Multivariate Statistical Methods - Lab 02

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Contents

1	Test of	Outliers	-
	1.1 Chi-	-Squared Approximation	
		Ferent Outlier Reasoning	
2	Test, Confidence Region and Confidence Intervals for a Mean Vector		
	2.1 Con	afidence Ellipse	
	2.2 Sim	ultaneous T2-intervals	
	2.3 Q-Q	plots and scatter plots	
3	Comparison of Mean Vectors (one-way MANOVA)		
	3.1 Exp	oloring the Data	
		ering of Mean Vectors	
		ifidence Intervals	
	3.3.		
	3.3.		
	3.3.		
	3.3.		
1	Course	Codo	1.

Focusing on the multivariate normal distribution, we will study methods for estimating, testing hypotheses about and comparing mean vectors. These methods are the multivariate generalizations of the univariate methods.

1 Test of Outliers

Consider again the data set from the T1-9.dat file, National track records for women. In the first assignment we studied different distance measures between an observation and the sample average vector. The most common multivariate residual is the Mahalanobis distance and we computed this distance for all observations.

1.1 Chi-Squared Approximation

The Mahalanobis distance is approximately chi–square distributed, if the data comes from a multivariate normal distribution and the number of observations is large. Use this chi–square approximation for testing each observation at the 0.1 percent significance level and conclude which countries can be regarded as outliers. Should you use a multiple–testing correction procedure? Compare the results with and without one. Why is (or maybe is not) 0.1 percent a sensible significance level for this task?

Answer: First we import, name and look at the track times.

```
country 100m 200m 400m 800m 1500m 3000m marathon
        ARG 11.57 22.94 52.50 2.05 4.25 9.19 150.32
## 1
        AUS 11.12 22.23 48.63 1.98 4.02 8.63
                                               143.51
## 2
## 3
        AUT 11.15 22.70 50.62 1.94 4.05 8.78
                                               154.35
        BEL 11.14 22.48 51.45 1.97 4.08 8.82
## 4
                                               143.05
        BER 11.46 23.05 53.30 2.07 4.29 9.81
## 5
                                               174.18
        BRA 11.17 22.60 50.62 1.97 4.17 9.04
## 6
                                               147.41
```

We reimport our function written in the previous lab for computing the Mahalanobis Distance as it is actually more convenient than the built-in function in ${\bf R}$.

```
sample_variance = function(X) {
    X = as.matrix(X)

    identity = diag(nrow(X))
    one_n = matrix(1, nrow=nrow(X), ncol=1)

    inter = identity - 1/nrow(X) * (one_n %*% t(one_n))

    return(1/nrow(X) * (t(X) %*% inter %*% X))
}

mahalanobis_distance = function(X) {
    X = as.matrix(X)

    V = sample_variance(X)
    ident = matrix(1, nrow=nrow(X), ncol=nrow(X))
    mu = 1/nrow(X) * (t(ident) %*% X)

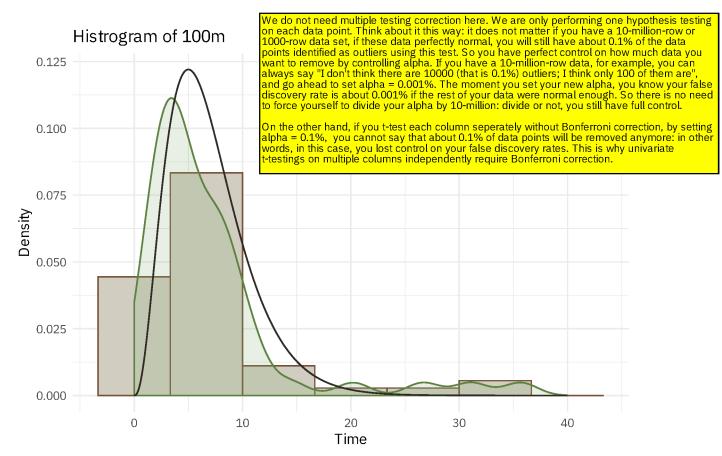
    X_centered = X - mu

    return(diag(X_centered %*% solve(V) %*% t(X_centered)))
}
```

We set the degrees of freedom for the $\chi^2(\nu)$ -distribution, which corresponds to the amount of features. We also calculate the Mahalanobis Distances. They should follow a $\chi^2(\nu)$ -distribution with 7 degrees of freedom.

```
nu = ncol(track_times) - 1
D = mahalanobis_distance(track_times[,2:8])
```

The following plot shows the histogram of the Mahalanobis Distances with the respective density. The real $\chi^2(\nu)$ -distribution with 7 degrees of freedom is outlined in black.



We define $\alpha = 0.001$ and we check for each observation if it lies within the $1 - \alpha$ percentile of the $\chi^2(\nu)$ -distribution with 7 degrees of freedom. Finally we check which countries are the outliers. Our findings match the results from the previous lab.

Below is the test without any multiple test correction. In this case the countries KORN, PNG and SAM are regarded as outliers.

```
alpha = 0.001
outlier_indeces = 1 - pchisq(D, nu) < alpha
track_times$country[outlier_indeces]</pre>
```

[1] KORN PNG SAM

54 Levels: ARG AUS AUT BEL BER BRA CAN CHI CHN COK COL CRC CZE DEN ... USA

Below is the test results with the correction for multiple tests. In this case the only country that is an outlier is SAM.

```
alpha = 0.001 / nrow(track_times)
outlier_indeces = 1 - pchisq(D, nu) < alpha
track_times$country[outlier_indeces]</pre>
```

[1] SAM

54 Levels: ARG AUS AUT BEL BER BRA CAN CHI CHN COK COL CRC CZE DEN ... USA

Multiple test correction should be used because we are doing multiple test at the same time, so the original significance level α should be adjusted to reflect that we are comparing multiple countries.

In this case the significance level $\alpha=0.1\%$ it depends on what is to be considered an outlier. In this case it seems reasonable since we want countries were its distance is way off from the other ones. So 0.01% seems reasonable. But this also depends on the application.

1.2 Different Outlier Reasoning

One outlier is North Korea. This country is not an outlier with the Euclidean distance. Try to explain these seemingly contradictory result.

The contradictory results arise from the fact that the euclidean distance doesn't normalize the data give the variations on the observations (variance and covariance) while the Mahalanobis distance does consider these variations. This is why we are getting contradictory results.

2 Test, Confidence Region and Confidence Intervals for a Mean Vector

Look at the bird data in file T5-12.dat and solve Exercise 5.20 of *Johnson*, *Wichern*. Do not use any extra R package or built—in test but code all required matrix calculations. You MAY NOT use loops!

2.1 Confidence Ellipse

```
library(ellipse)
##
## Attaching package: 'ellipse'
## The following object is masked from 'package:car':
##
##
       ellipse
## The following object is masked from 'package:graphics':
##
##
       pairs
birddata = read.table("T5-12.DAT")
# Male hook-billed kites
n = nrow(birddata)
p = dim(birddata)[2]
mu = data.frame("mu1" = 190, "mu2" = 275)
meanVec = NULL
x_bar = NULL
meanMu = NULL
x_bar = data.frame("X1" = mean(birddata$V1), "X2" = mean(birddata$V2))
sample_var = function(obs){
  X = as.matrix(obs)
                                                 You should divide by n-1 instead of n. Read the text
book carefully. But otherwise everything else are
  IdentityMat = diag(nrow(obs))
  one_n = matrix(1,nrow(X),nrow(X))
  # sample var formula from notes
  mix = (IdentityMat - (1/nrow(X)) *(one_n))
  sample_variance = (1/\text{nrow}(X)) * (t(X) %*% mix %*% X)
```

```
S = sample_var(birddata)
S = matrix(S, ncol=2, nrow=2)
eigenValVec = eigen(S)
meanMu = as.matrix(x_bar - mu)
# To check if mu is on the confidence region we need to check if t^2 is F_{n,n-p}
Tsq = n * (meanMu) %*% solve(S) %*% t(meanMu)
F_val = qf(0.95,df1=p, df=n-p) * (p*(n-1)/(n-p)) #5% significance level
cat("Is T^2 less than F_val? =",Tsq<F_val)</pre>
## Is T^2 less than F_{val} = TRUE
cat("\nThus, we do not reject the NULL Hypothesis(H_0)")
##
## Thus, we do not reject the NULL Hypothesis(H_0)
axes_len = function(lam,n,p){
  sqrt(lam)* sqrt((p*(n-1))/ (n*(n-p)) * qf(0.95,df1=p, df2=(n-p)))
}
11=axes_len(lam=eigenValVec$values[1],n=n,p=p)
12=axes_len(lam=eigenValVec$values[2],n=n,p=p)
# plot(birddata)
# lines(ellipse(mu, S, npoints = 200))
# dataEllipse(birddata, levels=.95)
# length(c(mu$mu1,mu$mu2))
```

Are these plaussible values for the mean tail length and wing length for the female birds? **Answer** It is quite plaussible as female hook-billed vary a lot in size just like the male hook-billed. Sometimes the juveniles are bigger than the size than they are supposed to be. And thus the mean size of tail length and wing length could be same for the female hook-billed kites.

```
rotate = function(a,b,c){
    res = list()

if (b==0 & a >= c){res$angle = 0}
    else if (b==0 & a<c) { res$angle = (pi/2)}
    else{ res$angle = atan2(eigenValVec$values[1]-a, b)}

res$lam1 = (a+c)/2 + sqrt(((a-c)/2)^2 + b^2)
    res$lam2 = (a+c)/2 - sqrt(((a-c)/2)^2 + b^2)

return(res)
}

# S = matrix(c(4,-2,-2,4),byrow = T, nrow = 2)
res = rotate(S[1,1],S[1,2],S[2,2])

chi95<-qchisq(.95,df=2)</pre>
```

This is not a confidence ellipse for the population mean. The confidence ellipse of the mean should be much, much smaller than this. The question means that you should plot the confidence ellipse for the population mean, and decide whether c(190, 275) is in the ellipse or not. So there is also no need to do the T2 test in this section. See equation 5-18 for the correct formula.

```
axes95 <- sqrt(eigenValVec$values) * sqrt(chi95)

In the ellipse or not. So there is also not the T2 test in this section. See equation correct formula.

library(ggforce)
ggplot()+
geom_point(aes(birddata$V1, birddata$V2))+
geom_ellipse(aes(x0=mu$mu1,y0=mu$mu2, a=axes95[1], b=axes95[2], angle = res$angle ))

300-

240-

170 180 190 200 210
```

2.2 Simultaneous T2-intervals

```
{\it\# Simultaneous\ T2-intervals\ for\ the\ component\ means\ as\ shadows\ of\ the\ confidence\ ellipse\ on\ the\ axes}
fsqrt = function(i){
  return(sqrt(p*(n-1)/ ((n-p)) * qf(0.95,df1=p, df=n-p)) * sqrt(S[i,i]/n))
                                                                                       Correct.
}
CI = NULL
CI$Lower = (x_bar$X1) - fsqrt(1)
CI$Upper =(x_bar$X1) + fsqrt(1)
CI = as.data.frame(CI)
temp = c ((x_bar_X^2) - fsqrt(2), (x_bar_X^2) + fsqrt(2))
CI = rbind(CI, temp)
rownames(CI) = c("Mu_1_tail","Mu_2_wing")
# kableExtra::kable(CI)
CI
                Lower
                          Upper
```

birddata\$V1

```
Correct.
## Mu_1_tail 189.4687 197.7758
## Mu_2_wing 274.3180 285.2375
# Bonferroni's Confidence intervals
set.seed(12345)
ben_alpha = 0.95
Ber_CI = NULL
Ber_CI$Lower = x_{a} = x_1 - abs(qt(0.05/2*p, df=n-1)) * sqrt(S[1,1]/n)
Ber_CI$Upper= x_{a} + abs(qt(0.05/2*p, df=n-1)) * sqrt(S[1,1]/n)
Ber_CI = as.data.frame(Ber_CI)
temp = c(x_bar^{X2} - abs(qt(0.05/2*p, df=n-1)) * sqrt(S[2,2]/n),
         x_bar$X2 + abs(qt(0.05/2*p, df=n-1)) * sqrt(S[2,2]/n))
Ber_CI = rbind(Ber_CI, temp)
rownames(Ber_CI) = c("Mu_1_tail", "Mu_2_wing")
Ber_CI
                Lower
                         Upper
## Mu_1_tail 190.9012 196.3432
## Mu_2_wing 276.2011 283.3544
```

 T^2 simultaneous CI is slightly wider than Bonferroni Intervals. Bonferroni method provides shorter intervals when m = p. Because they are easy to apply and provide the relatively short confidence intervals needed for inference.

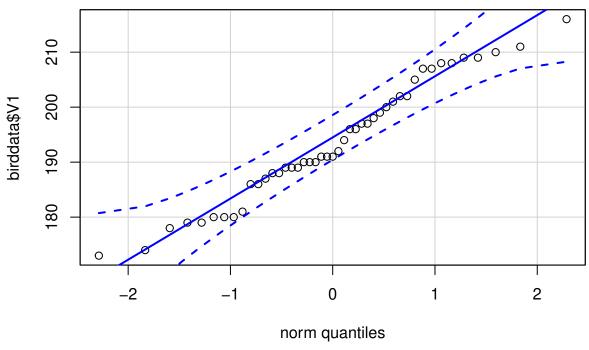
source: Applied multivariate Statistical Analysis - Pearson edition 2014

2.3 Q-Q plots and scatter plots

The data doesn't look normal from the QQ-plots.

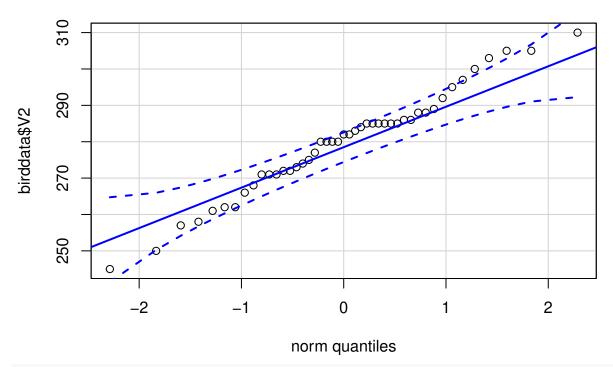
```
library(CARS)
qqPlot(birddata$V1, main ="QQ plot for X1: Tail length",id=F )
```

QQ plot for X1: Tail length



qqPlot(birddata\$V2, main="QQ plot for X2: Wing Length",id=F)

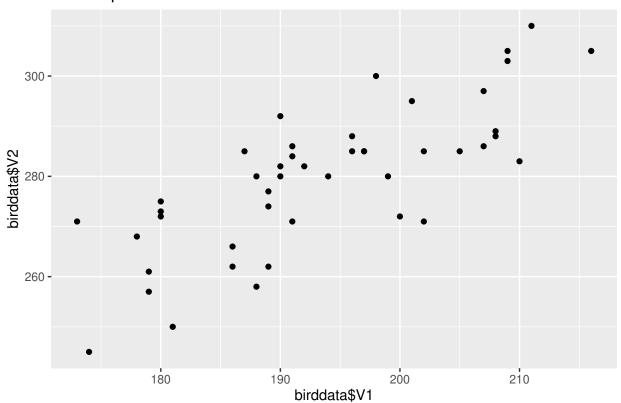
QQ plot for X2: Wing Length



Scat = ggplot()+
 geom_point(aes(x=birddata\$V1,y=birddata\$V2))+
 ggtitle(label = "Scatter plot X1 vs X2")



Scatter plot X1 vs X2



3 Comparison of Mean Vectors (one-way MANOVA)

We will look at a data set on Egyptian skull measurements (published in 1905 and now in heplots R package as the object Skulls). Here observations are made from five epochs and on each object the maximum breadth (mb), basibregmatic height (bh), basialiveolar length (bl) and nasal height (nh) were measured.

3.1 Exploring the Data

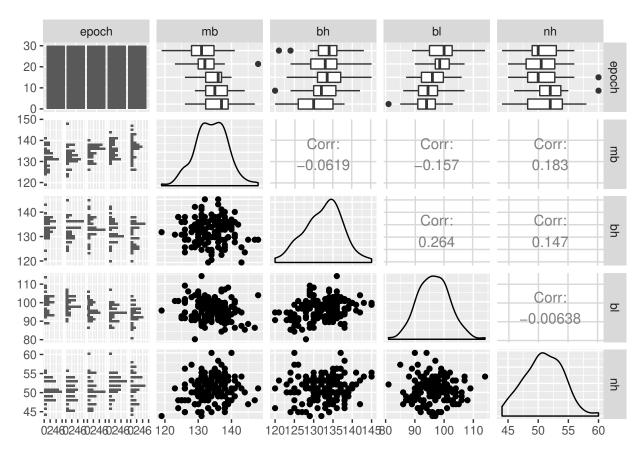
Explore the data first and present plots that you find informative.

Answer: We first take a glimpse at the data.

head(Skulls)

Next, we will look at the

ggpairs(Skulls)



Looking at the different distributions of our features we see that all of them are clustered around a mean. mb and bl seem to be symmetrical, whereas bh and nh seem not. Looking at the scatterplots and the corresponding correlations we seek that aprt from bh with mb and nh and bl all of them have a slight correlation. We also see that we have the same amount of epochs, so they're "distributed" normally. The epochs are:

```
unique(Skulls$epoch)
```

```
## [1] c4000BC c3300BC c1850BC c200BC cAD150
## Levels: c4000BC < c3300BC < c1850BC < c200BC < cAD150
```

Differing of Mean Vectors 3.2

Now we are interested whether there are differences between the epochs. Do the mean vectors differ? Study this question and justify your conclusions.

Task: The mean vectors do defer except nasal height. All other means are different between the epochs with a significante level between of 5 percent.

```
res = manova(cbind(mb, bh, bl, nh) ~ epoch, Skulls)
res
## Call:
##
          manova(cbind(mb, bh, bl, nh) ~ epoch, Skulls)
                                                                                                     Correct.
##
## Terms:
##
                                     epoch Residuals
                                                                                       You forgot to say that you should reject the hypothesis that all the in-group
                                  502.827 3061.067
## mb
                                                                                      means (per-epoch means) are equal.
You should do summary(res) and it will
give you the p-value described in Table
6.3 in the text book.
                                 229.907 3405.267
## bh
```

```
## bl
                  803.293 3505.967
                   61.200 1472.133
## nh
## Deg. of Freedom
                       4
                               145
## Residual standard errors: 4.59465 4.846091 4.917223 3.186321
## Estimated effects are balanced
summary.aov(res)
## Response mb :
##
               Df Sum Sq Mean Sq F value
                                           Pr(>F)
## epoch
               4 502.83 125.707 5.9546 0.0001826 ***
## Residuals 145 3061.07 21.111
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
##
   Response bh :
               Df Sum Sq Mean Sq F value Pr(>F)
##
              4 229.9 57.477 2.4474 0.04897 *
## epoch
## Residuals 145 3405.3 23.485
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
##
   Response bl :
##
               Df Sum Sq Mean Sq F value
                                          Pr(>F)
               4 803.3 200.823 8.3057 4.636e-06 ***
## epoch
## Residuals 145 3506.0 24.179
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
##
   Response nh :
               Df Sum Sq Mean Sq F value Pr(>F)
##
              4 61.2 15.300 1.507 0.2032
## epoch
## Residuals 145 1472.1 10.153
```

3.3 Confidence Intervals

If the means differ between epochs compute and report simultaneous confidence intervals. Inspect the residuals whether they have mean 0 and if they deviate from normality (graphically).

Tip: It might be helpful for you to read Exercise 6.24 of *Johnson*, *Wichern*. The function manova() can be useful for this question and the residuals can be found in the \$res field.

Answer: The means differ, so we calculate the confidence intervals for all groups and feature combinations. A row in teh result (combination, feature, group A and group B) is significant with $\alpha = 0.05$ if the confidence interval does **not** cover 0.

```
# k and l are the groups
# i is the feature

ci_custom = function(data, p, g, n, W, l, k, i, alpha = 0.05) {

# Define groups
group_k = unique(data$epoch)[k]
group_l = unique(data$epoch)[l]
group_data_k = data[data$epoch == group_k,]
```

```
group_data_1 = data[data$epoch == group_1,]
  n_k = nrow(group_data_k)
  n_1 = nrow(group_data_1)
  x_i = mean(group_data_k[,i+1]) - mean(group_data_l[,i+1])
  t_{crit} = qt(\frac{alpha/(p*g*(g-1))}{qt}, df = (n-g))
                                                        Should be qt(1 - alpha/(p*g*(g-1)), df=n-g). This way, you don't need the abs().
  var_i = sqrt(diag(W)[i]/(n-g) * (1/n_k + 1/n_l))
  abs_i = abs(t_crit * var_i)
  res = c(x_i - abs_i, x_i + abs_i)
  return(res)
scite = function(data, i) {
  # Static Parameters
  p = ncol(data) - 1
  g = length(unique(Skulls$epoch))
  n = nrow(data)
  # Calculation of W
  W = 0
  for (group in unique(Skulls$epoch)) {
    group_data = Skulls[Skulls$epoch == group,]
    S = sample_variance(group_data[,2:5])
    W = W + (nrow(group_data) - 1) * S
  df = data.frame()
  for (k in 2:length(unique(Skulls$epoch))) {
    for (1 in 1:max((k-1), 1)) {
      row = c(i, k, l, ci_custom(data=data, p=p, g=g, n, W=W, l=l, k=k, i=i))
      df = rbind(df, row)
  }
  colnames(df) = c("i", "k", "l", "lower", "upper")
  \#res = ci\_custom(data=data, p=p, g=g, n, \textit{W=W}, l=1, k=3, i=2)
  return(df)
}
```

You understand how to compute the intervals and what formula to use, so I am giving you a pass on this. But your numbers are slightly different from the correct ones. Please look for careless mistakes in your code.

Hint: The diagonal of the W matrix is the same as the "Residuals" column printed by `print(manova(...)) `.

```
df = scite(Skulls, 1)
df
     ikl
                 lower
                          upper
## 1 1 2 1 -2.83963613 4.839636
## 2 1 3 1 -0.73963613 6.939636
## 3 1 3 2 -1.73963613 5.939636
## 4 1 4 1 0.29369721 7.972969
## 5 1 4 2 -0.70630279 6.972969
## 6 1 4 3 -2.80630279 4.872969
## 7  1  5  1  0.96036387  8.639636
## 8 1 5 2 -0.03963613 7.639636
## 9 1 5 3 -2.13963613 5.539636
## 10 1 5 4 -3.17296946 4.506303
3.3.2 Variable i=2 (bh)
df = scite(Skulls, 2)
     i k l
               lower
                         upper
## 1 2 2 1 -4.949760 3.1497596
## 2 2 3 1 -3.849760 4.2497596
## 3 2 3 2 -2.949760 5.1497596
## 4 2 4 1 -5.349760 2.7497596
## 5 2 4 2 -4.449760 3.6497596
## 6 2 4 3 -5.549760 2.5497596
## 7 2 5 1 -7.316426 0.7830929
## 8 2 5 2 -6.416426 1.6830929
## 9 2 5 3 -7.516426 0.5830929
## 10 2 5 4 -6.016426 2.0830929
3.3.3 Variable i=3 (bl)
df = scite(Skulls, 3)
df
```

```
df

## i k l lower upper

## 1 3 2 1 -4.209203  4.0092027

## 2 3 3 1 -7.242536  0.9758694

## 3 3 3 2 -7.142536  1.0758694

## 4 3 4 1 -8.742536  -0.5241306
```

5 3 4 2 -8.642536 -0.4241306 ## 6 3 4 3 -5.609203 2.6092027

3.3.4 Variable i=4 (nh)

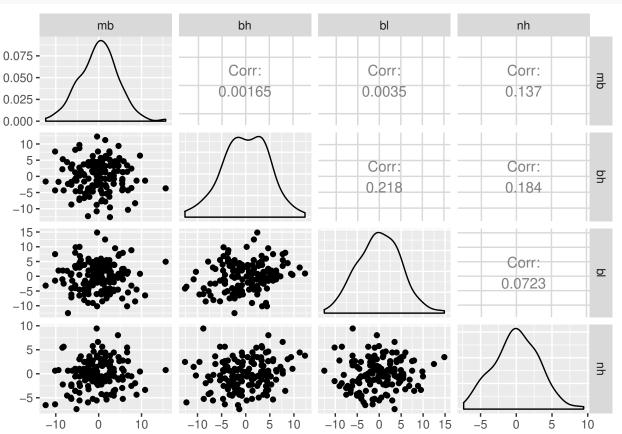
3.3.1 Variable i=1 (mb)

```
df = scite(Skulls, 4)
df
```

Hint: the correct values of this is (-9.846, -1.487)

Looking at the residuals we think that most of them look (almost) normal.

ggpairs(data.frame(res\$residuals))



4 Source Code

```
head(track_times)
sample_variance = function(X) {
  X = as.matrix(X)
  identity = diag(nrow(X))
  one_n = matrix(1, nrow=nrow(X), ncol=1)
  inter = identity - 1/nrow(X) * (one_n %*% t(one_n))
  return(1/nrow(X) * (t(X) %*% inter %*% X))
mahalanobis_distance = function(X) {
  X = as.matrix(X)
  V = sample_variance(X)
  ident = matrix(1, nrow=nrow(X), ncol=nrow(X))
  mu = 1/nrow(X) * (t(ident) %*% X)
  X_{centered} = X - mu
  return(diag(X_centered %*% solve(V) %*% t(X_centered)))
}
nu = ncol(track_times) - 1
nu = ncol(track_times) - 1
D = mahalanobis_distance(track_times[,2:8])
val = seq(0, 40, 0.01)
chi_sq_7 = dchisq(val, nu)
ggplot() +
  geom_histogram(aes(x = D, y=..density..),
                 color = "#755138", fill = "#D1CDC1",
                 bins = sqrt(nrow(track_times))) +
  geom_density(aes(x = D, y=..density..),
               color="#5C8240", fill="#8AB077", alpha = 0.2) +
  geom\_line(aes(x = val, y = chi\_sq\_7), color = "#2F2924") +
  labs(title = "Histrogram of 100m",
       y = "Density",
       x = "Time", color = "Legend") +
  scale_color_viridis(discrete=FALSE) +
  theme_minimal()
alpha = 0.001
```

```
outlier_indeces = 1 - pchisq(D, nu) < alpha</pre>
track_times$country[outlier_indeces]
alpha = 0.001 / nrow(track_times)
outlier_indeces = 1 - pchisq(D, nu) < alpha</pre>
track_times$country[outlier_indeces]
library(ellipse)
birddata = read.table("T5-12.DAT")
# Male hook-billed kites
n = nrow(birddata)
p = dim(birddata)[2]
mu = data.frame("mu1" = 190, "mu2" = 275)
meanVec = NULL
x_bar = NULL
meanMu = NULL
\# x - x_bar
x_bar = data.frame("X1" = mean(birddata$V1), "X2" = mean(birddata$V2))
sample_var = function(obs){
  X = as.matrix(obs)
  IdentityMat = diag(nrow(obs))
  one_n = matrix(1,nrow(X),nrow(X))
  # sample var formula from notes
  mix = (IdentityMat - (1/nrow(X)) *(one_n))
  sample_variance = (1/nrow(X)) * (t(X) %*% mix %*% X)
  return(sample_variance)
}
S = sample_var(birddata)
S = matrix(S, ncol=2, nrow=2)
eigenValVec = eigen(S)
meanMu = as.matrix(x_bar - mu)
# To check if mu is on the confidence region we need to check if t^2 is F_{n,n-p}
Tsq = n * (meanMu) %*% solve(S) %*% t(meanMu)
F_{val} = qf(0.95, df1=p, df=n-p) * (p*(n-1)/(n-p)) #5% significance level
cat("Is T^2 less than F_val? =",Tsq<F_val)</pre>
cat("\nThus, we do not reject the NULL Hypothesis(H_0)")
\# We conclude that it is in the region as \ Tsq is less than \ F\_val
axes_len = function(lam,n,p){
  sqrt(lam)* sqrt((p*(n-1))/ (n*(n-p)) * qf(0.95,df1=p, df2=(n-p)))
```

```
11=axes_len(lam=eigenValVec$values[1],n=n,p=p)
12=axes_len(lam=eigenValVec$values[2],n=n,p=p)
# plot(birddata)
\# lines(ellipse(mu, S, npoints = 200))
# dataEllipse(birddata, levels=.95)
# length(c(mu$mu1,mu$mu2))
rotate = function(a,b,c){
  res = list()
  if (b==0 \& a >= c){res$angle = 0}
  else if (b==0 \& a < c) \{ res sangle = (pi/2) \}
  else{ res$angle = atan2(eigenValVec$values[1]-a, b)}
  res\frac{1}{a+c} = \frac{a+c}{2} + \frac{((a-c)/2)^2}{b^2}
  res\frac{1am2}{(a-c)/2} - \frac{((a-c)/2)^2}{b^2}
  return(res)
\# S = matrix(c(4,-2,-2,4),byrow = T, nrow = 2)
res = rotate(S[1,1],S[1,2],S[2,2])
chi95 < -qchisq(.95,df=2)
axes95 <- sqrt(eigenValVec$values) * sqrt(chi95)</pre>
library(ggforce)
ggplot()+
  geom_point(aes(birddata$V1, birddata$V2))+
  geom_ellipse(aes(x0=mu$mu1,y0=mu$mu2, a=axes95[1], b=axes95[2], angle = res$angle ))
\# Simultaneous T2-intervals for the component means as shadows of the confidence ellipse on the axes
fsqrt = function(i){
  return(sqrt(p*(n-1)/((n-p)) * qf(0.95,df1=p, df=n-p)) * sqrt(S[i,i]/n))
}
CI = NULL
CI$Lower = (x_bar$X1) - fsqrt(1)
CI$Upper =(x_bar$X1) + fsqrt(1)
CI = as.data.frame(CI)
temp = c ((x_bar_X^2) - fsqrt(2), (x_bar_X^2) + fsqrt(2))
CI = rbind(CI, temp)
rownames(CI) = c("Mu_1_tail","Mu_2_wing")
\# \ kableExtra::kable(CI)
\# Bonferroni's Confidence intervals
set.seed(12345)
ben_alpha = 0.95
Ber_CI = NULL
```

```
Ber_CI$Lower = x_bar$X1 - abs(qt(0.05/2*p, df=n-1)) * sqrt(S[1,1]/n)
Ber_CI\$Upper= x_bar\$X1 + abs(qt(0.05/2*p, df=n-1)) * sqrt(S[1,1]/n)
Ber_CI = as.data.frame(Ber_CI)
temp = c(x_bar_{X2} - abs(qt(0.05/2*p, df=n-1)) * sqrt(S[2,2]/n),
         x_bar^{X2} + abs(qt(0.05/2*p, df=n-1)) * sqrt(S[2,2]/n))
Ber_CI = rbind(Ber_CI, temp)
rownames(Ber_CI) = c("Mu_1_tail", "Mu_2_wing")
Ber_CI
library(CARS)
qqPlot(birddata$V1, main ="QQ plot for X1: Tail length",id=F )
qqPlot(birddata$V2, main="QQ plot for X2: Wing Length",id=F )
Scat = ggplot()+
  geom_point(aes(x=birddata$V1,y=birddata$V2))+
  ggtitle(label = "Scatter plot X1 vs X2")
plot(Scat)
head(Skulls)
ggpairs(Skulls)
unique(Skulls$epoch)
res = manova(cbind(mb, bh, bl, nh) ~ epoch, Skulls)
res
summary.aov(res)
\# k and l are the groups
# i is the feature
ci_custom = function(data, p, g, n, W, l, k, i, alpha = 0.05) {
  # Define groups
  group_k = unique(data$epoch)[k]
  group_l = unique(data$epoch)[1]
  group_data_k = data[data$epoch == group_k,]
  group_data_1 = data[data$epoch == group_1,]
  n_k = nrow(group_data_k)
  n_1 = nrow(group_data_1)
  x_i = mean(group_data_k[,i+1]) - mean(group_data_l[,i+1])
  t_{crit} = qt(alpha/(p*g*(g-1)), df = (n-g))
  var_i = sqrt(diag(W)[i]/(n-g) * (1/n_k + 1/n_l))
  abs_i = abs(t_crit * var_i)
  res = c(x_i - abs_i, x_i + abs_i)
```

```
return(res)
scite = function(data, i) {
  # Static Parameters
  p = ncol(data) - 1
  g = length(unique(Skulls$epoch))
  n = nrow(data)
  # Calculation of W
  for (group in unique(Skulls$epoch)) {
    group_data = Skulls[Skulls$epoch == group,]
   S = sample_variance(group_data[,2:5])
   W = W + (nrow(group_data) - 1) * S
  df = data.frame()
  for (k in 2:length(unique(Skulls$epoch))) {
   for (1 in 1:max((k-1), 1)) {
     row = c(i, k, l, ci_custom(data=data, p=p, g=g, n, W=W, l=l, k=k, i=i))
      df = rbind(df, row)
   }
  }
  colnames(df) = c("i", "k", "l", "lower", "upper")
  \#res = ci\_custom(data=data, p=p, g=g, n, W=W, l=1, k=3, i=2)
  return(df)
}
df = scite(Skulls, 1)
df
df = scite(Skulls, 2)
df
df = scite(Skulls, 3)
df
df = scite(Skulls, 4)
df
```

ggpairs(data.frame(res\$residuals))