Individual Run Sheet - DAB

	d ID:			
	ocker: mary:			
	condary:			
1.	Incubate sections in temperature.	25% methanol so	lution for 30 minut	es at room
		Methanol	dH₂O	30% H ₂ O ₂
	1 tray (200 mL)	<mark>50 mL</mark>	150 mL	6 mL
2.	Wash sections in 1x	PBS for 5x6 minut	es. 🗆 🗆 🗆 🗆	
3.) +	- 0.4% PBS-Tx for 1 hour
	at room temperatur 1 mL solution	e.]
	per section	Blocker	0.4% PBS-Tx	
	1 section	0.1 mL	1 mL	
	sections	mL	mL	
4.	DO NOT RINSE			
5.	Incubate sections in primary () (dilution 1:
	+ 0.4% PBS-Tx for hours at 4°C on the shaker.			
	1 mL solution per section	Primary	0.4% PBS-Tx	
	1 section	μΙ	1 mL	
	sections	μΙ	mL	
6.	Wash sections in 1x	PBS for 5x6 minut	es. 🗆 🗆 🗆 🗆	•
7.	Incubate sections in	secondary () (dilution 1:
	+ blocker + 0.4% PB3	S-Tx for hour	· ·	ure.
	1 mL solution	Secondary	Blocker	0.4% PBS-Tx

per section			
1 section	μΙ	0.025 mL	1 mL
sections	μΙ	mL	mL

8. Wash sections in 1x PBS for 5x6 minutes. \square \square \square \square

9. Incubate sections in **ExtrAvidin** (1:1000) + 0.4% PBS-Tx for 30 minutes at room temperature.

1mL solution per 1 section	ExtrAvidin	0.4% PBS-Tx
1 section	1 μΙ	1 mL
sections	μΙ	mL

10. Wash sections in 1x PBS for 5x6 minutes. □ □ □ □ □

11. Prepare 4 trays with 1x PBS (20 mL each) for rinsing after DAB reaction.

12. Incubate sections in 0.025% DAB solution **without** 30% H_2O_2 for 12 minutes at room temperature.

	DAB	0.1M PBS	CoCl ₂
1 tray (200 mL)	2 aliquots (0.05g)	200mL	0.012g

Mix 0.1M PBS and CoCl₂ well before adding DAB aliquots.

13. Lift tissue out of tray and add $0.005\%~H_2O_2$ to DAB solution, stir quickly and place tissue back into tray.

	H ₂ O ₂	dH₂O
1 tray (200mL)	0.060mL	6mL

14. Allow tissue to react for 30s to 90s, depending on background. Once reaction is done, immediately place in a tray with 1x PBS (200mL) for a quick rinse.

15.	Wash sections in 1x PBS for 30 seconds, 1 minute, 2 minutes, and 6 minutes (x	(2)
	30s: □ □ 1min: □ □ 2min: □ □ 6min: □ □	

CLEAN ALL LAB SURFACES WITH BLEACH TO NEUTRALIZE DAB.