

BIOIMAGING
FINAL PROJECT

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05/14/2014

Introduction

Tomography imaging is the process where image slices of the object of interest are taken. These slices can then be reconstructed to produce a full image representing the original object.

Different kinds of tomography techniques are:

Physical Phenomenon	Type of tomogram
X-rays	CT
Gamma rays	SPECT
Radio Frequency waves	MRI
Electron positron annihilation	PET
Electron	Electron tomography

Computed Tomography (CT) is a widely used imaging service which has been in used for medical purposes since the early 1970s. The term *tomography* comes from the Greek words *tomos* (a cut, a slice, or a section) and *graphein* (to write or record). CT is used for a variety of medical diagnosis, such as cancer, circulatory (blood) system diseases and conditions, such as coronary artery disease (atherosclerosis), blood vessel aneurysms, and blood clots; spinal conditions; kidney and bladder stones; abscesses; inflammatory diseases, such as ulcerative colitis and sinusitis; and injuries to the head, skeletal system, and internal organs. CT can be a life-saving tool for diagnosing illness and injury in both children and adults [1].

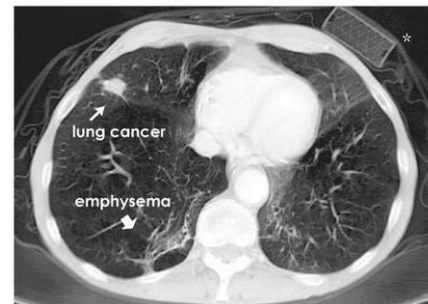
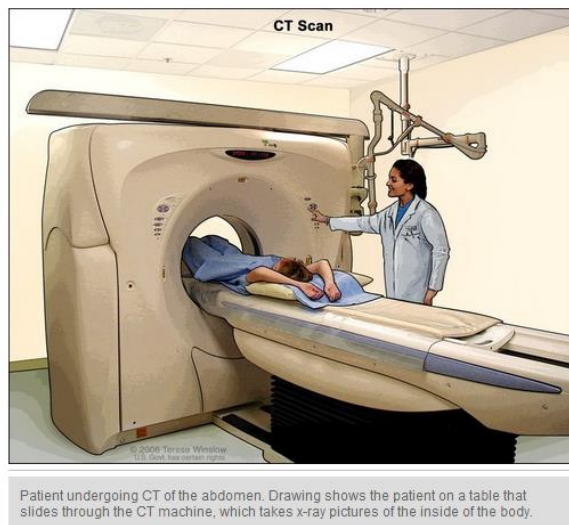


Fig. 1. Left: Patient undergoing CT of the abdomen. Left: CT image of Lung cancer

In this project, we will be using optical CT to diagnose MiniBots. Optical CT high-resolution, cross-sectional tomographic imaging of the internal microstructure in materials and biologic systems by measuring backscattered or back reflected light.

Phantoms have been used extensively for calibration of a new imaging technology or any new algorithm. In [2], fluorescence agent indocyanine green has been infused to a breast shaped phantom for the diagnosis of breast cancer. Companies such as Gammex has come up with innovative products such as Head/Body CT Phantom, which enables use of phantom without the need to fill in water [3]. CataPhan Inc. also provides phantoms for maximum obtainable performance of axial, spiral, multislice, conebeam and volume CT scanners [4].

Description of the lab and experiments performed

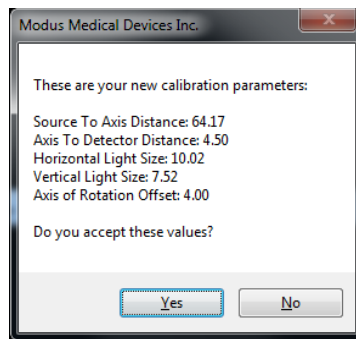
The goals of this project are follows:

- The overall goal of this project is to diagnose fabricated minibot in the best possible way using an DeskCat Optical CT scanner.
- To create a phantom that contains 3 tissue types : tumor, normal tissue and tear. The three should have different optical densities.
- Use the Optical CT scanner to measure the size of the tumors and the tear and verify with the actual size.
- Use the Optical CT scanner to measure the optical density of the tumor, tear and normal tissue.
- Export the most accurate 2D image to Matlab, add different levels of noise, and then explore the use of smoothing and edge detection filters to evaluate the tumor and tear.

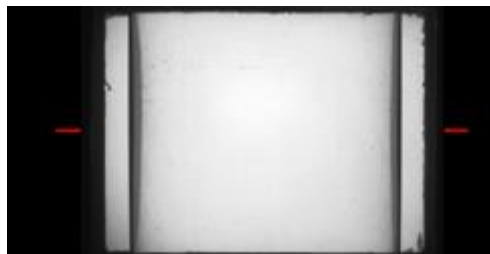
The first step of the experiment was to fabricate the phantom, which is described in detail in the next section.

Then we used DeskCat Optical CT Scanner to evaluate the 3 different tissue types. The steps for the experiment are as follows:

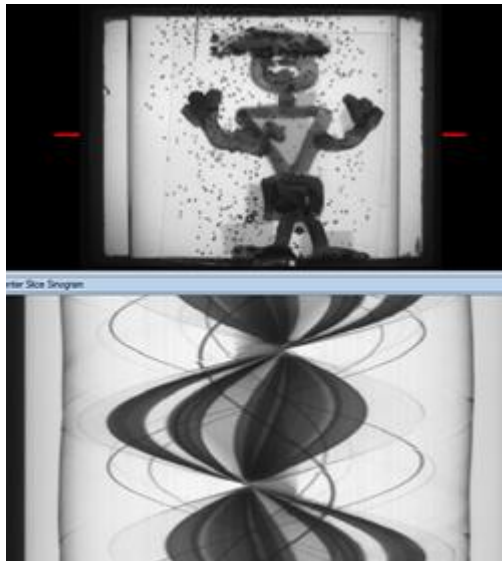
1. We checked motor connection, checked camera settings and calibrated without jar loaded.



2. Took the reference scan.



3. Checked the camera view, aligned for the reference view and the started the data scan.



4. Then we set voxel resolution to very high (0.25mm) and started the reconstruction of the images.

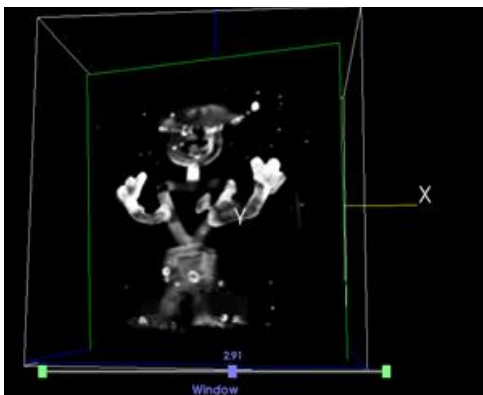


Fig. 2a. Reconstruction using Multiplanar Reconstruction (MPR)

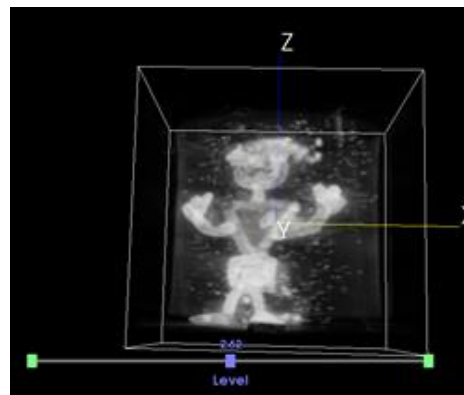


Fig. 2b. Reconstruction using Density Sum

5. From the reconstructed image, we took measurements of size and optical density of the tumor, tissue and tear.

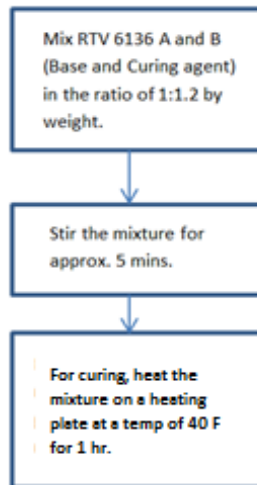
List of Materials Used

We used PDMS (Polydimethylsiloxane) as the normal tissue. We used Gelatin as the tumor tissue. As the tear in the normal tissue, we made an incision.

For PDMS, we used the following:

1. RTV6136 A and B (base and curing agent)
2. Digital Hot Plate

The procedure for making PDMS is as follows:



For making gelatin, we used warm water and red gelatin powder. We prepared the gelatin by mixing powder with warm water and left it in the refrigerator to cool down and solidify. We cut the gelatin into “fun” pieces and put them on thin transparent plastic. Following is how our phantom looks:



Fig. 3. The phantom made with soft PDMS and red gelatin.

For scanning the images we used the DeskCat Optical CT scanner. It is a portable imaging system that could be used interactively in the classroom or laboratory to demonstrate the principles of radiography and computed tomography (CT) using light rays instead of x-rays.

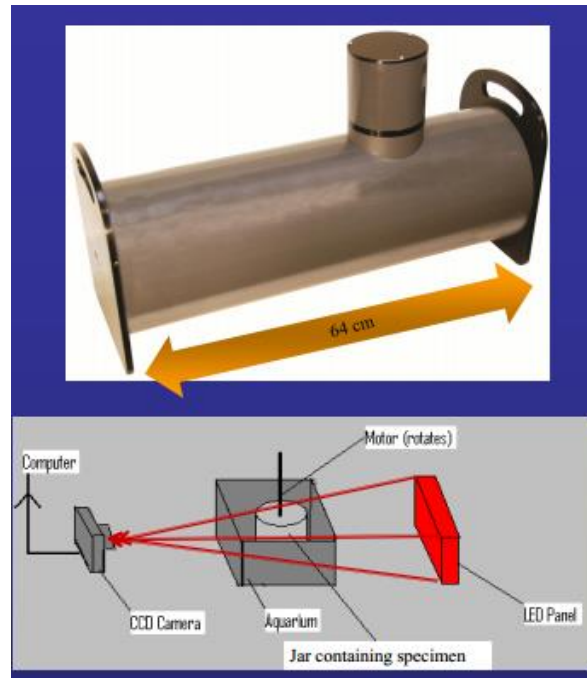


Fig. 4. DeskCAT optical CT scanner

1. **Figure out the appropriate background for the reference scan. Why did you use that type of reference scan? Use “very high” resolution, and obtain the reference scan and a 3D image of the phantom and MB (phantom can be separate or with the MB).**

Answer: For the reference scan we used a bottle filled with water. Our phantom’s background tissue is made of transparent silicone gel (PDMS). We used water for the background reference because it is close in optical properties to our silicone.

2. **Confirm the tissue types of the tumor and tear by comparison to the phantom. What are the optical densities measured to confirm that? Screenshot images to include in the lab report.**

Answer: We were able to differentiate the three tissue types (tumor, tear and normal) by measuring the optical density shown in the table below. The tumor had the highest attenuation coefficient, followed by the tear and the phantom.

Table 1. Results of density estimation

Tumor	Mean	S.D.
1	0.3428	0.0088
2	0.3816	0.0137
3	0.3431	0.0146
Tear		
1	0.0531	0.0002

2	0.0481	0.0022
3	0.0509	0.0006
Normal		
1	0.0157	0.0002
2	0.0191	0.008
3	0.0105	0.0081

Screen shots are presented next.

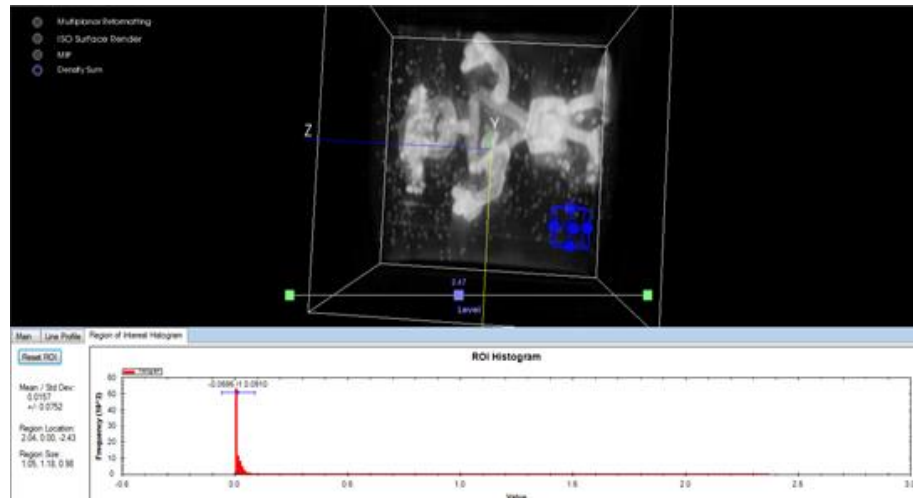


Fig. 5a. Measuring attenuation coefficient of normal tissue region

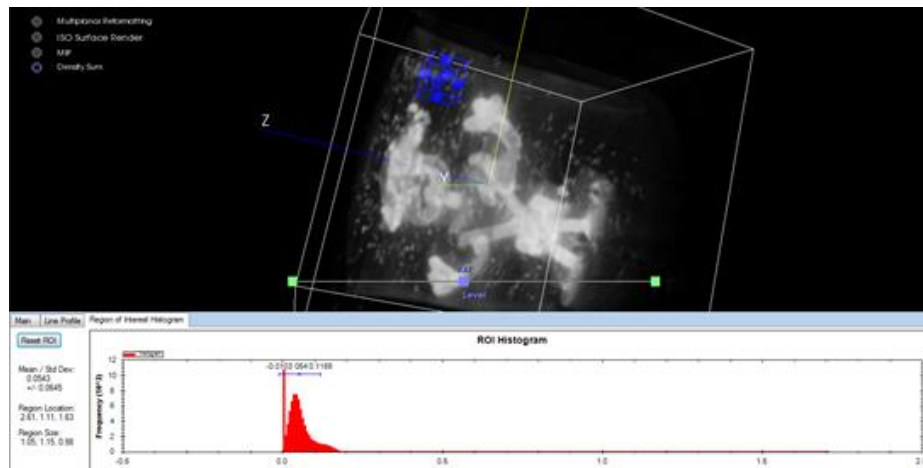


Fig. 5b. Measuring attenuation coefficient of tear region

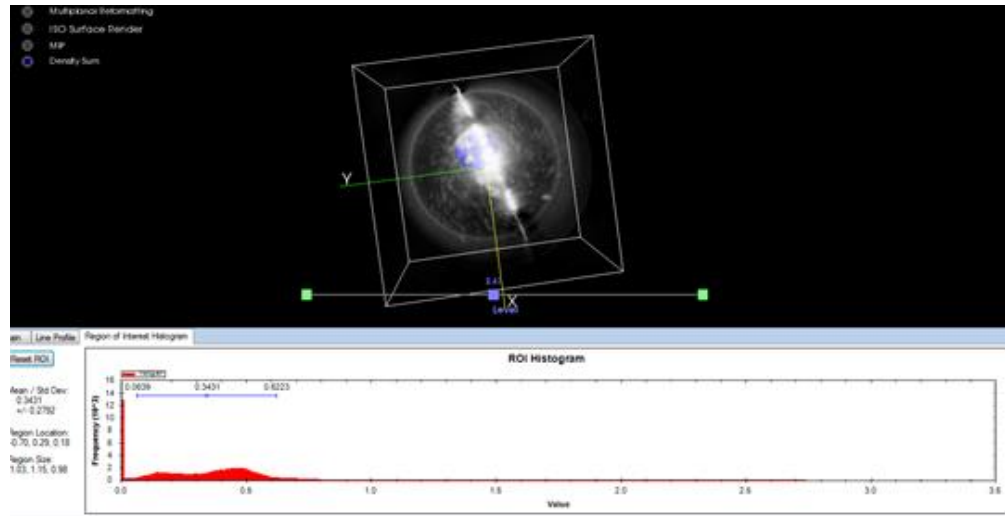


Fig. 5c. Measuring attenuation coefficient of tumor region

3. Obtain size measurements of the tumor and the tear (length) in five different 2D slice planes, and in the 3D image. *Screenshot images to include in the lab report.*

Answer: The original measured size of the tumor phantom is as follows:

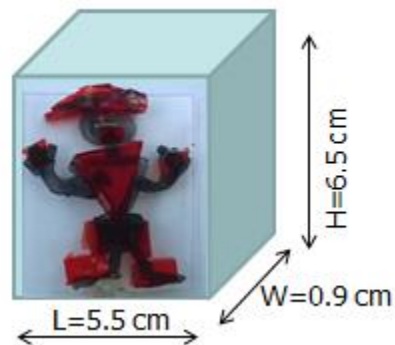


Fig. 6. The true dimensions of the phantom

The size measurements of the tumor and the tear in five different 2D slice planes, and 3D image presented in Table 2.

Table 2. Size measurements for MB and a tear.

2D	L	H	W	Tear
1	5.37	6.04	0.77	1.3
2	5.51	6.04	0.82	1.32
3	5.34	6.16	0.68	1.44

4	5.28	6.21	0.96	1.43
5	5.28	6.06	0.83	1.56
Avg.	5.35	6.10	0.81	1.41
3D	5.34	6.09	0.83	1.21
True values	5.50	6.50	0.90	1.60

Figure 7 show the our measurements process.

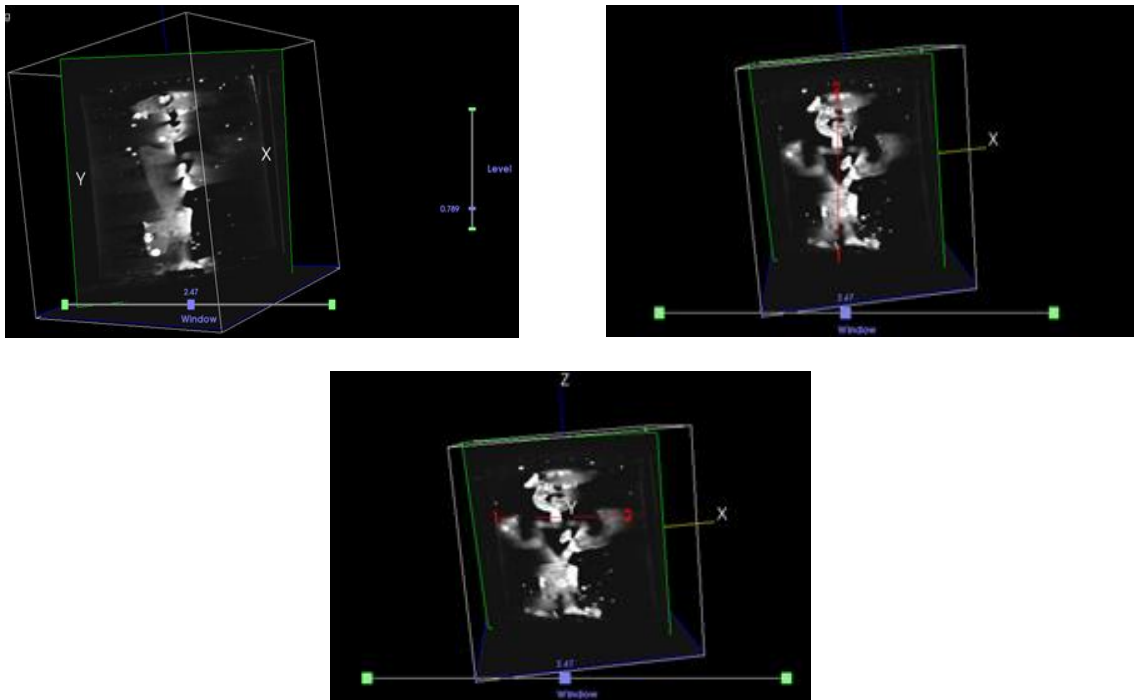


Fig. 7. Snapshots showing measurement of the size of the phantom.

4. You will create an MB as discussed in class. Since we're using the DeskCat, the MB has to be made of translucent materials. You will diagnose a tumor or plaque (i.e. , omething denser than the native tissue) in your MB. You will also diagnose an arterial or meniscal/cartilage tear in your MB.

Answer: We fabricated one phantom with both maladies. Background PDMS tissue was less stiff than minibot body components from gelatin. The phantom is shown on previous figures.

5. You'll also need to make a phantom that contains at least the three tissue types that are present in your MB. Alternatively, you can make three different phantoms with one tissue type each, but you need to have three tissue types to calibrate with all together. The three tissue types are the tumor, the tissue the tumor resides in, and the tissue that contains the tear, and they must have different optical densities. Be creative in what you construct your phantom and MB tissues from, but options include jello, gelatin, Knox Blox, gummy bears, silicone, paraffin, plastic, transparency sheets colored with markers, colored plastic cups, tissue paper, translucent

Answer: Our phantom was composed of three tissue types: tumor tissue, background tissue, and a tear. Tumor tissue made from dense red gelatin. Background tissue made of soft and transparent silicone gel (polydimethyl siloxane - PDMS). The tear was made by a vertical cut through with a sharp knife. The length of the tear 1.6 cm.

6. Decide which is the most accurate 2D image to determine size of the tumor and tear. Export these to Matlab

Answer: We chose to do raw images (.bmp) processing as well as from .vff file transfer for our image processing part of the report. One of the reasons was that the raw images showed the MB in full length along with a tear on the side in one vertical slice. In the images from .vff file we failed to find a region with a tear. PDMS is very elastic silicone and tear was almost invisible in horizontal slices from .vff file. That is why we present both results if image processing.

We consider two images for the image processing and analysis with raw images. Those are pretty noisy images. It was a challenge to work with them. One is the screenshot (640x480) taken by the DeskCAT instrument as shown in Fig. 8. Another one is a 2D slice image (320x320) extracted from .vff file. We consider Fig. 8 to be the most accurate 2D image (RGB) to determine size of the tumor and tear. This image was exported to MATLAB and converted to grayscale. In order to increase the signal to noise ratio, we subtracted this image from the background (reference) image and obtain Fig. 9.

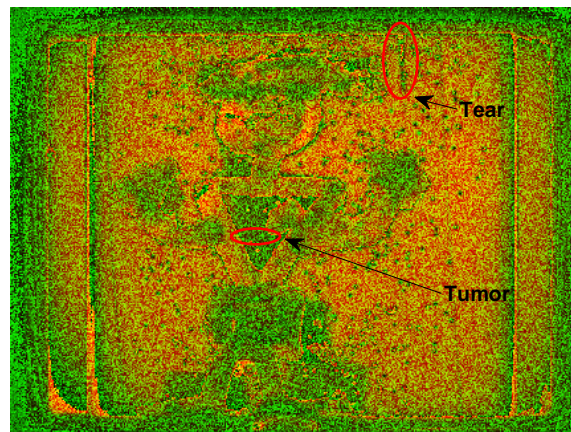
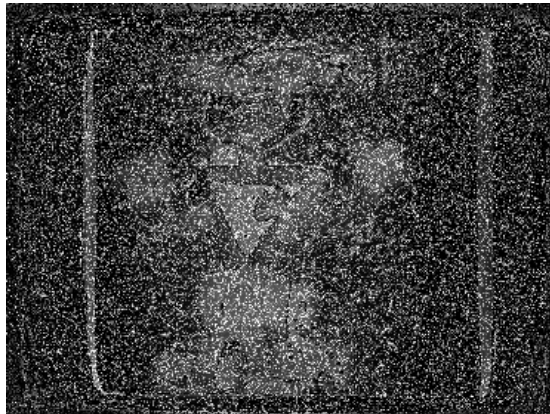


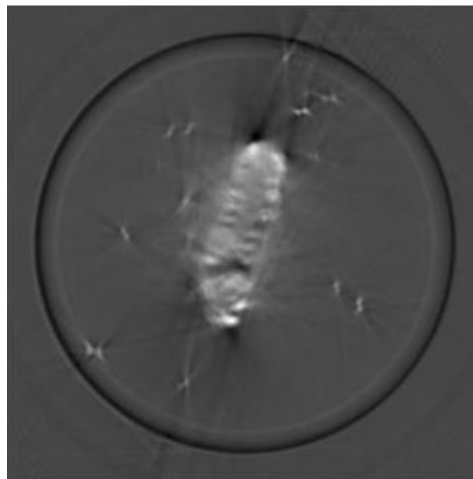
Fig. 8. 2D RGB image of phantom

Grayscale Image (Preprocessed)

**Fig. 9. Grayscale image of phantom**

We consider one image from .vff file transfer, which correspond to the “hat” region. As one can see from the Fig. 10, we have nicely defined “hat” region and some scatterings from small air bubbles.

Original Image

**Fig.10. 2D slice image extracted from the .vff file**

The 2D slice image corresponding to Fig. 10 was extracted from .vff file. This image contains less noise than image on Fig. 9 because it was reconstructed from the multiple view images. Reconstruction decreases noise level and increases signal to noise ratio (SNR).

- 7. In Matlab, generate random (Gaussian) noise using the RAND function, and add it to each of the 2D images. Add different levels of noise (either input standard deviation to change the amount of noise, or use a multiplier). Include screenshots or print out the images, whichever has better**

resolution. State what noise levels you tried, and at what point can you no longer discern features of image? Show images with “a little” and “a lot” of noise. What is the signal to noise ratio for these images? Is it the same in a homogeneous and nonhomogeneous region? Include numbers and explanation.

Answer: Raw images. We introduced Gaussian white noise with zero mean. The variance of the noise was varied to obtain different levels of noise. The variances are 0.0001 (a little noise), 0.001, 0.01, 0.1, and 1 (a lot noise). Fig. 11 shows the images with “a little” and “a lot” of noise.

Image with a LITTLE noise (mean=0,variance=0.0001) Image with a LOT noise (mean=0,variance=1)

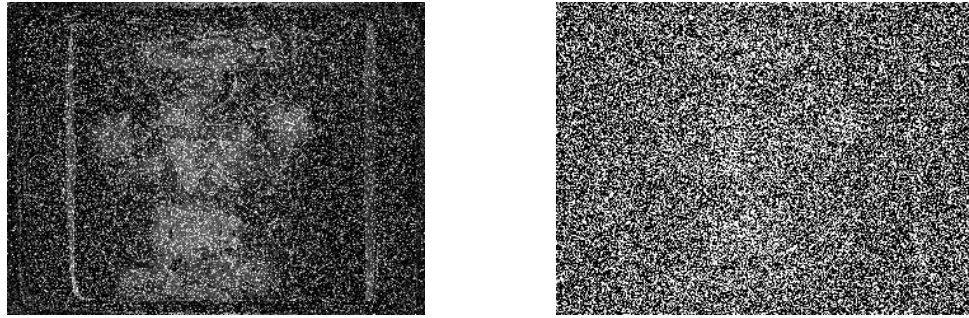


Fig.11. Image with “a little” and “a lot” noise

The signal to noise ratio values for each noise level are listed in Table 3.

Table 3. SNR for different levels of noise

Noise level	Zero mean Gaussian Random Noise with variance	SNR (dB)
1	0.0001	76.37
2	0.001	66.37
3	0.01	56.37
4	0.1	46.37
5	1	36.37

We cropped three regions of similar size (approximately 40×40 pixel) from a noisy image with noise variance 0.0001. Then we calculated the SNR of those image regions as listed in Table 4.

Table 4. SNR in different regions of phantom image

Region	Pixel Size	SNR (dB)
Phantom	640×480	76.37

Homogeneous (tissue)	40×40	75.75
Nonhomogeneous (tumor)	40×40	75.92
Nonhomogeneous (tear)	40×40	76.29

From Table 4, we see that the nonhomogeneous region of the phantom shows slightly higher SNR than the homogeneous region of the phantom. This result can be explained from the variances of the regions. The variances of the homogeneous, nonhomogeneous (tumor), nonhomogeneous (tear) are 3765, 3913, and 4258. The noise variance was same for all the regions. Therefore, SNR is proportional to the variances of the regions. Accordingly, the homogeneous regions with the lower signal variance show the lower SNR, and the nonhomogeneous regions show the higher SNR. However, all those differences are very small due to the high noise in the image.

For the .vff images, Figure 11b demonstrates how level of noise changes contrast of the image.

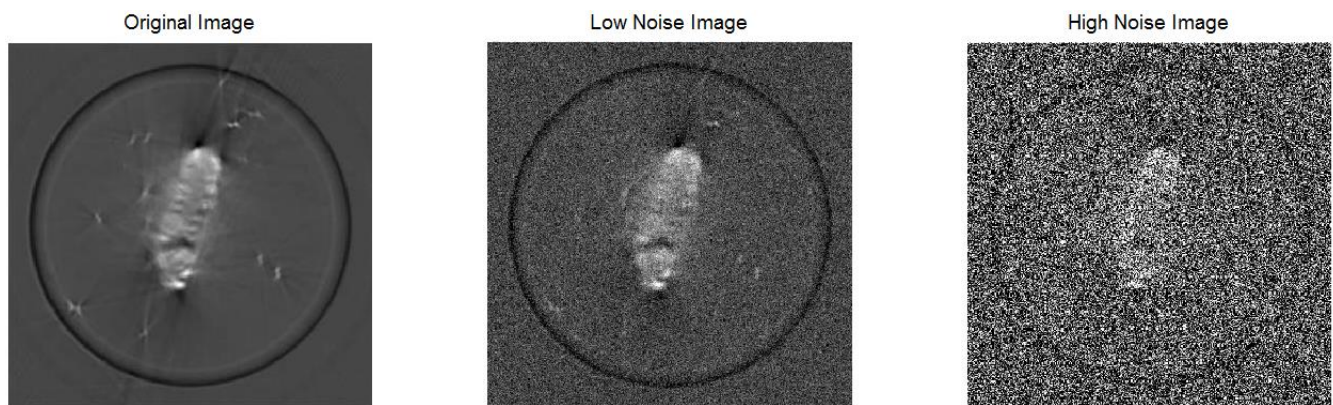


Fig.11b. Image (.vff) with “a little” and “a lot” noise

By looking on the Fig. 11b we can clearly see how noise covers information of the image.

8. **Apply the following filters to the image, and assess the effects of the filters: (*Extra credit points if you write your own filter code!*) Two smoothing filters of your choice. Do they work the same way? How? What’s different between the two filtered images? Describe the effect these filters have on the image. Does the answer depend on how noisy the image is? You can try iterative applications of filters to see the effect on the images as well. Two edge detection filters of your choice. Do they work the same way? How? What’s different between the two filtered images? Describe the effect these filters have on the image. Does the answer depend on how noisy the image is? Which filter is better to evaluate the tumor? What about to evaluate the tear?**

Answer:

We used two smoothing filters – Averaging filter and Gaussian filter [5]. Fig. 12 shows the images filtered with these two filters.

Effect of smoothing filter (raw images)

Image filtered with Averaging Filter

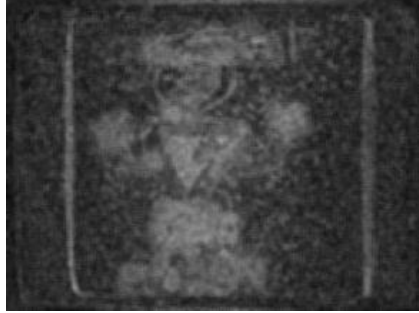
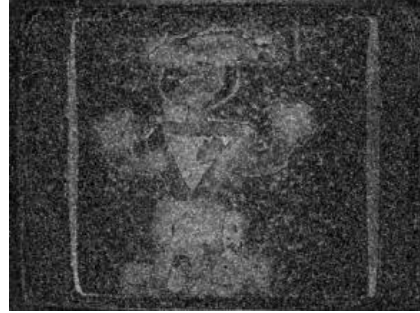


Image filtered with Gaussian Filter

**Fig.12. Filtered images with Averaging filter (left) and Gaussian filter (right)**

The two filters do not work the same way. The averaging filter replaces each pixel value in an image with the average value of its neighbor (defined by a $n \times n$ kernel) [5]. This filter can be considered as convolution filter. For example, a kernel of 3×3 gives the filter as:

$\frac{1}{9}$	$\frac{1}{9}$	$\frac{1}{9}$
$\frac{1}{9}$	$\frac{1}{9}$	$\frac{1}{9}$
$\frac{1}{9}$	$\frac{1}{9}$	$\frac{1}{9}$

Then this matrix is convolved with the original image to obtain a filtered image. The averaging signal reduces the variance noise, because the variance of noise in the average is smaller than variance of the pixel noise. In this case, a zero-mean Gaussian noise is assumed. The drawback of this filter is that it put the equal weights for all the pixel value, and therefore may not be the suitable filter where the contrast is desired.

On the other hand, the Gaussian filter uses unequal weights from a 2D Gaussian distribution [5]. In this case, the central pixels are given more weight than the neighboring weights. Gaussian filter is a low pass filter. Therefore, the filtered image is less affected with high frequency artifact.

Fig. 12 shows that the Gaussian filter smoothens the original image better than the averaging filter in terms of contrast.

The answer depends on how noisy the image is. We applied these two filters on a noisy image with noise variance 0.01. The result is shown in Fig. 13. It shows that the noise variance increase reduces the contrast. However, the Gaussian filter performed better than the averaging filter even in this noisy condition.

Image filtered with Averaging Filter

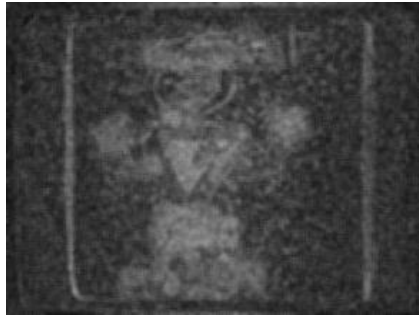


Image filtered with Gaussian Filter

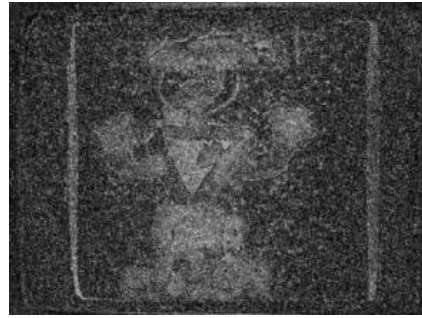


Fig.13. Filtered images with Averaging filter (left) and Gaussian filter (right) with higher noise variance

Iterative application of the filter reduces the quality of the images in this case. We applied the Averaging and Gaussian filter ten times iteratively on the same image. Fig. 14 shows that both filter performance deteriorates because of iterations.

Image filtered with Averaging Filter (Iteration=10)



Image filtered with Gaussian Filter (Iteration=10)

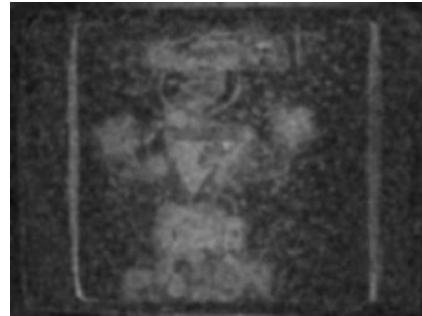


Fig.14. Filtered images with Averaging filter (left) and Gaussian filter (right) with 10 iterative application of filter

Image processing on 2D horizontal slice image from .vff file

We applied an averaging filter and a Gaussian filter described above on our original image to see changes from filtering (Fig. 15), on the image with low level of noise (Fig. 16), and on the image with high noise. We also proceed with three iterations of those two filters on the high noise image (Fig. 17).

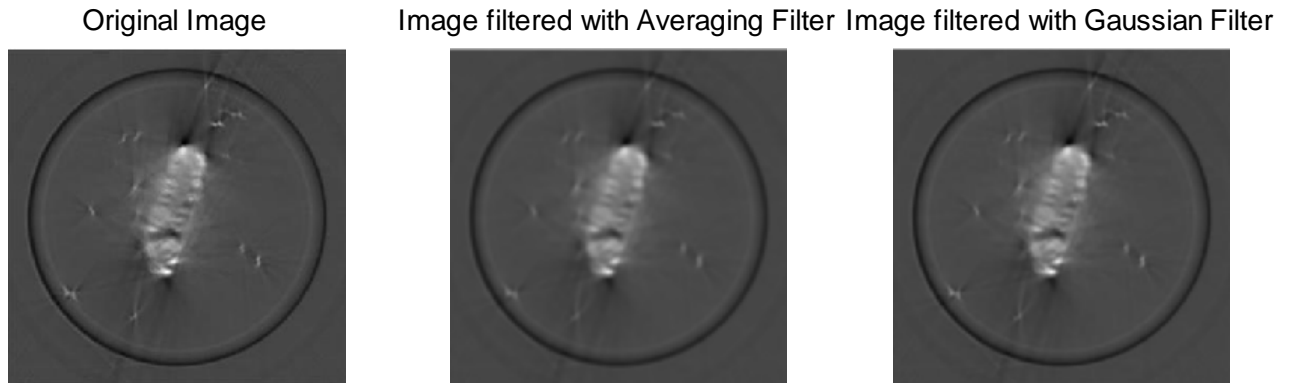


Fig.15. Original image (left), filtered images with Averaging filter (middle) and Gaussian filter (right)

Figure 15 shows little distortions from the application of averaging and Gaussian filter. Third image has sharper edges than the second image.

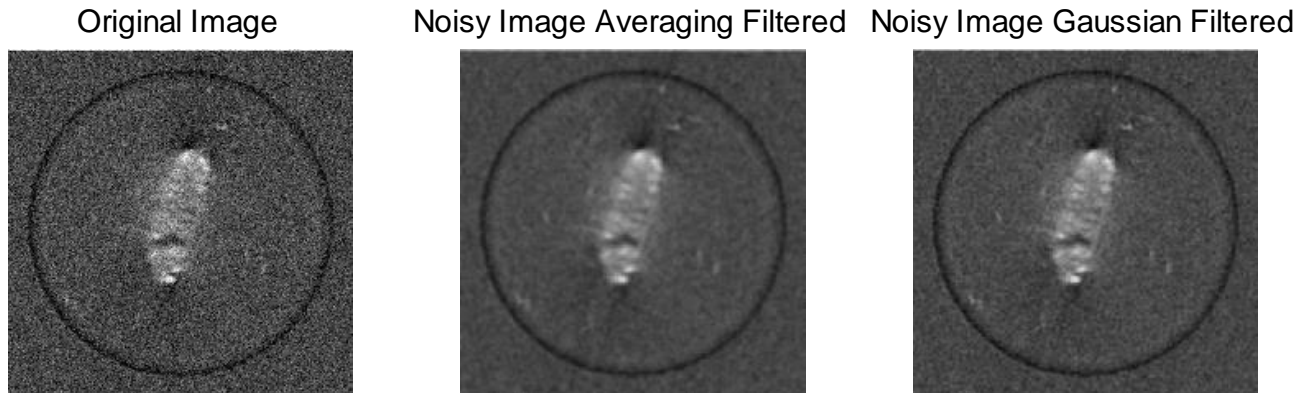


Fig.16. Noisy image (var=0.01) (left), filtered images with Averaging filter (middle) and Gaussian filter (right)

Figure 16 shows how those two filters filter image with low level of noise. And again, third image shows sharper edges than the second image.

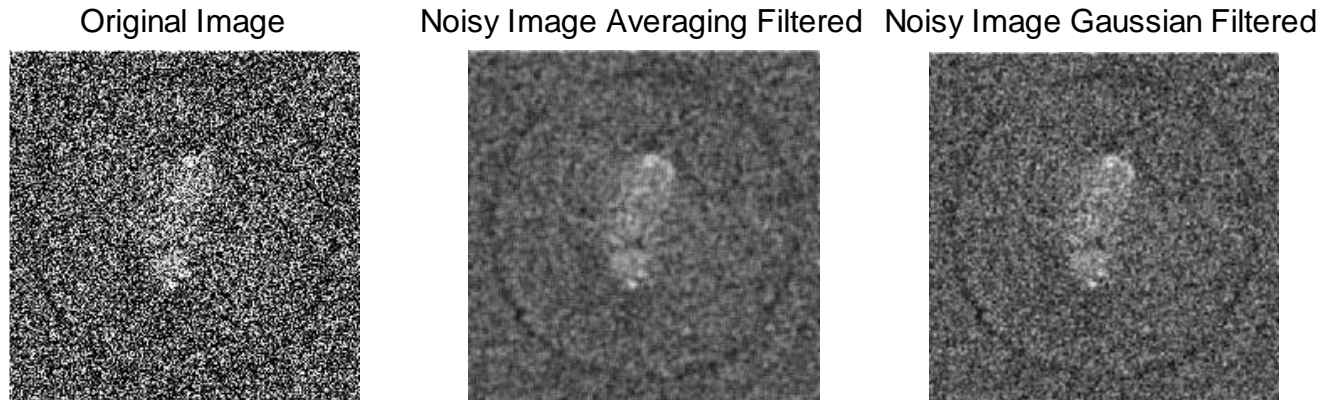


Fig.17. Noisy image (var=0.25) (left), filtered images with Averaging filter (middle) and Gaussian filter (right)

Figure 17 shows that both filters have to be more sophisticated to filter out the high level of Gaussian noise applied to the original image. The features of the image are hard to see.

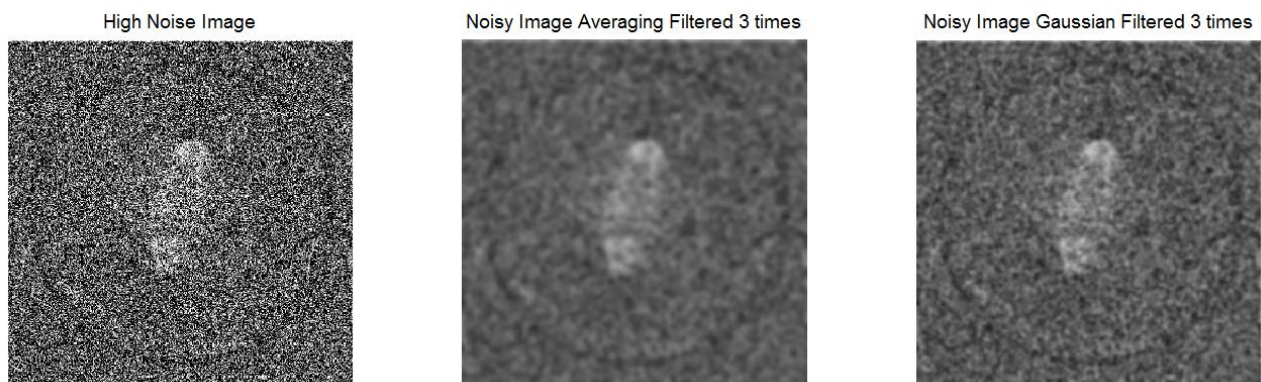


Fig.18. Noisy image (var=0.25) (left), filtered images with Averaging filter (middle) and Gaussian filter (right), iteration effect

Figure 18 shows that both filters did better after their sequential application to filter out the high level of Gaussian noise applied to the original image. The features of the image can be seen especially on the third image. The Gaussian filter seems to work better on the image than the averaging filter.

Effect of edge detection filters. Do they work the same way? How? What's different between the two filtered images? Describe the effect these filters have on the image. Does the answer depend on how noisy the image is?

Answer: We used two edge detection filters — Sobel filter and Canny filter [5]. Before applying these filters, we preprocess the grayscale images with morphological operations (*'strel'* and *'imerode'* functions in MATLAB). Fig. 19 shows the images filtered with the edge detection filters.

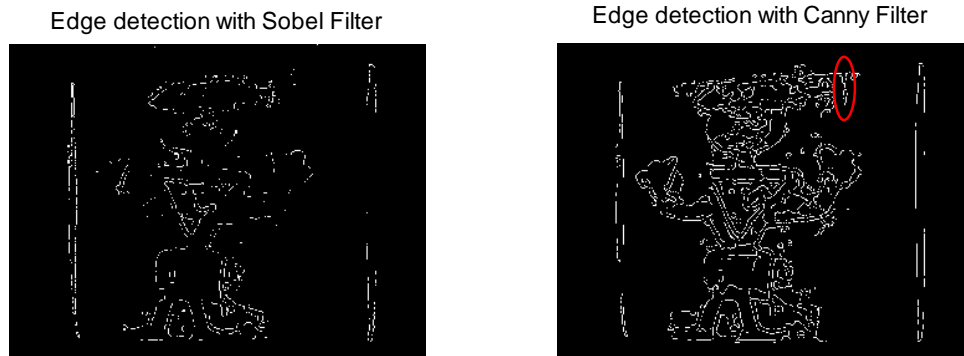


Fig.19. Edge detection with Sobel filter (left) and Canny filter (right)

The two edge detection filters do not work the same way. The input of the edge detection filter is a grayscale image. The output is a binary image where the pixels in the detected edge are 1 and the rest of the pixels are 0. The edge is defined as the place in the image where the pixel intensity varies rapidly. Therefore, the edge strength can be realized from the image gradient magnitude. In case of Sobel filter, this gradient or first derivative magnitude is found by Sobel approximation.

On the other hand, the Canny filter looks for the local maxima of the gradient of the image. A derivative of Gaussian filter is used to calculate the gradient. This method uses two thresholds to find strong and weak edges. We applied the Sobel filter with a threshold value of 0.2, and the Canny filter with threshold values of 0.2 and 0.08. The threshold values were obtained by trial and error. From Fig. 20, we observe that the Canny method was able to detect the edge of the minibot and the tear. However, the Sobel method showed poor performance. The noise affects the performance of the edge detection filters. We applied these two filters on a noisy image (noise variance = 0.001) and obtained Fig. 21.

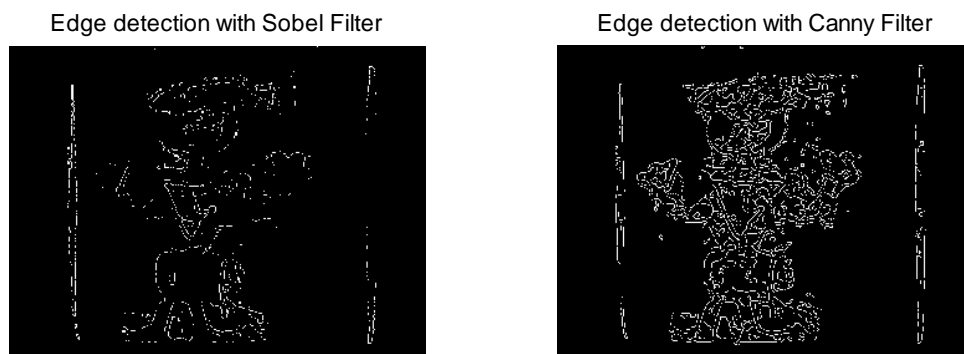
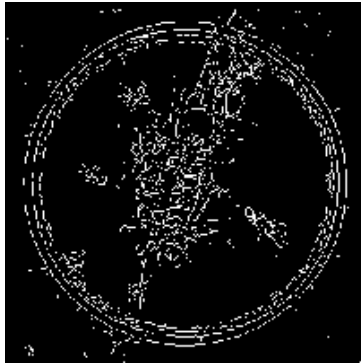
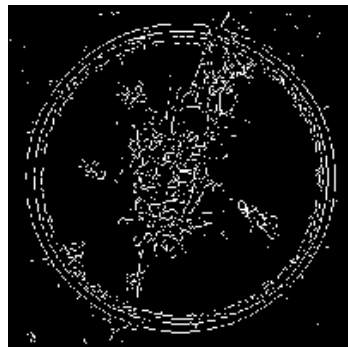


Fig.21. Edge detection with Sobel filter (left) and Canny filter (right) for noisy image

Edge detection on 2D slice from .vff file

Edge detection is a little difficult, yet was successful for the same two algorithms Sobel and Canny. There are multiple parameters in each filter to manipulate, so performance may be improved by trial and error. However, Canny filter seems to be more dynamic and adaptive. Figure 22 shows edge detection on original .vff image. Figure 23 shows edge detection on high noise image.

Edge detection with Sobel Filter**Edge detection with Canny Filter****Fig.22. Edge detection with Sobel filter (left) and Canny filter (right)****Edge detection with Sobel Filter****Edge detection with Canny Filter****Fig.23. Edge detection with Sobel filter (left) and Canny filter (right) for noisy image**

Which filter is better to evaluate the tumor? What about to evaluate the tear?

Answer: Based on raw images calculations, to evaluate tumor, the Gaussian filter is better. To evaluate the tear, the Canny edge detection filter is better.

Discussion

In this lab we were able to complete the following:

- Fabricated the phantom with the minibot, which had two tissue types (PDMS, Gelatin) and a tear.
- Measured size of the minibot and a tear.
- Differentiated minibot, tear and normal tissue by measuring their attenuation coefficients.
- Measured the size of the minibot, tear and normal tissue using DeskCat optical CT scanner.
- Performed image processing methods (smoothing, filtering, edge-detection) on the phantom. Gaussian filter worked better than Averaging filter for smoothing. Canny filter was better than Sobel for edge detection. Image processing methods were performed on raw images and on the .vff image.
- Results are presented as figures and tables.

Contributions

The data collection was performed by all the three members of the group. The phantom design was led by Vira Oleksyuk, supported by Amrita Sahu and Firdous Saleheen. The size and optical density calculations were led by Amrita Sahu, supported by Firdous Saleheen and Vira Oleksyuk. Matlab Analysis was led by Firdous Saleheen, supported by Vira Oleksyuk and Amrita Sahu. Presentation was led by Amrita Sahu, supported by Firdous Saleheen and Vira Oleksyuk. Writeup was led by Vira Oleksyuk and supported by Firdous Saleheen and Amrita Sahu.

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- [1] <http://www.cancer.gov/cancertopics/factsheet/detection/CT>
- [2] Diagnostic imaging of breast cancer using fluorescence-enhanced optical tomography: phantom studies. Godavarty A¹, Thompson AB, Roy R, Gurfinkel M, Eppstein MJ, Zhang C, Sevick-Muraca EM, J Biomed Opt. 2004 May-Jun; 9(3):488-96.
- [3] http://www.gammex.com/ace-images/DR_461A.pdf
- [4] <http://www.phantomlab.com/products/catphan.php>
- [5] Digital image processing, Rafael C. Gonzalez, and Richard E. Woods. Prentice Hall, Upper Saddle River, N.J., (2008)

Attachment

```

%% Work with VFF files
%% Program reads .vff file and outputs the required frames
%% Modification of the Shanrong Zhang and Samuel Lawman's codes
%% Vira Oleksyuk

%   This is a modification of the program described below

% %   This program can be used to open micro-CT VFF file in Matlab.
% %
% %   Shanrong Zhang, 12/17/2004
% %   Department of Radiology,
% %   University of Washington
% %   zhangs@u.washington.edu
% %

%   This allows 2D cross sections to be obtained from a 3D cube of data.
%   Easy modification will allow you to input 3D data into MatLAB from vff
%   files. Enjoy.

%   Samuel Lawman, 25/07/2008
%   School of Science and Technology
%   Nottingham Trent University

[filename pathname] = uigetfile('*.vff','Please select a vff file');

fid = fopen([pathname filename], 'r', 'b');

disp(filename)

num = 0;
done = false;

frame = 0;

line = fgetl(fid);
disp(line);
while (~isempty(line) && ~done)
    line = fgetl(fid);
    disp(line);
    if strmatch('size=', line)
        [token, rem] = strtok(line, 'size= ');
        M(1) = str2num(token);
        [token, rem] = strtok(rem);
        M(2) = str2num(token);
        M(3) = str2num(strtok(rem, ' '));
    end

    if strmatch('bits=', line)
        bits = str2num(strtok(line, 'bits= '));
    end
end

```

```

    num = num + 1;

    if num >= 14
        done = true;
    end
end
% ask = input(['Floating Point or Interger Data Type? (F/I)'], 's')
% if ask == 'I'
%     enctyp = 'int'
% else
%     enctyp = 'float'
% end

enctyp = 'float';
% while (frame < 1) | frame > M(3)
% fprintf(['\nWhich Frame number do you wish to see? 1-' num2str(M(3)) '\n'], 's')
% frame = input(['Frame?'])
%
% end
imagesBW = zeros(320,320);

for frame = 1:310

status = fseek(fid, -((M(1)).*(M(2)).*(M(3)).*bits./8) + (frame-
1)*M(1).*M(2).*bits./8, 'eof');

im = fread(fid,[M(1),M(2)],[enctyp num2str(bits)]);
imagesBW = [imagesBW im] ;

end

%% Display frames

for N=290:300 %chose which frame/frames you want to display

figure;
imshow(imagesBW(:,N*320:N*320+320), []);
colormap(gray);
title(filename);
%pause(0.2)
%close
end

%%
% end of code

%% BioImaging
%% Final Project

```

```

%% Image Processing on vff images
%% 5/13/14

close all
clear all
clc

%% Read image file
% % read rgb image file
% Irgb = imread('163.bmp');
% Iref = imread('163ref.bmp');
% % subtract the reference image from the data image
% I = Iref-Irgb;
% %figure, imshow(I)
% % convert rgb file to grayscale
% Igray = rgb2gray(I);
% figure,imshow(Igray);title 'Original Grayscale Image';
%% Read vff file
% in separate file

%% Read jpg files

Fig1=imread('Fig1.jpg');
Fig1 = rgb2gray(Fig1);
% Fig2=imread('Fig2.jpg');
% Fig2 = rgb2gray(Fig2);
% Fig3=imread('Fig3.jpg');
% Fig3 = rgb2gray(Fig3);
% Fig4=imread('Fig4.jpg');
% Fig4 = rgb2gray(Fig4);
% FigR=imread('FigReference.jpg');
% FigR = rgb2gray(FigR);

%% Image 1
hsize = [5 5];
H = fspecial('average',hsize);
figure;
%subplot(1,3,1);
imshow(Fig1);title('OriginalImage');

avIgray = imfilter(Fig1,H,'replicate');
%subplot(1,3,2);
figure;
imshow(avIgray);title('Image filtered with Averaging Filter');

%
sigma = 1;
H = fspecial('gaussian',hsize,sigma);
gaussIgray = imfilter(Fig1,H,'replicate');
%subplot(1,3,3);
figure;
imshow(gaussIgray);title('Image filtered with Gaussian Filter');

% CNR calculation
cnrIgray = cnrcalc(Fig1,avIgray);

```

```

cnrgaussIgray = cnrcalc(Fig1,gaussIgray);
%% Add noise and filter
V=0.01;
figure;
Fig1Noize1 = imnoise(Fig1,'gaussian',0,V) ;
subplot(1,3,1);
imshow(Fig1Noize1);
title('Original Image');

hsize = [5 5];
H = fspecial('average',hsize);
avIgray = imfilter(Fig1Noize1,H,'replicate');
subplot(1,3,2);
figure;
imshow(avIgray);title('Noisy Image filtered with Averaging Filter');

%
sigma = 1;
H = fspecial('gaussian',hsize,sigma);
gaussIgray = imfilter(Fig1Noize1,H,'replicate');
subplot(1,3,3);
figure;
imshow(gaussIgray);title('Noisy Image filtered with Gaussian Filter');

% CNR calculation
cnravIgray1 = cnrcalc(Fig1Noize1,avIgray);
cnrgaussIgray1 = cnrcalc(Fig1Noize1,gaussIgray);

%%
V=0.25;
figure;
Fig1Noize2 = imnoise(Fig1,'gaussian',0,V) ;
subplot(1,3,1);

imshow(Fig1Noize2);
title('Original Image');

figure;
hsize = [5 5];
H = fspecial('average',hsize);
avIgray = imfilter(Fig1Noize2,H,'replicate');
subplot(1,3,2);
imshow(avIgray);title('Noisy Image filtered with Averaging Filter');

%
sigma = 1;
H = fspecial('gaussian',hsize,sigma);
gaussIgray = imfilter(Fig1Noize2,H,'replicate');
subplot(1,3,3);
figure;
imshow(gaussIgray);title('Noisy Image filtered with Gaussian Filter');

% CNR calculation
cnravIgray2 = cnrcalc(Fig1Noize2,avIgray);

```



```

cnrgaussIgray2 = cnrcalc(Fig1Noize2,gaussIgray);

%% repeat filtering on noisy image 5 times

imshow(Fig1Noize2);
title('High Noise Image');

figure;
hsize = [5 5];

H = fspecial('average',hsize);

avIgray1 = imfilter(Fig1Noize2,H,'replicate');
avIgray1 = imfilter(avIgray1,H,'replicate');
avIgray1 = imfilter(avIgray1,H,'replicate');

%subplot(1,3,2);
imshow(avIgray1);title('Noisy Image filtered with Averaging Filter 3 times');

%
sigma = 1;
H = fspecial('gaussian',hsize,sigma);
gaussIgray1 = imfilter(Fig1Noize2,H,'replicate');
gaussIgray1 = imfilter(gaussIgray1,H,'replicate');
gaussIgray1 = imfilter(gaussIgray1,H,'replicate');

%subplot(1,3,3);
figure;
imshow(gaussIgray1);title('Noisy Image filtered with Gaussian Filter 3 times');

% CNR calculation
cnravIgray3 = cnrcalc(Fig1Noize2,avIgray1);
cnrgaussIgray3 = cnrcalc(Fig1Noize2,gaussIgray1);

%% %% Application of Edge Detection Filter
% Sobel filter and Canny filter
% preprocessing required before edge detection
Igray11 = Fig1;
se = strel('disk',1);
I2 = imerode(Igray11,se);
I3 = imadjust(I2);
imshow(I3);
%% Figure q
figure
edgeIgray = edge(I3,'sobel',0.02,'both');
subplot(1,2,1)
imshow(edgeIgray);title('Edge detection with Sobel Filter')
edgeIgray1 = edge(I3,'canny',0.08,'both');
subplot(1,2,2)
imshow(edgeIgray1);title('Edge detection with Canny Filter')
%%
% noise effect
figure

```

```

se = strel('disk',1);
I2 = imerode(I2,se);
I3 = imadjust(I2);
imshow(I3);

edgeIgray = edge(I3,'sobel',0.02,'both');
subplot(1,2,1)
imshow(edgeIgray);title('Edge detection with Sobel Filter')
edgeIgray1 = edge(I3,'canny',0.07,'both');
subplot(1,2,2)
imshow(edgeIgray1);title('Edge detection with Canny Filter')

%%

Raw Images Processing
%% Raw Images Processing

%% Firdous Saleheen

clear all; close all; clc;

%% Read image file
% read rgb image file
Irgb = imread('163.bmp');
Iref = imread('163ref.bmp');
% subtract the reference image from the data image
I = Iref-Irgb;
%figure, imshow(I)
% convert rgb file to grayscale
imshow(Irgb);
Igray = rgb2gray(I);
Igray = imadjust(Igray);
%Igray = wiener2(Igray,[3 3]);
figure,imshow(Igray);title 'Original Grayscale Image';

%% Add Gaussian Random Noise
% imnoise is a function that uses RAND to generate Gaussian random noises
% m and v are mean and variances of the noise distribution
m=0;
v1=0.0001;
v2=0.001;
v3=0.01;
v4=0.1;
v5=1;
J1 = imnoise(Igray,'gaussian',m,v1);
J2 = imnoise(Igray,'gaussian',m,v2);
J3 = imnoise(Igray,'gaussian',m,v3);
J4 = imnoise(Igray,'gaussian',m,v4);
J5 = imnoise(Igray,'gaussian',m,v5);
figure,imshow(J1);title 'Image with Gaussian Noise with \mu=0 and var=0.0001';
figure,imshow(J2);title 'Image with Gaussian Noise with \mu=0 and var=0.001';
figure,imshow(J3);title 'Image with Gaussian Noise with \mu=0 and var=0.01';
figure,imshow(J4);title 'Image with Gaussian Noise with \mu=0 and var=0.1';
figure,imshow(J5);title 'Image with Gaussian Noise with \mu=0 and var=1';

```

```

% Jarray = zeros(480,640,1,4);
% Jarray(:,:,,1) = J1;
% Jarray(:,:,,2) = J2;
% Jarray(:,:,,3) = J3;
% Jarray(:,:,,4) = J4;
% figure,montage(Jarray,'DisplayRange', []);
subplot(1,2,1);
imshow(J1); title 'Image with a LITTLE noise (mean=0,variance=0.0001)';
subplot(1,2,2);
imshow(J5); title 'Image with a LOT noise (mean=0,variance=1)';

%% Signal to Noise Ratio Calculation
% calculate variance of original image using std2 function
% noise variance is known
% SNR = signal variance/noise variance
% SNR(dB) = 10*log(SNR); 10 base log

SNR1 = 10*log10(std2(Igray).^2/v1);
SNR2 = 10*log10(std2(Igray).^2/v2);
SNR3 = 10*log10(std2(Igray).^2/v3);
SNR4 = 10*log10(std2(Igray).^2/v4);
SNR5 = 10*log10(std2(Igray).^2/v5);

%% SNR in homogeneous and nonhomogeneous region
% three small regions are cropped from J1 using imtool, tissue, tumor, tear
% imtool(J1); load homogeneous; load nonhomogeneous; load tear
%SNRhom = 10*log10(std2(homogeneous).^2/v1);
%SNRnonhom = 10*log10(std2(nonhomogeneous).^2/v1);
%SNRtear = 10*log10(std2(tear).^2/v1);

%% Application of Smoothing filter
% averaging filter and gaussian filter
hsize = [9 9];
H = fspecial('average',hsize);
avIgray = imfilter(Igray,H,'replicate');
subplot(1,2,1);
imshow(avIgray);title('Image filtered with Averaging Filter');

sigma = 1;
H = fspecial('gaussian',hsize,sigma);
gaussIgray = imfilter(Igray,H,'replicate');
subplot(1,2,2);
imshow(gaussIgray);title('Image filtered with Gaussian Filter');

% noise effect
hsize = [9 9];
H = fspecial('average',hsize);
avIgray = imfilter(J3,H,'replicate');
subplot(1,2,1);
imshow(avIgray);title('Image filtered with Averaging Filter');

sigma = 1;
H = fspecial('gaussian',hsize,sigma);
gaussIgray = imfilter(J3,H,'replicate');

```

```

subplot(1,2,2);
imshow(gaussIgray);title('Image filtered with Gaussian Filter');

%iterative application of filter
hsize = [9 9];
H = fspecial('average',hsize);
iter = 10;
avIgray = Igray;
for ii=1:iter
    avIgray = imfilter(avIgray,H,'replicate');
end

sigma = 1;
H = fspecial('gaussian',hsize,sigma);
gaussIgray = Igray;
for ii=1:iter
    gaussIgray = imfilter(gaussIgray,H,'replicate');
end
subplot(1,2,1);
imshow(avIgray);title('Image filtered with Averaging Filter (Iteration=10)');
subplot(1,2,2);
imshow(gaussIgray);title('Image filtered with Gaussian Filter (Iteration=10)');
%% Application of Edge Detection Filter
% Sobel filter and Canny filter
% preprocessing required before edge detection

se = strel('disk',3);
I2 = imerode(Igray,se);
I3 = imadjust(I2);
imshow(I3);

edgeIgray = edge(I3,'sobel',0.2,'both');
subplot(1,2,1)
imshow(edgeIgray);title('Edge detection with Sobel Filter')
edgeIgray1 = edge(I3,'canny',0.2,'both');
subplot(1,2,2)
imshow(edgeIgray1);title('Edge detection with Canny Filter')

% noise effect
se = strel('disk',3);
I2 = imerode(J2,se);
I3 = imadjust(I2);
imshow(I3);

edgeIgray = edge(I3,'sobel',0.2,'both');
subplot(1,2,1)
imshow(edgeIgray);title('Edge detection with Sobel Filter')
edgeIgray1 = edge(I3,'canny',0.2,'both');
subplot(1,2,2)
imshow(edgeIgray1);title('Edge detection with Canny Filter')

```