2.3	0.03820
Mammalia	Group 1 + Group 2	GO:0050688	negative regulation of adaptive immune response [...]	98	10	3.63	0.04352
Mammalia	Group 1 + Group 2	GO:0002718	regulation of defense response to virus	98	10	3.63	0.04352
Mammalia	Group 1 + Group 2	GO:0002367	regulation of cytokine production involved in immune response	35	9	1.3	3.7e-06
Mammalia	Group 1 + Group 2	GO:0007339	cytokine production involved in immune response	64	9	2.37	0.00055
Mammalia	Group 1 + Group 2	GO:0050729	binding of sperm to zona pellucida	64	9	2.37	0.00055
Mammalia	Group 1 + Group 2	GO:0042100	antimicrobial humoral immune response mediated by antimicrobial peptide	64	9	2.37	0.00055
Mammalia	Group 1 + Group 2	GO:0001580	killing of cells of another organism	63	9	3.55	0.00913
Mammalia	Group 1 + Group 2	GO:0002102	negative regulation of defense response to virus	59	9	2.18	0.01359
Mammalia	Group 1 + Group 2	GO:0002102	regulation of cytokine production involved in immune response	114	9	4.22	0.02862
Mammalia	Group 1 + Group 2	GO:0002102	flagellated sperm motility	119	9	4.41	0.03259
Mammalia	Group 1 + Group 2	GO:0061844	sperm motility	119	9	4.41	0.03259
Mammalia	Group 1 + Group 2	GO:0038061	non-canonical NF-kappaB signal transduction	120	9	4.44	0.03413
Mammalia	Group 1 + Group 2	GO:0072978	T cell migration	54	8	2.33	0.00078
Mammalia	Group 1 + Group 2	GO:0060759	regulation of response to cytokine stimulus	124	9	4.59	0.04078
Mammalia	Group 1 + Group 2	GO:0060294	cilium movement involved in cell motility	128	9	4.74	0.04826
Mammalia	Group 1 + Group 2	GO:0007342	fusion of sperm to egg plasma membrane involved in single fertilization	25	8	0.93	2.1e-06
Mammalia	Group 1 + Group 2	GO:0045071	negative regulation of viral genome replication	39	8	1.44	7.5e-05
Mammalia	Group 1 + Group 2	GO:0002218	activation of innate immune response	45	8	1.67	0.00022
Mammalia	Group 1 + Group 2	GO:0032757	positive regulation of interleukin-8 production	54	8	2	0.00078
Mammalia	Group 1 + Group 2	GO:0030888	regulation of B cell activation	54	8	2	0.00078
Mammalia	Group 1 + Group 2	GO:0060294	positive regulation of chemokine production	60	8	2.22	0.00158
Mammalia	Group 1 + Group 2	GO:0010921	regulation of phosphatase activity	65	8	2.41	0.00265
Mammalia	Group 1 + Group 2	GO:0070098	chemokine-mediated signaling pathway	69	8	2.55	0.00385
Mammalia	Group 1 + Group 2	GO:0002920	regulation of humoral immune response	39	8	1.33	0.00512
Mammalia	Group 1 + Group 2	GO:0043303	mast cell degranulation	43	8	1.59	0.00850
Mammalia	Group 1 + Group 2	GO:0002251	organ or tissue specific immune response	22	8	0.81	0.00652
Mammalia	Group 1 + Group 2	GO:0002886	regulation of myeloid leukocyte mediated immunity	45	8	1.67	0.00784
Mammalia	Group 1 + Group 2	GO:0043026	regulation of macrophage activation	46	8	1.7	0.00922
Mammalia	Group 1 + Group 2	GO:0002926	acute inflammatory response	83	8	3.07	0.01162
Mammalia	Group 1 + Group 2	GO:0032649	regulation of type II interferon production	88	8	3.26	0.01615

Tab. S10 continued from previous page

				Annotated	Significant	Expected	classicFisher
Dataset	Group of genes	GO-ID	Term	Annotated	Significant	Expected	classicFisher
Mammalia	Group 1 + Group 2	GO:0032609	type I interferon production	88	8	3.26	0.01615
Mammalia	Group 1 + Group 2	GO:0050691	regulation of defense response to virus by host	37	8	1.37	0.04975
Mammalia	Group 1 + Group 2	GO:0042267	natural killer cell mediated cytotoxicity	44	7	1.63	0.00107
Mammalia	Group 1 + Group 2	GO:008247	lymphocyte chemotaxis	49	7	1.81	0.00204
Mammalia	Group 1 + Group 2	GO:0042119	neutrophil activation	32	7	1.18	0.00786
Mammalia	Group 1 + Group 2	GO:002945	negative regulation of mononuclear cell proliferation	63	7	2.33	0.00845
Mammalia	Group 1 + Group 2	GO:002720	positive regulation of cytokine production involved in immune response	66	7	2.44	0.01083
Mammalia	Group 1 + Group 2	GO:0035456	response to interferon-beta	66	7	0.96	0.01252
Mammalia	Group 1 + Group 2	GO:0080853	B cell receptor signaling pathway	45	7	1.67	0.01683
Mammalia	Group 1 + Group 2	GO:0085526	cellular response to virus	78	7	2.89	0.02523
Mammalia	Group 1 + Group 2	GO:0084625	regulation of receptor signaling pathway via JAK-STAT	78	7	0.3253	0.03253
Mammalia	Group 1 + Group 2	GO:0002385	mucosal immune response	19	7	0.07	0.03395
Mammalia	Group 1 + Group 2	GO:0032755	positive regulation of interleukin-6 production	83	7	3.07	0.04679
Mammalia	Group 1 + Group 2	GO:1901222	regulation of non-canonical NF-kappaB signal transduction	89	7	3.3	0.22e-05
Mammalia	Group 1 + Group 2	GO:008245	eosinophil chemotaxis	63	6	0.63	0.00024
Mammalia	Group 1 + Group 2	GO:008240	sperm capacitation	25	6	0.93	0.00031
Mammalia	Group 1 + Group 2	GO:0070269	pyroptotic inflammatory response	26	6	0.96	0.00038
Mammalia	Group 1 + Group 2	GO:0002230	positive regulation of defense response to virus by host	27	6	1	0.00038
Mammalia	Group 1 + Group 2	GO:0031295	T cell costimulation	27	6	1	0.00058
Mammalia	Group 1 + Group 2	GO:0028285	positive regulation of T-helper 1 type immune response	38	6	1.07	0.00252
Mammalia	Group 1 + Group 2	GO:002691	regulation of cellular extravasation	41	6	1.52	0.00374
Mammalia	Group 1 + Group 2	GO:0140374	antiviral innate immune response	44	6	1.63	0.00535
Mammalia	Group 1 + Group 2	GO:0026359	positive regulation of immunoglobulin production	46	6	1.7	0.00667
Mammalia	Group 1 + Group 2	GO:0032731	positive regulation of interleukin-1 beta production	46	6	1.7	0.00742
Mammalia	Group 1 + Group 2	GO:1904894	positive regulation of receptor signaling pathway via STAT	47	6	1.74	0.01000
Mammalia	Group 1 + Group 2	GO:0034113	heterotypic cell-cell adhesion	50	6	1.85	0.01000
Mammalia	Group 1 + Group 2	GO:0025458	monocyte chemotaxis	54	6	2	0.01000
Mammalia	Group 1 + Group 2	GO:001961	positive regulation of cytokine-mediated signaling pathway	55	6	0.67	0.01079
Mammalia	Group 1 + Group 2	GO:0030449	regulation of complement activation	18	6	0.89	0.01099
Mammalia	Group 1 + Group 2	GO:002531	positive regulation of tyrosine phosphorylation of ST/AT protein	51	6	1.93	0.01205
Mammalia	Group 1 + Group 2	GO:0003337	type I interferon-mediated signaling pathway	52	6	1.93	0.01205
Mammalia	Group 1 + Group 2	GO:001357	cellular response to type I interferon	52	6	1.93	0.01256
Mammalia	Group 1 + Group 2	GO:0001914	regulation of T cell mediated cytotoxicity	54	6	2	0.01437
Mammalia	Group 1 + Group 2	GO:0051283	establishment of spindle-mediated localization	54	6	0.85	0.01626
Mammalia	Group 1 + Group 2	GO:0031873	killing by host of symbiont cells	23	6	2.22	0.02316
Mammalia	Group 1 + Group 2	GO:0061070	positive regulation of neuroinflammatory response	57	6	2.11	0.01841
Mammalia	Group 1 + Group 2	GO:0185077	response to type I interferon	28	6	1.04	0.01949
Mammalia	Group 1 + Group 2	GO:0034340	regulation of neuronflammatory response	58	6	2.15	0.01991
Mammalia	Group 1 + Group 2	GO:0051653	spindle localization	59	6	2.18	0.02150
Mammalia	Group 1 + Group 2	GO:001895	retina homeostasis	60	6	2.22	0.02316
Mammalia	Group 1 + Group 2	GO:0071260	cellular response to mechanical stimulus	60	6	2.22	0.02676
Mammalia	Group 1 + Group 2	GO:0080672	negative regulation of lymphocyte proliferation	62	6	2.3	0.02676
Mammalia	Group 1 + Group 2	GO:0032729	positive regulation of type II interferon production	62	6	2.3	0.02676
Mammalia	Group 1 + Group 2	GO:0042509	tyrosine phosphorylation of STAT protein	66	6	2.44	0.03500
Mammalia	Group 1 + Group 2	GO:0007260	tyrosine phosphorylation of STAT protein	11	5	0.41	2.6e-05
Mammalia	Group 1 + Group 2	GO:0002227	innate immune response in mucosa	31	5	0.52	0.00010
Mammalia	Group 1 + Group 2	GO:0028330	positive regulation of type 2 immune response	32	5	1.18	0.00010
Mammalia	Group 1 + Group 2	GO:0035435	response to interferon-alpha	14	5	0.52	0.00010
Mammalia	Group 1 + Group 2	GO:0033005	positive regulation of mast cell activation	17	5	0.63	0.00029
Mammalia	Group 1 + Group 2	GO:0061760	antifungal innate immune response	17	5	0.63	0.00029
Mammalia	Group 1 + Group 2	GO:0054536	cellular response to interferon-beta	21	5	0.78	0.00085
Mammalia	Group 1 + Group 2	GO:0046596	regulation of viral entry into host cell	31	5	1.15	0.00523
Mammalia	Group 1 + Group 2	GO:0007340	acrosome reaction	45	5	1.67	0.02467
Mammalia	Group 1 + Group 2	GO:0069698	cellular defense response	34	5	1.26	0.00782
Mammalia	Group 1 + Group 2	GO:001774	microglial cell activation	35	5	1.3	0.00895
Mammalia	Group 1 + Group 2	GO:002701	negative regulation of production of molecular mediator of immune response	47	5	1.3	0.00885
Mammalia	Group 1 + Group 2	GO:0140632	canonical inflammasome complex assembly	35	5	1.3	0.00885
Mammalia	Group 1 + Group 2	GO:006953	acute-phase response	38	5	1.41	0.01250
Mammalia	Group 1 + Group 2	GO:0032653	regulation of interleukin-10 production	45	5	1.67	0.02467
Mammalia	Group 1 + Group 2	GO:002613	regulation of phosphoprotein phosphatase activity	46	5	1.7	0.02686
Mammalia	Group 1 + Group 2	GO:003666	negative regulation of tumor necrosis factor production	47	5	1.74	0.02918
Mammalia	Group 1 + Group 2	GO:1903536	positive regulation of dephosphorylation	47	5	1.74	0.02918
Mammalia	Group 1 + Group 2	GO:0003556	negative regulation of tumor necrosis factor superfamily cytokine production	49	5	1.81	0.03417
Mammalia	Group 1 + Group 2	GO:0070228	regulation of lymphocyte apoptotic process	50	5	1.85	0.03686
Mammalia	Group 1 + Group 2	GO:002683	T cell extravasation	13	4	0.48	0.01012
Mammalia	Group 1 + Group 2	GO:002710	negative regulation of T cell mediated immunity	15	4	0.56	0.00183
Mammalia	Group 1 + Group 2	GO:005624	T helper 17 lineage commitment	17	4	0.63	0.00301
Mammalia	Group 1 + Group 2	GO:005624	positive regulation of T-cell differentiation	17	4	0.63	0.00301

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Mammalia	Group 1 + Group 2	GO:1903659	regulation of complement-dependent cytotoxicity	5	4	0.19	0.00398
Mammalia	Group 1 + Group 2	GO:0008228	opsonization	19	4	0.7	0.00462
Mammalia	Group 1 + Group 2	GO:0043302	positive regulation of leukocyte degranulation	20	4	0.74	0.00561
Mammalia	Group 1 + Group 2	GO:0001916	positive regulation of T cell mediated cytotoxicity	20	4	0.74	0.00561
Mammalia	Group 1 + Group 2	GO:0001862	response to protozoan	21	4	0.78	0.00673
Mammalia	Group 1 + Group 2	GO:0002717	negative regulation of natural killer cell mediated immunity	21	4	0.78	0.00673
Mammalia	Group 1 + Group 2	GO:1903601	negative regulation of viral life cycle	21	4	0.78	0.00673
Mammalia	Group 1 + Group 2	GO:0035821	modulation of process of another organism	21	4	0.78	0.00673
Mammalia	Group 1 + Group 2	GO:0043304	regulation of mast cell degranulation	23	4	0.85	0.00940
Mammalia	Group 1 + Group 2	GO:0032740	positive regulation of interleukin-17 production	23	4	0.85	0.00940
Mammalia	Group 1 + Group 2	GO:0043032	positive regulation of macrophage activation	23	4	0.85	0.00940
Mammalia	Group 1 + Group 2	GO:0070498	interleukin-1-mediated signaling pathway	24	4	0.89	0.01096
Mammalia	Group 1 + Group 2	GO:0010922	positive regulation of phosphatase activity	26	4	0.96	0.01454
Mammalia	Group 1 + Group 2	GO:0019884	antigen processing and presentation of exogenous antigen	28	4	1.04	0.01880
Mammalia	Group 1 + Group 2	GO:0002846	neutrophil mediated immunity	29	4	1.07	0.02118
Mammalia	Group 1 + Group 2	GO:0032743	positive regulation of interleukin-2 production	30	4	1.11	0.02375
Mammalia	Group 1 + Group 2	GO:200352	negative regulation of endothelial cell apoptotic process	30	4	1.11	0.02375
Mammalia	Group 1 + Group 2	GO:1900255	regulation of NLRP3 inflammasome complex assembly	30	4	1.11	0.02375
Mammalia	Group 1 + Group 2	GO:0043330	response to exogenous dsRNA	31	4	1.15	0.02649
Mammalia	Group 1 + Group 2	GO:0032733	positive regulation of interleukin-10 production	32	4	1.18	0.02942
Mammalia	Group 1 + Group 2	GO:0046006	regulation of activated T cell proliferation	32	4	1.18	0.02942
Mammalia	Group 1 + Group 2	GO:0032814	regulation of natural killer cell activation	32	4	1.18	0.02942
Mammalia	Group 1 + Group 2	GO:0045456	NLRP3 inflammasome complex assembly	33	4	1.22	0.03252
Mammalia	Group 1 + Group 2	GO:0050798	activated T cell proliferation	34	4	1.26	0.03582
Mammalia	Group 1 + Group 2	GO:0046636	negative regulation of alpha-beta T cell activation	35	4	1.3	0.03930
Mammalia	Group 1 + Group 2	GO:0043331	response to dsRNA	36	4	1.33	0.04297
Mammalia	Group 1 + Group 2	GO:0035307	positive regulation of protein dephosphorylation	36	4	1.33	0.04297
Mammalia	Group 1 + Group 2	GO:0030895	positive regulation of B cell proliferation	37	4	1.37	0.04682
Mammalia	Group 1 + Group 2	GO:0038095	Fc-receptor signaling pathway	37	5	1.37	0.04682
Mammalia	Group 1 + Group 2	GO:0045959	negative regulation of complement activation, classical pathway	5	3	0.19	0.00048
Mammalia	Group 1 + Group 2	GO:0038156	interleukin-3-mediated signaling pathway	7	3	0.19	0.00048
Mammalia	Group 1 + Group 2	GO:0051838	cytolysis by host of symbiont cells	7	3	0.26	0.00158
Mammalia	Group 1 + Group 2	GO:0006907	DNA damage response, signal transduction by p53 class mediator resulting in cell cycle arrest	8	3	0.3	0.00246
Mammalia	Group 1 + Group 2	GO:0097527	reciprocal signaling pathway	8	3	0.3	0.00246
Mammalia	Group 1 + Group 2	GO:0006924	activation-induced cell death of T cells	10	3	0.37	0.00499
Mammalia	Group 1 + Group 2	GO:1902563	regulation of neutrophil activation	10	3	0.37	0.00499
Mammalia	Group 1 + Group 2	GO:0010526	retrotransposon silencing	10	3	0.37	0.00499
Mammalia	Group 1 + Group 2	GO:0032754	positive regulation of interleukin-5 production	10	3	0.37	0.00499
Mammalia	Group 1 + Group 2	GO:200851	regulation of T-helper 2 cell cytokine production	10	3	0.37	0.00499
Mammalia	Group 1 + Group 2	GO:1901731	positive regulation of plakophilin aggregation	10	3	0.37	0.00499
Mammalia	Group 1 + Group 2	GO:0032736	positive regulation of interleukin-13 production	11	3	0.41	0.00667
Mammalia	Group 1 + Group 2	GO:0007343	egg activation	11	3	0.41	0.00667
Mammalia	Group 1 + Group 2	GO:0035723	interleukin-15-mediated signaling pathway	11	3	0.41	0.00667
Mammalia	Group 1 + Group 2	GO:0050079	negative regulation of neuroinflammatory response	12	3	0.44	0.00865
Mammalia	Group 1 + Group 2	GO:0060046	regulation of acrosome reaction	12	3	0.48	0.01094
Mammalia	Group 1 + Group 2	GO:0002323	natural killer cell activation involved in immune response	13	3	0.48	0.01094
Mammalia	Group 1 + Group 2	GO:0030889	T-helper 1-cell differentiation	13	3	0.48	0.01094
Mammalia	Group 1 + Group 2	GO:0006957	negative regulation of macrophage activation	15	3	0.56	0.01647
Mammalia	Group 1 + Group 2	GO:0002730	regulation of dendritic cell cytokine production	15	3	0.56	0.01647
Mammalia	Group 1 + Group 2	GO:0032516	changes to DNA methylation involved in embryo development	16	3	0.59	0.01973
Mammalia	Group 1 + Group 2	GO:0046597	negative regulation of viral entry into host cell	16	3	0.59	0.01973
Mammalia	Group 1 + Group 2	GO:0043045	epigenetic programming of gene expression	16	3	0.59	0.01973
Mammalia	Group 1 + Group 2	GO:0002371	dendritic-cell cytokine production	16	3	0.59	0.01973
Mammalia	Group 1 + Group 2	GO:1901538	defense response to protozoan	16	3	0.59	0.01973
Mammalia	Group 1 + Group 2	GO:0042832	MHC class II biosynthetic process	17	3	0.63	0.02331
Mammalia	Group 1 + Group 2	GO:0045342	positive regulation of embryonic development	17	3	0.63	0.02331
Mammalia	Group 1 + Group 2	GO:0040019	positive regulation of type I interferon-mediated signaling pathway	17	3	0.63	0.02331
Mammalia	Group 1 + Group 2	GO:0060340	positive regulation of cellular extravasation	18	3	0.67	0.02722
Mammalia	Group 1 + Group 2	GO:0045346	positive regulation of MHC class II biosynthetic process	18	3	0.67	0.02722
Mammalia	Group 1 + Group 2	GO:0042832	defense response to protozoan	19	3	0.7	0.03145
Mammalia	Group 1 + Group 2	GO:0045342	MHC class II biosynthetic process	19	3	0.7	0.03145
Mammalia	Group 1 + Group 2	GO:0040019	positive regulation of embryonic development	19	3	0.7	0.03145
Mammalia	Group 1 + Group 2	GO:0060340	positive regulation of type I interferon-mediated signaling pathway	19	3	0.7	0.03145

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Mammalia	Group 1 + Group 2	GO:0060333	type II interferon-mediated signaling pathway	19	3	0.7	0.03145
Mammalia	Group 1 + Group 2	GO:0032753	positive regulation of interleukin-4 production	20	3	0.74	0.03602
Mammalia	Group 1 + Group 2	GO:0045063	T-helper 1-cell differentiation	21	3	0.78	0.04090
Mammalia	Group 1 + Group 2	GO:0042104	positive regulation of activated T cell proliferation	22	3	0.81	0.04609
Mammalia	Group 1 + Group 2	GO:0002726	positive regulation of T cell cytokine production	22	3	0.81	0.04609
Mammalia	Group 1 + Group 2	GO:1903660	negative regulation of complement-dependent cytotoxicity	2	2	0.07	0.00137
Mammalia	Group 1 + Group 2	GO:0997528	executive phase of necrosis	2	2	0.07	0.00137
Mammalia	Group 1 + Group 2	GO:0030887	positive regulation of myeloid dendritic cell activation	2	2	0.07	0.00137
Mammalia	Group 1 + Group 2	GO:0009609	response to symbiotic bacterium	4	2	0.15	0.00781
Mammalia	Group 1 + Group 2	GO:0048006	antigen processing and presentation, endogenous lipid antigen via MHC class Ib	4	2	0.15	0.00781
Mammalia	Group 1 + Group 2	GO:0050859	negative regulation of B cell receptor signalling pathway	4	2	0.15	0.00781
Mammalia	Group 1 + Group 2	GO:00027165	immune response to signal transduction	5	2	0.19	0.01270
Mammalia	Group 1 + Group 2	GO:0036015	response to interleukin-3	5	2	0.19	0.01270
Mammalia	Group 1 + Group 2	GO:2000436	cellular response to interleukin-3	5	2	0.19	0.01270
Mammalia	Group 1 + Group 2	GO:0033634	regulation of eosinophil migration	5	2	0.19	0.01270
Mammalia	Group 1 + Group 2	GO:0045630	positive regulation of cell-cell adhesion mediated by integrin	5	2	0.19	0.01270
Mammalia	Group 1 + Group 2	GO:0042104	positive regulation of T-helper 2-cell differentiation	6	2	0.19	0.01270
Mammalia	Group 1 + Group 2	GO:0043313	regulation of neutrophil degranulation	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0033141	positive regulation of peptidyl-serine phosphorylation of STAT protein	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0002826	negative regulation of T-helper 1-type immune response	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0016300	protein-DNA covalent cross-linking repair	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0045341	MHC class I biosynthetic process	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0043307	regulation of MHC class I biosynthetic process	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0002733	antigen processing, presentation, exogenous lipid antigen via MHC class Ib	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0002735	positive regulation of response to type II interferon	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:000332	positive regulation of type II interferon-mediated signaling pathway	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0006559	regulation of interleukin-1-mediated signaling pathway	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0060345	MHC class I biosynthetic process	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0048007	positive regulation of necrotic cell cross-linking repair	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:00060332	positive regulation of type II interferon	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0006331	positive regulation of interleukin-1-mediated signaling pathway	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0006320	positive regulation of type II interferon-mediated signaling pathway	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0045343	positive regulation of necrotic cell cross-linking repair	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0048007	positive regulation of necrotic cell cross-linking repair	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0002735	positive regulation of type II interferon	6	2	0.22	0.01859
Mammalia	Group 1 + Group 2	GO:0003391	positive regulation of neutrophil extravasation	7	2	0.26	0.02539
Mammalia	Group 1 + Group 2	GO:0010940	positive regulation of necrotic cell death	7	2	0.26	0.02539
Mammalia	Group 1 + Group 2	GO:0060467	negative regulation of fertilization	7	2	0.26	0.02539
Mammalia	Group 1 + Group 2	GO:0060468	prevention of polyspermy	7	2	0.26	0.02539
Mammalia	Group 1 + Group 2	GO:0071358	cellular response to type III interferon	7	2	0.26	0.02539
Mammalia	Group 1 + Group 2	GO:0038196	type III interferon-mediated signaling pathway	7	2	0.26	0.02539
Mammalia	Group 1 + Group 2	GO:0002372	neutrophil degranulation	7	2	0.26	0.02539
Mammalia	Group 1 + Group 2	GO:0062172	myeloid dendritic cell cytokine production	7	2	0.26	0.02539
Mammalia	Group 1 + Group 2	GO:0001781	positive regulation of programmed necrotic cell death	7	2	0.26	0.02539
Mammalia	Group 1 + Group 2	GO:0043312	neutrophil apoptosis process	8	2	0.3	0.03304
Mammalia	Group 1 + Group 2	GO:0042796	snRNA transcription by RNA polymerase III	8	2	0.3	0.03304
Mammalia	Group 1 + Group 2	GO:0034342	response to type III interferon	8	2	0.3	0.03304
Mammalia	Group 1 + Group 2	GO:1003577	interleukin-4-mediated signaling pathway	8	2	0.3	0.03304
Mammalia	Group 1 + Group 2	GO:1003889	positive regulation of blood vessel endothelial cell proliferation involved in sprouting angiogenesis	8	2	0.3	0.03304
Mammalia	Group 1 + Group 2	GO:0003874	positive regulation of neutrophil extravasation	8	2	0.3	0.03304
Mammalia	Group 1 + Group 2	GO:0002638	negative regulation of acute inflammatory response	8	2	0.3	0.03304
Mammalia	Group 1 + Group 2	GO:0002650	negative regulation of immunoglobulin production	8	2	0.3	0.03304
Mammalia	Group 1 + Group 2	GO:0032610	regulation of interleukin-1-alpha production	8	2	0.3	0.03304
Mammalia	Group 1 + Group 2	GO:0045671	interleukin-4-mediated signaling pathway	8	2	0.3	0.03304
Mammalia	Group 1 + Group 2	GO:0001352	negative regulation of single stranded viral RNA replication via double stranded DNA intermediate	8	2	0.3	0.03304
Mammalia	Group 1 + Group 2	GO:2000553	cellular response to interleukin-2	8	2	0.3	0.03304
Mammalia	Group 1 + Group 2	GO:0038110	positive regulation of T-helper 2-cell cytokine production	8	2	0.3	0.03304
Mammalia	Group 1 + Group 2	GO:0033004	interleukin-2-mediated signaling pathway	8	2	0.3	0.03304
Mammalia	Group 1 + Group 2	GO:0070669	negative regulation of mast cell activation	8	2	0.33	0.04145
Mammalia	Group 1 + Group 2	GO:0061518	response to interleukin-2	9	2	0.33	0.04145
Mammalia	Group 1 + Group 2	GO:0033139	microglial cell proliferation	9	2	0.33	0.04145
Mammalia	Group 1 + Group 2	GO:0044006	regulation of peptidyl-serine phosphorylation of STAT protein	9	2	0.33	0.04145
Mammalia	Group 1 + Group 2	GO:0010918	adhesion of symbiont to host	9	2	0.33	0.04145
Mammalia	Group 1 + Group 2	GO:0033632	positive regulation of mitochondrial membrane potential	9	2	0.33	0.04145
Mammalia	Group 1 + Group 2	GO:0045625	regulation of cell-cell adhesion mediated by integrin	9	2	0.33	0.04145
Mammalia	Group 1 + Group 2	GO:0045628	regulation of T-helper 1-cell differentiation	9	2	0.33	0.04145
Mammalia	Group 1 + Group 2	GO:1901625	regulation of protein heterodimerization activity	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:1003497	regulation of activation of Janus kinase activity	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:1002569	negative regulation of IP-10 production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0044355	clearance of foreign intracellular DNA	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0071652	regulation of chemokine (C-C motif) ligand 1 production	1	1	0.04	0.03702

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Mammalia	Group 1 + Group 2	GO:0071654	positive regulation of chemokine (C-C motif) ligand 1 production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0002736	regulation of plasmacytoid dendritic cell cytokine production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0002737	negative regulation of plasmacytoid dendritic cell cytokine production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0071610	chemokine (C-C motif) ligand 1 production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0051673	disruption of plasma membrane integrity in another organism	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0150073	regulation of protein glutamine gamma-glutamyltransferase activity	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0150074	positive regulation of protein glutamine gamma-glutamyltransferase activity	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0032759	positive regulation of TNF</math> production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:1905151	NLRP1 inflammasome complex assembly	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:1600006	negative regulation of voltage-gated sodium channel activity	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0932723	Fc receptor-mediated immune complex endocytosis	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:1905154	positive regulation of connective tissue growth factor production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0032745	positive regulation of membrane invagination	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0016068	positive regulation of interleukin-21 production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:1904784	type I hypersensitivity	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0035397	helper T cell enhancement of adaptive immune response	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:1905223	epicardium morphogenesis	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0050902	leukocyte adhesive activation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0002518	lymphocyte chemotaxis across high endothelial venule	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0034156	negative regulation of toll-like receptor 7 signaling pathway	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:1904848	negative regulation of cell chemotaxis to fibroblast growth factor	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0120042	negative regulation of macrophage proliferation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0034125	negative regulation of MyD88-dependent toll-like receptor signaling pathway	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0060097	cytoskeletal rearrangement involved in phagocytosis, engulfment	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:2000422	regulation of eosinophil chemotaxis	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:2000424	positive regulation of eosinophil chemotaxis	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0090073	positive regulation of protein homodimerization activity	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0051977	lysophospholipid transport	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:032665	regulation of interleukin-21 production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0032679	regulation of TRAIL production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0032925	interleukin-21 production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0326339	TRAIL production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0001971	negative regulation of activation of membrane attack complex	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0036496	regulation of transnational initiation by eIF2 alpha dephosphorylation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0036497	eIF2alpha dephosphorylation in response to endoplasmic reticulum stress	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:2000229	regulation of pancreatic stellate cell proliferation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0051041	positive regulation of calcium-independent cell-cell adhesion	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:2000231	positive regulation of pancreatic stellate cell proliferation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:1900450	negative regulation of glutamate receptor signaling pathway	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0018003	peptidyl-lysine N6-acetylation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:1903916	regulation of endoplasmic reticulum stress-induced eIF2 alpha dephosphorylation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:1903917	positive regulation of endoplasmic reticulum stress-induced eIF2 alpha dephosphorylation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0090046	cleavage of foreign intracellular nucleic acids	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0150140	regulation of CD86 production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0150142	positive regulation of CD86 production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0150143	regulation of CD80 production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0140122	regulation of Lewy body formation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0140123	triglyceride acyl-chain remodeling	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0361653	acylglycerol acyl-chain remodeling	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0036155	negative regulation of adaptative immune memory response	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:1905675	regulation of chylomicron remnant clearance	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:00900320	positive regulation of chylomicron remnant clearance	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0150142	Lewy body formation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0140122	regulation of Lewy body formation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0060101	negative regulation of endothelial cell chemotaxis to fibroblast growth factor	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0070246	natural killer cell apoptotic process	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:1901251	regulation of natural killer cell apoptosis	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:1901247	positive regulation of lung goblet cell differentiation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:1901249	negative regulation of lung ciliated cell differentiation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:030186	regulation of lung goblet cell differentiation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0030187	melatonin metabolic process	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0072343	pancreatic stellate cell proliferation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0010186	regulation of cellular defense response	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0002384	positive regulation of cellular defense response	1	1	0.04	0.03702

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Mammalia	Group 1 + Group 2	GO:1903496	response to 11-deoxycorticosterone	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0002373	plasmacytoid dendritic cell cytokine production	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0098784	biomatrix organization	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0098786	biomatrix disassembly	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0002450	B cell antigen processing and presentation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0046603	negative regulation of mitotic centrosome separation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0002470	plasmacytoid dendritic cell antigen processing and presentation	1	1	0.04	0.03702
Mammalia	Group 1 + Group 2	GO:0072139	glomerular parietal epithelial cell differentiation	1	1	0.04	0.03702
Bivalvia	Group 1 + Group 2	GO:0080090	regulation of primary metabolic process	673	47	23.83	0.02305
Bivalvia	Group 1	GO:0019219	regulation of nucleobase-containing compound metabolic process	541	36	19.16	0.02702
Bivalvia	Group 1	GO:0051252	regulation of RNA metabolic process	517	32	18.31	0.03415
Bivalvia	Group 1	GO:0069500	response to stress	370	29	13.1	0.02704
Bivalvia	Group 1	GO:0033854	cellular response to stress	275	21	9.74	0.04194
Bivalvia	Group 1	GO:0051172	negative regulation of nitrogen compound metabolic process	117	15	4.14	0.00749
Bivalvia	Group 1	GO:0031325	positive regulation of cellular metabolic process	125	15	4.43	0.01000
Bivalvia	Group 1	GO:0006310	DNA recombination	66	13	2.34	0.00999
Bivalvia	Group 1	GO:0051173	positive regulation of nitrogen compound metabolic process	137	13	4.85	0.03359
Bivalvia	Group 1	GO:010629	negative regulation of gene expression	78	11	2.76	0.00344
Bivalvia	Group 1	GO:0048513	animal organ development	83	11	2.94	0.01719
Bivalvia	Group 1	GO:0045934	negative regulation of nucleobase-containing compound metabolic process	64	11	2.27	0.01863
Bivalvia	Group 1	GO:0006915	apoptotic process	95	9	3.36	0.00621
Bivalvia	Group 1	GO:0045892	negative regulation of DNA-templated transcription	59	9	2.09	0.01512
Bivalvia	Group 1	GO:1902679	negative regulation of RNA biosynthetic process	59	9	2.09	0.01512
Bivalvia	Group 1	GO:0051253	negative regulation of RNA metabolic process	61	9	2.16	0.01918
Bivalvia	Group 1	GO:0006417	regulation of translation	52	8	1.84	0.00043
Bivalvia	Group 1	GO:0051726	regulation of cell cycle	75	8	2.66	0.03901
Bivalvia	Group 1	GO:0065009	regulation of molecular function	114	8	4.04	0.04846
Bivalvia	Group 1	GO:0045893	positive regulation of DNA-templated transcription	67	7	2.37	0.00908
Bivalvia	Group 1	GO:0042981	regulation of apoptotic process	70	7	2.48	0.01145
Bivalvia	Group 1	GO:0043067	regulation of programmed cell death	72	7	2.55	0.01326
Bivalvia	Group 1	GO:0000122	negative regulation of transcription by RNA polymerase II	31	5	1.1	0.00426
Bivalvia	Group 1	GO:0006402	mRNA catabolic process	35	5	1.24	0.00725
Bivalvia	Group 1	GO:0045944	positive regulation of transcription by RNA polymerase II	38	5	1.35	0.01030
Bivalvia	Group 1	GO:0071310	cellular response to organic substance	52	5	1.84	0.03886
Bivalvia	Group 1	GO:0009628	response to abiotic stimulus	53	5	1.88	0.03850
Bivalvia	Group 1	GO:0051248	negative regulation of protein metabolic process	55	5	1.95	0.04413
Bivalvia	Group 1	GO:0303155	regulation of cell adhesion	11	4	0.39	0.00041
Bivalvia	Group 1	GO:0048868	embryonic organ development	12	4	0.42	0.00061
Bivalvia	Group 1	GO:0051607	defense response to virus	13	4	0.46	0.00085
Bivalvia	Group 1	GO:0010569	regulation of double-strand break repair via homologous recombination	15	4	0.18	0.00359
Bivalvia	Group 1	GO:0000902	cell morphogenesis	31	4	1.1	0.02274
Bivalvia	Group 1	GO:000280	nuclear division	38	4	1.35	0.04409
Bivalvia	Group 1	GO:0050769	positive regulation of neurogenesis	5	3	0.18	0.00042
Bivalvia	Group 1	GO:0001819	positive regulation of cytokine production	7	3	0.25	0.00138
Bivalvia	Group 1	GO:0070192	chromosome organization involved in meiotic cell cycle	7	3	0.25	0.00138
Bivalvia	Group 1	GO:0007368	determination of left/right symmetry	7	3	0.25	0.00138
Bivalvia	Group 1	GO:0001894	tissue homeostasis	10	3	0.35	0.00437
Bivalvia	Group 1	GO:0003007	heart morphogenesis	10	3	0.35	0.00437
Bivalvia	Group 1	GO:0008285	negative regulation of cell population proliferation	10	3	0.35	0.00437
Bivalvia	Group 1	GO:0051093	negative regulation of developmental process	11	3	0.39	0.00585
Bivalvia	Group 1	GO:0001501	skeletal system development	11	3	0.39	0.00585
Bivalvia	Group 1	GO:0007517	muscle organ development	13	3	0.46	0.00963
Bivalvia	Group 1	GO:0061982	meiosis I cell cycle process	14	3	0.5	0.01194
Bivalvia	Group 1	GO:0007127	meiosis I	14	3	0.5	0.01194
Bivalvia	Group 1	GO:0010257	NADH dehydrogenase complex assembly	17	3	0.6	0.02063
Bivalvia	Group 1	GO:0032981	mitochondrial respiratory chain complex I assembly	17	3	0.6	0.02063
Bivalvia	Group 1	GO:0051345	positive regulation of hydrodase activity	17	3	0.6	0.02063
Bivalvia	Group 1	GO:040013	meiotic nuclear division	18	3	0.64	0.02412
Bivalvia	Group 1	GO:0005976	polysaccharide metabolic process	19	3	0.67	0.02791
Bivalvia	Group 1	GO:048729	tissue morphogenesis	20	3	0.71	0.03200
Bivalvia	Group 1	GO:035295	tube development	21	3	0.74	0.03639
Bivalvia	Group 1	GO:0042274	ribosomal small subunit biogenesis	23	3	0.81	0.04603
Bivalvia	Group 1	GO:0022603	regulation of anatomical structure morphogenesis	23	3	0.81	0.04603
Bivalvia	Group 1	GO:0140053	mitochondrial gene expression	2	2	0.07	0.00125
Bivalvia	Group 1	GO:1905168	positive regulation of double-strand break repair via homologous recombination	2	2	0.07	0.00125
Bivalvia	Group 1	GO:032488	Cdc42 protein signal transduction	2	2	0.07	0.00125
Bivalvia	Group 1	GO:002280	digestive system process	2	2	0.07	0.00125
Bivalvia	Group 1	GO:032232	negative regulation of actin filament bundle assembly	2	2	0.07	0.00125
Bivalvia	Group 1	GO:033617	mitochondrial cytochrome c oxidase assembly	2	2	0.11	0.00366

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Bivalvia	Group 1	GO:0045910	negative regulation of DNA recombination	3	2	0.11	0.00366
Bivalvia	Group 1	GO:2000179	positive regulation of neural precursor cell proliferation	3	2	0.11	0.00366
Bivalvia	Group 1	GO:0061383	trabecula morphogenesis	3	2	0.11	0.00366
Bivalvia	Group 1	GO:0051701	biological process involved in interaction with host	3	2	0.11	0.00366
Bivalvia	Group 1	GO:0010833	telomere maintenance via telomere lengthening	3	2	0.11	0.00366
Bivalvia	Group 1	GO:0007095	mitotic G2 DNA damage checkpoint signaling	3	2	0.11	0.00366
Bivalvia	Group 1	GO:0000959	mitochondrial RNA metabolic process	3	2	0.11	0.00366
Bivalvia	Group 1	GO:0002064	epithelial cell development	3	2	0.11	0.00366
Bivalvia	Group 1	GO:0010001	glial cell differentiation	4	2	0.14	0.00714
Bivalvia	Group 1	GO:0046620	regulation of organ growth	4	2	0.14	0.00714
Bivalvia	Group 1	GO:0003190	atrioventricular-valve formation	4	2	0.14	0.00714
Bivalvia	Group 1	GO:0098781	ncRNA transcription	4	2	0.14	0.00714
Bivalvia	Group 1	GO:0030514	negative regulation of BMP signaling pathway	4	2	0.14	0.00714
Bivalvia	Group 1	GO:0035023	regulation of Rho protein signal transduction	4	2	0.14	0.00714
Bivalvia	Group 1	GO:0007416	synapse assembly	4	2	0.14	0.00714
Bivalvia	Group 1	GO:0001947	heart looping	5	2	0.18	0.01163
Bivalvia	Group 1	GO:0042026	protein refolding	5	2	0.18	0.01163
Bivalvia	Group 1	GO:0003143	embryonic heart tube morphogenesis	5	2	0.18	0.01163
Bivalvia	Group 1	GO:0003143	determination of heart left/right asymmetry	5	2	0.18	0.01163
Bivalvia	Group 1	GO:0061371	embryonic heart tube morphogenesis	5	2	0.18	0.01163
Bivalvia	Group 1	GO:0035050	determination of heart left/right asymmetry	5	2	0.18	0.01163
Bivalvia	Group 1	GO:0007162	embryonic heart tube morphogenesis	5	2	0.18	0.01163
Bivalvia	Group 1	GO:0007129	negative regulation of cell adhesion	5	2	0.18	0.01163
Bivalvia	Group 1	GO:0006360	homologous chromosome pairing at meiosis	6	2	0.21	0.01704
Bivalvia	Group 1	GO:0001889	transcription by RNA polymerase I	6	2	0.21	0.01704
Bivalvia	Group 1	GO:0001822	liver development	6	2	0.21	0.01704
Bivalvia	Group 1	GO:0045143	homologous chromosome segregation	6	2	0.21	0.01704
Bivalvia	Group 1	GO:0061008	hepatobiliary system development	6	2	0.25	0.02330
Bivalvia	Group 1	GO:0010212	response to ionizing radiation	7	2	0.25	0.02330
Bivalvia	Group 1	GO:0030490	maturative regulation of SSU-rRNA	7	2	0.25	0.02330
Bivalvia	Group 1	GO:0061448	connective tissue development	7	2	0.25	0.02330
Bivalvia	Group 1	GO:0048732	gland development	8	2	0.28	0.03036
Bivalvia	Group 1	GO:0001822	kidney development	8	2	0.28	0.03036
Bivalvia	Group 1	GO:0045596	negative regulation of cell differentiation	8	2	0.28	0.03036
Bivalvia	Group 1	GO:0000271	polysaccharide biosynthetic process	8	2	0.28	0.03036
Bivalvia	Group 1	GO:0001503	ossification	8	2	0.28	0.03036
Bivalvia	Group 1	GO:0045132	meiotic chromosome segregation	8	2	0.28	0.03036
Bivalvia	Group 1	GO:0016073	snRNA metabolic process	9	2	0.32	0.03813
Bivalvia	Group 1	GO:0072001	renal system development	9	2	0.32	0.03813
Bivalvia	Group 1	GO:0060862	epithelial tube morphogenesis	9	2	0.32	0.03813
Bivalvia	Group 1	GO:0048862	embryonic organ morphogenesis	9	2	0.32	0.03813
Bivalvia	Group 1	GO:0042326	negative regulation of phosphorylation	10	2	0.35	0.04657
Bivalvia	Group 1	GO:0001934	positive regulation of protein phosphorylation	10	2	0.35	0.04657
Bivalvia	Group 1	GO:0022604	regulation of cell morphogenesis	10	2	0.35	0.04657
Bivalvia	Group 1	GO:0034314	Arp2/3 complex-mediated actin nucleation	10	2	0.35	0.04657
Bivalvia	Group 1	GO:0010669	epithelial structure maintenance	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0046621	negative regulation of organ growth	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0097222	mitochondrial mRNA polyadenylation	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0061668	mitochondrial ribosome assembly	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0048799	animal organ maturation	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0044406	adhesion of symbiont to host	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0043247	telomere maintenance in response to DNA damage	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0061009	common bile duct development	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0096356	regulation of transcription by RNA polymerase I	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0000444	snRNA pseudouridine synthesis	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0031126	sn(s)RNA 3'-end processing	1	1	0.04	0.03541
Bivalvia	Group 1	GO:1002285	semaphorin-plexin signaling pathway involved in neuron projection guidance	1	1	0.04	0.03541
Bivalvia	Group 1	GO:1000045	adhesion of symbiont to host cell	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0090646	mitochondrial rRNA processing	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0016074	sn(s)RNA metabolic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0070977	protein insertion into membrane from inner side	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0031120	inhibitory synapse assembly	1	1	0.04	0.03541
Bivalvia	Group 1	GO:1004862	regulation of DNA recombination at telomere	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0002230	positive regulation of defense response to virus by host	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0008653	lipopolysaccharide metabolic process	1	1	0.04	0.03541

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Bivalvia	Group 1	GO:0045943	positive regulation of transcription by RNA polymerase I	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0052972	response to host immune response	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0051274	beta-glucan biosynthetic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0051278	fung-type cell wall polysaccharide biosynthetic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:1900220	semaphorin-peptid signaling pathway involved in bone trabecula morphogenesis	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0098625	extrinsic apoptotic signaling pathway via death domain receptors	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0909301	snRNA processing	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0015743	snRNA transport	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0032065	maintenance of protein location in cell cortex	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0032196	transposition	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0032197	retrotransposition	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0042149	cellular response to glucose starvation	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0052173	response to defenses of other organism	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0006074	(1->3)-beta-D-glucan metabolic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0006075	(1->3)-beta-D-glucan biosynthetic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0095136	response to host	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0099054	presynapse assembly	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0035618	protein localization to synapse	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0043931	osification involved in bone maturation	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0071966	fung-type cell wall polysaccharide metabolic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0099172	presynapse organization	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0033079	beta-H/ACA sn(s)RNA metabolic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:1002914	regulation of protein polyubiquitination	1	1	0.04	0.03541
Bivalvia	Group 1	GO:1002915	negative regulation of protein polyubiquitination	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0071947	protein deubiquitination involved in ubiquitin-dependent protein catabolic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0050691	regulation of defense response to virus by host	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0046493	lipid A metabolic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0017001	antibiotic catabolic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0071423	malate transmembrane transport	1	1	0.04	0.03541
Bivalvia	Group 1	GO:1901269	lipopolysaccharide metabolic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0070212	protein poly-ADP-ribosylation	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0070213	protein auto-ADP-ribosylation	1	1	0.04	0.03541
Bivalvia	Group 1	GO:1002414	protein localization to cell junction	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0001100	negative regulation of exit from mitosis	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0048144	fibroblast proliferation	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0070220	regulation of fibroblast proliferation	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0048145	negative regulation of fibroblast proliferation	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0048147	lipopolysaccharide biosynthetic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:1001271	intraluminal bile duct development	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0035622	beta-lactam antibiotic metabolic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0030653	beta-lactam antibiotic catabolic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0030655	response to host defenses	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0052200	mRNA pseudouridine synthesis	1	1	0.04	0.03541
Bivalvia	Group 1	GO:1000481	lipopolysaccharide biosynthetic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0009103	protection from non-homologous end joining at telomere	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0048634	bone trabecula morphogenesis	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0010826	retratransposon silencing	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0046402	O antigen metabolic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0042783	evasion of host immune response	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0043144	sno(s)RNA processing	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0016999	antibiotic metabolic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0072340	lactam catabolic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:1002041	regulation of extrinsic apoptotic signaling pathway via death domain receptors	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0020033	fung-type cell wall biogenesis	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0000863	mitochondrial RNA processing	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0007168	recently guanylyl cyclase signaling pathway	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0043628	regulatory ncRNA 3'-end processing	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0016999	protection from non-homologous end joining at telomere	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0061430	bone trabecula morphogenesis	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0048634	antigenic variation	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0010826	lactam metabolic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0009124	male meiotic nuclear division	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0003382	epithelial cell morphogenesis	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0009243	O antigen biosynthetic process	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0033687	osteoblast proliferation	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0033688	regulation of osteoblast proliferation	1	1	0.04	0.03541

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Bivalvia	Group 1	GO:0033689	negative regulation of osteoblast proliferation	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0030277	maintenance of gastrointestinal epithelium	1	1	0.04	0.03541
Bivalvia	Group 1	GO:0048239	negative regulation of DNA recombination at telomere	1	1	0.04	0.03541
Drosophila	Group 1	GO:0051276	chromosome organization	331	18	11.07	0.02925
Drosophila	Group 1	GO:0045132	meiotic chromosome segregation	64	7	2.14	0.00533
Drosophila	Group 1	GO:0071131	reciprocal meiotic recombination	37	6	1.24	0.00130
Drosophila	Group 1	GO:0931146	SCF-dependent proteasomal ubiquitin-dependent protein catabolic process	57	5	1.91	0.04125
Drosophila	Group 1	GO:0007141	male meiosis I	13	4	0.43	0.00069
Drosophila	Group 1	GO:1002275	regulation of chromatin organization	35	4	1.17	0.02847
Drosophila	Group 1	GO:0042078	germ-line stem cell division	36	4	1.2	0.03121
Drosophila	Group 1	GO:0010526	retrotransposon silencing	8	3	0.27	0.00183
Drosophila	Group 1	GO:0071218	cellular response to misfolded protein	12	3	0.4	0.00652
Drosophila	Group 1	GO:0034508	centromere complex assembly	19	3	0.64	0.02415
Drosophila	Group 1	GO:0007080	mitotic metaphase chromosome alignment	23	3	0.77	0.04004
Drosophila	Group 1	GO:0007020	microtubule nucleation	23	3	0.77	0.04004
Drosophila	Group 1	GO:0031445	regulation of heterochromatin formation	25	3	0.84	0.04953
Drosophila	Group 1	GO:0120261	regulation of heterochromatin organization	25	3	0.84	0.04953
Drosophila	Group 1	GO:0048136	male germ-line cyst formation	2	2	0.07	0.00112
Drosophila	Group 1	GO:0061964	negative regulation of entry into reproductive diapause	5	2	0.17	0.01043
Drosophila	Group 1	GO:0051382	kinetochore assembly	5	2	0.17	0.01043
Drosophila	Group 1	GO:0055116	entry into reproductive diapause	6	2	0.2	0.01531
Drosophila	Group 1	GO:0071712	ER-associated misfolded protein catabolic process	6	2	0.2	0.01531
Drosophila	Group 1	GO:0061963	regulation of entry into reproductive diapause	6	2	0.2	0.01531
Drosophila	Group 1	GO:0043984	histone H4-K16 acetylation	6	2	0.2	0.01531
Drosophila	Group 1	GO:0055115	entry into diapause	7	2	0.23	0.02096
Drosophila	Group 1	GO:1990834	response to odorant	7	2	0.23	0.02096
Drosophila	Group 1	GO:0042795	snRNA transcription by RNA polymerase II	8	2	0.27	0.02733
Drosophila	Group 1	GO:0022611	dormancy process	8	2	0.27	0.02733
Drosophila	Group 1	GO:0009301	snRNA transcription	8	2	0.27	0.02733
Drosophila	Group 1	GO:0071786	endoplasmic reticulum tubular network organization	9	2	0.3	0.03438
Drosophila	Group 1	GO:0030007	intracellular potassium ion homeostasis	9	2	0.3	0.03438
Drosophila	Group 1	GO:0051383	kinetochore organization	9	2	0.3	0.03438
Drosophila	Group 1	GO:0040020	regulation of meiotic nuclear division	9	2	0.3	0.03438
Drosophila	Group 1	GO:0042795	histone H4 acetylation	9	2	0.3	0.03438
Drosophila	Group 1	GO:0036376	sodium ion export across plasma membrane	9	2	0.3	0.03438
Drosophila	Group 1	GO:0006883	intracellular sodium ion homeostasis	9	2	0.3	0.03438
Drosophila	Group 1	GO:0001015	snRNA transcription by RNA polymerase II	1	1	0.03	0.03345
Drosophila	Group 1	GO:0010778	positive regulation of DNA repair synthesis involved in reciprocal meiotic recombination	1	1	0.03	0.03345
Drosophila	Group 1	GO:0093020	sn(s)RNA transcription	1	1	0.03	0.03345
Drosophila	Group 1	GO:0051308	male meiosis chromosome separation	1	1	0.03	0.03345
Drosophila	Group 1	GO:0051415	microtubule nucleation by interphase microtubule organizing center	1	1	0.03	0.03345
Mammalia	Group 1	GO:0006955	innate immune response	1297	120	38.75	0.00270
Mammalia	Group 1	GO:0045087	T cell activation	647	69	19.33	1.6e-06
Mammalia	Group 1	GO:0007978	positive regulation of immune response	419	47	12.52	0.00677
Mammalia	Group 1	GO:0022597	adaptive immune response	342	39	10.22	1.4e-05
Mammalia	Group 1	GO:0002694	regulation of leukocyte activation	456	39	13.62	0.00521
Mammalia	Group 1	GO:0001819	positive regulation of cytokine production	402	37	12.01	0.00251
Mammalia	Group 1	GO:0019221	cytokine-mediated signaling pathway	382	35	11.41	2.6e-05
Mammalia	Group 1	GO:0042742	defense response to bacterium	233	32	6.96	0.00036
Mammalia	Group 1	GO:0042110	T cell activation	432	31	12.91	0.01328
Mammalia	Group 1	GO:0002697	regulation of immune effector process	308	31	9.2	0.01876
Mammalia	Group 1	GO:0051607	defense response to virus	257	28	7.68	4.9e-06
Mammalia	Group 1	GO:0002703	myeloid leukocyte activation	195	26	5.83	0.02936
Mammalia	Group 1	GO:0001819	regulation of leukocyte mediated immunity	186	25	5.56	0.01358
Mammalia	Group 1	GO:0070661	leukocyte proliferation	273	24	4.57	0.02709
Mammalia	Group 1	GO:0031349	mononuclear cell differentiation	240	20	8.16	0.00474
Mammalia	Group 1	GO:0002699	regulation of inflammatory response	392	23	11.71	0.04822
Mammalia	Group 1	GO:0930217	T cell differentiation	300	21	8.96	0.00247
Mammalia	Group 1	GO:0045089	positive regulation of innate immune response	113	16	3.38	0.01390
Mammalia	Group 1	GO:0071222	cellular response to a lipopolysaccharide	164	15	4.9	0.00012
Mammalia	Group 1	GO:1003555	regulation of tumor necrosis factor superfamily cytokine production	137	15	4.09	0.00448
Mammalia	Group 1	GO:0010466	regulation of peptide activity	163	14	4.87	0.00039
Mammalia	Group 1	GO:0002429	immune response-activating cell surface receptor signaling pathway	164	14	4.9	0.00518
Mammalia	Group 1	GO:0032680	regulation of tumor necrosis factor production	133	14	3.97	0.01423

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Mammalia	Group 1	GO:0032640	tumor necrosis factor production	133	14	3.97	0.01423
Mammalia	Group 1	GO:0002705	positive regulation of leukocyte mediated immunity	113	14	3.38	0.03945
Mammalia	Group 1	GO:0050829	defense response to Gram-negative bacterium	66	13	1.97	6.1e-08
Mammalia	Group 1	GO:0002444	myeloid leukocyte mediated immunity	90	13	2.69	8.0e-05
Mammalia	Group 1	GO:0051091	positive regulation of DNA-binding transcription factor activity	227	13	6.78	0.01910
Mammalia	Group 1	GO:0002456	T cell mediated immunity	82	13	2.45	0.03948
Mammalia	Group 1	GO:0031241	regulation of cell killing	71	12	2.12	0.00420
Mammalia	Group 1	GO:0002695	negative regulation of leukocyte activation	148	12	4.42	0.01775
Mammalia	Group 1	GO:0002286	T cell activation involved in immune response	94	12	2.81	0.01940
Mammalia	Group 1	GO:0050866	negative regulation of cell activation	165	12	4.93	0.03509
Mammalia	Group 1	GO:0001818	negative regulation of cytokine production	225	12	6.72	0.03811
Mammalia	Group 1	GO:00830	defense response to Gram-positive bacterium	87	11	2.6	5.4e-05
Mammalia	Group 1	GO:0002275	myeloid cell activation involved in immune response	83	11	2.48	0.0097
Mammalia	Group 1	GO:0050909	sensory perception of taste	52	11	1.55	0.02888
Mammalia	Group 1	GO:0042102	positive regulation of T cell proliferation	85	10	2.54	0.00022
Mammalia	Group 1	GO:0043299	leukocyte degranulation	62	10	1.85	0.00050
Mammalia	Group 1	GO:0030101	natural killer cell activation	66	10	1.97	0.00092
Mammalia	Group 1	GO:0097696	cell surface receptor signaling pathway via STAT	139	10	4.15	0.00889
Mammalia	Group 1	GO:0016064	immunoglobulin mediated immune response	107	10	3.2	0.03212
Mammalia	Group 1	GO:0019724	myeloid cell activation involved in immune response	108	10	3.23	0.03362
Mammalia	Group 1	GO:0032760	B cell mediated immunity	84	9	2.51	0.00089
Mammalia	Group 1	GO:0006056	positive regulation of tumor necrosis factor production	47	9	1.4	0.00998
Mammalia	Group 1	GO:0050729	complement activation	122	9	3.64	0.01092
Mammalia	Group 1	GO:0051250	positive regulation of inflammatory response	124	9	3.7	0.01206
Mammalia	Group 1	GO:0050832	negative regulation of lymphocyte activation	48	9	1.43	0.01262
Mammalia	Group 1	GO:0002823	defense response to fungus	39	9	1.17	0.01741
Mammalia	Group 1	GO:0007259	negative regulation of adaptive immune response [...]	134	9	4	0.01917
Mammalia	Group 1	GO:0051092	cell surface receptor signaling pathway via JAK-STAT	136	9	4.06	0.02090
Mammalia	Group 1	GO:0002820	positive regulation of NF- κ B transcription factor activity	42	9	1.25	0.02592
Mammalia	Group 1	GO:0042100	negative regulation of adaptive immune response	76	9	2.27	0.02668
Mammalia	Group 1	GO:009620	B cell proliferation	58	9	1.73	0.03229
Mammalia	Group 1	GO:0035036	response to fungus	46	9	1.37	0.03998
Mammalia	Group 1	GO:0071887	sperm-egg recognition	98	9	2.93	0.04681
Mammalia	Group 1	GO:0007342	leukocyte apoptotic process	25	8	0.75	4.1e-07
Mammalia	Group 1	GO:0001580	fusion of sperm to egg plasma membrane involved in single fertilization	29	8	0.87	1.5e-06
Mammalia	Group 1	GO:0002218	detection of chemical stimulus involved in sensory perception of bitter taste	45	8	1.34	4.9e-05
Mammalia	Group 1	GO:0032649	activation of innate immune response	88	8	2.63	0.00475
Mammalia	Group 1	GO:0045576	regulation of type I interferon production	54	8	1.61	0.00632
Mammalia	Group 1	GO:0071346	type I interferon production	98	8	2.93	0.03990
Mammalia	Group 1	GO:0001959	cellular response to type I interferon	114	8	3.41	0.02094
Mammalia	Group 1	GO:00030317	regulation of cytokine-mediated signaling pathway	119	8	3.55	0.02628
Mammalia	Group 1	GO:0097722	flagellated sperm motility	119	8	3.55	0.02628
Mammalia	Group 1	GO:0002249	sperm motility	120	8	3.58	0.02746
Mammalia	Group 1	GO:0045576	male gonad development	121	8	3.61	0.02867
Mammalia	Group 1	GO:0046546	development of primary male sexual characteristics	124	8	3.7	0.03251
Mammalia	Group 1	GO:0060759	regulation of response to cytokine stimulus	128	8	3.82	0.03817
Mammalia	Group 1	GO:0060294	cilium movement involved in cell motility	35	7	1.05	6.6e-05
Mammalia	Group 1	GO:0007339	binding of sperm to zona pellucida	40	7	1.19	0.00016
Mammalia	Group 1	GO:0019731	antibacterial humor I response	54	7	1.61	0.00107
Mammalia	Group 1	GO:0032757	positive regulation of interleukin-8 production	60	7	1.79	0.00199
Mammalia	Group 1	GO:0032722	positive regulation of chemokine production	22	7	0.66	0.00357
Mammalia	Group 1	GO:0002251	organ or tissue specific immune response	78	7	2.33	0.00864
Mammalia	Group 1	GO:0098586	cellular response to virus	80	7	2.39	0.00988
Mammalia	Group 1	GO:0030593	neutrophil chemotaxis	43	7	1.28	0.01012
Mammalia	Group 1	GO:0043303	mast cell degranulation	103	7	3.08	0.03462
Mammalia	Group 1	GO:0002279	regulation of receptor signaling pathway via STAT	29	6	0.87	0.00018
Mammalia	Group 1	GO:1904892	regulation of defense response to virus by host	83	7	1.31	0.01141
Mammalia	Group 1	GO:0042825	regulation of dephosphorylation	45	7	1.34	0.01280
Mammalia	Group 1	GO:0042119	regulation of T-helper 1 type immune response	39	6	1.17	0.00097
Mammalia	Group 1	GO:0045071	neutrophil activation	44	6	1.31	0.01430
Mammalia	Group 1	GO:0002711	negative regulation of viral genome replication	44	6	1.31	0.00185
Mammalia	Group 1	GO:0002639	positive regulation of cell mediated immunity	44	6	1.31	0.00185
Mammalia	Group 1	GO:0042267	positive regulation of immunoglobulin production	44	6	1.31	0.00185
Mammalia	Group 1	GO:0032731	natural killer cell mediated cytotoxicity	46	6	1.37	0.00234
Mammalia	Group 1	GO:0060337	positive regulation of interleukin-1 beta production	52	6	1.55	0.00438
Mammalia	Group 1	GO:0030888	I-type interferon-mediated signaling pathway	54	6	1.61	0.00528

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Mammalia	Group 1	GO:0051293	establishment of spindle localization	54	6	1.61	0.00528
Mammalia	Group 1	GO:0032729	positive regulation of type II interferon production	62	6	1.85	0.01030
Mammalia	Group 1	GO:0010921	regulation of phosphatase activity	65	6	1.94	0.01285
Mammalia	Group 1	GO:0002720	positive regulation of cytokine production involved in immune response	66	6	1.97	0.01379
Mammalia	Group 1	GO:0002385	mucosal immune response	69	6	0.57	0.02179
Mammalia	Group 1	GO:2000106	regulation of leukocyte apoptotic process	75	6	2.24	0.02449
Mammalia	Group 1	GO:094625	regulation of receptor signaling pathway via JAK-STAT	78	6	2.33	0.02902
Mammalia	Group 1	GO:0032755	positive regulation of interleukin-6 production	83	6	2.48	0.03773
Mammalia	Group 1	GO:0061760	antifungal innate immune response	17	5	0.51	0.00011
Mammalia	Group 1	GO:0035458	cellular response to interferon-beta	21	5	0.63	0.00032
Mammalia	Group 1	GO:0048240	sperm capacitation	25	5	0.75	0.00075
Mammalia	Group 1	GO:0001914	regulation of T cell mediated cytotoxicity	25	5	0.75	0.00075
Mammalia	Group 1	GO:0002230	positive regulation of defense response to virus by host	27	5	0.81	0.00109
Mammalia	Group 1	GO:0002830	positive regulation of type 2 immune response	14	5	0.42	0.00192
Mammalia	Group 1	GO:0046596	regulation of viral entry into host cell	31	5	0.93	0.00208
Mammalia	Group 1	GO:0001774	microglial cell activation	35	5	1.05	0.00360
Mammalia	Group 1	GO:004632	canonical inflammasome complex assembly	38	5	1.14	0.00518
Mammalia	Group 1	GO:0006953	acute-phase response	41	5	1.22	0.00719
Mammalia	Group 1	GO:040374	antiviral innate immune response	42	5	1.25	0.00797
Mammalia	Group 1	GO:0001912	positive regulation of leukocyte mediated cytotoxicity	47	5	1.4	0.01273
Mammalia	Group 1	GO:0032720	negative regulation of tumor necrosis factor production	47	5	1.4	0.01273
Mammalia	Group 1	GO:0034113	heterotypic cell-cell adhesion	49	5	1.46	0.01508
Mammalia	Group 1	GO:1903556	negative regulation of tumor necrosis factor superfamily cytokine production	50	5	1.49	0.01635
Mammalia	Group 1	GO:0070288	regulation of lymphocyte apoptotic process	50	5	1.49	0.01635
Mammalia	Group 1	GO:0001961	positive regulation of cytokine-mediated signaling pathway	50	5	1.49	0.01635
Mammalia	Group 1	GO:0060760	positive regulation of cytokine-to cytokine stimulus	57	5	1.7	0.02732
Mammalia	Group 1	GO:0038034	signal transduction in absence of ligand	59	5	1.76	0.03114
Mammalia	Group 1	GO:0097192	extrinsic apoptotic signaling pathway in absence of ligand	59	5	1.76	0.03114
Mammalia	Group 1	GO:0034113	heterotypic cell-cell adhesion	60	5	1.79	0.03317
Mammalia	Group 1	GO:0071260	cellular response to mechanical stimulus	64	5	1.91	0.04210
Mammalia	Group 1	GO:0031640	killing of cells of another organism	64	5	1.91	0.04210
Mammalia	Group 1	GO:0061844	antimicrobial humoral immune response mediated by antimicrobial peptide	64	5	1.91	0.04210
Mammalia	Group 1	GO:0002227	innate immune response in mucosa	11	4	0.33	0.00022
Mammalia	Group 1	GO:035455	response to interferon-alpha	14	4	0.42	0.00062
Mammalia	Group 1	GO:0048245	eosinophil chemotaxis	17	4	0.51	0.00137
Mammalia	Group 1	GO:0072540	T-helper 17 cell lineage commitment	17	4	0.51	0.00137
Mammalia	Group 1	GO:0002717	positive regulation of natural killer cell mediated immunity	21	4	0.63	0.00314
Mammalia	Group 1	GO:0072740	positive regulation of interleukin-17 production	23	4	0.69	0.00443
Mammalia	Group 1	GO:0032740	positive regulation of macrophage activation	23	4	0.69	0.00443
Mammalia	Group 1	GO:0043032	interleukin-1-mediated signaling pathway	24	4	0.72	0.00519
Mammalia	Group 1	GO:0070498	pyroptotic inflammatory response	26	4	0.78	0.00697
Mammalia	Group 1	GO:0070269	T cell costimulation	27	4	0.81	0.00800
Mammalia	Group 1	GO:0031295	positive regulation of interleukin-17 production	28	4	0.84	0.00911
Mammalia	Group 1	GO:0019884	neutrophil mediated immunity	29	4	0.87	0.01033
Mammalia	Group 1	GO:0002446	negative regulation of endothelial cell apoptotic process	30	4	0.9	0.01164
Mammalia	Group 1	GO:2003032	GO:0070498 positive regulation of interleukin-2 production	30	4	0.9	0.01164
Mammalia	Group 1	GO:0032743	regulation of NLRP3 inflammasome complex assembly	30	4	0.9	0.01164
Mammalia	Group 1	GO:0002025	response to exogenous dsRNA	31	4	0.93	0.01306
Mammalia	Group 1	GO:0043330	regulation of activated T cell proliferation	32	4	0.96	0.01458
Mammalia	Group 1	GO:0046006	regulation of natural killer cell activation	32	4	0.96	0.01458
Mammalia	Group 1	GO:0032814	NLRP3 inflammasome complex assembly	33	4	0.99	0.01621
Mammalia	Group 1	GO:0044546	activated T cell proliferation	34	4	1.02	0.01795
Mammalia	Group 1	GO:0050798	cellular defense response	34	4	1.02	0.01795
Mammalia	Group 1	GO:0006968	negative regulation of production of molecular mediator of immune response	35	4	1.05	0.01980
Mammalia	Group 1	GO:0043331	response to dsRNA	36	4	1.08	0.02176
Mammalia	Group 1	GO:0002691	regulation of cellular extravasation	38	4	1.14	0.02604
Mammalia	Group 1	GO:2000351	regulation of interleukin-10 production	43	4	1.28	0.03879
Mammalia	Group 1	GO:0032613	interleukin-10 production	45	4	1.34	0.04473
Mammalia	Group 1	GO:0045624	positive regulation of T-helper cell differentiation	45	4	1.34	0.04473
Mammalia	Group 1	GO:0043666	regulation of phosphoprotein phosphatase activity	46	4	1.37	0.04788
Mammalia	Group 1	GO:1004894	positive regulation of receptor signaling pathway via STAT	46	4	1.37	0.04788
Mammalia	Group 1	GO:0038095	Fc-epsilon receptor signaling pathway	5	3	0.15	0.00025
Mammalia	Group 1	GO:0045959	negative regulation of complement activation, classical pathway	8	3	0.24	0.00133
Mammalia	Group 1	GO:0097527	cytolysis by host symbiont cells	8	3	0.24	0.00133
Mammalia	Group 1	GO:0006924	necrotic signaling pathway	10	3	0.3	0.00272
Mammalia	Group 1	GO:1001731	activation-induced cell death of T cells	10	3	0.3	0.00272
Mammalia	Group 1	GO:2000851	positive regulation of platelet aggregation	10	3	0.3	0.00272
Mammalia	Group 1	GO:0007343	regulation of T-helper 2 cell cytokine production	11	3	0.33	0.00365

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Mammalia	Group 1	GO:0035723	interleukin-15-mediated signaling pathway	11	3	0.33	0.00365
Mammalia	Group 1	GO:0050079	negative regulation of neuroinflammatory response	12	3	0.36	0.00476
Mammalia	Group 1	GO:0002323	natural killer cell activation involved in immune response	13	3	0.39	0.00606
Mammalia	Group 1	GO:0032197	retrotransposition	13	3	0.39	0.00606
Mammalia	Group 1	GO:0060046	regulation of acrosome reaction	13	3	0.39	0.00606
Mammalia	Group 1	GO:0072683	T cell extravasation	13	3	0.39	0.00606
Mammalia	Group 1	GO:0043306	positive regulation of mast cell degranulation	14	3	0.42	0.00754
Mammalia	Group 1	GO:0006957	complement activation, alternative pathway	14	3	0.42	0.00754
Mammalia	Group 1	GO:0050855	regulation of B cell receptor signaling pathway	14	3	0.42	0.00754
Mammalia	Group 1	GO:0023035	CD40 signaling pathway	14	3	0.42	0.00754
Mammalia	Group 1	GO:1093027	regulation of opsonization	14	3	0.42	0.00754
Mammalia	Group 1	GO:002710	negative regulation of T cell mediated immunity	15	3	0.45	0.00922
Mammalia	Group 1	GO:0043031	negative regulation of macrophage activation	15	3	0.45	0.00922
Mammalia	Group 1	GO:0046597	negative regulation of viral entry into host cell	16	3	0.48	0.01110
Mammalia	Group 1	GO:0002888	positive regulation of myeloid leukocyte mediated immunity	17	3	0.51	0.01318
Mammalia	Group 1	GO:0002827	positive regulation of type I helper 1-type immune response	18	3	0.54	0.01548
Mammalia	Group 1	GO:0045346	regulation of MHC class II biosynthetic process	18	3	0.54	0.01548
Mammalia	Group 1	GO:0042832	defense response to protozoan	19	3	0.57	0.01798
Mammalia	Group 1	GO:0040019	positive regulation of embryonic development	19	3	0.57	0.01798
Mammalia	Group 1	GO:0060340	negative regulation of type I interferon-mediated signaling pathway	19	3	0.57	0.01798
Mammalia	Group 1	GO:0060333	type II interferon-mediated signaling pathway	19	3	0.57	0.01798
Mammalia	Group 1	GO:0045342	MHC class II biosynthetic process	19	3	0.57	0.01798
Mammalia	Group 1	GO:0032753	positive regulation of interleukin-4 production	20	3	0.6	0.02069
Mammalia	Group 1	GO:0001916	positive regulation of T cell mediated cytotoxicity	20	3	0.6	0.02069
Mammalia	Group 1	GO:001562	response to protozoan	21	3	0.63	0.02362
Mammalia	Group 1	GO:1093001	negative regulation of viral life cycle	21	3	0.63	0.02362
Mammalia	Group 1	GO:0035821	modulation of process of another organism	21	3	0.63	0.02362
Mammalia	Group 1	GO:0002726	positive regulation of T cell cytokine production	22	3	0.66	0.02675
Mammalia	Group 1	GO:0042104	positive regulation of activated T cell proliferation	22	3	0.66	0.02675
Mammalia	Group 1	GO:0010743	regulation of macrophage derived foam cell differentiation	25	3	0.75	0.03742
Mammalia	Group 1	GO:0032673	positive regulation of interleukin-4 production	25	3	0.75	0.03742
Mammalia	Group 1	GO:0032633	interleukin-4 production	25	3	0.75	0.03742
Mammalia	Group 1	GO:0071354	cellular response to interleukin-6	26	3	0.78	0.04140
Mammalia	Group 1	GO:0035821	positive regulation of phosphatase activity	26	3	0.78	0.04140
Mammalia	Group 1	GO:1092019	regulation of cilium-dependent cell motility	28	3	0.84	0.04995
Mammalia	Group 1	GO:0070741	response to interleukin-6	28	3	0.84	0.04995
Mammalia	Group 1	GO:0060295	regulation of cilium movement involved in cell motility	28	3	0.84	0.04995
Mammalia	Group 1	GO:0042269	regulation of natural killer cell mediated cytotoxicity	28	3	0.84	0.04995
Mammalia	Group 1	GO:0097528	execution phase of necrotosis	2	2	0.06	0.00089
Mammalia	Group 1	GO:0010822	negative regulation of complement-dependent cytotoxicity	2	2	0.06	0.00089
Mammalia	Group 1	GO:0002726	regulation of cilium-dependent cell motility	2	2	0.06	0.00089
Mammalia	Group 1	GO:0042104	positive regulation of activated T cell proliferation	4	2	0.12	0.00513
Mammalia	Group 1	GO:0010743	regulation of natural killer cell mediated cytotoxicity	4	2	0.12	0.00513
Mammalia	Group 1	GO:0042269	immune response-inhibiting signal transduction	5	2	0.15	0.00839
Mammalia	Group 1	GO:1093060	positive regulation of T helper 2 cell differentiation	5	2	0.15	0.00839
Mammalia	Group 1	GO:0043313	regulation of neutrophil degranulation	6	2	0.18	0.01233
Mammalia	Group 1	GO:0040006	protein-DNA covalent cross-linking/repair	6	2	0.18	0.01233
Mammalia	Group 1	GO:0060545	positive regulation of necrotic process	6	2	0.18	0.01233
Mammalia	Group 1	GO:0002826	negative regulation of T-helper 1-type immune response	6	2	0.18	0.01233
Mammalia	Group 1	GO:0048006	antigen processing and presentation, endogenous lipid antigen via MHC class Ib	6	2	0.18	0.01233
Mammalia	Group 1	GO:0002765	immune response-regulation of response to type II interferon	6	2	0.18	0.01233
Mammalia	Group 1	GO:0045630	positive regulation of type II interferon-mediated signaling pathway	6	2	0.18	0.01233
Mammalia	Group 1	GO:0043313	antigen processing and presentation, exogenous lipid antigen via MHC class Ib	6	2	0.18	0.01233
Mammalia	Group 1	GO:0006559	regulation of interleukin-1-mediated signaling pathway	7	2	0.21	0.01693
Mammalia	Group 1	GO:0045341	MHC class I biosynthetic process	7	2	0.21	0.01693
Mammalia	Group 1	GO:0004647	negative regulation of fertilization	7	2	0.21	0.01693
Mammalia	Group 1	GO:0004648	prevention of polyspermy	7	2	0.21	0.01693
Mammalia	Group 1	GO:0023722	neutrophil apoptotic process	7	2	0.21	0.01693
Mammalia	Group 1	GO:0002733	myeloid dendritic cell cytokine production	7	2	0.21	0.01693
Mammalia	Group 1	GO:0002735	regulation of myeloid dendritic cell cytokine production	7	2	0.21	0.01693
Mammalia	Group 1	GO:0038196	positive regulation of myeloid dendritic cell cytokine production	8	2	0.24	0.02213
Mammalia	Group 1	GO:0043312	type III interferon-mediated signaling pathway	8	2	0.24	0.02213
Mammalia	Group 1	GO:1093089	positive regulation of blood vessel endothelial cell proliferation involved in sprouting angiogenesis	8	2	0.24	0.02213

Tab. S10 continued from previous page

Dataset	Group of genes	GO.ID	Term	Annotated	Significant	Expected	classicFisher
Mammalia	Group 1	GO:0006977	DNA damage response, signal transduction by p53 class mediator resulting in cell cycle arrest	8	2	0.24	0.02213
Mammalia	Group 1	GO:0038110	interleukin-2-mediated signaling pathway	8	2	0.24	0.02213
Mammalia	Group 1	GO:0002638	negative regulation of immunoglobulin production	8	2	0.24	0.02213
Mammalia	Group 1	GO:0032650	regulation of interleukin-1 alpha production	8	2	0.24	0.02213
Mammalia	Group 1	GO:0035771	interleukin-4-mediated signaling pathway	8	2	0.24	0.02213
Mammalia	Group 1	GO:0032610	interleukin-1 alpha production	8	2	0.24	0.02213
Mammalia	Group 1	GO:0071352	cellular response to interleukin-2	8	2	0.24	0.02213
Mammalia	Group 1	GO:2000853	positive regulation of T-helper 2 cell cytokine production	8	2	0.24	0.02213
Mammalia	Group 1	GO:0034342	response to type III interferon	8	2	0.24	0.02213
Mammalia	Group 1	GO:0044406	adhesion of symbiont to host	9	2	0.27	0.02789
Mammalia	Group 1	GO:0045625	regulation of T-helper 1 cell differentiation	9	2	0.27	0.02789
Mammalia	Group 1	GO:0033139	regulation of peptidyl-serine phosphorylation of STAT protein	9	2	0.27	0.02789
Mammalia	Group 1	GO:0061518	microglial cell proliferation	9	2	0.27	0.02789
Mammalia	Group 1	GO:0070669	response to interleukin-2	9	2	0.27	0.02789
Mammalia	Group 1	GO:0010891	positive regulation of mitochondrial membrane potential	9	2	0.27	0.02789
Mammalia	Group 1	GO:1902563	regulation of neutrophil activation	10	2	0.3	0.03419
Mammalia	Group 1	GO:0002430	adhesion of symbiont to host	10	2	0.3	0.03419
Mammalia	Group 1	GO:0044406	complement receptor mediated signaling pathway	10	2	0.3	0.03419
Mammalia	Group 1	GO:0032754	positive regulation of interleukin-5 production	10	2	0.3	0.03419
Mammalia	Group 1	GO:0010826	retrotransposon silencing	10	2	0.3	0.03419
Mammalia	Group 1	GO:0010828	regulation of transposition	10	2	0.3	0.03419
Mammalia	Group 1	GO:0010529	negative regulation of transposition	10	2	0.3	0.03419
Mammalia	Group 1	GO:1902563	regulation of neutrophil activation	10	2	0.3	0.03419
Mammalia	Group 1	GO:0050078	positive regulation of neuroninflammatory response	11	2	0.33	0.04097
Mammalia	Group 1	GO:0097011	cellular response to granulocyte macrophage colony-stimulating factor stimulus	11	2	0.33	0.04097
Mammalia	Group 1	GO:0097012	response to granulocyte macrophage colony-stimulating factor	11	2	0.33	0.04097
Mammalia	Group 1	GO:0002732	positive regulation of dendritic cell cytokine production	11	2	0.33	0.04097
Mammalia	Group 1	GO:0032736	positive regulation of interleukin-13 production	11	2	0.33	0.04097
Mammalia	Group 1	GO:1901857	positive regulation of neutrophil activation	11	2	0.33	0.04097
Mammalia	Group 1	GO:0061517	positive regulation of cellular respiration	11	2	0.33	0.04097
Mammalia	Group 1	GO:1900226	macrophage proliferation	11	2	0.33	0.04097
Mammalia	Group 1	GO:0051770	negative regulation of NLRP3 inflammasome complex assembly	11	2	0.33	0.04097
Mammalia	Group 1	GO:0032306	positive regulation of nitric-oxide synthase biosynthetic process	11	2	0.33	0.04097
Mammalia	Group 1	GO:0032308	regulation of prostaglandin secretion	12	2	0.36	0.04822
Mammalia	Group 1	GO:0042501	positive regulation of prostaglandin secretion	12	2	0.36	0.04822
Mammalia	Group 1	GO:0060330	serine phosphorylation of STAT protein	12	2	0.36	0.04822
Mammalia	Group 1	GO:0060334	regulation of response to type II interferon	12	2	0.36	0.04822
Mammalia	Group 1	GO:0018003	regulation of type II interferon-mediated signaling pathway	12	2	0.36	0.04822
Mammalia	Group 1	GO:0002384	peptidyl-lysine N-acetylation	1	1	0.03	0.02987
Mammalia	Group 1	GO:1905223	epicardium morphogenesis	1	1	0.03	0.02987
Mammalia	Group 1	GO:0002450	B cell antigen processing and presentation	1	1	0.03	0.02987
Mammalia	Group 1	GO:0002470	plasmacytoid dendritic cell antigen processing and presentation	1	1	0.03	0.02987
Mammalia	Group 1	GO:0072343	pancreatic stellate cell proliferation	1	1	0.03	0.02987
Mammalia	Group 1	GO:0010185	regulation of cellular defense response	1	1	0.03	0.02987
Mammalia	Group 1	GO:0010186	positive regulation of cellular defense response	1	1	0.03	0.02987
Mammalia	Group 1	GO:0098784	biofilm matrix organization	1	1	0.03	0.02987
Mammalia	Group 1	GO:0098786	biofilm matrix disassembly	1	1	0.03	0.02987
Mammalia	Group 1	GO:0030187	melatonin biosynthetic process	1	1	0.03	0.02987
Mammalia	Group 1	GO:0016068	melatonin metabolism	1	1	0.03	0.02987
Mammalia	Group 1	GO:0071660	positive regulation of IP-10 production	1	1	0.03	0.02987
Mammalia	Group 1	GO:0035397	cytoskeletal rearrangement involved in phagocytosis, engulfment	1	1	0.03	0.02987
Mammalia	Group 1	GO:0072139	helper T cell enhancement of adaptive immune response	1	1	0.03	0.02987
Mammalia	Group 1	GO:0140121	glomerular parietal epithelial cell differentiation	1	1	0.03	0.02987
Mammalia	Group 1	GO:0140122	Lewy body formation	1	1	0.03	0.02987
Mammalia	Group 1	GO:0140123	negative regulation of Lewy body formation	1	1	0.03	0.02987
Mammalia	Group 1	GO:0030186	melatonin metabolic process	1	1	0.03	0.02987
Mammalia	Group 1	GO:0030187	melatonin biosynthetic process	1	1	0.03	0.02987
Mammalia	Group 1	GO:0071660	positive regulation of IP-10 production	1	1	0.03	0.02987
Mammalia	Group 1	GO:0032759	cytoskeletal rearrangement of eosinophil chemotaxis	1	1	0.03	0.02987
Mammalia	Group 1	GO:0032723	positive regulation of connective tissue growth factor production	1	1	0.03	0.02987
Mammalia	Group 1	GO:1901251	positive regulation of lung goblet cell differentiation	1	1	0.03	0.02987
Mammalia	Group 1	GO:0032745	positive regulation of interleukin-21 production	1	1	0.03	0.02987
Mammalia	Group 1	GO:1901977	lysophosphatidyl transport	1	1	0.03	0.02987
Mammalia	Group 1	GO:1901247	negative regulation of lung ciliated cell differentiation	1	1	0.03	0.02987
Mammalia	Group 1	GO:1901249	regulation of lung goblet cell differentiation	1	1	0.03	0.02987
Mammalia	Group 1	GO:0070246	natural killer cell apoptotic process	1	1	0.03	0.02987
Mammalia	Group 1	GO:0070247	regulation of natural killer cell apoptosis	1	1	0.03	0.02987
Mammalia	Group 1	GO:1902310	positive regulation of peptidyl-serine dephosphorylation	1	1	0.03	0.02987

Tab. S10 continued from previous page

Dataset	Group of genes	GO ID	Term		Annotated	Significant	Expected	classicFisher	
Mammalia	Group 1	GO:2000229	regulation of pancreatic stellate cell proliferation		1	1	0.03	0.02987	
Mammalia	Group 1	GO:1904784	NLRP1 inflammasome complex assembly		1	1	0.03	0.02987	
Mammalia	Group 1	GO:2000231	positive regulation of pancreatic stellate cell proliferation		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0150140	positive regulation of CD36 production		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0150142	positive regulation of CD86 production		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0150143	regulation of CD80 production		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0150145	positive regulation of CD80 production		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0332665	regulation of interleukin-21 production		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0332679	regulation of TNFRL production		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0332679	interleukin-21 production		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0332639	TRAIL production		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0090320	regulation of chylomicron remnant clearance		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0090321	positive regulation of chylomicron remnant clearance		1	1	0.03	0.02987	
Mammalia	Group 1	GO:1904450	positive regulation of glutamate receptor signaling pathway		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0120042	negative regulation of macrophage proliferation		1	1	0.03	0.02987	
Mammalia	Group 1	GO:1903916	regulation of endoplasmic reticulum stress-induced eIF2 alpha dephosphorylation		1	1	0.03	0.02987	
Mammalia	Group 1	GO:1903917	positive regulation of endoplasmic reticulum stress-induced eIF2 alpha dephosphorylation		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0334496	regulation of translational initiation by eIF2 alpha dephosphorylation		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0334497	eIF2alpha dephosphorylation in response to endoplasmic reticulum stress		1	1	0.03	0.02987	
Mammalia	Group 1	GO:1901625	cellular response to estrogen		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0600006	FC receptor-mediated immune complex endocytosis		1	1	0.03	0.02987	
Mammalia	Group 1	GO:036153	triacylglyceride acyl-chain remodeling		1	1	0.03	0.02987	
Mammalia	Group 1	GO:036155	acylglycerol acyl-chain remodeling		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0046003	negative regulation of mitotic centrosome separation		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0056902	leukocyte adhesion activation		1	1	0.03	0.02987	
Mammalia	Group 1	GO:0341516	negative regulation of toll-like receptor 7 signaling pathway		1	1	0.03	0.02987	
Mammalia	Group 1	GO:1905151	negative regulation of voltage-gated sodium channel activity		1	1	0.03	0.02987	

Supplementary Table S11 – Accession numbers and general statistics of RNA-sequencing libraries of *Mytilus galloprovincialis* developmental stages.

SRA acc. no.	Time point (hpf)	Developmental stage	Biological replicate	No. of trimmed reads	No. of uniquely mapped reads	% of uniquely mapping reads	No. of uniquely mapped reads	% of uniquely mapped reads	No. of unmapped reads	% of unmapped reads
SRR25387458	0	Unfertilized Egg	R2	31,614,834	22,677,680	71.73%	4525340.00	14.31%	4,411,814	13.96%
SRR25387459	0	Unfertilized Egg	R1	31,229,282	22,930,909	73.43%	3631800.00	11.63%	4,666,573	14.94%
SRR25387436	4	Zygote	R2	31,573,942	22,238,537	70.43%	4747330.00	15.04%	4,588,075	14.53%
SRR25387447	4	Zygote	R1	31,215,045	22,657,854	72.59%	3980706.00	12.75%	4,576,485	14.66%
SRR25387434	8	Embryo	R2	31,679,197	22,361,698	70.59%	5224621.00	16.49%	4,092,878	12.92%
SRR25387435	8	Embryo	R1	31,137,192	22,231,464	71.40%	4558377.00	14.63%	4,347,351	13.96%
SRR25387432	12	Gastrula 1	R2	30,684,472	21,477,819	70.00%	5351506.00	17.44%	3,855,147	12.57%
SRR25387433	12	Gastrula 1	R1	31,006,745	22,158,048	71.46%	4808473.00	15.51%	4,040,224	13.03%
SRR25387430	16	Gastrula 2	R2	31,129,558	21,172,217	68.01%	5943113.00	19.10%	4,014,228	12.89%
SRR25387431	16	Gastrula 2	R1	31,108,790	22,366,338	71.90%	4797011.00	15.42%	3,945,441	12.68%
SRR25387456	20	Trochophore 1	R2	31,313,006	21,923,726	70.01%	5234475.00	16.71%	4,154,805	13.27%
SRR25387457	20	Trochophore 1	R1	31,153,366	22,818,864	73.25%	4258642.00	13.67%	4,075,860	13.08%
SRR25387454	24	Trochophore 2	R2	31,237,678	21,993,262	70.41%	5095540.00	16.32%	4,148,876	13.29%
SRR25387455	24	Trochophore 2	R1	30,568,998	22,102,616	72.30%	4376490.00	14.31%	4,089,892	13.38%
SRR25387452	28	Trochophore 3	R2	31,251,736	22,218,911	71.10%	4737140.00	15.16%	4,295,685	13.74%
SRR25387453	28	Trochophore 3	R1	30,151,120	21,916,806	72.69%	4155633.00	13.78%	4,078,681	13.53%
SRR25387450	32	Advanced Trophophore 1	R2	31,450,472	22,501,615	71.55%	4808549.00	15.29%	4,140,308	13.16%
SRR25387451	32	Advanced Trophophore 1	R1	30,344,616	22,168,913	73.06%	4088100.00	13.47%	4,087,603	13.47%
SRR25387448	36	Advanced Trophophore 2	R2	31,281,625	22,138,801	70.77%	5099606.00	16.30%	4,043,218	12.92%
SRR25387449	36	Advanced Trophophore 2	R1	30,446,461	22,282,130	73.18%	4164126.00	13.68%	4,000,205	13.13%
SRR25387445	40	Advanced Trophophore 3	R2	31,286,823	22,253,466	71.13%	4808187.00	15.37%	4,225,170	13.50%
SRR25387446	40	Advanced Trophophore 3	R1	30,427,358	22,021,178	72.37%	4139754.00	13.61%	4,266,426	14.02%
SRR25387443	44	D-veliger 1	R2	30,469,689	21,902,055	71.88%	4591610.00	15.06%	3,976,024	13.05%
SRR25387444	44	D-veliger 1	R1	30,486,877	22,346,398	73.30%	4184438.00	13.74%	3,953,041	12.96%
SRR25387441	48	D-veliger 2	R2	30,209,384	21,152,507	70.02%	5122804.00	16.95%	3,934,073	13.03%
SRR25387442	48	D-veliger 2	R1	30,492,559	22,356,764	73.32%	4132649.00	13.55%	4,003,146	13.12%
SRR25387439	52	D-veliger 3	R2	30,581,805	22,287,023	72.88%	4273235.00	13.98%	4,021,547	13.15%
SRR25387440	52	D-veliger 3	R1	30,540,062	22,508,657	73.70%	3938157.00	12.90%	4,093,248	13.40%
SRR25387437	72	Late D-veliger	R2	31,144,538	22,754,889	73.06%	4155579.00	13.35%	4,234,060	13.60%
SRR25387438	72	Late D-veliger	R1	31,504,212	23,356,674	74.14%	3830231.00	12.16%	4,317,307	13.70%

Supplementary Table S12 – Set of HCR probes generated with the ‘insitu_probe_generator’ script from the Ozpolat Lab (Kuehn et al., 2022).

Pool name	Sequence
B1_Mgal..10B017427..vasa..33..Dla0	GAGGAGGGCAGCAAACGgaaAAATGCTGTAAAAACAAAACGTATA TTAAAATGTTGAAAATGTTTAATAtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaAGAGTTCATCACGATTAAAGAAAGA ACTAAAAAACATTACATCATACAGAtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaGCACATGATTTAAGGTAGCACTACT AAACAATAAGAACTGGATTGATTtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaGACGAACCTCTCCAGACAGGCCG AAAAATAATGCCGCTTCCAATCAtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaTCAGCTGTTGGTTACTTGGTAAA CCAATAGTGATTACCTAAGTGAACAtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaGCAAGACATGGCCTGGTTATAGTT ATTATGCCCTAATGGTTTAGGTCGtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaACATTCCCAGTCTGAAGTCTGTTAG GTGACAACTGATTGATTATATGtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaCATTGAAGAATTATCTGCTGAGAGT TGACCCACTTTAGGTCAAACAtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaGCATTCAATGAAATGAGTGGAGT ACAATTCTGCTCCTCGCAGGTTAtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaAGAATGTGATTAATCCCAGGATTCC ATTTACAATGGTTGGAATAACAATGtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaTTGTCCTGGTTGGCTTTCTGAT CTCCGGAATCTCTGTGTTGGTTGTCtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaCTGAATCATCTGTTAGAAACTGGT GTAACAAAGCTTTGCAATTCCACCTaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaACATGTTAACAGAGGAATGTCATA ATAGATTGTTGAAAGTCATAATTAtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaGCAAGGAAGTCAGCATTCTTTCT GGAATCCAGACTGAGACAAGTAAGtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaATGCACATACTGTGCAACATCGTA TTGACGTTCTGATCACGGGAACCTtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaTTGCACTGAACATAAGAGTTGCT CAAGTCTCGGATTCTCTGGAtaGAAGACTTTCTTTAG GAGGAGGGCAGCAAACGgaaCTTCCCTTACCAATGACATCAATT TAAATATTTCAGCTTTCTAAACTGtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaCTACAGGTTAACATTGTGCCATG GTCCAAGAGGGTTCTCCATATAAtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaGAGAATGAACACTACCAGTCAGTCC ACTAAAGGCTGTGGCTCTGAACTAtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaGACTTTTACATTTAAAGAAAGG TGACAGGTGTTGGTCTATCAGTtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaTGCCTGTATGGTTTAAAATTCA TTTTTCATATTGTCAAAGTTTATGtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaGGGCCATATGTCCTTCTTATTACA AAAGACCTTCTGTTAGGACAATCtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaCACTTCTTACATTGAAGCAGTT CATTAGGACACTCTTGCCTGTGtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaCCTTGCCTCCACCACCTGATCCTC CCACTTCAACACATTGAAAGCAGTtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaCCTCCACCTTATTTCCTCAAATC CCCCCTTGTGCAAATCCACCATtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaAGATATGCTCATGTCCTTACCCCT AGGCTTCTCCTAAAAGCAGGGGTtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaTCCCTGAGCACGGCTCTACTTGGT TCTTCCAAACCCACCAGCCCCAAAtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaATTGACTGATTTAAGGTGTTGTT ATAACAATTTCATCCTCCAGGACTtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaACTTGTAAAATTTCATCAACCACGT AACTAAACTAAACATGCAAATTCTCAtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaGTGAAAACAAACGAATGTTAAAATT AAAAATATGGTACATGAACCTCAAtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGgaaTTGAAACTTTAGAACAAATTATA

Table S12 continued from previous page

Pool name	Sequence
B2_Mgal_10B093608_dmrt1l_32_Dla0	ACAGTGTAACTTTCTGCATGCCtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGGAaGTCAGTTATCTTCTTTCAAACAG CTTCTACTGCTGTATTGAAATATtaGAAGAGTCTTCCTTTAG GAGGAGGGCAGCAAACGGAaCAGACAGTTCCCTGCCTTTCTC TGTGTAGCCCCCTCAACCCCTTCGtaGAAGAGTCTTCCTTTAG CCTCGTAATCCTCATCaaaaaaaaggggggtattttttaaa AAAAAATAATGTCGATTTCAattaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaAACAAAGCTCTCAATATGATATATC taaccaCTTCCAGATATAACTTGAGGaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaAAGTAACACAgctaaaaattaaga CAGTTGAACCTTATTCCATACCTTaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCAattataacaattataacatTGACTG ttataccTCCCTCATCTTGATCCAaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaAAATCACCCCTGGTGGTGACAAA ACACATATTCCAACCTTATTGGaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaATGAATGTTGATAATCAAAGTACT CTATGATTATCATCAAGTATCATTaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaatcatgatatagggATATGAAGT TCAGACAAAGCATTGATCtttggaaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaTAACCTGTTGAAACATCCCTT TGACCTGGACTTTGCAAACCAAAATaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaaacattttttttgttcTCCGTA ACGGGCACCAAAACATGTCATTtcaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaATTTCAGACTCACATTCTATGT atttaaaacattatgttGTCACCTaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaatttttttttttttttttttttttt TTAAGACACACTATCTGGCTTGaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaACAGACAGaaacttttgatata TTTCAATGCAATTAAAGTTCTATAaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaCATGCAGACAGACACTCTAACAT tggatgtAAAAATTACAGACTCAaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaTCATTATTACTTCAAAGGTGTGC TCACCTCACCTCAACAAATAGCAAGaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaTTTGAATTGTTGtttttttttttt CTATAATTATTACCCCTGAAATTGaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaTTTCTCTTATGACAACATATCTT aatatttttagaaCATAAAAATAaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaacttttttttttttttttttttt TCTTGATTTAACATTtatcatcaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaAAATTCTACATTtgccatgtat acttt CCTCGTAATCCTCATCaaaAATGAAATATGCACTAATCaaaa tttaatt CCTCGTAATCCTCATCaaaTTCTCAAATATGGTGCCTTAAATCC tagtaatgttaATTGTAGCTTTGtaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaATGCATGACTACAGAGATTAAACT GAAAGTTCTAACAGATTCAATTAAaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaAGCCACGACTACAAAATTAAGTTTC AAAAACGTTCTCTTCTATCTTGtaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaGTACAGTTGAATGTGGAATATCATG CTTGTGCTAGTGGAGTTCCATAaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaATTGTACGGCATACTTGGTGTGGCA TGGTCCCTGCATATATCTGGGGTTaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaTGTAACAGTTGGTGGATAAAATCCA TGATGGTGGGTATGGTGTGGGATAaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaATTATTCTACACTCTGTGACCAAC TGGCTACACAGAGGACTGACAATaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaCTCGCACTAGATTTAACACCAGT CTCTGTATGGACAATCTCTGTGaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaTCTGTCAGATATTGGATTTGCTGT TTGACATCTACGGCAATACAGCTaaATCATCCAGTAAACCGCC CCTCGTAATCCTCATCaaaatttttttttttttttttttttttt TTCTTCTCTATTGCTTgttctgttaATCATCCAGTAAACCGCC

Table S12 continued from previous page

Pool name	Sequence
	CCTCGTAAATCCTCATCAAaCTGGCGTTAATACTAACACCTGA TTTTGAGCAGTAACTATCTTGTTaaATCATCCAGTAAACCGCC CCTCGTAAATCCTCATCAAaATCTAACATGGACACCAGAGCAAG AAAAACATCCAAGCTGTGACCTTaaATCATCCAGTAAACCGCC CCTCGTAAATCCTCATCAAaTGGCGGAATTGACGTTACTTC aaattacAACTTCTTCTTCTCTTaaATCATCCAGTAAACCGCC
	GTCCCCTGCCCTATATCTttCTTTCTCTTGAGACATCCACATT ATTAGTTTATCGCTTCCCTTttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttCTTGAGACATCCACATTAGTTTAT TTATCGTCTTCCCTTCTTCTTttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttCATCACATTAGTTATCGTCTT TTCTCCCTTTCTCTTCTTCTTGAAttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttATTTAGTTATCGTCTTCCCTT CTTTCTTCTTCTTGAAGACATCCttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttATATGCTCTACTAAAGACCTAGAGA TTTTCTACAGAAGTAGGTACAATttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttTTCTCTTCTTGAAGACATCCAC CAACATTAGTTATCGTCTTCCttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttTTCTCTTGAGACATCCACATTAGT TAGTTTATCGTCTTCTCCCTTCTtCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttGAGACATCCACATTAGTTATCGT TCGTCTTCTCCCTTCTTCTtCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttACATGTCGTCACGTGAGATTCTG CAATGGTATCATCTATTGGACCCCTtCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttGTCTCTGGATGTTCTCTCAGC TGCTGGGGAAACGGGAACGGCAAAttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttGCAAGCTCTGAATCATCGGTAG GTCTGACTCCGGGAAGAGTGGTAGttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttCTGTTCTTGAAGA TGCTGTGCTAAAGTGGCTATGttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttTCTTAGCTTTATCCTACTGG TTTTCTTGTCTCTGTGCTTCTtCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttGGGTGCTTGTGATCTTCTGACAT GTCAAGTATTGTCATAAGCAGACGTCttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttGCATTACAGGCTTGCTTCCAAATG TCATTGCTTTTCTTCTGttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttGACCACAGGCGCTACCCCTCGGA TGCTGGAAACAGCTTAAAGTCTCTCAattCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttTGATTATTGTGACATTGTCAGG GGTATTGAAACAGGAACGGTTGGttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttATCTGGAAACGTCTTGAAGAAG TGTAGTCAGGTATTCCCTGCTTGTGttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttGAATGGCTGTCGTACATGGGATAT ATTTcaTTCAAGAAATTCTGGCttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttGGACAGACAAATTGGTTACCTTGAG ATTGGTTGAACGATTGCTAGTCCTGttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttCATTACGGGAAATTGGTCTGTT GCATCGTTACAGGTTGAATAGGCAttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttATCCAGTACCTGGCATGAGAGTAG GAACTTTGACCAAGGTGTTCTATACGttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttTGTGACATTCCCTGAGGGTT tCCATCTGCTGACATAAAAGAAAATCttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttGCATTGCAAGTATCGGCTTGT GCTTGAAATGGTGTGCAACATTCAattCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttTTTAACTGTTCAATACTCCGTCC CAGGTTGGGTGAATGACATACAACTtCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTtttttttttttttttttttttttttt TTCTGTGTTAAATTAGccgaatttttCCACTCAACTTTAACCG GTCCCCTGCCCTATATCTttTTATCAGATGAAACATGTTCTGAA CCGGCGATAAGTTGCTGATTTtCCACTCAACTTTAACCG
B3_Mgal_10B014180_soxy_27_Dla0	CCTCAACCTACCTCCAACaagaaaaaaaataacaataatataat TTGCATGGTAAGAAATTGcccttaatTCTCACCATATTGCGCTTC

Table S12 continued from previous page

Pool name	Sequence
	CCTCAACCTACCTCCAACaaTAATTTCTTGTAGGCTCATAA ttaaaatataaaaaattctCGAtatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaCATTAGGTATGATACAATATCTCA TTTTCTGGCACAGAACATGTGACACatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaTCCAGTAGGTATAATGCACCGCCTC ACTGTATGGCAATTATTTACCTCTCatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaACTGGAGCCCCATGAACCTGAGTCG TTGTTGTCGTGAAGGAAAGGAAATatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaTTCACTTGATGGCTGATTGATTG AACTGGGAAATGTTGGTCCAGGATatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaCCGGTGGGGAACACGGGACTGGTT TATAACTACAGGCCGCTAAACTGGGatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaTGGAGATGCGAGAATTGGTCCAGA ACTGtataactgttattttgtACatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaTACGCTGGGGTGGAAAAATACGGTT CACGGCGAATATTGTGAATACTGAGatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaAGTGTCCGTCACAAATAATGGTT TTAAAGAAAATTGGTTGTATGGGAGatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaACGCATTCTCGTCACGTCGGTAG GAGAGATATCGATGCTCGTATGGTatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaGGGTCTAAAGTCAAATAATTCCCTT CCTTCTCAAACATGTCTCAAATGatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaTTACAAAACATTCAATTCACTAGCTT GTTCTCCCCACCTTCTTGGGACatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaCTTATTTCTCATAGTAGGGAAAT GTGACGAATACTATTGCCCACCTatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaAGTGTAACTCTTTATCATCGCATT TTCATGATATATTGATAAATTCCACatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaAAGAATATGGTGGTTGACATCGGG TAATTGCCATAGCAATAAGAGCAACatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaTTTGTCTCTTCTGGTTGTTT TTTTTCAACGGTTTATTACAGATTatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaCATTCTGTTGCTACTATTCTCCAT TCTTATCTCAAAGTAGATTATTatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaTTAACATGTTCAAGGAATTGAGGT TGCAGCGATTCCACATTTCGCTAatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaCTGAATTGGGATTCGAGAGCGGT CACATTTCGAGTCCATAAGTTTatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaCCGTCACACTGTatctaaatttg GTCTTAATTAAAATCCAAATCTGAatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaataatatttagATTACCAACACGC tatattcatgtttccatatttgtaatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaCATCGATTATtttttttgtgaa TCATAATGAAGTTATAatTGTCTTatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaTAACCTCTCATTTGTTGATTG CTCAGGTTTAGTGAACCTCAAACatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaTAGTCACCAATGTATTCTCATGG TTTCTCTCTTTAATTCTGTatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaCACTTCTCAGGATCTTAAACCTT GTCTTTCTTACTAAAGGTGACTatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaTCAAACCTATTGGCTGATCTGTAT TGTCTGATACACCAGGACAACCTCAatTCTCACCATATTGCTTC CCTCAACCTACCTCCAACaaCGTTGCTTGCTTTCTTGGTGG CGTTGTTGGTTGATTATTGCTTatTCTCACCATATTGCTTC

Supplementary Table S13 – Set of Vasa/Ddx4 and Ddx3 sequences used in the phylogenetic analysis. For each sequence, the species, the gene accession number (Gene ID), the orthology group and the gene names for reference species are shown. Reference species are marked with an asterisk.

Species	Gene ID	Orthology group	Gene name
<i>Phreagena okutanii</i>	DN44424.c0.g1.i2.p1	Vasa/Ddx4	-
<i>Calyptogena (Archivesica) marissinica</i>	Ama38729	Vasa/Ddx4	-
<i>Calyptogena (Archivesica) marissinica</i>	Ama38727	Vasa/Ddx4	-
<i>Corbicula fluminea</i>	DN125059.c0.g1.i19.p1	Vasa/Ddx4	-
<i>Mactra chinensis</i>	DN48157.c2.g2.i2.p1	Vasa/Ddx4	-
<i>Ruditapes decussatus</i>	DN22317.c4.g2.i1.p1	Vasa/Ddx4	-
<i>Ruditapes philippinarum</i>	XP_060562671.1	Vasa/Ddx4	-
<i>Mercenaria mercenaria</i>	XP_053394752.1	Vasa/Ddx4	-
<i>Cyclina sinensis</i>	Hic.asm.11.970.2	Vasa/Ddx4	-
<i>Pisidium coreanum</i>	DN31082.c0.g1.i7.p1	Vasa/Ddx4	-
<i>Dreissena polymorpha</i>	XP_052283635.1	Vasa/Ddx4	-
<i>Mya arenaria</i>	XP_052775885.1	Vasa/Ddx4	-
<i>Sinonovacula constricta</i>	Chr8.1697	Vasa/Ddx4	-
<i>Solen grandis</i>	DN2375.c0.g1.i12.p1	Vasa/Ddx4	-
<i>Tridacna squamosa</i>	DN1975.c0.g1.i5.p1	Vasa/Ddx4	-
<i>Panopea generosa</i>	DN2386.c0.g1.i4.p1	Vasa/Ddx4	-
<i>Cristaria plicata</i>	DN71694.c8.g1.i1.p1	Vasa/Ddx4	-
<i>Hyriopsis bialata (Unio delphinus)</i>	M00000006703	Vasa/Ddx4	-
<i>Potamilus streckersoni</i>	KAK3601505.1	Vasa/Ddx4	-
<i>Margaritifera margaritifera</i>	MMAM00000026330	Vasa/Ddx4	-
<i>Perna viridis</i>	s01977g89	Vasa/Ddx4	-
<i>Perumytilus purpuratus</i>	DN96437.c0.g1.i2.p1	Vasa/Ddx4	-
<i>Mytilus galloprovincialis</i>	VDI03911.1	Vasa/Ddx4	-
<i>Mytilus galloprovincialis</i>	VDI03912.1	Vasa/Ddx4	-
<i>Modiolus modiolus</i>	DN179.c0.g1.i2.p1	Vasa/Ddx4	-
<i>Argopecten irradians concentricus</i>	Contig172.33	Vasa/Ddx4	-
<i>Pecten maximus</i>	XP_033738807.1	Vasa/Ddx4	-
<i>Patinopecten yessoensis</i>	XP_021370692.1	Vasa/Ddx4	-
<i>Ostrea edulis</i>	XP_056020028.1	Vasa/Ddx4	-
<i>Magallana (Crassostrea) gigas</i>	XP_011437246.2	Vasa/Ddx4	-
<i>Crassostrea virginica</i>	XP_022316564.1	Vasa/Ddx4	-
<i>Saccostrea glomerata</i>	Sgl001349	Vasa/Ddx4	-
<i>Pinctada margaritifera</i>	DN36893.c1.g3.i1.p1	Vasa/Ddx4	-
<i>Atrina pectinata</i>	DN813.c0.g1.i1.p1	Vasa/Ddx4	-
<i>Tegillarca granosa</i>	KAJ8305640.1	Vasa/Ddx4	-
<i>Anadara (Scapharca) broughtonii</i>	EVM0008860.1	Vasa/Ddx4	-
<i>Drosophila melanogaster*</i>	NP_001260458.1	Vasa/Ddx4	vasa
<i>Homo sapiens*</i>	NP_077726.1	Vasa/Ddx4	DDX4
<i>Mus musculus*</i>	NP_001139357.1	Vasa/Ddx4	Ddx4
<i>Danio rerio*</i>	NP_571132.1	Vasa/Ddx4	ddx4
<i>Caenorhabditis elegans*</i>	NP_491876.1	Vasa/Ddx4	glh-2
<i>Caenorhabditis elegans*</i>	NP_491963.1	Vasa/Ddx4	glh-1
<i>Caenorhabditis elegans*</i>	NP_491681.1	Vasa/Ddx4	glh-3

Table S13 continued from previous page

Species	Gene ID	Orthology group	Gene name
<i>Caenorhabditis elegans</i> *	NP_491207.3	Vasa/Ddx4	<i>glh-4</i>
<i>Magallana (Crassostrea) gigas</i>	XP_011446924.2	Ddx3	-
<i>Magallana (Crassostrea) gigas</i>	XP_034330003.1	Ddx3	-
<i>Crassostra virginica</i>	XP_022337075.1	Ddx3	-
<i>Ostrea edulis</i>	XP_056006193.1	Ddx3	-
<i>Saccostrea glomerata</i>	Sgl003232	Ddx3	-
<i>Atrina pectinata</i>	DN371.c0.g4.i2.p1	Ddx3	-
<i>Pinctada margaritifera</i>	DN39745.c0.g1.i3.p1	Ddx3	-
<i>Perumytilus purpuratus</i>	DN34627.c0.g1.i16.p1	Ddx3	-
<i>Perna viridis</i>	s00037g119	Ddx3	-
<i>Mytilus galloprovincialis</i>	VDI00208.1	Ddx3	-
<i>Modiolus modiolus</i>	DN49076.c0.g1.i10.p1	Ddx3	-
<i>Tegillarca granosa</i>	KAJ8310842.1	Ddx3	-
<i>Argopecten irradians concentricus</i>	Contig829.57.3	Ddx3	-
<i>Pecten maximus</i>	XP_033759680.1	Ddx3	-
<i>Patinopecten yessoensis</i>	XP_021341010.1	Ddx3	-
<i>Hyriopsis bialata (Unio delphinus)</i>	M00000003015	Ddx3	-
<i>Cristaria plicata</i>	DN67742.c10.g2.i2.p1	Ddx3	-
<i>Megalonaia nervosa</i>	g136014.t1	Ddx3	-
<i>Potamilus streckersoni</i>	KAK3605786.1	Ddx3	-
<i>Margaritifera margaritifera</i>	MMAM00000009046	Ddx3	-
<i>Ruditapes decussatus</i>	DN22481.c1.g4.i1.p1	Ddx3	-
<i>Ruditapes philippinarum</i>	XP_060588962.1	Ddx3	-
<i>Cyclina sinensis</i>	Hic.asm.6.43.1	Ddx3	-
<i>Calyptogena (Archivesica) marissinica</i>	Ama32770	Ddx3	-
<i>Phreagena okutanii</i>	DN58569.c0.g1.i6.p1	Ddx3	-
<i>Mactra chinensis</i>	DN49476.c41.g3.i1.p1	Ddx3	-
<i>Corbicula fluminea</i>	DN126815.c0.g1.i7.p1	Ddx3	-
<i>Dreissena polymorpha</i>	XP_052217061.1	Ddx3	-
<i>Mya arenaria</i>	XP_052782518.1	Ddx3	-
<i>Pisidium coreanum</i>	DN3392.c0.g2.i6.p1	Ddx3	-
<i>Sinonovacula constricta</i>	Chr9.1187	Ddx3	-
<i>Sinonovacula constricta</i>	Chr9.1230	Ddx3	-
<i>Solen grandis</i>	DN51.c4.g1.i12.p1	Ddx3	-
<i>Panopea generosa</i>	DN13909.c0.g3.i2.p1	Ddx3	-
<i>Tridacna squamosa</i>	DN33643.c0.g2.i2.p1	Ddx3	-
<i>Danio rerio</i> *	NP_001119895.1	Ddx3	<i>ddx3xa</i>
<i>Danio rerio</i> *	NP_571016.2	Ddx3	<i>ddx3xb</i>
<i>Mus musculus</i> *	NP_149068.1	Ddx3	<i>Pl10</i>
<i>Homo sapiens</i> *	NP_001180346.1	Ddx3	<i>DDX3X</i>
<i>Drosophila melanogaster</i> *	NP_001262379.1	Ddx3	<i>bel</i>
<i>Pisidium coreanum</i>	DN29220.c0.g1.i2.p1	Ddx3	-
<i>Caenorhabditis elegans</i> *	NP_001021793.1	Ddx3	<i>vbh-1</i>
<i>Caenorhabditis elegans</i> *	NP_001254859.1	Ddx3	<i>laf-1</i>
<i>Pisidium coreanum</i>	DN19615.c0.g1.i2.p1	Ddx3	-

Data availability

All the supplementary figures and supplementary tables, as well as high-resolution figures, will be accessible online at my GitHub personal page.

Activity report

This is the report of the activities carried out during my 3-year PhD course (2021–2024).

Research activity

Here are the research activities not directly related to the main topic of the PhD thesis.

- Manual curation of long interspersed nuclear element (LINE) libraries of several bivalve species;
- comparative genomics analysis of Hox and ParaHox genes in brachiopod crustaceans;
- comparative genomics analysis of brachiopod crustaceans to investigate the molecular underpinnings of morphological stasis and genome size variations;
- molecular phylogenetics and Bayesian dating of brachiopod crustaceans;
- preparation of mRNA sequences of genes involved in body segmentation in *Triops cancriformis* (Pancrustacea, Branchiopoda), to be used to generate probes for mRNA *in-situ* HCR on larvae (in collaboration with the Patel Lab; Marine Biology Lab, Woods Hole, MA, USA);
- collection, fixation, and storing of juvenile stages of several stick insect (Insecta, Phasmida) species, to be used for mRNA *in-situ* HCR to investigate the temporal and spatial transcription of genes involved in wing morphogenesis (in collaboration with the Patel Lab; Marine Biology Lab, Woods Hole, MA, USA);
- preparation of a review on the evolutionary causes and consequences of trait loss reversals;
- preparation of mitotic chromosome plates in the red wood ant *Formica paralugubris* from cerebral ganglia of pre-pupae.

Visiting scholar

- Nuzhdin Lab (University of Southern California, Los Angeles, CA, UA; Aug 20, 2023–Feb 20, 2024), to accomplish the abroad period of my PhD;
- Juan Pasantes' lab (University of Vigo, Vigo, Spain; Jan 12–22, 2023), for a specific training on chromosome mitotic plate preparation in bivalve species.

Teaching activity

- Practical class "CAFE: estimating gene family turnover across a phylogenetic tree" (Apr 23, 2024) for first-year students of the course "Molecular phylogenetics" pursuing a Master degree in "Bioinformatics" at the University of Bologna (Italy);
- Practical invertebrate zoology class (Sep 2022–Jan 2023) for first-year students pursuing a Bachelor degree in "Biological Sciences" at the University of Bologna (Italy).

Co-supervised thesis

- *Evaluation of different calibration methods on Branchiopoda (Crustacea) phylogeny.* Niccolò Righetti. Master degree in "Biodiversità ed evoluzione", University of Bologna, Bologna (Italy). Supervisor: Andrea Luchetti. Co-supervisor: Filippo Nicolini. AA 2022/2023;
- *Filogenesi molecolare di alcune famiglie dell'ordine Phasmatodea con enfasi sulla famiglia Heteropterygidae (Bacilloidea).* Giacomo Orsini. Bachelor degree in "Scienze biologiche", University of Bologna, Bologna (Italy). Supervisor: Andrea Luchetti. Co-supervisor: Simona Corneti, Filippo Nicolini. AA 2021/2022;
- *Filogenesi molecolare di specie appartenenti alle famiglie Heteropterygidae e Anisacanthidae (Phasmatodea, Bacilloidea).* Alessandro Siragusa Camacho. Bachelor degree in "Scienze biologiche", University of Bologna, Bologna (Italy). Supervisor: Andrea Luchetti. Co-supervisor: Simona Corneti, Filippo Nicolini. AA 2021/2022;
- *Filogenesi molecolare di specie della famiglia Pseudophasmatidae.* Giovanni Amedeo Paselli. Bachelor degree in "Scienze biologiche", University of Bologna, Bologna (Italy). Supervisor: Barbara Mantovani. Co-supervisors: Simona Corneti, Filippo Nicolini. AA 2020/2021.

Courses and workshops

- *Establishing state-of-the-art mollusc genomics.* EMBO Workshop. Namur, Belgium. May 28–31, 2024;
- *Art (Science) Attack.* Physalia Courses. Online. May 20–23, 2024;
- *Introduction to Python for biologists.* Physalia Courses. Online. Sep 25–28, 2023;
- *ITA *PHY phylogenetics workshop.* Trento, Italy. Jun 6–9, 2023;
- *Sex chromosome evolution.* Physalia Courses. Online. Jan 23–27, 2023.

Awards and scholarships

- Travel grant to attend the "Evoluzione2024" congress in Naples (Italy). Stazione Zoologica Anton Dohrn. Sep 8–11, 2024;
- Travel grant to attend the EMBO workshop *Establishing state-of-the-art mollusc genomics* in Namur (Belgium). EMBO. May 28–31, 2024;
- Laura Bassi scholarship for editorial assistance to postgraduates and junior academics. Editing Press. Apr 13, 2023.

Presentations at congresses

Oral presentations

- Nicolini F, Iannello M, Piccinini G, Ghiselli F, Luchetti A, Milani L. (2024). Advancing the study of bivalve sex determination in the light of comparative genomics. Establishing state-of-the-art mollusc genomics (EMBO workshop). Namur (Belgium). May 27-30, 2024;
- Nicolini F, Ghiselli F, Milani L, Luchetti A. (2023). Contrasting patterns of amino acid evolution and shared ancestry between putative sex-determining genes in bivalve molluscs. EVOLMAR 2023. Online. Nov 14-17, 2023;
- Nicolini F, Ghiselli F, Milani L, Luchetti A. (2023). Sex-determination related genes in bivalves: novel acquisitions and high rates of sequence evolution. Evolution 2023 (Ernst Mayr Award symposium). Online. Jun 2-3, 2023.

Poster presentations

- Nicolini F, Iannello M, Piccinini G, ghiselli F, Nuzhdin S, Luchetti A, Milani L. (2024). How to detect sex-determining genes through molecular evolution: bivalves a a case study. Evoluzione 2024. Naples, Italy. Sep 8–11, 2024;
- Nicolini F, Ghiselli F, Milani L, Luchetti A. (2022). Clues of accelerated molecular evolution in gene families associated wit sex determination in bivalves. SMBE 2023. Ferrara, Italy. Jul 24–27, 2023;
- Nicolini F, Ghiselli F, Milani L, Luchetti A. (2022). Clues of accelerated molecular evolution in gene families associated wit sex determination in bivalves. SIBE/ISEB 2022. Ancona, Italy. Sep 4–7, 2022;

- Nicolini F, Martelossi J, Forni G, Mantovani B, Luchetti A. (2021) First insights and comparative genomics of Hox and ParaHox genes in tadpole shrimps. EuroEvoDevo 2022. Naples, Italy. May 31–Jun 3, 2022.

Invited talks

- *From comparative genomics to fluorescence imaging: a multi-disciplinary approach to study bivalve sex determination.* Auer Lab. University of Fribourg, Fribourg. Jul 26, 2024.

Outreach activity

- Editor and web writer for BioPills – the Italian community of life sciences (biopills.net/). Jul 2017–ongoing;
- Presenter at the European Researchers' Night 2024, University of Bologna, Bologna (Italy). Sep 27, 2024;
- Presenter at the BiGeA Day 2023, University of Bologna, Bologna (Italy). May 27, 2023;
- Opening Days, University of Bologna, Bologna (Italy). Nov 18, 2022.

Scientific publications

* equal contribution

- Righetti N*, Nicolini F*, Forni G, & Luchetti A. (2024). Towards a time-tree solution for Branchiopoda diversification: a jackknife assessment of fossil age priors. *Submitted for peer-review*
- Nicolini F, Ghiselli F, Luchetti A, & Milani L. (2023). Bivalves as emerging model systems to study the mechanisms and evolution of sex determination: a genomic point of view. *Genome Biology and Evolution*, 15(10), evad181. doi: 10.1093/gbe/evad181
- Martelossi J, Nicolini F, Subacchi S, Pasquale D, Ghiselli F, & Luchetti A. (2023). Multiple and diversified transposon lineages contribute to early and recent bivalve genome evolution. *BMC Biology*, 21(1), 1–23. doi: 10.1186/s12915-023-01632-z
- Nicolini F, Martelossi J, Forni G, Savojardo C, Mantovani B, & Luchetti A. (2023). Comparative genomics of Hox and ParaHox genes among major lineages of Branchiopoda with emphasis on tadpole shrimps. *Frontiers in Ecology and Evolution*, 11, 23. doi: 10.3389/fevo.2023.1046960

- Forni G, Cussigh A, Brock PD, Jones BR, Nicolini F, Martelossi J, Luchetti A, & Mantovani B. (2023). Taxonomic revision of the Australian stick insect genus *Candovia* (Phasmida: Necrosciinae): insight from molecular systematics and species-delimitation approaches. *Zoological Journal of the Linnean Society*, 197(1), 189–210. doi: 10.1093/zoolinnean/zlac074