

Julia: a natural language for computational biology

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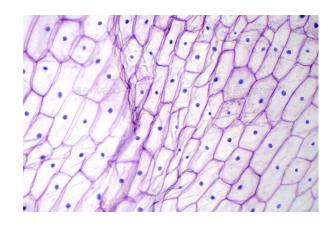
Talk outline

- Computational biology
- Julia
- BioJulia
- BioStructures.jl
- Bio3DView.jl
- Molly.jl
 - Simulating an ideal gas
 - Simulating diatomic molecules
 - Simulating proteins
- Differentiable molecular simulation
- Assessing Julia
- The future



What is biology?

"Biology is the natural science that studies life and living organisms, including their physical structure, chemical processes, molecular interactions, physiological mechanisms, development and evolution." - Wikipedia



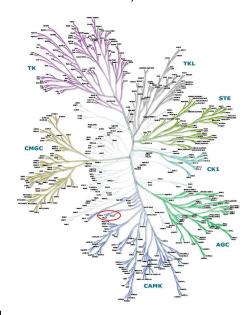


What *is* biology?

- Complex systems
- Multi-scale systems
- Lots of data
- Incomplete, noisy data
- Rules have exceptions

Computational biology addresses this by developing and applying data analysis, mathematical modeling and simulation techniques to biology.

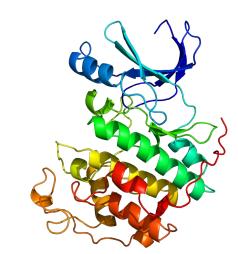
From Manning et al, *Science*, 2002



Protein structure prediction

- My day job involves predicting the structure of proteins from the sequence of amino acids that makes them up
- We do this using deep learning (CNNs/ResNets) on evolutionary information
- https://github.com/psipred/DMPfold

MENFQKVEKIGEGTYGVVYKARNKLTGEVVALKKIRLDTETEGVPSTAIREISLLKE LNHPNIVKLLDVIHTENKLYLVFEFLHQDLKKFMDASALTGIPLPLIKSYLFQLLQG LAFCHSHRVLHRDLKPQNLLINTEGAIKLADFGLARAFGVPVRTYTHEVVTLWYRAP EILLGCKYYSTAVDIWSLGCIFAEMVTRRALFPGDSEIDQLFRIFRTLGTPDEVVWP GVTSMPDYKPSFPKWARQDFSKVVPPLDEDGRSLLSQMLHYDPNKRISAKAALAHPF FQDVTKPVPHLRL



Julia

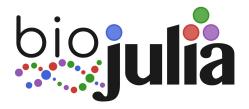
- High-level programming language
- Fast → addresses the two-language problem
- Free, open source
- Strong community
- Dynamically typed
- Multiple dispatch



```
[6]: abstract type Animal end
     struct Dog <: Animal
         name::String
     end
     struct Cat <: Animal
         name::String
         secret name::String
     end
     sayname(a::Cat) = println("Maiow ", a.name)
     sayname(a::Dog) = println("Woof ", a.name)
     function meet(a::Animal, b::Animal)
         println(a.name, " meets ", b.name)
     end
     dog = Dog("Buster")
     cat = Cat("Salem", "???")
     sayname(dog)
     Woof Buster
[7]: sayname(cat)
     Maiow Salem
[8]: meet(dog, cat)
     Buster meets Salem
```

BioJulia

- Fast, open, easy, software for biology
- The bioinformatics infrastructure for the Julia language



Efficient sequence type

```
In [1]: using BioSequences
In [2]: d = DNASequence("TTANC")
Out[2]: 5nt DNA Sequence:
    TTANC
In [3]: complement(d)
Out[3]: 5nt DNA Sequence:
    AATNG
```

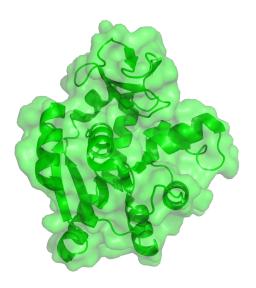
BioJulia

File parsers

```
In [4]: reader = FASTA.Reader(open("myseq.fa", "r"))
for record in reader
    # Do something
end
close(reader)
```

Alignments

- Read, write and manipulate macromolecular structures
- https://github.com/BioJulia/BioStructures.jl



PDB format

ATOM	1	N	MET	Α	1	26.981	53.977	40.085	1.00 40.83	N	
ATOM	2	CA	MET	Α	1	26.091	52.849	39.889	1.00 37.14	С	
ATOM	3	C	MET	Α	1	26.679	52.163	38.675	1.00 30.15	С	
ATOM	4	0	MET	Α	1	27.020	52.865	37.715	1.00 27.59	0	
ATOM	5	CB	MET	Α	1	24.677	53.310	39.580	1.00 38.06	С	
ATOM	6	CG	MET	Α	1	23.624	52.189	39.442	1.00 46.67	С	
ATOM	7	SD	MET	Α	1	21.917	52.816	39.301	1.00 61.54	S	
ATOM	8	CE	MET	Α	1	21.930	53.926	37.910	1.00 51.17	С	
ATOM	9	N	ARG	Α	2	26.861	50.841	38.803	1.00 28.23	N	
ATOM	10	CA	ARG	Α	2	27.437	49.969	37.786	1.00 25.76	С	

mmCIF format

```
ATOM 1 N N . MET A 1 1 ? 26.981 53.977 40.085 1.00 40.83 ? 1 MET A N 1
ATOM 2 C C A . MET A 1 1 ? 26.091 52.849 39.889 1.00 37.14 ? 1 MET A CA 1
ATOM 3 C C . MET A 1 1 ? 26.679 52.163 38.675 1.00 30.15 ? 1 MET A C 1
ATOM 4 0 0 . MET A 1 1 ? 27.020 52.865 37.715 1.00 27.59 ? 1 MET A 0 1
ATOM 5 C CB . MET A 1 1 ? 24.677 53.310 39.580 1.00 38.06 ? 1 MET A CB 1
ATOM 6 C CG . MET A 1 1 ? 23.624 52.189 39.442 1.00 46.67 ? 1 MET A CG 1
ATOM 7 S SD . MET A 1 1 ? 21.917 52.816 39.301 1.00 61.54 ? 1 MET A CG 1
ATOM 8 C CE . MET A 1 1 ? 21.930 53.926 37.910 1.00 51.17 ? 1 MET A CE 1
ATOM 9 N N . ARG A 1 2 ? 26.861 50.841 38.803 1.00 28.23 ? 2 ARG A N 1
ATOM 10 C CA . ARG A 1 2 ? 27.437 49.969 37.786 1.00 25.76 ? 2 ARG A CA 1
```

Read, write and manipulate macromolecular structures

```
In [25]: using BioStructures
In [26]: struc = read("1AKE.pdb", PDB)
Out[26]: ProteinStructure 1AKE.pdb with 1 models, 2 chains (A,B), 428 residues, 3804 atoms
In [27]: struc[1]["A"]
Out[27]: Chain A with 214 residues, 242 other molecules, 1954 atoms
In [28]: countresidues(struc[1]["A"], standardselector)
Out[28]: 214
In [29]: for at in collectatoms(struc, calphaselector)[1:5]
             println(atomname(at), " ", coords(at))
         end
         CA [26.091, 52.849, 39.889]
         CA [27.437, 49.969, 37.786]
         CA [24.961, 47.988, 35.671]
         CA [25.194, 44.925, 33.36]
         CA [22.428, 44.503, 30.712]
```

Spatial calculations

```
In [39]: # Print the PDB records for all C\alpha atoms within 4 Å of residue 38
         for at in collectatoms(struc['A'], calphaselector)
             if distance(struc['A'][38], at) < 4.0 && resnumber(at) != 38
                 println(pdbline(at))
             end
         end
         ATOM
                     CA ALA A 37
                                        33.778 51.895 15.373 1.00 27.13
         MOTA
                     CA VAL A
                                        36.426
                                               48.279
                                                      12.266
                                                              1.00 21.10
         ATOM
                     CA SER A 41
                                        38.507 53.491 11.905
         MOTA
                     CA GLY A 42
                                        40.955 50.617 11.185 1.00 29.51
         ATOM
                329
                     CA GLY A 46
                                        39.438 45.942 17.303 1.00 17.17
         MOTA
                    CA LYS A 47
                                        39.679 46.226 13.467 1.00 25.84
```

• Spatial calculations

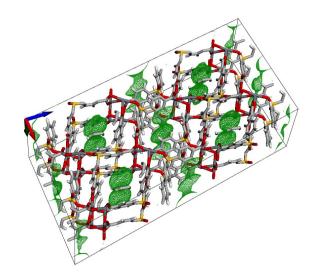
```
In [33]: dists = DistanceMap(collectatoms(struc['A'], cbetaselector))
Out[33]: Distance map of size (214, 214)
In [34]: using Plots
    plot(dists)
Out[34]:
                            16 31 47 62 77 92 108 123 138 153 168 184 199 214
                                                                                   - 45
                       31
                                                                                   - 40
                       47
                                                                                  - 35
                       77
                                                                                   - 30
                                                                                  - 25
                      107
                      123
                                                                                  - 20
                      138
                                                                                  - 15
                      153
                      168
                                                                                  - 10
                      184
```

Access the Protein Data Bank (PDB)

```
In []: # Calculate the cysteine fraction of every structure in the PDB
l = pdbentrylist()
for p in l
    downloadpdb(p, file_format=MMCIF) do fp
    s = read(fp, MMCIF)
    nres = countresidues(s, standardselector)
    if nres > 0
        frac = countresidues(s, standardselector, x -> resname(x) == "CYS") / nres
        println(p, " ", round(frac, digits=2))
    end
end
end
```

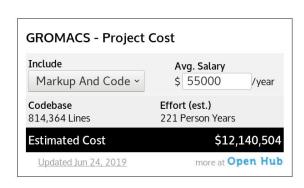
Bio3DView.jl

- Visualisation of macromolecular structures
- Wrapper round 3Dmol.js (Rego and Koes, Bioinformatics, 2015)
- Works in IJulia or Blink popup window
- https://github.com/jgreener64/Bio3DView.jl

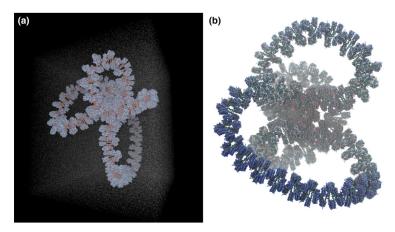


Molly.jl

- Molecular dynamics (MD) is a computational technique used to explore the movement and interaction of molecules
- Molly.jl is a proof-of-concept for MD in Julia
- https://github.com/jgreener64/Molly.jl



From gromacs.org



From Jung et al, J Comp Chem, 2019

Create some atoms with the relevant parameters defined

```
In [1]: using Molly
            n atoms = 100
            mass = 10.0
            atoms = [Atom(mass=mass, \sigma=0.3, \epsilon=0.2) for i in 1:n atoms]
Out[1]: 100-element Array{Atom,1}:
              Atom("", "", 0, "", 0.0, 10.0, 0.3, 0.2)
              Atom("", "", 0, "", 0.0, 10.0, 0.3, 0.2)
              Atom("", "", 0, "", 0.0, 10.0, 0.3, 0.2)
             Atom("", "", 0, "", 0.0, 10.0, 0.3, 0.2)
Atom("", "", 0, "", 0.0, 10.0, 0.3, 0.2)
Atom("", "", 0, "", 0.0, 10.0, 0.3, 0.2)
Atom("", "", 0, "", 0.0, 10.0, 0.3, 0.2)
             Atom("", "", 0, "", 0.0, 10.0, 0.3, 0.2)
Atom("", "", 0, "", 0.0, 10.0, 0.3, 0.2)
Atom("", "", 0, "", 0.0, 10.0, 0.3, 0.2)
              Atom("", "", 0, "", 0.0, 10.0, 0.3, 0.2)
              Atom("", "", 0, "", 0.0, 10.0, 0.3, 0.2)
```

Define some starting coordinates and velocities

```
In [2]: box size = 2.0 \# nm
        coords = [Coordinates(rand(3) .* box size) for i in 1:n atoms]
        temperature = 298 \# K
        velocities = [Velocity(mass, temperature) for i in 1:n atoms]
Out[2]: 100-element Array{Velocity,1}:
         [-3.62082, -4.11298, -1.80747]
         [-6.06135, 2.48857, -0.968609]
         [3.5725, -1.64122, 2.61146]
         [-8.20442, -1.96685, 12.5213]
         [6.7406, 8.06838, 2.47006]
         [6.26042, -5.08734, 7.4645]
         [-5.41582, 7.2642, -0.0616184]
         [10.4321, -2.95435, -1.25541]
         [4.51567, 6.09565, 4.18389]
         [-5.45313, 1.96933, -1.17946]
         [5.50508, -5.97522, -2.35687]
         [-0.969843, 2.73667, 4.7403]
         [-2.38129, 4.79214, -6.1126]
```

- Now we can define our dictionary of general interactions, i.e. those between most or all atoms
- Because we have defined the relevant parameters for the atoms, we can use the built-in Lennard Jones type

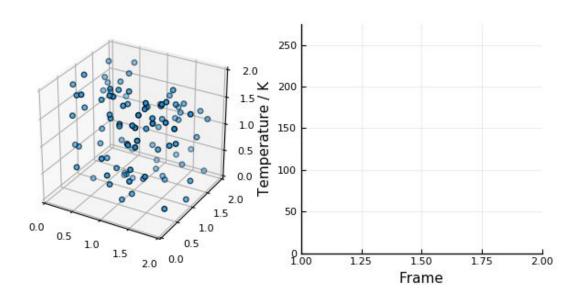
- Define and run the simulation
- Use an Andersen thermostat to keep a constant temperature
- Log the temperature and coordinates every 100 steps

Starting simulation

Progress: 99%| ETA: 0:00:00

 Plot the simulation

```
In [26]: using Plots
         pyplot(leg=false)
         coords = s.loggers[2].coords
         temps = s.loggers[1].temperatures
         splitcoords(coord) = [c[1] for c in coord], [c[2] for c in coord], [c[3] for c in coord]
         @gif for (i, coord) in enumerate(coords)
             l = @layout [a b{0.7h}]
             cx, cy, cz = splitcoords(coord)
             p = scatter(cx, cy, cz,
                 xlims=(0, box size),
                  ylims=(0, box size),
                  zlims=(0, box size),
                  layout=l
             plot!(p[2],
                  temps[1:i],
                  xlabel="Frame",
                  ylabel="Temperature / K",
                 xlims=(1, i),
                  vlims=(0.0, maximum(temps[1:i]))
         end
```



Simulating a diatomic molecule

- We can define specific interactions between groups of atoms, e.g. bonds
- Use the built-in bond type to place a harmonic constraint between paired atoms

```
In [13]: bonds = [Bond((i * 2) - 1, i * 2, 0.1, 300_000) for i in 1:(n_atoms / 2)]
    specific_inter_lists = Dict("Bonds" => bonds)
```

```
In [12]: coords = Coordinates[]
    for i in 1:(n_atoms / 2)
        c = rand(3) .* box_size
        push!(coords, Coordinates(c))
        push!(coords, Coordinates(c + [0.1, 0.0, 0.0]))
    end

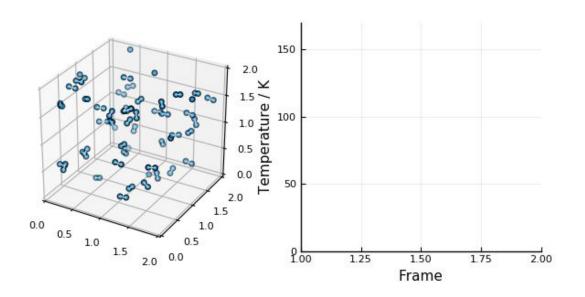
velocities = [Velocity(mass, temperature) for i in 1:n_atoms]
```

Simulating a diatomic molecule

- Use a neighbour list to speed up the Lennard Jones calculation
- We will use the provided neighbour finder but you could write your own

```
In [15]: neighbour_finder = DistanceNeighbourFinder(trues(n_atoms, n_atoms), 10, 1.2)
```

Simulating a diatomic molecule



Simulating a protein

- In addition to Lennard Jones forces and bonds, proteins also have angles, dihedral angles (torsions) and electrostatics
- Molly.jl has basic functionality to read in topology and forcefield data from GROMACS format files

Simulating a protein

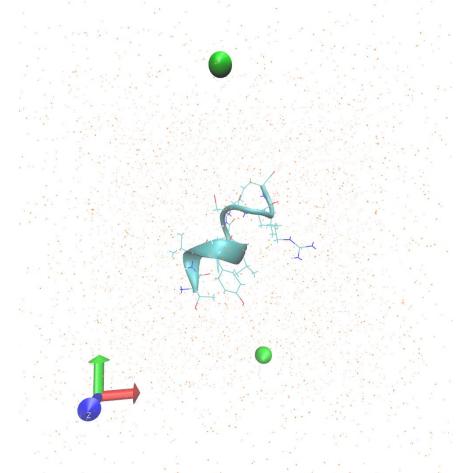
Simulate as before

Starting simulation

Progress: 5%| | | | | ETA: 0:11:04

Simulating a protein

 1 ps simulation viewed with VMD



Defining a force

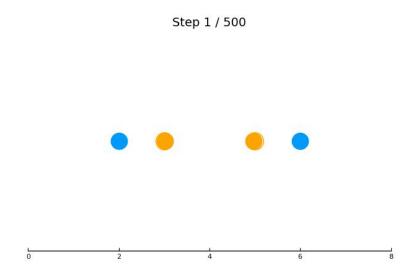
The entire definition for the electrostatic (Coulomb) force

```
In [ ]: struct Coulomb <: GeneralInteraction</pre>
            nl only::Bool
In [ ]: @fastmath @inbounds function update accelerations!(accels::Vector{Acceleration},
                                                     inter::Coulomb,
                                                     s::Simulation,
                                                     i::Integer,
                                                     i::Integer)
            dx = vector1D(s.coords[i].x, s.coords[j].x, s.box size)
            dy = vector1D(s.coords[i].y, s.coords[j].y, s.box size)
            dz = vector1D(s.coords[i].z, s.coords[j].z, s.box size)
            r2 = dx * dx + dy * dy + dz * dz
            if r2 > sqdist cutoff nb
                 return
            end
            f = (coulomb const * s.atoms[i].charge * s.atoms[j].charge) / sqrt(r2 ^ 3)
            accels[i].x += -f * dx
            accels[i].v += -f * dv
            accels[i].z += -f * dz
            accels[i].x += f * dx
            accels[i].v += f * dv
            accels[j].z += f * dz
        end
```

Differentiable molecular simulation

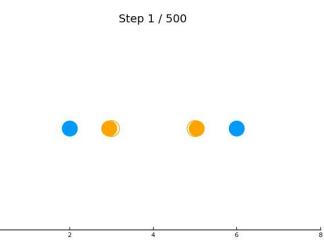
 Recent developments with Flux.jl make it possible to run molecular simulations and obtain gradients of parameters, e.g. forcefield parameters

```
[66]: b0learn = param(1.0)
      b0true = 4.0
      k = 0.01
      nsteps = 500
      function simulate(nsteps)
          coords = [3.0, 5.0]
          coords last = copy(coords) + randn(2) * 0.01
          for i in 1:nsteps
              dist = abs(coords[2] - coords[1])
              force = k * (dist - b0learn) * abs(dist - b0learn)
              dir = coords[1] < coords[2] ? 1.0 : -1.0
              coords next = 2 * coords - coords last + [dir * force, -dir * force]
              coords = coords next
              coords last = coords
          end
          return coords
      end
      simulate(nsteps)
```



Differentiable molecular simulation

```
[74]: function loss(b0true, nsteps)
          coords = simulate(nsteps)
          dist = abs(coords[2] - coords[1])
          return (dist - b0true) ^ 2
      end
      gs = Tracker.gradient(() -> loss(b0true, nsteps), params(b0learn))
      qs[b0learn]
      -5.773390700589583 (tracked)
[75]: for i in 1:500
          gs = Tracker.gradient(() -> loss(b0true, nsteps), params(b0learn))
          update!(b0learn, -0.01 * qs[b0learn])
          i % 50 == 0 && println("Epoch ", i, " - b0learn ", b0learn)
      end
      Epoch 50 - b0learn 2.6801969855276746 (tracked)
      Epoch 100 - b0learn 3.574286748329535 (tracked)
      Epoch 150 - b0learn 3.9043047080232394 (tracked)
      Epoch 200 - b0learn 4.025176930121734 (tracked)
      Epoch 250 - b0learn 4.069391236713611 (tracked)
      Epoch 300 - b0learn 4.0855609902908805 (tracked)
      Epoch 350 - b0learn 4.09147724816171 (tracked)
      Epoch 400 - b0learn 4.093635443806043 (tracked)
      Epoch 450 - b0learn 4.094422175776826 (tracked)
      Epoch 500 - b0learn 4.094708733260627 (tracked)
```



Julia for computational biology

Pros

- High-level language with nice syntax
- Fast
- Prototype and code in the same language
- Sane design
- Great community
- Making packages is relatively easy
- There is a rabbit hole to go down

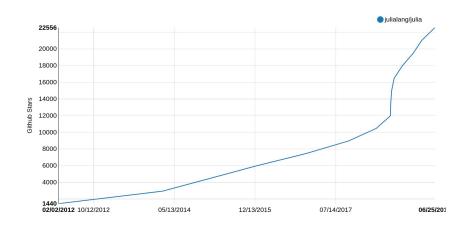
Cons

- Compile time requires a workflow change
- Performance gotchas
- Effort required to shift from OOP
- Plotting is still getting there
- Libraries still being implemented
- Sometimes you have to go down the rabbit hole



The future

- Popularity of the language increasing
- Increasingly used in teaching and research
- Stability of Julia v1.0 has made things easier for package developers
- Exciting developments in the language:
 - Improved multithreading
 - GPU/TPU programming
 - Deep learning/differentiable programming
 - Static compilation



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