Lab 2 - Gr. 14 - Bioinformatics (732A93)

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Assignment 1

Question 1.1

Starting from 33 DNA sequence of various species of casque-headed lizard (Basiliscus basiliscus), other 33 sequences of nucleotides have been generated. The sampling probabilities are the same of the real proportions of the original dataset.
After the artificial DNA has been created, the base frequencies are compared in Table 1. As expected, the observed proportions of the generated data closely resamble the theoretical ones.

Question 1.2

- Created one phylogenetic tree with 33 tips
- For each original DNA sequence of the 33 available, used the function `simSeq(.)` from package `phangorn` to simulate the sequences.
- Result: 33 phylogenetic tree, one for DNA sequence, each with 33 tips.

Is the result Krzysztof wanted? I've read the suggested materials and the lecture slides, but I am still not sure how to use the tree in order to simulate the sequences.

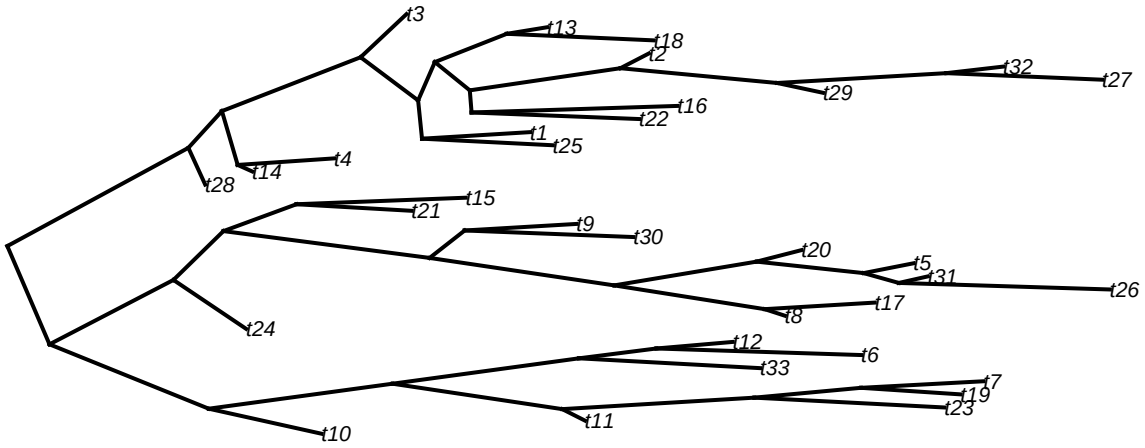

Table 1: Base frequencies of the 33 original and generated DNA sequences.

Base	Original frequency	Simulated frequency
a	0.3121	0.3120
c	0.2052	0.2045
g	0.2307	0.2329
t	0.2519	0.2505

Table 2: Base frequencies of the 33 original DNA sequences and of the 33 simulated phylogenetic trees.

Base	Original frequency	Simulated frequency
a	0.3121	0.3126
c	0.2052	0.2053
g	0.2307	0.2307
t	0.2519	0.2514

Table 3: Lenght of the 33 original DNA sequences and of the 33 simulated phylogenetic trees.

Base	Original frequency	Simulated frequency
1	997	1012
2	2709	2597
3	2891	2634
4	1006	1061
5	1474	1369
6	1091	1059
7	2891	2762
8	1004	904
9	1044	1068
10	2920	3174
11	2816	2958
12	2886	3135
13	2748	2807
14	2835	2731
15	2744	2629
16	1003	965
17	999	1023
18	2825	2977
19	2845	2573
20	1008	1068
21	1472	1380
22	2892	2945
23	1444	1350
24	1000	936
25	2837	2940
26	2506	2623
27	1005	1073
28	2886	2772
29	2890	2683
30	1060	1117
31	2737	2993
32	1003	1027
33	931	983

Assignment 2

2.1

2.2

2.3

Assignment 3

3.1

3.2

Appendix

```
knitr::opts_chunk$set(fig.width = 7, fig.height = 3, echo = FALSE)

library(dplyr)
library(tidyr)
library(magrittr)
library(ape)          # This is a general R-package for phylogenetics and comparative methods
library(seqinr)       # This is an specialized package for nucleotide sequence management
library(phangorn)
library(kableExtra)

source("732A51_BioinformaticsHT2018_Lab02_GenBankGetCode.R")
# ------------------------------------------------------------------------------
# Question 1.1
# ------------------------------------------------------------------------------

lizards_format_sequences <- read.fasta(file = "lizard_seqs.fasta")  # Alternative version of the file
# Useful in some ways?

n <- length(lizards_accession_numbers)  # Number of sequences to reproduce
p <- base.freq(lizards_sequences)  # Probability of the base sequences
simulated_lizards <- list()  # Object that will contain our simulated data

# The names of the simulated data are the original names + "_sim"
# NOTE: it does not follow the format from GenBank
simulated_names <- paste(lizards_accession_numbers, "_sim", sep = "")

set.seed(1535)  # Set seed in order to reproduce the experiment
for(i in 1:n) {  # Cycle through every single object of the lizard_sequences
  len_seq <- length(lizards_sequences[[i]])  # Lenght of each sequence
  simulated_lizards[[ simulated_names[i] ]] <-
    sample(c("a", "c", "g", "t"), len_seq, replace = T, prob = p)
  # Creating the artificial sequence sampling with probabilities p equal to the original ones
  # NOTE: we use the general distribution for every single sequence
}
```

```r
# Save as fasta file
write.dna(simulated_lizards, file = "simulated_lizards.fasta", format = "fasta", append = F,
          nbcol = 6, colsep = " ", colw = 10)

# Table with simulated base frequency
df_table <- data.frame("Base" = c("a", "c", "g", "t"),
                       "Original\nfrequency" = p,
                       "Simulated\nfrequency" = base.freq(as.DNAbin(simulated_lizards)),
                       row.names = NULL)
kable(df_table, booktabs = T, align = c("r", "l", "l"), digits = c(NA, 4, 4),
      col.names = c("Base", "Original\nfrequency", "Simulated\nfrequency"), format = "latex",
      caption = "Base frequencies of the 33 original and generated DNA sequences.")
# ------------------------------------------------------------------------------
# Question 1.2
# ------------------------------------------------------------------------------

################################################################################
# Create the tree
################################################################################

# Set the random seed for reproducibility once again
set.seed(1545)
# Simulate the tree: it has to be with 33 tips. We try to generate a tree with uniform distribution
# (default function)
simulated_tree <- rtree(33)

# Plot the tree:
par(lend = 2)
par(mar = rep(2, 4))
par(cex = 0.8)

plot(simulated_tree, type = "cladogram", edge.width = 2)  # Cladogram template, the easiest to
                                                          # understand for this tree


################################################################################
# Simulate the sequences
################################################################################

# Object where to store the randomic sequences
simulated_tree_seq <- list()
# The names of the simulated data are the original names + "_sim_tree"
# NOTE: it does not follow the format from GenBank
simulated_names_tree <- paste(lizards_accession_numbers, "_sim_tree", sep = "")


# We now define a function to compute a partially random transiction matrix:
# It first calculate first-order Markov transition matrix where each *row*
# corresponds to a single run of the Markov chain. It then add some random noise.

trans.matrix <- function(DNA_seq) {

  # Retrieve unique elements
```

```
  DNA_unique <- unique(DNA_seq)

  # Create an empty matrix
  matrix <- matrix(0, ncol = length(DNA_unique), nrow=length(DNA_unique))

  # Fill it: to count i and element i + 1 and add one in the corresponding cell of the matrix.
  for (i in 1:(length(DNA_seq) - 1)) {
    index_of_i <- DNA_unique == DNA_seq[i]
    index_of_i_plus_1 <- DNA_unique == DNA_seq[i + 1]
    matrix[index_of_i, index_of_i_plus_1] = matrix[index_of_i, index_of_i_plus_1] + 1
  }

  # Normalize it
  matrix <- matrix / rowSums(matrix)

  # Add random noise, keeping the rowSum equal to 1
  # Proceed row by row
  for(i in nrow(matrix)) {
    noise <- rnorm(1, mean = 0,  sd = 0.01)   # Random quantity to ADD/SUBtract
    ind_add <- floor(runif(1, 1, nrow(matrix)+1))  # Column index where to add
    ind_sub <- floor(runif(1, 1, nrow(matrix)+1))  # Column index where to subtract
    matrix[i,ind_add] <- matrix[i,ind_add] + noise  # Add noise to one prob...
    matrix[i,ind_sub] <- matrix[i,ind_sub] - noise  # ...and subtract from another
  }

  return(matrix)
}


# Time to generate! Set the seed again (better safe than sorry)
set.seed(1545)
# Here we will store all the newly generated lenght of the sequences
len_generated_sequences <- rep(NA, 33)
len_original_sequences <- rep(NA, 33)

# Create one sequence for each one of the originals
for(i in 1:n) {
  # Problem: original sequences include unknown nucleotides --> delete them
  known_nucleotides <- lizards_sequences[[i]][lizards_sequences[[i]] %in% c(18, 28, 48, 88)]

  # Decide lenght randomically: we take the lenght of the original known sequence +
  # uniform random number equal max to 10% of the original lenght
  len_seq <- length(known_nucleotides)
  len_original_sequences[i] <- len_seq
  len_seq <- len_seq + floor(runif(1, -floor(len_seq/10), floor(len_seq/10)))
  len_generated_sequences[i] <- len_seq

  # We re-use the same probabilities computed before, referring to the original dataset.
  # We also compute the transiction matrix using the function defined above.
  simulated_tree_seq[[ simulated_names_tree[i] ]] <-
    simSeq(simulated_tree, l = len_seq, bf = p, Q = trans.matrix(known_nucleotides))
}
```

```r
# Convert the 33 generated trees as DNAbin
simulated_tree_seq2 <- as.DNAbin(simulated_tree_seq)

# Save as fasta file
write.dna(simulated_tree_seq2, file = "simulated_lizards_tree.fasta", format = "fasta", append = F,
          nbcol = 6, colsep = " ", colw = 10)

# Table with simulated base frequency
df_table <- data.frame("Base" = c("a", "c", "g", "t"),
                       "Original\nfrequency" = p,
                       "Simulated\nfrequency" = base.freq(simulated_tree_seq2),
                       row.names = NULL)

kable(df_table, booktabs = T, align = c("r", "l", "l"), digits = c(NA, 4, 4),
      col.names = c("Base", "Original\nfrequency", "Simulated\nfrequency"), format = "latex",
      caption = "Base frequencies of the 33 original DNA sequences and of the 33 simulated phylogenetic
# Table with simulated lenght vs original
df_table <- data.frame("Sequence n." = 1:33,
                       "Original\nlenght" = len_original_sequences,
                       "Simulated\nlenght" = len_generated_sequences,
                       row.names = NULL)

kable(df_table, booktabs = T, align = c("r", "l", "l"),
      col.names = c("Base", "Original\nfrequency", "Simulated\nfrequency"), format = "latex",
      caption = "Lenght of the 33 original DNA sequences and of the 33 simulated phylogenetic trees.")
# ------------------------------------------------------------------------------
# Question 2.1
# ------------------------------------------------------------------------------

# ------------------------------------------------------------------------------
# Question 2.2
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# ------------------------------------------------------------------------------
# Question 2.2
# ------------------------------------------------------------------------------

# ------------------------------------------------------------------------------
# Question 3.1
# ------------------------------------------------------------------------------

# ------------------------------------------------------------------------------
# Question 3.2
# ------------------------------------------------------------------------------
```